REVIEW

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Recent progress in chemical approaches for the development of novel neuraminidase inhibitors

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Influenza virus is the main cause of an infectious disease called influenza affecting the respiratory system including the throat, nose and lungs. Neuraminidase inhibitors are reagents used to block the enzyme called neuraminidase to prevent the influenza infection from spreading. Neuraminidase inhibitors are widely used in the treatment of influenza infection, but still there is a need to develop more potent agents for the more effective treatment of influenza. Complications of the influenza disease lead to death, and one of these complications is drug resistance; hence, there is an urgent need to develop more effective agents. This review focuses on the recent advances in chemical synthesis pathways used for the development of new neuraminidase agents along with the medicinal aspects of chemically modified molecules, including the structure–activity relationship, which provides further rational designs of more active small molecules.

1. Introduction

A highly infectious disease called influenza or flu is caused by influenza virus, and has symptoms such as fever, headache, muscle pains and runny nose, causing 250 000 to 500 000 deaths.[1] Importantly, there are three types of flu: A and B viruses are responsible for annual influenza epidemics, whereas C virus has less severe symptoms. The two classes of obtainable antiviral agents involve M2 inhibitors, which have potency only against virus A. Two classes of obtainable antiviral agents are involving the M2 inhibitors which have only potency against influenza A virus but with side effect including short time of effeteness and viral strain resistance[2–7] and the neuraminidase inhibitors (NAIs) which have potency against influenza A and B viruses.[8] Influenza is an enveloped RNA virus, which is a highly contagious pathogen worldwide and affects millions of people, animals and birds each year in seasonal epidemics.[9,10] Neuraminidase inhibitors are drugs that inactivate viral neuraminidase (NA) protein. Blocking sialic acid receptors and inhibiting virus–host cell interactions have been the best possible way to control and prevent the infection. This stops the release of viruses and prevents new host cells from being infected.[11] Therefore, inactivating NA using neuraminidase inhibitors offers the best possible method to inhibit sialic acid receptors. Oseltamivir and zanamivir are NA inhibitors, which inhibit all the subtypes of neuraminidase enzymes, and therefore, are effective against influenza viruses A and B. The role of neuraminidase inhibitors is important in the treatment of influenza, which act *via* blocking the release of viruses from the infected host cell and prevent the new host cells from infection. The role of antiviral agents is to block the neuraminidase enzymes from being more effective against A and B viruses. Neuraminidase inhibitors act by preventing the cleavage of sialic acid by a glycoprotein enzyme called neuraminidase and inhibiting

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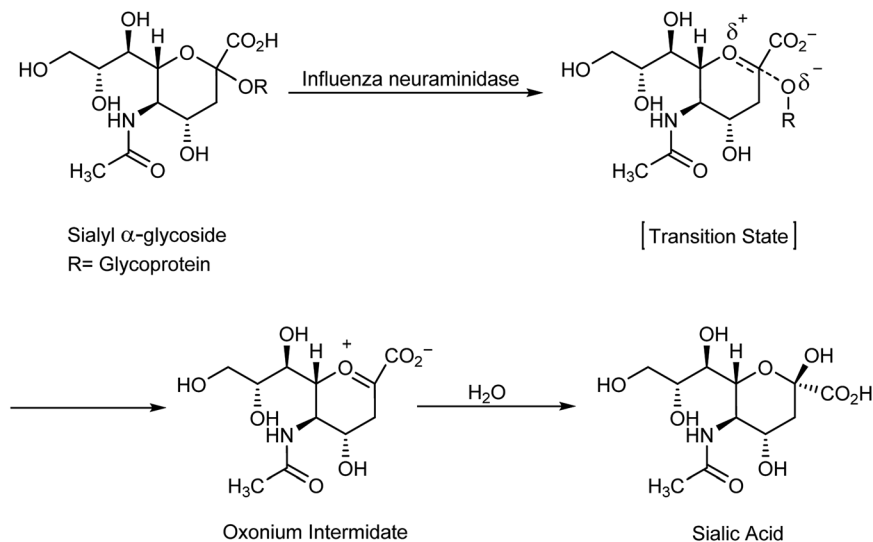
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Scheme 1 Hydrolysis step of sialylglycosides.

the distribution of viruses to neighboring cells[12,13] (Scheme 1).

The antiviral agents are considered to be alternative choices of the use of vaccines since there are many disadvantages including the time of production storage and effective time limitation. There are many approved neuraminidase drugs in the market worldwide including 2-deoxy-2,3-dehydro-*N*-acetylneuraminic acid (DANA), zanamivir (Relenza),[14] oseltamivir phosphate (Tamiflu),[15,16] lani-namivir[17] and peramivir,[18] as shown in Fig. 1.

Zanamivir has low bioavailability and can be administrated by inhalation and was first synthesized from methyl 5-acet-amido-4,7,8,9-tetra-*O*-acetyl-2,6-anhydro-3,5-dideoxy-D-*glycero*-D-galacto-non-2-enonate (**1**),[19–22] *N*-acetylneuraminic acid (**2**, NANA),[23] *N*-acetylneuraminic acid **2** (ref. 24) and D-glucono-δ-lactone **3** (ref. 25) (Fig. 2).

Oseltamivir phosphate is an oral active prodrug that requires ester hydrolysis to form active oseltamivir carboxylate as an inhibitor against influenza virus. Many strategies were described for the synthesis of this inhibitor including Gilead starting from (−)-shikimic acid (**4**)[26] and (−)-quinic acid (**5**),[27] F. Hoffmann-La Roche Ltd starting from (−)-shikimic acid (**4**) and (−)-quinic acid (**5**),[28] epoxide (**6**),[29] furan (**7**) and ethyl acrylate (**8**),[30] 2,6-dimethoxyphenol (**9**) and mesylate (**10**),[30] Roche Colorado Corporation starting from epoxide (**6**),[31] Corey starting from 1,3-butadiene (**11**) and 2,2,2-trifluoroethyl acrylate (**12**),[32] Okamura starting from *N*-nosyl-3-hydroxy-2-pyridone (**13**) and ethyl acrylate (**14**),[33] Shibasaki starting form *N*-3,5-dini-trobenzoylaziridine (**15**),[34,35] azide (**16**),[35] 1-(trimethylsiloxy)-1,3-butadiene (**17**) and fumaryl chloride (**18**),[36] 1-(trimethylsiloxy)-1,3-butadiene (**17**) and dimethyl fumarate (**19**),[37] Yao starting
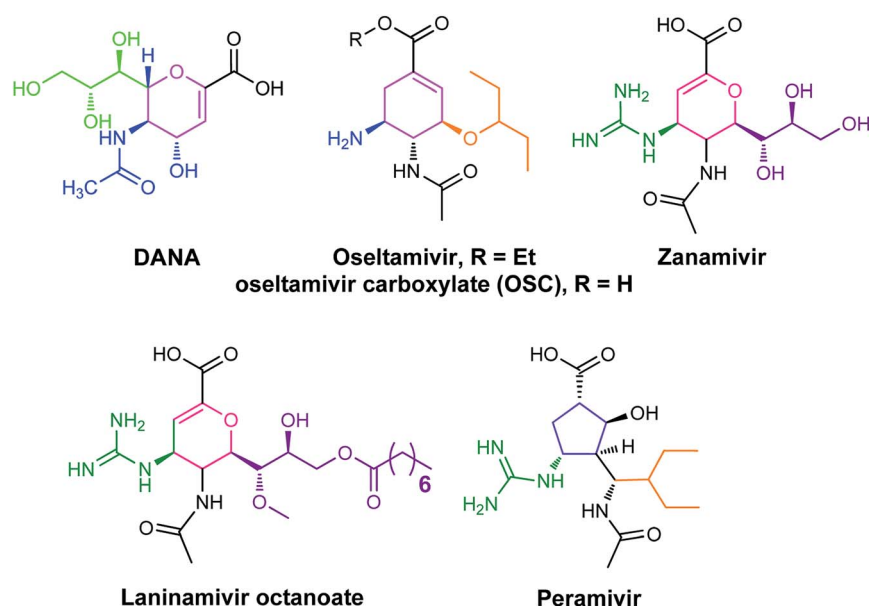

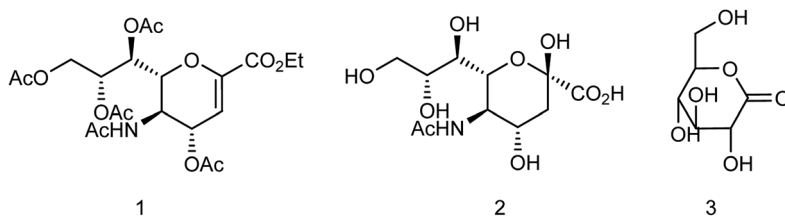**Fig. 1** DANA, zanamivir and oseltamivir phosphate structures.

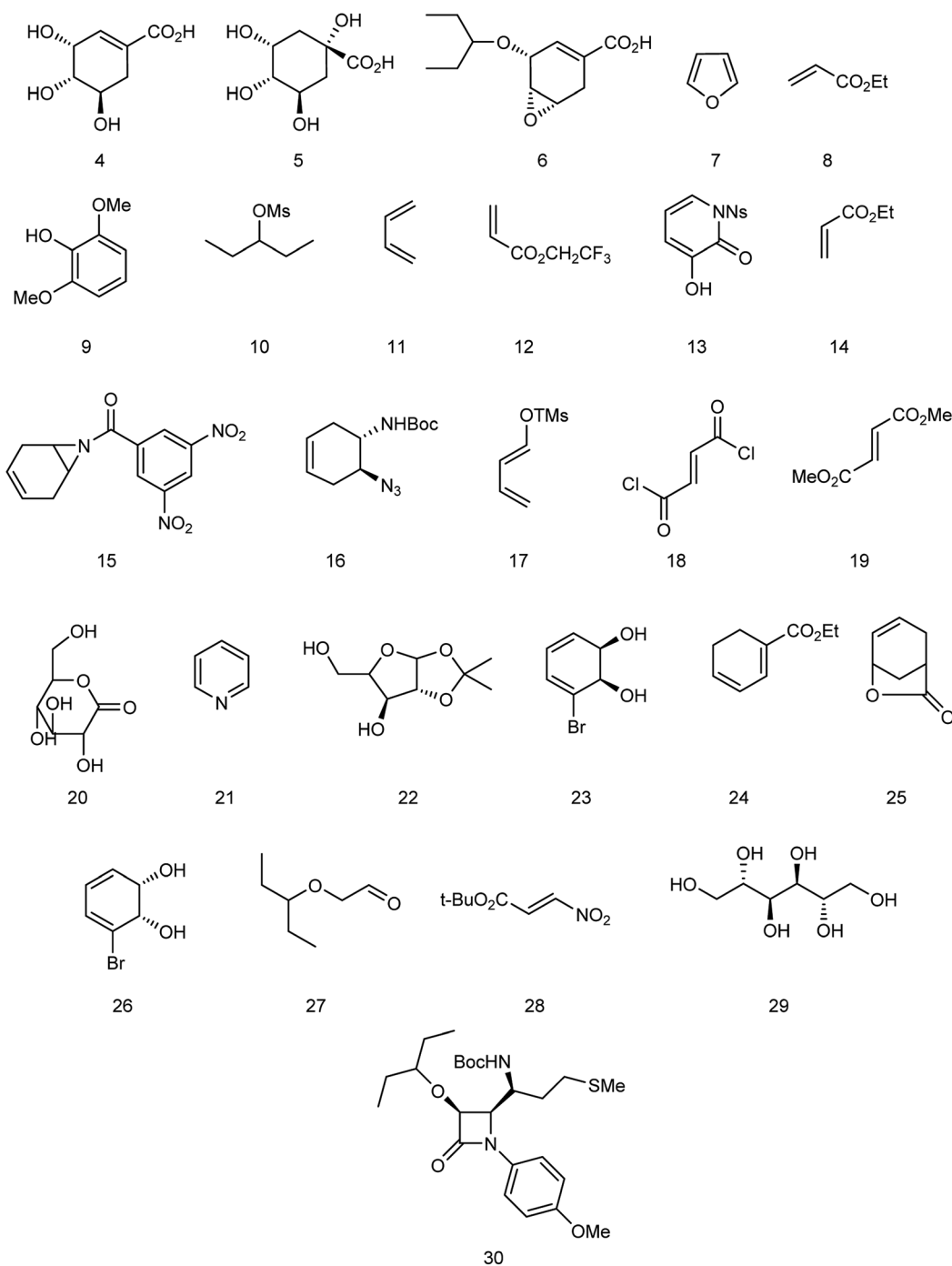**Fig. 2** Structures of starting materials for the synthesis of zanamivir.

Fig. 3 Structures of starting materials for the synthesis of oseltamivir phosphate.
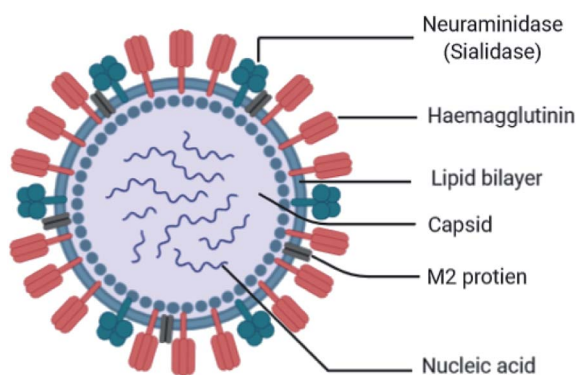
Fig. 4 Structure of influenza virus.

from D-glucono-δ-lactone (**20**),[38] Fukuyama starting from pyridine (**21**),[39,40] Fang starting from D-xylose (**22**)[41] and *cis*-1,2-dihydrodiol (**23**),[42] Kann starting from cyclohexadienoic acid ethyl ester (**24**),[43] Trost starting from lactone (**25**),[44] Banwell starting from *cis*-1,2-dihydrodiol (**26**),[45] Shi starting from (−)-shikimic acid (**4**),[46,47] Hayashi starting from aldehyde (**27**) and nitroalkene (**28**),[48] Mandai starting from D-mannitol (**29**)[49] and L-methionine derivative (**30**)[50] (Fig. 3).

2. Types of influenza virus

Influenza is an RNA virus that exists mostly in spherical shape 80–120 nm in diameter. They belong to the Orthomyxoviridae family of viruses, which contain a negative-stranded RNA genome made up of eight RNA segments that encode ten viral

Table 1 Types of influenza virus

Types	Name	Year of finding	Origin	Key hosts	Infection type	Ref.
XType-A (8 RNA segments) (antigenic shift)	H1N1	1918, 1976 (swine), 1977 (Russian flu), 2009	Spanish flu, Alberta, Canada	Mutated avian influenza virus, duck	Most common, most severe and can mutate Highly pathogenic Infects humans and animals	51–54
	H2N2	1957 Asian flu	Singapore	Birds and pigs	Pathogenic to humans	54 and 57
	H3N2	1968	America	Human	Most common, most severe and can mutate	54
	H3N2v	2011	America	Pigs	Infects humans and animals	
	H5N1	1997 (avian) (bird flu)	China	Geese, poultry	Pathogenic to humans	54
	H5N5	2008	China	Ducks, poultry	Zoonotic	56 and 58
	H5N8	2010	China	Poultry	Zoonotic	56
	H7N7	2003	Europe	Poultry	Zoonotic	54 and 59
	H7N9	2013 (avian)	China novel avian influenza a ducks, poultry, and wild birds	Poultry	Pathogenic to humans Humans and birds	55
	H7N3	2003 (bird flu)	Taiwan, British Columbia, North America	Migratory birds, poultry	Zoonotic	60
	H9N2	1966 (bird flu)	America, Turkey	Chicken, duck, pigeon	Potentially pathogenic to humans	61
	H10N8	2012 (bird flu)	China	Ducks	Potentially pathogenic to humans	62
Type-B (8 RNA segments) (antigenic drift)	Have HA and NA, two types Yamagata and Victoria	1988	Yamagata, Japan	Humans	Less common and doesn't mutate as often Infects only humans	63
Type-C (7 RNA segments) (antigenic drift)	No HA, and NA, but have hemagglutinin-esterase fusion (HEF) glycoprotein	1982	Sao Paulo, Brazil	Pigs, cows	Least common and least likely to mutate Infects children and pigs	64
Type-D (7 RNA segments) (antigenic drift)	No HA, and NA, but have hemagglutinin-esterase fusion (HEF) glycoprotein	2011	America	Pigs, cattle	No human infections reported	65

proteins. It is an enveloped virus (lipoprotein of host). The envelope contains two different varieties of glycoproteins: HA (hemagglutinin attachment to the host) and NA (neuraminidase for release, Fig. 4).

These are the main exposed segments of the virus responsible for virus entry. Inside the envelope, the matrix protein is present, which gives additional support to the envelope. Inside the matrix, the influenza genome is encapsidated. The influenza genome consists of 8 strands of RNA. These are the genetic materials combined with the capsid called nucleocapsid. The doted mark is known as RNA polymerase, which replicates RNA. The virus can survive on surfaces for few hours. It may enter into the body through mouth and nose by inhaling the droplets containing viruses from the sneeze and cough of an infected person within a range of 6 ft. When it enters the body, it binds sialic acid sugars of epithelial cells of the upper respiratory tract using hemagglutinin.

The influenza viruses are classified into four types (A, B, C, and D) based on the host predilections, ability to undergo genetic reassortment, and the severity of the disease they cause.[51] Influenza A virus has several variants based on the presence of different HA (1–18) and NA (1–11) types. The virus undergoes major genetic variations (antigenic shift) and adapts itself in different hosts including humans, pigs, horses, birds, reptiles, and others resulting in pandemics. The influenza type B viruses are exclusively seen in humans and cause mild to moderate illness. They cause seasonal diseases, undergo minor genetic changes (antigenic drift) and are responsible for epidemic flu-like illnesses. The type C influenza viruses usually cause infections in pigs. They are not known to undergo genetic changes and are rarely reported in humans. Various types of influenza viruses and their infection types are listed in Table 1 for better understanding.[51–65]

3. Symptoms

The common symptoms are acute-onset fever, headache, runny nose, sore throat, and dry cough. Other symptoms noted during the flu include myalgia, fatigue/weakness, vomiting, and diarrhea. The incubation period ranges from 1 to 4 days and uncomplicated infection resolves within 1 week. The complications are acute otitis media, bronchiolitis, croup, sinusitis, pneumonia, myocarditis, and secondary microbial infections. Young children, pregnant women, adults above 65 years and persons having heart or lung disease are at higher risk of developing complications.[66–68]

4. Diagnosis

Influenza viral infections are diagnosed based on the clinical symptoms as discussed earlier. Virus can be cultured from nasal, nasopharyngeal, and throat swabs taken from the infected and suspected persons. Laboratory diagnosis by enzyme-linked immunosorbent assay (ELISA) can be performed for detecting the IgM antibodies in acute infections, and IgG antibodies from the convalescent persons. Diagnosis of flu can also be done by the complement fixation test, hemagglutination, and hemagglutination inhibition tests. Rapid influenza

diagnostic tests (detect type does not strain) and polymerase chain reaction (detects viral RNA) are preferred for preliminary diagnosis and confirmation of the disease, respectively.[69–71]

5. Mechanism

5.1. Virus entry mechanism

The influenza viruses adhere to the host cells with the help of spike-like projections protruding from the viral envelope called HA (Fig. 5A). Later, the viruses identify appropriate sites on the host cells, known as *N*-acetylneuraminic acid (sialic acid) residue-containing areas to initiate binding/attachment. The terminal α-sialic acid residues are oligosaccharide chains of N- and O-linked glycoproteins and lipids, which bind to the underlying galactose by a-2,3 or a-2,6 glycosidic linkages. This linkage and the resulting structural consequences influence how well the virus can bind to its receptor. After successfully attaching to the host cell, the virus enters the cell by using the clathrin-dependent endocytosis or the micropinocytosis. The host factors like the C-type lectins, Annexin V, 6-Sulfo sialyl Lewis X, dynamin, actin, clathrin, espin-1, EGFR, c-met kinase, and PLC-γ1 facilitate the successful attachment, and internalization of virus to the host cell. After entering into the host cell, the endosome fuses with the viral HA and increases the pH (acidity) within the virion. This causes release of the viral nucleic acid into the host cell cytoplasm. The host cell factors including Rab5, Rab7, PKC βII, Cullin 3, HDAC8, vATPase, CD81, and ITCH facilitate the endosomal trafficking, acidification, fusion, and uncoating of the viral nucleic acid. The viral nucleic acid then uses the host cell nuclear import pathways and importin-α, and importin-β proteins, to enter the nucleosome. The host factors that contribute to the import of viral RNA into the nucleosome include karyopherin (α1; α3; α5), Ran, p 10, and CSE1.[72,73]

5.2. Neuraminidase inhibition mechanism

Because of the significance of sialic acid receptors in the attachment and entry of viruses into the host cells, blocking these receptors and inhibiting virus–host cell interactions have been the best possible way to control and prevent the infection.
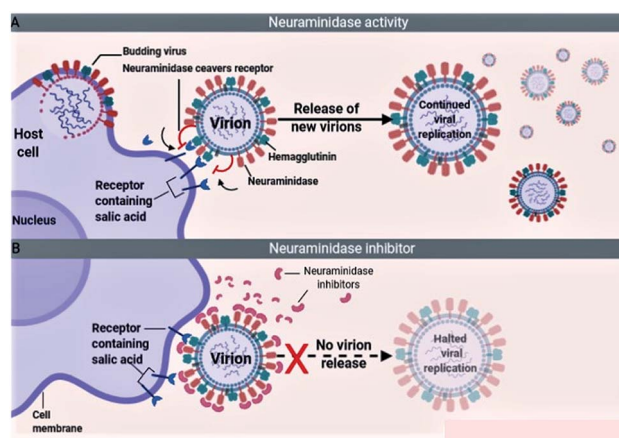

Fig. 5 Mechanism of virus entry (A) and neuraminidase inhibitors (B).

For the virus to successfully attach to the host cell, sialic acid must first be cleaved to expose the site of attachment for the virus. This function is performed by the NA proteins present on the virus envelope as projections similar to HAs. Therefore, inactivating NA using the neuraminidase inhibitors offers the best possible method to inhibit sialic acid receptors (Fig. 5B). The most efficient drugs in the treatment of flu caused by influenza A and pathogenic influenza B virus including oseltamivir and zanamivir are NA inhibitors.[74–76]

6. Neuraminidase inhibitors (NAIs)

6.1. Synthetic neuraminidase inhibitors

Several aryl and alkyl substituted 3-hydroxypyridin(1H)-2-ones were studied by LaVoie and co-workers[77] for their ability to inhibit IAV endonuclease activity. Their endonuclease inhibitory activities are provided. The effect of *para*-substituents on the phenyl group substituted at position 5 was assessed, which indicates the importance of having an acidic functional group at this site. A number of 2,3-difluorosialic acids and their derivatives (Fig. 6) were synthesized chemoenzymatically by Chen and

co-workers.[78] PmAldolase was found to catalyze the formation of a C5-azido analogue of 3-fluoro(a)-sialic acid, whereas its *Escherichia coli* homologue (EcAldolase) did not. Both EcAldolase and PmAldolase could catalyze the synthesis of 3-fluoro(a/e)-sialic acids and their C-9 analogues, but PmAldolase was more efficient.

Ma and co-workers[79] have designed a dual-functional luminescent probe and NA inhibitor, a key influenza target (Fig. 7). The lead iridium(III) compound displayed appreciated inhibition against NA compared to oseltamivir (FDA-approved drug), and could detect NA even in the presence of an auto-fluorescent background.

Hu and co-workers[80] have designed twenty-seven novel chalcone derivatives (Fig. 8), and their inhibitory activities were evaluated against NA of influenza A virus *in vitro*. As per the study, the derivatives having a pyran ring displayed better NA inhibitory activity than those without a pyran ring. The molecular docking studies revealed that some compounds are in the good binding mode with zanamivir binding sites. Moreover, the structure–activity relationship was also reported.

Hu and co-workers[81] have evaluated the neuraminidase (NA) inhibitory activity of a number of thiazolylhydrazone derivatives
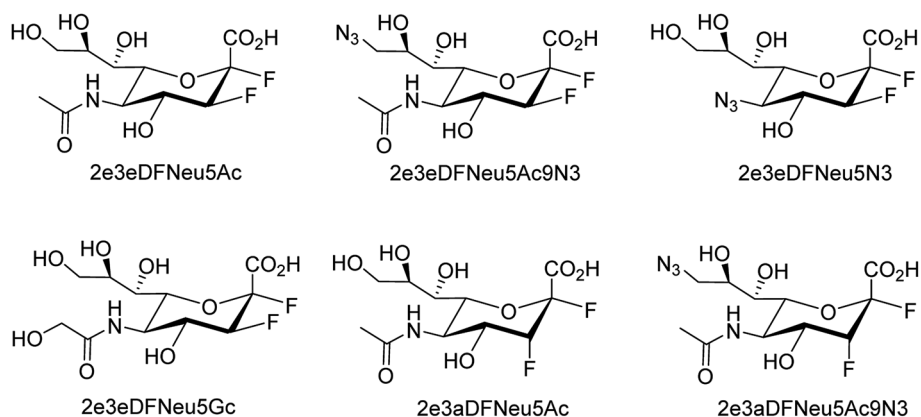

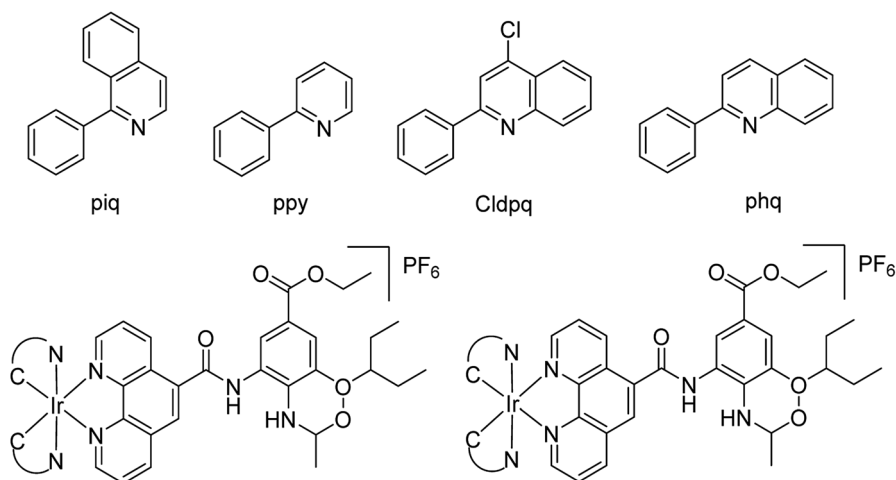**Fig. 6** Structures of 2,3-difluorosialic acid analogues.

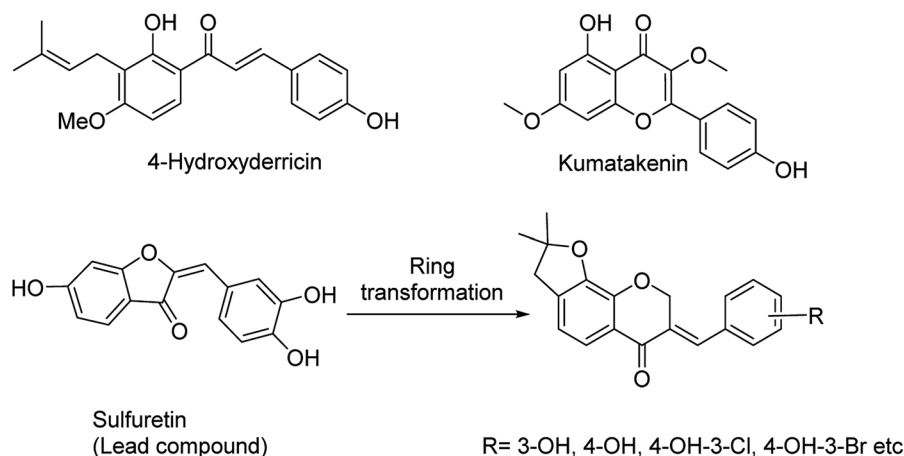**Fig. 7** Structures of iridium(III) complexes.

Fig. 8 Structures of chalcone derivatives.

(Fig. 9) against H1N1 (influenza virus) *in vitro*. Most of the compounds exhibited moderate-to-good inhibitory activity. The NA inhibitory activity of the most active compound was found to be $IC_{50} = 7.12$ μg mL^{-1}. Furthermore, a molecular docking study was performed to understand the possible interactions between the compound and the active site of NA. In addition, DFT calculations and SAR were derived and discussed.

hPIV causes acute respiratory tract infections and becomes life-threatening to young children and immunocompromised individuals. As there are no anti-hPIV drugs, it is important to develop specific antiviral therapies to decrease the mortality. HN is the hPIV surface protein playing critical roles in binding and cleavage. von Itzstein and co-workers[82] have discussed the structural features of the HN protein that are being exploited for structure-guided inhibitor designs (Fig. 10), which will be very helpful for antiviral drug design.

NA is used for the treatment of influenza A and B infections, as it plays a vital role in the replication and transmission of the virus. Wu and co-workers[83] have discovered a new lead neuraminidase inhibitor by receptor-based virtual screening, ligand-based virtual screening, MD simulation, and bioassay validation. A series of acylhydrazone NA inhibitors (Fig. 11) have also

been synthesized based on the lead compound. The compound 6e exerted $IC_{50} = 2.37 \pm 0.5$ mM (lower than OC) against NA.

Yang and co-workers[84] have prepared a series of divalent guanidino oseltamivir and oseltamivir analogues *via* a click reaction (Fig. 12). The SAR study suggested an appropriate distance between two oseltamivir monomers, which result in highly effective NA inhibitors against three strains of influenza virus. This study provides a basis for the multivalent modification on oseltamivir.

The combined therapy improves the treatment outcomes and reduces the emergence of drug-resistant variants. Kiso and co-workers[85] have infected immunocompromised nude mice with influenza A virus and treated them with viral polymerase (favipiravir) and/or neuraminidase (oseltamivir and laninamivir) inhibitors. The combination therapy (for 28 days) increased the survival times compared with monotherapy, but the mice died after the treatment was terminated. This study explained that combination therapy increases survival times in immunocompromised hosts, but does not suppress the emergence of neuraminidase inhibitor-resistant variants. The designing of novel neuraminidase inhibitors is very essential, which may withstand the challenges of resistance. Malbari and co-
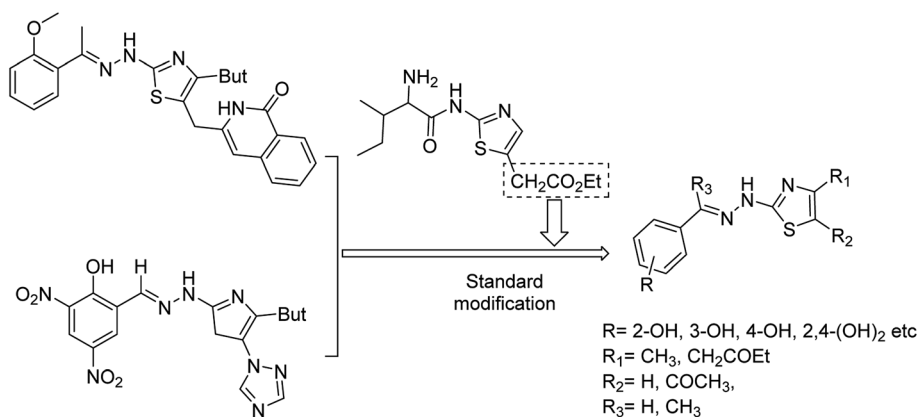

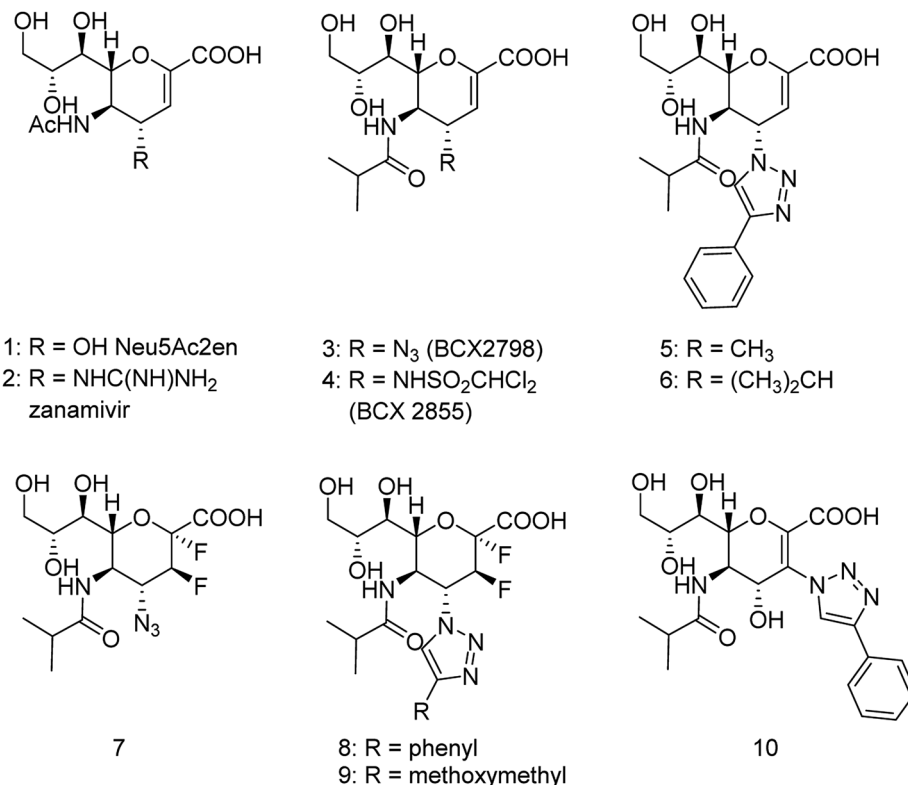**Fig. 9** Design and synthesis of 2-thiazolylhydrazone derivatives.

1: R = OH Neu5Ac2en
2: R = NHC(NH)NH₂ zanamivir

3: R = N₃ (BCX2798)
4: R = NHSO₂CHCl₂ (BCX 2855)

5: R = CH₃
6: R = (CH₃)₂CH

7

8: R = phenyl
9: R = methoxymethyl

10

Fig. 10 Structures of Neu5Ac2en-derived hPIV-3 HN inhibitors.

6a: R = 2-methoxy
6b: R = 3,4,5-trimethoxy
6c: R = 3-nitryl-4-hydroxyl

6d: R = 2-hydroxy
6e: R = 3,4-dimethoxy
6f: R = 3-methoxy-4-hydroxyl
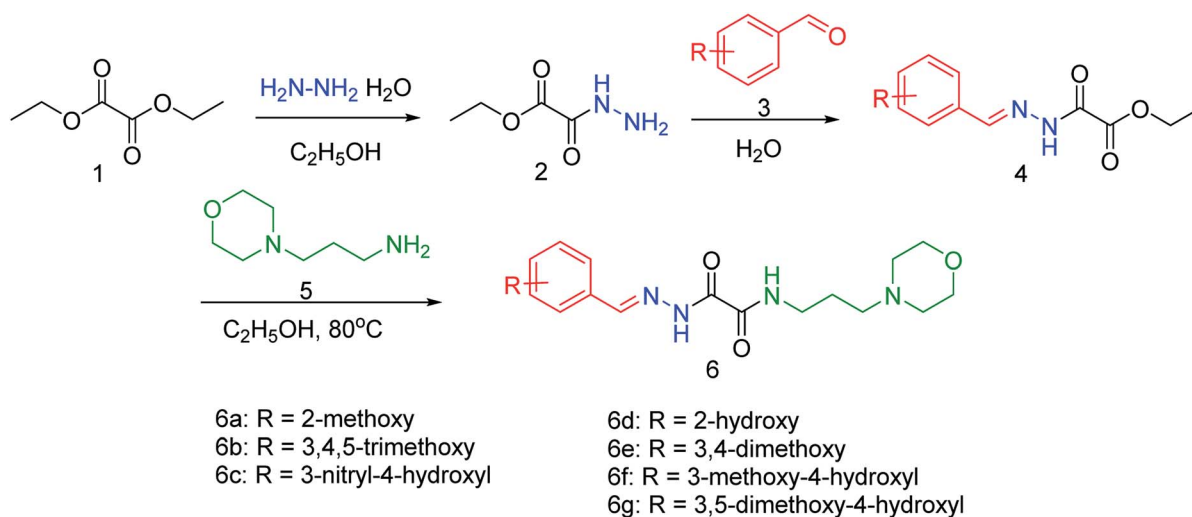6g: R = 3,5-dimethoxy-4-hydroxyl

Fig. 11 Synthesis of acylhydrazone NA inhibitors.

workers[86] have designed (Fig. 13) five series of scaffolds (aurones, chalcones, cinnamic acid analogues, pyrimidine analogues and cinnamic acid linkages) based on virtual screening against H1N1 virus. The molecular modelling studies revealed that the reported compounds occupied 430-loop cavity of neuraminidase. The favourable compounds have synthesized and evaluated for their cytotoxicity and cytopathic effect inhibition by H1N1 virus. The study indicated that aurones will be treated as potential neuraminidase inhibitors against the pandemic H1N1 virus.

Prasath and co-workers[87] have analysed the anti-influenza agent LGN using FT-Raman and FT-IR spectra. The experimental results were compared with DFT calculations, which are in good agreement with the computational one. The suitability of a drug candidate was evaluated for human intake by ADMET properties. According to the study, LGN molecules exhibited
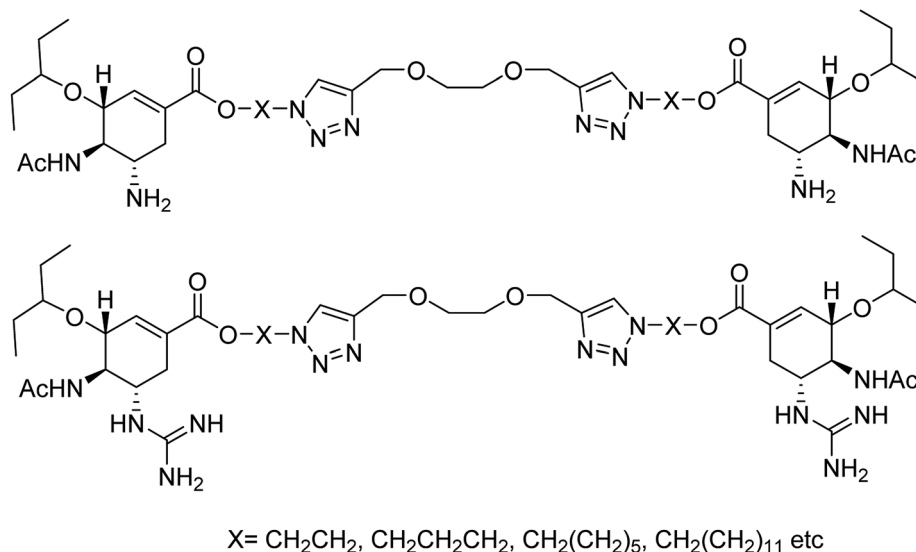
Fig. 12   Structures of divalent oseltamivir analogues.

X= CH₂CH₂, CH₂CH₂CH₂, CH₂(CH₂)₅, CH₂(CH₂)₁₁ etc

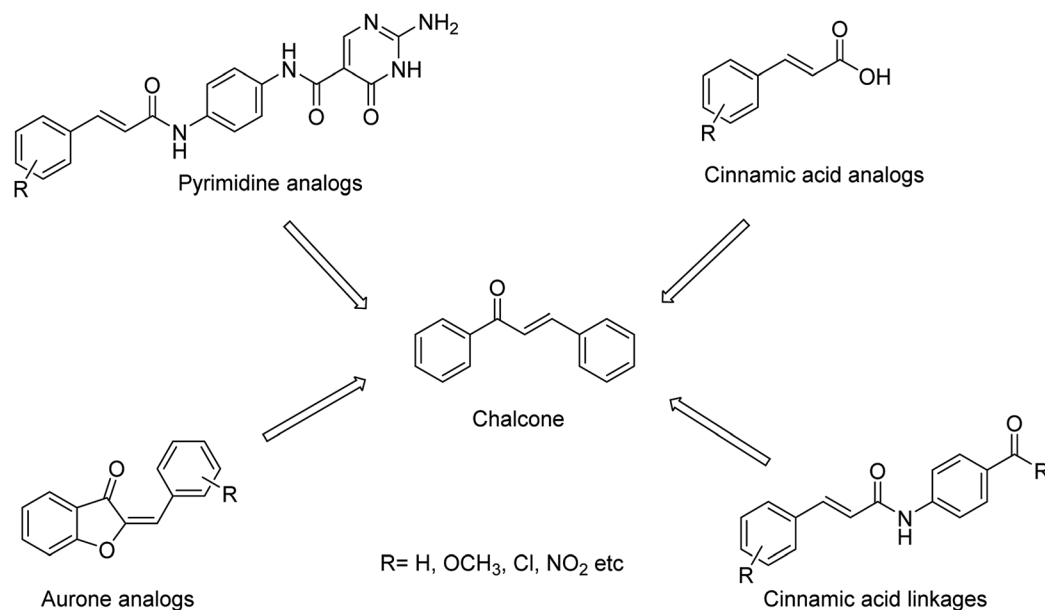X= CH_2CH_2, $CH_2CH_2CH_2$, $CH_2(CH_2)_5$, $CH_2(CH_2)_{11}$ etc

Fig. 13   Structures of aurones, chalcones, cinnamic acid analogues, pyrimidine analogues and cinnamic acid linkages.

good bioactive score and less toxicity. Moreover, molecular docking study was carried out to determine the binding affinity in the active site. The correct configuration in sialosides at C2 is a major challenge due to the absence of anomeric hydrogen. Therefore, the correct configuration of 3,4-unsaturated derivatives of Neu5Ac could be assigned as per the reported 3,4-unsaturated 2O-methyl sialosides. In this context, Anastasia and co-workers[88] have tried to resolve this issue through the synthesis of the corresponding unreported unsaturated 1,7-lactones (Fig. 14).

Ebaraa and co-workers[89] have synthesized (Fig. 15) and modified NA-resistant sialoside (having unnatural S-glycoside bonds) on a 3WJ DNA to display complementary distribution to its binding sites on a HA trimer. This modified sialoside maintained high binding affinity and displayed certain NA resistance. Most importantly, this study may help in the development of NA-resistant sialoside derivatives for a broad spectrum of viruses.
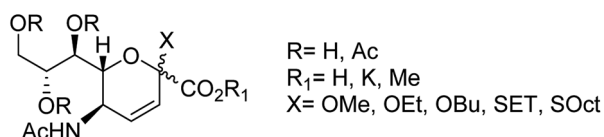

R= H, Ac
R₁= H, K, Me
X= OMe, OEt, OBu, SET, SOct

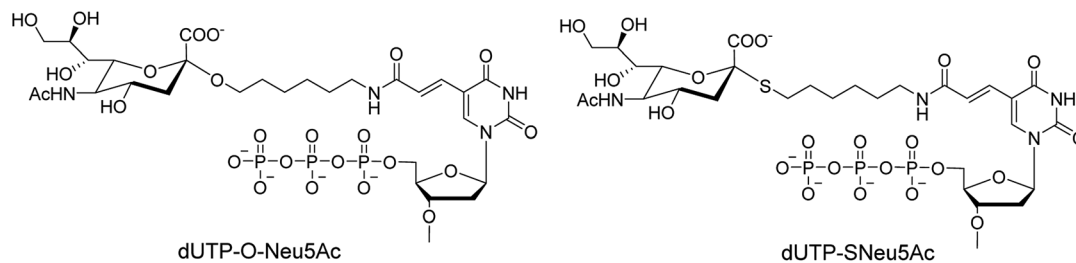Fig. 14   Structures of 3,4-unsaturated Neu5Ac derivatives.

Fig. 15 Structures of dUTP-ONeu5Ac and dUTP-SNeu5Ac derivatives.

We all know that influenza viruses are causing severe upper respiratory illness in humans. Moreover, the current IAV strains have developed resistance to neuraminidase inhibitors. Manicassamy and co-workers[90] have summarized the existing antiviral strategies for combating influenza viruses. This report may provide ample references for the treatment of influenza virus disease and the development of antiviral resistance. Fang and co-workers[91] have reported the anti-influenza and NA inhibition activity of some OC, GOC (by replacing 5-amino group with a guanidino group) and their carboxyl bioisosteres (Fig. 16). The GOC congeners showed better anti-influenza and NA inhibition activity than the corresponding OC congeners. The GOC sulfonic acid congener (EC_{50} = 2.2 nM) is the most potential influenza agent against the wild-type H1N1 virus.

Due to the potential therapeutic properties of calixarene derivatives, Hamid and co-workers[92] have reported the antibacterial and antiviral potentials of some azo-based derivatives of calix[4]arene (Fig. 17). The mono azo products of sulfaguanidine, sulphanilamide, and 2-methyl-4-aminobenzoic acid displayed good activity against some selected bacterial strains. The use of sodium acetate trihydrate and calix[4]arene in 3 : 1 molar ratio helped in partial substitution. Moreover, a molecular docking study was carried out to understand the interactions with bacterial and viral receptors.

Abdalsalam[93] has reported virtual screening and docking analysis of several compounds obtained from the ZINC database against the H1N13TI6 protein (Fig. 18) by using Autodock Vina.

Here, the exhibited data are compared with the co-crystallized drug oseltamivir. The novel compounds were successfully identified, which could serve as a potential lead compound for the development of a new anti-neuraminidase drug. Oseltamivir resistance against H1N1 influenza virus has been reported lately. Tambunan and co-workers[94] have reported the docking simulation of disulfide cyclic peptide ligands (DNY, LRL, NNT) (Fig. 19) along with zanamivir and oseltamivir as the standard ligands using the MOE 2008.10 software. The study showed that all disulfide cyclic peptide ligands have lower ΔG binding than the standard ligands, with the lowest for DNY (ΔG binding = −7.8544 kcal mol^{-1}). Furthermore, these ligands had better interactions with neuraminidase than the standards, hence may be used as potential neuraminidase H1N1 inhibitors.

The new strains of influenza virus, resistant to current inhibitors (oseltamivir, amantadine, *etc.*), are making serious threats to the public health. Hence, it is essential to develop better and more effective vaccines and therapeutics. With this in view, Yassine and co-workers[95] have designed a computational study to identify potential flavonoid inhibitors that bind to the contact epitopes of the HA stem that are targeted by bNAb. The QSAR study was performed for 100 natural compounds along with molecular docking analysis. According
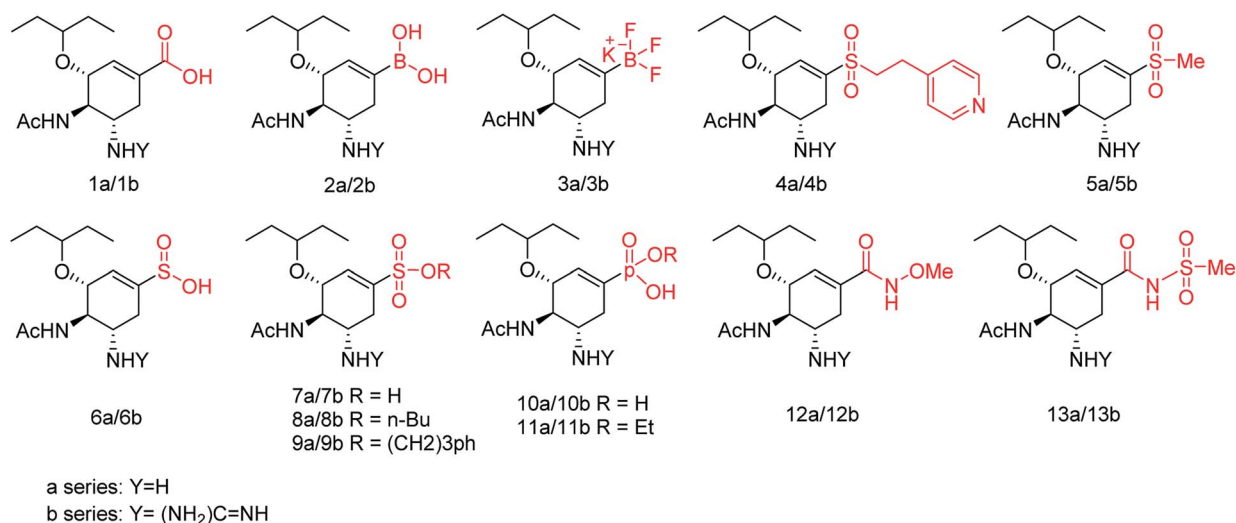

Fig. 16 Structures of oseltamivir carboxylic acid (**1a**, OC), guanidino OC (**1b**, GOC) and their carboxyl bioisosteres.
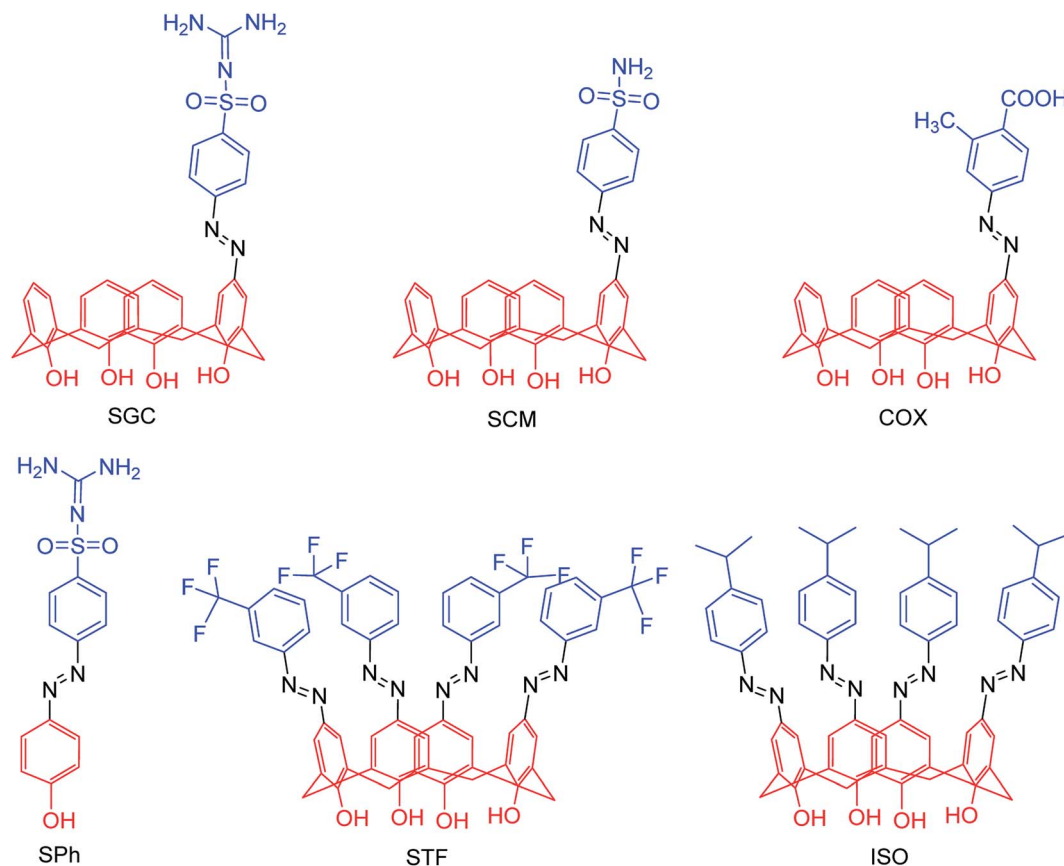
Fig. 17 Structures of azo derivatives.

to the study, 18 lead flavonoids have strong binding abilities to bNAb epitopes of various HA subtypes. Cairo and co-workers[96] have reported a library of modified DANA analogues (Fig. 20) and evaluated them against four human neuraminidase isoenzymes (NEU1-4). The best NEU1 inhibitor was C5-hexanamido–C9-acetamido–DANA having 340-fold selectivity ($K_i = 53 \pm 5$ nM). Moreover, it was demonstrated that C5-modifications

Fig. 18 Superimposition between the docked conformation (yellow) and the crystal structure (green) of H1N1 3TI6.

combined with a C4-guandino group provided the most potent NEU2 inhibitor.

Onul and co-workers[97] have reported the synthesis of S-substituted perhalo-2-nitrobuta-1,3-dienes (**3a,b**) from the reaction of allyl mercaptan with polyhalo-2-nitrobuta-1,3-dienes (**1a,b**). The addition of bromine to S-substituted polyhalo-2-nitrobuta-1,3-diene (**3b**) in CCl$_4$ yielded 1-(2,3-dibromopropanethio)-4-bromo-1,3,4-trichloro-2-nitrobuta-1,3-diene (**4**). The reaction of thiosubstituted polyhalonitrobutadienes (**3a,b**, and **4**) with m-CPBA in CHCl$_3$ yielded sulfoxides (**5a,b**, and **6**). The structures of these newly reported compounds were confirmed by ^{1}H NMR, ^{13}C NMR, FTIR and MS data. These reported compounds (Fig. 21) exhibited antixanthine oxidase, antielastase, antityrosinase, and anti-neuraminidase activities.

To overcome the drug–resistance activity, the biological nanoparticles are playing significant rules in the development of novel anti-influenza drugs. In this context, selenium nanoparticles with low toxicity have attracted increasing attention for biomedical intervention. Zhu and co-workers[98] have designed the surface decoration of selenium NPs by amantadine. Se@AM has less toxicity and inhibited the ability of H1N1 to infect host cells through suppression of neuraminidase activity. Moreover, Se@AM could prevent H1N1 from infecting the MDCK cell line. In another work, Zhu and co-workers[99] have also reported the synthesis of oseltamivir (OTV) surface-modified SeNPs with superior antiviral properties. Se@OTV had less toxicity and inhibited H1N1 infection. Se@OTV interfered with H1N1 virus
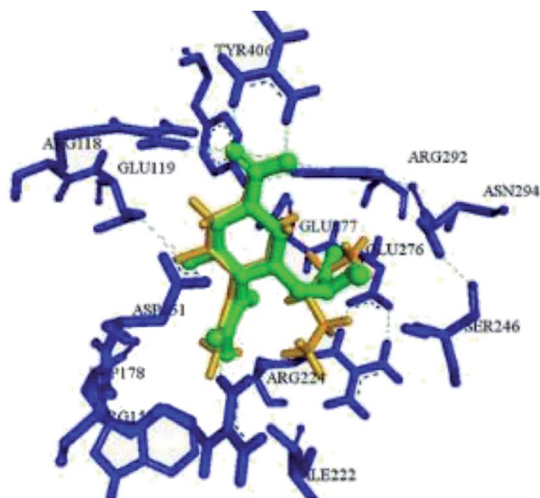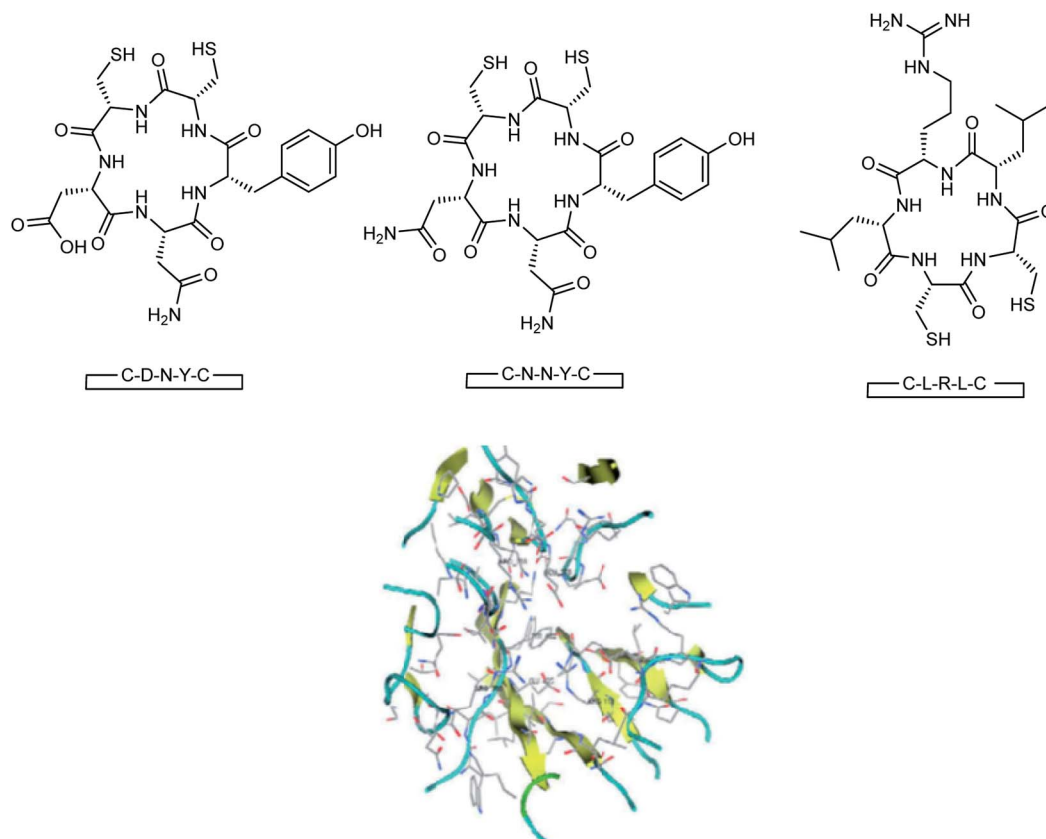
Fig. 19 Structures of cyclic peptides and the visualization of the catalytic site of influenza A (H1N1) neuraminidase.
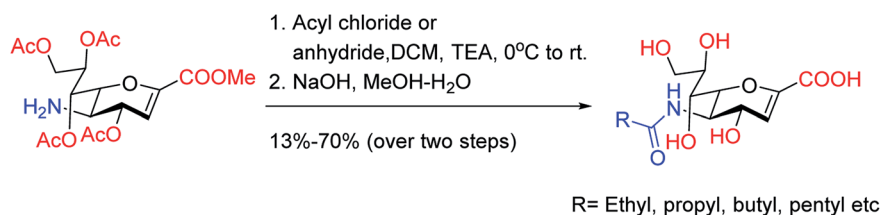

Fig. 20 Synthesis of C5-modified DANA analogues.

to host cells through suppression of neuraminidase and hemagglutinin activity. The study demonstrated that Se@OTV is a promising efficient antiviral pharmaceutical for H1N1. Cheng and co-workers[100] have studied a series of zanamivir derivatives as potential NA inhibitors by using molecular docking, 3D-QSAR, and MD simulation techniques. Out of which, two selected compounds were synthesized. Both the compounds were biologically evaluated by virus inhibition and NA inhibition assays. One compound exhibited excellent anti-viral activity (IC$_{50}$ = 0.670 μM, SI > 149) against A/WSN/33H1N1, superior to zanamivir (IC$_{50}$ = 0.873 μM, SI > 115). This study may provide significant contribution for the development of new anti-influenza drugs. Sialidases and neuraminidases are becoming the main targets for drug development as they are playing important roles in bacterial and viral infections. Several derivatives of Neu5Ac2en (DANA) have been designed and synthesized by Chen and co-workers[101] (Fig. 22) as triazole-linked transition state analogs. The inhibition studies confirmed that E-(TriazoleNeu5Ac2en)-AKE and (TriazoleNeu5Ac2en)-A were selective inhibitors against *V. cholerae* sialidase, while (TriazoleNeu5Ac2en)-AdE selectively inhibited *V. cholerae* and *A. ureafaciens* sialidases.

On the basis of molecular docking study, Pratama and co-workers[102] have reported the relationship between artemisinins and neuraminidase (NA). The docking study was performed using AutoDock 4.2.3 toward NA in complexes with oseltamivir as a co-crystal ligand. Artesunate provided most negative free ΔG (−9.55 kcal mol^{-1}) and lowest K_i (100.66 nM) toward NA. Artesunate showed higher affinity than oseltamivir with inter-actions between artesunate and amino acids at position 246, which had important influences on artesunate affinity toward NA from H5N1. The study indicated that artesunate could be
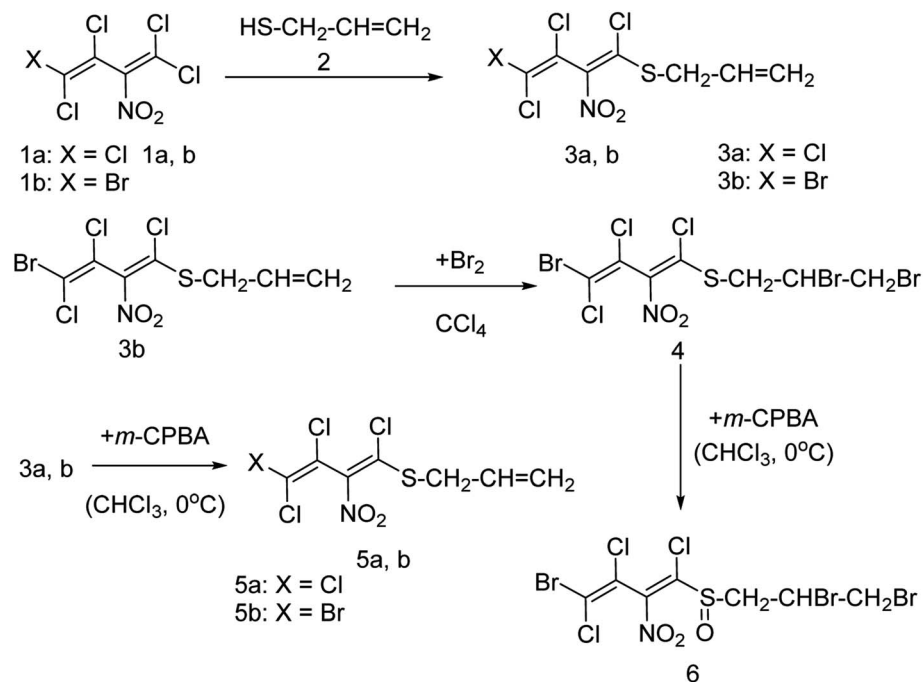
Fig. 21 Synthesis of S-substituted perhalonitrobuta-1,3-diene derivatives.

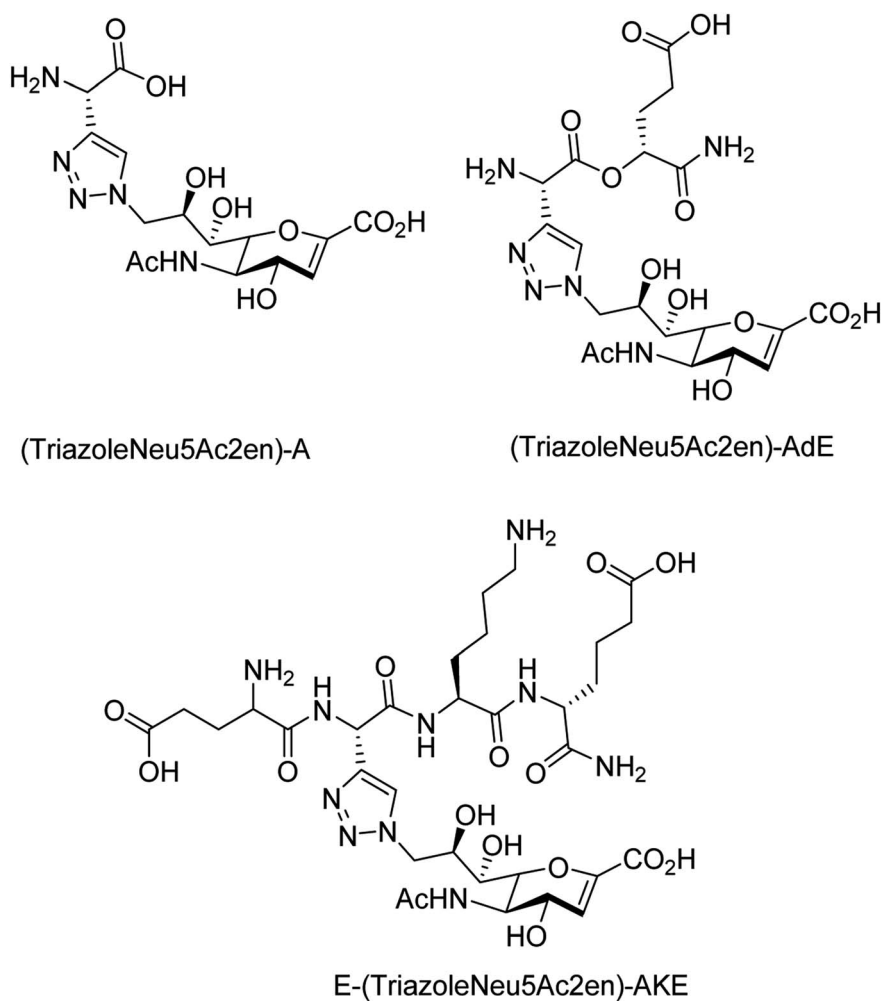

(TriazoleNeu5Ac2en)-A

(TriazoleNeu5Ac2en)-AdE

E-(TriazoleNeu5Ac2en)-AKE

Fig. 22 Structures of (TriazoleNeu5Ac2en)-A, (TriazoleNeu5Ac2en)-AdE and E-(TriazoleNeu5Ac2en)-AKE.

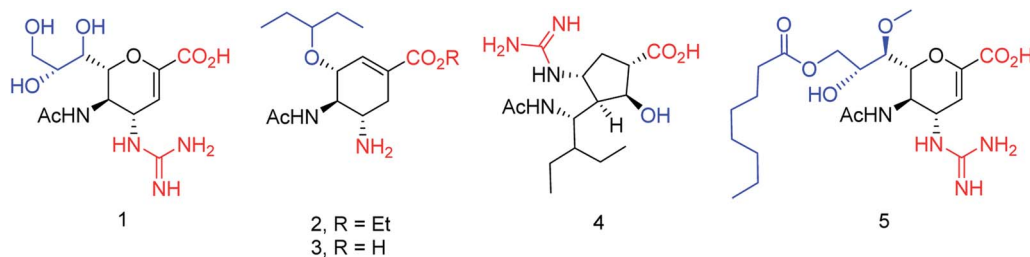© 2021 The Author(s). Published by the Royal Society of Chemistry

Fig. 23 Structures of carboxyl-modified oseltamivir analogues.

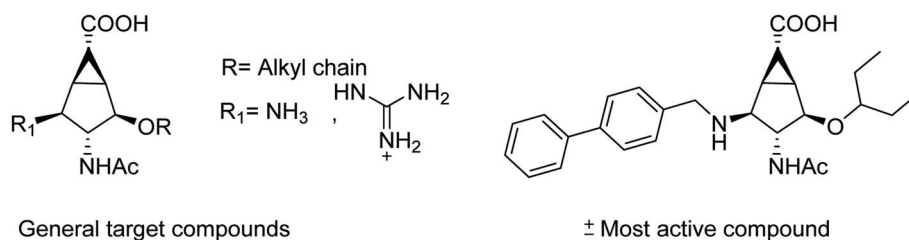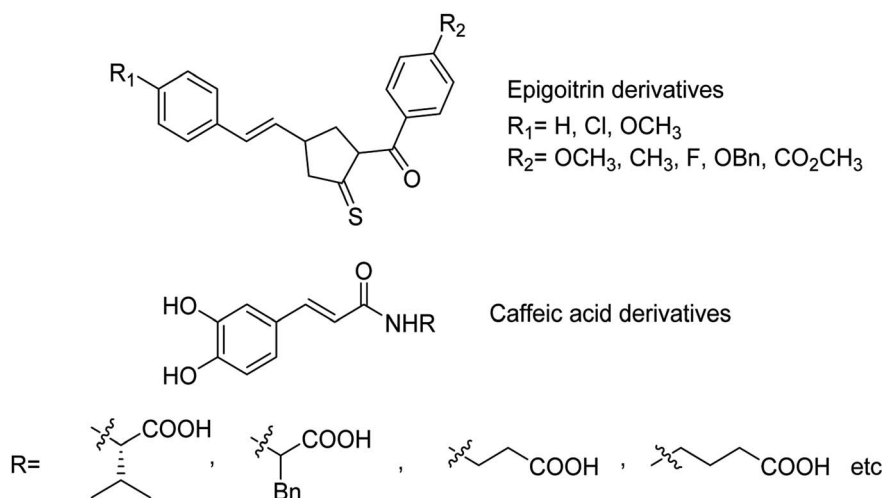**Fig. 24** Structures of bicyclo[3.1.0]hexane analogues of sialic acid.

Fig. 25 Structures of caffeic acid and epigoitrin derivatives.

considered as a NA inhibitor and should have potential to be developed as an anti-influenza drug particularly to H5N1 with oseltamivir resistance. OSMIP@silica gel was prepared by Yang and co-workers[103] to improve the chromatographic performance. NIP@silica gel has been made for comparison. These were characterized through FT-IR, SEM, SSAA and porosity measurements. The OSMIP@silica gel column displayed good affinity and selectivity for template OS and peramivir, and hence, it could be employed to search more active neuraminidase inhibitor analogues from traditional herbs. Li and co-workers[104] have studied the inhibitory activities of a series of carboxyl-modified oseltamivir analogs against neuraminidase from H5N1 virus. One compound has shown potent inhibitory activity (IC$_{50}$ = 1.30 ± 0.23 μM) and is more lipophilic (log D = −0.12) than oseltamivir carboxylate (log D = −1.69) at pH of 7.4.

There results may be helpful in identifying neuraminidase inhibitors in the exploration of 430-cavity with optimal lipophilicity. The active ingredients of four neuraminidase inhibitors are shown in Fig. 23.

Igdoura and co-workers[105] have evaluated the effect of sialidase NEU1 on atherogenesis. They have generated ApoE-deficient mice and found that hypomorphic NEU1 expression in male mice reduces serum levels of VLDL and LDL cholesterol. The reduction decreases atherosclerosis *via* its significant effects on lipid metabolism and inflammation. From the study, it was concluded that NEU1 may represent a promising target for managing atherosclerosis. Colombo and co-workers[106] have reported a mini library of new bicyclic compounds (Fig. 24) and characterized them by enzyme inhibition assays. None of the synthesized compounds presented fair activity against
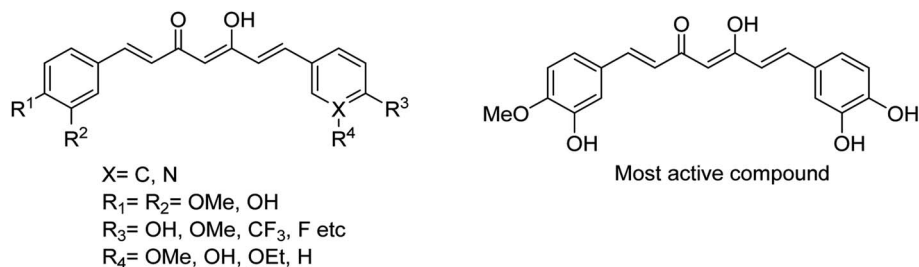
X= C, N
R₁= R₂= OMe, OH
R₃= OH, OMe, CF₃, F etc
R₄= OMe, OH, OEt, H

Most active compound

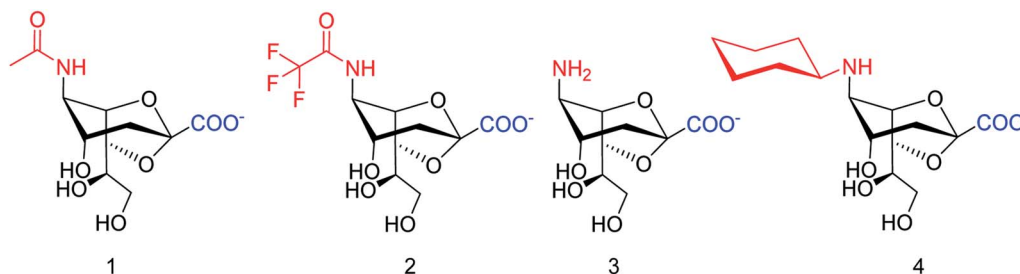**Fig. 26**   Structures of curcumin analogues.

Fig. 27   Structures of enzymatically synthesized 2,7-anhydro-Neu5Ac (**1**) and its chemoenzymatically synthesized derivatives **2–4**.

influenza A neuraminidases at concentrations less than 2 mM. According to them, choice and positioning of the functional groups still need to be properly tuned.

The sialidase's activity was correlated with the pathogenicity of many pathogenic microorganisms. Sialidases efficiently and specifically cleave the glycosidic bond in sialo-glycoconjugates. Xing and co-workers[107] have reported a sialic acid-coated TPE4S with good stability, excellent hydrophilicity, high sensitivity and unique selectivity towards sialidases. Moreover, TPE4S was conveniently employed for the screening of sialidase inhibitors and diagnosis of bacterial vaginosis. Molecular docking and 3D-QSAR methods were used by Yin co-workers[108] to explore the SAR with caffeic acid derivatives[109] (Fig. 25). Ten epigoitrin derivatives have been outlined on the basis of computational

results and analysed. The computational studies governed the designation of epigoitrin derivatives as novel neuraminidase inhibitors.

Lee and co-workers[110] have reported some potential agents using turmeric and curcumin analogues for treating bacterial infections caused by *S. pneumoniae* Nan A. The SAR studies were described, and the synthesised derivatives (Fig. 26) were potent inhibitors of *S. pneumoniae* sialidase (IC$_{50}$ = 0.2 ± 0.1 μM). The most potent compound displayed a 3.0-fold improvement in inhibitory activity over that of curcumin. This study suggested that the curcumin derivatives may be used for the treatment of sepsis by bacterial infections.

Mistry and co-workers[111] have analyzed the biological activities of substituted piperazine-based berberine analogues

R₁= H, Met, Et
R₂= MeEt, Phenyl, Acetyl etc
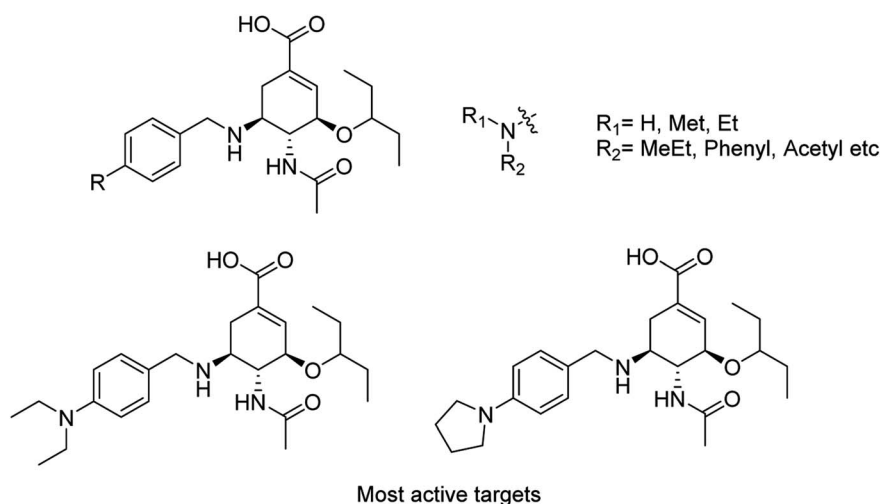
Most active targets

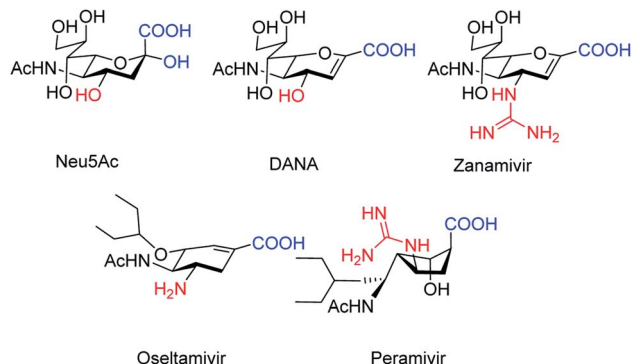**Fig. 28**   Structures of N-substituted oseltamivir derivatives.

Fig. 29 Structures of tested compounds.

conjugated through a pentyloxy side chain. All the analogs have been screened for their *in vitro* antiviral activity against some influenza strains by using SRB and CPE assays. Moreover, the cytotoxicity was evaluated using MDCK cell lines. In addition, the anti-influenza activities of BPD were examined. As per the study, these compounds were found to be potential neuraminidase inhibitors that merge to the active site of neuraminidase. Smith and co-workers[112] have described the recent progress of the biotin/(strept)avidin self-assembly system and the modern design rules for creating synthetic mimics. Moreover, a series of case studies have been given on synthetic derivatives of cucurbiturils, cyclodextrins and various organic cyclophanes (calixarenes, pillararenes, tetralactams and deep cavitands).

SpNanB is responsible for streptococcal infection of the upper and lower respiratory tract. Chen and co-workers[113] have reported the development of a potent OPME system for synthesizing 2,7-anhydro-Neu5Ac and its derivatives (Fig. 27). An *N*-cyclohexyl derivative of 2,7-anhydro-neuraminic acid was synthesized and found to be a selective inhibitor against SpNanB and SpNanC. The study demonstrated a unique strategy of synthesizing 2,7-anhydro-sialic acids and potential applications of their derivatives as selective sialidase inhibitors.

Liu and co-workers[114] have reported some potent oseltamivir derivatives on the basis of their earlier discovery of N1-selective inhibitors (Fig. 28). Some of the synthesized derivatives were exceptionally active against group-1 and group-2 NAs. These compounds displayed greater inhibitory activity than OSC towards E119V and H274Y variant. Moreover, they showed significant potency toward viruses (H5N2, H5N1, H5N6, and

H5N8) in cellular assays. Furthermore, molecular dynamics and computational modeling studies have also been discussed to understand the role of R group. This study will be an important breakthrough in the development of more potent anti-influenza agents.

Cairo and co-workers[115] have tested the inhibitory activity of DANA, oseltamivir, zanamivir, and peramivir against the human isoenzymes (Fig. 29). These compounds showed poor activity against the human neuraminidase enzymes except DANA and zanamivir. Ordered water is important for inhibitor design according to molecular docking simulation of NEU2 and NEU3.

Toshimaa and co-workers[116] have chemically synthesized a series of fucoidan derivatives to bind HAs of influenza virus and inhibit the hemagglutination activity. Moreover, a synthesized ligand with three sulfated oligofucoside moieties showed attractive hemagglutination inhibition activity. One of the reported compounds effectively inhibited influenza virus infection. Zanamivir and GOC are the potent inhibitors against influenza neuraminidase. Fang and co-workers[117] have investigated the use of zanamivir and GOC acylguanidine derivatives (Fig. 30) as possible oral prodrugs. The derivatives have synthesized by coupling with either (*S*)-naproxen or *n*-octanoic acid. According to molecular docking studies, the straight octanoyl chain could extend to 150-cavity and 430-cavity of NA to gain extra hydrophobic interactions. Mice taking a zanamivir octanoylguanidine derivative survived better than those treated with zanamivir. Moreover, a GOC octanoylguanidine derivative could be orally administered to treat mice with efficacy equal to oseltamivir. This study may be helpful for the development of oral drugs for influenza therapy.

Yin and co-workers[118] have synthesized a series of 1-amino-2-alkanols. According to the study, (*E*)-1-amino-4-phenylbut-3-en-2-ol had better inhibitory activities than 2-amino-1-arylethan-1-ol derivatives. The sulfonation of (*E*)-1-amino-4-phenylbut-3-en-2-ol with 4-methoxybenzenesulfonyl chloride afforded more active inhibitor II. Moreover, molecular docking and SAR analysis also aided valuable information towards the discovery of (*E*)-1-amino-4-phenylbut-3-en-2-ol derivatives as potent and novel NA inhibitors. Kobayashi and co-workers[119] have reported the MGO suppressed influenza A virus replication in a strain-independent manner. In this study, the anti-influenza activity of methylglyoxal against influenza B strains by using MDCK cells was evaluated. The susceptibilities of influenza A and B viruses were compared. This study confirmed that
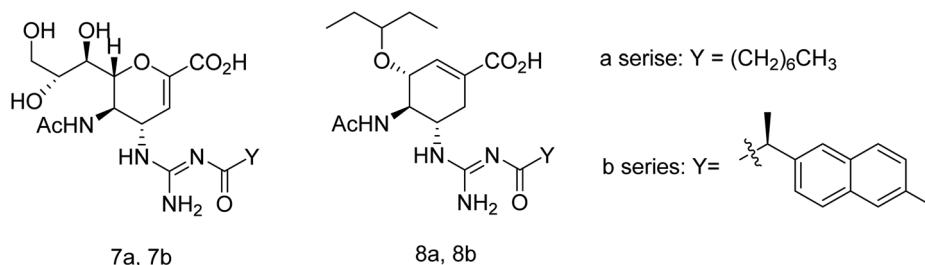

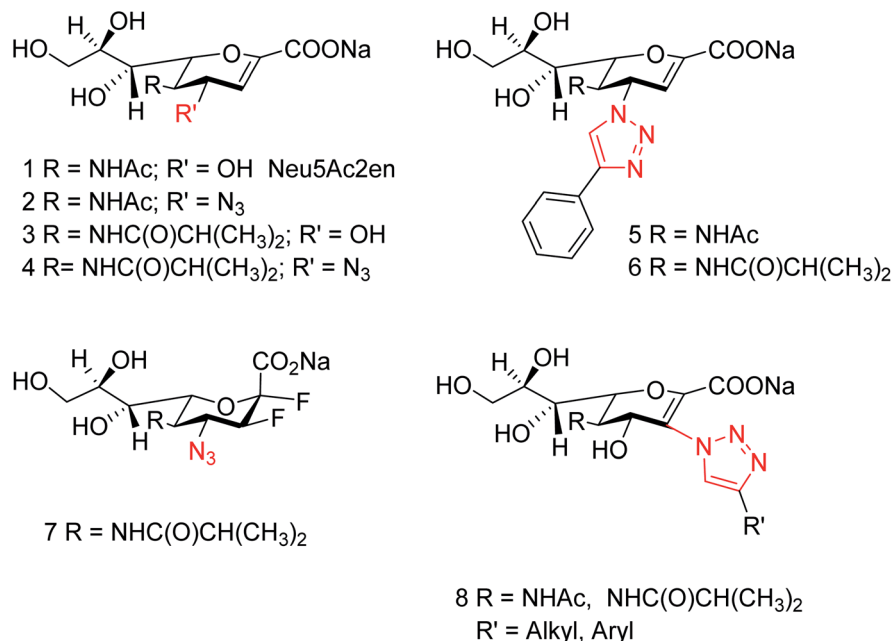Fig. 30 Structures of acylguanidine derivatives.

1 R = NHAc; R' = OH Neu5Ac2en
2 R = NHAc; R' = N_3
3 R = NHC(O)CH(CH$_3$)$_2$; R' = OH
4 R= NHC(O)CH(CH$_3$)$_2$; R' = N_3

5 R = NHAc
6 R = NHC(O)CH(CH$_3$)$_2$

7 R = NHC(O)CH(CH$_3$)$_2$

8 R = NHAc, NHC(O)CH(CH$_3$)$_2$
R' = Alkyl, Aryl

Fig. 31 Structures of reference hPIV-3 HN inhibitors (**1–7**) and the potential new target inhibitors (**8**).

methylglyoxal has potent inhibitory activity against influenza B viruses. A novel approach to hPIV-3 inhibitor design (Fig. 31) has been evaluated by von Itzstein and co-workers[120] with the introduction of nitrogen-based functionalities on the neuraminidase inhibitor template N-acyl-Neu2en. Due to introduction of triazole substituents, the compounds showed a 48-fold improved potency over the corresponding C-3 unsubstituted analogue. However, C-3-triazole N-acyl-Neu2en derivatives were less active against the virus' haemagglutinin function and showed insignificant in vitro antiviral activity.

To enhance the therapeutic effects, the azo (N=N) linkage is very much useful for linking two bioactive moieties. Based on this, Narasimhan and co-workers[121] have discussed the recent developments on diazenyl derivatives as antimicrobial agents during the last five years. This study has provided useful information to develop novel antimicrobial agents by slight modifications in active derivatives. Due to poor solubility in saline, baicalin has limited clinical use. In this context, Dou and co-workers[122] have prepared sodium baicalin (Fig. 32), which showed more solubility in saline. It was evaluated against oseltamivir-resistant mutant H1N1-H275Y in vitro and in vivo.

The treatment alleviated lung injury and body weight loss. This study confirmed that sodium baicalin is effective on wild-type and oseltamivir-resistant viral strains.

Swine flu is caused by influenza A virus in both swine and human. Neuraminidase plays a significant role in the infection of progeny virions inside the respiratory tract of the host body. Hafeez and co-workers[123] have predicted possible neuraminidase inhibitors from the online available tools using a bioinformatics software. The active site has been predicted and then allowed for docking studies. According to docking results, indacaterol have minimum free energy (-8.2 kcal mol^{-1}). An extremely potent inhibitor of influenza virus sialidase (Fig. 33) was identified by von Itzstein and co-workers.[124] The synthesised 3-guanidino sialosyl α-sulfonate inhibits virus replication in vitro comparable to that of zanamivir. As per protein X-ray crystallography, the sialosyl α-sulfonate template binds inside the sialidase active site in 1C4 chair conformation. This sulfonozanamivir analogue will bring a fruitful step towards the anti-influenza drug development.

Jia and co-workers[125] have reported the progress of small molecular inhibitors such as HA inhibitors, NA inhibitors,
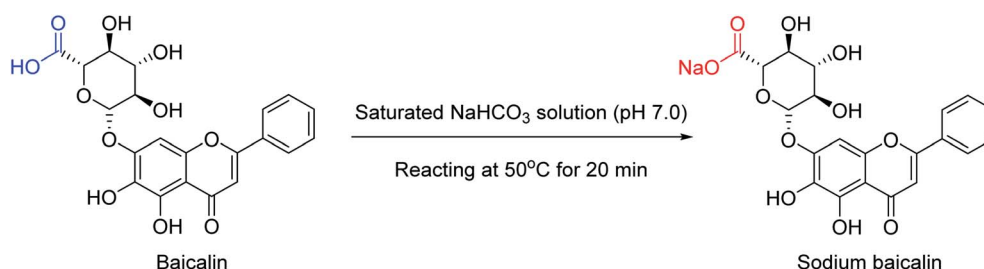

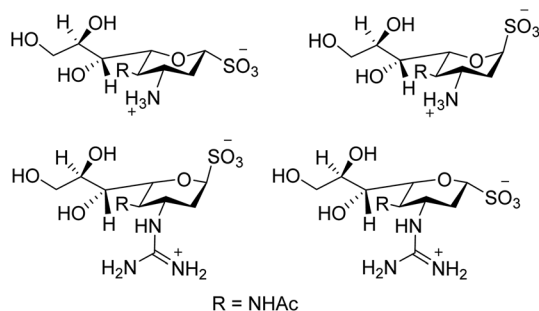**Fig. 32** Synthesis of sodium baicalin.

Fig. 33 Structures of 3-amino sialosyl sulfonates and 3-guanidino sialosyl α-sulfonates.

Fig. 35 Structures of C-1-modified oseltamivir derivatives.

RdRp inhibitors and M2 ion channel protein inhibitors acting as antiviral agents. They also summarized some newly reported potential targets and strategies to develop new anti-influenza drugs. Ivanova and co-workers[126] have reported the anti-influenza activity of some pyrazole derivatives (Fig. 34). In this study, 1-(β-d-ribofuranosyl)-3-polyfluoroalkyl-5-phenyl-1H-pyrazoles were synthesized from regiospecific ribosylation of 3-polyfluoroalkyl-5-phenylpyrazoles and 1,2,3,5-tetra-O-acetyl-b-d-ribofuranose in the presence of SnCl4 followed by deacetylation.

Liu and co-workers[127] have synthesized a group of novel oseltamivir derivatives with C-1 modification, targeting the 430-cavity. These reported compounds (Fig. 35) showed robust anti-influenza potencies against H5N1 and H5N6 viruses. Moreover, the molecular modeling study has also been carried out. The study will be helpful in the discovery of potent inhibitors against group-1 and group-2 NAs.

Cairo and co-workers[128] have designed a series of DANA analogues (Fig. 36) with modifications at C4 and C9 positions. They discovered selective inhibitors targeting the human NEU3 isoenzyme. The most selective inhibitor ($K_i = 320 \pm 40$ nM) has a 15-fold selectivity over other human neuraminidase isoenzymes. It blocks the glycolipid processing by NEU3 *in vitro*. Some esters were also synthesized from the best inhibitors. Few
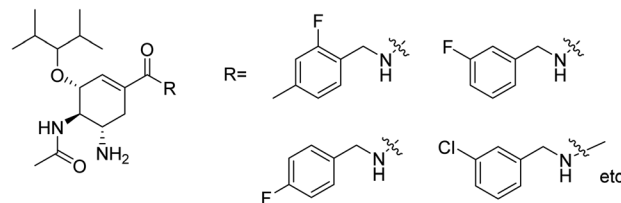
synthesized compounds displayed selective inhibition of NEU orthologues from the murine brain.

Due to the drug-resistant activity, it is an essential and urgent need to discover some novel antiviral drugs. *Artemisia rupestris* L. is a folk medicine used for the treatment of cold. Huang and co-workers[129] have modified rupestonic acid (Fig. 37), a component of *A. rupestris*, to synthesize a series of fifteen 2-substituted rupestonic acid methyl esters. The compounds were characterized by using several spectral tools. These compounds (**3b** and **3c**) displayed potent activities against H1N1 and may be used for the treatment of influenza.

6.2. Natural neuraminidase inhibitors

The *Laggera pterodonta* herb may inhibit influenza virus infection but its anti-influenza components remain unknown. Yang and co-workers[130] have isolated the components (Fig. 38) from the plant by column chromatography. NA inhibition was determined by chemiluminescence assay. The anti-inflammation and anti-virus effects were measured by dual-luciferase reporter assay, quantitative real-time PCR, immuno-fluorescence, and luminex assay. It was suggested that pterodontic acid (isolated from *L. pterodonta*) might be a potential antiviral agent against influenza A virus.

Kolodziej and co-workers[131] have assessed the inhibitory activity of plant tannins (Fig. 39) using a MUNANA-based activity assay on both bacterial and viral model neuraminidase. The

Fig. 34 Synthesis of 1-(β-d-ribofuranosyl)-3-polyfluoroalkyl-5-phenyl-1H-pyrazoles.

2a,3a; R = N(CH₃)₂
2b,3b; R = NHAc
2c,3c; R = NH₂
2d,3d; R = CH₃
2e,3e; R = OCH₃
2f,3f; R = F
2g,3g; R = CF₃
2h; R = COOME
7h; R = COOH
2i,3i; R = Ph
2j,3j; R = OPh

Fig. 36 Synthetic route to C9-phenyltriazole DANA analogues.

Fig. 37 Structures of isolated rupestonic acid and rupestonic acid derivatives.

Fig. 38 Structure of pterodontic acid.

Fig. 40 Structure of simpterpenoid A.

evaluated potency was compared with OC and zanamivir against H1N1 and VCNA. All the tested polyphenols presented a weak inhibition of the viral enzyme but higher potency on the bacterial neuraminidase. In addition, SAR analyses were performed to understand which groups are crucial for inhibitory activity.

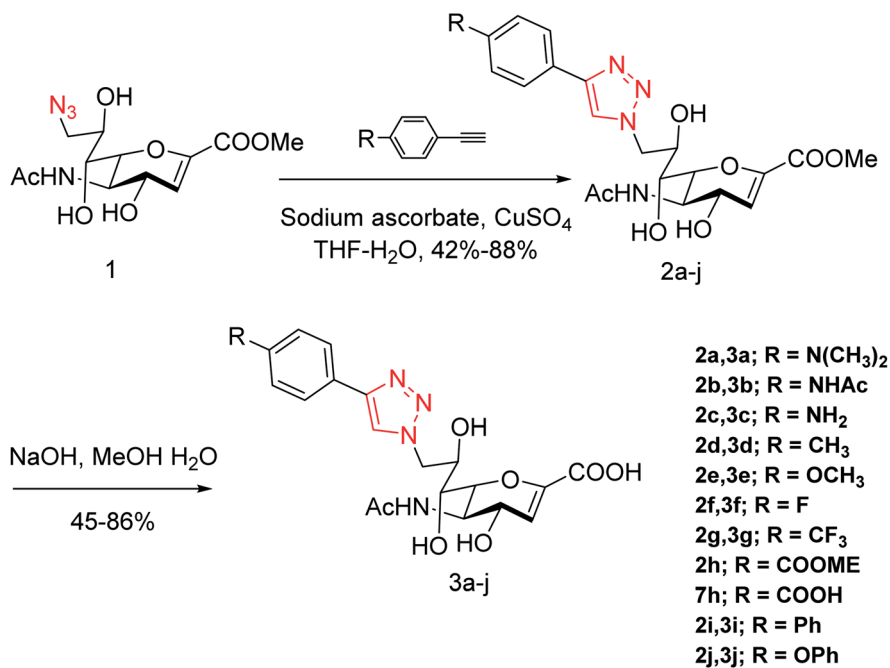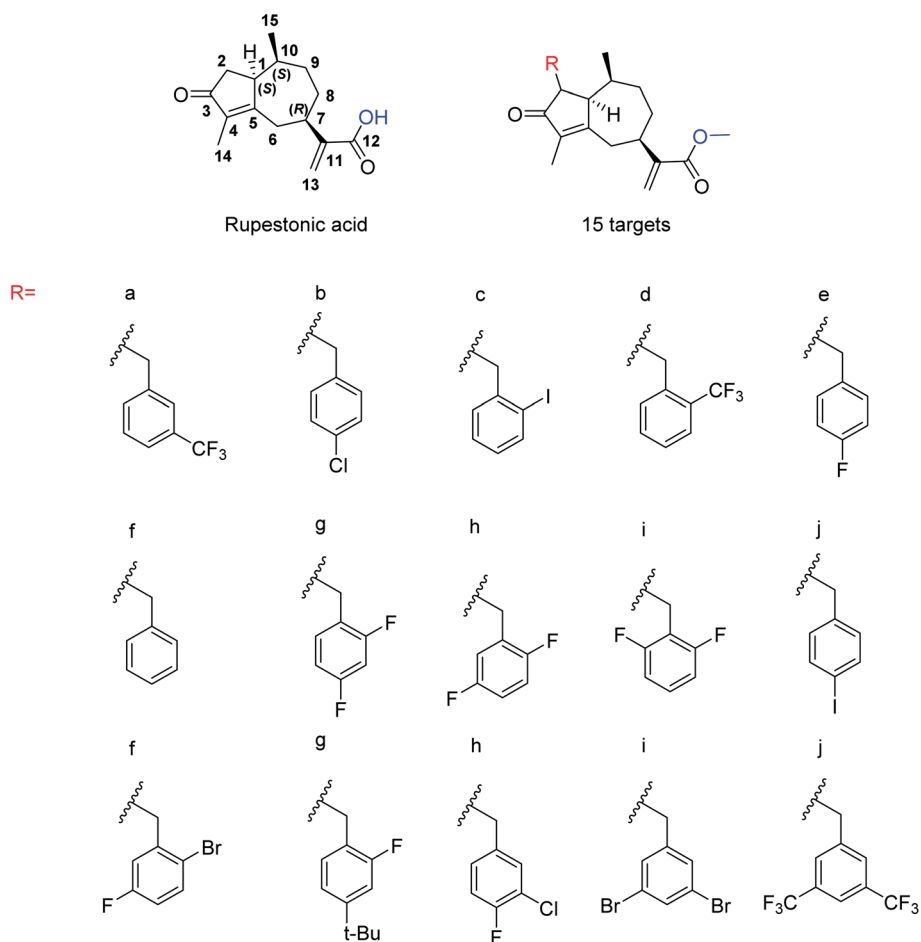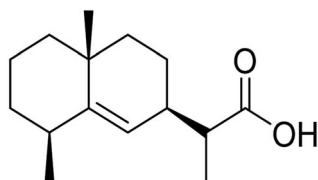Marine microbials have attracted tremendous attention in the last two decades. Wang and co-workers[132] have reported simpterpenoid A (Fig. 40) containing a highly functionalized cyclohexadiene moiety (ring C), identified from mangrove-derived *Penicillium simplicissimum* MA-332. The X-ray crystallographic analysis was also reported to understand the unambiguous assignment of the planar structure and steric configuration. Simpterpenoid A showed inhibitory activities against influenza neuraminidase in nanomolar quantities.

Kanyalkar and co-workers[133] have designed a series of chalcone derivatives based on isoliquiritigenin (natural NA inhibitor). Molecular modeling studies suggested that isoliquiritigenin and its analogs occupied a 430-loop cavity of NA. Moreover, the favorable compounds were synthesized and evaluated. The inhibitory effect was also quantified by H1N1-NA inhibition and HA assay. The marine-derived microbes are biologically active natural products. Keeping this in mind, Wang and co-workers[134] have reported ten citrinin analogues from the EtOAc extracts of a coculture of two marine algal-derived endophytic fungal strains: *Penicillium citrinum* (EN-535) and *Aspergillus sydowii* (EN-534). The structures were confirmed by NMR, ECDs, HRESIMS, optical rotation, and XRD data. The antimicrobial and influenza neuraminidase inhibitory activities of these compounds were also assessed. Due to
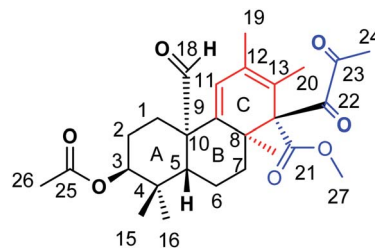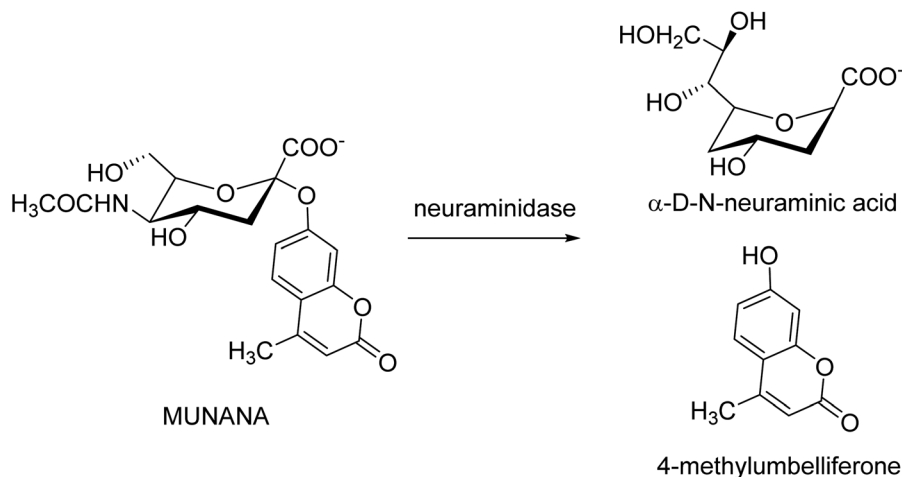
the drug resistance activity of unfamiliar strains to the available drugs, there is a rapid demand for the discovery of antiviral drugs. Neuraminidase is a crucial enzyme for viral replication and is the most promising target for the new drugs. Chandra and co-workers[135] have reported the update on neuraminidase enzyme highlighting its structure, function, mechanism and inhibition by natural products, which will be helpful in the discovery of antiviral agents in the future. Some experimental studies have reported that tea catechins may inhibit influenza viral adsorption and suppressed replication and neuraminidase activity. In addition, tea catechins may be helpful to enhance immunity against viral infection. Moreover, regular consumption of green tea decreases the infection rates of some cold symptoms, and gargling with tea catechin may protect against influenza infection. Yamada and co-workers[136] have briefly discussed the effect of tea catechins on common cold and influenza infection with a focus on clinical studies. Park and co-workers[137] have reported the isolation of phenolic metabolites (Fig. 41) from the methanol extract of *Usnea longissima* by phytochemical investigation. All these compounds were characterized by several spectral techniques. Most of the compounds displayed dose-dependent inhibition when assessed against BNA. The compounds **2** and **3** (IC_{50} = 7.8, 8.2 μM) had three-fold greater potency than quercetin (IC_{50} = 21.4 μM). Furthermore, compounds **2** and **3** showed reversible and mixed type-I inhibitory behaviors (K_i = 6.8 and 7.2 μM, respectively).

α-D-N-neuraminic acid

MUNANA

4-methylumbelliferone

Fig. 39 Neuraminidase-catalyzed hydrolysis of 20-(4-methylumbelliferyl)-α-D-*N*-acetylneuraminic acid (MUNANA).
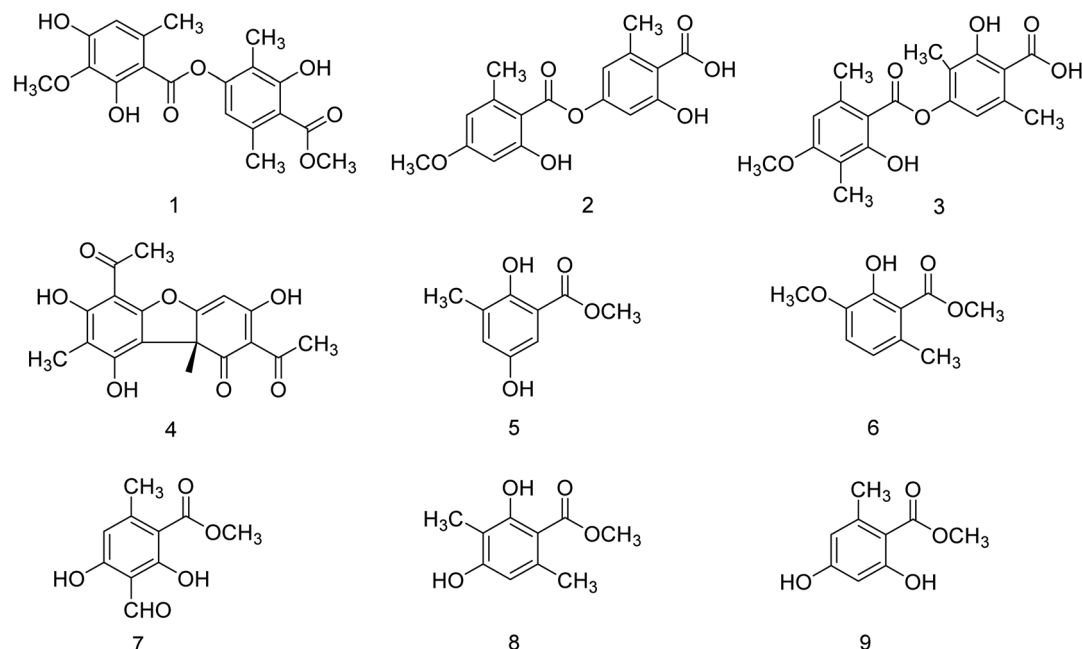
Fig. 41 Structures of isolated compounds from *U. longissima*.

BF-30 (peptide) isolated from the venom of *Bungarus fasciatus* (snake) is suggested to have antitumor and antibacterial activities. Dou and co-workers[138] have suggested that BF-30 could inhibit influenza virus strains (H1N1 and H3N2) and oseltamivir-resistant H1N1, *in vitro*. Mice treated with BF-30 exhibited 50% survival at a dosage of 4 μM, with an approximately 2 log viral titer decrease in the lung. According to the study, peptide BF-30 showed an effective inhibitory activity against influenza A virus, and hence, it will be a promising candidate for influenza therapy. Flavonoids containing *Scutellaria baicalensis* root are traditionally used to treat against fever, common cold, and influenza. The anti-influenza and anti-inflammatory activities of FESR and the main flavonoids (Fig. 42) were assessed by Chen and co-workers[139] *in vitro*. Baicalin (BG) was given as a reference control. According to the assays, FESR has a great potential against ALI induced by IAV.

Moreover, FESR exhibited more potent therapeutic effects than BG.

DS is a traditional herb available in various Asian countries. Nile and co-workers[140] have reported the extraction of bioactive compounds and their biological activities from the EA, Bu and DW extracts of *Dianthus superbus*. The antiviral, antioxidant and anticancer activities were determined. EA extracts exhibited strong anticancer activities on NCL-H1299, SKOV, and Caski cancer cell lines. The Bu extracts showed the strongest antiviral activity with respect to both influenza A and B viruses. The isorhamnetin 3-glucoside and quercetin 3-rutinoside exhibited higher neuraminidase inhibition activity in a dose-dependent manner. This study may provide vital information for the effective utilization of DS for food, medicinal and therapeutic purposes. Geniposide extracted from *Gardenia jasminoides* Ellis (GJ) fruit, has antiviral and anti-inflammation activities.
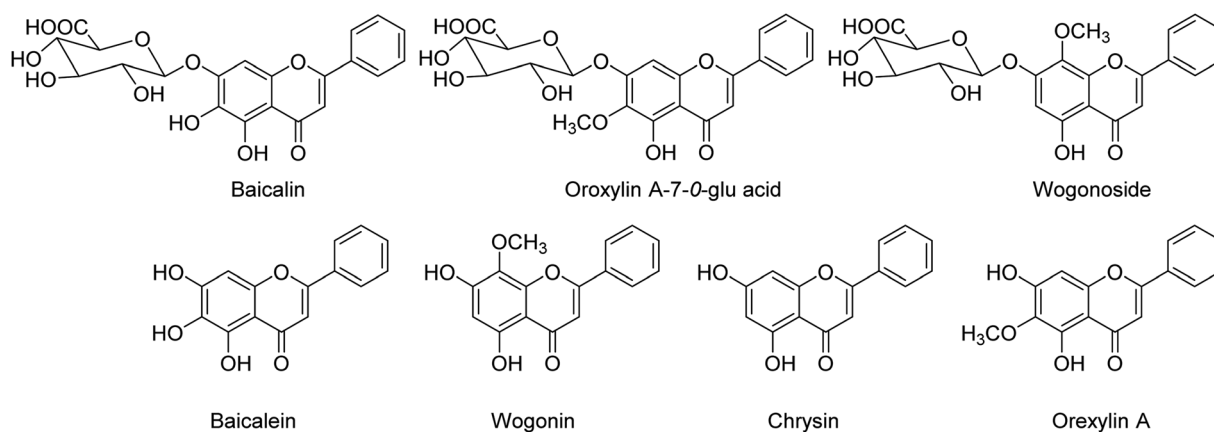

Fig. 42 Structures of baicalin, oroxylin A-7-*O*-glu acid, wogonoside, baicalein, wogonin, chrysin and oroxylin A.

Fig. 43 Structures of honokiol and honokiol derivatives.

Fig. 44 Structures of isolated chromenone derivatives from *Flemingia philippinensis*.

Ganzhu and co-workers[141] have reported the antiviral and cytotoxicity activity of geniposide by an MTT assay. The influenza respiratory tract infection murine model was studied with A/Jiangsu/1/2009 (H1N1) virus. Geniposide substantially inhibited the virus-induced alveolar wall changes. The study confirmed that geniposide effectively inhibited the cell damage caused by A/Jiangsu/1/2009 (H1N1) virus and mitigated virus-induced acute inflammation. A natural polyphenol (Honokiol) having NA inhibitory activity was structurally modified by Hu and co-workers[1] (Fig. 43). The authors have reported 23 compounds with their ortho-effects in epoxidation and hydrolyzation reactions. The benzoylhydrazone derivatives exhibited much better anti-NA activity than honokiol. According to SAR analysis, polyphenols showed better anti-NA activity than monophenols and biphenols. Moreover, the most active compound had potent anti-influenza virus activity.
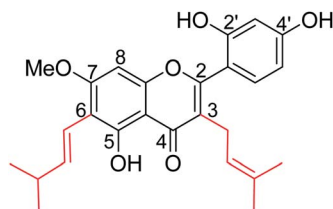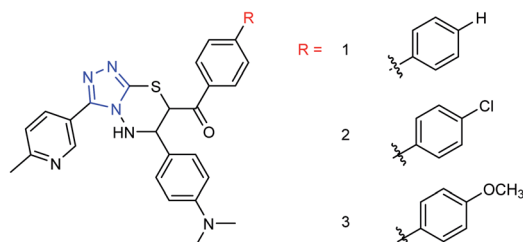
Fig. 45 Structure of artocarpin.

Fig. 46 Structures of 4-dimethylaminobenzaldehyde derivatives.

The chromenone derivatives (Fig. 44) obtained from *Flemingia philippinensis* exhibit significant inhibition against bacterial NA and play an important role in the pathogenesis of bacterial infection.[142] As per the kinetic study, 8,8-diprenyl compounds showed a competitive inhibitory mode. Moreover, inhibitors (**1** and **2**) were simple, reversible and slow-binding against bacterial neuraminidase.

The inhibition of tyrosinase and melanogenesis are two skin-whitening activities of artocarpin (Fig. 45), an isoprenyl flavone from *Artocarpus* species. It protects UVB-induced skin damage. Besides this, artocarpin possesses other pharmacological properties such as antimicrobial, anti-inflammatory, antioxidant, antiplasmodial, antitubercular, cytotoxic, and neuraminidase inhibitory activities. Chan and co-workers[143] have reported the chemistry and pharmacology of artocarpin.

6.3. Patents study

Ye and co-workers[144] have synthesized three novel compounds (Fig. 46) from 4-dimethylaminobenzaldehyde with the yields ranging from 59 to 89%. These compounds could inhibit neuraminidase to varying degrees. The anti-influenza virus
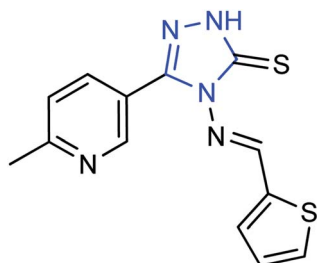

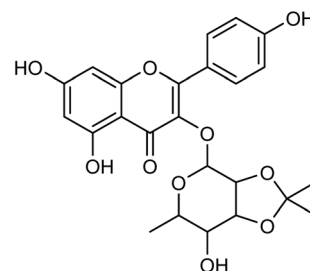**Fig. 47** Structure of 3-(6-methylpyridine-3-yl)-4-[(thiophene-2-yl) methylene amino]-*H*-1,2,4-triazol-5(4*H*)-thione.

Fig. 48 Structure of kaempferol-3-0-[(4″5″-0-isoprotylidene)-*a*-L-rhamnopyranoside.

aminotransferase activity of these compounds was detected in the enzyme reaction system.

A new compound (Fig. 47), 3-(6-methylpyridine-3-yl)-4-[(thiophene-2-yl) methylene amino]-1*H*-1,2,4-triazol-5(4*H*)-thione, was synthesized from 6-methylnicotinic acid methyl ester by Ye and co-workers[145] with a yield of 73%. At a concentration of 40 μg mL^{-1}, the inhibition rate of the compound was 42.19%. It was proved that the compound has promising activities as an anti-influenza neuraminidase inhibitor and can be used for further preparation of neuraminidase inhibitors.

Cui and co-workers[146] have used the branches, leaves or fruits of Cassia trees as raw materials, and separated a kind of flavonoid compounds (Fig. 48) by organic solvent ultrasonic extraction, organic solvent extraction, silica gel column chromatography and liquid chromatography. The molecular formula of the flavonoid is $C_{24}H_{24}O_{10}$, named kaempferol-3-0-[(4″5″-0-isoprotylidene)-*a*-L-rhamnopyranoside]. The results indicated that the compound exhibited good neuraminidase inhibitory activity with an IC$_{50}$ value of 187.40 μML^{-1}. It can be used as a leading compound of anti-influenza drugs.

Peptide inhibitors are widely used in the prevention and treatment of viral infection. Ning and co-workers[147] have used molecular docking to screen small molecules, which can dock with neuraminidase from a small molecular library. According to the docking score, the best 20 protein sequences were selected, and the method of increasing or reducing 2–4 amino acids was used to obtain peptide inhibitors with a higher antiviral bioactivity. *In vitro* experiments have investigated that the screened 10 peptides showed low IC$_{50}$ values against influenza virus (H1N1). Among them, two peptides showed good inhibitory activities against H1N1 with IC$_{50}$ values of 2.24 and 2.99 μM, and the results indicated a potent strong inhibitory effect
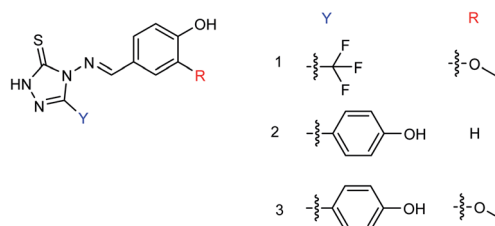

Fig. 49 Structure of 4-(arylimino)-3-alkyl-1*H*-1,2,4-triazol-5(4*H*)-thioketone derivatives.
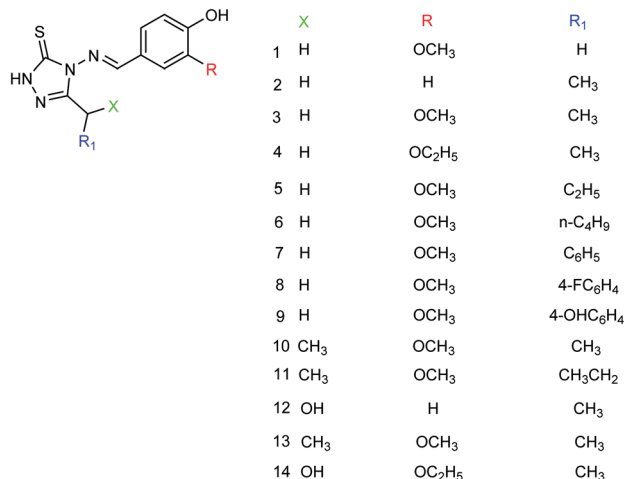
	X	R	R$_1$
1	H	OCH$_3$	H
2	H	H	CH$_3$
3	H	OCH$_3$	CH$_3$
4	H	OC$_2$H$_5$	CH$_3$
5	H	OCH$_3$	C$_2$H$_5$
6	H	OCH$_3$	n-C$_4$H$_9$
7	H	OCH$_3$	C$_6$H$_5$
8	H	OCH$_3$	4-FC$_6$H$_4$
9	H	OCH$_3$	4-OHC$_6$H$_4$
10	CH$_3$	OCH$_3$	CH$_3$
11	CH$_3$	OCH$_3$	CH$_3$CH$_2$
12	OH	H	CH$_3$
13	CH$_3$	OCH$_3$	CH$_3$
14	OH	OC$_2$H$_5$	CH$_3$

Fig. 50 Structures of 4-(4-hydroxyphenylmethyleneamino)-1,2,4-triazol-5-thione derivatives.

Fig. 52 Structures of oxazolamide derivatives.

against virus-infected cells. Their protein sequences were errKPAQP and hsFHHKPAK respectively, and the non-toxic effect of peptide inhibitors on HEK 293T was investigated. Three derivatives of 4-(arylimino)-3-alkyl-1H-1,2,4-triazol-5(4H)-thioketone (Fig. 49) were designed and synthesized by Yan and co-workers[148] with the yields ranging from 60 to 75%. *In vitro* experiments indicated that the synthesized compound showed good activity against influenza virus neuraminidase, which can be used to prepare further neuraminidase inhibitors of influenza virus. The activity of compound 3 is the best, the IC$_{50}$ value is 19 μg mL^{-1}, and the inhibition rate of compound 3 to neuraminidase H1N1 is 70%.

Hu and co-workers[149] designed and prepared a series of 4-(4-hydroxyphenyl-methyleneamino)-1,2,4-triazol-5-thione derivatives (Fig. 50) with the yields ranging from 55 to 82%. The most potent derivative was compound 3 with an IC$_{50}$ value of 14.68 μg mL^{-1}.

Ye and co-workers[150] designed and synthesized two N-thiazolylpyridine formamide derivatives A and B as neuraminidase inhibitors (Fig. 51) with yields of 67% for both of them. Compounds A and B showed good inhibition rates of 82.7 and 83.6% respectively at a concentration of 40.0 μg mL^{-1}. Compounds A and B showed potent inhibitory activities with IC$_{50}$ values of 44.7 μM and 33.9 μM respectively.

Two oxazolamide derivatives were synthesized (Fig. 52) by Ye and co-workers[151] with the yields of 43.5 to 63.8% respectively. *In vitro* results showed that the inhibition rates of the compounds against A/PR/8/34 (H1N1) are 72.5% and 69.8%
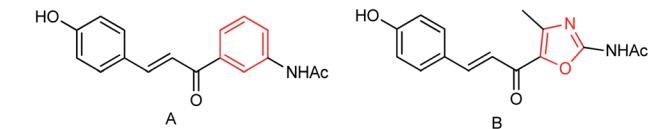
respectively at a concentration of 40.0 μg mL^{-1}. The compounds showed inhibitory activity with IC$_{50}$ values of 67.9 and 73.3 μM respectively. 4-Alkyl-5-(3-phenylacryloyl)-2-alkanoyl-aminoxazole showed good anti-influenza activity, which can be used to prepare potent neuraminidase inhibitors.

Baicalein and its derivatives including flavonoid have significant inhibitory effects on influenza virus; therefore, Li and co-workers[152] designed and synthesized a series of baicalein derivatives (Fig. 53) bearing a novel structure with the yields ranging from 25 to 40%. In order to play the role of prevention and treatment to influenza virus H1N1, type I neuraminidase was chosen as the target. The compounds showed better inhibitory activity than baicalein as a positive control (the half inhibitory concentration of type I neuraminidase was lower than the baicalein control). *In vitro* results indicated good inhibitory activity of compound IB-3 with IC$_{50}$ 4.4 μg mL^{-1}. Compound IB-3 showed good inhibitory activity with an IC$_{50}$ value of 4.4 μg mL^{-1} and in general baicalein phenol hydroxyl esterified derivatives showed good activity and can play an important pharmacological role against H1N1 influenza virus for the treatment of influenza A. The synthesis process is simple and easy to implement and the reaction conditions are stable, controllable and easy to promote and apply.

Another study of Li and co-workers[153] was to establish a small library of baicalein phenol hydroxyl etherified derivatives (Fig. 54) and a series of eighty-two derivatives were synthesized with the yields ranging from 20.2 to 41.4%. *In vitro* study showed the best activity of compound id-5 with IC$_{50}$ of 4.5 μg mL^{-1} and the inhibitory activity was the same as the positive drug zanamivir. In addition, the activity of all compounds was better than baicalin, which has good inhibitory effects on type I neuraminidase.

Nine derivatives of 5-(3-phenylacryloyl)-2-benzoylaminothiazole (Fig. 55) were synthesized by Ye and co-workers[154] with the yields ranging from 44 to 88%. Compound 4-methyl-5-acetyl-2-(3-methyl-4-nitrobenzoylamino)thiazole was selected by the national drug screening center of the Institute of Medicine, Chinese Academy of Medical Sciences as a positive drug for *in vitro* experiment. Compound J showed potent
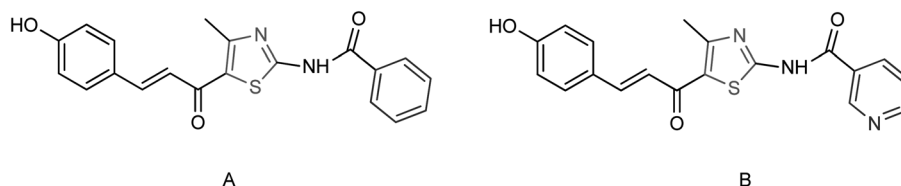

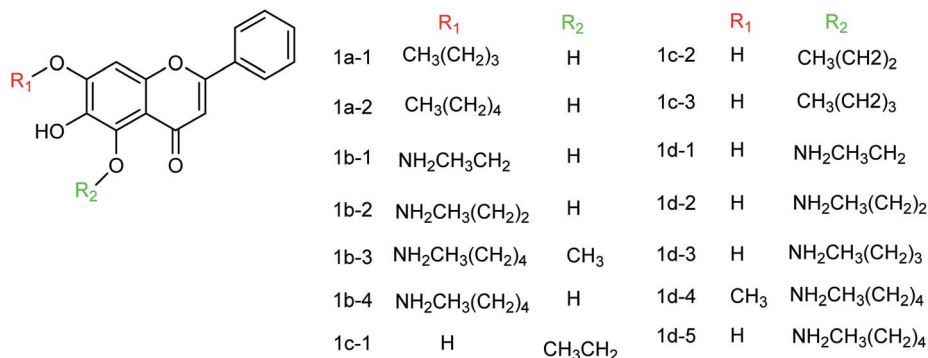Fig. 51 The synthesized N-thiazolylpyridine formamide derivatives.

	R_1	R_2		R_1	R_2
1a-1	$CH_3(CH_2)_3$	H	1c-2	H	$CH_3(CH_2)_2$
1a-2	$CH_3(CH_2)_4$	H	1c-3	H	$CH_3(CH_2)_3$
1b-1	$NH_2CH_3CH_2$	H	1d-1	H	$NH_2CH_3CH_2$
1b-2	$NH_2CH_3(CH_2)_2$	H	1d-2	H	$NH_2CH_3(CH_2)_2$
1b-3	$NH_2CH_3(CH_2)_4$	CH_3	1d-3	H	$NH_2CH_3(CH_2)_3$
1b-4	$NH_2CH_3(CH_2)_4$	H	1d-4	CH_3	$NH_2CH_3(CH_2)_4$
1c-1	H	CH_3CH_2	1d-5	H	$NH_2CH_3(CH_2)_4$

Fig. 53 Structures of baicalein derivatives.

Fig. 54 Structure of compound ld-5.

R=
A H
B 4-OH
C 4-OCH₃
D 4-F
E 4-CN
F 4-CF₃
G 4-NO₂
H 3-NO₂
J 3-CH₃-4-NO₂

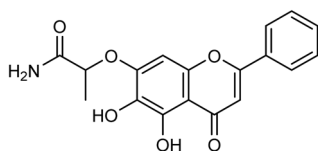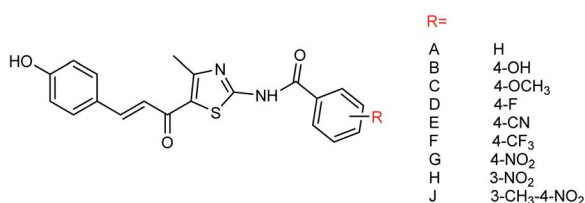**Fig. 55** Structures of 5-(3-phenylacryloyl)-2-benzoylaminothiazole derivatives.

inhibitory activities among all of the synthesized compounds with an IC$_{50}$ value of 14.5 μM at a concentration of 40.0 μg mL^{-1}.

There are many advantages in the preparation of neuraminidase compounds by microbial fermentation because there is no reduction of species and no pollution. Li and co-workers[155] used ethyl acetate to extract the fermentation products of the marine fungus Aspergillus WENTII and purified them by gradient elution with reduced pressure silica gel column chromatography (Fig. 56). Four new diterpenoids with significant neuraminidase inhibitory activities were found.
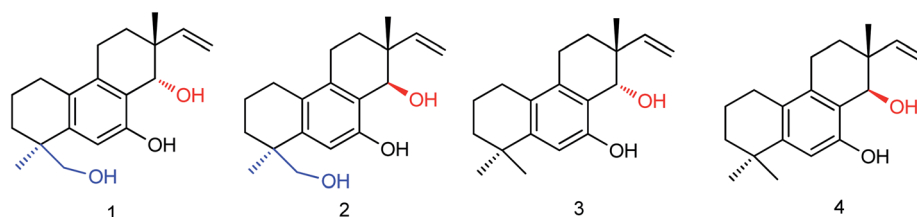
Compounds **1**–**4** showed significant inhibitory activities with IC$_{50}$ values of 18.2, 10.9, 14.2 and 16.7 μM respectively.

Thirteen derivatives of 2-(5-nitrothiazol-2-yl)-imino-4-thiazolinone (Fig. 57) were designed and synthesized by Ye and co-workers[156] with the yields ranging from 52 to 89%. Most of the compounds have good inhibitory activities at a concentration of 40.0 μg mL^{-1}. Among them, 2-(4-*tert*-butyl-5-nitrothiazole-2-yl)imino-4-thiazoline showed the best activity with an IC$_{50}$ value of 12.50 μg mL^{-1}.

Ding and co-workers[157] have extracted Chinese gallnut with ethyl acetate and *n*-butanol to obtain the corresponding ester and alcohol extracts. Gallnut possesses good inhibitory activity against H1N1 influenza virus, effectively controls the infectivity of the virus, down regulates the expression of inflammatory cytokines TNF-α, and improves the lung inflammation of influenza virus-infected mice. In order to further clarify the active parts of gallnut against influenza virus, the authors have carried out *in vitro* experiments on the components of gallnut. The alcohol extract and gallate extract of gallnut showed different degrees of inhibition, while the effect of the gallate extract was particularly obvious with the inhibition rate up to 83%. There are sesquiterpenes, chromogenic ketones, phenylpropanoids, alkaloids and other components in traditional Chinese medicine, which have a wide range of pharmacological activities. Qin and co-workers[158] have studied the inhibitory activity of the extracts from dried Flos Farfarae which is also known as Kuandonghua in China. The results of pharmacodynamic experiments showed that the *n*-butanol extract of Flos Farfarae exhibited significant inhibitory activities. Among them, the *n*-butanol part (d) showed a strong inhibitory activity with IC$_{50}$ = 27.64 ± 0.184 μg mL^{-1}, indicating that the *n*-

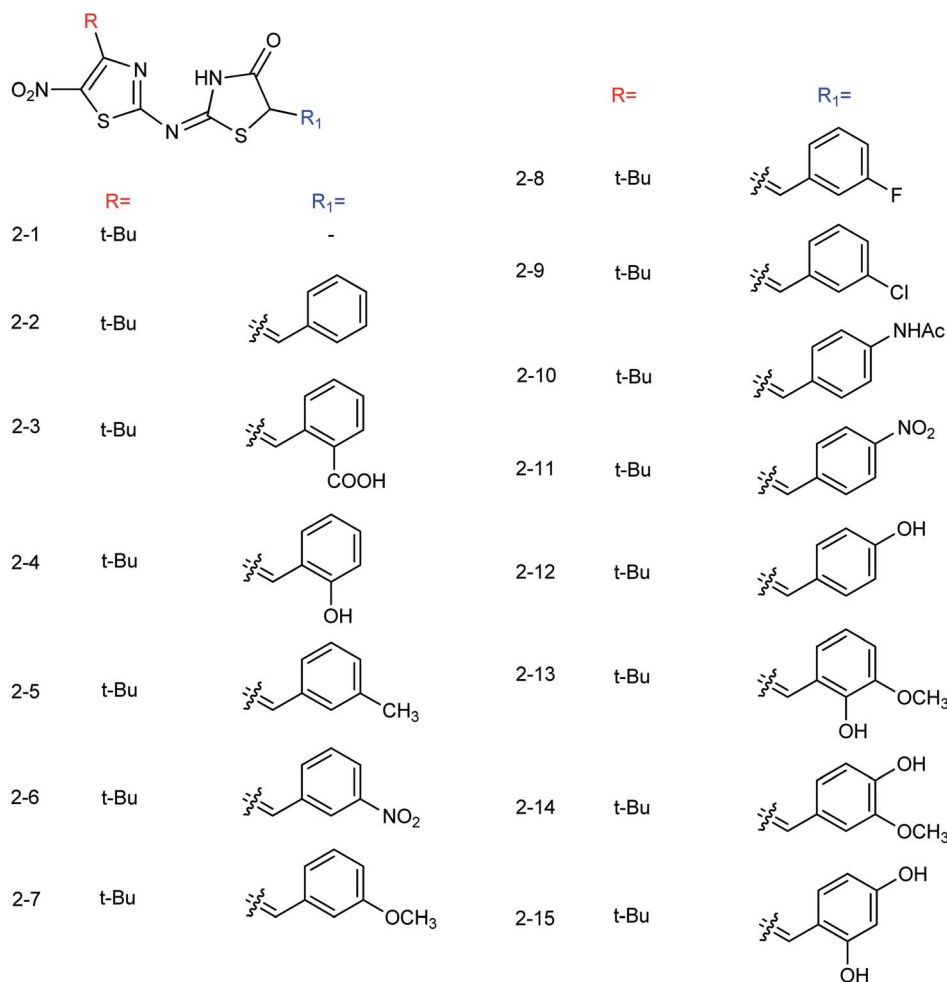**Fig. 56** Structures of diterpenoids.

Fig. 57 Structures of 2-(5-nitrothiazol-2-yl)-imino-4-thiazolinone derivatives.

butanol part (d) is the effective part to inhibit H1N1 neuraminidase, which can be used to prepare further drugs for the prevention and treatment of H1N1 influenza. Jia and coworkers[159] have synthesized a series of 5-(3-phenylacryloyl) thiazole derivatives (Fig. 58) with the yields ranging from 52.2 to 89.7%. Compound 5-(3-phenylacryloyl)thiazole derivatives

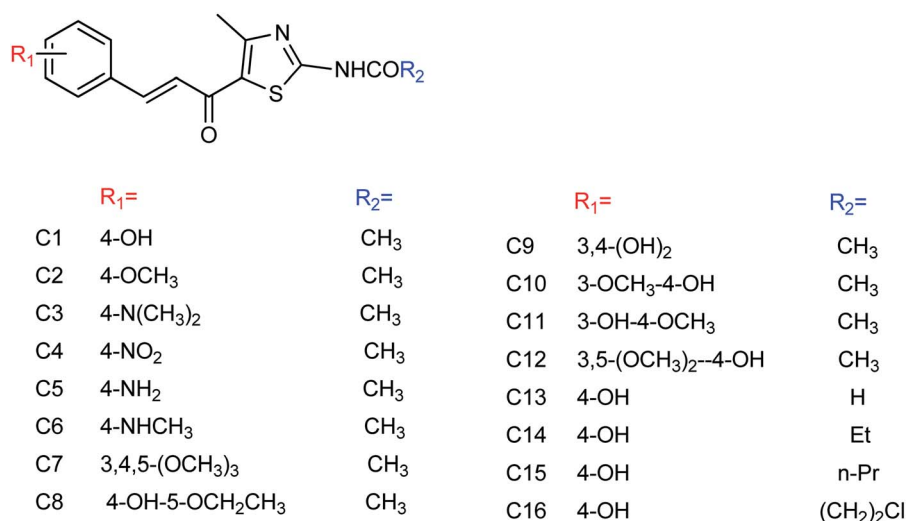|	R₁=	R₂=		R₁=	R₂=
C1	4-OH	CH₃	C9	3,4-(OH)₂	CH₃
C2	4-OCH₃	CH₃	C10	3-OCH₃-4-OH	CH₃
C3	4-N(CH₃)₂	CH₃	C11	3-OH-4-OCH₃	CH₃
C4	4-NO₂	CH₃	C12	3,5-(OCH₃)₂--4-OH	CH₃
C5	4-NH₂	CH₃	C13	4-OH	H
C6	4-NHCH₃	CH₃	C14	4-OH	Et
C7	3,4,5-(OCH₃)₃	CH₃	C15	4-OH	n-Pr
C8	4-OH-5-OCH₂CH₃	CH₃	C16	4-OH	(CH₂)₂Cl

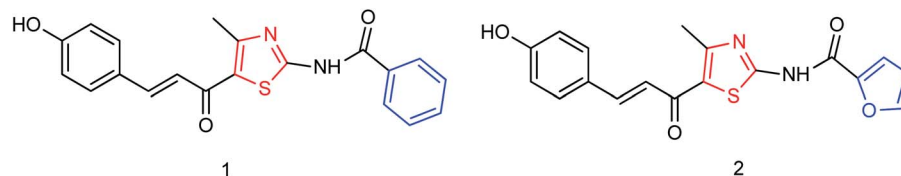**Fig. 58** Structures of 5-(3-phenylacryloyl)thiazole derivatives.

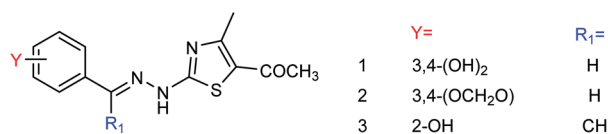**Fig. 59** Structures of furan formamide derivatives.

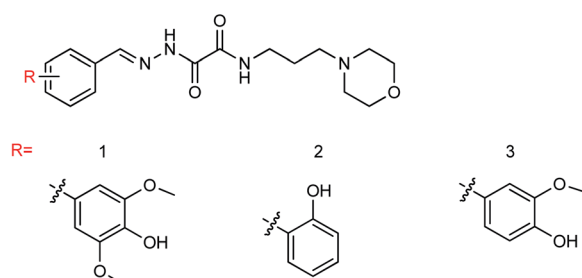**Fig. 60** Structures of (4-alkyl-5-acyl-2-thiazolyl)hydrazone derivatives.

Fig. 61 Structures of hydrazide neuraminidase inhibitors.

showed good inhibitory activities and compound **C1** showed the best activity with an IC$_{50}$ value of 27.1 $\pm$ 0.1 µM. At the same time, the author compared the anti-influenza virus neuraminidase activity of reference compounds **A** and **B** (Institute of medicine, Chinese Academy of Medical Sciences) with compound **C1**. The results indicated that the inhibition rates of compounds **A** and **B** were 62.59% and 79.25% at a concentration of 40 µg mL^{-1}, while the inhibition rate of compound **C1** was 92.32%.

Two furan formamide derivatives **1** and **2** (Fig. 59) were designed and synthesized by Li and co-workers[160] with yields of 67.7% and 57.7% respectively. The bioactivity of the two

compounds was subsequently evaluated. The two compounds have a certain anti-influenza virus neuraminidase activity with IC$_{50}$ values of 44.7 µM and 33.5 µM respectively.

Ye and co-workers[161] prepared three (4-alkyl-5-acyl-2-thiazolyl)hydrazone derivatives (Fig. 60) from acetylacetone with yields ranging from 61.1 to 81.7%. *In vitro* study showed that these compounds displayed good inhibitory activity on the A/PR/8/34 (H1N1) cell line. Among them, compound **2** showed the best activity with an IC$_{50}$ value of 21.07 µg mL^{-1} and an inhibition rate of 71.68%.

Neuraminidase is one of the targets of anti-influenza drugs. In order to find a new neuraminidase inhibitor, Li and co-workers[162] designed and synthesized three novel hydrazide neuraminidase inhibitors (Fig. 61) based on the structure of neuraminidase using computer virtual screening technology and computer-aided drug design. Three compounds showed good neuraminidase inhibitory activity with IC$_{50}$ values of 0.075 µM, 0.46 µM and 0.21 µM respectively, which have higher activity than the OS group (IC$_{50}$ = 33.75 µM).

The emergence of oseltamivir resistant strains, such as H274Y, I117V, E119A, R292K and other neuraminidase mutants, has decreased the effectiveness of neuraminidase inhibitors; therefore, it is urgent to develop new neuraminidase inhibitors. Huang and co-workers[163] designed and synthesized thirteen new oseltamivir derivatives (Fig. 62) with yields ranging from 27 to 69%. *In vitro* study showed good inhibitory activity of the synthesized compounds against H1N1, H3N2 and H259Y (H1N1) influenza viruses. Among them, compound **12** showed potent inhibitory activities with IC$_{50}$ values of 1.43 µM (H1N1), 3.64 µM (H3N2) and 14.47 µM (H259Y) respectively.

The secondary metabolites of marine microorganisms are rich in structural diversity and significant biological activity, which are important sources of drug leading compounds. The
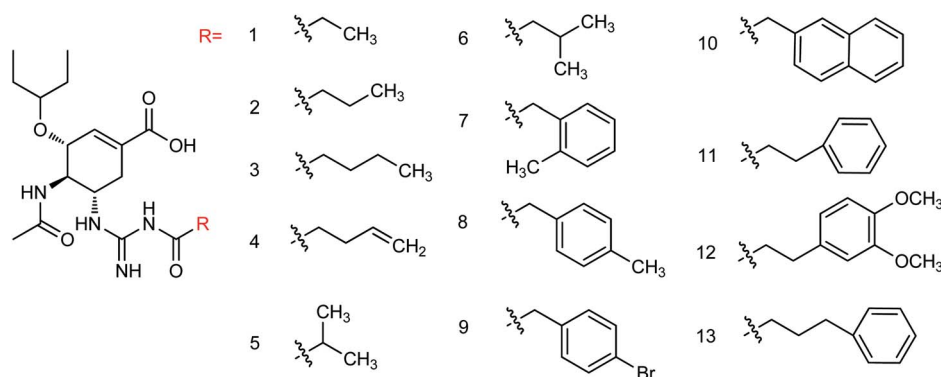

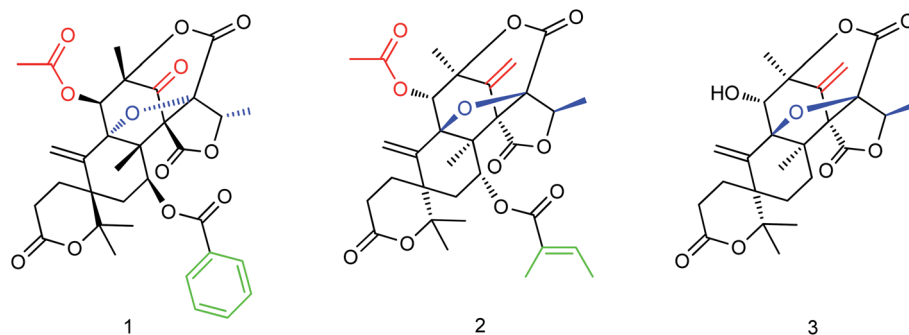**Fig. 62** Structures of oseltamivir derivatives.
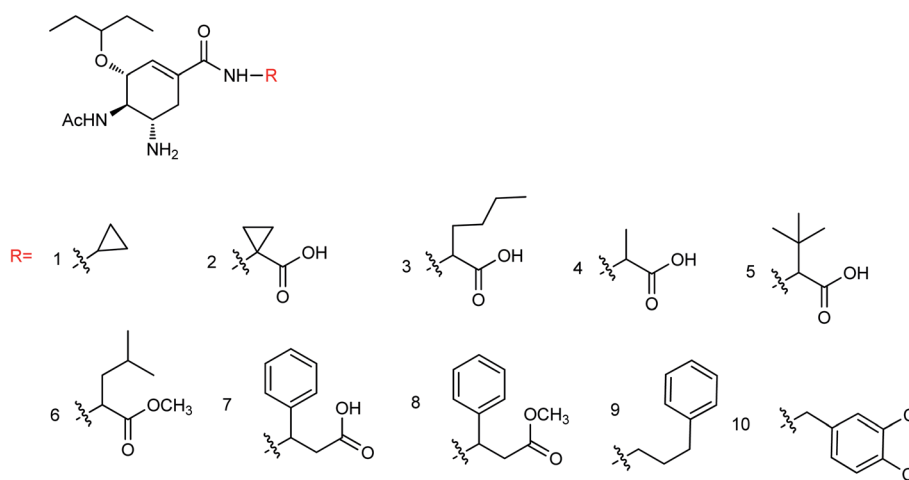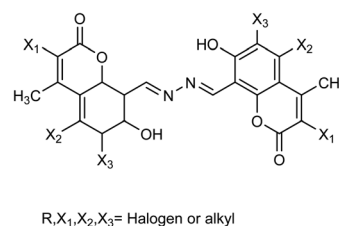
Fig. 63 Structures of terpenoids.

Fig. 64 Structures of oseltamivir derivatives.

mixed terpenes with austin skeleton structure are a kind of bioactive compounds produced by fungi which have certain inhibitory activities on aquatic pathogenic fungi, bacteria and Artemia. Li and co-workers[164] isolated and purified three new terpenoids (Fig. 63) from the fermentation products of marine fungus *Aspergillus ustus*. The molecular formulas of compounds **1–3** are $C_{33}H_{34}O_{12}$, $C_{32}H_{38}O_{11}$, and $C_{25}H_{28}O_8$. *In vitro* bioactivity experiments showed significant inhibitory activities for these terpenoids with IC_{50} values of 5.28 μM, 8.95 μM and 10.72 μM respectively, which are expected to become new potential anti-influenza drugs.

Ma and co-workers[165] found that the neuraminidase inhibitors oseltamivir phosphate and zanamivir displayed effective therapeutic effects on myocarditis model mice. After treatment with oseltamivir phosphate or zanamivir, the serum cTnI concentration was significantly reduced and the degree of myocardial injury was improved. The score of myocardial cases was significantly reduced and the degree of myocardial lesion was relieved; therefore, oseltamivir phosphate and zanamivir can be used to prepare drugs for the treatment of myocarditis. Ten novel neuraminidase inhibitors (Fig. 64) were prepared by the modification of ethyl carboxylate at the C-1 position of oseltamivir by Li and co-workers[166] with the yields ranging from 60.4 to 85.3%. The results indicated good inhibitory activities

against neuraminic acid A/Anhui/1/2005 (H5N1) for all deriva-tives and compounds **1–5** showed strong inhibitory activity. Compound **5** showed potent inhibitory activities among all the synthesized derivatives with an inhibition rate of 90.5%.

The most effective way to obtain effective antiviral drugs is from direct extraction of antiviral drugs from natural products or chemical modification of the extracts. Coumarins have excellent antiviral, antitumor and anticoagulant properties. Li[167] has used 4-methyl-7-hydroxy-8-formylcoumarin as a start-ing material reacting with hydrazine (NH_2–NH_2) at room temperature to yield *N*,*N*′-bis(7-hydroxycoumarin-8-methylene) hydrazine (Fig. 65). The synthesis was carried out in ethanol as a solvent without any catalyst at room temperature. *In vitro*

R,X₁,X₂,X₃= Halogen or alkyl

Fig. 65 Structures of *N*,*N*′-bis(4-methyl-7-hydroxycoumarin-8-methylene)hydrazine derivatives.
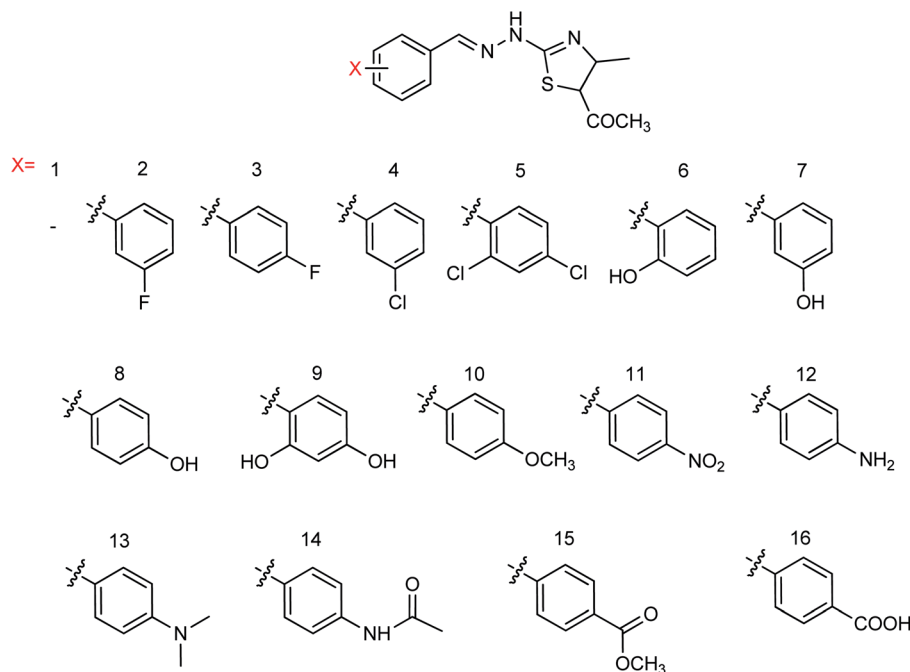
Fig. 66 Structures of 2-(2-benzylhydrazide)-5-acylthiazole derivatives.

study showed that the inhibition rate was 42.3% for N,N'-bis(4-methyl-7-hydroxycoumarin-8-methylene)hydrazine at a concentration of 30.0 μg mL^{-1}. The results indicated that N,N'-bis(4-methyl-7-hydroxycoumarin-8-methylene)hydrazine showed good inhibition activity and could be used to prepare further anti-influenza drugs.

Sixteen 2-(2-benzylhydrazide)-5-acylthiazole derivatives (Fig. 66) were designed and synthesized by Jia and co-workers[168] with the yields ranging from 52.4 to 92.1%. Compound **15** showed potent inhibitory activities with IC$_{50}$ values of 8.62 (H1N1) and 15.69 (H3N2) μg mL^{-1} at a concentration of 40.0 μg mL^{-1}.

Based on vanillin and its isomers, three novel 2-(2-benzylhydrazinyl)-5-acylthiazole derivatives (Fig. 67) bearing hydroxy and methoxy groups were designed and synthesized from acetylacetone by Wei and co-workers[169] with the yields ranging from 51.2 to 83.3%. The inhibition rate of compounds was determined at a concentration of 40.0 μg mL^{-1}. The results
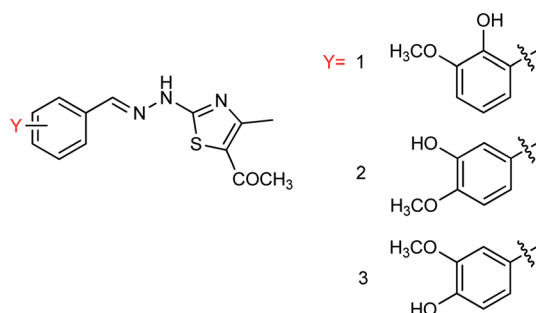
indicated that compound **3** has the best activity with an IC$_{50}$ value of 7.13 μg mL^{-1} and an inhibition rate up to 93.33%.

Tiliroside (Fig. 68), a flavonoid isolated and purified from traditional Chinese medicine *Duchesnea indica*, has a wide range of anti-inflammatory and antioxidant activities. However, there are few studies on the anti-influenza virus activity. Sheng co-workers[170] have obtained 4 mg tiliroside from 1 kg of *Duchesnea indica* plant. An *in vitro* study revealed that it can inhibit the oseltamivir-sensitive and drug-resistant influenza virus strains and reduce the replication of oseltamivir-sensitive and drug-resistant influenza virus strains. Tiroside showed inhibitory activity against oseltamivir-sensitive (197.9 μM L^{-1}) and drug-resistant neuraminidase (125.4 μM L^{-1}). At doses of 40 μM and 200 μM, the replication of oseltamivir sensitive influenza virus strain PR8 and resistant influenza virus strain H274Y was reduced respectively. At the same time, it was found that tiliroside did not produce cytotoxicity to MDCK at a dose of 1 mM. Tiliroside can be used as a safe and effective natural product to provide a theoretical basis for the preparation of
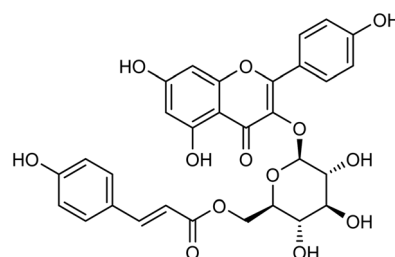

Fig. 67 Structures of 2-(2-benzylhydrazinyl)-5-acylthiazole derivatives.

Fig. 68 Structure of tiliroside.

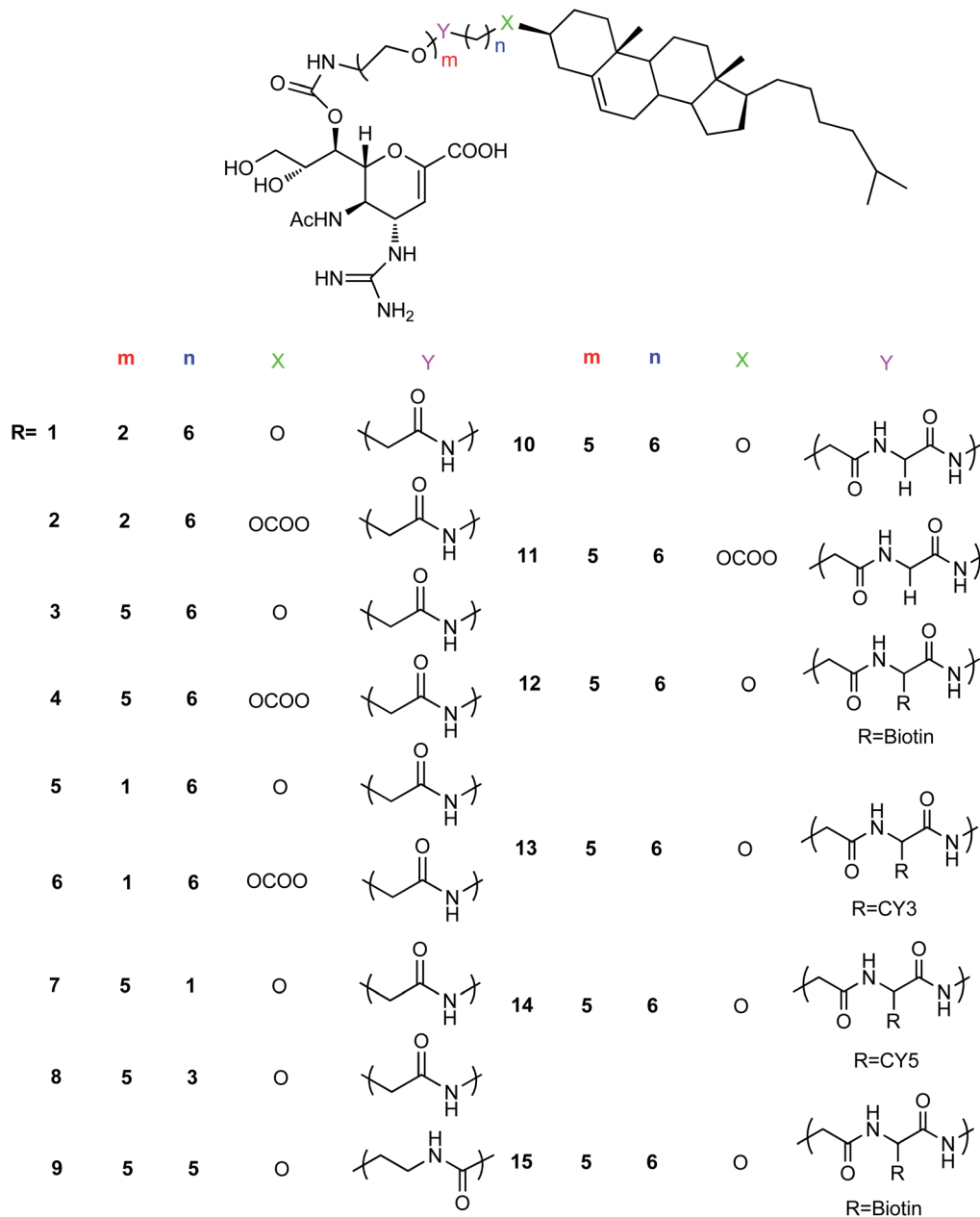| R=	m	n	X	Y		m	n	X	Y
1	2	6	O		10	5	6	O	
2	2	6	OCOO		11	5	6	OCOO	
3	5	6	O		12	5	6	O	R=Biotin
4	5	6	OCOO		13	5	6	O	R=CY3
5	1	6	O		14	5	6	O	R=CY5
6	1	6	OCOO		15	5	6	O	R=Biotin
7	5	1	O						
8	5	3	O						
9	5	5	O						

Fig. 69 Structures of anti-influenza derivatives.

influenza prevention and treatment drugs. It is expected to be promoted and used as a new neuraminidase inhibitor.

Fifteen new anti-influenza compounds (Fig. 69) were designed and synthesized by Chen and co-workers.[171] The synthesized derivatives showed good inhibitory activity against influenza virus (N2 (H3N2, E119V)) resistant to zanamivir. The fat water partition coefficient experiment showed that the lipid water partition coefficient of these compounds was significantly higher in comparison to zanamivir. It is expected to significantly improve the pharmacokinetics of zanamivir to be good potential agents.

In order to solve the problem of drug resistance caused by virus variants, Xiao and co-workers[172] designed and synthesized 15 novel acylhydrazone neuraminidase inhibitors (Fig. 70) with the yields ranging from 86 to 93%. *In vitro* study showed the best inhibitory activity for the compounds L8, L10, L14 and L15 with IC_{50} values of 0.60 μM, 7.31 μM, 6.20 μM and 5.29 μM respectively, which were significantly higher than oseltamivir acid (IC_{50} = 17.00 μM). SAR study has proved the importance of substitution for the improvement of the activity by introducing the same substituent at different positions; therefore, the activity of position 4 was higher than that of the substituent at position 2 such as compounds L1 and L5, compounds L4 and L15, compounds L7 and L8 and compounds L12 and L14 due to the binding mode and the number of hydrogen bonds formed between inhibitors and substrate amino acids.
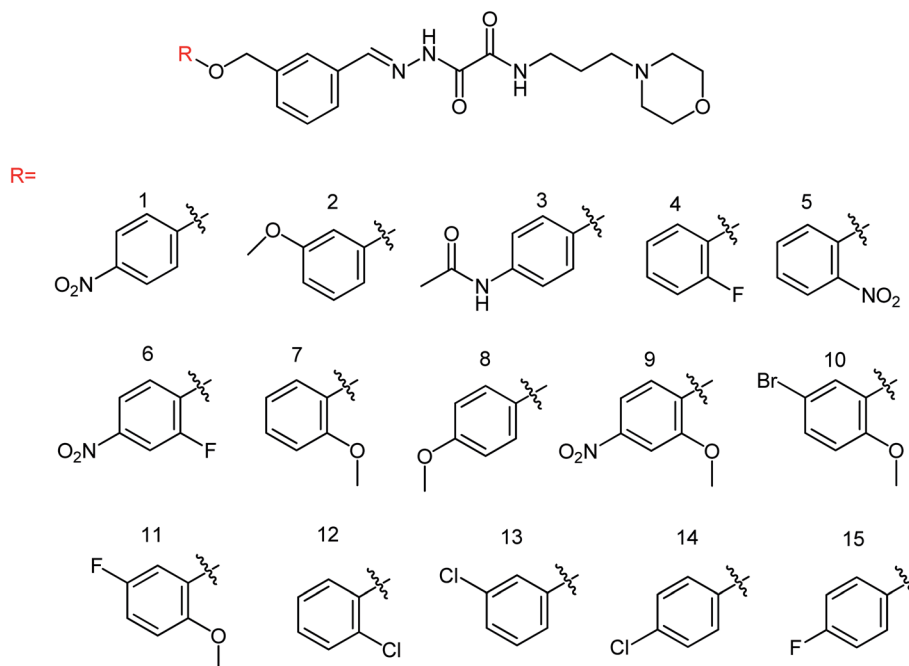
Fig. 70 Structures of acylhydrazone derivatives.

Naturally occurring anti-influenza compounds have received much attention in recent years due to their applications in medicinal chemistry and drug discovery. Actinomycetes produce active secondary metabolites with a novel structure and a unique mechanism due to their special living environment. Therefore, researchers pay more attention to find active ingredients from actinomycetes to prevent and treat the influenza disease. Streptomyces, the largest genus of actinomycetes, has attracted much attention due to their abundant secondary metabolites with significant bioactivities. A class of sek15 polyketides with neuraminidase inhibitory activities were isolated from *Streptomyces* sp. by Liu and co-workers.[173] These three compounds (Srepolyketide B, Srepolyketide C and SEK15)

showed inhibitory activities with IC_{50} values of 85.6 μM, 58.2 μM and 51.3 μM respectively (Fig. 71).

It has been proved that thiourea compounds have inhibitory activities against influenza virus. On this basis, Hu and co-workers[174] have designed and synthesized three thiourea derivatives (Fig. 72) and determined their bioactivity against influenza virus (H1N1). The inhibitory rate of these three compounds were 70.13 ± 7.05, 68.3 ± 5.66 and $56.59 \pm 6.21\%$ respectively with the IC_{50} values of 22.78 ± 3.73, 21.76 ± 5.22 and 32.69 ± 10.46 μM respectively.

Five vanillin derivatives (Fig. 73) were designed and synthesized by Hu and co-workers[175] with the yields ranging from 79.6 to 95.0%. The synthesized derivatives showed potent inhibitory
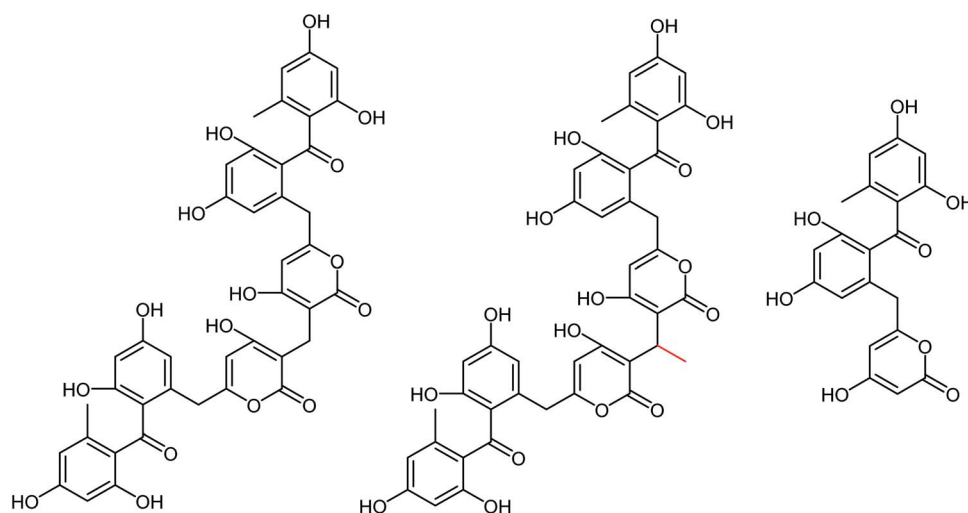

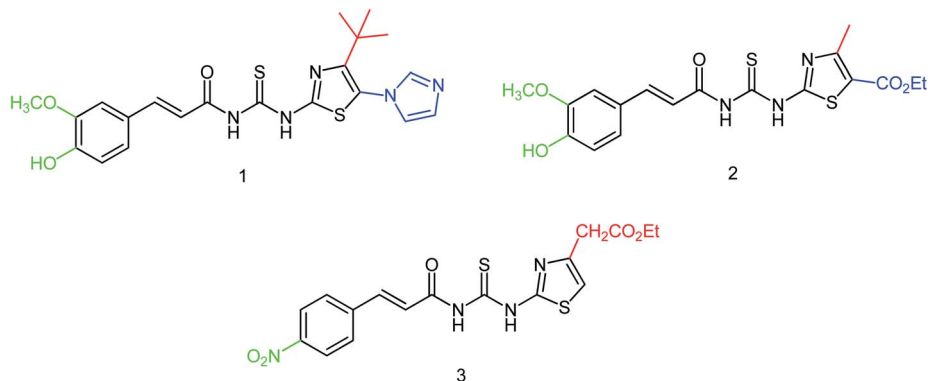**Fig. 71** Structures of Srepolyketide B, Srepolyketide C and SEK15.

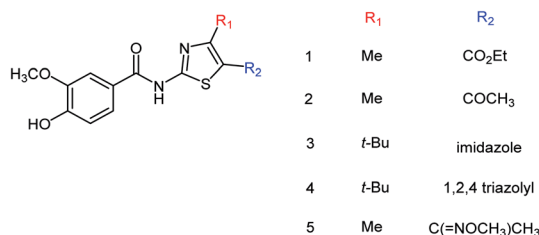**Fig. 72** Structures of thiourea derivatives.

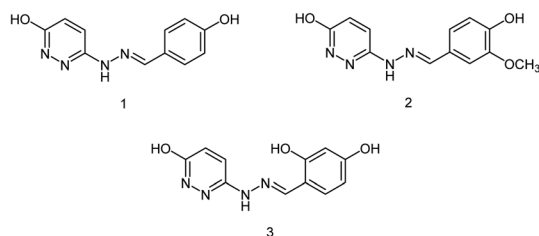**Fig. 73** Structures of vanillin derivatives.

Fig. 74 Structures of pyridazine hydrazone derivatives.

activities and may be used to prepare further inhibitors. Compound 5 showed the best activity with an IC_{50} value of 12.33 $\pm$ 3.01 μM.

In another study, Hu and co-workers[176] have designed and synthesized three pyridazine hydrazone derivatives (Fig. 74), and *in vitro* determination of their anti-influenza H1N1 activity has been investigated. The yields were ranging from 71.9 to 86.9%. Compound 3 showed significant inhibitory activities with the inhibition rate of 84.52 $\pm$ 1.32% and an IC_{50} value of 11.17 $\pm$ 0.34 μM.

7. Conclusions

Influenza virus (IFV) is a respiratory pathogen that can cause life-threatening diseases such as pneumonia and encephalitis when aggravated. Due to influenza infections, occasional, seasonal, and pandemic influenza transmission occurs in humans as well as animals and spreads easily *via* droplets and contact. Neuraminidase inhibitors are widely used in the treatment of influenza infection, and there is an urgent need to develop more potent agents due to drug resistance activities. This review has discussed various developments in the field of neuraminidase inhibitors along with their medicinal properties. It also focuses on the recent advances of chemical synthesis pathways used for the development of new neuraminidase agents along with medicinal aspects of chemically modified molecules. Hence, this review presents a strong intellectual background in the said field, which gives an in-depth insight into several aspects of this global pandemic and will provide ample references for the researchers, which may be useful for the ongoing development of therapeutic agents and vaccines.

Abbreviations

ALI	Acute lung injury
ADMET	Absorption, distribution, metabolism, excretion and toxicity
ApoE	Apolipoprotein E
BNA	Bacterial neuraminidase
BPD	Berberine-piperazine derivatives
BCCAO	Bilateral common carotid artery occlusion
bNAb	Broadly neutralizing antibodies
Bu	butanol
DFT	Density functional theory
DANA	2-Deoxy-2,3-didehydro-*N*-acetylneuraminic acid
DNA	Deoxyribonucleic acid
DS	*Dianthus superbus*
DW	Distilled water
FESR	Flavonoids-enriched extract of *S. baicalensis*
GOC	Guanidino OC
GOC	Guanidino-oseltamivir carboxylic acid
HN	Haemagglutinin-neuraminidase
HA	Hemagglutinin
HEK 293T	Human embryonic kidney cells 293
hPIV	Human para influenza virus
hPIV-3	Human parainfluenza virus type-3
IAV	Influenza A virus
LGN	Liquiritigenin
MDCK	Madin–Darby canine kidney

MGO	Methylglyoxal
MUNANA	20-(4-Methylumbelliferyl)-α-D-N-acetylneuraminic acid
MD	Molecular dynamics
MIP	Molecularly imprinted polymer
M2	Matrix protein 2
Neu5Ac	N-Acetylneuraminic acid
N-Acyl-Neu2en	N-Acyl-2,3-dehydro-2-deoxy-neuraminic acid
NPs	Nanoparticles
NA	Neuraminidase
NEU1	Neuraminidase 1
NEU2	Neuraminidase 2
NEU3	Neuraminidase 3
NIP@silica gel	Non-imprinted polymer with silica gel
OPME	One-pot multienzyme
OS	Oseltamivir
OTV	Oseltamivir
OC	Oseltamivir carboxylate
OSMIP	Oseltamivir molecularly imprinted polymer
Se@OTV	OTV decoration of SeNPs
PmAldolase	*Pasteurella multocida* sialic acid aldolase
RNA	Ribonucleic acid
RdRp	RNA-dependent RNA polymerase
SEM	Scanning electron microscopy
OSMIP@silica gel	Silica gel coated with an MIP layer for OS
SSAA	Specific surface area analysis
SpNanB	*Streptococcus pneumoniae* sialidase
SAR	Structure–activity relationship
TPE4S	Tetraphenylethene luminogen
3WJ	Three–way junction
VLDL	Very-low-density lipoprotein
VCNA	*Vibrio cholerae* neuraminidase

Conflicts of interest

The authors declare that they have no conflict of interests in this work.

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References

1 D. Lin, Y.-J. Yi, M.-W. Xiao, J. Chen, J. Ye, A.-X. Hu, W.-W. Lian, A.-L. Liu and G.-H. Du, *J. Asian Nat. Prod. Res.*, 2019, **21**, 1052–1067, DOI: 10.1080/10286020.2018.1509854.

2 A. J. Hay, A. J. Wolstenholme, J. J. Skehel and M. H. Smith, *EMBO J.*, 1985, **4**, 3021–3024, DOI: 10.1002/j.1460-2075.1985.tb04038.x.

3 A. J. Hay, *Semin. Virol.*, 1992, **3**, 21–30.

4 F. G. Hayden, A. Minocha, D. A. Spyker and H. E. Hoffman, *Antimicrob. Agents Chemother.*, 1985, **28**, 216–221, DOI: 10.1128/AAC.28.2.216.

5 F. G. Hayden and R. B. Couch, *Rev. Med. Virol.*, 1992, **2**, 89–96, DOI: 10.1002/rmv.1980020205.

6 F. G. Hayden, E. B. Belshe, R. D. Clover, A. J. Hay, M. G. Oakes and W. Soo, *N. Engl. J. Med.*, 1989, **321**, 1696–1702, DOI: 10.1056/NEJM198912213212502.

7 S. Grambas, M. S. Bennett and A. J. Hay, *Virology*, 1992, **191**, 541–549, DOI: 10.1016/0042-6822(92)90229-i.

8 J. L. McKimm-Breschkin, Influenza Other Respir, *Viruses*, 2012, **7**, 25–36, DOI: 10.1111/irv.12047.

9 Y. Li, Z. Lin, M. Zhao, T. Xu, C. Wang, L. Hua, H. Wang, H. Xia and B. Zhu, *ACS Appl. Mater. Interfaces*, 2016, **8**, 24385–24393.

10 C.-Z. Wang, H.-H. Han, X.-Y. Tang, D.-M. Zhou, C. Wu, G.-R. Chen, X.-P. He and H. Tian, *ACS Appl. Mater. Interfaces*, 2017, **9**, 25164–25170.

11 A. Moscona, *N. Engl. J. Med.*, 2005, **353**, 1363–1373.

12 A. F. Abdel-Magid, C. A. Maryanoff and S. J. Mehrman, *Curr. Opin. Drug Discovery Dev.*, 2001, **4**, 776–791.

13 V. Y. Glanz, V. A. Myasoedova, A. V. Grechko and A. N. Orekhov, *Drug Des., Dev. Ther.*, 2018, **12**, 3431–3437, DOI: 10.2147/DDDT.S176220.

14 J. Magano, *Chem. Rev.*, 2009, **109**, 4398–4438, DOI: 10.1021/cr800449m.

15 V. Farina and J. D. Brown, *Angew. Chem., Int. Ed.*, 2006, **45**, 7330–7334, DOI: 10.1002/anie.200602623.

16 M. Shibasaki and M. Kanai, *Eur. J. Org. Chem.*, 2008, 1839–1850, DOI: 10.1002/ejoc.200800033.

17 N. Anuwongcharoen, W. Shoombuatong, T. Tantimongcolwat, V. Prachayasittikul and C. Nantasenamat, *PeerJ*, 2016, **4**, e1958, DOI: 10.7717/peerj.1958.

18 A. Moscona, *N. Engl. J. Med.*, 2009, **360**, 953–956, DOI: 10.1056/NEJMp0900648.

19 M. Itzstein, W.-Y. Wu and B. Jin, *Carbohydr. Res.*, 1994, **259**, 301–305, DOI: 10.1016/0008-6215(94)84065-2.

20 M. Itzstein, B. Jin, W.-Y. Wu and M. Chandler, *Carbohydr. Res.*, 1993, **244**, 181–185, DOI: 10.1016/0008-6215(93)80014-6.

21 E. Schreiner, E. Zbiral, R. G. Kleineidam and R. Schauer, *Liebigs Ann. Chem.*, 1991, **1991**, 129–134, DOI: 10.1002/jlac.199119910124.

22 K. Kim, Y.-T. Lin and H. S. Mosher, *Tetrahedron Lett.*, 1988, **29**, 3183–3186, DOI: 10.1016/0040-4039(88)85116-5.

23 J. Scheigetz, R. Zamboni, M. A. Bernstein and B. Roy, *Org. Prep. Proced. Int.*, 1995, **27**, 637–644, DOI: 10.1080/00304949509458521.

24 M. Chandler, M. J. Bamford, R. Conroy, B. Lamont, B. Patel, V. K. Patel, I. P. Steeples, R. Storer, N. G. Weir, M. Wright and C. Williamson, *J. Chem. Soc., Perkin Trans. 1*, 1995, 1173–1180, DOI: 10.1039/P19950001173.

25 K.-G. Liu, S. Yan, Y.-L. Wu and Z.-J. Yao, *Org. Lett.*, 2004, **6**, 2269–2272, DOI: 10.1021/ol0491890.

26 C. U. Kim, W. Lew, M. A. Williams, H. Liu, L. Zhang, S. Swaminathan, N. Bischofberger, M. S. Chen, D. B. Mendel, C. Y. Tai, W. G. Laver and R. C. Stevens, *J. Am. Chem. Soc.*, 1997, **119**, 681–690, DOI: 10.1021/ja963036t.

27 J. C. Rohloff, K. M. Kent, M. J. Postich, M. W. Becker, H. H. Chapman, D. E. Kelly, W. Lew, M. S. Louie, L. R. McGee, E. J. Prisbe, L. M. Schultze, R. H. Yu and L. Zhang, *J. Org. Chem.*, 1998, **63**, 4545–4550, DOI: 10.1021/jo980330q.

28 M. Federspiel, R. Fischer, M. Hennig, H.-J. Mair, T. Oberhauser, G. Rimmler, T. Albiez, J. Bruhin, H. Estermann, C. Gandert, V. Göckel, S. Götzö, U. Hoffmann, G. Huber, G. Janatsch, S. Lauper, O. Röckel-Stäbler, R. Trussardi and A. G. Zwahlen, *Org. Process Res. Dev.*, 1999, **34**, 266–274, DOI: 10.1021/op9900176.

29 M. Karpf and R. Trussardi, *J. Org. Chem.*, 2001, **66**, 2044–2051, DOI: 10.1021/jo005702l.

30 S. Abrecht, P. Harrington, H. Iding, M. Karpf, R. Trussardi, B. Wirz and U. Zutter, *Chimia*, 2004, **58**, 621–629, DOI: 10.2533/000942904777677605.

31 P. J. Harrington, J. D. Brown, T. Foderaro and R. C. Hughes, *Org. Process Res. Dev.*, 2004, **8**, 86–91, DOI: 10.1021/op0302107.

32 Y. S. Hong and E. J. Corey, *J. Am. Chem. Soc.*, 2006, **128**, 6310–6311, DOI: 10.1021/ja0616433.

33 N. T. Kipassa, H. Okamura, K. Kina, T. Hamada and T. Iwagawa, *Org. Lett.*, 2008, **105**, 815–816, DOI: 10.1021/ol7029646.

34 Y. Fukuta, T. Mita, N. Fukuda, M. Kanai and M. D. Shibasaki, *J. Am. Chem. Soc.*, 2006, **128**, 6312–6313, DOI: 10.1021/ja061696k.

35 T. Mita, N. Fukuda, F. X. Roca, M. Kanai and M. Shibasaki, *Org. Lett.*, 2007, **9**, 259–262, DOI: 10.1021/ol062663c.

36 K. Yamatsugu, S. Kamijo, Y. Suto, M. Kanai and M. Shibasaki, *Tetrahedron Lett.*, 2007, **48**, 1403–1406, DOI: 10.1016/j.tetlet.2006.12.093.

37 K. Yamatsugu, L. Yin, S. Kamijo, Y. Kimura, M. Kanai and M. Shibasaki, *Angew. Chem., Int. Ed.*, 2009, **48**, 1070–1076, DOI: 10.1002/anie.200804777.

38 X. Cong and Z.-J. Yao, *J. Org. Chem.*, 2006, **71**, 5365–5368, DOI: 10.1021/jo060633h.

39 N. Satoh, T. Akiba, S. Yokoshima and T. Fukuyama, *Angew. Chem., Int. Ed.*, 2007, **46**, 5734–5736, DOI: 10.1002/anie.200701754.

40 N. Satoh, T. Akiba, S. Yokoshima and T. Fukuyama, *Tetrahedron*, 2009, **65**, 3239–3245, DOI: 10.1016/j.tet.2008.09.103.

41 J.-J. Shie, J.-M. Fang, S.-Y. Wang, K.-C. Tsai, Y.-S. E. Cheng, A.-S. Yang, S.-C. Hsiao, C.-Y. Su and C.-H. Wong, *J. Am. Chem. Soc.*, 2007, **129**, 11892–11893, DOI: 10.1021/ja073992i.

42 J.-J. Shie, J.-M. Fang and C.-H. Wong, *Angew. Chem., Int. Ed.*, 2008, **47**, 5788–5791, DOI: 10.1002/anie.200801959.

43 K. M. Bromfield, H. Graden, D. P. Hagberg, T. Olsson and N. Kann, *Chem. Commun.*, 2007, **30**, 3183–3185, DOI: 10.1039/B703295A.

44 B. M. Trost and T. Zhang, *Angew. Chem., Int. Ed.*, 2008, **47**, 3759–3761, DOI: 10.1002/anie.200800282.

45 M. Matveenko, M. G. Banwell and A. C. Willis, *Tetrahedron Lett.*, 2008, **49**, 7018–7020, DOI: 10.1016/j.tetlet.2008.09.130.

46 L.-D. Nie and X.-X. Shi, *Tetrahedron: Asymmetry*, 2009, **20**, 124–129, DOI: 10.1016/j.tetasy.2008.11.027.

47 L.-D. Nie, X.-X. Shi, K. H. Ko and W.-D. Lu, *J. Org. Chem.*, 2009, **74**, 3970–3973, DOI: 10.1021/jo900218k.

48 H. Ishikawa, T. Suzuki and Y. Hayashi, *Angew. Chem., Int. Ed.*, 2009, **48**, 1304–1307, DOI: 10.1002/anie.200804883.

49 T. Mandai and T. Oshitari, *Synlett*, 2009, 783–786, DOI: 10.1055/s-0028-1087941.

50 T. Oshitari and T. Mandai, *Synlett*, 2009, 787–789, DOI: 10.1055/s-0028-1087940.

51 F. Krammer, G. J. D. Smith, R. A. M. Fouchier, M. Peiris, K. Kedzierska, P. C. Doherty, P. Palese, M. L. Shaw, J. Treanor, R. G. Webster and A. García-Sastre, Influenza, *Nat Rev Dis Primers.*, 2018, **4**, 3, DOI: 10.1038/s41572-018-0002-y.

52 M. Billings, *The Influenza Pandemic of 1918*, 2005, http://virus.stanford.edu/uda/.

53 Centers for Disease Control and Prevention (CDC), *2009 H1N1 Pandemic (H1N1pdm09 virus)*, 2019, https://www.cdc.gov/flu/pandemic-resources/2009-h1n1-pandemic.html.

54 European Centre for Disease Prevention and Control, *Expert opinion on neuraminidase inhibitors for the prevention and treatment of influenza – review of recent systematic reviews and meta-analyses*, ECDC, Stockholm, 2017, https://www.ecdc.europa.eu/sites/portal/files/documents/Scientific-advice-neuraminidase-inhibitors-2017.pdf.

55 E. Chen, Y. Chen, L. Fu, Z. Chen, Z. Gong, H. Mao, D. Wang, M. Y. Ni, P. Wu, Z. Yu, T. He, Z. Li, J. Gao, S. Liu, Y. Shu, B. J. Cowling, S. Xia and H. Yu, *Eurosurveillance*, 2013, **18**, 20616.

56 K. Zhao, M. Gu, L. Zhong, Z. Duan, Y. Zhang, Y. Zhu, G. Zhao, M. Zhao, Z. Chen, S. Hu, W. Liu, X. Liu, D. Peng and X. Liu, *Vet. Microbiol.*, 2013, **163**, 351–357, DOI: 10.1016/j.vetmic.2012.12.025.

57 U. Joseph, M. Linster, Y. Suzuki, S. Krauss, R. A. Halpin, D. Vijaykrishna, T. P. Fabrizio, T. M. Bestebroer, S. Maurer-Stroh, R. J. Webby, D. E. Wentworth, R. A. M. Fouchier, J. Bahl and G. J. D. Smith, *J. Virol.*, 2015, **89**, 2442–2447, DOI: 10.1128/JVI.02590-14.

58 M. Gu, W. Liu, Y. Cao, D. Peng, X. Wang, H. Wan, G. Zhao, Q. Xu, W. Zhang, Q. Song, Y. Li and X. Liu, *Emerging Infect. Dis.*, 2011, **17**, 1060–1063, DOI: 10.3201/eid1706.101406.

59 H. Liu, C. Xiong, J. Chen, G. Chen, J. Zhang, Y. Li, Y. Xiong, R. Wang, Y. Cao, Q. Chen, D. Liu, H. Wang and J. Chen, *Emerging Microbes Infect.*, 2018, **7**, 62, DOI: 10.1038/s41426-018-0064-7.

60 N. Beerens, R. Heutink, F. Harders, A. Bossers, G. Koch and B. Peeters, *J. Virol.*, 2020, **94**, e01818–19, DOI: 10.1128/JVI.01818-19.

61 T. H. P. Peacock, J. James, J. E. Sealy and M. Iqbal, *Viruses*, 2019, **11**, 620, DOI: 10.3390/v11070620.

62 E. Montomoli and C. M. Trombetta, *Pathog. Global Health*, 2014, **108**, 213, DOI: 10.1179/2047772414Z.000000000215.

63 P. A. Rota, T. R. Wallis, M. W. Harmon, J. S. Rota, A. P. Kendal and K. Nerome, *Virology*, 1990, **175**, 59–68, DOI: 10.1016/0042-6822(90)90186-U.

64 G. Yuanji, J. Fengen, W. Ping, W. Min and Z. Jiming, *J. Gen. Virol.*, 1983, **64**, 177–182.

65 B. M. Hause, M. Ducatez, E. A. Collin, Z. Ran, R. Liu, Z. Sheng, A. Armien, B. Kaplan, S. Chakravarty, A. D. Hoppe, R. J. Webby, R. R. Simonson and F. Li, *PLoS Pathog.*, 2013, **9**, e1003176, DOI: 10.1371/journal.ppat.1003176.

66 *Flu Symptoms & Complications*, https://www.cdc.gov/flu/symptoms/symptoms.htm.

67 S. W. Boktor and J. W. Hafner, Influenza. [Updated 2020 May 26], in *StatPearls [Internet]. Treasure Island (FL)*, StatPearls Publishing; 2020 Jan, https://www.ncbi.nlm.nih.gov/books/NBK459363/.

68 V. Kumar, *Indian J. Pediatr.*, 2017, **84**, 139–143, DOI: 10.1007/s12098-016-2232-x.

69 V. K. Trombetta, Y. L. Chan and M. J. Bankowski, *Hawaii J. Med. Public Health*, 2018, **77**, 226–230.

70 E. Egilmezer, G. J. Walker, P. Bakthavathsalam, J. R. Peterson, J. J. Gooding, W. Rawlinson and S. Stelzer-Braid, *Rev. Med. Virol.*, 2018, **28**, e1995, DOI: 10.1002/rmv.1995.

71 S. H. Choi, J. W. Chung, T. Kim, K. H. Park, M. S. Lee and Y. G. Kwak, *Korean J. Intern. Med.*, 2018, **33**, 391–396, DOI: 10.3904/kjim.2016.226.

72 D. Dou, R. Revol, H. Östbye, H. Wang and R. Daniels, *Front. Immunol.*, 2018, **9**, 1581, DOI: 10.3389/fimmu.2018.01581.

73 T. O. Edinger, M. O. Pohl and S. Stertz, *J. Gen. Virol.*, 2014, **95**, 263–277, DOI: 10.1099/vir.0.059477-0.

74 Y. A. Shtyrya, L. V. Mochalova and N. V. Bovin, *Acta Naturae*, 2009, **1**, 26–32.

75 A. G. Winquist, K. Fukuda, C. B. Bridges and N. J. Cox, *Morb. Mortal. Wkly. Rep.*, 1999, **48**, 1–9.

76 A. J. Burnham, T. Baranovich and E. A. Govorkova, *Antiviral Res.*, 2013, **100**, 520–534, DOI: 10.1016/j.antiviral.2013.08.023.

77 E. J. LaVoie, H. Y. Sagong, J. D. Bauman, A. Nogales, L. Martínez-Sobrido and E. Arnold, *ChemMedChem*, 2019, **14**, 1204–1223, DOI: 10.1002/cmdc.201900084.

78 W. Li, A. Santra, H. Yu, T. J. Slack, M. M. Muthana, D. Shi, Y. Liu and X. Chen, *J. Org. Chem.*, 2019, **84**, 6697–6708, DOI: 10.1021/acs.joc.9b00385.

79 C. Wu, K.-J. Wu, J.-B. Liu, X.-M. Zhou, C.-H. Leung and D.-L. Ma, *Chem. Commun.*, 2019, **55**, 6353–6356, DOI: 10.1039/c9cc02189b.

80 C. Aiyu, L. Yongdong, Y. Jiao, H. Aixi, L. Wenwen, L. Ailin and D. Guanhua, *Chem. Res. Chin. Univ.*, 2019, **35**, 395–402, DOI: 10.1007/s40242-019-8346-8.

81 M.-Y. Cui, J.-X. Nie, Z.-Z. Yan, M.-W. Xiao, D. Lin, J. Ye and A.-X. Hu, *Med. Chem. Res.*, 2019, **28**, 938–947, DOI: 10.1007/s00044-019-02343-3.

82 V. P. Chibanga, L. Dirr, P. Guillon, I. M. El-Deeb, B. Bailly, R. J. Thomson and M. von Itzstein, *Antiviral Res.*, 2019, **167**, 89–97, DOI: 10.1016/j.antiviral.2019.04.001.

83 Z. X. Zhao, L. P. Cheng, M. Li, W. Pang and F. H. Wu, *Eur. J. Med. Chem.*, 2019, **173**, 305–313, DOI: 10.1016/j.ejmech.2019.04.006.

84 Z.-L. Yan, A.-Y. Liu, X.-X. Wei, Z. Zhang, L. Qin, Q. Yu, P. Yu, K. Lu and Y. Yang, *Carbohydr. Res.*, 2019, **477**, 32–38, DOI: 10.1016/j.carres.2019.03.012.

85 M. Kiso, T. J. S. Lopes, S. Yamayoshi, M. Ito, M. Yamashita, N. Nakajima, H. Hasegawa, G. Neumann and Y. Kawaoka, *J. Infect. Dis.*, 2018, **217**, 887–896, DOI: 10.1093/infdis/jix606.

86 K. D. Malbari, A. S. Chintakrindi, L. R. Ganji, D. J. Gohil, S. T. Kothari, M. V. Joshi and M. A. Kanyalkar, *Mol. Diversity*, 2019, **23**, 927–951, DOI: 10.1007/s11030-019-09919-6.

87 B. Sathya and M. Prasath, *Res. Chem. Intermed.*, 2019, **45**, 2135–2166, DOI: 10.1007/s11164-018-03727-7.

88 P. L. Rocca, P. Rota, M. Piccoli, F. Cirillo, M. Orioli, A. Ravelli, P. Allevi and L. Anastasia, *J. Org. Chem.*, 2019, **84**, 5460–5470, DOI: 10.1021/acs.joc.9b00431.

89 M. Yamabea, A. Fujitaa, K. Kaihatsub and Y. Ebara, *Carbohydr. Res.*, 2019, **474**, 43–50, DOI: 10.1016/j.carres.2019.01.008.

90 J. Han, J. Perez, A. Schafer, H. Cheng, N. Peet, L. Rong and B. Manicassamy, *Curr. Med. Chem.*, 2018, **25**, 5115–5127.

91 B.-T. Hong, Y.-S. E. Cheng, T.-J. Cheng and J.-M. Fang, *Eur. J. Med. Chem.*, 2019, **163**, 710–721, DOI: 10.1016/j.ejmech.2018.12.027.

92 Y. Ali, N. M. Bunnori, D. Susanti, A. M. Alhassan and S. A. Hamid, *Front. Chem.*, 2018, **6**, 210, DOI: 10.3389/fchem.2018.00210.

93 A. A. A. Abdalsalam, *J. Appl. Pharm. Sci.*, 2017, **7**, 024–033.

94 R. P. Putra, R. Imaniastuti, M. A. F. Nasution, D. Kerami and U. S. F. Tambunan, *IOP Conf. Ser.: Mater. Sci. Eng.*, 2018, **349**, 012052, DOI: 10.1088/1757-899X/349/1/012052.

95 S. Mathew, A. A. Al-Thani and H. M. Yassine, *PLoS One*, 2018, **13**, e0203148, DOI: 10.1371/journal.pone.0203148.

96 T. Guo, R. Héon-Roberts, C. Zou, R. Zheng, A. V. Pshezhetsky and C. W. Cairo, *J. Med. Chem.*, 2018, **61**, 11261–11279.

97 N. Onul, O. Ertik, N. G. Mermer and R. Yanardag, *J. Chem.*, 2018, 4386031, DOI: 10.1155/2018/4386031.

98 Y. Li, Z. Lin, M. Guo, M. Zhao, Y. Xia, C. Wang, T. Xu and B. Zhu, *Int. J. Nanomed.*, 2018, **13**, 2005–2016.

99 Y. Li, Z. Lin, M. Guo, Y. Xia, M. Zhao, C. Wang, T. Xu, T. Chen and B. Zhu, *Int. J. Nanomed.*, 2017, **12**, 5733–5743.

100 L. P. Cheng, T. C. Wang, R. Yu, M. Li and J. W. Huang, *Bioorg. Med. Chem. Lett.*, 2018, **28**, 23–24, DOI: 10.1016/j.bmcl.2018.10.040.

101 T. J. Slack, W. Li, D. Shi, J. B. McArthur, G. Zhao, Y. Li, A. Xiao, Z. Khedri, H. Yu, Y. Liu and X. Chen, *Bioorg. Med. Chem.*, 2018, **26**, 5751–5757, DOI: 10.1016/j.bmc.2018.10.028.

102 M. R. F. Pratama and T. Gusdinar, *Asian J. Pharm. Clin. Res.*, 2017, **10**, 304–308, DOI: 10.22159/ajpcr.2017.v10i8.18667.

103 Y.-J. Yang, X.-W. Liu, X.-J. Kong, Z. Qin, Z.-H. Jiao, S.-H. Li and J.-Y. Li, *Molecules*, 2018, **23**, 1881, DOI: 10.3390/molecules23081881.

104 B. Wang, K. Wang, P. Meng, Y. Hu, F. Yang, K. Liu, Z. Lei, B. Chen and Y. Tian, *Bioorg. Med. Chem. Lett.*, 2018, **26**, 5751–5757, DOI: 10.1016/j.bmcl.2018.09.014.

105 E. J. White, G. Gyulay, S. Lhoták, M. M. Szewczyk, T. Chong, M. T. Fuller, O. Dadoo, A. E. Fox-Robichaud, X. R. C. Austin, B. L. Trigatti and S. A. Igdoura, *J. Biol. Chem.*, 2018, **293**, 14689–14706.

106 C. Colombo, Č. Podlipnik, L. Lo Presti, M. Niikura, A. J. Bennet and A. Bernardi, *PLoS One*, 2018, **13**, e0193623, DOI: 10.1371/journal.pone.0193623.

107 G. Liu, B. Wang, Y. Zhang, G. Xing, X. Yang and S. Wang, *Chem. Commun.*, 2018, **54**, 10691–10694, DOI: 10.1039/C8CC06300A.

108 F. Meng, Y. Yin, C. Lu, Y. Duan, Y. Zhu, C. Huang, Y. Zhou, X. Xu and F. Wu, *Monatsh. Chem.*, 2018, **149**, 2037–2046, DOI: 10.1007/s00706-018-2245-4.

109 Y.-C. Xie, B. Huang, K.-X. Yu, F.-Y. Shi, T.-Q. Liu and W.-F. Xu, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 3556–3560, DOI: 10.1016/j.bmcl.2013.04.033.

110 B. R. Kim, J.-Y. Park, H. J. Jeong, H.-J. Kwon, S.-J. Park, I.-C. Lee, Y. B. Ryu and W. S. Lee, *J. Enzyme Inhib. Med. Chem.*, 2018, **33**, 1256–1265, DOI: 10.1080/14756366.2018.1488695.

111 G. Enkhtaivan, D. H. Kim, G. S. Park, M. Pandurangan, D. A. Nicholas, S. H. Moon, A. A. Kadam, R. V. Patel, H.-S. Shin and B. M. Mistry, *Int. J. Biol. Macromol.*, 2018, **119**, 1204–1210, DOI: 10.1016/j.ijbiomac.2018.08.047.

112 W. Liu, S. K. Samanta, B. D. Smith and L. Isaacs, *Chem. Soc. Rev.*, 2017, **46**, 2391–2403, DOI: 10.1039/c7cs00011a.

113 A. Xiao, T. J. Slack, Y. Li, D. Shi, H. Yu, W. Li, Y. Liu and X. Chen, *J. Org. Chem.*, 2018, **83**, 10798–10804.

114 J. Zhang, V. Poongavanam, D. Kang, C. Bertagnin, H. Lu, X. Kong, H. Ju, X. Lu, P. Gao, Y. Tian, H. Jia, S. Desta, X. Ding, L. Sun, Z. Fang, B. Huang, X. Liang, R. Jia, X. Ma, W. Xu, N. A. Murugan, A. Loregian, B. Huang, P. Zhan and X. Liu, *J. Med. Chem.*, 2018, **61**, 6379–6397.

115 M. R. Richards, T. Guo, C. D. Hunter and C. W. Cairo, *Bioorg. Med. Chem.*, 2018, **26**, 5349–5358.

116 S. Kosono, A. Kasai, S. Komaba, T. Matsubara, T. Sato, D. Takahashi and K. Toshima, *Chem. Commun.*, 2018, **54**, 7467–7470, DOI: 10.1039/C8CC03865A.

117 P.-H. Hsu, D.-C. Chiu, K.-L. Wu, P.-S. Lee, J.-T. Jan, Y.-S. E. Cheng, K.-C. Tsai, T.-J. Cheng and J.-M. Fang, *Eur. J. Med. Chem.*, 2018, **154**, 314–323, DOI: 10.1016/j.ejmech.2018.05.030.

118 C. Lu, Y. Yin, F. Meng, Y. Dun, K. Pei, C. Wang, X. Xu and F. Wu, *Bioorg. Med. Chem. Lett.*, 2018, **28**, 2003–2007.

119 S. Charyasriwong, T. Haruyama and N. Kobayashi, *Drug Discoveries Ther.*, 2016, **10**, 201–210.

120 M. Pascolutti, L. Dirr, P. Guillon, A. V. DenBergh, T. Ve, R. J. Thomson and M. von Itzstein, *ACS Chem. Biol.*, 2018, **13**, 1544–1550, DOI: 10.1021/acschembio.8b00150.

121 H. Kaur and B. Narasimhan, *Curr. Top. Med. Chem.*, 2018, **18**, 3–21.

122 J. Jin, Y. Chen, D. Wang, L. Ma, M. Guo, C. Zhou and J. Dou, *Arch. Pharmacal Res.*, 2018, **41**, 664–676, DOI: 10.1007/s12272-018-1022-6.

123 A. Hafeez, M. Roy, P. K. Pal, R. Kumar, P. Gangwar and A. Mohan, *Int. J. Biol., Pharm. Allied Sci.*, 2015, **4**, 5277–5283.

124 Á. Hadházi, L. Li, B. Bailly, A. Maggioni, G. Martin, L. Dirr, J. Dyason, R. Thomson, G. Gao, A. Borbás, T. Ve, M. Pascolutti and M. von Itzstein, *ChemMedChem*, 2018, **13**, 785–789, DOI: 10.1002/cmdc.201800092.

125 X. Wu, X. Wu, Q. Sun, C. Zhang, S. Y. Yang, L. Li and Z. Jia, *Theranostics*, 2017, **7**, 826–845, DOI: 10.7150/thno.17071.

126 A. E. Ivanova, Y. V. Burgart, V. I. Saloutin, Y. R. Orshanskaya and V. V. Zarubaev, *Mendeleev Commun.*, 2018, **28**, 52–54.

127 H. Ju, J. Zhang, Z. Sun, Z. Huang, W. Qi, B. Huang, P. Zhan and X. Liu, *Eur. J. Med. Chem.*, 2018, **146**, 220–231.

128 T. Guo, P. Datwyler, E. Demina, M. R. Richards, P. Ge, C. Zou, R. Zheng, A. Fougerat, A. V. Pshezhetsky, B. Ernst and C. W. Cairo, *J. Med. Chem.*, 2018, **61**, 1990–2008.

129 M. Obul, X. Wang, J. Zhao, G. Li, H. A. Aisa and G. Huang, *Mol. Diversity*, 2019, **23**, 1–9, DOI: 10.1007/s11030-018-9840-5.

130 W. Guan, J. Li, Q. Chen, Z. Jiang, R. Zhang, X. Wang, Z. Yang and X. Pan, *Molecules*, 2017, **22**, 1738, DOI: 10.3390/molecules22101738.

131 S. Quosdorf, A. Schuetz and H. Kolodziej, *Molecules*, 2017, **22**, 1989, DOI: 10.3390/molecules22111989.

132 H.-L. Li, R. Xu, X.-M. Li, S.-Q. Yang, L.-H. Meng and B.-G. Wang, *Org. Lett.*, 2018, **20**, 1465–1468.

133 A. S. Chintakrindi, D. J. Gohil, S. T. Kothari, A. S. Chowdhary and M. A. Kanyalkar, *Med. Chem. Res.*, 2018, **27**, 1013–1025, DOI: 10.1007/s00044-017-2124-2.

134 S.-Q. Yang, X.-M. Li, X. Li, H.-L. Li, L.-H. Meng and B.-G. Wang, *Phytochem. Lett.*, 2018, **25**, 191–195.

135 N. Singh, N. Anjum and R. Chandra, *Phytochem. Rev.*, 2019, **18**, 69–107, DOI: 10.1007/s11101-018-9581-1.

136 D. Furushima, K. Ide and H. Yamada, *Molecules*, 2018, **23**, 1795, DOI: 10.3390/molecules23071795.

137 M. Ullah, Z. Uddin, Y. H. Song, Z. P. Li, J. Y. Kim, Y. J. Ban and K. H. Park, *S. Afr. J. Bot.*, 2018, **120**, 326–330, DOI: 10.1016/j.sajb.2018.10.020.

138 J. Xu, S. Chen, J. Jin, L. Ma, M. Guo, C. Zhou and J. Dou, *Peptides*, 2019, **112**, 14–22, DOI: 10.1016/j.peptides.2018.10.004.

139 H.-J. Zhi, H.-Y. Zhu, Y.-Y. Zhang, Y. Lu, H. Li and D.-F. Chen, *Phytomedicine*, 2018, **57**, 105–116, DOI: 10.1016/j.phymed.2018.12.009.

140 D. H. Kim, G. S. Park, A. S. Nile, Y. D. Kwon, G. Enkhtaivan and S. H. Nile, *Food Chem. Toxicol.*, 2019, **125**, 313–321, DOI: 10.1016/j.fct.2019.01.013.

141 Z. Yunshi, Y. Jing, Q. Xian, L. Xing, L. Xieqin and F. Ganzhu, *Antiviral Ther.*, 2017, **22**, 599–611, DOI: 10.3851/IMP3152.

142 Y. Wang, J. Y. Kim, Y. H. Song, Z. P. Li, S. H. Yoon, Z. Uddin, Y. J. Ban, K. W. Lee and K. H. Park, *Int. J. Biol. Macromol.*, 2019, **128**, 149–157, DOI: 10.1016/j.ijbiomac.2019.01.105.

143 E. W. C. Chan, S. K. Wong, J. Tangah and H. T. Chan, *Syst. Rev. Pharm.*, 2018, **9**, 58–63.

144 A.-X. Hu, Q.-Y. Tao, Y.-K. Yuan and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 109305979 A 20190205, 2019.

145 A.-X. Hu, K.-Y. Yuan and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 109293644 A 20190201, 2019.

146 X.-M. Gao, H. Zhu, S.-S. Wang, Z. Zhang and D. Cui, Faming Zhuanli Shenqing, Design Patent, CN 109265503 A 20190125, 2019.

147 J.-M. Chen, X.-Y. An, G. Gao, J.-K. Zhang, X.-Y. Huang, J.-L. Ge, Y. Zhang and X.-H. Ning, Faming Zhuanli Shenqing, Design Patent, CN 109160937 A 20190108, 2019.

148 A.-X. Hu, L. Liu, J. Ye and Y. Zhu, Faming Zhuanli Shenqing, Design Patent, CN 109053607 A 20181221. 2018.

149 J. Ye, M. He, L. Liu, Y. Zhu and A.-X. Hu, Faming Zhuanli Shenqing, Design Patent, CN 109053606 A 20181221, 2018.

150 A.-X. Hu, Z.-Z. Yan, M. Zhang and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 108864073 A 20181123, 2018.

151 A.-X. Hu, Y.-J. Yi, M. Zhang, X.-Q. Wei and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 108863972 A 20181123, 2018.

152 Y.-L. Zhao, H.-T. Li, X.-L. Zhou, S.-C. Wei, J.-Y. Li and K. Li, Faming Zhuanli Shenqing, Design Patent, CN 108794441 A 20181113, 2018.

153 Y.-L. Zhao, H.-T. Li, X.-L. Zhou, S.-C. Wei, J.-Y. Li and K. Li, Faming Zhuanli Shenqing, Design Patent, CN 108794442 A 20181113, 2018.

154 A.-X. Hu, Y.-J. Yi, M. Zhang and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 108774193 A 20181109, 2018.

155 B.-G. Wang, X. Li, X.-D. Li and X.-M. Li, Faming Zhuanli Shenqing, Design Patent, CN 108774115 A 20181109, 2018.

156 A.-L. Liu, A.-X. Hu, W.-W. Lian, M.-W. Xiao and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 108689961 A 20181023, 2018.

157 L. Li, F.-G. Lu, H.-H. Huang, L.-J. Chen, K. Wei, Y. Ning, J. Hu and H. Ding, Faming Zhuanli Shenqing, Design Patent, CN 108578439 A 20180928, 2018.

158 Z.-Y. Li, S.-T. Yu, G.-H. Du, A.-L. Liu and X.-M. Qin, Faming Zhuanli Shenqing, Design Patent, CN 108578458 A 20180928, 2018.

159 A.-X. Hu, M. Zhang, X.-Q. Wei, J. Ye, A.-L. Liu and H. Jia, Faming Zhuanli Shenqing, Design Patent, CN 108546254 A 20180918, 2018.

160 Y.-W. Li, M. Zhang, A.-X. Hu and W.-S. Li, Faming Zhuanli Shenqing, Design Patent, CN 108530439 A 20180914, 2018.

161 A.-X. Hu, W. Chen, J.-X. Nie, M.-Y. Cui and J. Ye, Faming Zhuanli Shenqing, Design Patent, CN 108503604 A 20180907, 2018.

162 L.-P. Cheng, Z.-X. Zhao, W. Pang, R. Ye and Y.-P. Li, Faming Zhuanli Shenqing, Design Patent, CN 108383805 A 20180810, 2018.

163 W. Xiao, Z.-L. Li, Y.-C. Meng, Z.-Q. Meng, G. Ding, Z.-Z. Wang and W.-Z. Huang, Faming Zhuanli Shenqing, Design Patent, CN 108299246 A 20180720, 2018.

164 B.-G. Wang, L. Li and X.-M. Li, Faming Zhuanli Shenqing, Design Patent, CN 108299462 A 20180720, 2018.

165 L.-W. Qi, L. Zhang, X.-J. Yin, J. Li and G.-X. Ma, Faming Zhuanli Shenqing, Design Patent, CN 108125942 A 20180608, 2018.

166 Y.-S. Tian, K.-L. Wang, B.-Y. Wang, L.-J. Li, D.-Y. Liu and G.-G. Li, Faming Zhuanli Shenqing, Design Patent, CN 108101804 A 20180601, 2018.

167 H.-P. Li, Faming Zhuanli Shenqing, Design Patent, CN 108078980 A 20180529, 2018.

168 A.-X. Hu, J.-X. Nie, J. Ye, A.-L. Liu and H. Jia, Faming Zhuanli Shenqing, Design Patent, CN 108047160 A 20180518, 2018.

169 A.-X. Hu, W.-X. Xie, J.-X. Nie and X.-Q. Wei, Faming Zhuanli Shenqing, Design Patent, CN 107987033 A 20180504, 2018.

170 H.-Y. Tian, Z.-J. Jiang, S.-F. Luo, C.-F. Li and C.-M. Shen, Faming Zhuanli Shenqing, Design Patent, CN 111265539 A 20200612, 2020.

171 X.-B. Li, X. Lv, P.-F. Wang and J.-Z. Chen, Faming Zhuanli Shenqing, Design Patent, CN 111233962 A 20200605, 2020.

172 L.-P. Chen, M. Li, Z.-J. Zhong, W. Yu, L.-P. Shi, W. Pang and X.-Z. Xiao, Faming Zhuanli Shenqing, Design Patent, CN 111233790 A 20200605, 2020.

173 Z.-J. Ma, Y.-J. Jiang and M.-X. Liu, Faming Zhuanli Shenqing, Design Patent, CN 111205252 A 20200529, 2020.

174 Z.-L. Wu, M.-Y. Cui and A.-X. Hu, Faming Zhuanli Shenqing, Design Patent, CN 111153898 A 20200515, 2020.

175 P. Chen, M.-Y. Cui and A.-X. Hu, Faming Zhuanli Shenqing, Design Patent, CN 111138377 A 20200512, 2020.

176 Z.-L. Wu, A.-Y. Chen and A.-X. Hu, Faming Zhuanli Shenqing, Design Patent, CN 111100074 A 20200505, 2020.