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# Synthesis of green and pure copper oxide nanoparticles using two plant resources *via* solid-state route and their phytotoxicity assessment

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Among the conventional methods in synthesizing nanoparticles, the methods that use biological resources, as reducing and stabilizing agents, can be considered eco-friendly methods. In this study, the leaf tissue of green tea (*Camellia sinensis* L.) and lavender (*Lavandula angustifolia*) were utilized by the solid-state method as a one-step and low-cost method for the biosynthesis of copper oxide nanoparticles (CuO NPs). The results of the X-ray Diffraction (XRD), field emission scanning electron spectroscopy (FESEM) and transmission electron microscopy (TEM) showed that lavender is more productive in the synthesis of pure and uniform CuO NPs (50 nm). Comparing biogenic synthesized CuO NPs with chemically synthesized CuO NPs in terms of induction of phytotoxicity, exposed in treatments with concentrations of 40, 400 and 4000  $\mu\text{g ml}^{-1}$ , green CuO NPs had less inhibitory effects on the seed germination factors (*i.e.*, germination percentage, germination rate, shoot and root length, *etc.*) of lettuce (*Lactuca sativa* L.), and tomato (*Solanum lycopersicum* L.) seeds. However, both green/chemically synthesized CuO NPs at their lowest concentrations (4  $\mu\text{g ml}^{-1}$ ), had an effective role in root and shoot expansion of lettuce and tomato seedlings.

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## Introduction

Nanotechnology and application of nanoscale materials play an essential role in serving humans.<sup>1</sup> The shape and size of nanoparticles as the two main factors determine the unique magnetic, electronic, optical and catalytic properties of nanoparticles.<sup>2</sup> Copper oxide nanoparticles (CuO NPs) due to their unique electric, thermal, mechanical, catalytic and magnetic properties, are used in various fields such as agricultural, environmental, industrial and medical.<sup>3,4</sup> In general, all nanoparticles synthesis protocols are sorted into three main categories; physical, chemical and biological; and the naming of each of these methods is based on the reducing agent or electron source involved in the nanoparticle synthesis process.<sup>5</sup> So that, in the physical method, the electron source is a physical source such as the electric current; in the chemical method, it is a chemical source (*e.g.*, sodium borohydride and hydrazine); and in the biological method it is an organism or a biomolecule.<sup>6–9</sup> The noteworthy, both of the physical and chemical methods are potentially hazardous for the environment and living organisms due to the usage of toxic reducing and

stabilizing agents in the synthesis process. Additionally, these methods are time-consuming, costly and consume high amounts of energy.<sup>10</sup> Therefore, the usage of biological methods to reduce environmental hazards, costs and energy consumption can be an appropriate alternative to the above-mentioned methods.<sup>11</sup> Recently, green synthesis of nanoparticles, as a one-step and eco-friendly method has attracted the attention of researchers and industries. The basis of this low-cost method is the use of plant extracts and organic compounds in the extracts that play the major role as reducing agents.<sup>12,13</sup>

Various methods were employed in the past to synthesize different copper-based nanoparticles, such as chemical synthesis, electrochemical synthesis, solvothermal route and phytosynthesis.<sup>14–17</sup> In this regard, several studies have reported the usage of herbal extracts in the green synthesis of Cu NPs such as *Garcinia mangostana* leaf extract, *Rubus glaucus* Benth, fruit and leaf extract, extract of seedless dates, leaves extract of *Ocimum sanctum* L., *Punica granatum* peel extract, root and leaf extract of *Asparagus adscendens* Roxb., fruit extract of *Ziziphus spina-christi* L. and leaf aqueous extract of *Nerium oleander*.<sup>2,13,18–23</sup> In particular, the green synthesis of CuO NPs and their various applications in the biomedical, agricultural and environmental fields have been recently reviewed.<sup>24,25</sup>

Considering this background into account, in this work, we investigate the formation of CuO NPs using *Lavandula angustifolia* and *Camellia sinensis* L. under solid-phase procedures as a very facile, inexpensive and eco-friendly method. Although no report has been published on the synthesis of CuO NPs in the

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solid-phase procedures, successful reports of the use of this method in the synthesis of Ag NPs have persuaded us to use this method in the synthesis of CuO NPs.<sup>26,27</sup>

Lavender (*Lavandula angustifolia*), also known as medicinal lavender, is a shrub of the family *Lamiaceae*, native to the Mediterranean region, but is grown in many other climates of the world.<sup>28</sup> This plant is a natural source of organic compounds such as coumarin, coumaric acid, ursolic acid, valeric acid, anthocyanins, essential oils, tannins, phytosterols, herniarin and glycolic acid.<sup>29</sup> Tea (*Camellia sinensis* L.), belongs to the family Theaceae, is the oldest caffeine-containing non-alcoholic beverage in the world.<sup>30</sup> The biochemical compositions of tea leaves, such as alkaloids (theobromine, caffeine, etc.) and polyphenols (catechins and flavonoids), polysaccharides, volatile oils, vitamins and inorganic elements, have antimicrobial, antioxidant, anti-inflammatory and antiallergic activity.<sup>31</sup> According to previous reports, the use of lavender in the biosynthesis of CuO NPs in this study is innovative and the only use of the tea plant in the biosynthesis of copper nanoparticles has been just in the aqueous phase, not in the solid phase.<sup>4,32–34</sup> In the following, through phytotoxicity assessment, green synthesized CuO NPs were evaluated in comparison with the chemically synthesized CuO NPs. Hence, evaluating the phytotoxicity effects of both green and chemically synthesized nanoparticles was performed using both seed germination and seedling growth assay of two plant species lettuce (*Lactuca sativa* L.), and tomato (*Solanum lycopersicum* L.).

## Materials and methods

### Chemicals

For the synthesis of CuO NPs, copper sulfate pentahydrate ( $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ ) was purchased from Sigma-Aldrich and the absolute ethanol was commercially obtained from Merck (Germany).

### Plant materials

The dry specimens of lavender (*L. angustifolia*) and green tea (*C. sinensis*) were purchased from local market of Karaj, Iran. Then, unwanted impurities were removed from the samples and the leaves of both plants were separated from the prepared heterogeneous samples. Finally, in order to obtain the desired powder sample, the leaf samples were crushed *via* a mortar and pestle.

### Synthesis of CuO NPs

For the synthesis of green CuO NPs using the solid-state route, the dry specimens of lavender and green tea leaves were thoroughly powdered using mortar and pestle. In the following, different amounts of  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$  were added to the certain mass of lavender powder and different samples with different ratios ( $m_{\text{CuSO}_4 \cdot 5\text{H}_2\text{O}} : m_{\text{lavender powder}} = 1 : 1, 1 : 2.5$  and  $1 : 5$ ) were prepared, which were coded as Cu-L1, Cu-L2.5, and Cu-L5. Then the resulting mixtures were well-grounded again, attaining homogenized powder samples. Similarly, three additional samples were prepared by green tea leave powder so that the

copper sulfate was mixed with green tea in different weight ratios ( $m_{\text{CuSO}_4 \cdot 5\text{H}_2\text{O}} : m_{\text{green tea powder}} = 1 : 1, 1 : 2.5$  and  $1 : 5$ ) and named Cu-GT1, Cu-GT2.5 and Cu-GT5. Next, all the as-prepared samples were placed in a furnace and heated up to 600 °C, and after 4 h, they were allowed to cool down to ambient temperature. The precipitates were collected and suspended in absolute ethanol using vortex for washing. Then, synthesized CuO NPs were separated using a centrifuge at 5000 rpm for 4 min, the supernatant was discarded and the pellet was re-suspended in deionized water. The washing process with water was repeated three times and the black CuO NPs precipitates were dried at 80 °C and stored for further utilization.

### Characterization of CuO NPs

To monitor particle size and chemical composition of the current green-synthesized CuO NPs, X-ray diffraction (XRD, Philips model: PW1730,  $\lambda = 1.54056 \text{ \AA}$  (Cu K $\alpha$  irradiation),  $2\theta = 20\text{--}70^\circ$ ) was employed. For calculation of the crystallite sizes ( $D$ ) of the samples Scherrer's equation were employed:<sup>3</sup>

$$D = K\lambda/(\beta \times \cos \theta)$$

where,  $K$  is the shape factor constant (0.9),  $\lambda$  is the wavelength of the X-ray (0.1546 nm),  $\beta$  is the broadening of the diffraction line at half-maximum in radians, and  $\theta$  is the Bragg's diffraction angle. Moreover, field emission scanning electron microscopy (FESEM, a TESCAN MIRA II, with the 20 kV voltage) and transmission electron microscopy (TEM, EM208S Philips microscope with accelerating voltage of 100 kv) were utilized for evaluating the surface topology and morphology of the green-synthesized CuO NPs. For depicting the particle size histogram, the size of more than 100 particles in the TEM image was estimated.

### Phytotoxicity study of CuO NPs

Tests were conducted using two plant species seeds, lettuce (*Lactuca sativa* L.) and tomato (*Solanum lycopersicum* L.), purchased from a local nursery. Test seeds were screened to remove broken, small and immature seeds. In the following, to remove the biological contamination, the seeds were sterilized using 5% sodium hypochlorite solution and rinsed thrice using distilled water. Nanoparticles were suspended directly in distilled water and discrete by ultra-sonic vibration for 30 min. Concentrations of 0, 4, 40, 400, 4000  $\mu\text{g ml}^{-1}$  were prepared from green and chemically synthesized nanoparticles and distributed in the plates containing 25 seeds and the plates were placed in a germinator at 22 °C. Treatments were arranged in a complementary randomized design (CRD) with three replications. In order to evaluate the potential of nanoparticles toxicity on two studied plant species, six important attributes including germination percentage, germination rate, abnormal seedling, stem length (mm) and root length (mm) were recorded.

### Statistical analysis

The experiments were conducted based on a completely randomized design (CRD) with three replications and the data



were statistically analyzed using ANOVA to determine significant differences (defined as  $p \leq 0.05$ ) by using SAS software package. Fitting standard errors of the means ( $\pm$ SEM) were calculated for presentation of graphs. Fisher's least significant difference (LSD) was applied to compare the potential significant differences between treatments.

## Results and discussion

### Characterization of green as-synthesized CuO NPs

The microstructure and purity of the nanoparticles, the samples were characterized by XRD analysis (Fig. 1). XRD patterns of the samples produced using both plants in the  $m_{\text{CuSO}_4 \cdot 5\text{H}_2\text{O}} : m_{\text{plant}}$  ratio of (1 : 5) (i.e., Cu-L5, Cu-GT5) showed diffraction peaks around  $2\theta = 32.7^\circ$ ,  $35.8^\circ$ ,  $39.1^\circ$ ,  $49.2^\circ$ ,  $53.9^\circ$ ,  $58.2^\circ$ ,  $61.7^\circ$ ,  $66.8^\circ$  and  $68.7^\circ$  corresponding to the planes (110), (111), (200), (202), (020), (202), (113), and (022), respectively, and all revealed clearly the synthesis of the pure crystalline particles of monoclinic CuO NPs.<sup>34</sup> While decreasing the weight ratio of copper salt to plant powder up to 2.5 resulted in no significant changes in the XRD pattern of Cu-L2.5 sample, but it caused presence of some additional peaks in  $2\theta$  range between  $20$  to  $30^\circ$  in the case of Cu-GT2.5 sample, which were attributed to  $\text{Cu}_2\text{O}$  impurity.<sup>35,36</sup> When the ratio of  $m_{\text{CuSO}_4 \cdot 5\text{H}_2\text{O}} : m_{\text{plant}}$  was fixed at (1 : 1), XRD pattern of Cu-GT1 displayed diffraction peaks at  $2\theta = 26.8$  and  $29.7$  corresponding to  $\text{Cu}_2\text{O}$ , and  $2\theta = 31.2$  attributing to  $\text{Cu}(\text{OH})_2$ , along with CuO NPs signals. The XRD pattern of Cu-L1 sample similarly showed the impurity peaks, but the peak's intensities of the impurities were lower than Cu-GT1 sample. Moreover, according to the Scherrer's equation, the crystalline

sizes were found to be 94.94, 124.34, 152.01, 52.44, 25.56, and 104.91 nm for Cu-GT5, Cu-GT2.5, Cu-GT1, Cu-L5, Cu-L2.5, and Cu-L1, respectively. Altogether, XRD results confirmed that with increasing the amount of plant powder, more pure CuO NPs could be produced, and besides lavender powder could lead to the more pure CuO NPs compared to the green tea plant.

In the next step, SEM analyses were employed to compare and contrast the dispersity and morphology of the as-synthesized CuO NPs (Fig. 2). SEM images showed that both plant resource type and the ratio of  $m_{\text{CuSO}_4 \cdot 5\text{H}_2\text{O}} : m_{\text{plant}}$  played crucial roles in the morphology of the samples. For example, Cu-GT1 sample mostly consisted of large, rod shape stacked particles, but in Cu-GT2.5 sample, aggregated particles with polygonal shapes could be seen with the average particle size of 100 to 300 nm. In contrast, spherical CuO NPs (50 to 100 nm) were observed in the SEM image of the Cu-GT2.5 sample, which were to some extent agglomerated. SEM images of the samples produced using lavender plant (Cu-L2.5 and Cu-L5) showed tiny nanoparticles tightly stuck together. Interestingly, Cu-L1 sample displayed flower-like CuO NPs in its SEM image, in which growing rod shape particles probably formed a designed structure like flowers.

Moreover, for better clarification of the morphology of CuO NPs, TEM analysis was utilized. Based on the XRD and SEM results, Cu-L2.5 sample was selected for TEM analysis. The TEM images of this sample confirmed that CuO NPs were well-formed using lavender leaf powder through solid-state route (Fig. 3). It could be evidently seen uniform and spherical CuO NPs which were aggregated to some extent.

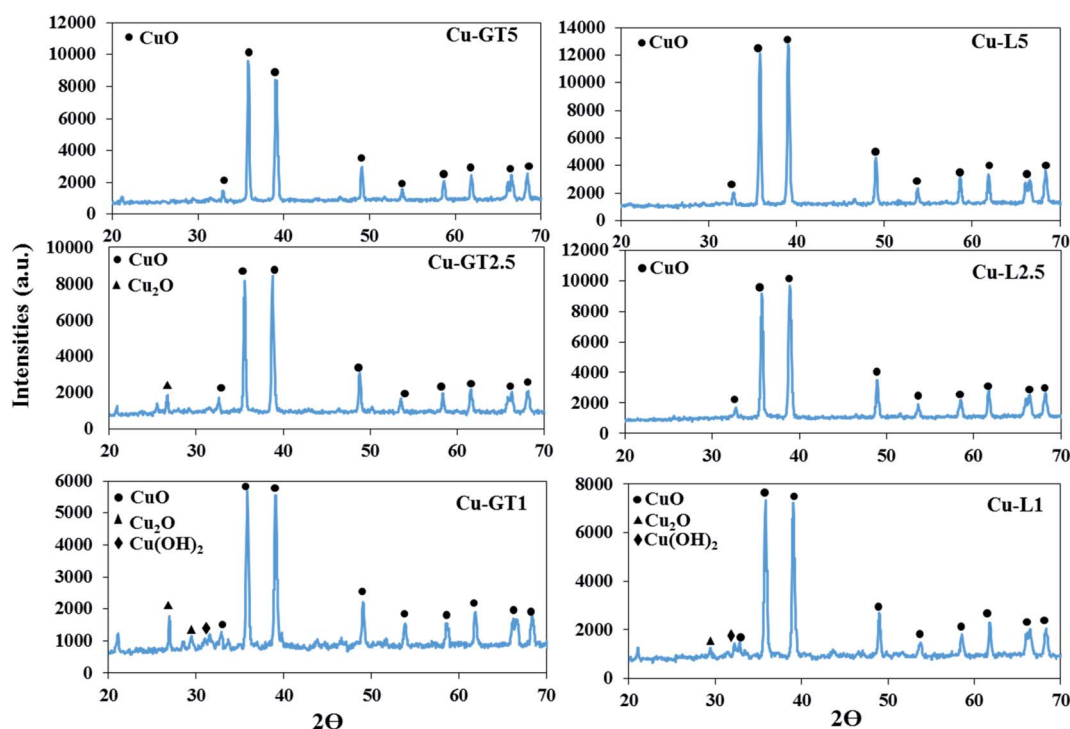


Fig. 1 XRD patterns of CuO NPs synthesized using different amounts of lavender and green tea leaf powders via solid-state method.





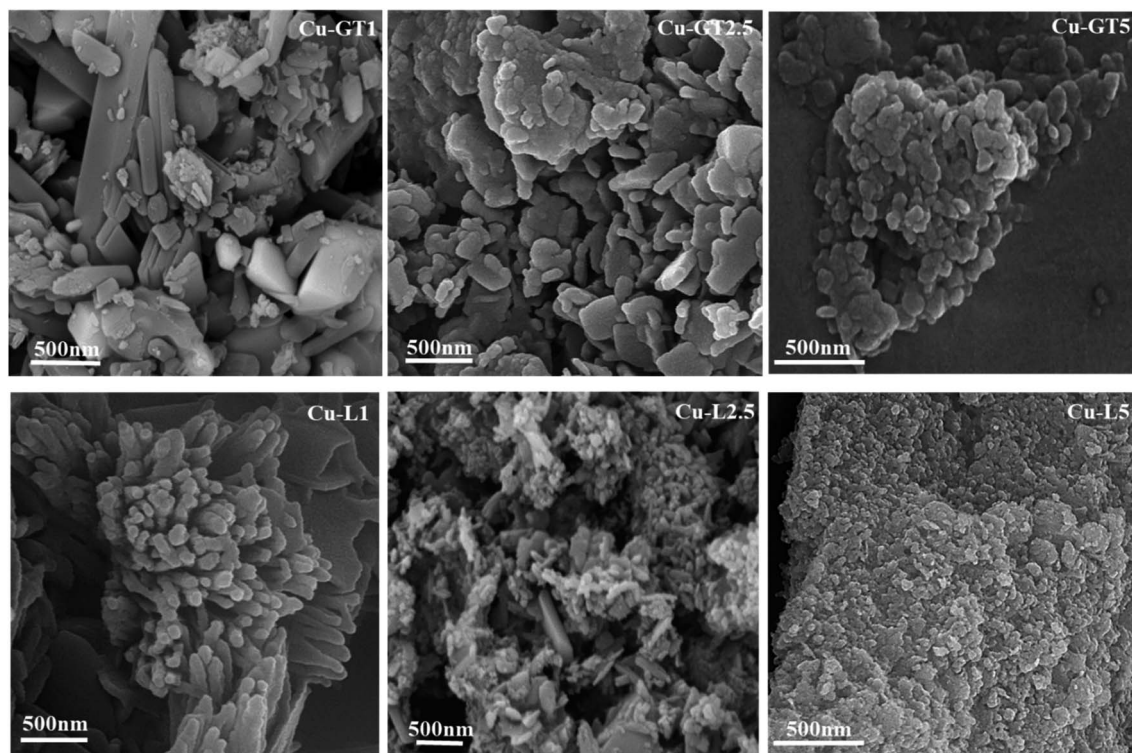


Fig. 2 FESEM images of CuO NPs synthesized using different amounts of lavender and green tea leaf powders *via* solid-state method.

### Phytotoxicity of green and chemically synthesized CuO NPs

The phytotoxicity assessment results showed that both green and chemically synthesized CuO NPs at the lowest concentration were effective on root and shoot development of seedlings of both plants species but at higher concentrations had a limiting effect on all investigated germination factors. The results of the phytotoxicity assessment are presented in the form of pictures and diagrams (Fig. 4 and 5).

### Germination percentage (GP)

Following the evaluation of the phytotoxicity criteria of the green and chemically synthesized CuO NPs in this study, it was observed that germination percentage, as the first inspected

factor, was affected by the toxicity of the nanoparticles. Taking tomato into account, the negative effect of green CuO NPs on the metric quantity was only visible at the highest concentration ( $4000 \mu\text{g ml}^{-1}$ ). However, the lower concentrations ( $4$ ,  $40$  &  $400 \mu\text{g ml}^{-1}$ ) did not have a significant negative effect on the germination percentage. With respect to chemically synthesized nanoparticles, decreasing oscillations began at a lower concentration ( $40 \mu\text{g ml}^{-1}$ ) and at concentrations of  $40$ ,  $400$  &  $4000 \mu\text{g ml}^{-1}$  GP values of  $65.33$ ,  $42.66$  and  $18.67\%$ , were acquired, respectively (Fig. 4). The effect of nanoparticles on the pattern of germination of the lettuce seeds was similar to that of previous plant. The only effective concentration of green CuO NPs on germination was observed at concentrations of  $4000 \mu\text{g ml}^{-1}$ , while in relation to chemically synthesized CuO NPs

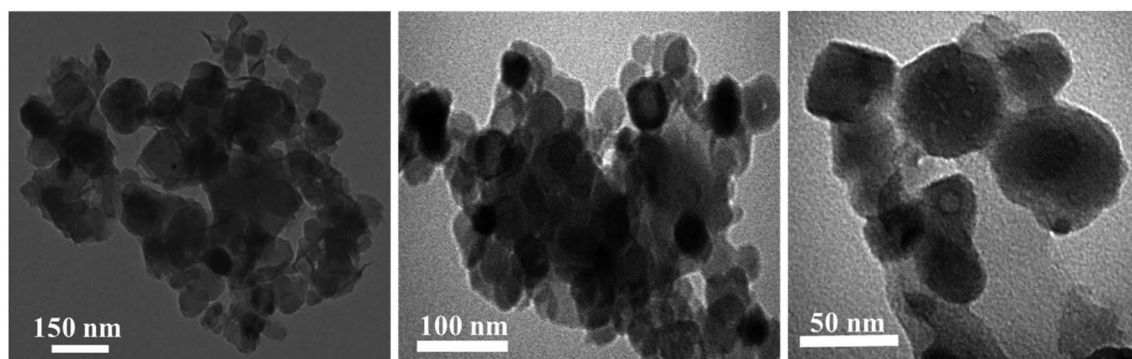


Fig. 3 TEM images of Cu-L2.5 sample in different magnifications.

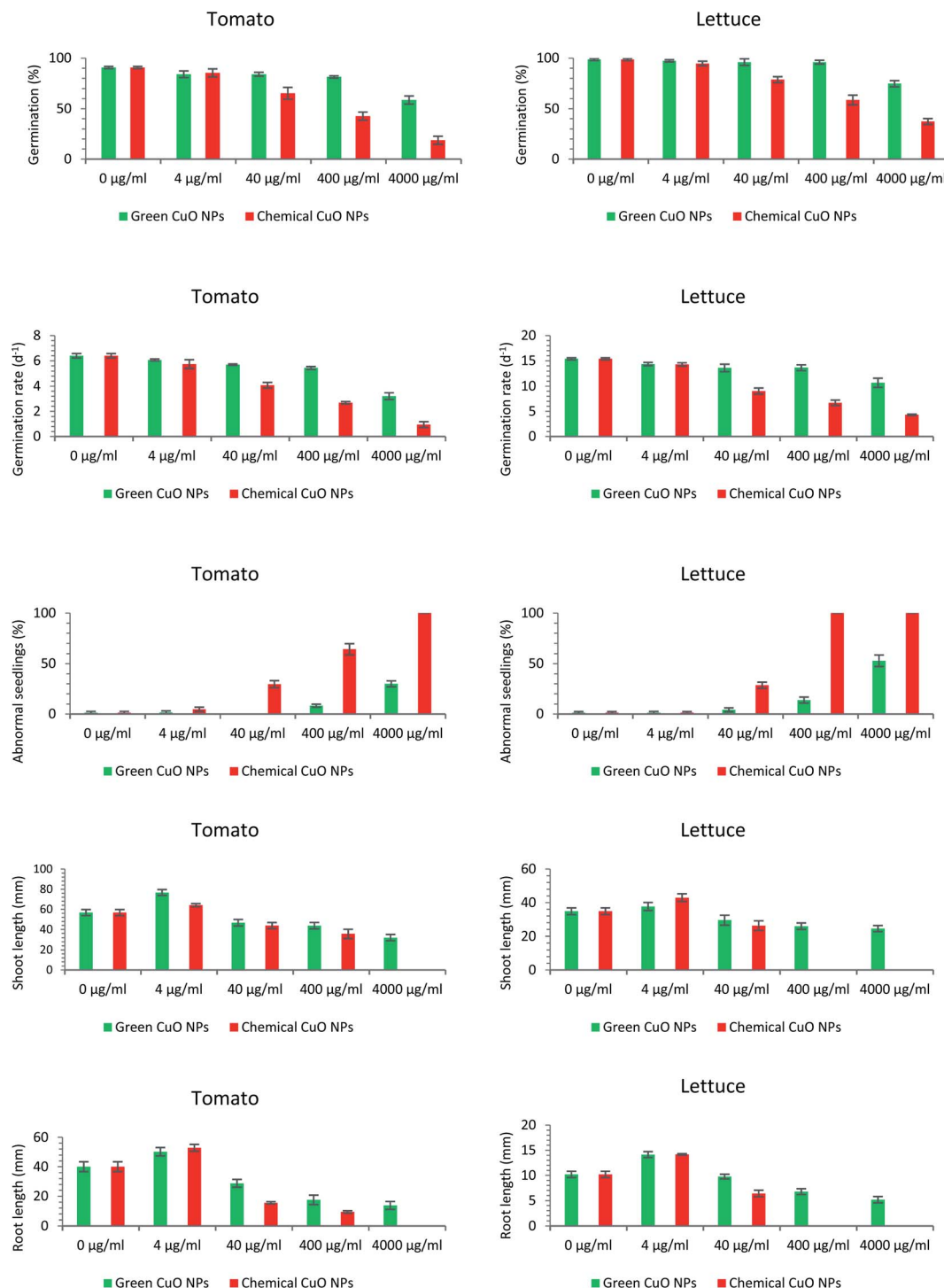


Fig. 4 Effect of green and chemically synthesized copper oxide nanoparticles on five factors of seed germination of *L. sativa* and *S. lycopersicum* seeds (from top to bottom, germination percentage, germination rate, seedling length, root length and abnormal seedling percentage).

concentrations of 40, 400 & 4000  $\mu\text{g ml}^{-1}$ , the germination percentage was reduced to 78.67, 58.67 and 37.33%, respectively (Fig. 4). When comparing the germination of seeds of two plants, it was noted that the tomato seeds showed more sensitivity to nanoparticle toxicity. Comparing nanoparticles in terms of their effects on seed germination, observations

disclosed that the chemically synthesized CuO NPs have a greater inhibitory effect than green CuO NPs.

#### Germination rate

Observations showed that the germination rate of tomato and lettuce seeds was also considerably affected by both kinds of the



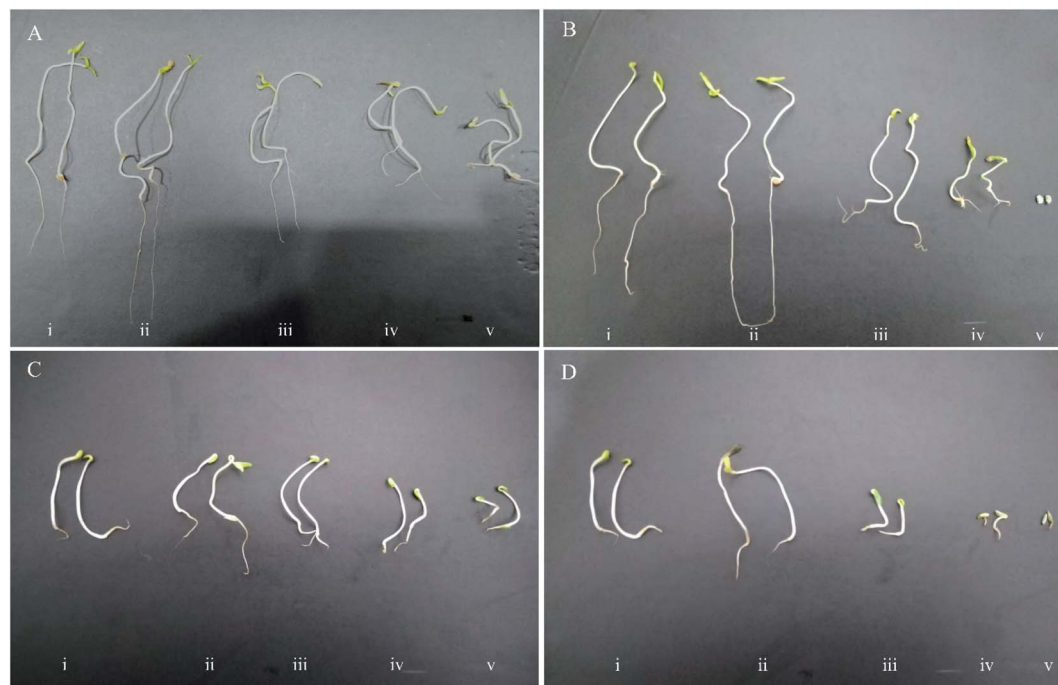


Fig. 5 Effect of green CuO NPs (left) and chemically synthesized CuO NPs (right) on seed germination of *S. lycopersicum* (A and B) and *L. sativa* (C and D) under different concentrations of 0 (i), 4 (ii), 40 (iii), 400 (iv) and 4000 (v)  $\mu\text{g ml}^{-1}$ .

CuO NPs. Regarding the first plant species, tomato, increasing the concentration of the chemically synthesized CuO NPs caused a downward trend in the germination rate. So that, the germination rate decreased from 6.4 (in  $0 \mu\text{g ml}^{-1}$  CuO NPs) to 0.95 (in  $4000 \mu\text{g ml}^{-1}$  CuO NPs). Green CuO NPs except at  $4000 \mu\text{g ml}^{-1}$  concentration had no striking effect on germination rate of tomato seeds. In fact, chemically and green synthesized CuO NPs had a similar effect on the germination rate of both plant species. In lettuce, the calculated value of 15.39 in control treatment was abridged to 14.27, 9.04, 6.71 & 4.32 under the influence of increasing the concentration of chemically synthesized CuO NPs to 4, 40, 400 &  $4000 \mu\text{g ml}^{-1}$ , respectively (Fig. 4). However, the only substantial inhibitory effect of green CuO NPs was recorded at the concentration of  $4000 \mu\text{g ml}^{-1}$ , which reduced the index value to 10.68.

#### Abnormal seedling

Similar to the germination percentage and the germination rate criteria, it was found that a large proportion of the tomato and lettuce seedlings were more negatively influenced by the high concentrations of chemically synthesized CuO NPs more than green CuO NPs. The abnormal seedlings mainly showed the symptoms of short and necrotic roots as well as short and springy shoots (Fig. 5). The green CuO NPs suspensions at concentrations of 400 &  $4000 \mu\text{g ml}^{-1}$  induced abnormal seedlings with abundance 8.25% and 30% in tomato seedlings, whereas these quantities in lettuce seedlings were 13.84 and 52.93%, respectively. The observations suggested that chemically synthesized CuO NPs encompass more side effects on the appearance of abnormal seedlings. So that, the increment

abnormal symptoms began under the influence of chemically synthesized CuO NPs at a concentration of  $40 \mu\text{g ml}^{-1}$  in both of the plant species seedlings. Interestingly, no normal tomato seedlings were observed in the last treatment (*i.e.*,  $4000 \mu\text{g ml}^{-1}$ ) and actually, the extremely toxic treatments for lettuce seedlings were 400 and  $4000 \mu\text{g ml}^{-1}$  (Fig. 4).

#### Shoot and root length

Remarkably, it was inferred that both green and chemically synthesized CuO NPs at their lowest concentration ( $4 \mu\text{g ml}^{-1}$ ), stimulated the shoot and root elongation in the seedlings of both plant species (Fig. 5). Contrary to this positive effect, the results indicated that higher concentrations of both green and chemically synthesized CuO NPs were toxic for seedling destruction. As the concentration of both nanoparticles increased, the recorded lengths of root and shoot for both plants declined dramatically. Finally, the higher toxicity of the chemically synthesized CuO NPs compared to the green form as well as the higher vulnerability of the root compared to the shoot against the inhibitory effect of nanoparticles on elongation were clearly visible (Fig. 4).

The effect of nanoparticles on plants can be very diverse based on NPs characteristics such as type, size, concentration, chemical and physical properties as well as plant-specific features.<sup>37</sup> Some reports indicated an influential role of metallic nanoparticles in damaging living organism's DNA, but with unknown molecular mechanisms.<sup>38</sup> The most prominent observations in this study were the lower phytotoxic effects of green synthesized CuO NPs compared to the chemically synthesized CuO NPs, on the germination indices. In the last



two decades, a number of reports have also presented the inhibitory effects of copper nanoparticles on plant growth factors; For example, the results of a study revealed the growth of *Triticum aestivum* and *Phaseolus radiatus* seedlings was limited under treatments of 200–1000 mg l<sup>-1</sup> of Cu NPs.<sup>39</sup> The 15 nm sized Cu NPs synthesized by polyols process method reduced stem and root elongation in *Eruca sativa* plant under the treatment of Murashige and Skoog (MS) medium containing 30 µg ml<sup>-1</sup> Cu NPs.<sup>40</sup> In another study, the root and shoot elongation of two important plant species, *Glycine max* and *Cicer arietinum*, significantly decreased in concentration of >100 ppm of copper oxide nanoparticles.<sup>41</sup> According to the copper oxide NPs phytotoxicity results, *Glycine max* seedlings were negatively affected by concentrations more than 50 mg l<sup>-1</sup>. However, the roots showed superior sensitivity to copper oxide NPs toxicity compared with the shoots.<sup>42</sup>

Despite the reports of the mentioned researches, comparing the toxic effect of green and chemically synthesized CuO NPs on plant growth characteristics has not been reported so far.

Similar to the results of this study, there are reports of the positive effect of nanoparticles on the growth attributes of some plant species. In a recent study, 25 and 50 mg l<sup>-1</sup> copper nanoparticles treatments significantly stimulated the soybean primary roots elongation.<sup>43</sup> Furthermore, it has been reported that concentrations of less than 1 ppm (0.2–1.0 ppm) of copper nanoparticles in wheat seedling medium led to a significant effect on increasing root dry weight and leaf area.<sup>44</sup> However, the massive accumulation of nanoparticles in shoots and roots could lead to phytotoxicity, the lower concentration of nanoparticles induced activity of antioxidant enzyme system, rubisco and chloroplast; therefore, the activity of these biosynthetic systems under low concentrations of nanoparticles may be effective in stimulating plant growth.<sup>44–47</sup>

In general, the toxicity of a nanoparticle can be attributed to the properties of the nanoparticle (size, surface area and intrinsic catalytic activity) and the reducing and stabilizing agents used in its synthesis and since the green nanoparticles at least lacks potential toxic reducing and stabilizing agents, these behave more gently in inducing phytotoxicity. Comparing green CuO NPs to chemically synthesized CuO NPs, some research reported that the green NPs induced less phytotoxicity whereas the chemically synthesized NPs showed more severe inhibitory effects. The results of trial using green and chemically synthesized Ag NPs with concentrations of 0, 100, 200, 400 and 600 ppm on the seed germination percentage and seedling length of *Matricaria chamomilla* and *Ocimum basilicum* revealed that, the green Ag NPs was significantly less toxic than chemically synthesized Ag NPs.<sup>26</sup> According to another report, the green Ag NPs nanoparticles synthesized by *Laminaria japonica* algal extract had no significant effect on the germination of *Triticum aestivum* and *Phaseolus mungo* seeds at the concentration of 0–80 ppm, whereas the non-toxic effect of nanoparticles on shoot and root elongation was factual at concentrations of below 30 ppm.<sup>48</sup>

Generally, CuO NPs have potentially inhibitory effects on germination features (e.g., germination percentage, shoot and root elongation, etc.). Nevertheless, depending on their

synthesis process whether green or chemically synthesized, different intensities of induced toxicity can be observed. Thus, green CuO NPs with the effects of toxicity much less than the chemically synthesized CuO NPs could be recognized as more eco-friendly NPs. Interestingly, CuO NPs can have a dual performance depending on their concentration, a nanotoxicant at high concentrations or a nanonutrient at low concentrations. At a constant concentration, each of the germination indices compared to the other exhibits various responses from inattention to sensitivity or stimulation. Different plant species could show variability in response patterns in the same CuO NPs treatments.

## Conclusions

From the outcome of our investigation, it is possible to conclude that, lavender as an organic and green source has been fruitful in synthesis of CuO NPs. Comparison of bio-synthesized CuO NP with chemically synthesized CuO in relation to phytotoxicity through seed germination test disclosed the biosynthesized CuO NP has little inhibitory effects on germination factors. However, in contrast, the adverse effects of chemically synthesized CuO NP on seed germination factors were more severe. Despite the phytotoxicity and limiting effects on seed germination and seedling growth due to the use of the CuO NPs, it was observed that both green and chemical CuO NPs at low concentrations (4 µg ml<sup>-1</sup>) effectively stimulated root and shoot elongation of lettuce and tomato seedlings. The use of lavender as a biogenic reducing and stabilizing agent, solid-state route as a low cost, safe and convenient method and the low toxicity effect of green CuO NPs made in this research can be considered as an eco-friendly research. The proposed method can be readily used in practice and can be successfully used for a number of fields related to nanotechnology.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 E. K. Elumalai, T. N. Prasad, J. Hemachandran, S. V. Therasa, T. Thirumalai and E. David, *J Pharm Sci Res*, 2010, **2**, 549–554.
- 2 Y. T. Prabhu, K. V. Rao, V. S. Sai and T. Pavani, *J. Saudi Chem. Soc.*, 2017, **21**, 180–185.
- 3 S. Sukumar, A. Rudrasenan and D. Padmanabhan Nambiar, *ACS Omega*, 2020, **5**, 1040–1051.
- 4 J. Sarkar, N. Chakraborty, A. Chatterjee, A. Bhattacharjee, D. Dasgupta and K. Acharya, *Nanomaterials*, 2020, **10**, 312.
- 5 S. Shende, A. P. Ingle, A. Gade and M. Rai, *World J. Microbiol. Biotechnol.*, 2015, **31**, 865–873.



- 6 R. Zhou, X. Wu, X. Hao, F. Zhou, H. Li and W. Rao, *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. Atoms*, 2008, **266**, 599–603.
- 7 Q. M. LIU, D. B. ZHOU, Y. Yamamoto, R. Ichino and M. Okido, *Trans. Nonferrous Metals Soc. China*, 2012, **22**, 117–123.
- 8 X. Su, J. Zhao, H. Bala, Y. Zhu, Y. Gao, S. Ma and Z. Wang, *J. Phys. Chem. C*, 2007, **111**, 14689–14693.
- 9 R. Cuevas, N. Durán, M. C. Diez, G. R. Tortella and O. Rubilar, *J. Nanomater.*, 2015, 2015.
- 10 A. K. Mittal, Y. Chisti and U. C. Banerjee, *Biotechnol. Adv.*, 2013, **31**, 346–356.
- 11 K. Vithiya and S. Sen, *Int. J. Pharm. Sci. Res.*, 2011, **2**, 2781.
- 12 P. Parikh, D. Zala and B. A. Makwana, *Open Access Library*, 2014, **1**, 1–15.
- 13 M. Gopinath, R. Subbaiya, M. M. Selvam and D. Suresh, *Int. J. Curr. Microbiol. Appl. Sci.*, 2014, **3**, 814–818.
- 14 S. P. Mondal, K. Das, A. Dhar and S. K. Ray, *Nanotechnology*, 2007, **18**, 095606.
- 15 D. Mo, J. Liu, H. J. Yao, J. L. Duan, M. D. Hou, Y. M. Sun, Y. F. Chen, Z. H. Xue and L. Zhang, *J. Cryst. Growth*, 2008, **310**, 612–616.
- 16 J. S. Jang, U. A. Joshi and J. S. Lee, *J. Phys. Chem. C*, 2007, **111**, 13280–13287.
- 17 R. C. Kasana, N. R. Panwar, R. K. Kaul and P. Kumar, *Environ. Chem. Lett.*, 2017, **15**, 233–240.
- 18 B. Kumar, K. Smita, L. Cumbal, A. Debut and Y. Angulo, *J. Saudi Chem. Soc.*, 2017, **21**, S475–S480.
- 19 E. A. Mohamed, *Heliyon*, 2020, **6**, e03123.
- 20 S. Usha, K. T. Ramappa, S. Hiregoudar, G. D. Vasanthkumar and D. S. Aswathanarayana, *Int. J. Curr. Microbiol. Appl. Sci.*, 2017, **6**, 2219–2228.
- 21 P. Kaur, R. Thakur and A. Chaudhury, *Green Chem. Lett. Rev.*, 2016, **9**, 33–38.
- 22 S. Thakur, S. Sharma, S. Thakur and R. Rai, *Int. J. Curr. Microbiol. Appl. Sci.*, 2018, **7**, 683–694.
- 23 R. Khani, B. Roostaei, G. Bagherzade and M. Moudi, *J. Mol. Liq.*, 2018, **255**, 541–549.
- 24 S. A. Akintelu, A. S. Folorunso, F. A. Folorunso and A. K. Oyebamiji, *Heliyon*, 2020, **6**, e04508.
- 25 K. S. Siddiqi and A. Husen, *Biomater. Res.*, 2020, **24**, 1–15.
- 26 J. Nasiri, M. Rahimi, Z. Hamezadeh, E. Motamedi and M. R. Naghavi, *J. Cleaner Prod.*, 2018, **192**, 514–530.
- 27 M. R. Moghadas, E. Motamedi, J. Nasiri, M. R. Naghavi and M. Sabokdast, *Heliyon*, 2020, **6**, e04730.
- 28 R. Prusinowska and K. B. Śmigielski, *Herba Pol.*, 2014, **60**, 56–66.
- 29 K. B. Śmigielski, R. Prusinowska, K. Krosowiak and M. Sikora, *J. Essent. Oil Res.*, 2013, **25**, 291–299.
- 30 Y. S. Lin, Y. J. Tsai, J. S. Tsay and J. K. Lin, *J. Agric. Food Chem.*, 2003, **51**, 1864–1873.
- 31 A. B. Sharangi, *Food Res. Int.*, 2009, **42**, 529–535.
- 32 M. I. Din, F. Arshad, Z. Hussain and M. Mukhtar, *Nanoscale Res. Lett.*, 2017, **12**, 638.
- 33 P. Sutradhar, M. Saha and D. Maiti, *J. Nanostruct. Chem.*, 2014, **4**, 86.
- 34 A. H. Keihan, H. Veisi and H. Veasi, *Appl. Organomet. Chem.*, 2017, **31**, e3642.
- 35 R. Borah, E. Saikia, S. J. Bora and B. Chetia, *RSC Adv.*, 2016, **6**, 100443–100447.
- 36 F. Nishino, M. Jeem, L. Zhang, K. Okamoto, S. Okabe and S. Watanabe, *Sci. Rep.*, 2017, **7**, 1–11.
- 37 T. A. Shalaby, Y. Bayoumi, N. Abdalla, H. Taha, T. Alshaal, S. Shehata, M. Amer, É. Domokos-Szabolcsy and H. El-Ramady, *Nanoscience in Food and Agriculture*, 2016, **1**, 283–312.
- 38 D. H. Atha, H. Wang, E. J. Petersen, D. Cleveland, R. D. Holbrook, P. Jaruga, M. Dizdaroglu, B. Xing and B. C. Nelson, *Environ. Sci. Technol.*, 2012, **46**, 1819–1827.
- 39 W. M. Lee, Y. J. An, H. Yoon and H. S. Kweon, *Environ. Toxicol. Chem.*, 2008, **27**, 1915–1921.
- 40 M. Zaka, B. H. Abbasi, L. U. Rahman, A. Shah and M. Zia, *IET Nanobiotechnol.*, 2016, **10**, 134–140.
- 41 T. Adhikari, S. Kundu, A. K. Biswas, J. C. Tarafdar and A. S. Rao, *J. Agric. Sci. Technol. A*, 2012, **2**, 815.
- 42 P. M. G. Nair and I. M. Chung, *Biol. Trace Elem. Res.*, 2014, **162**, 342–352.
- 43 T. H. Pham, C. M. Nguyen, L. Le Quynh, K. B. Ninh, V. M. Chau, H. C. Nguyen, Q. B. Ngo, T. H. Dao, T. V. Nguyen, T. H. Le Thi and M. L. Tran, *Int. J. Agric. Biol.*, 2018, **20**, 1562–1568.
- 44 A. Hafeez, A. Razzaq, T. Mahmood and H. M. Jhanzab, *J. Nanosci Adv Technol*, 2015, **1**, 6–11.
- 45 G. F. Nekrasova, O. S. Ushakova, A. E. Ermakov, M. A. Uimin and I. V. Byzov, *Russ. J. Ecol.*, 2011, **42**, 458.
- 46 F. Gao, F. Hong, C. Liu, L. Zheng, M. Su, X. Wu, F. Yang, C. Wu and P. Yang, *Biol. Trace Elem. Res.*, 2006, **111**, 239–253.
- 47 F. Hong, J. Zhou, C. Liu, F. Yang, C. Wu, L. Zheng and P. Yang, *Biol. Trace Elem. Res.*, 2005, **105**, 269–279.
- 48 D. Y. Kim, R. G. Saratale, S. Shinde, A. Syed, F. Ameen and G. Ghodake, *J. Cleaner Prod.*, 2018, **172**, 2910–2918.

