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# Superior anti-infective potential of eugenol–casein nanoparticles combined with polyethylene glycol against *Colletotrichum musae* infections

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The aim of this study was to improve the stability of eugenol–casein nanoparticles (EL–CS-NPs) through polyethylene glycol (PEG) modification. The results show that modifying the EL–CS-NPs with PEG after loading with eugenol (EL) gives PEG–EL–CS-NPs, with increased stability. The NPs modified with higher-molecular-weight PEG showed better stability. A CS/PEG ratio of 200 : 1 (w/w) yielded the NPs with the best stability. A PEG<sub>20 K</sub>–EL–CS-NP dispersion remained stable in cold storage for over one year, and also exhibited stronger inhibitory effects against *Colletotrichum musae* inoculated on bananas than an EL–CS-NP dispersion, since it showed more prolonged sustained release of EL than the EL–CS-NP dispersion. Lyophilized PEG<sub>20 K</sub>–EL–CS-NP powder showed better effectiveness against mold on bread than lyophilized EL–CS-NPs powder. Using PEG to modify CS-NPs shows potential for improving the stability of CS-NPs loaded with hydrophobic substances for delivery in the fields of food and agriculture.

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## 1. Introduction

The transportation of hydrophobic compounds through hydrophilic food matrices remains a challenge, particularly in the case of bioactive molecules, such as antioxidants, vitamins, fatty acids, and probiotics, which are susceptible to changes in pH and ionic strength, and to oxidation. Nanostructured carriers are considered promising vehicles for delivering hydrophobic substances, allowing for the transportation of hydrophobic compounds while remaining stable in the food matrix to ensure safe, functional, and targeted delivery.<sup>1–3</sup> The encapsulation of sensitive ingredients into nanostructures increases their stability and maintains their bioavailability, protecting them against external degradation factors and ensuring controlled release.<sup>4,5</sup>

Protein-based food materials have been shown to be safe and they are considered appropriate for delivering bioactive compounds and maintaining their functionality in the body. Caseins have been proven to be one of the most reliable nanovehicles in terms of cost, nutrition, and safety.<sup>6–9</sup> Curcumin has been encapsulated in casein micelles to form nanoparticles with an average size distribution of <200 nm, with the potential to provide an oral dose of curcumin.<sup>10</sup> Docosahexaenoic acid (DHA) entrapped in casein micelles has been found to be protected against oxidation, and the micelles have good colloidal stability when stored at 4 °C.<sup>11</sup> The encapsulation of vitamin D<sub>3</sub>

in casein micelles has been found to stabilize vitamin D<sub>3</sub> in aqueous systems, improve the accuracy and uniformity of distribution, protect it during thermal treatment (80 °C, 1 min) and 28 days of cold storage,<sup>12</sup> shield it from harmful UV radiation damage,<sup>13</sup> prevent its degradation in the stomach,<sup>14</sup> and improve its bioavailability.<sup>15,16</sup> Casein nanoparticles formed from re-assembled casein micelles have been used as nanovehicles for β-carotene, enabling water-insoluble β-carotene to be applied during different processing technologies in the food industry; this is because encapsulating β-carotene reduces its exposure to peroxy radical oxidation and improves its bioaccessibility.<sup>17,18</sup> Thymol encapsulated in sodium caseinate is more effective than unencapsulated thymol in inhibiting food-borne pathogens in milk owing to its enhanced distribution and solubility.<sup>19</sup> Spray-dried zein/casein nanoparticles in which thymol and eugenol are co-encapsulated can be easily rehydrated to produce a stable dispersion. The encapsulated essential oils show controlled release over 24 h, exhibiting enhanced bactericidal and bacteriostatic effects in milk whey against *E. coli* O157:H7 and *L. monocytogenes*, respectively.<sup>20</sup> However, based on existing studies, obtaining stable target substance-loaded nanoparticles is critical for applying casein nanoparticles as nanocarriers for functional substances.<sup>21</sup> More fundamental research is therefore required to fully understand and control the assembly and disassembly mechanisms of structures and the stability of nanoparticles during processing and storage.<sup>22</sup>

Eugenol (4-allyl-2-methoxyphenol), a major constituent of clove essential oil, has been found to inhibit various pathogens. Eugenol (EL) has been considered as an ideal antibacterial agent in the food industry since it is classified as “generally

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regarded as safe".<sup>23,24</sup> However, the efficiency of EL is substantially limited due to its high volatility, low water solubility, and high susceptibility to environmental stress.<sup>25</sup> Embedding and encapsulation technologies have been used for improving the physicochemical stabilities of volatile active agents.<sup>26</sup> Recently, several studies have also revealed that the encapsulation of EL into nanoparticles can enhance its stability, permeability, and antimicrobial activity.<sup>27–29</sup>

Polyethylene glycol is a chain molecule that can be synthesized with different chain lengths depending on requirements. The long chain of polyethylene glycol may bind to nanoparticles with polar surfaces, increasing their stability. In the present study, we exhibit the effects of polyethylene glycol on the stability of eugenol–casein nanoparticles, which exhibit an improvement in their effectiveness at inhibiting pathogenic microorganisms in bananas and bread.

## 2. Materials and methods

### 2.1. Chemicals and reagents

Casein sodium (C8654-1KG) and polyethylene glycol (PEG, HO(CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>H) with average molecular weights of 2000 (2 K), 4000 (4 K), 8000 (8 K), 20 000 (20 K), and 35 000 (35 K) Da were purchased from Sigma-Aldrich (Shanghai, China). Eugenol (C<sub>10</sub>H<sub>12</sub>O<sub>2</sub>), high-performance liquid chromatography (HPLC)-grade acetonitrile and methanol, analytical-grade ethyl acetate, formic acid, NaCl, and MgCl<sub>2</sub> were purchased from Shanghai Yuanye Biotechnology Co., Ltd (Shanghai, China).

The banana pathogen *Colletotrichum musae* used in this study has been reported in previous work,<sup>29</sup> and it was used as the indicator microorganism for antifungal assays. Stock cultures were then maintained on potato dextrose agar (PDA) slants at 25 ± 2 °C for further study. The fungi were transferred to fresh PDA and incubated at room temperature before any experiments were carried out.

### 2.2. Preparation of PEG–EL–CS–NP dispersions

PEG–EL–CS–NP dispersions were prepared using casein sodium at a concentration of 20 mg mL<sup>-1</sup>. EL was pre-dissolved in absolute ethanol at a concentration of 100 mg mL<sup>-1</sup>. A known amount of the resulting solution was then added dropwise to the protein micellar dispersion under stirring at 25 °C. Following the addition of the EL solution, the dispersion was ultrasonicated for 3 min in an ice bath, then a given amount of aqueous PEG solution at a concentration of 20 mg mL<sup>-1</sup> was added to the dispersion, and ultrasonication in the ice bath was continued for a further 3 min. The dispersion was filtered through a 0.46 μm film to yield the PEG–EL–CS–NP dispersion.

### 2.3. Effects of CS–NP PEG modification on the release of loaded EL

The % entrapment efficiency (EE) of EL in PEG–EL–CS–NPs exposed to heat (37 °C) was determined to evaluate the effectiveness of PEG in controlling EL release from casein NPs, since EL remained in the EL–CS–NPs at 37 °C for approximately 1 day.

### 2.4. Nanoparticle physicochemical characterization

The mean particle size, size distribution, and zeta potential of the nanoparticles were measured using a Zetasizer Nano ZSE instrument (Malvern Instruments, Malvern, U.K.), employing a nominal 5 mW He–Ne laser operated at a wavelength of 633 nm and a scattering angle of 173°, as described in a previous report.<sup>30</sup> For characterization, 1 mL of the NP dispersion was diluted with 10 mL of water at room temperature. All data are expressed as mean values of three independent batches of samples. Transmission electron microscopy (TEM) was used for the morphological characterization of PEG–EL–CS–NPs using a process described previously. Samples were prepared *via* dilution, and they were then placed on a film-coated 200-mesh copper specimen grid for 10 min, stained with 3% phosphotungstic acid and dried prior to TEM observations using a Hitachi H-9500E electron microscope.

### 2.5. Entrapment efficiency (EE) of EL in NPs

The amount of EL remaining in the NPs was determined using a previously reported procedure.<sup>31</sup> Briefly, a NP dispersion was centrifuged at 10 000g to separate the free EL. Then, the clear aqueous phase (50 μL) was added to 450 μL of methanol to denature the protein. After removing the protein through filtering, the methanol solution was then subjected to high-performance liquid chromatography (HPLC) analysis for EL. HPLC analysis was performed using a Shimadzu HPLC system (Shimadzu, Kyoto, Japan) with two LC-10A pumps, an SIL-10Avp autosampler, an SPD-M10Avp UV detector, and a Symmetry C18 (5 m, 4.6 mm × 250 mm) column. The column was eluted using a mobile phase composed of water–methanol (30 : 70) at a flow rate of 1 mL min<sup>-1</sup> at 30 °C. The chromatograms were recorded *via* UV absorbance at 280 nm. The EE (%) of EL in the NPs was calculated from the results of HPLC analysis using the equation:

$$\text{EE (\%)} = \left( \frac{\text{EL concentration of the clear aqueous phase}}{\text{concentration of the dispersion}} \right) \times 100$$

### 2.6. Evaluation of the storage stability of the NP dispersion

To confirm the stability of PEG<sub>20 K</sub>–EL–CS–NP during storage, NP dispersions were kept in sealed amber-colored glass vials in a cold environment (4–10 °C). The particle size, polydispersity index (PDI), zeta potential (ZP), and EE of the dispersions were then measured at predetermined time intervals and compared with the values obtained from fresh dispersions.

### 2.7. The lyophilization of NP dispersions and calculations of the remaining EL

The PEG<sub>20 K</sub>–EL–CS–NP and EL–CS–NP dispersions were pre-frozen to –80 °C and then lyophilized using a lyophilizer with a cold trap at –80 °C (Beijing Biocool Experimental Instrument Co., Ltd, China). Some of the powder was subjected to EL extraction with methanol. The methanol solution was injected into the HPLC system for EL determination. The amount of EL



remaining in the lyophilized powder was calculated based on the EL concentration of the methanol solution.

## 2.8. Determination of EL release from lyophilized NP powder

EL release from NP powder samples at room temperature (20–25 °C) was determined *via* the following procedure. Mini bags (0.5 cm × 1 cm) containing lyophilized PEG<sub>20 K</sub>-EL-CS-NP or EL-CS-NP powder (5 mg) were placed in empty bread packages without being completely sealed. After a given number of days of treatment, the remaining EL in the lyophilized powder was extracted using methanol for HPLC analysis. The release of EL was calculated based on the remaining EL in the lyophilized powder.

## 2.9. Determination of the inhibitory effects of NP dispersions on *C. musae* on banana

The mycelium strain *Colletotrichum musae* was pre-cultured on Luria-Bertani (LB, g L<sup>-1</sup>; 10 g of tryptone, 5 g of yeast extract, and 10 g of NaCl) agar media at 28 °C for 7 d as a seed culture. A spore suspension was prepared *via* washing 20 day-old seed culture with sterile water containing 0.01% (v/v) Tween-80 and then diluting this to 1 × 10<sup>5</sup> spores per mL with the aid of a hemocytometer. The minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) values of the prepared nanoparticles were investigated *via* turbidimetric and micro-dilution methods. Seed cultures were inoculated with 5% (v/v) inoculum in fresh LB media with final eugenol amounts of 0.05–2.50 mg mL<sup>-1</sup> from eugenol, EL-CS-NPs, and PEG<sub>20 K</sub>-EL-CS-NPs, respectively. Additionally, dilution separation methods were used to examine the presence of bacteria in the original MIC tubes *via* transferring 50 μL of solution from the non-turbid tubes to the appropriate broth medium and permitting growth after overnight incubation.<sup>32</sup> The minimum

concentration in a test tube with no *C. musae* cell growth gave the MFC value.

The inhibitory activity of PEG-EL-CS-NP dispersions toward *C. musae* on bananas was determined using a previously reported procedure.<sup>29</sup> Briefly, 10 μL of *C. musae* spore suspension was injected into a site under the peel of one half of a banana, and pure water was injected into a site under the peel on the other half as a control. The treated bananas were coated with the NP dispersion at an EL concentration of 0.1 g L<sup>-1</sup> *via* brushing. After treatment, all bananas were stored at 25 °C in a breathable plastic basket. The lesions were observed and recorded after peeling at intervals of 2 days. Each treatment had three replicates, and the entire experiment was repeated three times.

## 2.10. Determination of the effectiveness of the lyophilized powder against mold growth on bread

The preparation of bread samples (without preservative) was carried out according to the procedure described by Gao *et al.*<sup>33</sup> Briefly, bread dough was prepared *via* mixing ingredients in a dough mixer for 1 min at 45 rpm and 5 min at 100 rpm. After resting for 15 min, the bread dough was divided into 25 g pieces, which were rounded and proofed at 40 °C and 85% RH for 70 min. After proofing, the dough pieces were baked at 200 °C for 10 min, and then the bread was cooled down at ambient temperature for 1 h. Mini bags (0.5 cm × 1 cm) containing lyophilized PEG<sub>20 K</sub>-EL-CS-NPs, EL-CS-NPs, or CS-NPs (control) powder (1 mg) were placed under the bread, which was packaged in a hemispherical plastic package with an incomplete seal. A free-EL sample was prepared *via* dissolving EL in acetone, dropping this solution onto a piece of filter paper, and then removing the acetone *via* spontaneous evaporation. Bread was placed in packages without fungal inoculation in order to determine the effects of the lyophilized powder against naturally occurring mold in bread. The packed bread

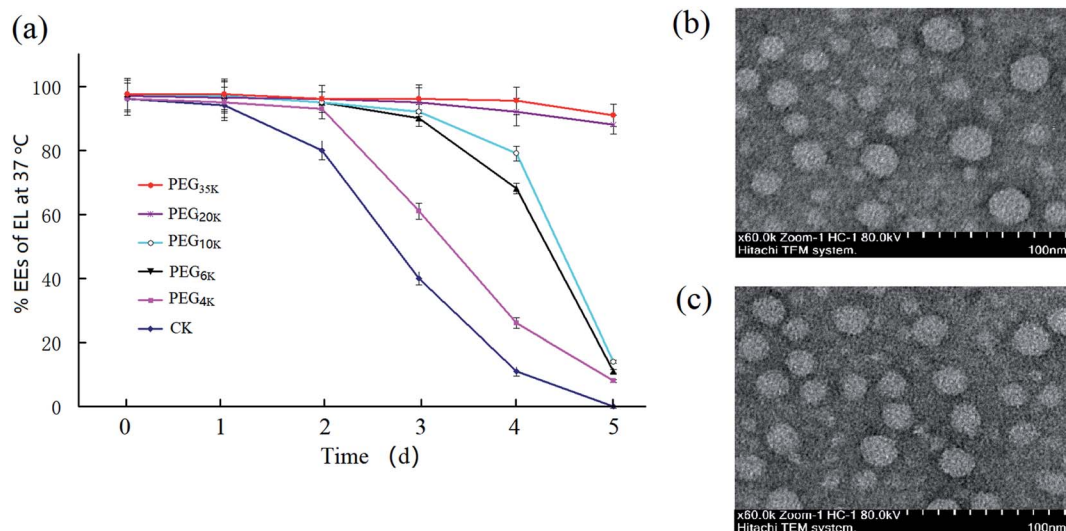


Fig. 1 (a) The EEs (%) of EL in PEG-EL-CS-NPs with PEG of different molecular weights at 37 °C during a period of 5 days (a), and TEM images of (b) PEG<sub>35 K</sub>-EL-CS-NPs and (c) EL-CS-NPs (CK: EL-CS-NPs).



**Table 1** Diameters of NPs loaded with different amounts of eugenol (EL)<sup>a</sup>

Composition of NPs	Diameter (nm) at different EL : CS ratios		
	1 : 10	1.5 : 10	3 : 10
EL-CS-NPs	180.4 ± 2.7 <sup>a</sup>	142.9 ± 1.6 <sup>a</sup>	117.4 ± 1.8 <sup>a</sup>
PEG <sub>35 K</sub> -EL-EL-NPs	162.8 ± 2.1 <sup>b</sup>	125.8 ± 1.4 <sup>b</sup>	96.4 ± 1.1 <sup>b</sup>

<sup>a</sup> Values in a column followed by different letters are significantly different ( $p < 0.05$ ).

samples were stored at  $25 \pm 2$  °C and 75% RH until mold was observed on the last group. The experiments were done in triplicate.

### 2.11. Statistical analysis

All samples were prepared and analyzed in triplicate. The data are presented as mean ± standard-deviation values from three determinations. Statistical differences between two groups were evaluated using Student's *t*-test. Multiple comparisons of means were done *via* LSD (least significant difference) testing. A probability value of <0.05 was considered significant. All computations were carried out using the SAS system for Windows V8.

## 3. Results and discussion

### 3.1. Effects of PEG molecular weight on PEG-EL-CS-NPs stability

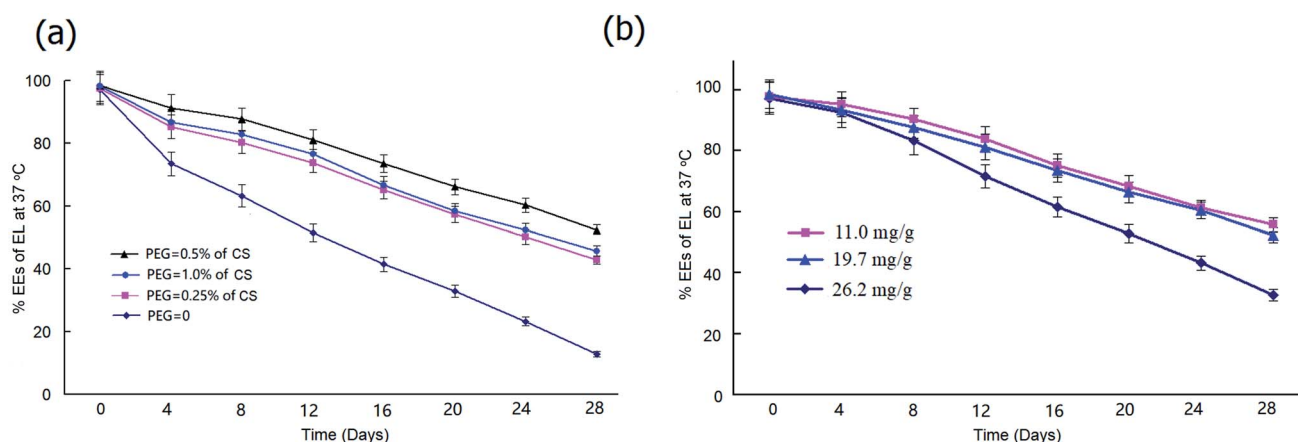
The EL-CS-NPs are only stable for one day at 37 °C, which limits the use of these NPs for EL delivery in food-related processes. To address this limitation, we modified EL-CS-NPs with different types of PEG. According to pre-experiments, an EL : casein-sodium (20 mg mL<sup>-1</sup>) ratio of 1 : 10 yielded EL-CS-NPs with an ideal EE (>90%). Thus, we used PEG samples with different molecular weights (MWs) to modify the NPs to improve their stability. The results showed that when PEG was used, the obtained PEG-EL-CS-NPs was more stable at 37 °C than EL-CS-

NPs (Fig. 1a). The clear increase in stability upon increasing the PEG MW from 4 K to 20 K Da indicates that the molecular chain length of PEG plays a key role in stabilizing the NPs, which implies that PEG can fill spaces on the surfaces of the NPs. The PEG<sub>35 K</sub>-EL-CS-NPs exhibit a slight increase in stability over PEG<sub>20 K</sub>-EL-CS-NPs. Interestingly, when the loading amount of EL was increased, the diameter of PEG<sub>35 K</sub>-EL-CS-NPs decreased (Table 1). Therefore, we suggest that EL displaced water from inside the NPs to yield a more compact structure, and PEG filled the spaces on the surfaces of the NPs as a result of interactions between PEG and the polar residues on the surfaces of the NPs (Fig. 1b and c). PEG encapsulation effectively blocked the leakage of eugenol from NP holes,<sup>12</sup> thus effectively improving the slow-release efficiency of eugenol nanoparticles and enhancing their antibacterial efficiency.<sup>29</sup>

To improve the stabilities of casein-nanoparticle-loaded substances and the entrapment efficiencies, dextran, pectin, and chitosan have been used for co-assembly with caseins. A casein-graft-dextran copolymer was used for β-carotene encapsulation, resulting in stable β-carotene nanoparticles, although the encapsulation efficiency was relatively low.<sup>19</sup> Casein was conjugated with maltodextrin and co-assembled with hydrophobic nutraceuticals to create nanovehicles for the enrichment of clear beverages.<sup>14</sup> Moreover, nanocomplexes were prepared with sodium caseinate/pectin *via* acidification, using glucono-δ-lactone and thermal treatment to yield stable compact spherical nanocomplexes, in which pectin delayed the hydrolysis of casein by pepsin under gastric conditions for the controlled release of rutin.<sup>34</sup> Therefore, we think that PEG<sub>20 K</sub> is an excellent material for enhancing the stability of casein-nanoparticles since PEG<sub>20 K</sub> possesses the properties of being food-safe and inexpensive.

### 3.2. Effects of the amount of PEG used and the EL-loading level on the stability of PEG-EL-CS-NPs

To determine the most effective stabilization approach, we prepared PEG<sub>20 K</sub>-EL-CS-NPs with different amounts of PEG<sub>20 K</sub>. Stability tests showed that particles containing PEG<sub>20 K</sub> at a level that was 0.5% of the amount of CS exhibited better NP



**Fig. 2** The effects of the % PEG composition (a) and EL loading amounts (b) on NPs stability.



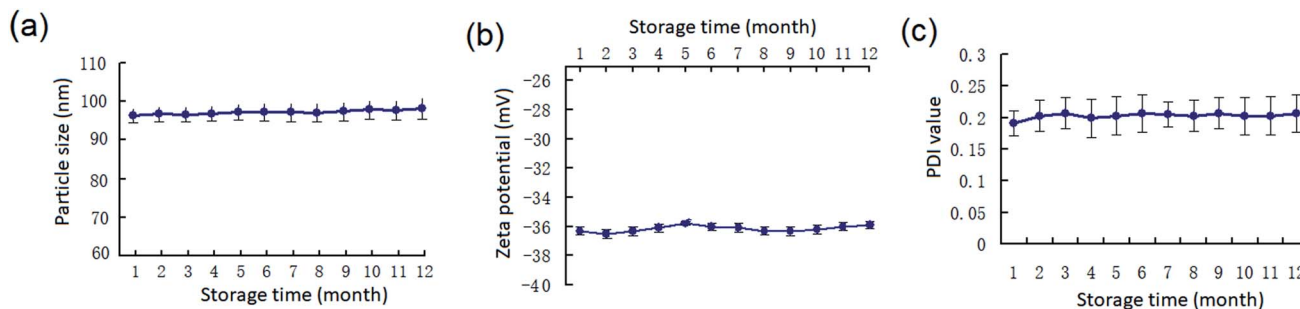


Fig. 3 (a) Changes in the particle size, (b) zeta potential, and (c) PDI values of the PEG<sub>20K</sub>-EL-CS-NPs during cold storage (4–10 °C).

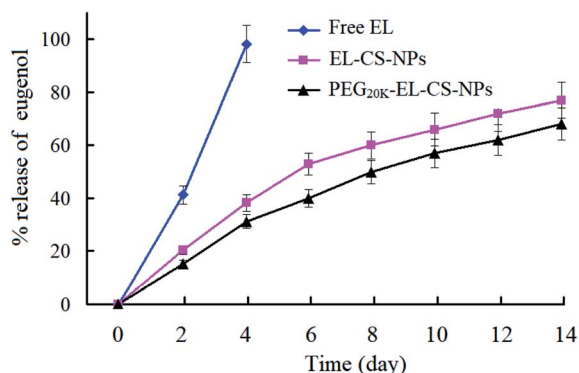


Fig. 4 EL release from the lyophilized NP powder samples compared with free EL from filter paper.

stability than those containing PEG<sub>20K</sub> at levels that were 0.25% and 1.0% of the amount of CS (Fig. 2a), although particles containing 1.0% PEG<sub>20K</sub> were more stable than those containing 0.25%. Under heating at 37 °C, PEG<sub>20K</sub>-EL-CS-NPs yielded long-term sustained release.

We also determined the stabilities of NPs with different EL-loading amounts. The results showed that high EL loading led to a significant reduction in the stability of the NPs. As shown in Fig. 2b, the stability of PEG<sub>20K</sub>-EL-CS-NPs with a loading amount of 26.2 mg g<sup>-1</sup> sharply decreased compared to samples with loading amounts of 14.9 mg g<sup>-1</sup> and 19.7 mg g<sup>-1</sup>.

### 3.3. Stability of PEG-EL-CS-NPs dispersions during cold storage (4–10 °C)

The long-term stability of PEG<sub>20K</sub>-EL-CS-NPs dispersions in cold storage (4–10 °C), which is critical for their eventual

Table 2 Minimal inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) values for eugenol, EL-CS-NPs, and PEG<sub>20K</sub>-EL-CS-NPs against *C. musae*<sup>a</sup>

	MIC (mg mL <sup>-1</sup> )	MFC (mg mL <sup>-1</sup> )
Eugenol (EL)	1.35 ± 0.11 <sup>a</sup>	1.85 ± 0.15 <sup>a</sup>
EL-CS-NPs	1.05 ± 0.09 <sup>b</sup>	1.30 ± 0.12 <sup>b</sup>
PEG <sub>20K</sub> -EL-CS-NPs	0.85 ± 0.07 <sup>c</sup>	0.95 ± 0.09 <sup>c</sup>

<sup>a</sup> Values in a column followed by different letters are significantly different ( $p < 0.05$ ).

application, was determined. The results demonstrate that PEG<sub>20K</sub>-EL-CS-NPs was stable for more than 12 months based on changes in the particle size, zeta potential, and PDI values of the NPs (Fig. 3). Therefore, the PEG-EL-CS-NPs dispersion could be used to deliver EL to prevent the fungal decay of fresh fruits and vegetables.

### 3.4. Remaining EL in and EL-release from lyophilized NP powder

The amounts of EL remaining in EL-CS-NPs lyophilized powder and PEG<sub>20K</sub>-EL-CS-NPs lyophilized powder were 63.1 ± 4.2% and 86.7 ± 6.3%, respectively, compared with the initial usage of EL for the preparation of the NP dispersions. It is clear that PEG<sub>20K</sub> significantly prevented the volatilization of EL during the preparation and lyophilization processes. The release levels of EL from PEG<sub>20K</sub>-EL-CS-NPs lyophilized powder and EL-CS-NPs lyophilized powder were determined and compared with the release of free EL from filter paper. The results showed that more than 98% of the free EL volatilized after 4 days while the EP lyophilized powder samples exhibited the sustained release of EL (Fig. 4).

### 3.5. Enhanced inhibitory effects of a PEG<sub>20K</sub>-EL-CS-NP dispersion against *C. musae* on bananas

The inhibitory activities of the EL-CS-NPs and PEG<sub>20K</sub>-EL-CS-NPs against *C. musae* were tested. Antibacterial analysis showed that the MIC and MFC values of PEG<sub>20K</sub>-EL-CS-NPs against *C. musae* were distinctly lower than those of eugenol casein nanoparticles without PEG assistance (Table 2), indicating that stronger antibacterial activity arose from the PEG coating of nanoparticles. *C. musae* on bananas causes decay. Two days after the bananas were inoculated with *C. musae*, the control group showed decay at the site of inoculation. However, the decay of the groups treated with the EL-CS-NP and PEG<sub>20K</sub>-EL-CS-NP dispersions was delayed until the sixth and eighth days, respectively (Fig. 5). The results demonstrate that the PEG<sub>20K</sub>-EL-CS-NP dispersion yielded enhanced inhibitory activity against *C. musae* compared with the EL-CS-NP formulation. In the above experiments, we demonstrated that PEG<sub>20K</sub>-EL-CS-NPs exhibited the better sustained release of EL than the EL-CS-NPs since the PEG<sub>20K</sub>-EL-CS-NP are more stable than the EL-CS-NPs. However, the release of EL from the dispersion was



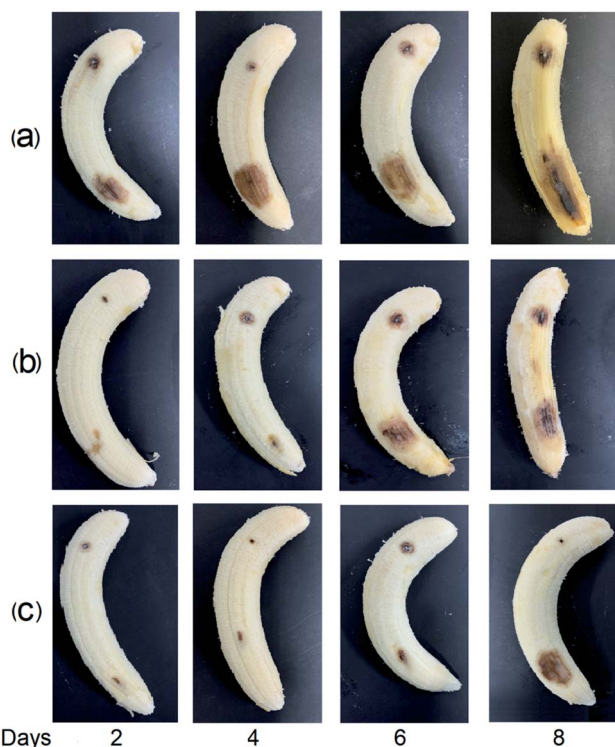


Fig. 5 The inhibition of fungal colony growth on bananas inoculated with *C. musae*: (a) treated with water, (b) treated with an EL-CS-NPs dispersion, and (c) treated with a PEG<sub>20K</sub>-EL-CS-NPs dispersion. The lower site of each banana was inoculated with *C. musae*, while the upper site was treated with water as a control.

very slow (Fig. 2a), and this was not synchronous with the anti-fungal effects on the banana. This result indicates that the stability of the NPs decreased due to the evaporation of water from the dispersions brushed onto the banana, which

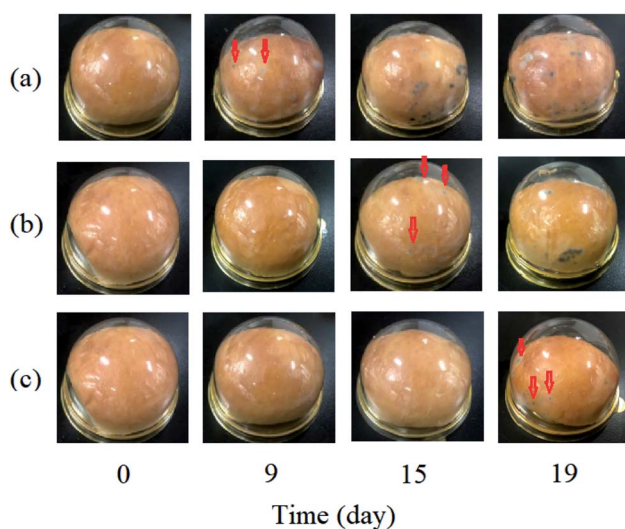


Fig. 6 The inhibition of mold growth using lyophilized NP powder compared with free EL (EL on filter paper): (a) free EL, (b) EL-CS-NP lyophilized powder, and (c) PEG<sub>20K</sub>-EL-CS-NP lyophilized powder. The red arrows indicate the sites of mold growth.

accelerated the release of EL. Nevertheless, the PEG<sub>20K</sub>-EL-CS-NPs exhibited more effective release than the EL-CS-NPs. Pathogenic microorganisms play a significant role in affecting the quality of food products; in particular, they can lead to food poisoning and the production of other mycotoxins, with associated health risks. The eco-friendly encapsulation system showed obvious controlled release behavior, thereby extending the duration and extent of the microbial inhibition effect. Thus, the PEG<sub>20K</sub>-EL-CS-NPs may have promising potential for applications relating to the control of anthracnose in food storage.

### 3.6. The enhanced inhibitory effects of PEG<sub>20K</sub>-EL-CS-NPs lyophilized powder against mold on bread

The anti-mold effectiveness of the PEG<sub>20K</sub>-EL-CS-NP lyophilized powder was evaluated and compared with free EL (control) and EL-CS-NP lyophilized powder through being used for the packaging of bread. Visually observed shelf-lives and mold-counts on bread during storage are shown in Fig. 6. Mold growth on bread packed with EL on filter paper, EL-CS-NP lyophilized powder, and PEG<sub>20K</sub>-EL-CS-NP lyophilized powder was visually observed after 9, 15, and 19 days of storage, respectively. The results indicated that the PEG<sub>20K</sub>-EL-CS-NPs effectively prolonged the shelf-life of bread, compared with free EL and the EL-CS-NPs. Previous studies have demonstrated that lyophilized AgCl nanoparticles remain as intact spheres with an outer covering of poly(vinyl alcohol), which preserves their spatial integrity.<sup>35</sup> Therefore, that the lyophilized PEG<sub>20K</sub>-EL-CS-NP powder in this study maintained its antibacterial activity can probably be attributed to the PEG coating on the nanoparticles.

## 4. Conclusions

In conclusion, this study was aimed at improving the stability and antimicrobial efficiency of eugenol-casein nanoparticles via PEG modification. The formulated eugenol-entrapping casein nanoparticles modified with PEG of various molecular weights showed that the PEG molecular weight was positively correlated with the stability of PEG-EL-CS-NPs. The PEG<sub>20K</sub>-EL-CS-NP dispersion obtained using PEG with a MW of 20 000 Da exhibited remarkable stability for over one year at low temperature, and it possesses a satisfactory sustained-release gradient, enhancing the inhibitory effects against *C. musae* on bananas and mold on bread. This work shows potential for improving the stability of casein NPs loaded with hydrophobic substances for delivery in the food and agriculture fields; however, further studies are needed, such as clarifying the interaction mechanism between PEG and casein NPs, and testing any improvements in the stabilities of casein NPs loaded with other hydrophobic substances.

## Author contributions

Conceptualization: Q. D., X. H. and P. J.; methodology: X. H., S. H., D. Q. and Q. D.; software: X. H., S. H. and D. Q.; validation: Q.



D., X. H. and P. J.; formal analysis: X. H., S. H., D. Q. and Y. Z.; investigation: X. H. and S. H.; resources: Q. D.; data curation: Q. D., X. H. and P. J.; writing – original draft preparation: Q. D. and P. J.; writing – review and editing: Q. D. and P. J.; visualization: Q. D. and P. J.; supervision: Q. D.; project administration: Q. D.; funding acquisition: D. Q. and Q. D. All authors have read and agreed to the published version of the manuscript.

## Conflicts of interest

Authors declare no conflicts of interest.

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## References

- 1 F. Rehan, N. Ahemad, R. A. Islam, M. Gupta, S. H. Gan and E. H. Chowdhury, Optimization and formulation of nanostructured and self-assembled caseinate micelles for enhanced cytotoxic effects of paclitaxel on breast cancer cells, *Pharmaceutics*, 2020, **12**, 984.
- 2 J. Zhang, Y. Liu, X. Liu, Y. Li, X. Yin, M. Subirade, P. Zhou and L. Liang, The folic acid/ $\beta$ -casein complex: characteristics and physicochemical implications, *Food Res. Int.*, 2014, **57**, 162–167.
- 3 S. M. Jafari, M. Fathi and I. Mandala, Emerging product formation, in *Food Waste Recovery: Processing Technologies and Industrial Techniques*, Elsevier Inc., 2015, pp. 293–317.
- 4 E. Assadpour, S. M. Jafari and Y. Maghsoudlou, Evaluation of folic acid release from spray dried powder particles of pectin-whey protein nano-capsules, *Int. J. Biol. Macromol.*, 2017, **95**, 238–247.
- 5 E. Assadpour, Y. Maghsoudlou, S. M. Jafari, M. Ghorbani and M. Aalami, Evaluation of folic acid nano-encapsulation by double emulsions, *Food Bioprocess Technol.*, 2016, **9**, 2024–2032.
- 6 G. M. Tavares, T. Croguennec, A. F. De Carvalho and S. Bouhallab, Milk proteins as encapsulation devices and delivery vehicles: applications and trends, *Trends Food Sci. Technol.*, 2014, **37**, 5–20.
- 7 F. Kimpel and J. J. Schmitt, Review: milk proteins as nanocarrier systems for hydrophobic nutraceuticals, *J. Food Sci.*, 2015, **80**, 2361–2366.
- 8 Y. D. Livney, Milk proteins as vehicles for bioactives, *Curr. Opin. Colloid Interface Sci.*, 2010, **15**, 73–83.
- 9 M. H. Tunick and D. L. Van Hekken, Dairy products and health: recent insights, *J. Agric. Food Chem.*, 2015, **63**, 9381–9388.
- 10 A. Sahu, N. Kasoju and U. Bora, Fluorescence study of the curcumin-casein micelle complexation and its application as a drug nanocarrier to cancer cells, *Biomacromolecules*, 2008, **9**, 2905–2912.
- 11 P. Zimet, D. Rosenberg and Y. D. Livney, Re-assembled casein micelles and casein nanoparticles as nano-vehicles for  $\omega$ -3 polyunsaturated fatty acids, *Food Hydrocolloids*, 2011, **25**, 1270–1276.
- 12 M. Haham, S. Ishshalom, M. Nodelman, I. Duek, E. Segal, M. Kustanovich and Y. D. Livney, Stability and bioavailability of vitamin D nanoencapsulated in casein micelles, *Food Funct.*, 2012, **3**, 737–744.
- 13 E. Semo, E. Kesselman, D. Danino and Y. D. Livney, Casein micelle as a natural nano-capsular vehicle for nutraceuticals, *Food Hydrocolloids*, 2007, **21**, 936–942.
- 14 G. Markman and Y. D. Livney, Maillard-conjugate based core-shell co-assemblies for nanoencapsulation of hydrophobic nutraceuticals in clear beverages, *Food Funct.*, 2012, **3**, 262–270.
- 15 Y. Levinson, S. Ishshalom, E. Segal and Y. D. Livney, Bioavailability, rheology and sensory evaluation of fat-free yogurt enriched with VD3 encapsulated in re-assembled casein micelles, *Food Funct.*, 2016, **7**, 1477–1482.
- 16 Y. Cohen, S. Ishshalom, E. Segal, O. Nudelman, A. Shpigelman and Y. D. Livney, The bioavailability of vitamin D3, a model hydrophobic nutraceutical, in casein micelles, as model protein nanoparticles: human clinical trial results, *J. Funct. Foods*, 2017, **30**, 321–325.
- 17 J. Yi, T. I. Lam, W. Yokoyama, L. W. Cheng and F. Zhong, Beta-carotene encapsulated in food protein nanoparticles reduces peroxy radical oxidation in Caco-2 cells, *Food Hydrocolloids*, 2015, **43**, 31–40.
- 18 Y. Zhang, S. He, Y. Ma, W. Xu and H. Tang, Characterization and bioaccessibility of  $\beta$ -carotene in re-assembled casein, *RSC Adv.*, 2015, **5**, 77595–77600.
- 19 X. Pan, P. Yao and M. Jiang, Simultaneous nanoparticle formation and encapsulation driven by hydrophobic interaction of casein-graft-dextran and  $\beta$ -carotene, *J. Colloid Interface Sci.*, 2007, **315**, 456–463.
- 20 H. Chen, Y. Zhang and Q. Zhong, Physical and antimicrobial properties of spray-dried zein-casein nanocapsules with co-encapsulated eugenol and thymol, *J. Food Eng.*, 2015, **144**, 93–102.
- 21 S. Haratifar and A. Guri, 5-Nanocapsule formation by caseins A2-Jafari, Seid Mahdi, in *Nanoencapsulation Technologies for the Food and Nutraceutical Industries*, Academic Press, 2017, pp. 140–164.
- 22 D. S. Horne, Casein micelle structure and stability, in *Milk proteins: From expression to food*, ed. M. B. A. Thompson and H. Singh, Academic Press/Elsevier, Paris-Amsterdam-Boston, 2020, pp. 133–162.
- 23 S. Ribes, M. Ruiz-Rico, É. Pérez-Estève, A. Fuentes, P. Talens, R. Martínez-Mañez and J. M. Barat, Eugenol and thymol immobilised on mesoporous silica-based material as an innovative antifungal system: application in strawberry jam, *Food Control*, 2017, **81**, 181–188.
- 24 P. Suwanamornlert, S. Sangchote, W. Chinsirikul, A. Sane and V. Chonhenchob, Antifungal activity of plant-derived compounds and their synergism against major postharvest



- pathogens of longan fruit in vitro, *Int. J. Food Microbiol.*, 2018, **271**, 8–14.
- 25 L. Gong, T. Li, F. Chen, X. Duan, Y. Yuan, D. Zhang and Y. Jiang, An inclusion complex of eugenol into  $\beta$ -cyclodextrin: preparation, and physicochemical and antifungal characterization, *Food Chem.*, 2016, **196**, 324–330.
- 26 Y. Bai, Y. Cui, G. C. Paoli, C. Shi, D. Wang and X. Shi, Nanoparticles affect PCR primarily via surface interactions with PCR components: using amino-modified silica-coated magnetic nanoparticles as a main model, *ACS Appl. Mater. Interfaces*, 2015, **7**, 13142–13153.
- 27 K. A. Abd-Elsalam and A. R. Khokhlov, Eugenol oil nanoemulsion: antifungal activity against *Fusarium oxysporum* f. sp. *vasinfectum* and phytotoxicity on cottonseeds, *Appl. Nanosci.*, 2015, **5**, 255–265.
- 28 S. Woranuch and R. Yoksan, Eugenol-loaded chitosan nanoparticles: I. Thermal stability improvement of eugenol through encapsulation, *Carbohydr. Polym.*, 2013, **96**, 578–585.
- 29 P. Jin, R. Yao, D. Qin, Q. Chen and Q. Du, Enhancement in antibacterial activities of eugenol-entrapped ethosome nanoparticles via strengthening its permeability and sustained release, *J. Agric. Food Chem.*, 2019, **67**, 1371–1380.
- 30 Z. Hong, Y. Q. Xu, J. F. Yin, J. Jin and Q. Du, Improving the effectiveness of (–)-epigallocatechin gallate (EGCG) against rabbit atherosclerosis by EGCG-loaded nanoparticles prepared from chitosan and polyaspartic acid, *J. Agric. Food Chem.*, 2014, **62**, 12603–12609.
- 31 L. Xv, X. Qian, Y. Wang, C. Yu, D. Qin, Y. Zhang, P. Jin and Q. Du, Structural modification of nanomicelles through phosphatidylcholine: the enhanced drug-loading capacity and anticancer activity of celecoxib-casein nanoparticles for the intravenous delivery of celecoxib, *Nanomaterials*, 2020, **10**, 451.
- 32 W. B. Suleiman, *In vitro* estimation of superfluid critical extracts of some plants for their antimicrobial potential, phytochemistry, and GC-MS analyses, *Ann. Clin. Microbiol. Antimicrob.*, 2020, **19**, 29.
- 33 J. Gao, J. X. Wong, J. C. Lim, J. Henry and W. Zhou, Influence of bread structure on human oral processing, *J. Food Eng.*, 2015, **167**, 147–155.
- 34 Y. Luo, K. Pan and Q. Zhong, Casein/pectin nanocomplexes as potential oral delivery vehicles, *Int. J. Pharm.*, 2015, **486**, 59–68.
- 35 N. D. Trinh, T. T. B. Nguyen and T. H. Nguyen, Preparation and characterization of silver chloride nanoparticles as an antibacterial agent, *Adv. Nat. Sci.: Nanosci. Nanotechnol.*, 2015, **6**, 045011.

