


Cite this: *RSC Adv.*, 2021, 11, 14700

# Highly sensitive and selective colorimetric detection of dual metal ions ( $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ ) in water: an eco-friendly approach†

Rintumoni Paw,<sup>ac</sup> Moushumi Hazarika,<sup>a</sup> Purna K. Boruah,<sup>d</sup>  
Amlan Jyoti Kalita,<sup>b</sup> Ankur K. Guha,<sup>b</sup> Manash R. Das<sup>d</sup>  
and Chandan Tamuly<sup>\*ac</sup>

Application of an alliin-based precursor for the synthesis of silver nanoparticles (Ag NPs) which is an emerging, reliable and rapid sensor of heavy metal ion contaminants in water is reported here. The Ag NPs were characterized by using UV-visible spectroscopy, X-ray diffraction, transmission electron microscopy, X-ray photoelectron spectroscopy and Raman spectroscopy analysis techniques. The Ag NPs simultaneously and selectively detect  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions from aqueous solution. The sensitivity and selectivity of the prepared Ag NPs towards other representative transition-metal ions, alkali metal ions and alkaline earth metal ions were also studied. For more precise evidence, a density functional theory study was carried out to understand the possible mechanism and interaction in the detection of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  by Ag NPs. The limits of detection for  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions were found as 15.7 nM and 11.25 nM, respectively. This assay indicates the possible use of garlic extract-synthesized Ag NPs for sensing  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  in aqueous solution very significantly. So, the simple, green, eco-friendly and easy method to detect the dual metal ions may further lead to a potential sensor of heavy metal ion contaminants in water of industrial importance.

Received 23rd November 2020  
Accepted 29th March 2021

DOI: 10.1039/d0ra09926k

rsc.li/rsc-advances

## Introduction

Heavy metals usually refer to elements with atomic weight and density higher than 5 times that of water. Some of these metallic compounds are considered highly toxic to humans and other organisms. Extensive use of these metals or their compounds in industrial, agricultural, domestic and health sectors releases them in the environment, raising health and environmental concerns.<sup>1</sup> Mercury (Hg) is a heavy metal that is reported to be hazardous in some forms. However, metallic Hg is still used in thermometers, barometers, batteries, electrical appliances, dental fillings, some religious remedies *etc.* Some inorganic mercury compounds are used as fungicides, cosmetics, antiseptics, paints, tattoo dyes *etc.*<sup>2,3</sup> All Hg compounds that are discharged in the environment ultimately end up in soil or surface water.<sup>1</sup> Likewise, plant products

may also contain Hg derived from some sprays with Hg compounds as one of the components.<sup>4</sup>

Elemental Sn has significant applications with other elements like Al, Fe, Cu *etc.* in brass, bronze and solder. One of the major applications of Sn compounds is as intermediate materials for the synthesis of organo-Sn compounds and for electroplating composites. Organo-Sn compounds are extensively used as stabilizing agents in the preparation of materials like PVC, as homogenous catalysts for polyurethane foam formation, for glass coating, and as pesticides and disinfectants. However, use of certain organo-Sn compounds is discontinued due to their toxicity and hazardous nature in the environment.<sup>5,6</sup> Naturally occurring Me-Sn are formed from inorganic Sn by the action of mainly soil- and sediment-dwelling bacteria and other microorganisms.<sup>7,8</sup>

Stannous chloride ( $\text{SnCl}_2$ ) is most commonly available in packed food, cold beverages, pesticides, toothpaste *etc.* It interrupts ossification in the fetal skeleton and may cause complete post-implantation loss.<sup>9</sup> Certain organo-Sn compounds like trimethyl-Sn and triethyl-Sn are reported as highly neurotoxic.<sup>10</sup> Butyl-Sn and octyl-Sn are known to impair the immune system in different animal groups. Likewise, anaemia is reported in rats with oral exposure to some of the organo-Sn compounds like dibutyltin dichloride, tributyltin oxide, or dioctyltin dichloride which also causes teratogenic effects in rat embryo and induces carcinogenicity.<sup>11</sup>

<sup>a</sup>Natural Product Chemistry Section, CSIR-North East Institute of Science and Technology, Itanagar, Arunachal Pradesh-791110, India. E-mail: c.tamuly@gmail.com

<sup>b</sup>Dept of Chemistry, Cotton University, Guwahati, Assam-781001, India

<sup>c</sup>Academic of Scientific & Innovative Research, CSIR-North East Institute of Science and Technology, Jorhat, Assam-785006, India

<sup>d</sup>Material Science and Technology Division, CSIR-North East Institute of Science and Technology, Jorhat, Assam-785006, India

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra09926k



Therefore, monitoring of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ion level which varies with source is important to protect human and environmental health. Many techniques including HPLC, electrochemical, fluorescence and colorimetric methods are used to detect heavy and hazardous metals in the environment and water.<sup>12–14</sup> Many of these can detect even trace amounts of contaminants. But, these are expensive and need complicated setups or involve tedious processes. Colorimetric sensors are cheaper with comparatively easier processing methods.<sup>15</sup>

Developing a robust system for detecting  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  with high sensitivity and selectivity against other common metal ions dissolved in water has been a challenge in recent years. From an environmental point of view, the development of a low-cost, simple, selective and sensitive technique of detection of these ions becomes very important. Therefore, the present study attempts to assess selective and sensitive detection of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions using phyto-synthesized silver nanoparticles (Ag NPs). Here, garlic (*Allium sativum*), a common spice and medicinal plant, was used to synthesize the Ag NPs. Garlic is widely consumed as spice and medicine throughout the world in many cultures and traditions over centuries of human civilization. It is also used in Ayurvedic medicine for treatment of different diseases.<sup>16</sup> Garlic contains various organo-S compounds, responsible for its specific flavour, aroma and medicinal properties. Alliin is the major S compound present in garlic. The alliin content in garlic cloves is reported as  $\sim 8 \text{ g kg}^{-1}$ .<sup>17</sup> Garlic has several advantages compared to other reducing agents. It is commonly available throughout the year and widely available. It is also available worldwide. It is easy to preserve at normal temperature. It has a significant advantage compared to other biomaterials for reduction of  $\text{Ag}^+$  ions. However, it is transformed to allicin(diallylthiosulfinate) rapidly by the action of the enzyme alliinase after crushing or cutting garlic cloves.<sup>18,19</sup> Accordingly, hot extraction was carried out of garlic paste in our study to avoid this transformation at about  $60^\circ\text{C}$ .

Here, it is proposed to synthesize Ag NPs using garlic extract that contains alliin as the major bioactive ingredient. The synthesized Ag NPs were further used for highly selective sensing of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  metal ion contaminants in water. In addition, a density functional theory (DFT) study was carried out to support the experimental evidence and to reveal a possible mechanism for the sensing of the metal ions. For precise application or practical utility of our method, the experiment was done for water samples collected from three ponds of different locations of Arunachal Pradesh.

## Results and discussion

### Formation of Ag NPs

The formation of Ag NPs was characterized by using UV-visible spectra. The Ag NPs were synthesized using garlic extract (Fig. 1). After addition of garlic extract, the colourless  $\text{AgNO}_3$  solution turned to a brownish colour. This indicates the action of the garlic extract as a reducing agent to form Ag NPs.<sup>20</sup> The absorption spectrum of the Ag NPs was bell-shaped with a peak at a wavelength of 420 nm (Fig. 2A). This band corresponds to the typical surface plasmon resonance (SPR) of conducting electrons from the

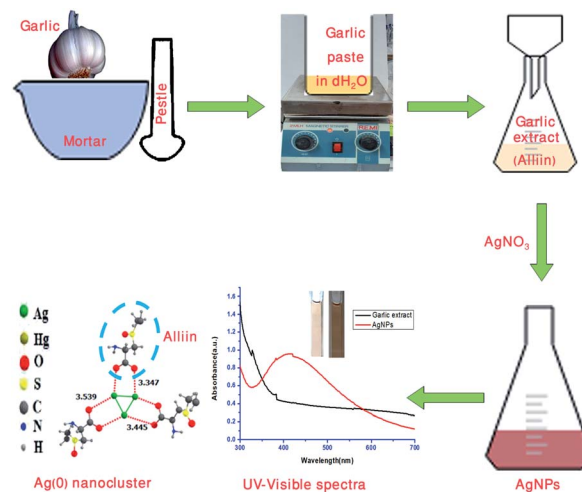


Fig. 1 Schematic diagram of formation of Ag NPs from garlic extract.

surface of Ag NPs and was used to confirm the reduction of  $\text{Ag}^+$  to  $\text{Ag}^0$ .<sup>21–24</sup> The SPR peak for synthesized Ag NPs was also reported at a wavelength of 420 nm in previous studies.<sup>25,26</sup>

### X-ray diffraction (XRD)

The crystalline nature of the Ag NPs was characterized by using the XRD technique. There were four well-defined characteristic

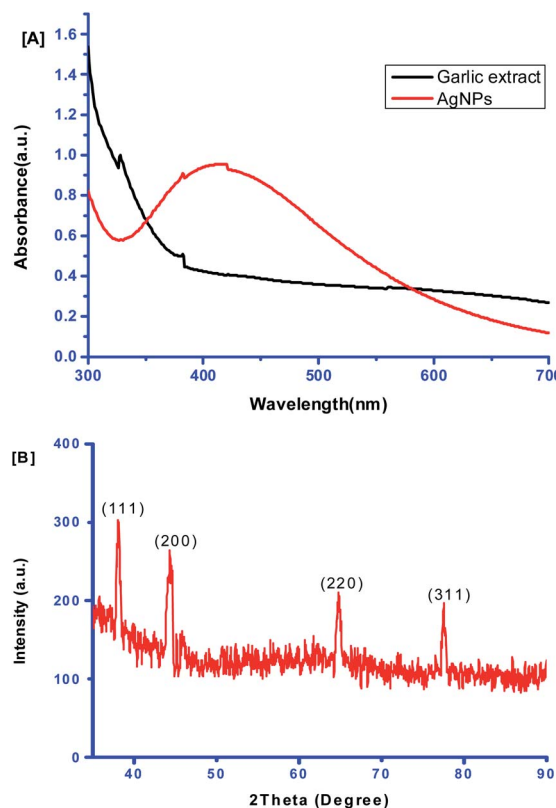


Fig. 2 (A) UV-visible spectra and (B) XRD pattern of formation of Ag NPs.



diffraction peaks at  $2\theta$  values of  $38.16^\circ$ ,  $44.54^\circ$ ,  $64.82^\circ$  and  $77.56^\circ$  (Fig. 2B). These characteristic peaks revealed the reflection planes (111), (200), (220) and (311) of face-centred cubic (FCC) structure of crystalline metallic Ag as per JCPDS file no. 04-0783.<sup>27,28</sup> This is also supported by our previously reported results.<sup>22</sup> In the presence of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions the peaks at  $38.16^\circ$ ,  $44.54^\circ$ ,  $64.82^\circ$  and  $77.56^\circ$  disappear due to the disappearance of Ag NPs. This is shown in Fig. S1 [ESI<sup>†</sup>].

### Raman spectroscopy

The Raman spectrum of the Ag NPs was recorded to find out the possible surface-bound functional groups of the active compounds in garlic extract (alliin) associated with the stabilization of Ag NPs (Fig. 3). The broad band at  $1580\text{ cm}^{-1}$  revealed asymmetric C=O stretching vibrations of carboxyl (O=C=O) group. This peak indicates that the active molecule of garlic extract is bound to the surface of the Ag NPs through formation of chemical bond between Ag and carboxylate ( $-\text{COO}^-$ ) group of alliin. The shoulder peak at  $2350\text{ cm}^{-1}$  may be due to C=C stretching.<sup>29,30</sup> Raman analysis was also carried out for Ag NPs +  $\text{Hg}^{2+}$  and Ag NPs +  $\text{Sn}^{2+}$  mixtures. The spectra are shown in Fig. 3. The peak at  $1585\text{ cm}^{-1}$  for Ag NPs +  $\text{Hg}^{2+}$  and Ag NPs +  $\text{Sn}^{2+}$  revealed the presence of C=O stretching vibration of carboxylate ion. The peak  $2350\text{ cm}^{-1}$  may be due to C=C stretching of alliin molecule. The peak is sharper compared to that of Ag NPs synthesized *via* alliin. This may be due to surface alliin molecules becoming more labile and  $\text{Ag}^0$  state disappearing in the presence of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ .<sup>31,32</sup>

### Transmission electron microscopy (TEM) analysis

The Ag NPs were further characterized by TEM analysis. The TEM analysis revealed that the Ag NPs are spherical in shape and with sizes in the range  $2.0 \pm 0.3$  to  $17.6 \pm 0.6\text{ nm}$ . The average size of the NPs is  $10.3\text{ nm}$ . This is supported by reported literature.<sup>21–24</sup> Fig. 4A indicates the presence of spherical Ag NPs in the system. The inset of Fig. 4A shows the HR-TEM image with a scale of  $2\text{ nm}$  which reveals the spherical surface pattern of Ag NPs. The diffraction pattern of the Ag nanocrystal surface

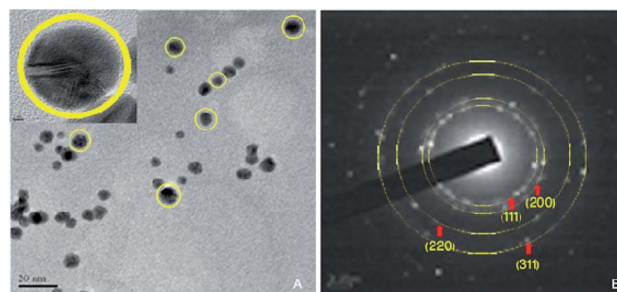


Fig. 4 (A) TEM image of Ag NPs with scale bare of  $20\text{ nm}$ . (B) Diffraction pattern of Ag NPs.

is shown in Fig. 4B. The diffraction pattern reveals the (111), (200), (220) and (311) planes of Ag crystalline surface.<sup>21,22</sup>

### X-ray photoelectron spectroscopy (XPS)

XPS was used to interpret the chemical nature and surface constituents of the Ag NPs synthesized using aqueous garlic extract. The survey scan shows the existence of Ag, O, C, S and N elements. The atomic composition of elements was found as: C  $1s - 58.78\%$ , O  $1s - 34.4\%$ , N  $1s - 5.51\%$ , Ag  $3d - 0.37\%$  and S  $2p - 0.05\%$ . Fig. 5A presents an XPS spectrum of the Ag NPs with a range of  $0-1300\text{ eV}$ . The HR-XPS core-level spectra of S(2s), C(1s), Ag(3d), N(1s), and O(1s) are shown in Fig. 5B–F. The Ag 3d region shows two characteristic peaks at  $372.9\text{ eV}$  and  $366.9\text{ eV}$  (Fig. 5B) that occur due to the spin orbital splitting representing Ag  $3d_{3/2}$  and Ag  $3d_{5/2}$  core levels, respectively.<sup>33,34</sup> The result

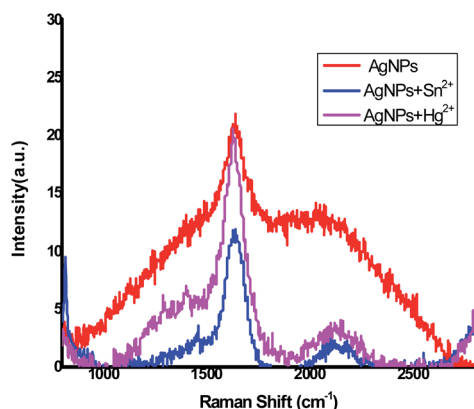


Fig. 3 Raman spectra of Ag NPs, AgNPs +  $\text{Hg}^{2+}$  and AgNPs +  $\text{Sn}^{2+}$ .

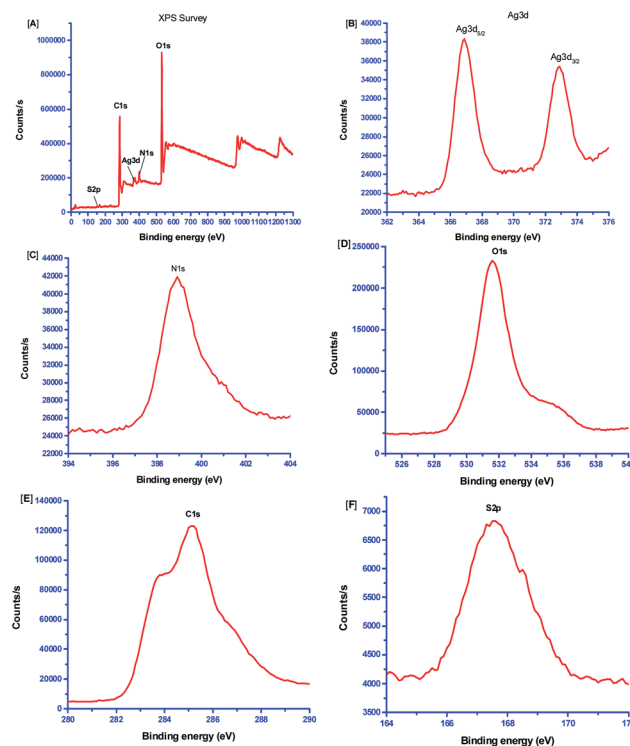


Fig. 5 (A) XPS spectrum of Ag NPs and high-resolution XPS spectra of (B) Ag 3d core levels, (C) N (1s), (D) O (1s), (E) C (1s), (F) S (2p) regions.



suggests the formation of Ag NPs. Fig. 5C presents the HR-XPS spectrum for the N(1s) region. The binding energy was recorded at 398.9 eV. Fig. 5D presents the HR-XPS spectrum of the O(1s) region. Here, a peak at 532.2 eV was observed for the Ag–O bond.<sup>35</sup> The Ag–O bond was formed due to strong interaction between a vacancy in the d-orbital of Ag and the active biomolecule present in the extract of *Allium sativum*. The peak seen at binding energy of 531.6 eV shows the existence of S=O and O=C=O bonds.<sup>34</sup> Fig. 5E depicts the C(1s) spectrum that refers to bonds with C atoms. Here, peaks were noted at 283.8 eV and 285.2 eV ascribed to C–O and C=O, respectively.<sup>34</sup> Fig. 5F presents the S(2p) spectrum showing the existence of S at 167.5 eV for S 2p<sub>3/2</sub>. The binding energy of S corresponds to established data for S 2p.<sup>36</sup>

### Metal ion sensitivity of the Ag NPs

The aqueous solution of the synthesized Ag NPs was brown in colour which may be ascribed to the excited vibrations of the SPR band. The SPR peak was recorded at 420 nm. The colour of the freshly prepared Ag NPs solution turns from brown to colourless gradually on addition of Hg<sup>2+</sup> and Sn<sup>2+</sup> ions. Response of Ag NPs to other metal ions did not show changes in the SPR band (Fig. 6). The metal ions Fe<sup>2+</sup>, Ni<sup>2+</sup>, Pd<sup>2+</sup>, Pb<sup>2+</sup>, Na<sup>+</sup>, Al<sup>3+</sup>, Zn<sup>2+</sup>, K<sup>+</sup>, Cu<sup>2+</sup>, Ca<sup>2+</sup>, Ba<sup>2+</sup>, Sn<sup>2+</sup> and Hg<sup>2+</sup> were taken to evaluate the sensitivity of Ag NPs. But, except Hg<sup>2+</sup> and Sn<sup>2+</sup>, the other metal ions did not show any sensitivity towards the as-synthesized Ag NPs. In this case, 0.2 ml of 1 mM salt solution of metal ion was added to a 3 ml solution of Ag NPs at similar experimental conditions. Fresh solution of Ag NPs was used for detecting the metal ions. The study revealed that the colour of the Ag NPs solution turns from brownish to colourless on addition of freshly prepared Hg<sup>2+</sup> and Sn<sup>2+</sup> ions.

The results show that the presence of standard solution of Fe<sup>2+</sup>, Ni<sup>2+</sup>, Pd<sup>2+</sup>, Pb<sup>2+</sup>, Na<sup>+</sup>, Al<sup>3+</sup>, Zn<sup>2+</sup>, K<sup>+</sup>, Cu<sup>2+</sup>, Ca<sup>2+</sup> and Ba<sup>2+</sup> ions does not cause apparent changes in the SPR band and colour of Ag NPs solution, in contrast to Hg<sup>2+</sup> and Sn<sup>2+</sup> that turn the colour of the solution from brown to colourless. Experiments to detect Hg<sup>2+</sup> and Sn<sup>2+</sup> ions were performed using UV-

visible spectroscopic and XRD methods. The change in colour with respect to time is shown in Fig. 6. The colour of the solution is yellowish brown in the absence of Hg<sup>2+</sup> and Sn<sup>2+</sup> ions. The Hg<sup>2+</sup> ion has a closed-shell d<sub>10</sub> and this configuration does not exhibit significant optical spectroscopic property. In the presence of 10 µl of 1 mM Hg<sup>2+</sup> and Sn<sup>2+</sup> ions the colour of the Ag NPs solution disappears. This is accompanied by broadening of the SPR band. The colourless solution did not show the peaks at 38.04°, 45.82°, 64.62° and 77.4° corresponding to the reflection planes (111), (200), (220) and (311) of the FCC structure of Ag NPs [Fig. S1, ESI†]. This indicates that the Ag NPs disappeared in the presence of Hg<sup>2+</sup> and Sn<sup>2+</sup> ions.

### Possible mechanism of Ag NPs synthesis and binding with Hg<sup>2+</sup> and Sn<sup>2+</sup> ions

In present study, the garlic paste was boiled immediately after grinding. This destroyed the enzyme activity of alliinase to convert alliin to allicin.<sup>17,18</sup> Therefore, alliin was involved as stabilizing agent of the synthesized Ag NPs. In this regard, HPLC analysis was done for further confirmation about the presence of major bioactive constituent (alliin) which may play a major role in synthesis and stabilization of the Ag NPs. The HPLC results clearly revealed the presence of alliin instead of allicin in this study (Fig. 7A and B). The peak at retention time of 2.45 min in Fig. 7A revealed the presence of alliin in garlic extract as major constituent. The HPLC trace of standard alliin is shown in Fig. 7B where the retention time is found at 2.45 min. In both cases the HPLC was run with the solvent system consisting of A : B (80 : 20) (acetonitrile : water with 0.2% phosphoric acid).

Hence, alliin is the major chemical constituent of hot aqueous garlic extract. So, it plays significant role in the

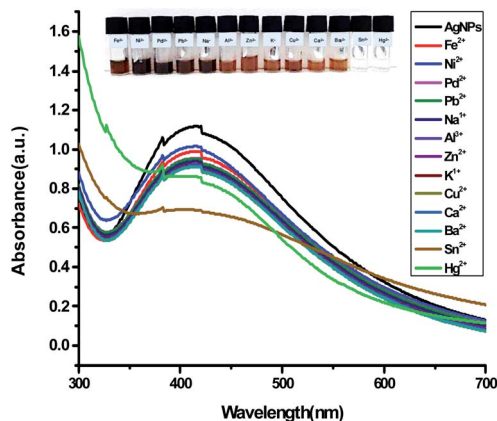


Fig. 6 UV-visible spectra of Ag NPs after addition of different metal ion solutions. The colour of the solutions is shown in the inset photograph.

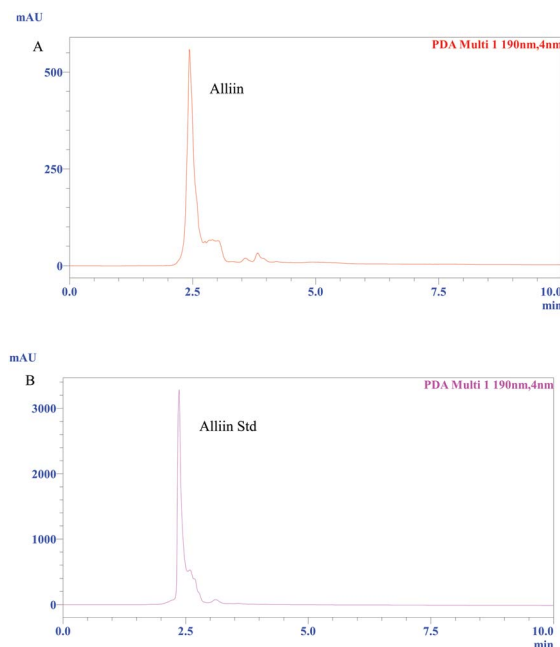


Fig. 7 HPLC traces of (A) garlic extract and (B) alliin standard.



synthesis of Ag NPs as reported in Fig. 8. Here, one  $\text{Ag}^+$  ion directly attaches with one carboxylate ion of alliin which may further form a cluster of three  $\text{Ag}^0$  ions with three carboxylate ions of alliin. After addition of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions, the solution becomes colourless. This results in degradation of the Ag NPs in the solution as indicated by UV-visible spectroscopy and XRD study. Based on these observations, it may be stated that these phenomena may be due to a redox reaction between  $\text{Ag}^0$  and  $\text{Hg}^{2+}$  with standard potentials of 0.8 eV ( $\text{Ag}^+/\text{Ag}$ ) and 0.85 eV ( $\text{Hg}^{2+}/\text{Hg}$ ), respectively.<sup>37</sup> Similar phenomena may also be attributed to  $\text{Sn}^{2+}$  ion. It is also expected that added  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions to the freshly prepared Ag NPs replaced the stabilizing compound alliin.

For more precise evidence, a DFT study was carried out to understand the possible interaction of alliin with Ag ions during formation of Ag NPs. A similar study was also conducted for analysing the optimized nanocluster in sensing  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions. The calculation reveals that the interaction of  $\text{Ag}^0$  nanocluster with alliin stabilizes the cluster. The calculated stabilization energy is 18.4 kcal mol<sup>-1</sup>. This energy is sufficient to hold together the  $\text{Ag}^0$  nanocluster. This nanocluster also interacts with  $\text{Hg}^0$  in its oxidized form, i.e.,  $\text{Hg}^{2+}$  state. Fig. 9 shows the optimized geometry of the  $\text{Ag}^+$  nanocluster interacting with each of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ . It is also to be noted that while interacting with  $\text{Hg}^{2+}$ , alliin separates from the  $\text{Ag}^+$  nanocluster. The interaction energy of  $\text{Ag}^+$  nanocluster with  $\text{Hg}^{2+}$  is calculated to be 7.6 kcal mol<sup>-1</sup> while it is 8.3 kcal mol<sup>-1</sup> for  $\text{Sn}^{2+}$ .

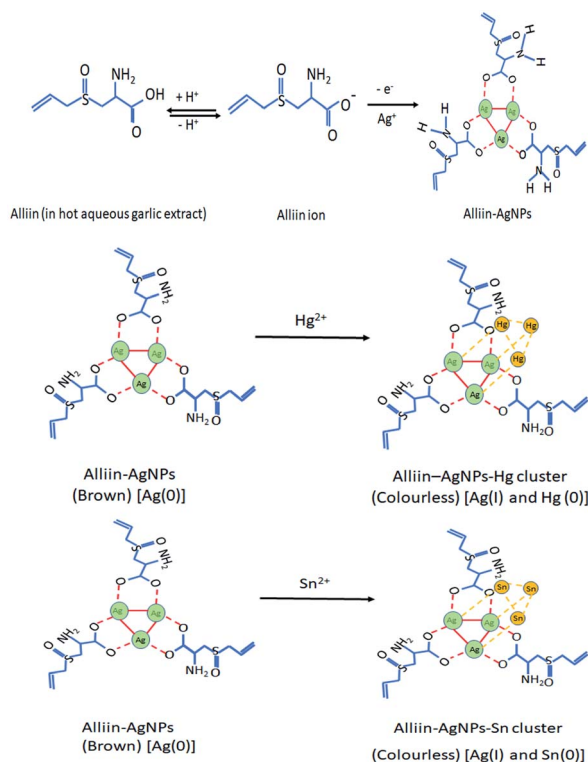


Fig. 8 Schematic illustration of proposed synthesis of Ag NPs using hot aqueous garlic extract and analytical detection of mercury ion ( $\text{Hg}^{2+}$ )/stannous ion ( $\text{Sn}^{2+}$ ) using synthesized Ag NPs.

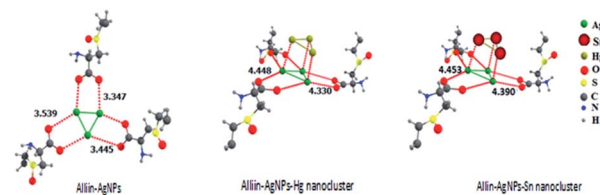


Fig. 9 Optimized geometry of  $\text{Ag}(0)$  nanocluster stabilized by alliin ion and the  $\text{Ag}(\text{I})$ - $\text{Hg}(0)$  and  $\text{Ag}(\text{I})$ - $\text{Sn}(0)$  interactions. Kinetics for binding Ag NPs with  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ .

Similar phenomena are also expected for interaction between Ag NPs and the  $\text{Sn}^{2+}$  ion.

To estimate the kinetics of the binding of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions with Ag NPs, we studied the time-dependent absorbance from the UV-visible absorption spectra of the Ag NPs in the presence of different concentrations of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions. We recorded the time-dependent UV-visible spectra at intervals of every 5 min. The absorbance of Ag NPs +  $\text{Hg}^{2+}$  and Ag NPs +  $\text{Sn}^{2+}$  was found to decrease with increasing time (Fig. 10A and B) in similar reaction conditions. This also corresponds to a decrease

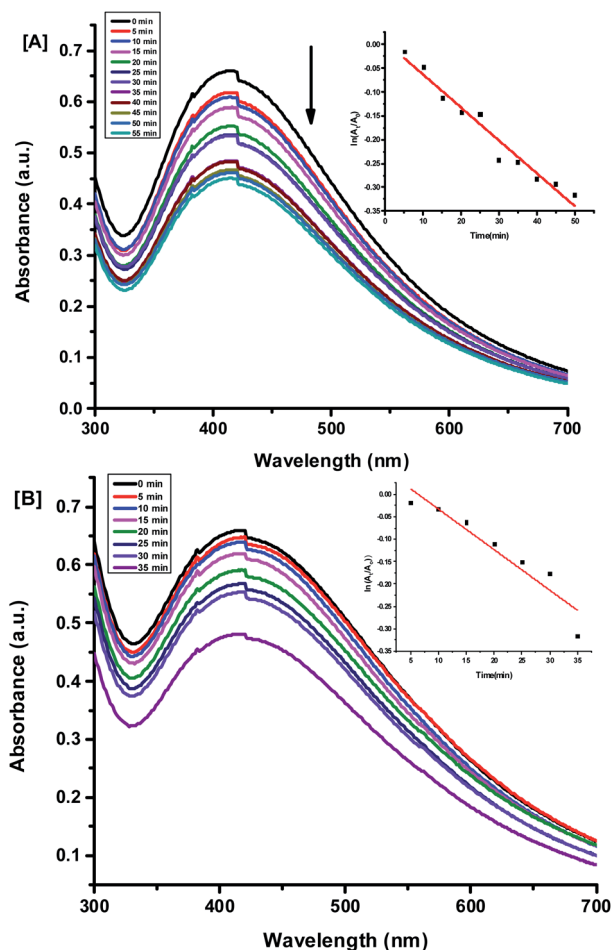


Fig. 10 Effect of kinetics on UV-visible absorption spectra of Ag NPs in the presence of (A) 5 nM  $\text{Hg}^{2+}$  and (B) 5 nM  $\text{Sn}^{2+}$  and at time intervals of 5 minutes.

**Table 1** Rate constant for binding of Ag NPs with  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  at different concentrations

No.	Concentration	Rate constant ( $K$ ) ( $\text{min}^{-1}$ )
1	10 nM $\text{Hg}^{2+}$	0.01434
2	5 nM $\text{Hg}^{2+}$	0.00689
3	1 nM $\text{Hg}^{2+}$	0.00508
4	10 nM $\text{Sn}^{2+}$	0.00907
5	5 nM $\text{Sn}^{2+}$	0.00791
6	1 nM $\text{Sn}^{2+}$	0.00674

of the rate constant (Table 1) with decreasing concentration of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions. When the concentration of metal ion increases, the Ag NPs released the bioactive compound, *i.e.* alliin, simultaneously, then formed a cluster with  $\text{Ag}^0$  ion. The mechanism has already been discussed in the previous section. The same phenomenon was observed in the case of  $\text{Sn}^{2+}$  ions.

### Effect of pH in sensing of $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ ions

The effect of pH in sensing the metal ions, *i.e.*  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ , was investigated in Ag NPs solution at 4.0, 7.0 and 9.0, respectively. At pH = 4.0 the Ag NPs solution turned colourless faster after addition of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ . This was indicated by a rapid decrease in absorbance of the solution. At pH = 9.0 the Ag NPs solution became colourless slowly compared to at pH 4.0 and 7.0 for both metal ions. The results indicated a decrease in rate of the reaction with increase of pH. The pH-specific colorimetric change was also noted for other Ag NPs.<sup>38</sup> The rate constant of the reaction is shown in Table 2.

### Limit of detection (LoD) of $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ ions by Ag NPs

The sensitivity and the minimum detectable concentration of  $\text{Hg}^{2+}$  in aqueous solution were evaluated by colour change of the system, and monitoring the UV-visible absorption. Different concentrations of aqueous solutions of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions were added to a solution of the Ag NPs (3 ml). The experiment was carried out at room temperature (25 °C). The LoD for  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  was determined using the SPR peak of Ag NPs at 420 nm.

From Fig. 11 and 12, it was observed that the absorption peak of Ag NPs decreases with increasing concentration of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions and there is a linear relationship ( $Y = 0.0366X + 0.2301$ ,  $R^2 = 0.8821$ ; and  $Y = 0.0234X + 0.1170$ ,  $R^2 = 0.9386$ , respectively) between the changes of absorbance and the

**Table 2** Rate constant for binding of Ag NPs with  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  at different pH

No.		pH	Rate constant ( $K$ ) ( $\text{min}^{-1}$ )
1	$\text{Hg}^{2+}$	4.0	0.00924
2		7.0	0.00621
3		9.0	0.00168
4	$\text{Sn}^{2+}$	4.0	0.00853
5		7.0	0.00560
6		9.0	0.00147

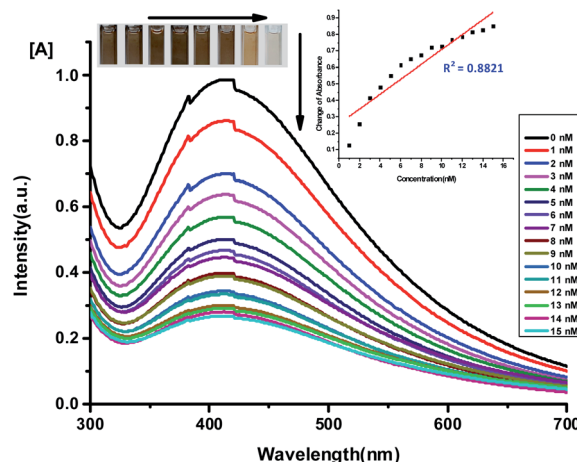


Fig. 11 UV-visible absorption spectra of Ag NPs upon addition of  $\text{Hg}^{2+}$  solution of various concentrations (1.0–16 nM). The inset shows the absorbance versus concentration of  $\text{Hg}^{2+}$ .

concentration of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions over the range 1–15 nM at 403 nm and 1–11 nM at 405 nm, respectively (Fig. 11 and 12). The synthesized Ag NPs can be used for the colorimetric detection of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  with LoD of 15.70 nM and 11.25 nM, respectively, which are very significant results compared to reported data (Tables 3 and 4).

### Determination of $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ in water samples

To study the practical utility of the developed method for sensing and detection of heavy metal ion contaminants in water, three water samples were collected from ponds of different locations of Arunachal Pradesh. Detection of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  in pond water with added  $\text{Fe}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Pd}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Al}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{K}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Ca}^{2+}$  and  $\text{Ba}^{2+}$  ions showed mean absorbance of  $0.486 \pm 0.007$  nm for solution with added  $\text{Hg}^{2+}$ ,  $0.3789 \pm 0.005$  nm for solution with added  $\text{Sn}^{2+}$ , and  $0.4884 \pm 0.0089$  for solution containing both  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ . The relative standard

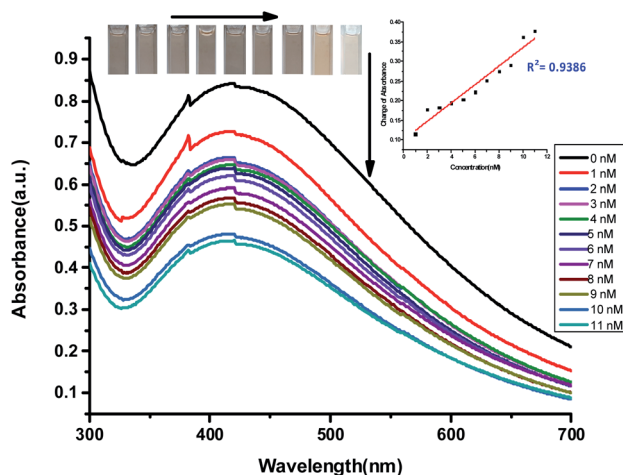


Fig. 12 UV-visible absorption spectra of Ag NPs upon addition of  $\text{Sn}^{2+}$  solution of various concentrations (1.0–11 nM). The inset shows the absorbance versus concentration of  $\text{Sn}^{2+}$ .



**Table 3** Comparison of LoD for Hg(II) analysis using the proposed Ag NPs protocol with previously reported methods

Probe	LoD/M (reference)
<b>A. Ligand spectrophotometry</b>	
(i) Rhodamine derivative	77 nM (ref. 39)
(ii) Ruthenium complexes	100 nM (ref. 40)
<b>B. Electrochemical detection</b>	
(i) AgNPs	8.43 $\mu$ M (ref. 41)
<b>C. Nanomaterial spectrophotometry</b>	
(i) AuNPs	750 nM (ref. 40)
(ii) AuNPs	55 nM (ref. 42)
(iii) AuNPs	10 000 nM (ref. 43)
(iv) AuNPs	20 000 nM (ref. 44)
(v) AgNPs	800 nM (ref. 45)
(vi) AgNPs	2200 nM (ref. 46)
(vii) AgNPs	85 000 nM (ref. 47)
(viii) AgNPs	1500 nM (ref. 48)
(ix) AgNPs	300 nM (ref. 49)
(x) AgNPs	5 nM (ref. 50)
(xi) AgNPs	17 nM (ref. 51)
(xii) AgNPs	0.007 mg L <sup>-1</sup> (ref. 52)
(xiii) AgNPs	6.2 ppb (ref. 53)
(xiv) AgNPs	100 nM (ref. 54)
(xv) AgNPs	6.25 ppb (ref. 55)
(xvi) AgNPs	15.70 nM (present study)

deviations for water samples from the three ponds were calculated as 1.488%, 1.430% and 1.638%, respectively, for these solutions. A *t*-test also showed a statistically significant difference for these absorbance values at <0.01% confidence level ( $F = 288.74$ ). These results show that the proposed method has great potential for the sensing of Hg<sup>2+</sup> and Sn<sup>2+</sup> in environmental samples.

## Experimental

### Chemicals and reagents

Garlic was purchased from Naharlagun market, Arunachal Pradesh, India. Silver nitrate (AgNO<sub>3</sub>) and alliin were purchased from Sigma-Aldrich (USA). Metal salts of Al(NO<sub>3</sub>)<sub>3</sub>, AgNO<sub>3</sub>, Cr<sub>2</sub>O<sub>3</sub>, FeCl<sub>3</sub>, KCl, FeSO<sub>4</sub>·7H<sub>2</sub>O, CuCl<sub>2</sub>, NaCl, NiCl<sub>2</sub>, Zn(NO<sub>3</sub>)<sub>2</sub>, and Pb(NO<sub>3</sub>)<sub>2</sub> were purchased from Merck India Ltd.

**Table 4** Comparison of LoD for Sn(II) analysis using the proposed Ag NPs protocol with previously reported methods

Probe	LoD/M (reference)
<b>A. HPLC</b>	
(i) Reversed phase HPLC	1.4 ppb (ref. 56)
<b>B. Inductively coupled plasma optical emission spectrometry (ICP OES)</b>	
(i) Direct method	0.0036 mg L <sup>-1</sup> (ref. 57)
(ii) Hydride generation	0.0006 mg L <sup>-1</sup> (ref. 57)
<b>C. Nanomaterial spectrophotometry</b>	
(i) AgNPs	0.036–0.040 $\mu$ M (ref. 58)
(ii) AgNPs	11.25 nM (present study)

### Preparation of garlic extract

A paste was prepared by grinding 40 g of peeled garlic bulb and mixing in 400 ml deionized water. The mixture was shaken using a magnetic stirrer at 60 °C for 45 minutes. The mixture was cooled to room temperature (24–25 °C) and filtered using Whatman filter paper (Cat no 1004 150) with pore size 20–25  $\mu$ m. The filtrate part was used for further experiments.

### Synthesis of Ag NPs

The filtrate (hereafter called garlic extract) was used for synthesizing Ag NPs. 10 ml of 1 mM AgNO<sub>3</sub> solution was mixed with 2 ml garlic extract. After 48 h, the solution turned brownish indicating the formation of Ag NPs. In order to study the formation of Ag NPs, UV-visible absorption spectra were recorded using a spectrophotometer at wavelengths of 300–700 nm (Fig. 1).

### High-performance liquid chromatography (HPLC) analysis

For determining the major bioactive molecules present in the aqueous extract, HPLC experiment was performed. The garlic extract was prepared in water as mentioned above. The flow rate was 1 ml min<sup>-1</sup> and the injection volume was 20  $\mu$ l. The mobile phase was a binary solvent system consisting of A : B (80 : 20) (acetonitrile : water with 0.2% phosphoric acid) which was sonicated for almost 15 minutes and degassed. The temperature of the column was maintained at 30 °C. The compounds in the chromatographic analysis were identified by comparing with standard compounds using the retention time and the absorption spectrum profile.

### Characterization of the synthesized Ag NPs

The Ag NPs were characterized using various techniques such as UV-visible spectroscopy, XRD, TEM, and Raman spectroscopy. The formation of Ag NPs was monitored using UV-visible spectroscopy in which UV-visible spectra were recorded in the range between 300 and 700 nm (model: Lambda 25, Switzerland). The HPLC experiment was done using a Shimadzu Prominence HPLC instrument (LC-20AP), with PDA detector (200–800 nm) and an analytical column of modified silica gel (C<sub>18</sub> column, 5  $\mu$ m pore size). XRD analysis was done using a Rigaku X-ray diffractometer (model: ULTIMA IV, Rigaku, Japan) with a Cu K X-ray source (1.54056 Å) at a generator voltage of 40 kV, current 40 mA with scanning rate of 2° min<sup>-1</sup>. High-resolution TEM images were captured using a JEOL model 2100 EX (Japan) operated at a voltage of 200 kV. XPS analysis was carried out with an ESCALAB Xi + e (Thermo Fisher Scientific Pvt. Ltd, UK) to determine the elemental composition, empirical formula, chemical state and electronic state of the elements within the nanoparticles. Lastly, Raman spectra were obtained with a Thermo-Scientific DXR2 smart Raman at an excitation wavelength of 532 nm.

### Detection of metal ions using synthesized Ag NPs

To study the metal ion detection and sensing capacity of the green-synthesized freshly prepared Ag NPs, 1 mM Fe<sup>2+</sup>, Ni<sup>2+</sup>,



Table 5 Comparison of calculated amount of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  with actual value

Parameter	AgNPs + 2 nM $\text{Hg}^{2+}$	AgNPs + 2 nM $\text{Sn}^{2+}$
Change in absorption compared to AgNPs ( $Y$ )	0.299	0.170
Calculated concentration (nM) ( $X$ )	1.882	2.261
Regression model	$Y = 0.0366X + 0.2301$	$Y = 0.0234X + 0.1170$
$R^2$	0.8821	0.9386
Pearson's coefficient	0.9436	0.9719

$\text{Pd}^{2+}$ ,  $\text{Pb}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Al}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{K}^+$ ,  $\text{Cu}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Ba}^{2+}$ ,  $\text{Sn}^{2+}$ ,  $\text{Hg}^{2+}$  were added to 5 ml Ag NPs solution. The changes in the UV-visible absorption spectra were monitored at room temperature (25 °C) accordingly.

### Estimation of LoD for $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ ions

The sensitivity and the minimum detectable concentration of the Ag NPs were studied by using UV-visible absorption spectra at different concentrations of  $\text{Hg}^{2+}$  (1–15 nM) and  $\text{Sn}^{2+}$  (1–11 nM) in aqueous solution at room temperature (25 °C). Total of 3 ml of Ag NPs solution diluted with deionized water (0.2 times) was taken for the detection of both metal ions. The respective amount of metal ion solution was added to the solution. The absorption was determined at 403 nm and 405 nm, respectively. The LoD was estimated using the UV-visible absorption spectra of Ag NPs with characteristic peak at 420 nm. Changes in absorption were monitored with variation of concentration: 1–15 nM and 1–11 nM for  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$ , respectively. The LoD was calculated using the standard formula:<sup>59</sup>

$$\text{LoD} = 3.3 \times \text{SD}/S$$

where SD = standard deviation of response (peak area) and  $S$  = slope of the calibration curve.

### Kinetics for binding of $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ with Ag NPs

The reaction kinetics and rate were studied using absorption spectra of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  solutions at different time intervals and concentrations. The rate constant was estimated from the slope of the regression plot with time *versus*  $\ln(A_t/A_0)$ , where  $A_t$  and  $A_0$  are the absorbance at time  $t$  and 0, respectively (Table 5).<sup>60</sup>

### Impact of pH on detection of $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$ ions

The pH value of the synthesized Ag NPs solution was 6.4. It was tuned to 4.0, 7.0 and 9.0 by adding buffer capsule of 4.0, 7.0 and 9.0, respectively, in the Ag NPs solution. The experiment was carried out at room temperature, *i.e.* 25 °C. Briefly, 5.00 ml of Ag NPs solution was taken. 10  $\mu\text{l}$  of 1 mM  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ion solution was added where pH was maintained using a buffer tablet. The absorbance was measured at each of 5, 10, 15, 20, 25 and 30 min.

### Synthesis and sensing mechanism for binding $\text{Hg}^{2+}$ and $\text{Sn}^{2+}$

A plausible mechanism of the synthesis of Ag NPs and detection of metal ions using the nanomaterial was predicted from

previous literature, characteristics of the synthesized Ag NPs and theoretical DFT study of optimized geometry.

### Computational details

All the structures were fully optimized without any symmetry constraint at the M06-2X level of theory.<sup>61</sup> The effective core potential basis set LANL2DZ was used for Ag, Hg and Sn atoms, while the 6-31+G\* basis set was used for all other main group elements. Frequency calculations were also performed to check the nature of the stationary states. All structures were found to be at their local minima with all real values of the Hessian matrix. All these calculations were performed using the GAUSSIAN 16 suite of programs.<sup>62</sup>

### Preparation of water samples

Water samples were collected from Lekhi (27°12'17.23" N, 93°73'35.73" E, altitude 281 m), Doimukh (27°14'03.52" N, 93°78'28.39" E, altitude 283 m) and Naharlagun (27°06'16.81" N, 93°41'42.65" E, altitude 200 m), Arunachal Pradesh. Suitable amounts of 1 mM standard solutions of  $\text{Hg}^{2+}$ ,  $\text{Sn}^{2+}$  and other metal ions were added to these collected samples, mixed thoroughly and filtered. The water samples were analysed following the procedures described above.

## Conclusions

A very simple, cost-effective, highly selective and sensitive, rapid colorimetric method for detection of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  using alliin-mediated garlic extract-synthesized Ag NPs has been exploited in this work. The reported probe (Ag NPs) showed high selectivity and sensitivity for  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions over several other metal ions. The  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions successfully induced the transformation of  $\text{Ag}^+$  to  $\text{Ag}^0$  NPs *via* complex formation yielding a blue shift in the SPR peak accompanying a colour change from brown to colourless. The LoD for  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions were found as 15.7 nM and 11.25 nM, respectively, which is very significant compared to reported information so far. The kinetics for the binding of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ions has also been reported in this study. The prepared probe (Ag NPs) represents a very significant game-changing analytical method for detection of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  ion contaminants in water. The mechanism of the protocol is strongly supported by DFT study. The result shows that the proposed method has great potential for the sensing of  $\text{Hg}^{2+}$  and  $\text{Sn}^{2+}$  in environmental samples.





## Author contributions

Rintumoni Paw: conceptualization, data curation, investigation, methodology, formal analysis, resources, visualization, writing – original draft, writing – review & editing. Moushumi Hazarika: validation. Purna K. Boruah: characterization. Amlan Jyoti Kalita: software, data analysis. Ankur K. Guha: validation, software. Manash R. Das: characterization. Chandan Tamuly: conceptualization, methodology, supervision, visualization, writing – review & editing.

## Conflicts of interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Acknowledgements

The authors acknowledge the Director, CSIR-North East Institute of Science & Technology Jorhat Assam for providing facilities and valuable suggestions (ref NMN-202145) and R. Paw thanks CSIR New Delhi for the Junior Research Fellowship/ Financial Assistance to perform the presented work. C. Tamuly thanks to Dr Hiren Gogoi, Assist Prof., Dept of Zoology, RGU, Doimukh, Arunachal Pradesh, SEED Division, New Delhi and NMPB, New Delhi for their support and help.

## References

- 1 P. B. Tchounwou, C. G. Yedjou, A. K. Patlolla and D. J. Sutton, in *Molecular, Clinical and Environmental Toxicology* ed. A. Luch. Springer, Basel, 2012, *Experientia Supplementum*, vol. 101, pp. 133–164.
- 2 G. Guzzi and C. A. La Porta, *Toxicol.*, 2008, **244**, 1–12.
- 3 K. M. Rice, E. M. Walker Jr, M. Wu, C. Gillette and E. R. Blough, *J. Prev. Med. Public Health*, 2014, **47**, 74–83.
- 4 Y. S. Hong, Y. M. Kim and K. E. Lee, *J. Prev. Med. Public Health*, 2012, **45**, 353–363.
- 5 ATSDR (Agency for Toxic Substances and Disease Registry), *Toxicological Profile for Tin and Tin Compounds*. U.S. Department of Health and Human Services, Atlanta, 2005.
- 6 T. Gajda and A. Jancso, in *Metal Ions in Life Sciences* ed. A. Sigel, H. Sigel and R. K. O. Sigel, Royal Society of Chemistry, Cambridge, 2010, ch. 4, vol. 7, pp. 111–151.
- 7 E. Dopp, L. M. Hartmann, A. M. Florea, A. W. Rettenmeier and A. V. Hirner, *Crit. Rev. Toxicol.*, 2004, **34**, 301–333.
- 8 A. V. Hirner and A. W. Rettenmeier, *Met. Ions Life Sci.*, 2010, **7**, 465–521.
- 9 A. I. El-Makawy, S. M. Girgis and W. K. B. Khalil, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2008, **657**, 105–110.
- 10 B. Michalke, S. Halbach and V. Nischwitz, *J. Environ. Monit.*, 2009, **11**, 939–954.
- 11 E. Dopp, A. W. Rettenmeier, R. H. Kretsinger and V. N. Uversky, in *Encyclopedia of Metalloproteins*, ed. E. A. Permyakov, Springer, New York, 2013.
- 12 D. Martín-Yerga, M. B. González-García and A. Costa-García, *Talanta*, 2012, **116**, 1091–1104.
- 13 S. Lin, W. Zhu, Y. Jin and K. B. Crozier, *Nano Lett.*, 2013, **13**, 559–563.
- 14 Y. Lu, S. Feng, X. Liu and L. Chen, *J. Nanomater.*, 2013, 984831, DOI: 10.1155/2013/984831, 1–8.
- 15 G. Chen, Z. Guo, G. Zeng and L. Tang, *Analyst*, 2015, **140**, 5400–5443.
- 16 E. Block, J. Dane, S. Thomas and R. Cody, *J. Agric. Food Chem.*, 2010, **58**, 4617–4625.
- 17 T. Miron, T. Bercovici, A. Rabinkov, M. Wilchek and D. Mirelman, *Anal. Biochem.*, 2004, **331**, 364–369.
- 18 S. H. Omar and N. A. Al-Wabel, *Saudi Pharm. J.*, 2010, **18**, 51–58.
- 19 A. Shang, S. Cao, X. Xu, R. Ren-You Gan, G. Tang, H. Corke, V. Mavumengwana and H. Li, *Foods*, 2019, **8**, 246.
- 20 A. L. González, C. Noguez, J. Beránek and A. S. Barnard, *J. Phys. Chem. C*, 2014, **118**, 9128–9136.
- 21 C. Tamuly, M. Hazarika, M. Bordoloi and M. R. Das, *Mater. Lett.*, 2013, **102–103**, 1–4.
- 22 C. Tamuly, M. Hazarika, M. Bordoloi, P. K. Bhattacharyya and R. Kar, *Spectrochim. Acta, Part A*, 2014, **132**, 687–691.
- 23 A. Hebeish, M. H. El-Rafie, F. A. Abdel-Mohdy, E. S. Abdel Halim and H. E. Emam, *Carbohydr. Polym.*, 2010, **82**, 933–941.
- 24 Z. A. Ali, R. Yahya, S. D. Sekaran and R. Puteh, *Adv. Mater. Sci. Eng.*, 2016, 4102196.
- 25 I. Barwal, P. Ranjan, S. Kateriya and S. C. Yadav, *J. Nanobiotechnol.*, 2011, **9**, 56.
- 26 W. Yu and H. Xie, *J. Nanomater.*, 2012, 435873.
- 27 S. Asghari, S. A. Johari, J. H. Lee, Y. S. Kim, Y. B. Jeon, H. J. Choi, M. C. Moon and I. J. Yu, *J. Nanobiotechnol.*, 2012, **10**, 14.
- 28 R. A. Raza, Z. Kanwal, A. Rauf, A. N. Sabri, S. Riaz and S. Naseem, *Nanomater.*, 2016, **6**, 74.
- 29 A. J. Kora, R. B. Sashidhar and J. Arunachalam, *Carbohydr. Polym.*, 2010, **82**, 670–679.
- 30 N. Biswas, S. Kapoor, H. S. Mahal and T. Mukherjee, *Chem. Phys. Lett.*, 2007, **444**, 338–345.
- 31 J. A. Buledi, S. Amin, S. I. Haider, M. I. Bhanger and A. R. Solangi, *Environ. Sci. Pollut. Res.*, 2020, DOI: 10.1007/s11356-020-07865-7.
- 32 S. Singh, A. Halder, O. Sinha, P. K. Sarkar, P. Singh, A. Banerjee, S. A. Ahmed, A. Alharbi, R. Obaid, S. K. Ghosh, A. Mitra and S. K. Pal, *PLoS One*, 2020, **15**(1), e0227584, DOI: 10.1371/journal.pone.0227584.
- 33 J. A. Adekoya, E. O. Dare, M. A. Mesubi, A. A. Nejo, H. C. Swart and N. Revaprasadu, *Results Phys.*, 2014, **4**, 12–19.
- 34 S. Liang, G. Zhang, J. Min, J. Ding and X. Jiang, *J. Nanomater.*, 2014, 684251.
- 35 S. Kumar-Krishnan, E. Prokhorov, M. Hernández-Iturriaga, J. D. Mota-Morales, M. Vázquez-Lepe, Y. Kovalenko, *et al.*, *Eur. Polym. J.*, 2015, **67**, 242–251.
- 36 S. Ivanov, A. Barylyak, K. Besaha, A. Bund, Y. Bobitski, R. Wojnarowska-Nowak, I. Yaremchuk and M. Kus-Liśkiewicz, *Nanoscale Res. Lett.*, 2016, **11**, 140.



- 37 A. J. Bard, R. Parsons and J. Jordan, *Standard Potentials in Aqueous Solution*, International Union of Pure and Applied Chemistry, Marcel Dekker Inc., New York, 1985.
- 38 G. Wu, C. Dong, Y. Li, Z. Wang, Y. Gao, Z. Shen and A. Wu, *RSC Adv.*, 2015, 5(26), 20595–20602, DOI: 10.1039/C5RA00001G.
- 39 H. El Kaoutit, P. Estévez, F. C. García, F. Serna and J. M. García, *Anal. Methods*, 2013, 5, 54–58.
- 40 E. Coronado, J. R. Galán-Mascarós, C. Martí-Gastaldo, E. Palomares, J. R. Durrant, R. Vilar, M. Gratzel and M. K. Nazeeruddin, *J. Am. Chem. Soc.*, 2005, 127, 12351–12356.
- 41 E. Eksin, A. Erdem, T. Fafal and B. Kivcak, *Electroanalysis*, 2019, 31, 1–9.
- 42 X. Yang, H. Liu, J. Xu, X. Tang, H. Huang and D. Tian, *Nanotechnol.*, 2011, 22, 275503.
- 43 S. Boopathi, S. Senthilkumar and K. L. Phani, *J. Anal. Methods Chem.*, 2012, 348965.
- 44 S. Si, A. Kotal and T. K. Mandal, *J. Phys. Chem. C*, 2007, 111, 1248–1255.
- 45 S. Bothra, J. N. Solanki and S. K. Sahoo, *Sens. Actuators, B*, 2013, 188, 937–943.
- 46 K. Farhadi, M. Forough, R. Molaei, S. Hajizadeh and A. Rafipour, *Sens. Actuators, B*, 2012, 161, 880–885.
- 47 M. L. Firdaus, I. Fitriani, S. Wyantuti, Y. W. Hartati, R. Khaydarov, J. A. Mcalister, H. Obata and T. Gamo, *Anal. Sci.*, 2017, 33, 831–837.
- 48 F. Tanvir, A. Yaqub, S. Tanvir, R. An and W. A. Anderson, *Materials*, 2019, 12, 1533.
- 49 A. Tirado-Guizar, G. Rodriguez-Gattorno, F. Paraguay-Delgado, G. Oskam and G. E. Pina-Luis, *MRS Commun.*, 2017, 7, 695–700.
- 50 U. Khan, A. Niaz, A. Shah, M. I. Zaman, M. A. Zia, F. J. Iftikhar, J. Nisar, M. N. Ahmed, M. S. Akhter and A. H. Shah, *New J. Chem.*, 2018, 42, 528–534.
- 51 Y. Wang, F. Yang and X. Yang, *ACS Appl. Mater. Interfaces*, 2010, 2, 339–342.
- 52 S. Puchum, R. Meelapsom, S. S. Muniandy, H. L. Lee, S. Pencharee, M. Amatongchai, K. Suttisintong and P. Jarujamrus, *Int. J. Environ. Anal. Chem.*, 2019, 99, 139–156.
- 53 G. M. Sangaonkar, M. P. Desai, T. D. Dongale and K. D. Pawar, *Sci. Rep.*, 2020, 10, 2037.
- 54 A. Nain, S. R. Barman, S. Jain, A. Mukherjee and J. Satija, *Appl. Nanosci.*, 2017, 7, 299–307.
- 55 S. Ichinoki, H. Iwase, F. Arakawa, K. Hirano and Y. Fujii, *J. Liq. Chromatogr. Relat. Technol.*, 2003, 26, 3129–3139.
- 56 M. Fišera, S. Kráčmar, H. Velichová, L. Fišerová, P. Burešová and P. Tvrzník, *Potr. S. J. F. Sci.*, 2019, 13, 269–377.
- 57 J. K. Salem, I. M. El Nahhal and M. H. Shurrab, *Int. J. Environ. Anal. Chem.*, 2020, 1726901.
- 58 B. T. Alquadeib, *Saudi Pharm. J.*, 2019, 27, 66–70.
- 59 F. Ali, S. B. Khan, T. Kamal, K. A. Alamry and A. M. Asiri, *Sci. Rep.*, 2018, 8, 6260.
- 60 M. J. Frisch, G. W. Trucks, H. B. Schlegel and G. E. Scuseria *et al.*, *FOX GAUSSIAN 16 (Revision A.03)*, Gaussian Inc, Wallingford CT, 2008.
- 61 Y. Zhao and D. G. Truhlar, *Theor. Chem. Acc.*, 2008, 120, 215–241.
- 62 G. M. Sangaonkar, M. P. Desai, T. D. Dongale and K. D. Pawar, *Sci. Rep.*, 2020, 10, 2037, DOI: 10.1038/s41598-020-58844-4.

