


 Cite this: *RSC Adv.*, 2021, 11, 1901

Effects of amino acid ionic liquids with different cations ([N₂Py], [N₂₂₂₂], [P₂₂₂₂], and [C₂mim]) on wheat seedlings

Zhonglin Chen, Hong Xiao, Siqi Wu, Jie Wang, * Jiayang Ji and Xiao Qin

The ecotoxicity of four ionic liquids with different cations (*N*-ethyl-pyridine alanine [N₂Py][Ala], tetraethyl phosphine L- α -amino propionic acid salt [P₂₂₂₂][Ala], 1-ethyl-3-methyl-imidazolium alanine [C₂mim][Ala], and tetraethyl ammonium L- α -amino propionic acid salt [N₂₂₂₂][Ala]) was assessed in hydroponically-grown wheat seedlings at concentrations from 200–1200 mg L⁻¹. The results showed that type of cation has a significant influence on the growth, chlorophyll and nutrient uptake of wheat seedlings ($P < 0.05$). We observed decreased dry weight and shorter roots and shoots in the treated seedlings with increasing IL concentrations. The contents of Chl *a* and Chl *b* in wheat seedlings exposed to ILs showed the trend of firstly increasing followed by a decrease with increasing IL concentrations, but they peaked at different concentrations of ILs. In addition, the exposure of wheat seedling to ILs containing different cations (200–1200 mg L⁻¹) led to first an increase and then a decrease of nitrogen content, and reduced the content of phosphorus and potassium. Moreover, the cellular structures, including nuclei, mitochondria, chloroplasts, cell membranes, and the cell walls of wheat leaf and root were affected to varying degrees by 600 mg L⁻¹ ILs. The negative impacts of ILs on wheat seedlings ranked from high to low were: [N₂Py][Ala] > [N₂₂₂₂][Ala] > [P₂₂₂₂][Ala] > [C₂mim][Ala]. In this work, the relatively stronger toxicity of [N₂Py][Ala] was likely contributed by ethanol, which was used to dissolve [N₂Py][Ala]. Therefore, it is not recommended to use *N*-ethyl-pyridine alanine ([N₂Py][Ala]) widely in practical applications.

 Received 12th October 2020
 Accepted 14th December 2020

DOI: 10.1039/d0ra08350j

rsc.li/rsc-advances

1. Introduction

Ionic liquids (ILs) are organic salts composed of organic cations and organic or inorganic anions that are liquid at room temperature.¹ They are considered new alternative solvents that can be widely applied, especially as an important medium, in the field of synthesis and catalysis. Different cations and anions can be combined to design a wide variety of ionic liquids. The cations of ionic liquids mainly include imidazole, pyridine, quaternary ammonium, and quaternary phosphine. The anions are mainly ionic halide salts, such as Cl⁻ and Br⁻, and ionic non-halide salts, such as BF₄⁻, PF₆⁻, CF₃SO₃⁻, and SbF₆⁻.²

Existing studies have shown that ILs have harmful effects on fish, bacteria, algae, and higher plants.^{3,4} The toxicity of ILs is determined by the type of cations and anions and the length of the alkyl substituent. Tot *et al.* investigated the acute developmental toxicity, antioxidant activity, and membrane lipid peroxidation level of snails exposed to [C_{*n*}mim]Br ($n = 6, 8, 10, 12$), and found the snail death rate increased with the length of the carbon chain.⁵ Ma found that several imidazole-type ionic liquids with varying alkaline chain length suppressed the

growth of maize seedlings, and the longer the carbon chain, the greater the toxicity.⁶ Other researchers found similar results in wheat, using [C_{*n*}mim][OAc] ($n = 2, 4, 6$) ILs, again finding that the longer the carbon chain, the greater the IL toxicity to the plants.^{7,8} Habibur *et al.* also got similar results that the toxic effects of ILs on rice growth depends on the alkyl chain length: [C₈mim]Br > [C₄mim]Br > [C₂mim]Br.⁹

There have been extensive studies on the contribution of anions to the overall toxicity of ILs.^{10–12} Liu *et al.*¹⁰ explored the effects of three imidazole-type ionic liquids containing different anions on broad bean seedlings. The results showed that the effects of toxicity on broad bean seedlings were as follows: BF₄⁻ > Br⁻ > Cl⁻. Imidazole-type ionic liquids containing different anions have increasing toxicity on hydrogen-producing bacteria, as the number of fluorine atoms in anions increases.¹¹ Moreover, the toxicity of Imidazole-type ILs with different anions on wheat seedling growth was assessed in a previous study, and the toxicity of different anions followed [TfO]⁻ > [Cl]⁻ > [BF₄]⁻ > [Lact]⁻ > [Ala]⁻.¹²

However, there have been few studies on the effects of cations on IL toxicity.¹³ Couling *et al.* evaluated the toxicity of 23 ILs on the aquatic organisms *Vibrio cheri* and *Daphnia magna* using the quantitative structure–property relationship. When the substitution side chain and anions were the same, the

School of Environmental Science, Liaoning University, Shenyang 110036, PR China.
 E-mail: 94266689@qq.com



toxicity of the following five types of cationic cores on the two aquatic organisms showed similar patterns: tetrazole cations > triazole cations > imidazole cations > pyridine cations > quaternary ammonium salts cations.¹³ Liu found that imidazole-type ILs with different cations had greater toxicity than pyridine-type ILs on the freshwater green alga *Scenedesmus obliquus*.¹⁴ And the cytotoxicity of ionic liquids mainly depends on their ability to permeabilize/disrupt the cellular membrane.¹⁵ Notably, these studies of cations on IL toxicity to organisms have only involved aquatic species; the toxicity of different cations on higher plants has not been reported.

Therefore, we tested the effect of four types of ILs with different cations on wheat seedlings: [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala], and [N₂₂₂₂][Ala]. The influence on the growth, chlorophyll contents, and nutrients uptake of wheat seedlings, was assessed in this study. Furthermore, we examined the ultra-microcellular structures of the root and leaf tip cells by transmission electron microscopy. This study provides data support and an important reference for assessing the ecotoxicological effects and environmental safety of ILs.

2. Materials and methods

2.1 Ionic liquids

The ionic liquids [N₂Py][Ala] (25% purity, dissolved in ethanol), [P₂₂₂₂][Ala] (97% purity, dissolved in water), [C₂mim][Ala] (97% purity, dissolved in water), and [N₂₂₂₂][Ala] (97% purity, dissolved in water), were attained from Lanzhou Institute of Chemical Physics, Gansu, China. In the experiment, seven different concentrations (0, 200, 400, 600, 800, 1,000, and 1200 mg L⁻¹) of each IL as used. Furthermore, the same dose of ethanol series concentration as the solvent of [N₂Py][Ala] was prepared, to analyze the interference effect of ethanol on the toxicity of [N₂Py][Ala] (Fig. 1).

2.2 Wheat culture

Wheat seeds were purchased from Liaoning Academy of Agricultural Sciences, Liaoning, China. Following Chen *et al.*,¹⁶ wheat seeds of the same size were sterilized prior to use with 0.1% HgCl₂ solution for 15 min and then rinsed with deionized water until the surface mucus was washed away. After filtering and drying, 30 seeds were dispersed evenly across an 80 mm diameter Petri dish for each experimental group and then covered with three layers of gauze. Each treatment group was provided with three replicates. We added a small amount of distilled water to the control group, and the same amount of solutions of four ILs or ethanol with different concentrations

(200, 400, 600, 800, 1,000, and 1200 mg L⁻¹, prepared with distilled water) to each treatment group. The samples were incubated at 28 °C and allowed to germinate for 24 h. Additional treatment solution was added every 8 h to keep the seeds moist. Once the seeds were evenly germinated, they were transferred to a plastic beaker tied with a nylon mesh, and the beaker had been pre-filled with the Hoagland nutrient solution (Control) or Hoagland solutions containing four ILs or ethanol with the appropriate concentrations (200, 400, 600, 800, 1,000, and 1200 mg L⁻¹).¹⁷ It should be noted that, nutrient solution was added to each beaker such that the root tips of the germinated wheat seeds were just submerged.⁷ The beakers were exposed to natural light with a 10 h/14 h light/dark photoperiod for 15 d with 22 °C and 17 °C day and night temperatures. The appropriate Hoagland solution was added as needed to maintain the growth of wheat plants.

2.3 Determination methods

2.3.1 Growth indexes. After 15 d, when the wheat seedlings grew to one heart and two leaves, four growth indicators, including plant height, root length, aerial and underground biomass, were measured. Following protocols from Chen *et al.*,¹⁶ we randomly selected three seedlings from each treatment group. We divided the seedlings into the aerial and underground portions. The endosperm was removed and surface moisture was carefully dried with filter paper. A vernier caliper was for plant height and root length measurement. Subsequently, the samples were then placed in a drying oven set at a constant temperature. The samples were dried at 100 °C for 30 min and then at 80 °C until a constant weight was achieved.¹⁶ After drying, an electronic analytical balance (BS 224S, 0.0001 g, Beijing Sartorius Instrument Co., Ltd. China) was used to measure the dry weights of the aerial and underground parts.

2.3.2 Photosynthetic pigments. After 8 d exposure, photosynthetic pigments were determined using fresh leaves (0.1 g, accurate to 0.0001 g) ground into a fine powder, which were suspended in 20 mL 80% of acetone and incubated in the dark for 24 h. The contents of chlorophyll pigments (Chl *a*, Chl *b*) were determined using a spectrophotometer (UV5500, Shanghai Metash Instruments Co., Ltd. China) at 663 and 645 nm. Calculations were performed following the method described by Zhang.¹⁸

2.3.3 Transmission electron microscopy (TEM). The mesophyll and root tip cells of the control and wheat plants treated by 600 mg L⁻¹ were examined with a transmission electron microscopy (HITACHI-H7650, Japan) on 15th d. At first, the leaf and root tips were collected, and the veins were removed. The tissues were cut into rectangular segments with a length of 1–2 cm and a width of 2–3 mm and fixed in 2.5% glutaraldehyde for 72 h at 4 °C.¹⁹ After fixation, the samples were processed three times by phosphate buffer (0.1 M, pH7.0), with 15 min per wash. Next, they were fixed in 1% osmium tetroxide and washed three times in phosphate buffer (0.1 M, pH7.0). Afterwards, the samples were dehydrated by a series of ethanol solutions (50%, 70%, 80%, 90%, 95% and 100%) for 20 min at each concentration before being treated in pure acetone.

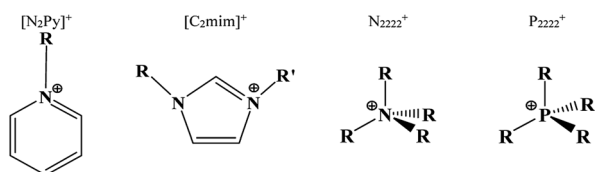


Fig. 1 The cationic structure of ILs.



Subsequently, samples were transferred to a mixture solutions (pure acetone : embedding medium = 1 : 1, calculated by volume) for 1 h, followed by transferring to a 1 : 3 (v/v) mixture of pure acetone and Spurr's resin for 3 h at room temperature and then they were placed in pure embedding medium overnight. After that, the samples were heated at 70 °C for 12 h approximately and then the ultrathin sections were cut with a diamond knife. Later, they were stained with 50% uranyl acetate for 30 min followed by 30% lead citrate for 15 min.¹⁹ At last, the samples were scanned through TEM.

2.3.4 Determination of N, P and K. Samples (0.1 g, accurate to 0.0001 g) of the dried and finely ground leaves (0.5 mm sieve) were digested using concentrated H₂SO₄-H₂O₂ method (NY/T 2017-2011).²⁰ At first, they were put into the bottom of 100 mL sterilizing tube, and moistened with 1 mL of water, then added 5 mL of concentrated H₂SO₄ followed by shaking well. Later, the samples were added H₂O₂ (30%) solution in two times, 2 mL each time, and capped with a small funnel after shaking well. At the end of the intense reaction, they were heated on the digester to make the solid disappear into a solution. When the concentrated H₂SO₄ released white smoke and the solution turned brown, heating was stopped. After that, they were removed from the digester, and added 10 drops H₂O₂ (30%) after cooling slightly, and the tube was shaken thoroughly. Subsequently, the solution was reheated to a slight boil for 5-10 min followed by adding H₂O₂ (30%) 5-10 drops after a little cooling, which was repeated several times until the solution was colorless or clear. Then, the solution should be reheated for about 10 min to remove the remaining H₂O₂. The discolouring solutions were transferred harmlessly with water into a volumetric flask (100 mL) and cooled to room temperature. At last, the solution was filtrated with dry filter paper to determine N, P and K. The concentrations of N, P and K were determined by the Kjeldah method (KDN-08C, Shanghai Xinrui Instrumentation Co., Ltd. China), ammonium molybdate method (UV5500, Shanghai Metash Instruments Co., Ltd, China), and using an atomic absorption spectrophotometer (TAS 990F, Beijing Purkinje General Instrument Co., Ltd, China) respectively.²⁰

2.4 Data analysis

A least significant difference (LSD) test of univariate analysis of variance (ANOVA) was used to examine the differences between different groups in SPSS 17.0 (IBM Corp., Armonk, NY, USA). Origin 7.0 (OriginLab Corp. Northampton, Massachusetts, USA) was used for plotting.

3. Results and discussion

3.1 Effects of the ILs with different cations ([N₂Py], [P₂₂₂₂], [C₂mim] and [N₂₂₂₂]) on the growth of wheat seedlings

When subjected to biotic or abiotic stress, plants often show reduced dry weight, plant height, and root length, which are among the most direct growth indicators.²¹ Wheat growth was inhibited with increasing [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala], [N₂₂₂₂][Ala], and ethanol concentrations in all five treatment

groups (Fig. 2 and 3). Especially, the plants grew very short when the both treatments (N₂Py [Ala] and ethanol) were given dosages greater than 800 mg L⁻¹. Tot *et al.* also found that, under strong ionic liquid stress ([Im][Cl], [Bmim][Cl], [C₂OC₂mIm][Cl], [C₁OC₂mIm][Cl], [OHC₃eIm][Cl], [OHC₃mIm][Cl], [OHC₂mIm][Cl], 1000 mg L⁻¹) adverse effects of ionic liquids on plant seedling growth is visible.⁵ The growth indices of ethanol and [N₂Py][Ala] treatment groups showed the largest inhibited effects, which is consistent with the published research.¹⁶

As shown in Fig. 4, different cationic ionic liquids had significant effects on the growth of wheat seedlings (*P* < 0.05). The aerial dry weight, underground dry weight, plant height, and root length of the treatment groups were lower than corresponding controls, except for the underground dry weight of the wheat seedlings under the 200-400 mg L⁻¹ [C₂mim][Ala] treatment, which were higher than control. Compared with control, the inhibited effects of the treatments of [N₂Py]Ala, [P₂₂₂₂]Ala, [C₂mim]Ala, [N₂₂₂₂]Ala and ethanol on wheat growth was the strongest at dosage of 1200 mg L⁻¹, the corresponding inhibition rates for plant height were 83.3%, 37.0%, 43.4%, 52.7% and 87.5% respectively, the corresponding inhibition rates for root length were 96.1%, 67.6%, 50.1%, 72.9% and 78.3% respectively, the corresponding inhibition rates for aerial dry weight were 79.1%, 43.0%, 49.0%, 51.9% and 88.4% respectively, the corresponding inhibition rates for underground dry weight were 87.6%, 37.8%, 42.1%, 49.3% and 78.2% respectively. These results indicate that the growth of wheat seedlings was negatively affected by four ILs in general (Fig. 4). The increased underground dry weight under the 200-400 mg L⁻¹ [C₂mim][Ala] treatment was likely a result of the fibrous roots system of wheat plants, which had an increased number of roots despite a reduced root length under this treatment.

The treatments of ethanol-dissolved [N₂Py][Ala] and pure ethanol decreased wheat growth the most among all treatments, indicating that [N₂Py][Ala] and ethanol were the most toxic to wheat growth. However, the two exerted opposite effects on the underground and aboveground parts, with the aboveground part more strongly affected by ethanol than [N₂Py][Ala] and the underground part more severely affected by [N₂Py][Ala]

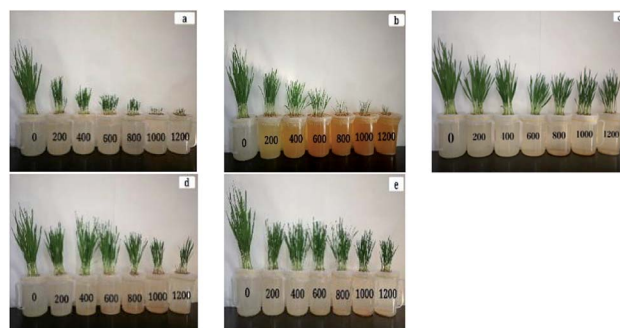


Fig. 2 Effects of different concentrations of treatment (0, 200, 400, 600, 800, 1000 and 1200 mg L⁻¹) of wheat seedlings on the growth under ILs stress after 15 d. (a) Ethanol, (b) [N₂Py][Ala], (c) [P₂₂₂₂][Ala], (d) [C₂mim][Ala], (e) [N₂₂₂₂][Ala].



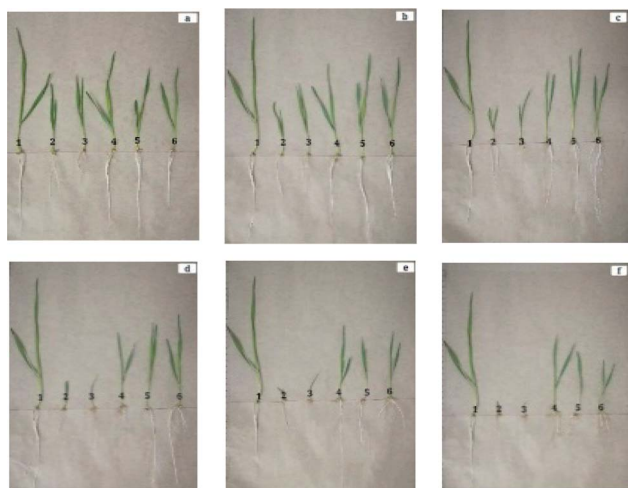


Fig. 3 Effects of different concentrations of treatment on the growth of wheat seedlings under IL and ethanol stress after 8 d. (a) 200 mg L⁻¹; (b) 400 mg L⁻¹; (c) 600 mg L⁻¹; (d) 800 mg L⁻¹; (e) 1000 mg L⁻¹; (f) 1200 mg L⁻¹. 1–6 were respectively control, ethanol, [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala] and [N₂₂₂₂][Ala].

than ethanol, suggesting a greater impact of [N₂Py][Ala] on root length. The results are consistent with the study of Liu *et al.* on the effects of 1-octyl-3-methylimidazolium chloride ionic liquid on rice seedlings.²¹ Moreover, we found in the experiment that the roots of wheat seedlings grown in [N₂Py][Ala] solution showed more serious erosion phenomenon. Habibul *et al.* also found the similar phenomenon in wheat root exposure to [C₈mim]Ala.⁹ The results reveal that [N₂Py][Ala] was the most toxic among the four ILs, one possible reason was that its structure was well maintained in ethanol, which exacerbated the toxicity of [N₂Py][Ala], the another reason may be that cations possessing aromatic head-groups are more toxic than the non-aromatic ones.²² And the ranked degree of toxicity from high to low was: [N₂Py][Ala] > [N₂₂₂₂][Ala] > [P₂₂₂₂][Ala] > [C₂mim][Ala].

3.2 Effects of ILs with different cations ([N₂Py], [P₂₂₂₂], [C₂mim] and [N₂₂₂₂]) on the chlorophyll content of wheat seedlings

Chlorophyll is the main pigment for plant photosynthesis, the content of chlorophyll in plant leaves has a direct influence on the synthesis of organic matter in the plant body and then on the growth and development of the plant.¹⁸ The effects of four ILs on Chl *a* and Chl *b* productions in the leaves of wheat seedlings is shown in Fig. 5.

As shown in Fig. 5, the effect of different ILs on the chlorophyll of wheat seedlings was significantly different at the same concentration ($P < 0.05$). In general, the contents of Chl *a* and Chl *b* in wheat seedlings exposed to ILs ([P₂₂₂₂][Ala], [C₂mim][Ala] and [N₂₂₂₂][Ala]) showed the same trend of slow increase followed by a slight decrease with increasing ILs concentrations. The chlorophyll (Chl *a*, Chl *b*) peaked at 600 mg L⁻¹ ILs. At the same concentration, their contents of Chl *a* were 36.9%, 41.0%, and 61.2% higher than those in the control plants, respectively,

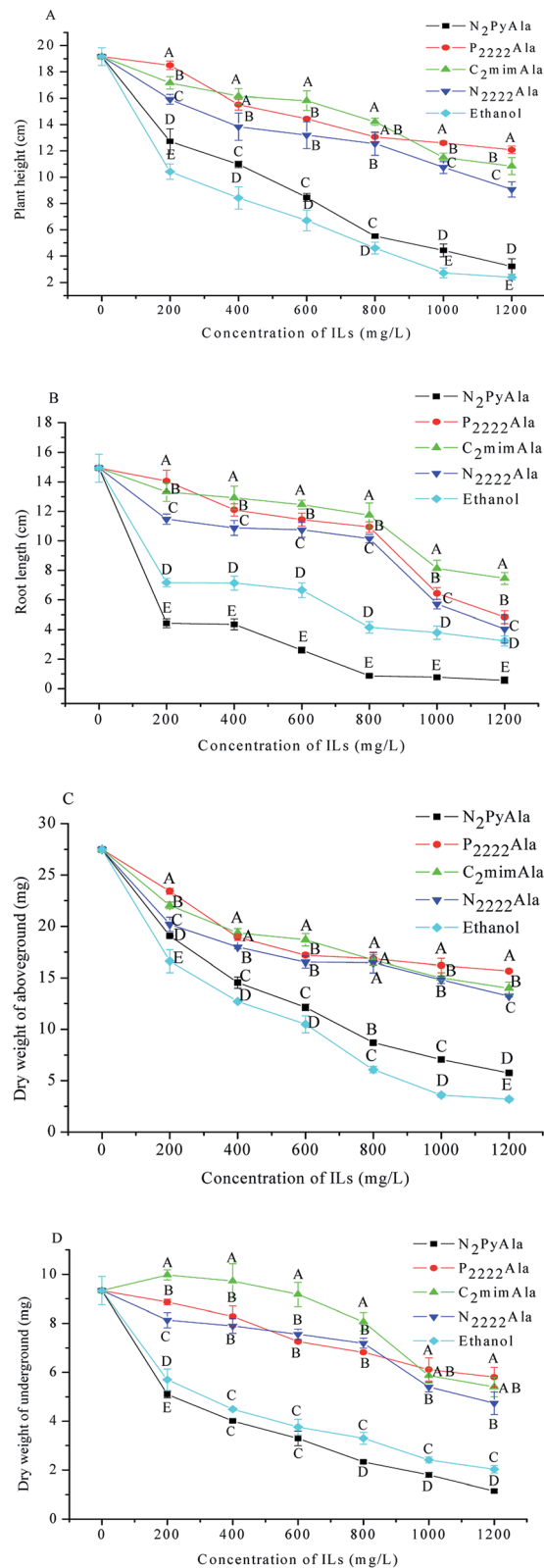


Fig. 4 Effects of ethanol, [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala] and [N₂₂₂₂][Ala] on (A) plant height, (B) root length, (C) aboveground dry weight, and (D) underground dry weight of wheat seedlings. Note: different capital letters represent significant differences between treatments with different ILs of the same concentrations at 0.05 level. The same below.



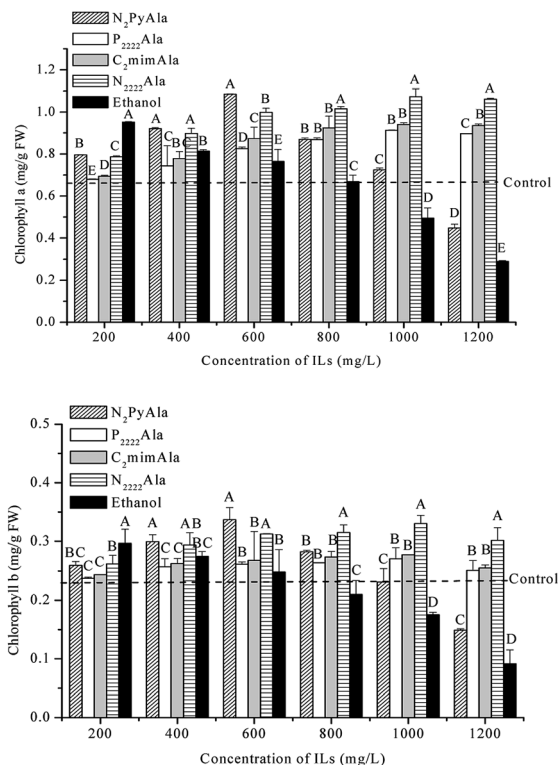


Fig. 5 Effects of ethanol, $[N_2Py][Ala]$, $[P_{2222}][Ala]$, $[C_2mim][Ala]$ and $[N_{2222}][Ala]$ on the chlorophyll *a* and chlorophyll *b* of wheat seedlings.

whereas their contents of Chl *b* were 18.3%, 21.3%, and 44.5%, respectively, of those observed in control. This may be because the use of low concentration of ILs can induce the accumulation of pigment. However, with the increase of concentration of ILs, the amount of ILs entering the plant increases, and ILs could increase membrane permeability by relaxing the bilayer, cause damage to the chloroplast membrane structure and inhibit chlorophyll synthesis.²³

On the other hand, the treatments of $[N_2Py][Ala]$ and ethanol had the similar effects on chlorophyll in wheat seedlings. With the increase of $[N_2Py][Ala]$ or ethanol dose, chlorophyll (Chl *a*, Chl *b*) contents in wheat seedlings increased at first and then decreased sharply, they were the largest at 600 mg L⁻¹ for $[N_2Py][Ala]$ and 200 mg L⁻¹ for ethanol, respectively. At the same concentration, their contents of Chl *a* were 62.7% and 47.3% higher than those in control, respectively, whereas their contents of Chl *b* were 47.3% and 29.6% higher than those in control. However, when the concentration of $[N_2Py][Ala]$ or ethanol was 1200 mg L⁻¹, the chlorophyll content was significantly lower than that of the control group. The reason for the results was that, plants grown in a solution containing ethanol can only breathe anaerobically, which makes it difficult for plants to absorb nutrients, leading to disruption of membrane physiological functions and, consequently, to the increased toxicity.⁵ Zhu *et al.* also found that with the increasing of the concentration of IL, the number of viable cells decreased tremendously.²⁴ Moreover, some studies suggest that, under saline-alkali stress, the content of chlorophyll *b* is decreased

and the content of chlorophyll *a* is increased in order to maintain the absorption of water and nutrients.²⁵ Whereas, Liu *et al.* found that, chlorophyll synthesis was severely inhibited in *A. thaliana* by the three ILs ($[C_8mim]Cl$, $[C_{10}mim]Cl$, and $[C_{12}mim]Cl$).²⁶ In our study, the contents of Chl *a* and Chl *b* in wheat seedlings showed different trends with the increase of ionic liquid, but the chlorophyll *a/b* values had the increasing trends except for individual treatments. This may be because different types and concentrations of ionic liquids have different effects on plants.

3.3 Effects of ILs with different cations ($[N_2Py]$, $[P_{2222}]$, $[C_2mim]$ and $[N_{2222}]$) on the nutrient uptake of wheat seedlings

Nitrogen (N), phosphorus (P) and potassium (K) are essential macronutrients for all organisms and referred as the “three essential elements” for plant growth.²⁷ As shown in Fig. 6, The contents of N, P and K in wheat seedlings were significantly ($P < 0.05$) different between treatments of ILs at the same concentration. Firstly, the contents of N in wheat seedlings exposed to $[P_{2222}][Ala]$, $[C_2mim][Ala]$ and $[N_{2222}][Ala]$ showed the trends of slow

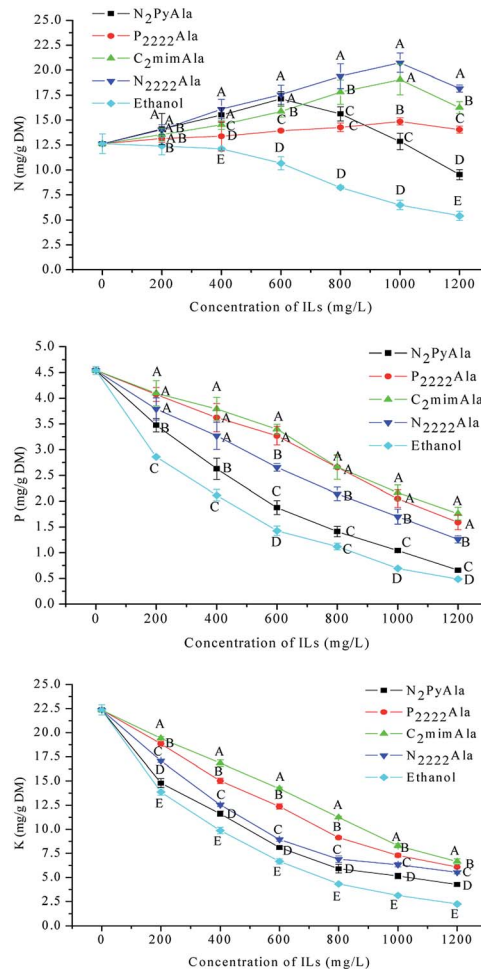


Fig. 6 Effects of ethanol, $[N_2Py][Ala]$, $[P_{2222}][Ala]$, $[C_2mim][Ala]$ and $[N_{2222}][Ala]$ on the nutrients' absorption of wheat seedlings.



increase followed by a slight decrease with increasing ILs concentrations, and peaked at 1000 mg L⁻¹ ILs. At the same concentration, their contents of N were 17.8%, 51.1%, and 64.3% higher than those in the control plants, respectively. Whereas, with the increase of [N₂Py]Ala dosage, nitrogen contents in wheat seedlings increased at first and then decreased sharply, they were the largest at 600 mg L⁻¹, and was 35.74% higher than those in control. Moreover, with the increase of ethanol concentration, the contents of N in wheat seedlings decreased gradually. Specially, the exposure to 1200 mg L⁻¹ [N₂Py]Ala and ethanol decreased N contents in plant by 24.6% and 57.3%, respectively, compared to control plants.

However, the contents of P showed a downward trend with increasing ILs or ethanol concentrations, which was similar to the variation trends of growth indexes. Compared with the control, the contents of P in wheat seedlings exposure to 200–1200 mg L⁻¹ ILs ([N₂Py]Ala, [P₂₂₂₂]Ala, [C₂mim]Ala, [N₂₂₂₂]Ala and ethanol) declined by 23.3–85.4%, 10.1–64.9%, 9.6–61.2%, 16.3–72.2%, and 36.9–89.3%, respectively.

Meanwhile, the same variation trend of P was observed in the K contents in wheat seedlings exposure to ILs. Compared with the control, The K content in wheat seedlings exposure to the ILs with different cations ([N₂Py], [P₂₂₂₂], [C₂mim] and [N₂₂₂₂]) and ethanol at 200–1200 mg L⁻¹ were reduced by 33.9–80.9%, 15.7–72.9%, 13.1–70.3%, 23.6–75.2%, and 38.0–89.9%, respectively.

Above all, the exposure of wheat seedling to ILs containing different cations (200–1200 mg L⁻¹), led to the first increase and then decrease of nitrogen contents, and reduced the content of phosphorus and potassium. These may be because the ILs used in this experiment contains alanine ions, which could translocate into the plant and increased the nitrogen contents correspondingly. As for the decline of N contents at the dosage of being greater than 1000 mg L⁻¹, one possible explanation was that ILs with high concentration harms plant cells and inhibits its absorption of nutrients.⁹ Meanwhile, the results showed that the different chemical structure and initial concentration of ILs result in the different effect on plant.⁹ On the other hand, different cationic ionic liquids caused different degrees of toxicity to the wheat seedlings, which resulted in the hindrance of the absorption of P and K. And the inhibited effect ranged from high to low was [N₂Py][Ala] > [N₂₂₂₂][Ala] > [P₂₂₂₂][Ala] > [C₂mim][Ala]. Ionic liquid is a low temperature molten salt, it may affect plants through ion toxicity, osmotic stress or the interaction of the two, and the former will lead to plant physiological drought and ion imbalance.²⁸ Further more, the responses of different plant species, organs and growth periods to salt stress are also different.²⁹ Song *et al.* have shown that, when salt content increases, leaf P content decreases, leaf N content and leaf N:P increase,²⁹ whose results are consistent with us.

3.4 Effects of ILs with different cations ([N₂Py], [P₂₂₂₂], [C₂mim] and [N₂₂₂₂]) on the subcellular structure of the wheat leaves and roots

Environmental stress damages the ultrastructure of plant cells, thereby affecting plant growth and development.¹⁴ At

600 mg L⁻¹ ILs, the morphological differences of the ultrastructure of mesophyll (photos labelled “1”) and root tip cells (photos labelled “2”) of the control group and different treatment groups are shown in Fig. 6.

As shown in Fig. 7, the mesophyll cells of wheat seedlings in the control samples were morphologically intact with a clear structure, tight binding of cell wall and membrane, and smooth membrane (CK1). The nucleus was intact, the nuclear membrane was clear, and the chromatin was abundant and evenly distributed. However, upon [C₂mim][Ala] exposure, the cell wall, membrane, and nuclear membrane became vague (A1), and after exposures to [P₂₂₂₂][Ala] (B1) and [N₂₂₂₂][Ala] (C1), the cell walls and cell membranes became loosely bound, the plasma membranes shrank, and local wall separation occurred. In addition, the nucleus became irregularly-shaped, the chromatin distribution became uneven, and cell contents were significantly reduced.³⁰ The nuclear membrane even began to dissolve in [N₂Py][Ala]-treated (D1) and ethanol-treated (E1) wheat seedling, suggesting that nuclei were severely damaged.³¹ These changes indicate that exposure to ILs affected plant cells to varying degrees.

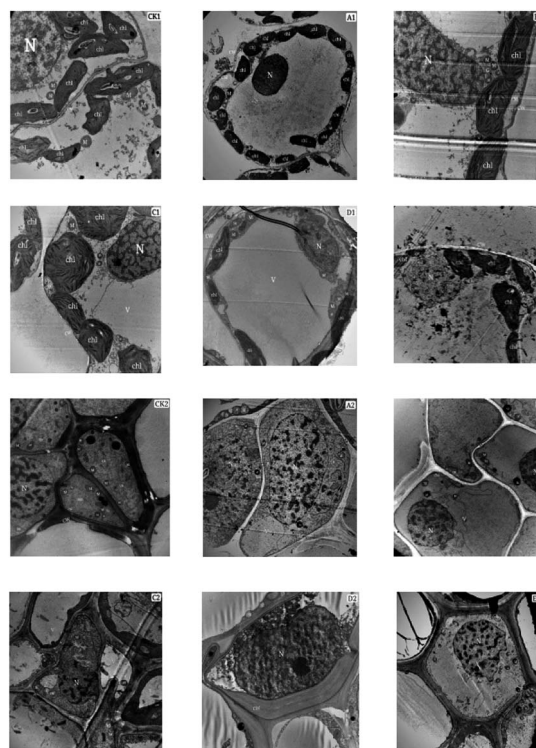


Fig. 7 Transmission electron micrographs of root cells and wheat leaf with ethanol, [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala] and [N₂₂₂₂][Ala] treatments. (The control group – CK1, CK2; [C₂mim][Ala]-A1, A2; [P₂₂₂₂][Ala]-B1, B2; [N₂₂₂₂][Ala]-C1, C2; [N₂Py][Ala]-D1, D2; ethanol – E1, E2). CK1 – control leaf cells; CK2 – control root cells; A1, B1, C1, D1, E1 – leaf cells treated with 600 mg L⁻¹ ethanol, [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala] and [N₂₂₂₂][Ala]; A2, B2, C2, D2 and E2 – root cells treated with 600 mg L⁻¹ [N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala] and [N₂₂₂₂][Ala]. Abbreviations: CM – cell membrane; CW – cell wall; N – nucleus; Mitochondrion; thy – thylakoid; Chl – chloroplast; ER – rough endoplasmic reticulum; G – Golgi apparatus; R – ribosome; V – vacuole).



Moreover, the chloroplasts in control samples were spindle-shaped, closely attached to the cell wall, and neatly and orderly distributed. The membrane structure of the chloroplast was intact and clear, and the thylakoids and grana lamellae in the chloroplasts were arranged neatly and parallel to the chloroplast long axis (CK1). By contrast, the thylakoids and grana lamellae were scattered and the chloroplasts were partially empty in [C₂mim][Ala]-treated wheat seedlings (A1). The chloroplasts swelled even more, and the thylakoids and grana lamellae were loosely arranged and disordered in the [P₂₂₂₂][Ala] (B1) and [N₂₂₂₂][Ala] (C1) treatment groups. The thylakoids and grana lamellae became disordered and the chloroplasts were irregularly-shaped in the [N₂Py][Ala] (D1) and ethanol (E1) treatment groups, suggesting damaged chloroplast structure. In addition, fewer chloroplasts were observed and some chloroplasts moved toward the center of the cell.

We observed a considerable number of mitochondria in the control (CK1) and [C₂mim][Ala]-treated (A1) samples. The mitochondria were of normal configuration, contained intact membranes and clearly visible cristae, and distributed around the chloroplasts. By contrast, in the [P₂₂₂₂][Ala] (B1) and [N₂₂₂₂][Ala] (C1) treatments, the number of mitochondria decreased, and the cristae became vague or even dissolved. Therefore, the [N₂Py][Ala] and ethanol treatments had likely hindered protein synthesis in mitochondria, which caused mitochondrion swelling and intimal rupture and resulted in reduced cellular metabolism and plant growth.¹⁶

The root tip cells of the control seedlings were intact, the cell wall and membrane were tightly attached and smooth, and the nuclear membrane was intact, and the chromatin was uniformly-coloured (CK2). The mitochondria had a regular shape, the inner cristae were intact (CK2). Compared with the control, [C₂mim][Ala]-treated wheat seedlings (A2) displayed swollen mitochondria but no significant change in the nuclear membrane. [P₂₂₂₂][Ala]- (B2) and [N₂₂₂₂][Ala]-treated wheat seedlings (C2) showed abnormally-stained and condensed chromosomes, unclear nuclear membrane, and fewer and scattered mitochondria with unclear crista structures. These observations indicate that the impact on mitochondria increased with increasing concentration as described above. In wheat seedlings subjected to the ethanol treatment (E2), we observed cell vacuolization, a reduced number of mitochondria, dissolution of the cristae, and scattered distribution of ribosomes. In addition, the cell walls were severely damaged and cell contents disappeared (D2).⁸

Compared with the control group, ILs with different cations ([N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala], and [N₂₂₂₂][Ala]) showed different degrees of toxicity to the cell structure of wheat seedlings, and the toxicity to root tip cells of wheat seedlings was more serious than that of mesophyll cells. This may be due to the preferential root accumulation of ILs in plant.⁹ Meanwhile, our results are consistent with Liu *et al.*²¹ and Xia *et al.*¹⁴ who found that ILs caused damage to the ultrastructure of plants. Chen *et al.*¹⁶ also found that ILs with different anions mainly damage chloroplasts, mitochondria, cell walls, cell membranes, and nuclei but have minimal on the Golgi in root cells and wheat leaves, which is in good agreement with the results of this

study. The ILs used in this study have different chemical structures compared with those employed in previous studies but caused similar damages to the plants on the subcellular level. Among all treatments tested in this study, ethanol and pyridine caused the greatest damage to mesophyll and apical organelles, which was in good agreement with the changes in growth indicators. Caution should be taken when interpreting the relatively stronger toxicity of pyridine compared with other ILs, which might be partly contributed by the toxicity of ethanol.

4. Conclusion

This paper revealed that ILs with different cations ([N₂Py][Ala], [P₂₂₂₂][Ala], [C₂mim][Ala], and [N₂₂₂₂][Ala]) at the dosage of 200–1200 mg L⁻¹ had significant inhibition on the wheat seedling growth ($P < 0.05$), led to the first increase and then decrease of nitrogen contents, and reduced the content of phosphorus and potassium significantly ($P < 0.05$). These may be because the ILs used in this experiment contains alanine ions, which could translocate into the plant and increased the nitrogen contents correspondingly. Above all, the inhibitory effect of ILs on the growth, chlorophyll contents, and nutrients uptake of wheat seedlings ranked from high to low was: [N₂Py][Ala] > [N₂₂₂₂][Ala] > [P₂₂₂₂][Ala] > [C₂mim][Ala]. Especially, the inhibitory effect of four ILs on wheat roots was significantly greater than that on shoots. Furthermore, the IL treatments exhibited toxicity to root tip and mesophyll organelle of wheat seedlings and altered their the structure to varying degrees, and the toxicity to root tip cells of wheat seedlings was more serious than that of mesophyll cells. Specially, [N₂Py][Ala] is only chemically-stable and structurally-unaffected in ethanol, which we found was particularly toxic to plants. For this reason, we do not recommend the use of *N*-ethyl-pyridine alanine ([N₂Py][Ala]) widely in practical applications.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The project was supported by the National Natural Science Foundation of China (31600311), college teaching reform and research of Liaoning university (LNDXJG20183014), Peoples Republic of China and Liaoning University “College Students’ Innovation and Entrepreneurship Training Program” (D201904152238479387).

Notes and references

- 1 Z. Q. Dong, L. Wei, S. W. Xu, *et al.*, *Ecotoxicol. Environ. Saf.*, 2018, **147**, 480–486.
- 2 L. Y. Lu, Y. J. Zhang, J. J. Chen, *et al.*, *Chin. J. Chem. Phys.*, 2017, **30**, 423–428.
- 3 S. A. S. Amiril, E. A. Rahim and S. Syahrullail, *J. Cleaner Prod.*, 2017, **168**, 1571–1589.
- 4 N. Meksi and A. Moussa, *J. Cleaner Prod.*, 2017, **161**, 105–126.



- 5 A. Tot, M. Vraneš, I. Maksimović, *et al.*, *Ecotoxicol. Environ. Saf.*, 2018, **147**, 401–406.
- 6 J. G. Ma, X. Y. Dong, Q. Fang, *et al.*, *J. Biochem. Mol. Toxicol.*, 2014, **28**, 69–75.
- 7 M. Yang, J. He, Y. Z. Sun, *et al.*, *J. Agro-Environ. Sci.*, 2017, **36**, 1719–1725.
- 8 Z. L. Chen, B. Dai, W. C. Zhang, *et al.*, *RSC Adv.*, 2016, **6**, 96908.
- 9 N. Habibul, M. Ilmurat, Z. Habibul, *et al.*, *Chemosphere*, 2020, **242**, 125228.
- 10 T. Liu, L. S. Zhu, J. H. Wang, *et al.*, *Sci. Rep.*, 2015, **5**, 18444.
- 11 H. Wang, S. V. Malhotra and A. J. Francis, *Chemosphere*, 2011, **82**, 1597–1603.
- 12 Z. L. Chen, Q. Zhou, F. Leng, *et al.*, *Environmental Engineering Research*, 2017, **22**, 281–287.
- 13 D. J. Couling, R. J. Beront, K. M. Docherty, *et al.*, *Green Chem.*, 2006, **8**, 82–90.
- 14 Y. L. Xia, D. D. Liu, Y. Dong, *et al.*, *Chemosphere*, 2018, **195**, 437–447.
- 15 K. Cook, K. Tarnawsky, A. J. Swinton, *et al.*, *Biomolecules*, 2019, **9**(6), 251.
- 16 Z. L. Chen, Q. Zhou, W. Guan, *et al.*, *Chemosphere*, 2018, **194**, 20–27.
- 17 D. R. Hoagland and D. I. Arnon, *Calif. Agric. Exp. Stn., Circ.*, 1937, **347**, 357–359.
- 18 X. Z. Zhang, *Method of crop physiology research*, Agricultural Press, Beijing, 1992, pp. 74–77.
- 19 H. J. Liu and M. Y. Xiong, *Aquat. Toxicol.*, 2009, **93**, 100–106.
- 20 J. B. Fang, R. L. Pang, L. L. Guo, *et al.*, *Agricultural trade standards of the People's Republic of China*, NY/T 2017–2011, pp. 1–5.
- 21 H. J. Liu, S. X. Zhang, X. N. Hu, *et al.*, *Environ. Pollut.*, 2013, **181**, 242–249.
- 22 N. Kaur, M. Fischer, S. Kumar, *et al.*, *J. Colloid Interface Sci.*, 2021, **581**, 954–963.
- 23 R. Biczak, M. Śnioszek, A. Telesiński, *et al.*, *Ecotoxicol. Environ. Saf.*, 2017, **139**, 463–471.
- 24 J. J. Zhu, Q. Kong, S. S. Zheng, *et al.*, *Int. J. Biol. Macromol.*, 2020, **158**, 800–810.
- 25 Y. Y. Zhao, Z. H. Lu, L. He, *et al.*, *Appl. Biochem. Biotechnol.*, 2014, **173**, 1680–1691.
- 26 H. J. Liu, Y. L. Xia, H. Y. Fan, *et al.*, *J. Hazard. Mater.*, 2018, **358**, 327–336.
- 27 S. C. Yan, Y. Wu, J. L. Fan, *et al.*, *Field Crops Research*, 2020, **250**, 107767.
- 28 L. Nielsen, M. A. Brock, K. Crossl, *et al.*, *Freshwater Biol.*, 2003, **48**(12), 2214–2223.
- 29 M. L. Song, Q. Chai, X. Z. Li, *et al.*, *Plant Soil*, 2015, **387**, 153–165.
- 30 M. A. Preston, R. K. Bhaduri and A. Klein, *Phys. Today*, 1976, **29**, 56–57.
- 31 D. Trachootham, W. Zhang and P. Huang, *Cancer Cell*, 2009, **7**, 137–175.

