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The metabolomic analysis of five *Mentha* species: cytotoxicity, anti-*Helicobacter* assessment, and the development of polymeric micelles for enhancing the anti-*Helicobacter* activity

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Mentha species are medicinally used worldwide and remain attractive for research due to the diversity of their phytoconstituents and large therapeutic indices for various ailments. This study used the metabolomics examination of five *Mentha* species (*M. suaveolens*, *M. sylvestris*, *M. piperita*, *M. longifolia*, and *M. viridis*) to justify their cytotoxicity and their anti-*Helicobacter* effects. The activities of species were correlated with their phytochemical profiles by orthogonal partial least square discriminant analysis (OPLS-DA). Tentatively characterized phytoconstituents using liquid chromatography high-resolution electrospray ionization mass spectrometry (LC-HR-ESI-MS) included 49 compounds: 14 flavonoids, 10 caffeic acid esters, 7 phenolic acids, and other constituents. *M. piperita* showed the highest cytotoxicity to HepG2 (human hepatoma), MCF-7 (human breast adenocarcinoma), and CACO2 (human colon adenocarcinoma) cells using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assays. OPLS-DA and dereplication studies predicted that the cytotoxic activity was related to benzyl glucopyranoside-sulfate, a lignin glycoside. Furthermore, *M. viridis* was effective in suppressing the growth of *Helicobacter pylori* at a concentration of 50 mg mL⁻¹. OPLS-DA predicted that this activity was related to a dihydroxytrimethoxyflavone. *M. viridis* extract was formulated with Pluronic® F127 to develop polymeric micelles as a nanocarrier that enhanced the anti-*Helicobacter* activity of the extract and provided minimum inhibitory concentrations and minimum bactericidal concentrations of 6.5 and 50 mg mL⁻¹, respectively. This activity was also correlated to tentatively identified constituents, including rosmarinic acid, catechins, carvone, and piperitone oxide.

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1. Introduction

The genus *Mentha* in the family Lamiaceae (Labiatae) is highly complex, including 61 species and hundreds of subspecies, varieties, and cultivars.^{1,2} Mint has been used worldwide since ancient eras in its fresh or dried forms as it displays numerous beneficial properties.^{3,4} This wide and significant bioactivity is typically correlated with the diversity of the secondary metabolites of this genus.^{3,5} Chemical profiling of *Mentha* species confirms the presence of volatile constituents and polyphenolics, including flavones, flavonols, flavanones, phenolic acids, and tannins.^{6–9} Hydroxycinnamic acid derivatives, chiefly rosmarinic acid, are widely spread in the family Lamiaceae and predominate other phenolics. These phytoconstituents show important biological and pharmacological activities, including antioxidant, antiviral, antiobesity, and antibacterial effects.^{10–13} Many investigations of *Mentha* species are available; however, the interest in studying mint and its application in



pharmaceutical formulations is continuous. All species in this genus display multiple bioactive constituents that show no side effects, even in doses ten times higher than their therapeutic doses. Therefore, they are highly safe in therapy, and investigation of their possible activities is of high importance.

Plant metabolomics is crucial for improving our understanding of metabolite levels, facilitating differentiation, quantitation, and comparing the large numbers of secondary metabolites.¹⁴ Further, metabolomics contributes significantly to the study of differences between plant phenotypic and genotypic physiology and biology when combined with functional genomics.¹⁵

Cancer is considered to be a significant global health problem, accounting for many thousands of deaths; therefore, exploring new drugs and chemotherapeutics to improve treatment are of high priority with a main target to overcome their fatal side effects. Therefore, a current focus on the potential of natural products as anticancer candidates and investigation of their activity is favored to minimize the side effects of chemically derived drugs and benefit from their high therapeutic value.^{16,17}

Moreover, peptic ulcers are considered a major global health problem, and one of their main causes is infection with *Helicobacter pylori* bacteria. This bacterium is a flagellated, Gram-negative coccobacillus that affects 50% of the world's population. Its prevalence is higher in developing countries than in developed countries.¹⁸ *H. pylori* cases are associated with severe pathologies, including gastric cancer and peptic ulcer.¹⁹ Different treatment regimens are proposed to eradicate *H. pylori*, including the use of two or more antibiotics coupled with a proton pump inhibitor.²⁰ Antimicrobial resistance is a leading cause of standard triple therapy failure in most countries, which highlights the need for alternative compounds with proven antimicrobial activity. Natural products are excellent sources that may allow the discovery of synthesized active constituents or derivatized secondary metabolites to provide new antimicrobials that successfully overcome bacterial resistance.

However, the conventional administration of natural drugs is hindered due to their low solubility, permeability, or bioavailability. Finding suitable pharmaceutical dosage forms to increase their absorption and their potency is in great demand. Nanonization may be an appropriate strategy to address these problems and improve the physicochemical properties of natural drug preparations.²¹ Polymeric micelles, as core/shell nanoscale dispersion systems formed by self-assembly of various amphiphilic block copolymers, may contribute as a dosage form to help solve this problem. These micelles are characterized by small sizes (~10–200 nm), biodegradability for easy elimination, ability to encapsulate poorly water-soluble drugs, high drug loading capacity and reproducibility, and low cost.^{22–24} Nanoparticles enhance formulation parameters and improve the therapeutic potential of hydrophobic drugs.^{25–27}

Pluronic® F127 is a biodegradable and biocompatible polymer that is used successfully to develop nanoscale polymeric micelles. The polymer promotes the drug encapsulation, permeation and therapeutic potential of various drugs.^{28–31}

The conventional use of *Mentha* species for the treatment of gastrointestinal symptoms and its inclusion in German Commission E lists for treatment of gastrointestinal

disorders,³² in most cases without identification of phenotypic plasticity and genetic variability because of extensive hybridization, triggered our interest in comparing the chemical profiles of five *Mentha* species cultivated in Egypt: *M. suaveolens*, *M. sylvestris*, *Mentha* × *piperita*, *M. longifolia*, and *M. viridis*. Chemical components were assessed using liquid chromatography with high-resolution electrospray ionization mass spectrometry (LC-HR-ESI-MS), followed by principal component analysis (PCA). The objectives were to assess the cytotoxic and anti-*Helicobacter* activities of different extracts and compare them by orthogonal partial least squares discriminant analysis (OPLS-DA) as well as to develop Pluronic® F127 polymeric micelles as a nanocarrier for the most effective *Mentha* extract. Further, in the study, we characterized the prepared systems to investigate their improved antimicrobial activity against *H. pylori* as an adjuvant treatment for peptic ulcer.

2. Material and experimental methods

2.1. Plant material and extraction procedure

The medicinal plants studied were *Mentha suaveolens*, *M. sylvestris* and *M. longifolia* (L.) Huds sub-spp. schimperii Briq. The voucher samples were kindly authenticated by Dr Gemma L.C. Bramley, Curator of the Lamiaceae collection Herbarium Department, Library, Art & Archives Directorate, Royal Botanic Gardens, Kew, Richmond Surrey, U.K., Voucher No. M-20/313, 20-01-2020 II, and 25-6-2015, respectively. *M. piperita* and *M. viridis* were kindly authenticated by Dr Mohamed El-Gebaly, Senior Botanist at El-Orman Botanic Garden, Egypt (Voucher No. 20-01-2020-I, 20-01-2020 V respectively). The leaves of each plant under study were collected from the Experimental Station of Medicinal and Aromatic Plants, Department of Pharmacognosy, Faculty of Pharmacy, Cairo University, Cairo, Egypt, in March 2019. Voucher specimens were deposited in the Herbarium of the Department of Pharmacognosy, Faculty of Pharmacy, Cairo University, Cairo, Egypt. The fresh leaves were air-dried at room temperature, then ground into a fine powder. 100 g of each sample was extracted with 70% ethanol (3 × 200 mL) for 30 min in a water bath at 50 °C.³³ The samples were then cooled and filtered, and the solvent was evaporated at 45 °C using a rotary evaporator (Buchi R-300, USA) and stored at 4 °C until further use.

2.2. Anti-proliferative assay

2.2.1. Cytotoxic activity. The cytotoxicity of the 70% ethanolic extracts was evaluated in cell lines using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay.³⁴ HepG2 (human hepatoma), MCF-7 (human breast adenocarcinoma), and CACO2 (human colon adenocarcinoma) cells were obtained from the American Type Culture Collection (ATCC, Rockville, MD, USA; HPACC, Salisbury, UK) and were maintained in RPMI medium (Merck, Darmstadt, Germany) supplemented with 10% fetal bovine serum (FBS). Cancer cells were cultured at 37 °C, 5% (v/v) CO₂ in RPMI1640 medium supplemented with 5% (v/v) fetal bovine serum (FBS), 1% (w/v) L-glutamine, 1% sodium pyruvate and 0.4% (w/v) antibiotics (50 U mL⁻¹



penicillin, 50 mg mL⁻¹ streptomycin). Cells were routinely sub-cultured twice per week. All chemicals and reagents were purchased from Sigma Aldrich (Darmstadt, Germany). To normalize the cell viability values, each plate included a triplicate of cells treated with the compound carrier DMSO to define 100% viable cells as well as a triplicate of cells incubated with a cytotoxic mixture (200 ng mL⁻¹ Tumor Necrosis Factor (TNF), 200 ng mL⁻¹ CD95L (Fas ligand), 200 ng mL⁻¹ TNF-related apoptosis-inducing ligand (TRAIL), 25 g mL⁻¹ cycloheximide (CHX), 1% (w/v) sodium azide) to define maximal cell death and, thus, 0% viability. All other viability values were normalized according to the averages of these triplicates and analyzed by Graph Pad Prism 5 software (La Jolla, CA, USA).

2.3. Screening of the anti-*Helicobacter* activity of *Mentha* species

2.3.1. Strain of *Helicobacter pylori* and culturing. *Helicobacter pylori* ATCC® 43504™ was kindly obtained from the Microbiology Department (Faculty of Medicine) of October 6 University Hospital, Giza, Egypt, on chocolate agar plates. Pure colonies were sub-cultured directly onto supplemented Columbia blood agar with DENT after a positive urease test; then, Gram staining of a smear prepared from the pure colonies was performed. The cultures were incubated for a minimum of 3 days and a maximum of 7 days at 37 °C under microaerophilic conditions (CO₂ 5%). The *H. pylori* strain was identified by colony morphology, observing the characteristic spiral shape in the Gram staining (Gram-negative), and positive oxidase and catalase tests. The isolates were stored at -80 °C for 5 to 10 months in Columbia broth medium containing 15% glycerol.

2.3.2. Disc diffusion test. Kirby-Bauer disc diffusion was used to analyze the susceptibility of *H. pylori* ATCC® 43504™ to different *Mentha* species extracts.^{35,36} Each extract residue (50 mg) was dissolved in one mL of dimethyl sulfoxide (DMSO), sterilized through a Millipore filter (0.22 µm) and then loaded on a sterile filter paper disc (6 mm in diameter). Ten mL of supplemented Columbia blood agar (CBA) medium was poured into sterile Petri dishes (as a basal layer), followed by surface inoculation of *H. pylori* suspensions (100 mL of broth medium/3 mL of 10⁸ CFU) to attain a 0.5 MacFarland turbidity standard (0.5 × 10⁸ CFU mL⁻¹). Sterile filter paper discs loaded with *Mentha* species extracts (50 mg mL⁻¹) were placed on top of the supplemented Columbia blood agar plates. Discs (6 mm diameter, Hi-Media, India) with 10 µg per disc of metronidazole and DMSO were used as a positive and a negative control against *H. pylori*, respectively. The inhibition zones were measured by a Vernier caliper, recorded and considered as an indication of antibacterial activity.³⁷ The antimicrobial activity was determined in triplicate.

2.4. LC-MS analysis

An Agilent 1100 HPLC Series system (Agilent, USA) equipped with a degasser, binary gradient pump, column thermostat, autosampler, and U.V. detector was used. The HPLC system was coupled with an Agilent 1100 mass spectrometer (LC-MS analysis Ion Trap V.L.). For the separation, a reverse-phase analytical column was employed (Zorbax SB-C18, 100 × 3.0 mm i.d., 3.5 µm particle);

the work temperature was 48 °C. The detection of the compounds was performed in both U.V. and M.S. mode. The U.V. detector was set at 330 nm until 17.5 min; then, it was set at 370 nm. The MS system was operated using an electrospray ion source in negative and positive modes. The chromatographic data were processed using ChemStation and DataAnalysis software from Agilent, USA. The mobile phase was a binary gradient: methanol and acetic acid 0.1% (v/v). The elution started with a linear gradient, beginning with 5% methanol and ending at 42% methanol, for 35 minutes; then, 42% methanol was used for the next 3 minutes. The flow rate was 1 mL min⁻¹ and the injection volume was 5 µL. Compounds were qualitatively identified based on spectral matching of the signals and spectra of each polyphenol with the available library and literature.

2.5. Preparation of the most effective *Mentha* extract polymeric micelles

Preliminary screening using different concentrations of the most effective *Mentha* extract and polymer ratios were performed. Extract polymeric micelles with a 100 mg mL⁻¹ concentration and an extract-to-polymer ratio of 1 : 3 were finally developed. The formulae were prepared using the thin-film hydration method. Weights of 500 mg extract and 1.5 g Pluronic® F127 were completely dissolved separately in 5 mL methanol. Then, the two solutions were combined, and the mixture was stirred for a further 15 min. The solvent was evaporated to form a dried thin film of the extract and polymer matrix using either of the following methods. In the first method, the mixture was transferred to a porcelain dish and then was placed in a domestic microwave (Microwave, LG, 2450 MHz, China) operating at a power of 1000 W for a short pulse of 1 min for 10 cycles to achieve complete drying. For the second method, the mixture was transferred to a round flask connected to a rotary evaporator (Rotavapor, Heidolph VV 2000, Burladingen, Germany) rotated at a rotation speed of 120 rpm and 50 °C for one h under vacuum. The obtained dry film for each case was hydrated with 5 mL double distilled water and stirred for 30 min to obtain a homogeneous micellar dispersion (100 mg mL⁻¹). The prepared formulae were stored in tightly closed containers. They were coded MvP-M for the formula prepared using a microwave and MvP-R for the formula designed using the rotary evaporator.

2.6. Characterization of the prepared *Mentha* polymeric micelles

2.6.1. Visual inspection and determination of particle size (PS), polydispersity index (PDI) and zeta potential (ZP). The prepared formulae were examined visually for appearance and homogeneity.

The PS and PDI of 100-fold diluted formulae were determined at 25 °C by dynamic light scattering (DLS) using a Zetasizer (Nano Z.S., Malvern, U.K.). Also, the ZP was measured to investigate the physical stability. Measurements were performed in triplicate at 25 °C.

2.6.2. Screening of anti-*Helicobacter* activity. The anti-*Helicobacter* activity of the prepared formula was determined using the disc diffusion test described in Section 2.4.2 using



a concentration of 100 mg mL⁻¹ for the most potent *Mentha* extract and its formulae.

2.6.3. Determination of minimum inhibitory concentrations (MICs) and minimum bactericidal concentrations (MBCs) of the most effective *Mentha* extract and polymeric micelles. The most effective *Mentha* extracts (dissolved in DMSO), MvP-M and MvP-R, were manipulated to determine their MICs and MBCs. Broth microdilution was performed with 96-well U-shaped bottom microtiter plates following Clinical and Laboratory Standards Institute (CLSI) guidelines (2017).^{38,39}

The tested samples were used in an initial concentration of 100 mg mL⁻¹ and then serially diluted two-fold to obtain concentrations in the range of 0.098–50 mg mL⁻¹. The inoculum of *H. pylori* was prepared in Columbia broth, with the OD_{600 nm} adjusted to 0.5. Equal volumes (50 µL) of broth and different concentrations of each sample were incubated at 37 °C for 72 h under microaerophilic conditions. Bacteria-free broth and 50 µL DMSO were used as a negative control. The bacterial density was read at 620 nm using an ELISA reader. The lowest concentration at which no bacterial density was determined was the MIC value. The experiment was performed in triplicate.⁴⁰

For MBC determination, 10 µL of mixtures containing *Mentha* extract and *H. pylori* as well as MvP-M and MvP-R (different concentrations) were inoculated on respective agar plates (as detailed above), and the bacterial counts were determined and expressed as CFU mL⁻¹ after incubation at 37 °C for 72 h. The MBC was determined as the lowest concentration at which the incubated *H. pylori* strain showed no detectable colonies on the respective agar plates. The experiment was performed three independent times.

2.6.4. Surface morphology studies by transmission electronic microscopy (TEM). Based on the previous characterization results, the formula with the best criteria was selected. The morphology of the selected formula was investigated by TEM (JEM-2100, Jeol® Tokyo, Japan). TEM offers valuable information on the inner structure of a sample, such as its crystal structure, morphology, and stress state information.^{41,42}

A drop of a diluted aqueous dispersion of the formula was dropped onto copper-coated carbon grids, dried, and stained with 2% w/v phosphotungstic acid solution; then, the excess was absorbed by filter paper.

2.7. Statistical analysis

Statistical analysis of the data was performed using one-way ANOVA followed by Turkey multiple range tests. All the data are presented as the mean ± SE of 3 determinations. The level of significance was set at $p < 0.05$. Statistical calculations were performed using SPSS version 15.0 software. The p value significance is represented as an asterisk (*) for $p < 0.05$, two asterisks (**) for $p < 0.01$ and three asterisks (***) for $p < 0.001$.

3. Results and discussion

3.1. Cytotoxic activity

The five extracts were tested for their cytotoxic activity against three human cancer cell lines (Table 1). Only *M. piperita* was

Table 1 Cytotoxic activities of the crude extracts (IC₅₀ µg mL⁻¹)^a

Tested sample (70% ethanolic extracts)	HepG-2 (liver)	MCF-7 (breast)	CACO2 (colon)
<i>M. suaveolens</i> ***	52.19 ± 5.6	55.07 ± 4.9	61.06 ± 4.8
<i>M. sylvestris</i> ***	51.30 ± 3.9	52.81 ± 5.5	58.11 ± 6.3
<i>M. piperita</i> *	11.52 ± 4.2	7.97 ± 5.6	13.72 ± 3.3
<i>M. longifolia</i> ***	54.23 ± 2.3	56.54 ± 7.2	61.25 ± 5.9
<i>M. viridis</i> ***	58.61 ± 4.3	51.74 ± 6.4	62.36 ± 4.3
Doxorubicin	5.72 ± 0.7	5.17 ± 0.6	5.81 ± 0.5

^a The results were obtained from three independent replicate experiments and expressed as means ± standard deviation. The statistical significance of the results was tested using one-way Analysis of Variance (ANOVA) and Tukey–Kramer multiple Comparisons Test between Doxorubicin and the different plant extracts. The p value significance was represented as an asterisk (*) for $p < 0.05$ and three asterisks (***) for $p < 0.001$.

able to induce cytotoxicity towards the tested cell lines, with IC₅₀ values ranging from 7–13 µg mL⁻¹. Previous studies showed the powerful anticancer effect against breast cell carcinoma of *M. piperita* leaf extract ethyl acetate, with cellular apoptosis, inhibition of proliferation, perturbation of oxidative balance, and upregulation of Bax gene.⁴³ Additionally, the reported phenolics were demonstrated to scavenge ·OH and superoxide free radicals and to inhibit lipid peroxidation.^{44,45}

3.2. Anti-*Helicobacter* activity of *Mentha* species

Different *Mentha* species were investigated for their effects against the *H. pylori* strain using the disc diffusion method. The results are recorded in Table 2. *M. viridis*, *M. piperita*, and *M. suaveolens* caused suppression of bacterial growth at a concentration of 50 mg mL⁻¹, with inhibition zones of 14 mm, 12 mm, and 10 mm, respectively. *M. viridis* was the most effective extract in retarding microbial growth of all the tested isolates. In contrast, extracts of *M. sylvestris* and *M. longifolia* were not effective (no zone of inhibition). Essential oils of *M. viridis* and *M. piperita* were reported for their anti-*Helicobacter* activities.⁴⁶

3.3. Tentative identification of the major phytoconstituents in *Mentha* species

Metabolites were identified in the five *Mentha* species using LC/MS in both positive and negative modes, where the peaks were identified using an in-house library. Forty-seven compounds belonging to different classes have been tentatively identified, including flavonoids (13), hydroxycinnamic esters (12), organic acids, phenolic acids, ascorbic acid, and riboflavin (22). The identities, retention times, and observed molecular and fragment ions of the tentatively identified components are presented in Table 3. The caffeic acid esters identified were salvianolic acid, chlorogenic acid, salvianolic acid B, sagerenic acid, rosmarinic acid, salvianolic acid A, I, and dihydrodrosalvianolic acid, which are widespread in all the studied plants. Other metabolites which were common in all species include isomenlactone (perillic acid), naringin, ladanin (dihydroxy, dimethoxyflavone), vanillyl alcohol, 11-cisretinoic acid,



Table 2 Antibacterial activity tests of *Mentha* species (50 mg mL⁻¹) against *Helicobacter pylori*^{a,b}

	<i>Helicobacter pylori</i> ATCC® 43504™
Plant extracts	Zone of inhibition (mm)
<i>M. suaveolens</i> (50 mg mL ⁻¹)**	10 ± 2.0
<i>M. sylvestris</i> (50 mg mL ⁻¹)	—
<i>M. piperita</i> (50 mg mL ⁻¹)**	12 ± 1.5
<i>M. longifolia</i> (50 mg mL ⁻¹)	—
<i>M. viridis</i> (50 mg mL ⁻¹)**	14 ± 1.5
Positive control (10 µg per disc of metronidazole)	17 ± 0

^a $n = 3$, mean ± SD. ^b The results were obtained from three independent replicate experiments and expressed as means ± standard deviation. The statistical significance of the results was tested using one-way Analysis of Variance (ANOVA) and Tukey–Kramer multiple Comparisons Test. The p value significance was represented two asterisks (**) for $p < 0.01$.

carvone, eugenol, riboflavin, antheraxanthin, 9-*cis* retinol acetate, and piperitone oxide. Those results were in agreement with those reported for other *Mentha* species.^{47–49} *M. viridis*, which showed the best anti-*Helicobacter* activity, was tentatively identified to contain a variety of active constituents, including protocatechuic, shikimic, homovanillic, ferulic, salvianolic, sagerinic, and rosmarinic acids, in addition to the presence of flavonoids, such as gallo catechin, hesperidin, epicatechin, and naringin, and monoterpenes, such as carvone and piperitone oxide. Catechins have been reported to eradicate *H. pylori* when in combination with sialic acid.⁵⁰ Rosmarinic acid was also proved to have an excellent drug-like profile in an *in silico* screening of shikimate kinase, showing a possible synergistic effect with many antibiotics.⁵¹ Additionally, monoterpenes have been proved to have gastroprotective and possible anti-*Helicobacter* effects.⁵² Meanwhile, *M. piperita*, which showed the best cytotoxic activity, was distinguished by its phenolic acid as well as its flavonoid contents in addition to the presence of salvadoside, a lignan glycoside previously identified in *Salvadora persica*;⁵³ these epoxy carotenoids represent lipophilic-active compounds of xanthophylls in foods and play a great role in cancer prevention,^{54,55} including violaxanthin, previously identified in *Mentha* species,⁴⁸ and the main constituent in *Dunaliella tertiolecta*, which demonstrated a powerful antiproliferative effect against breast cell carcinoma⁵⁶ in addition to antheraxanthin.

3.4. Metabolomics analysis

To widen the coverage of the metabolome of *Mentha*, the positive and negative mass spectral data were combined into one data matrix. The data were processed via MZmine2 according to a method developed previously in our lab.⁵⁷ The processed data were then moved into an in-house database Excel file with a built-in Dictionary of Natural Products Database (DNP) for dereplication purposes.⁵⁸ Principle component analysis (PCA) was performed to test the similarity and/or the variation in the chemical profiles among the tested species. PCA is an unsupervised multivariate data analysis that aims to reduce the dimensionality of the data to reveal clusters, groups, and/or outliers among the observations.⁵⁹ The PCA score plot (Fig. 1A) showed respective total variances of 81% and 10% for

PC1 and PC2 and demonstrated the clustering of *M. suaveolens* and *M. sylvestris* extracts (UA-62 and UA-64, respectively). This result indicated the similar chemical fingerprints of those extracts. Moreover, PCA showed a dispersal of *M. viridis* (UA-66), *M. piperita* (UA-65) and *M. longifolia* (UA-63) extracts, which demonstrated their unique chemical profiles. The PCA loading plot (Fig. 1B) highlighted the metabolites that contributed to this variation, as *M. viridis* (UA-66) extract was characterized by molecules at m/z (retention time in minutes) 345.097 $[M + H]^+$ (t_R 5.65) and 305.069 $[M - H]^-$ (t_R 2.48). The former corresponds to a dihydroxytrimethoxyflavone, equivalent to a molecular formula of $C_{18}H_{17}O_7^+$, while the latter is equivalent to the molecular formula of $C_{15}H_{13}O_7^-$, which corresponds to gallo catechin in the DNP database. Moreover, *M. piperita* extract (UA-65) was distinguished by the presence of discriminatory metabolites at m/z $[M - H]^-$ 349.059 (t_R 2.22) and 408.042 (t_R 1.75). They are respectively equivalent to $C_{13}H_{18}O_9S$, which corresponds to benzyl glucopyranoside-sulfate or salvadoside. Sulphated glycosides have been previously reported in *Mentha* × *piperita*.⁴⁷ To investigate which compounds are mediating the biological activity of *M. piperita* extract, a supervised multivariate data analysis was carried out. Orthogonal partial least squares discriminant analysis (OPLS-DA) was applied to highlight the molecules that are highly correlated with the biological activity. The OPLS-DA score plot (Fig. 2A) demonstrated complete discrimination between the active extract *M. piperita* (UA-65) and the other inactive *Mentha* species. The model's measures for the goodness of fit $R^2 = 0.91$ and prediction $Q^2 = 0.67$ indicate a strong fit model with a high predictive power. The coefficient of variation plot (Fig. 2B) is a very useful tool to compare the variable magnitude against its reliability, where regression coefficients related to center-scaled X-variables are displayed. This scaling of the data enables comparison of the coefficients. Thus, these coefficients express how strongly Y (biological activity) is correlated to the systematic part of each of the X-variables. The molecules highly correlated with *M. piperita* activity were checked, and only those with high coefficients of variation, with high VIP (variable importance), and whose 95% confidence level limits did not cross zero were chosen. The descriptive statistics of the model, including the coefficient of





Table 3 List of identified metabolites in different *Mentha* species

t_R	$M + H^+ / M - H^-$	MS/MS	Error (ppm)	Formula	Name	Class	<i>M. suaveolens</i>	<i>M. longifolia</i>	<i>M. sylvestris</i>	<i>M. piperita</i>	Ref.
0.4	169.0414	169 (125)	0.0272	$C_7H_5O_5^-$	Gallic acid	Phenolic acid	—	—	+	—	70
0.66	191.055	111, 173	0.0563	$C_6H_7O_7^-$	Citric acid	Organic acid	+	—	+	+	71
0.67	165.0584		0.034	$C_{10}H_{13}O_2^-$	Isomenlactone (perillic acid)	Phenolic acid	+	+	+	+	72
0.77	167.055		0.02	$C_8H_7O_4^-$	Homoprotocatechuic acid (3,4-dihydroxyphenyl acetic acid)	Phenolic acid	—	+	+	+	
1.77	153.0453		0.026	$C_7H_5O_4^-$	Protocatechuic acid (2,3-dihydroxybenzoic acid)	Phenolic acid	—	+	+	+	73
1.88	173.0637		0.0182	$C_7H_{11}O_5^+$	Shikimic acid, 3,4,5-trihydroxy-cyclohex-1-ene-1-carboxylic acid	Phenolic acid	—	+	+	+	48
1.89	377.1614		0.0158	$C_{17}H_{21}N_4O_6^+$	Riboflavin	Flavins	+	+	+	+	48
2.01	175.0791		0.0543	$C_6H_7O_6^-$	Ascorbic acid	Organic acid	—	+	+	+	48
2.03	353.0867	191, 179, 173	0.0011	$C_{16}H_{17}O_9^-$	Chlorogenic acid	Caffeic acid and its esters	+	+	—	—	48 and 70
2.12	165.0913		0.003	$C_{10}H_{13}O_2^+$	Eugenol	Phenylpropanoids	+	+	+	+	1 and 74
2.22	349.059	325, 309, 293, 265, 232, 218	0.0054	$C_{13}H_{17}O_9S^-$	Salvadoside (benzyl B-D-glucopyranoside sulphate)	Glycosides	—	—	+	—	47
2.25	181.0693		0.0187	$C_9H_9O_4^-$	Homovanillic acid	Phenolic acid	—	—	+	+	75 and 76
2.40	197.0444	179	0.0011	$C_9H_{10}O_5$	Salvianilic acid (vanillylmandelic acid)	Caffeic acid and its esters	+	—	+	+	47
2.48	305.0694	225, 179	0.0020	$C_{15}H_{13}O_7^-$	Gallocatechin	Caffeic acid ester	+	+	—	+	76
3.09	593.1507	285	0.0005	$C_{27}H_{29}O_{15}^-$	Dihydroxyflavone, rutinoid (luteolin rutinoside)	Caffeic acid and its esters	+	+	—	—	47
3.16	329.1609	307, 293, 265, 189, 171	0.0149	$C_{20}H_{25}O_4^-$	Carnosol	Diterpene lactones	—	+	—	+	77
3.28	581.1767		0.0098	$C_{27}H_{33}O_{14}^+$	Naringin	Flavonoid-7-O-glycosides	+	+	+	+	76 and 78
3.32	579.1711	556, 481, 447, 337, 365, 297	0.0002	$C_{27}H_{31}O_{14}^+$	Apigenin-7-rutinoside	Flavone-O-glycoside	—	+	+	—	47
3.41	717.1461	519, 359, 321	0.000	$C_{36}H_{29}O_{16}^-$	Salvianolic acid B	Caffeic acid and its esters	+	—	—	—	4
3.49	169.1213		0.001	$C_{10}H_{17}O_2^+$	Piperitone oxide	Monoterpene oxide	+	+	+	+	79
3.49	169.1607		0.001	$C_{11}H_{21}O^+$	Mentha-3-en-8-ol	Monoterpene alcohols	+	+	+	—	
3.52	609.1848	297, 279, 217, 167	0.0034	$C_{28}H_{33}O_{15}^+$	Diosmetin rutinoside	Flavone-O-glycoside	—	+	—	—	47
3.61	715.1922	337, 319, 293	0.0618	$C_{36}H_{27}O_{16}^-$	Didehydrosalvianolic acid B	Caffeic acid and its esters	—	—	—	+	47
3.63	611.1991	600, 556, 37, 297, 243, 217	0.0587	$C_{28}H_{35}O_{15}^+$	Hesperidin	Flavanone-O-glycosides	+	+	—	+	47
3.68	719.1609	537, 493, 359, 295	0.0033	$C_{36}H_{31}O_{16}^-$	Sagerenic acid	Caffeic acid der	+	—	+	+	4
3.77	165.0627/163.0778		0.0166	$C_9H_9O_3^+$	Coumaric acid	Phenolic acid	—	+	+	+	73
3.96	359.0762	197, 179, 161	0.0010	$C_{18}H_{15}O_8^-$	Rosmarinic acid	Caffeic acid der.	+	+	+	+	4
3.98	193.0919	134, 149, 179	0.0412	$C_{10}H_9O_4^-$	Ferulic acid	Caffeic acid der.	—	+	+	+	80
4.08	537.1025	493, 359, 295	0.0013	$C_{27}H_{21}O_{12}^-$	Salvianolic acid I (lithospermic acid)	Caffeic acid der.	+	—	—	—	4



Table 3 (Contd.)

t_R	$M + H^+ / M - H^-$	MS/MS	Error (ppm)	Formula	Name	Class	<i>M. suaveolens</i>	<i>M. longifolia</i>	<i>M. sylvestris</i>	<i>M. piperita</i>	<i>M. viridis</i>	Ref.
4.10	493.1132	359, 313, 295	0.0008	$C_{26}H_{21}O_{10}^-$	Salvanolic acid A	Caffeic acid deriv. Flavones	+	—	—	—	—	4
4.52	299.0848	284, 265, 239, 233, 171, 163	0.0287	$C_{16}H_{11}O_6^-$	Trihydroxymethoxyflavone		+	+	+	—	+	9
4.62	301.091	299, 284, 265, 171, 163	0.0193	$C_{16}H_{13}O_6^-$	Hesperitine	Flavanone-O-glycosides	—	+	+	—	—	47
4.65	151.1108		0.001	$C_{10}H_{15}O^+$	Carvone	Menthane monoterpenoids	+	+	+	+	+	81
4.76	181.1048	173, 167, 158	0.0173	$C_6H_{13}O_6^+$	Caffeic acid	Caffeic acid	+	+	+	—	+	80
4.99	271.044		0.0172	$C_{15}H_{11}O_5^-$	Naringenin	Flavanones	—	+	—	—	—	47
5.17	344.0955	329, 309, 293, 265	0.9661	$C_{17}H_{13}O_8^-$	Axillarin	6-O-methylated flavonoids	—	—	—	—	+	82
5.65	345.0971		0.0002	$C_{18}H_{17}O_7^+$	Dihydroxytrimethoxyflavone	Flavones	+	—	—	—	+	9
6.229	315.0813	261, 217, 167, 158	0.005	$C_{17}H_{15}O_6^+$	Ladanin (dihydroxy, dimethoxyflavone)	Flavonoids	+	+	+	+	+	9
6.64	287.1198	285, 268, 171, 163	0.0637	$C_{15}H_{11}O_6^-$	Dihydrokaempferol	Flavonoids	—	+	—	—	—	83
7.01	199.1693		0.000	$C_{12}H_{23}O_2^+$	Menthyl acetate	Monoterpene alcohols	+	+	+	—	+	84
7.19	157.1599		0.0012	$C_{10}H_{21}O^+$	Menthol	Monoterpene alcohols	—	+	—	—	—	4
7.98	601.3801		0.0443	$C_{40}H_{57}O_4^+$	Violaxanthin	Xanthophylls	+	+	—	+	+	48
9.76	585.4408		0.0106	$C_{40}H_{57}O_3^+$	Antheraxanthin	Xanthophylls	+	+	+	+	+	48
9.77	301.1919		0.0243	$C_{20}H_{29}O_2^+$	11- <i>cis</i> -Retinoic acid	Retinoids	+	+	+	+	+	48
9.96	291.1443	261, 229, 217, 187	0.0580	$C_{15}H_{15}O_6^+$	Epicatechin	Flavonoids	—	—	—	+	+	78
10.39	329.2513		0.0467	$C_{22}H_{33}O_2^+$	9- <i>cis</i> -Retinol acetate	Retinoids	+	+	+	+	+	48
11.59	569.3819	556, 297, 261	0.0534	$C_{40}H_{57}O_2^+$	Zeaxanthin	Xanthins	—	—	—	+	—	48

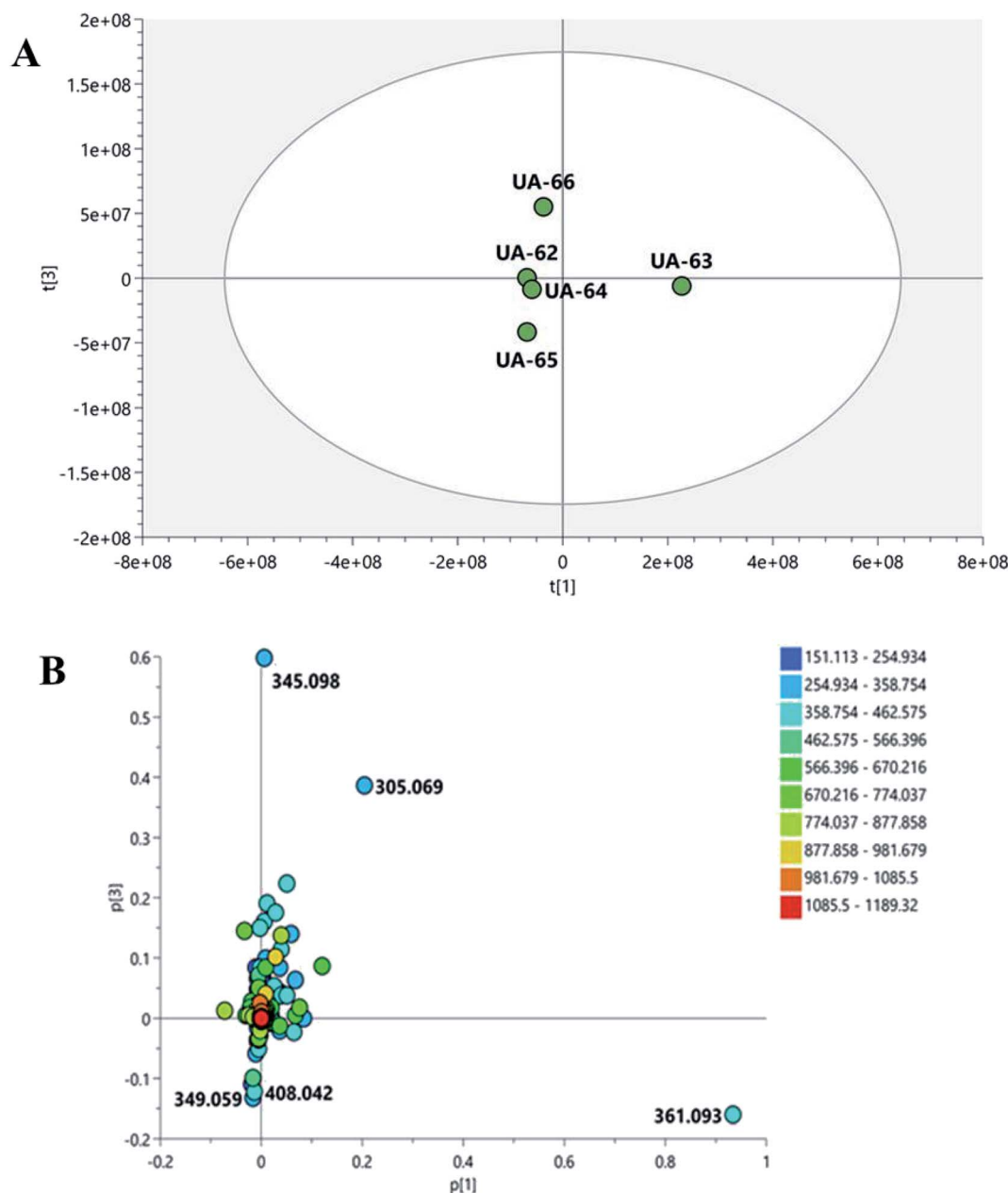


Fig. 1 Principal component analysis (PCA) of different *Mentha* species: score plot (A) and loading plot (B) of the high-resolution mass spectrometry (LC-HRMS) data.

variation, VIP, and 95% confidence limits, led to the significant elements at m/z $[M - H]^-$ 349.059 (VIP 5.35, t_R 2.22) and 408.042 (VIP 5.15, t_R 1.75), which were highlighted previously in the principle component analysis as distinguishing for *M. piperita*. The molecule at m/z 349.059, equivalent to $C_{13}H_{17}O_9S^-$, was dereplicated by DNP as benzyl glucopyranoside-sulfate or salvadoside. This lignin glycoside was reported previously in *Salvadora persica*,⁵³ *Tribulus terrestris*,⁶⁰ *Tricalysia dubia*,⁶¹ and *Dillenia philippinensis*.⁶² It was only tested against *Leishmania* and A549 human lung adenocarcinoma cells and showed no significant effect.⁶² The OPLS-DA score plot (Fig. 3A) demonstrated complete discrimination between the active extract *M. viridis* (UA-66). The molecules highly correlated with *M. viridis*

activity were checked, and only those with high coefficients of variation, with high VIP (variable importance), and whose 95% confidence level limits did not cross zero were chosen. The descriptive statistics of the model, including the coefficient of variation, VIP and 95% confidence limits, led to the identification of significant elements at m/z $[M + H]^+$ 345.097 (VIP 10.38, t_R 5.65) identified as dihydroxytrimethoxyflavone and equivalent to $C_{18}H_{17}O_7^+$, in addition to 375.108 (VIP 4.98, t_R 6.008) and 359.114 (VIP 5.18, t_R 6.91) Methoxylated flavones have shown their possible potential as anti-*Helicobacter* agents⁶³ and were highlighted previously in principle component analysis of *M. viridis* and other species.

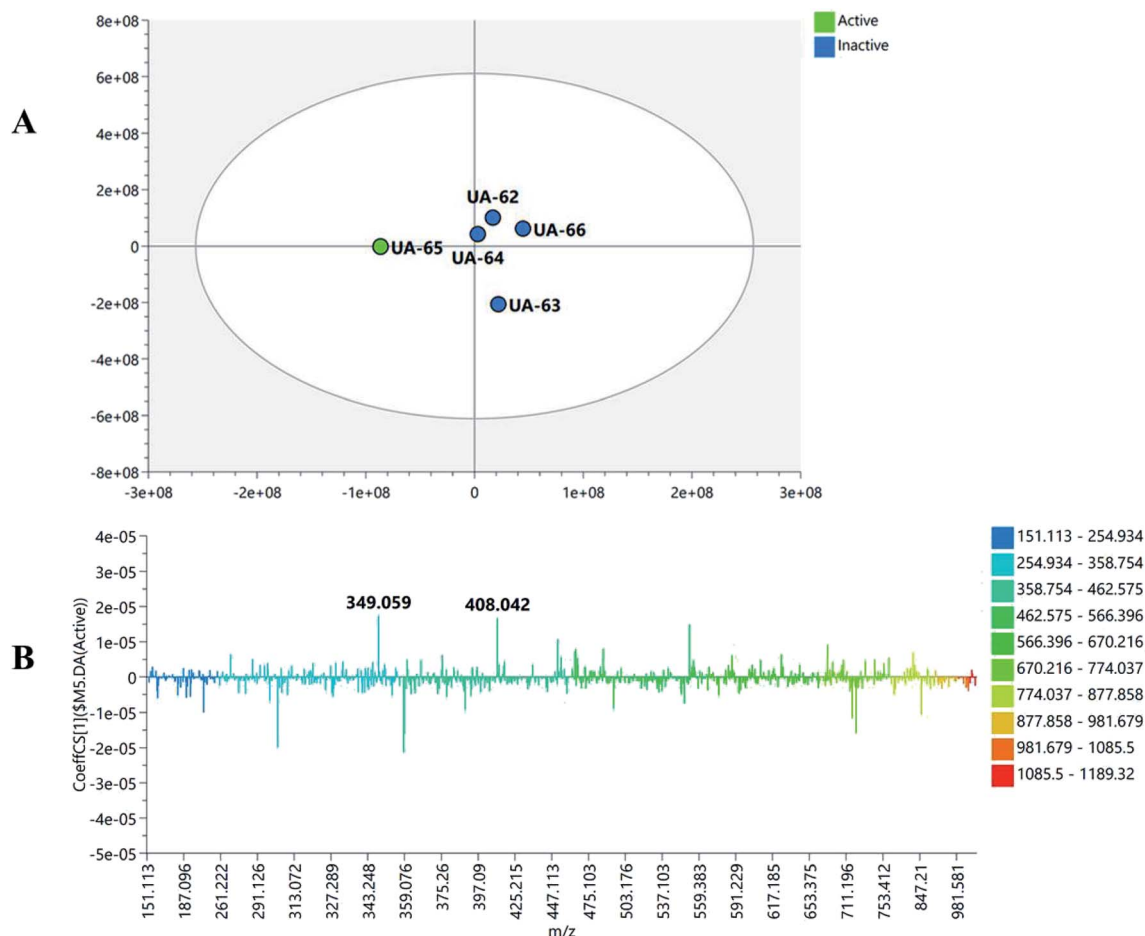


Fig. 2 (A) Orthogonal partial least squares discriminant analysis (OPLS-DA) score plot of the active fractions (green circle) versus inactive fractions (blue circle). (B) S-Loading plot showing the putatively active metabolites for cytotoxic activity.

3.5. Preparation of *M. viridis* polymeric micelles

Two Pluronic® F127 micellar formulae loaded with *M. viridis* extract were prepared at a concentration of 100 mg mL⁻¹ with an extract-to-polymer ratio of 1 : 3 to improve the solubility and antimicrobial activity of the extract. Two methods were used to form the dried thin film of the extract and polymer mixture for preparation of the micelles: a microwave method and a rotary evaporator method. The rotary evaporator method is commonly used for the preparation of polymeric micelles.^{29–31} However, the introduction of microwave offers a faster alternative mode of drying.⁶⁴ The microwave was used successfully for the development of different formulations.^{65–67}

3.6. Characterization of *M. viridis* polymeric micelles and their anti-*Helicobacter* potential

Visual inspection revealed that both MvP-M and MvP-R were fluid homogeneous dispersions showing no agglomerations or phase separation. The results of the mean size diameter, PDI, zeta potential, and antimicrobial assessment are recorded in Table 4. Both formulae showed particle sizes in the nanometer range with PDI values less than 0.4, indicating good size distributions. Also, both formulae had high zeta potentials, indicating good physical stability of the prepared micelles.

However, a significant difference (at $p < 0.05$) was recorded for the antimicrobial activity data, as illustrated by a higher zone of inhibition and lower MIC and MBC for MvP-R compared with both MvP-M and the plain *M. viridis* extract.

The MIC and MBC results confirmed the potency of *M. viridis* extract against the *H. pylori* strain. Meanwhile, incorporation of the extract in Pluronic® F127 polymeric micelles enhanced this activity significantly; it displayed lower MIC and MBC values of 6.25 and 50 mg mL⁻¹ against *H. pylori*, whereas the extract showed values of 25 and 100 mg mL⁻¹, respectively.

The ability of the polymeric micelles formula (MvP-R) to improve the antimicrobial activity is due to the increased drug solubility and the size of the nanocarriers, which enhance the penetration of the extract micelles through the bacterial cell wall and improve their efficacy against the target microorganism.^{68,69} Unfortunately, MvP-M showed lower antimicrobial activity than the plain extract, as the usage of the microwave in preparation of this formula could cause an increase of temperature during drying of extract/polymer dispersion. The elevated temperature may cause partial degradation of the extract constituents and, consequently, a reduction of their antimicrobial activity.⁶⁴

The morphology of the MvP-R formula under TEM (Fig. 4) shows spherical structures of the prepared *M. viridis* polymeric



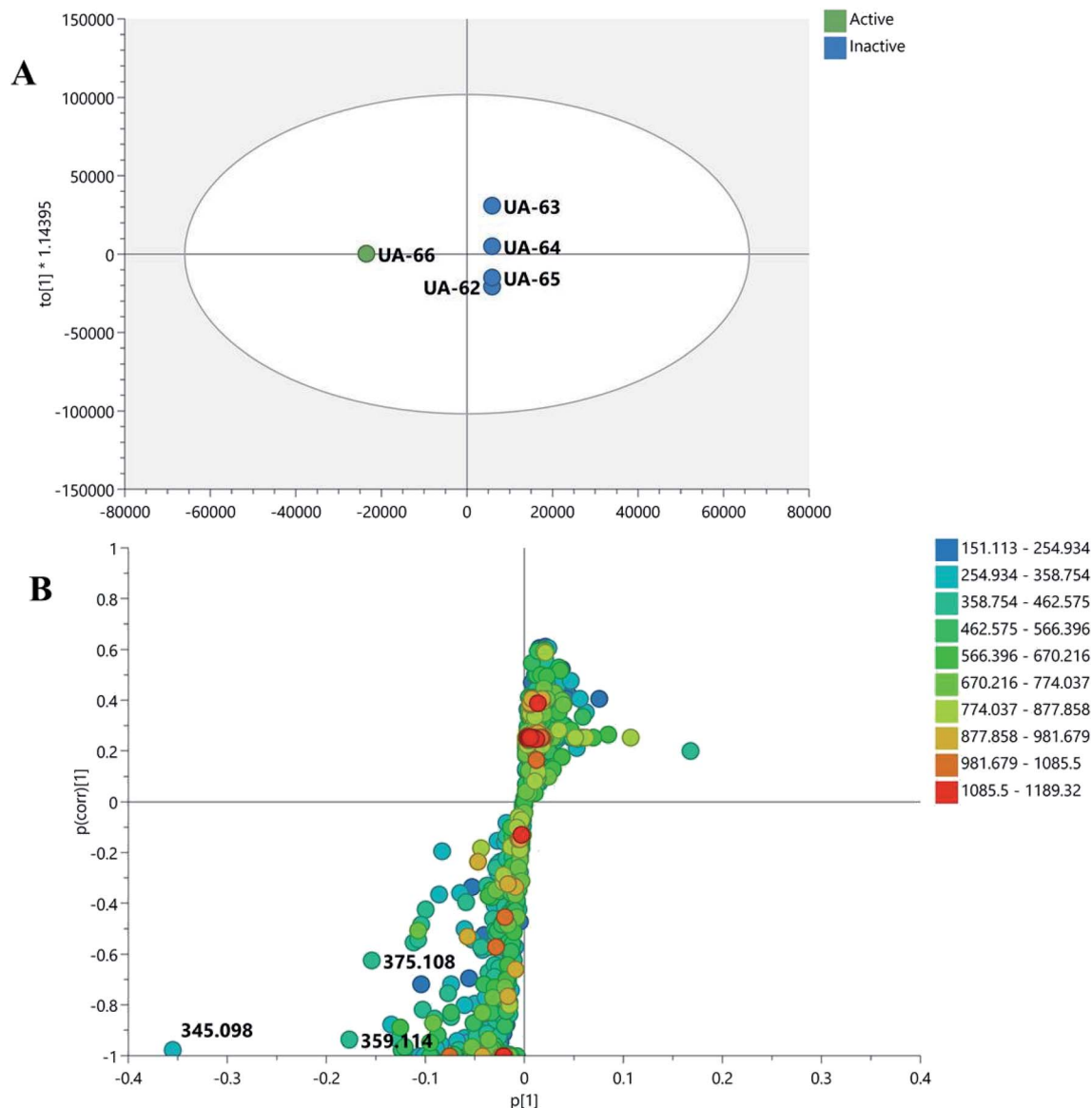


Fig. 3 (A) Orthogonal partial least squares discriminant analysis (OPLS-DA) (green circle) versus inactive fractions (blue circle). (B) S-Loading plot showing the putatively active metabolites for anti-*Helicobacter* activity.

micelles with a mean diameter of about 65 nm, which is smaller than the results obtained by DLS (112 nm). The size difference can be explained by the fact that DLS measures the hydrodynamic diameter of micelles, whereas TEM measures the size of the dried particles.²³

4. Conclusion

Metabolic profiling of five *Mentha* species cultivated in Egypt was studied via LC-ESI-MS, whereas the discrimination was based on PCA analysis. Characterization of those phytoconstituents showed

Table 4 Characterizations of the prepared *M. viridis* polymeric micelles^{b,c}

Formula	PS (nm)	PDI	Zeta potential (mV)	Zone of inhibition (mm)	MIC (mg mL ⁻¹)	MBC (mg mL ⁻¹)
Pure extract (100 mg mL ⁻¹)	ND ^a	ND	ND	16 ± 1.5	25 ± 0.0	100 ± 0.0
MV-M**	114.9 ± 7.98	0.228 ± 0.006	-29.6 ± 0.31	13 ± 1.5	>100	ND
MV-R**	112.2 ± 1.72	0.37 ± 0.005	-31.0 ± 0.60	20 ± 2.0	6.25 ± 0.0	50 ± 0.0

^a ND: not determined. ^b $n = 3$, mean ± SD. ^c The results were obtained from three independent replicate experiments and expressed as means ± standard deviation. The statistical significance of the results was tested using one-way Analysis of Variance (ANOVA) and Turkey-Kramer multiple Comparisons Test between pure extract and the prepared formula and the tested formulae together. The p value significance was represented as an asterisk (*) for $p < 0.05$ and two asterisks (**) for $p < 0.01$.



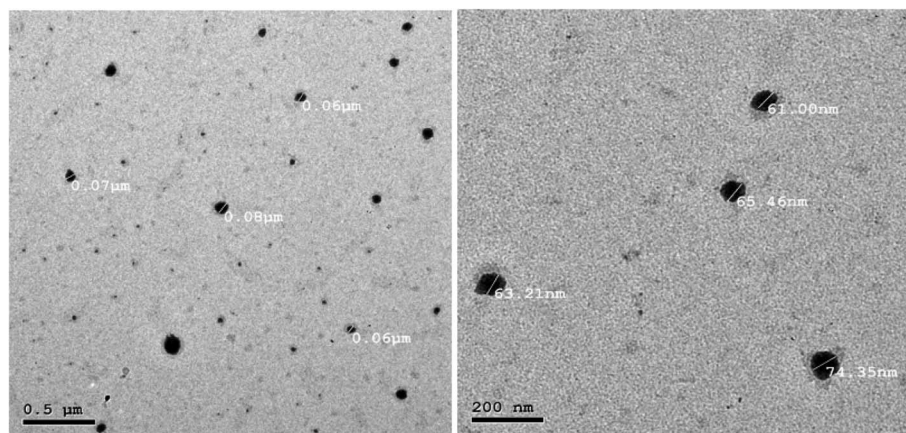


Fig. 4 Transmission electronic microscopy (TEM) images of the *M. viridis* polymeric micelles (MvP-R).

a diversity of compounds, including flavonoids, phenolic acids, and caffeic acid esters. Cytotoxic and anti-*Helicobacter* activities were assessed for the five species, where *M. piperita* and *M. viridis* exhibited the most prominent effects, respectively. Those activities were correlated with the chemical profile using OPLS-DA, where a lignin glycoside and a methoxylated flavone were responsible for those effects, respectively. Polymeric micelles were prepared for *M. viridis*, the most active anti-*Helicobacter* species, using two methodologies, namely rotary evaporator and microwave; similar sizes and zeta potentials were obtained, but higher antimicrobial activity was observed with the micelles prepared using a microwave. Pluronic® F127 polymeric micelles were found to present promise as nanocarrier for *Mentha* extract and to improve its antibacterial activity against *H. pylori* compared with the plain extract, and they could be used as adjuvant therapy for ulcer treatment.

Author contributions

Collection, drying and extraction of different *Mentha* species, in addition to tentative identification of the characterized compounds through LC/MS analysis, collection of data, writing and revising the final manuscript: R. O. B., M. A. A. F., A. M. E., M. A. A., H. A. E. Chemometric study, OPLS-DA and cytotoxic activity, conceptualization, methodology: U. R. A., N. T., A. T and M. F. Anti-*Helicobacter* activity (W. A. A.); nanoparticle formulation and characterization (S. M. F., S. M. S. E.); writing, reviewing, and editing, all authors.

Conflicts of interest

The authors declare there is no conflict of interest.

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References

- 1 B. Salehi, Z. Stojanović-Radić, J. Matejić, F. Sharopov, H. Antolak, D. Kręgiel, S. Sen, M. Sharifi-Rad, K. Acharya, R. Sharifi-Rad, M. Martorell, A. Sureda, N. Martins and J. Sharifi-Rad, *Plants*, 2018, **7**, 70.
- 2 A. O. Tucker and R. F. Naczi, in *Mint: the genus Mentha*, ed. B. M. Lawrence, CRC Press, 2007, pp. 1–40.
- 3 Z. Sun, H. Wang, J. Wang, L. Zhou and P. Yang, *PLoS One*, 2014, **9**(12), e114767.
- 4 M. Cirlini, P. Mena, M. Tassotti, K. A. Herrlinger, K. M. Nieman, C. Dall'Asta and D. Del Rio, *Molecules*, 2016, **21**, 15.
- 5 F. Anwar, A. Abbas, T. Mehmood, A. H. Gilani and N. ur Rehman, *Phytother. Res.*, 2019, 1–23.
- 6 A. Sato and H. Tamura, *Fitoterapia*, 2015, **102**, 74–83.
- 7 M. Mahboubi, *J. Tradit. Complement. Med.*, 2018, 1–7.
- 8 M. S. Ali-Shtayeh, R. M. Jamous, S. Y. Abu-Zaitoun, A. I. Khasati and S. R. Kalbouneh, *Evidence-Based Complementary Altern. Med.*, 2019, **2019**, 3834265.
- 9 F. Zaidi, B. Voirin, M. Jay and M. R. Viricel, *Phytochemistry*, 1998, **48**, 991–994.
- 10 S. Sumbul, M. A. Ahmad, M. Asif and M. Akhtar, *J. Pharm. BioAllied Sci.*, 2011, **3**, 361–367.
- 11 J. Teixeira, A. Gaspar, E. M. Garrido, J. Garrido and F. Borges, *BioMed Res. Int.*, 2013, **2013**, 251754.
- 12 M. A. Alam, N. Subhan, H. Hossain, M. Hossain, H. M. Reza, M. M. Rahman and M. O. Ullah, *Nutr. Metab.*, 2016, **13**, 27.
- 13 I. Cocan, E. Alexa, C. Danciu, I. Radulov, A. Galuscan, D. Obistioiu, A. A. Morvay, R. M. Sumalan, M. A. Poiana, G. Pop and C. A. Dehelean, *Exp. Ther. Med.*, 2018, **15**, 1863–1870.
- 14 M. Creydt and M. Fischer, *Anal. Chem.*, 2017, **89**, 10474–10486.
- 15 J. Hong, L. Yang, D. Zhang and J. Shi, *Int. J. Mol. Sci.*, 2016, **17**(6), 767.
- 16 D. O. Ochwang'i, C. N. Kimwele, J. A. Oduma, P. K. Gathumbi, J. M. Mbaria and S. G. Kiama, *J. Ethnopharmacol.*, 2014, **151**, 1040–1055.
- 17 M. Greenwell and P. K. S. M. Rahman, *Int. J. Pharma Sci. Res.*, 2015, **6**, 4103.
- 18 R. M. Peek and M. J. Blaser, *Nat. Rev. Cancer*, 2002, **2**, 28–37.
- 1 B. Salehi, Z. Stojanović-Radić, J. Matejić, F. Sharopov, H. Antolak, D. Kręgiel, S. Sen, M. Sharifi-Rad, K. Acharya,



- 19 S. Martini, C. D'Addario, A. Colacevich, S. Focardi, F. Borghini, A. Santucci, N. Figura and C. Rossi, *Int. J. Antimicrob. Agents*, 2009, **34**, 50–59.
- 20 A. T. B. Abadi and J. G. Kusters, *BMC Gastroenterol.*, 2016, **16**, 94–98.
- 21 Y. D. Taghipour, R. Bahramsoltani, A. M. Marques, R. Naseri, R. Rahimi, P. Haratipour, A. I. Panah, M. H. Farzaei and M. Abdollahi, *Daru, J. Pharm. Sci.*, 2018, **26**, 229–239.
- 22 A. R. Fares, A. N. Elmeshad and M. A. A. Kassem, *Drug Delivery*, 2018, **25**, 132–142.
- 23 A. S. Deshmukh, P. N. Chauhan, M. N. Noolvi, K. Chaturvedi, K. Ganguly, S. S. Shukla, M. N. Nadagouda and T. M. Aminabhavi, *Int. J. Pharm.*, 2017, **532**, 249–268.
- 24 C. Sun, W. Li, P. Ma, Y. Li, Y. Zhu, H. Zhang, M. Adu-Frimpong, W. Deng, J. Yu and X. Xu, *Food Chem. Toxicol.*, 2020, **137**, 111126.
- 25 M. U. Akbar, K. M. Zia, A. Nazir, J. Iqbal, S. A. Ejaz and M. S. H. Akash, *AAPS PharmSciTech*, 2018, **19**, 2719–2739.
- 26 M. Abedanzadeh, M. Salmanpour, F. Farjadian, S. Mohammadi and A. M. Tamaddon, *J. Drug Delivery Sci. Technol.*, 2020, **58**, 101793.
- 27 F. Molavi, M. Barzegar-jalali, H. Hamishehkar and H. Hamishehkar, *J. Control. Release.*, 2020, **320**, 265–282.
- 28 Y. Chen, W. Zhang, J. Gu, Q. Ren, Z. Fan, W. Zhong, X. Fang and X. Sha, *Int. J. Pharm.*, 2013, **452**, 421–433.
- 29 J. K. Valenzuela-Oses, M. C. García, V. A. Feitosa, J. A. Pachioni-Vasconcelos, S. M. Gomes-Filho, F. R. Lourenço, N. N. P. Cerize, D. S. Bassères and C. O. Rangel-Yagui, *Mater. Sci. Eng., C*, 2017, **81**, 327–333.
- 30 M. Jaiswal, M. Kumar and K. Pathak, *Colloids Surf., B*, 2015, **130**, 23–30.
- 31 J. Oyama, D. S. S. L. Lera-Nonose, Á. C. F. H. Ramos-Milare, F. B. Padilha Ferreira, C. F. de Freitas, W. Caetano, N. Hioka, T. G. V. Silveira and M. V. C. Lonardoní, *Acta Trop.*, 2019, **192**, 11–21.
- 32 M. Blumenthal, W. Busse, A. Goldberg, J. Gruenwald, T. Hall and C. Riggins, *The Complete German Commission E Monographs: Therapeutic Guide to Herbal Medicines | Annals of Internal Medicine | American College of Physicians*, American Botanical Council, 1999.
- 33 A. C. Martin, A. D. Pawlus, E. M. Jewett, D. L. Wyse, C. K. Angerhofer and A. D. Hegeman, *RSC Adv.*, 2014, **4**, 26325–26334.
- 34 C. Cheng, E. M. Othman, H. Stopper, R. A. Edrada-Ebel, U. Hentschel and U. R. Abdelmohsen, *Mar. Drugs*, 2017, **15**, 383–391.
- 35 A. W. Bauer, W. M. Kirby, J. C. Sherris and M. Turck, *Am. J. Clin. Pathol.*, 1966, **45**, 493–496.
- 36 J. A. Nazeam, W. A. AL-Shareef, M. W. Helmy and A. E. El-Haddad, *Food Chem.*, 2020, **326**, 126993.
- 37 T. A. Nagy, L. E. Wroblewski, D. Wang, M. B. Piazuelo, A. Delgado, J. Romero-Gallo, J. Noto, D. A. Israel, S. R. Ogden, P. Correa, T. L. Cover and R. M. Peek, *Gastroenterology*, 2011, **141**, 553–564.
- 38 A. El Haddad, W. AL-Shareef and S. Eid, *J. Adv. Pharm. Res.*, 2019, **3**, 143–149.
- 39 J. Patel, M. Weinstein, G. Eliopoulos, S. Jenkins, B. Limbago, A. Mathers, T. Mazzulli, R. Patel, S. Richter, M. Satlin, J. Swenson, M. Traczewski, J. Turnidge and B. Zimmer, *M100: Performance Standards for Antimicrobial Susceptibility Testing An informational supplement for global application developed through the Clinical and Laboratory Standards Institute consensus process*, 2017.
- 40 M. H. Taleb, N. F. Abdeltawab, R. N. Shamma, S. S. Abdelgayed, S. S. Mohamed, M. A. Farag and M. A. Ramadan, *Molecules*, 2018, **23**(9), 2164.
- 41 Q. Yao, Y. Liu, L. Kou, Y. Tu, X. Tang and L. Zhu, *Nanomedicine*, 2019, **19**, 71–80.
- 42 W. Zhang, Y. Shi, Y. Chen, J. Ye, X. Sha and X. Fang, *Biomaterials*, 2011, **32**, 2894–2906.
- 43 D. Jain, N. Pathak, S. Khan, G. V. Raghuram, A. Bhargava, R. Samarth and P. K. Mishra, *Int. J. Toxicol.*, 2011, **30**, 225–236.
- 44 Y. X. Li, Y. B. Liu, A. Q. Ma, Y. Bao, M. Wang and Z. L. Sun, *Food Sci. Biotechnol.*, 2017, **26**, 1675–1683.
- 45 M. O. Arruda, S. J. F. Mendes, S. A. Teixeira, L. S. S. De Mesquita, M. N. De Sousa Ribeiro, S. D. S. L. Galvão, M. N. Muscará, E. S. Fernandes and V. Monteiro-Neto, *J. Immunol. Res.*, 2017, **2017**, 2078794.
- 46 H. Imai, K. Osawa, H. Yasuda, H. Hamashima, T. Arai and M. Sasatsu, *Microbios*, 2001, **106**, 31–39.
- 47 A. Bodalska, A. Kowalczyk, M. Włodarczyk and I. Fecka, *Molecules*, 2020, **25**, 69–88.
- 48 Y. J. Park, S. A. Baek, Y. Choi, J. K. Kim and S. U. Park, *Molecules*, 2019, **24**, 258.
- 49 H. J. D. Dorman, M. Koşar, K. H. C. Başer and R. Hiltunen, *Nat. Prod. Commun.*, 2009, **4**, 535–542.
- 50 J. C. Yang, H. C. Yang, C. T. Shun, T. H. Wang, C. T. Chien and J. Y. Kao, *Evidence-Based Complementary Altern. Med.*, 2013, **2013**, 248585.
- 51 P. Fong, C. H. Hao, C. C. Io, P. I. Sin and L. R. Meng, *Molecules*, 2019, **24**(19), 3608.
- 52 L. L. Périco, M. T. Emílio-Silva, R. Ohara, V. P. Rodrigues, G. Bueno, J. M. Barbosa-Filho, L. R. M. da Rocha, L. M. Batista and C. A. Hiruma-Lima, *Biomolecules*, 2020, **10**, 265.
- 53 M. S. Kamel, K. Ohtani, M. H. Assaf, R. Kasai, M. A. El-Shanawani, K. Yamasaki, A. A. Ali and O. Tanaka, *Phytochemistry*, 1992, **31**, 2469–2471.
- 54 D. B. Rodriguez-Amaya, *Food Carotenoids*, John Wiley & Sons, Ltd, Chichester, UK, 2015.
- 55 A. R. Khuda-Bukhsh, S. K. Saha and S. Das, in *Cancer: Oxidative Stress and Dietary Antioxidants*, Elsevier Inc., 2014, pp. 77–89.
- 56 V. Pasquet, P. Morisset, S. Ihammouine, A. Chepied, L. Aumailley, J. B. Berard, B. Serive, R. Kaas, I. Lanneluc, V. Thiery, M. Lafferriere, J. M. Piot, T. Patrice, J. P. Cadoret and L. Picot, *Mar. Drugs*, 2011, **9**, 819–831.
- 57 A. Tawfike, E. Z. Attia, S. Y. Desoukey, D. Hajjar, A. A. Makki, P. J. Schupp, R. A. Edrada-Ebel and U. R. Abdelmohsen, *AMB Express*, 2019, **9**, 12.
- 58 A. Tawfike, M. Romli, C. Clements, G. Abbott, L. Young, M. Schumacher, M. Diederich, M. Farag and R. A. Edrada-



- Ebel, J. *Chromatogr. B: Anal. Technol. Biomed. Life Sci.*, 2019, **1106**, 71–73.
- 59 A. Tawfike, R. Tate, G. Abbott, L. Young, C. Viegmann, M. Schumacher, M. Diederich and R. Edrada-Ebel, *Chem. Biodiversity*, 2017, **14**, e1700040.
- 60 V. G. Nebieridze, A. V. Skhirtladze, E. P. Kemertelidze and M. Ganzera, *Chem. Nat. Compd.*, 2018, **54**, 63–65.
- 61 J. Shitamoto, S. Sugimoto, K. Matsunami, H. Otsuka, T. Shinzato and Y. Takeda, *Chem. Pharm. Bull.*, 2011, **59**, 72–77.
- 62 R. A. S. Macahig, K. Matsunami and H. Otsuka, *Chem. Pharm. Bull.*, 2011, **59**, 397–401.
- 63 T. Isobe, M. Doe, Y. Morimoto, K. Nagata and A. Ohsaki, *Biol. Pharm. Bull.*, 2006, **29**, 1039–1041.
- 64 T. Wong, *Curr. Drug Delivery*, 2008, **5**, 77–84.
- 65 M. Alle, B. reddy G, T. H. Kim, S. H. Park, S. H. Lee and J. C. Kim, *Carbohydr. Polym.*, 2020, **229**, 115511.
- 66 H. Ashraf, T. Anjum, S. Riaz and S. Naseem, *Front. Microbiol.*, 2020, **11**, 238.
- 67 J. Isaac, S. Kaity, S. Ganguly and A. Ghosh, *J. Pharm. Pharmacol.*, 2013, **65**, 219–229.
- 68 V. D. Wagh and O. J. Deshmukh, *ISRN Pharm.*, 2012, **2012**, 1–7.
- 69 E. Haba, S. Bouhdid, N. Torregio-Solana, A. M. Marqués, M. J. Espuny, M. J. García-Celma and A. Manresa, *Int. J. Pharm.*, 2014, **476**, 134–141.
- 70 J. Sun, F. Liang, Y. Bin, P. Li and C. Duan, *Molecules*, 2007, **12**, 679–693.
- 71 K. Kapp, E. Hakala, A. Orav, L. Pohjala, P. Vuorela, T. Püssa, H. Vuorela and A. Raal, *Food Res. Int.*, 2013, **53**, 758–766.
- 72 R. Kodama, T. Yano, K. Furukawa, K. Noda and H. Ide, *Xenobiotica*, 1976, **6**, 377–389.
- 73 M. B. Bahadori, G. Zengin, S. Bahadori, L. Dinparast and N. Movahhedine, *Int. J. Food Prop.*, 2018, **21**, 183–193.
- 74 D. Lu, X. Yuan, S.-J. Kim, J. V. Marques, P. P. Chakravarthy, S. G. A. Moinuddin, R. Luchterhand, B. Herman, L. B. Davin and N. G. Lewis, *Plant Biotechnol. J.*, 2017, **15**, 970–981.
- 75 M. J. Magera, A. L. Stoor, J. K. Helgeson, D. Matern and P. Rinaldo, *Clin. Chim. Acta*, 2001, **306**, 35–41.
- 76 B. Fatih, K. Madani, M. Chibane and P. Duez, in *Aromatic and Medicinal Plants - Back to Nature*, InTech, 2017.
- 77 P. Mena, M. Cirilini, M. Tassotti, K. A. Herrlinger, C. Dall'Asta and D. Del Rio, *Molecules*, 2016, **21**(11), 1576.
- 78 A. Taamalli, D. Arráez-Román, L. Abaza, I. Iswaldi, A. Fernández-Gutiérrez, M. Zarrouk and A. Segura-Carretero, *Phytochem. Anal.*, 2015, **26**, 320–330.
- 79 M. Božovic, A. Piroli and R. Ragno, *Molecules*, 2015, **20**, 8605–8633.
- 80 Z. Özer, *J. Turk. Chem. Soc., Sect. A*, 2018, **5**, 445–456.
- 81 D. P. De Sousa, E. Raphael and T. J. Brocksom, *Z. Naturforsch., B: J. Chem. Sci.*, 2010, **65**, 1381–1383.
- 82 F. A. Tomás-Barberán, S. Z. Husain and M. I. Gil, *Biochem. Syst. Ecol.*, 1988, **16**, 43–46.
- 83 G. H. Jang, H. W. Kim, M. K. Lee, S. Y. Jeong, A. R. Bak, D. J. Lee and J. B. Kim, *Saudi J. Biol. Sci.*, 2018, **25**, 1622–1631.
- 84 N. R. Desam, A. J. Al-Rajab, M. Sharma, M. M. Mylabathula, R. R. Gowkanapalli and M. Albratty, *J. King Saud Univ., Sci.*, 2019, **31**, 528–533.

