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Quantitative proteomics approach to investigate the antibacterial response of *Helicobacter pylori* to daphnetin, a traditional Chinese medicine monomer†

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Helicobacter pylori is a Gram-negative bacterium related to the development of peptic ulcers and stomach cancer. An increasing number of infected individuals are found to harbor antibiotic-resistant *H. pylori*, which results in treatment failure. Daphnetin, a traditional Chinese medicine, has a broad spectrum of antibacterial activity without the development of bacterial resistance. However, the antibacterial mechanisms of daphnetin have not been elucidated entirely. To better understand the mechanisms of daphnetin's effect on *H. pylori*, a label-free quantitative proteomics approach based on an EASY-nLC 1200 system coupled with an Orbitrap Fusion Lumos mass spectrometer was established to investigate the key protein differences between daphnetin- and non-daphnetin-treated *H. pylori*. Using the criteria of greater than 1.5-fold changes and adjusted *p* value < 0.05, proteins related to metabolism, membrane structure, nucleic acid and protein synthesis, ion binding, *H. pylori* colonization and infection, stress reaction, flagellar assembly and so on were found to be changed under daphnetin pressure. And the changes of selected proteins in expression level were confirmed by targeted proteomics. These new data provide us a more comprehensive horizon of the proteome changes in *H. pylori* that occur in response to daphnetin.

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1. Introduction

Helicobacter pylori (*H. pylori*) is a microaerophilic Gram-negative bacterium with extremely demanding growth conditions which is linked to the development of duodenal ulcers and stomach cancer.^{1,2} Increasing evidence suggests that *H. pylori* may be associated with multiple chronic disorders, such as neurological disorders, cerebrovascular and cardiovascular disorders, etc.^{3,4} In 2017, the WHO published its first ever list of antibiotic-resistant “priority pathogens”, and *H. pylori* was regarded as one of the 12 high priorities.⁵ Standard triple therapy has been the recommended treatment for *H. pylori* eradication for the past two decades,⁶ leading to the rising of antibiotic-resistant rate. Due to the decreased efficacy of treatment and increased antibiotic resistance, a large amount of research has focused on the discovery of new antimicrobials.

Daphnetin (7,8-dihydroxycoumarin) is a natural coumarin derivative with anti-inflammation, anti-microorganism and antioxidative properties.^{7,8} To date, neither toxic effects nor genetic toxicity have been observed in the treatment of

daphnetin.^{9,10} Therefore, inclusive research interests were attached to study the activity and mechanism of daphnetin against multidrug resistant (MDR) *H. pylori*. In our previous research, daphnetin showed good activities against MDR *H. pylori* ATCC 43504, with minimal inhibit concentration (MIC) values as 25 µg mL⁻¹. And preliminary exploration of the antibacterial mechanisms of daphnetin is presumed to be the increased translocation of phosphatidylserine, nucleic acid damage, up-regulated RecA expression, and decreased *H. pylori* adhesion,¹¹ but a lot of work need to do to get a more comprehensive explanation of the mechanism.

Quantitative proteomics based on mass spectrometry (MS) has emerged as a preferred strategy to discover condition-related proteins, and few comparative analysis studies of *H. pylori* by 2-DE/MS or immunoproteomics have been performed to look for differences in protein expression.^{12,13} Moreover, quantitative proteomics has been employed to study antibiotic reactions in various bacteria,¹⁴ which helped us explain the mechanisms of antibacterial drug actions by differences in protein expression. As the data-dependent acquisition (DDA)-based label-free method has the properties of simplified sample preparation and low costs due to no isotope labeling. And parallel reaction monitoring (PRM) is widely used in targeted proteomics, with a tolerance of approximately 10–20 ppm.¹⁵ In this study, label-free quantitative proteomics and

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PRM methods were used to discover the differentially expressed proteins under daphnetin pressure and dig the mechanisms of daphnetin's action on *H. pylori*. Based on previous research methods,^{14,16} we applied subinhibitory doses of daphnetin (1/2 MIC) to give the bacteria an opportunity to respond to daphnetin and to continue to grow. The plate culture method was chosen to culture the bacteria, as culturing with *H. pylori* is very demanding.

2. Materials and methods

2.1 Cell culture and antibiotic treatment

Frozen *H. pylori* ATCC 43504 was thawed and recovered on Columbia blood agar plates. *H. pylori* cells for proteomics analysis were incubated with or without 12.5 $\mu\text{g mL}^{-1}$ daphnetin (Meilun Biotech Co., Ltd., Dalian, China) for 3 days at 37 °C under microaerobic conditions as previously described,¹⁷ then collected, adjusted to the same amount and washed in PBS twice by centrifugation, stored at –80 °C.

2.2 Sample and peptide preparation

H. pylori cells were resuspended in the lysis buffer (50 mM NH_4HCO_3 , 150 mM NaCl, 2% sodium deoxycholate, pH 8.5) and sonicated at 4 °C to increase the extraction efficiency.¹⁸ For label-free proteomics, proteins were reduced with 10 mM dithiothreitol (DTT) at 95 °C for 5 min. Protein samples were digested by trypsin (1 : 50 w/w, Promega, Madison, WI) overnight at 37 °C. Then, formic acid was added to a final concentration of 1% to remove sodium deoxycholate. Last, the sample was desalted by C18 reverse-phase tips (Reprosil-Pur Basic C18, 5 μm , Dr Maisch GmbH). All samples were dried with a CHRIST Freeze Dryer ALPHA 2-4D (CHRIST, Vaihingen, Germany) and stored at –80 °C before use.

2.3 Label-free quantification analysis

Standard bottom-up proteomics technique was used to analyze the samples on a Thermo Scientific Orbitrap Fusion Lumos platform coupled with an EASY-nLC 1200.¹⁹ Peptides were enriched on a trap column (Reprosil-Pur Basic C18 (3 μm , Dr Maisch GmbH, Germany), 20 \times 0.15 mm) and separated on a C18 column (Reprosil-Pur Basic C18 (1.9 μm , Dr Maisch GmbH, Germany) 150 \times 0.15 mm) at a flow rate of 600 $\mu\text{L min}^{-1}$. The solvent buffer A consists of water with 0.1% formic acid, and that of solvent B was 80% acetonitrile with 0.1% formic acid. After sample loading, the gradient started from 9% of buffer B and then from 9 to 12% of buffer B for 3 min. Then, the gradient increased from 12 to 28% of buffer B for 39 min and from 28 to 36% for 11 min. Finally, the gradient ascended to 95% of buffer B and was maintained for 7 min. Three biological replicates were analyzed and the scan sequence consist of an MS1 spectrum (Orbitrap analysis; resolution, 120 000; mass range, 350–1550 m/z ; RF lens, 50%; AGC target, 4×10^5 ; maximum injection time, 50 ms) and an MS2 analysis with HCD accompanied by the following conditions: collision energy, 32%; AGC, 5×10^4 ; maximum injection time, 22 ms; Orbitrap solution, 15 000; and isolation window, 1.6 Da. For

database analyses, these initial data were searched against *H. pylori* ATCC 43504 and ATCC 26695 combined databases from UniProt by the Thermo Scientific™ Proteome Discoverer™ version 2.2 (PD2.2) software. Trypsin was specified for protein digestion with two missed cleavages allowed for each peptide. The search parameters were set as previously described.²⁰

2.4 Bioinformatics analysis

The results were exported from PD 2.2 and analyzed by “Wu Kong” platform powered by R language (<https://www.omicsolution.org/wkomics/main/>). Both experiments were repeated in triplicate to generate three biological replicates and three technical duplications. Groups with values greater than or equal to 2/3 were included. Missing values left were imputed by the KNN method. For relative quantification, the label-free quantification intensities were transformed with \log_2 function. Proteins with a CV (coefficient of variance) value below 20% in three biological replicates were subjected to quantitative analysis and one-way ANOVA, and p value was adjusted by q value method. To identify significant differences, proteins with adjusted $p < 0.05$ and fold change > 1.5 was considered to be up- or down-regulated. Functional annotation and clustering enrichment of differentially expressed proteins were conducted by using the Gene Ontology (GO) enrichment tool. Due to the lack of *H. pylori* ATCC 43504-related database in the KEGG pathway database, we replaced the gene name with the corresponding gene name of *H. pylori* ATCC 26695 by the Blast method to perform KEGG pathway analysis. For the analysis of interactions between proteins, differentially expressed proteins were searched against the STRING database (<http://string-db.org>) by the same ID-changed method used in the KEGG pathway analysis.

2.5 Mass spectrometry for label-free PRM analysis

PRM was used to confirm the differentially expressed proteins. The sample processing method was the same as described above. Both experiments were repeated in triplicate. The precursor m/z , charge state and retention time of peptides were referred to in the prior label-free experiments. The same Orbitrap Fusion Lumos platform and nanoLC parameters were applied for the PRM study. The targeted MS1 parameters were as follows: resolution, 60 000; AGC target, 2.0×10^5 ; and maximum injection time, 50 ms. PRM scanning was performed at 30 000 resolution, 1×10^5 AGC target, 54 ms maximum injection time and 0.7 m/z isolation window with peptides in ± 5 min retention time windows. Fragmentation was performed with a normalized collision energy of 32.²¹

2.6 Target peptides quantification

Data from PRM analysis were analyzed by Skyline,²² with spectral libraries built by DDA results. Briefly, the FASTA sequences were downloaded from the UniProtKB database, and imported into Skyline while fragmentation and selection are performed if they can match peptides in the spectral libraries. 2 or 3 peptides per protein were selected for quantification. Experiments were repeated in triplicate to generate three biological replicates. The data were normalized by total ionic abundance and processed



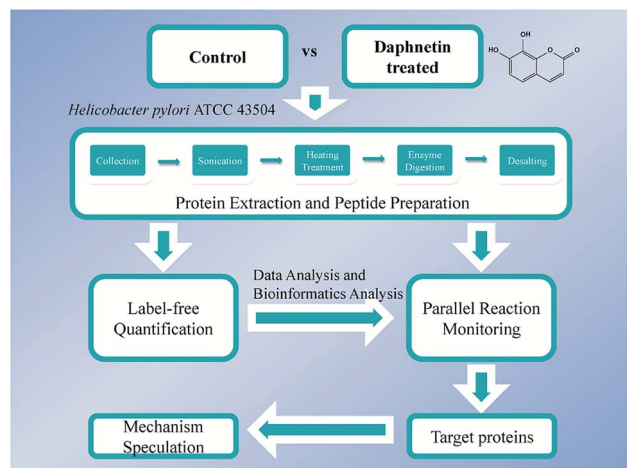


Fig. 1 Quantitative proteomics workflow applied to daphnetin-treated and untreated *H. pylori* ATCC 43504.

using a \log_2 scale. Peptides were selected by one-way ANOVA with p -value < 0.05 and fold change > 1.5 .

3. Results

3.1 The overall proteomic response of *H. pylori*

Our previous study found that daphnetin inhibited *H. pylori* ATCC 43504 with MIC of $25 \mu\text{g mL}^{-1}$.¹¹ Because full growth inhibition would be impractical for proteomic research, we applied a sublethal concentration of daphnetin and monitored

changes. In order to avoid the gap in the amount of bacteria introducing by daphnetin, before the pretreatment, we adjusted the amount of bacteria in different groups to be consistent. As shown in Fig. 1, label-free and PRM quantitative proteomics were applied to analyze the protein expression level of daphnetin-treated *H. pylori* ATCC 43504. In total, we identified 1282 bacterial proteins in three biological replicates, covering approximately 78% of the *H. pylori* proteome. Proteins with a large proportion of missing values and a CV value greater than 20% were excluded, 1207 proteins were used to perform correlation analysis. Replicates in the same group presented a linear correlation with $R^2 > 0.9$ (see Fig. S1 in the ESI[†]), indicating good repeatability. Based on adjusted p value ($p < 0.05$) and max fold change > 1.5 ,¹⁴ the expression of 152 proteins were finally identified to be significantly changed by treatment with daphnetin (Fig. 2A). Among which, there were 55 proteins exhibited decreased accumulation and 97 proteins exhibited increased accumulation (Table S1 in the ESI[†]). The hierarchical clustering of values is shown in Fig. 2B and demonstrates that the response to daphnetin was reproducible.

3.2 Differentially expressed proteins in the daphnetin-treated *H. pylori* compared to control groups

By using GO annotation enrichment analysis, KEGG pathway analysis and STRING analysis as mentioned above, we further divided these differentially expressed proteins into different functional categories. As shown in Fig. S2 in the ESI[†], the most enriched proteins in the GO annotation enrichment analysis

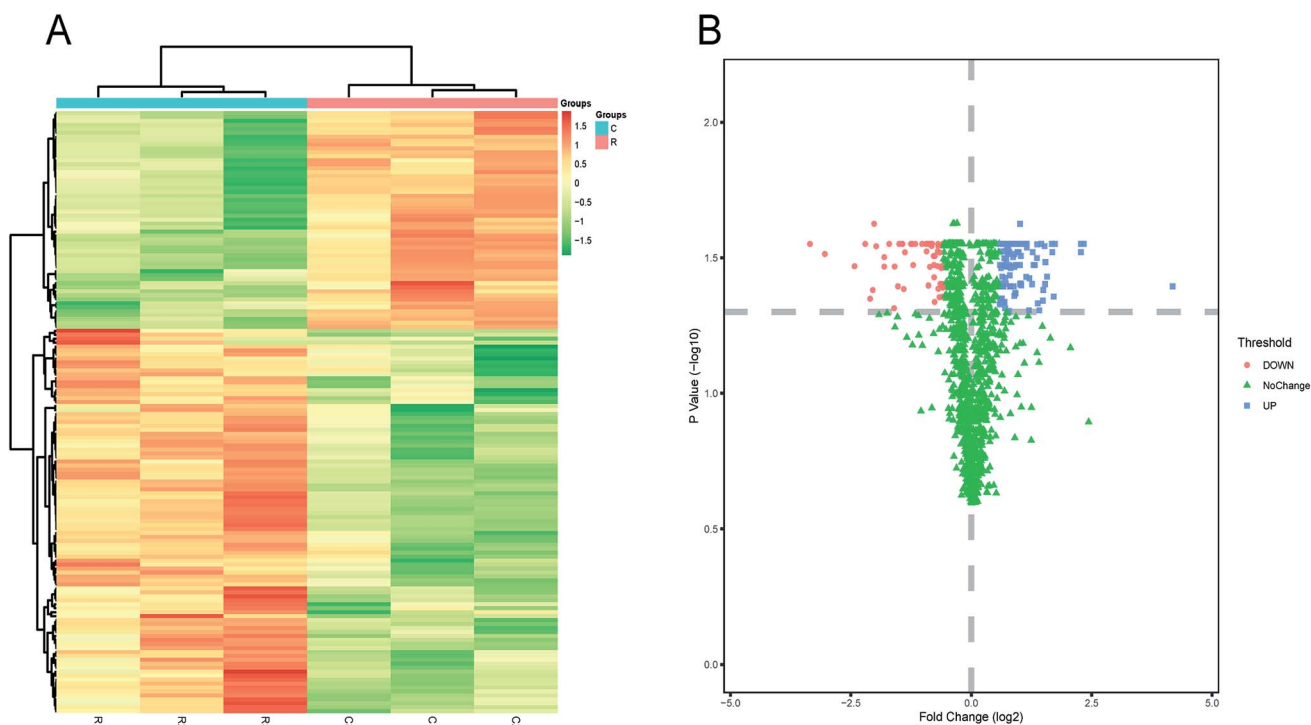


Fig. 2 Protein profiles of *H. pylori* without and with daphnetin treatment. (A) Hierarchical clustering of the proteins' test after hypothetical test from three biological replicates. Red represents the highest relative expression, and green represents the lowest. C1, C2, and C3 (*H. pylori* controls); R1, R2, and R3 (daphnetin treated *H. pylori*). (B) Volcano distribution of \log_2 average normalized and scaled values for proteins from *H. pylori* with daphnetin treatment divided by $p < 0.05$ and fold change > 1.5 , red represents down-regulated and blue represents up-regulated.

results of STRING analysis (Fig. 3) showed that, there were clusters of genes which have strong interactions with each other, and genes such as *tuf*, *tsf*, *rpsR*, *pheT*, *etc.* constituted the hub of the interaction network, indicating the vital roles played by these genes. Based on these analysis results, proteins were chosen to perform targeted proteomics.

In order to ensure reliable and repeatable measurements of protein expression in all the samples and to improve the sensitivity of the measurements, we applied a targeted MS-based method for defined protein quantification based on a PRM approach to validate protein expression differences. Peptides preparation process was the same as mentioned above. Three biological replicates were performed in all experiments. Protein candidates were selected from our label-free

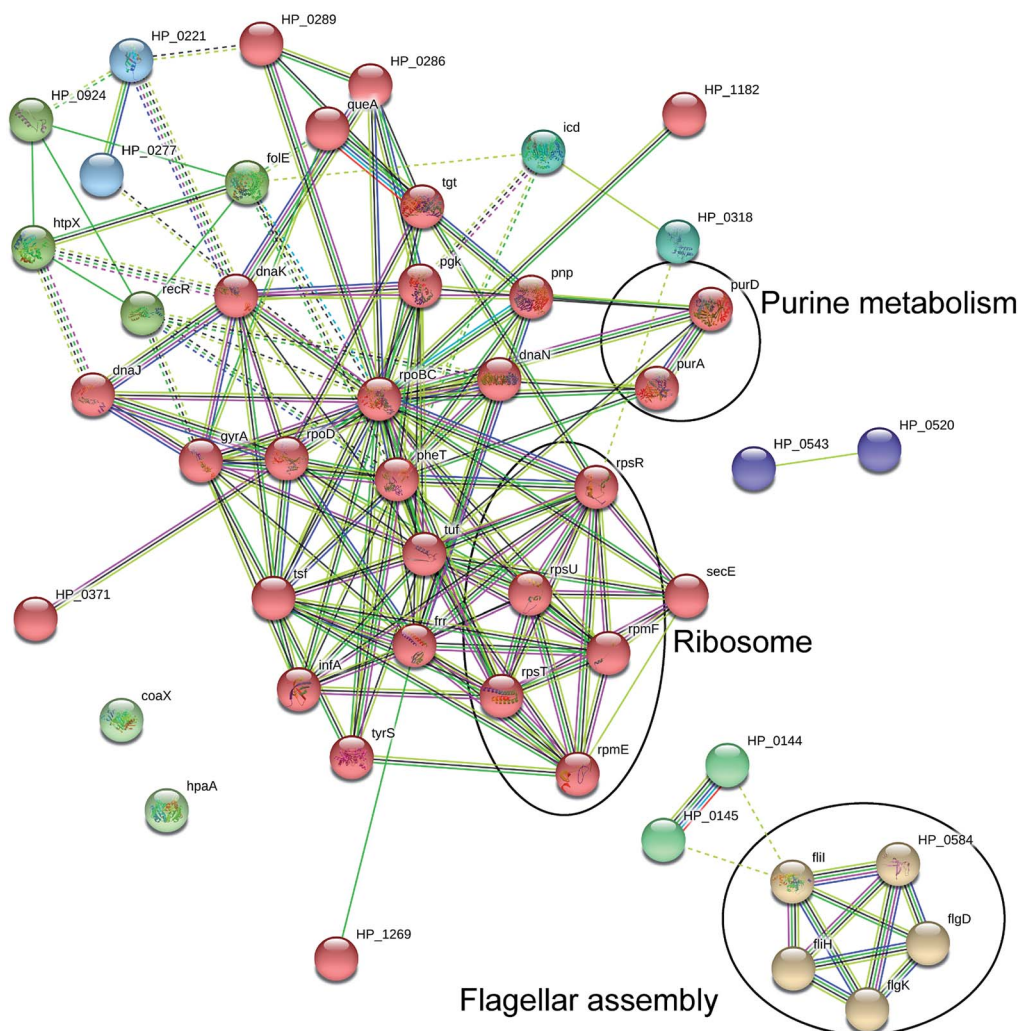


Fig. 3 Protein–protein interaction networks of partial differentially expressed proteins under daphnetin stress. Interacting proteins were identified by using STRING with a high confidence score and clustered by MCL clustering, and homologous gene names in *H. pylori* ATCC 26695 were used due to the lack of an *H. pylori* ATCC 43504 database. Nodes represent proteins, while edges represent protein–protein interactions. Colored nodes represent clusters. Colored edges: known interactions from: teal, curated databases; magenta, experimentally determined; predicted interactions: green, gene neighborhood; red, gene fusions; blue, gene co-occurrence; other: yellow, text mining; black, co-expression; purple, protein homology. Partial KEGG pathways were marked by circle.

quantification measurements, and 2 or 3 unique peptides were chosen to quantify one protein. All raw data were analyzed and processed with Skyline software with spectral library built by DDA results. Peptides containing amino acids that are easily modified and missed cleavage were excluded from the analysis. A minimum dotp value of 0.7 was used to filter precursors to remove the noisy peaks and interfering fragment ions. The peptide peak areas were exported into Excel for data normalization by total ionic intensity and statistical analysis. In total, we reliably quantified 11 proteins with 25 peptides (see Table S2 in the ESI†) as significantly impacted proteins (adjusted p value <0.05 and fold change >1.5).

4. Discussion

In light to the latest severe drug resistance situation of *H. pylori*, discovering antibacterial drugs with new structures or new mechanisms is extremely imperative. Our previous studies have demonstrated that daphnetin has anti-*H. pylori* activity, regardless of the resistance patterns to other antibiotics. And the effect of daphnetin on *H. pylori* ATCC 43504 was initially explored. However, the antibacterial mechanisms of daphnetin have not been elucidated entirely. With the development of protein identification techniques, quantification proteomics by LC-MS has been gradually used in the discovery of drug-target and mechanism.^{14,23,24} In this study, the application of high-resolution mass spectrometry improved the accuracy and sensitivity of this approach. Three biological replicates are necessary because of the poor homogeneity of the microbial population. Normalization by total iron intensity is necessary in bacteria proteome analysis as the amount of bacteria is difficult to control accurately. PRM can solve problems when no antibody is available to help verify or when many protein needs to be quantified. By means of quantification proteomics, we found the mechanism of action of daphnetin was extensive and complex.

4.1 Extensive impacts of daphnetin on *H. pylori* ATCC 43504

As shown in Fig. 4, interestingly, among the differentially expressed proteins, membrane structure-related proteins

accounted for the largest proportion, and approximately 1/3 of these proteins were down-regulated, while approximately 2/3 of those were up-regulated, which further verified the membrane structure changes induced by daphnetin found in the previous study.¹¹ Apart from membrane structure-related proteins, the expression of proteins related to acid metabolic and nucleotide metabolic processes were stimulated, while many biosynthetic processes were suppressed by daphnetin, leading to the imbalance between synthesis and metabolism. What's more, up-regulated expression of ion binding related proteins and down-regulated expression of RNA binding, structural constituent of ribosome and structural molecule activity related proteins were also observed in daphnetin-treated groups, indicating the declining of the cell activities.

Pathway enrichment analysis results showed that the expression levels of most proteins in the metabolic pathways, biosynthesis of secondary metabolites, microbial metabolism in diverse environments and carbon metabolism were up-regulated. And all the expression levels of the five proteins related to flagellar assembly pathway were up-regulated, while all the expression levels of proteins related to epithelial cell signaling in *H. pylori* infection pathway were down-regulated. Expression levels of four proteins related to ribosome were down-regulated, while those of four proteins related to purine metabolism pathway were up-regulated. Interestingly, five genes related to flagellar assembly pathway were proved to have strong interaction with each other by STRING analysis (Fig. 3), which was also reported in previous study.²⁵ The expression levels of *HP_0144* and *HP_0145*, genes related to cytochrome c oxidase, were down-regulated and have strong interaction with each other. As observed from Fig. 3 genes with important roles in the basic physical activities occupied the hub of interaction net constructed by String analysis. Among the hub genes, the expression levels of *tgt*, *rpoBC*, *frr*, *rpsR*, *rpsT*, *rpmF*, *pheT*, *gyrA*, *queA*, *pnp*, *tuf* and *rpsU*, related to gene function or protein expression, and *purA*, *HP_0221*, *HP_1182*, *HP_1269*, *icd* and *pgk* related to metabolism were all down-regulated. What's more, the expression levels of *dnaJ* and *dnaK* (heat shock proteins), combined with *recR* (recombination protein), and *hugZ* (heme

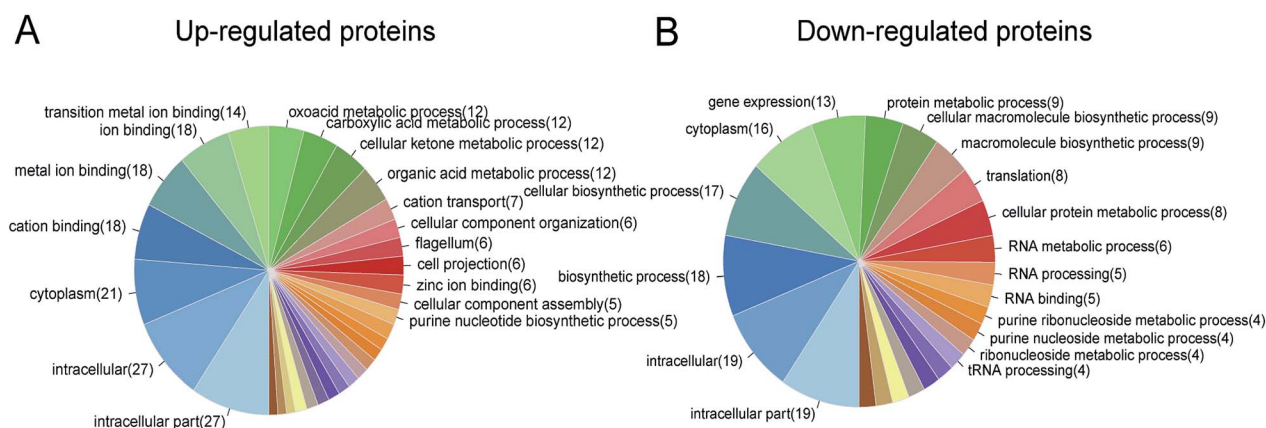


Fig. 4 Pie charts of GO enrichment classification of significantly changed proteins (counts) under daphnetin stress. (A) GO enrichment classification of up-regulated proteins under daphnetin stress. (B) GO enrichment classification of down-regulated proteins under daphnetin stress.



oxygenase) were up-regulated under daphnetin pressure. As members of epithelial cell signaling in *H. pylori* infection pathway, interactions between *HP_0520* (Cag1) and *HP_0543* (Cag22) or *hpaA* were also observed in the interaction network as previously reported.^{26,27}

4.2 Effects on epithelial cell signaling pathway during *H. pylori* infection

H. pylori secretes a number of proteinaceous factors which are essential for initial colonization and subsequent persistence in the host's stomach. I0ZHH0, a putative neuraminylactose-binding hemagglutinin-like protein HpaA, can bind to the surface receptors of gastric epithelial cells and further damage the gastric mucosa.²⁸ Vacuolating cytotoxin A (VacA), one of the major protein toxins, can bind to host cells and causes severe "vacuolation".^{29,30} Therefore, the decreased putative vacuolating cytotoxin (VacA)-like protein (I0ZBK7) definitely affects the ability of the bacteria to colonize the stomach and the development of gastric disease. Furthermore, many Gram-negative pathogens have the type IV secretion systems (T4SS) which can translocate bacterial virulence factors into host cells

to hijack cellular processes while delivering the CagA oncoprotein into gastric epithelial cells.³¹ As a member of Cag family, studies have found that CagF (I0ZJ86) might be a protein which is involved in the early steps of CagA recognition and delivery into the T4SS channel.³² In our study, the down-regulation of CagF and Cag1 (I0ZH94) may influence the translocation of virulence factor CagA into host cells. According to the KEGG pathway and string results, it is worth noting that HpaA, type-IV secretion system (T4SS), CagA, VacA together with BabA and UreI found in our previous study¹¹ have vital roles in the epithelial cell signaling pathway during *H. pylori* infection (Fig. 5). Apart from the vacuolation effect mentioned above, it is reported that VacA can indirectly induce ulcerogenesis in mice. And CagA has been shown to interact with more and more host signaling molecules in phosphorylation-dependent and-independent ways such as MAPK signaling pathway. Our results proved that daphnetin reduced the expression levels of several important proteins in this pathway, suggesting the inhibition of *H. pylori* colonization and infection, which further explained our previous results that daphnetin exposure remarkably decreased the

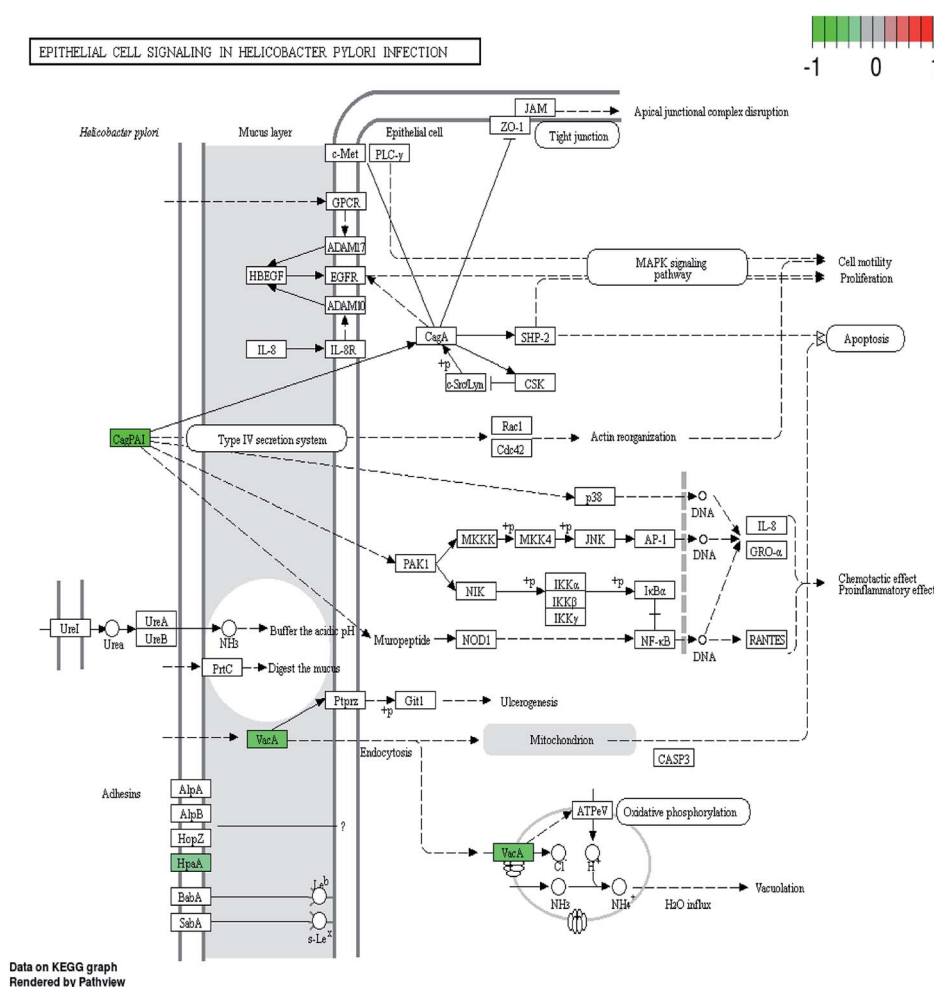


Fig. 5 The epithelial cell signaling pathways in *H. pylori* infection. Proteins with decreased accumulation under daphnetin pressure are shown in green.

adherence of *H. pylori* to GES-1 cells compared with the control condition.¹¹

4.3 Effects on membrane structure, ion binding, nucleic acid synthesis and metabolism

Among the proteins with significantly different expression levels induced by daphnetin, the large amount of proteins related to membrane represents huge changes in the structure of the cell membrane. There are many proteins in membrane proteins that play important roles for the infection and colonization ability of *H. pylori*. The outer membrane protein OipA (I0ZBG0) is involved in the induction of IL-8 secretion and is related to the development of peptic ulcers and gastric cancer.³³ Reduced OipA expression levels represent changes in the membrane structure and decreased infection ability. Studies have demonstrated a high popularity of strains with oipA, and significant associations have been found between functional oipA and cagA or vacA genotypes.³⁴ The effects of daphnetin on these proteins reveal its potential capability to disrupt the *H. pylori* infection process. What's more, the expression of a considerable amount of proteins related to ion binding changed under daphnetin pressure. In *H. pylori*, the ability to acquire ion such as iron is essential for the successful colonization on the gastric niche and the subsequent infection in a mouse model.³⁵ Thus the daphnetin-induced down-regulation of the iron(III) dicitrate transport protein FecA (I0ZG47) makes the colonization and infection process difficult.

Daphnetin influenced the expression level of a large amount of enzymes related to gene duplication, transcription and

translation, fundamentally leading to internal synthesis and metabolism disorders. Apart from the down-regulated proteins related to ribosome, studies have demonstrated that the unique fused beta- and beta'-like subunits of DNA-dependent RNA polymerase (I0ZE29, ropBC) in *H. pylori* conduces to its capability to colonize the human gastric mucosa.³⁶ Therefore, the suppression of ropBC by daphnetin will affect the colonization of *H. pylori*. In addition, the decreased expression of DNA gyrase subunit A, gyrA (I0ZIK8), represents a decline in DNA synthesis ability or DNA damage. Translation elongation factor Tu (I0ZE21, EF-Tu) is an important component of the protein translation system.³⁷ Recently, EF-Tu was reported to be a new secretory adherence factor in *H. pylori* and is related to *H. pylori* adaptation for host-pathogen interaction during pathogenesis.³⁸ The down-regulated expression of EF-Tu not only affects protein translation but also the interaction with host cells.

The maximum amount of proteins, which changed significantly under daphnetin's pressure, belongs to the metabolism group, covering various metabolic and synthetic pathways. Among them, adenylosuccinate synthetase, AdSS (I0ZBQ4), is an enzyme that regulates purine metabolism of *H. pylori*. AdSS is a promising drug target in pathogenic organisms that utilize only the purine salvage pathway, the decreased expression level (shown in Fig. 6) under daphnetin pressure will influence the salvage of purine.³⁹

4.4 Upregulated proteins

The ubiquitous molecular chaperones can control the dynamic balance of cell proteins, such as stressful growth conditions. In

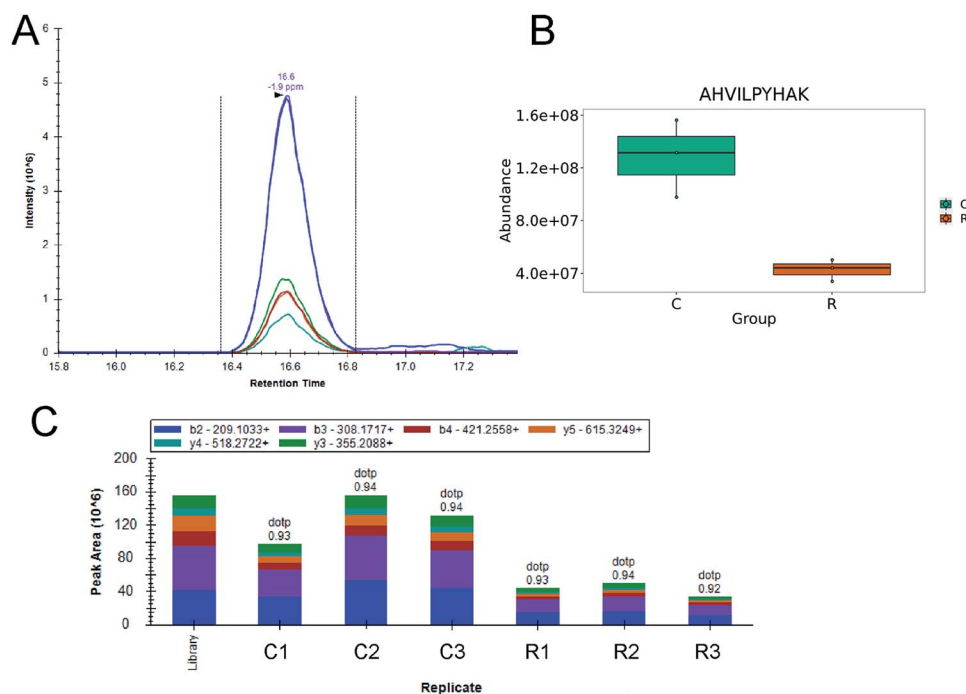


Fig. 6 AHVILPYHAK peptide (I0ZBQ4) quantification results by PRM. One way ANOVA statistical test and *q* value method was used, and adjusted *p* < 0.05. (A) Extracted ion chromatogram of PRM fragment ions for AHVILPYHAK; (B) plot map of ion abundance comparison, C, control groups (marked green); R, daphnetin treated groups (marked orange); (C) the corresponding peak area replicates of daphnetin treated and untreated groups. Control groups: C1, C2, C3; daphnetin treated groups: R1, R2, R3.

our study, the expression levels of highly conserved chaperone proteins DnaK (HSP70) and DnaJ (HSP40) were up-regulated under daphnetin pressure. DnaK is supposed to play an important role in promoting proliferation and antiapoptosis. Recent studies have shown that the expression level of HSP70 will affect the proliferation of gastric epithelial cells to a certain extent.⁴⁰ Faced with the pressure of daphnetin, *H. pylori* increased expression levels of heat shock proteins to respond to stressors and slowed the proliferation of gastric epithelial cells. Apart from heat shock proteins, the expression levels of recombinational repair protein RecR were up-regulated for mediating recombinational repair of DNA damage caused by daphnetin.⁴¹ In addition to stress-responsive proteins, it is surprising that the expression of five proteins related to the flagellar assembly pathway were up-regulated. The phenomenon was also observed in acid treated *H. pylori*, as the flagellar protein and movement were activated.⁴²

4.5 Speculation on the antibacterial mechanisms of daphnetin against *H. pylori*

As previous studies have elaborated, daphnetin possesses notable antioxidative, anti-inflammatory and anticoagulant activities in diabetic nephropathy, neurodegenerative disorders, and non-alcoholic fatty liver disease treatment through suppressing the LPS-induced activation of JAK/STAT cascades and ROS production, regulating p38 MAPK and JNK signaling pathway, enhancing the expression of HSP70 via ERK signaling pathway, etc.^{43–45} But little is known about the mechanisms of daphnetin's action on bacteria. In this study, quantitative proteomics was used to explore the possible mechanism of daphnetin on *H. pylori* ATCC 43504. And proteins related to metabolism, membrane structure, nucleic acid and protein synthesis, ion binding, *H. pylori* colonization and infection, stress reaction, flagellar assembly and so on were considered responsible for the drug action, indicating that daphnetin exerted its efficacy through multiple targets. Furthermore, among these proteins, VacA toxin, and AdSS were chosen as promising targets for inhibitor screening,^{39,46,47} and HpaA, VacA, Cag, gyrA and FecA were administered as vaccine antigens in animal models^{48–50} in other studies. All the results confirm that daphnetin is a promising antibacterial with specific and extensive antibacterial mechanisms. Although the *in vitro* antibacterial activity is not outstanding, we are full of hope for its *in vivo* efficacy, especially its inhibition of host-pathogen colonization and the infection process. With increasing multi-drug resistance rate of *H. pylori*, drugs with multiple targets can effectively slow the development of drug resistance. Additionally, to effectively inhibit the emergence of multidrug resistance, traditional Chinese medicine extracts with multiple targets, such as daphnetin, are worthy of further research and discovery.

5. Conclusions

The application of label-free quantitative proteomics and PRM provides an alternative method for seeking targets and

mechanisms in antibiotic-treated bacteria. From the perspective of proteomics, the protein variation in bacteria becomes more intuitive and abundant. In general, the antibacterial mechanism of daphnetin to *H. pylori* are related to metabolism, membrane structure, nucleic acid and protein synthesis, ion binding, *H. pylori* colonization and infection, stress reaction, flagellar assembly and so on. These data suggest that daphnetin has great potential to be an effective anti-*H. pylori* clinical agent with potential *in vivo* activity and provide possible targets for anti-*H. pylori* compounds screening.

Conflicts of interest

The authors declare that they have no competing interests.

Acknowledgements

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References

- 1 J. R. Warren and B. Marshall, *Lancet*, 1983, **321**, 1273–1275.
- 2 B. E. Dunn, H. Cohen and M. J. Blaser, *Clin. Microbiol. Rev.*, 1997, **10**, 720–741.
- 3 J. D. de Korwin, G. Ianaro, G. Gibiino and A. Gasbarrini, *Helicobacter*, 2017, **22**, e12411.
- 4 G. J. Finn, B. S. Creaven and D. A. Egan, *Biochem. Pharmacol.*, 2004, **67**, 1779–1788.
- 5 WHO Publishes List of Bacteria for Which New antibiotics Are Urgently Needed, 2017, Available online: <http://www.who.int/mediacentre/news/releases/2017/bacteria-antibiotics-needed/en/>.
- 6 V. Papastergiou, S. D. Georgopoulos and S. Karatapanis, *World J. Gastroenterol.*, 2014, **20**, 9898.
- 7 H. M. Chawla, K. Chakrabarty, S. S. Chibber, A. N. Kalia and N. C. Chaudhury, *Indian J. Pharm. Sci.*, 1980, **42**, 138–139.
- 8 H. Fukuda, S. Nakamura, Y. Chisaki, T. Takada, Y. Toda, H. Murata, K. Itoh, Y. Yano, K. Takata and E. Ashihara, *Biochem. Biophys. Res. Commun.*, 2016, **471**, 63–67.
- 9 Q. S. Wenwei Zou, Y. Fu and Z. Xia, *Chin. Tradit. Pat. Med.*, 2013, **35**, 39–45.
- 10 G. J. Finn, E. Kenealy, B. S. Creaven and D. A. Egan, *Cancer Lett.*, 2002, **183**, 61–68.
- 11 G. Wang, J. Pang, X. Hu, T. Nie, X. Lu, X. Li, X. Wang, Y. Lu, X. Yang, J. Jiang, C. Li, Y. Q. Xiong and X. You, *Int. J. Mol. Sci.*, 2019, **20**, 850.



- 12 P. R. Jungblut, F. Schiele, U. Zimny-Arndt, R. Ackermann, M. Schmid, S. Lange, R. Stein and K. P. Pleissner, *Proteomics*, 2010, **10**, 182–193.
- 13 S. C. Baik, K. M. Kim, S. M. Song, D. S. Kim, J. S. Jun, S. G. Lee, J. Y. Song, J. U. Park, H. L. Kang and W. K. Lee, *J. Bacteriol.*, 2004, **186**, 949–955.
- 14 L. Xiaofen, H. Yingwei, P. Pei-Jing, C. Daijie and L. Henry, *J. Proteome Res.*, 2013, **13**, 1223–1233.
- 15 J. Woo, D. Han, J. I. Wang, J. Park, H. Kim and Y. Kim, *J. Proteome Res.*, 2017, **16**, 3419–3432.
- 16 Y. Sung-Ho, C. Chi-Won, K. Sang-Oh, P. Gun Wook, C. Kun, K. Kyung-Hoon, K. J. Young, Y. Jong Shin, L. Je Chul and C. Jong-Soon, *J. Proteome Res.*, 2011, **10**, 459–469.
- 17 J. M. Whitmire and D. S. Merrell, *Methods Mol. Biol.*, 2012, **921**, 17–27.
- 18 B. Leng, J. Zeng, Z. Xiong, W. Lv and Y. Wan, *J. Proteome Res.*, 2006, **5**, 2547–2553.
- 19 R. Karlsson, K. Thorell, S. Hosseini, D. Kenny, C. Sihlbom, Å. Sjöling, A. Karlsson and I. Nookaew, *Front. Microbiol.*, 2016, **7**, 1757.
- 20 G. Espadas, E. Borràs, C. Chiva and E. Sabidó, *Proteomics*, 2017, **17**, 201600416.
- 21 A. C. Peterson, J. D. Russell, D. J. Bailey, M. S. Westphall and J. J. Coon, *Mol. Cell. Proteomics*, 2012, **11**, 1475.
- 22 B. MacLean, D. M. Tomazela, N. Shulman, M. Chambers, G. L. Finney, B. Frewen, R. Kern, D. L. Tabb, D. C. Liebler and M. J. MacCoss, *Bioinformatics*, 2010, **26**, 966–968.
- 23 I. Vranakis, I. Goniotakis, A. Psaroulaki, V. Sandalakis, Y. Tselentis, K. Gevaert and G. Tsiotis, *J. Proteomics*, 2014, **97**, 88–99.
- 24 B. Cooper, N. Islam, Y. Xu, H. S. Beard, W. M. Garrett, G. Gu and X. Nou, *Proteomics*, 2018, **18**, e1700461.
- 25 M. C. Lane, P. W. O'Toole and S. A. Moore, *J. Biol. Chem.*, 2006, **281**, 508–517.
- 26 D. M. Pinto-Santini and N. R. Salama, *J. Bacteriol.*, 2009, **191**, 7343–7352.
- 27 R. Hauser, A. Ceol, S. V. Rajagopala, R. Mosca, G. Sizler, N. Wermke, P. Sikorski, F. Schwarz, M. Schick, S. Wuchty, P. Aloy and P. Uetz, *Mol. Cell. Proteomics*, 2014, **13**, 1318–1329.
- 28 C. Elisabet, N. M. Johanna, B. L. Ingrid, C. L. Nilsson and S. Ann-Mari, *Infect. Immun.*, 2006, **74**, 920–926.
- 29 S. L. Palframan, T. Kwok and K. Gabriel, *Front. Cell. Infect. Microbiol.*, 2012, **2**, 92.
- 30 A. Link, C. Langner, W. Schirrmeister, W. Habendorf and P. Malfertheiner, *World J. Gastroenterol.*, 2017, **23**, 4712.
- 31 S. Backert, N. Tegtmeyer and W. Fischer, *Future Microbiol.*, 2015, **10**, 955–965.
- 32 C. Marc Roger, T. Elizabetta, M. Cesare and S. Markus, *Infect. Immun.*, 2006, **74**, 273.
- 33 Y. Yamaoka, D. H. Kwon and D. Y. Graham, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 7533–7538.
- 34 T. Ando, R. M. Peek, D. Pride, S. M. Levine, T. Takata, Y. C. Lee, K. Kusugami, A. van der Ende, E. J. Kuipers, J. G. Kusters and M. J. Blaser, *J. Clin. Microbiol.*, 2002, **40**, 239–246.
- 35 A. Danielli, S. Romagnoli, D. Roncarati, L. Costantino, I. Delany and V. Scarlato, *J. Bacteriol.*, 2009, **191**, 3717–3725.
- 36 N. Zakharova, P. S. Hoffman, D. E. Berg and K. Severinov, *J. Biol. Chem.*, 1998, **273**, 19371–19374.
- 37 D. Wang, B. Luo, W. Shan, M. Hao, X. Sun and R. Ge, *Metallomics*, 2013, **5**, 888–895.
- 38 K. H. Chiu, L.-H. Wang, T.-T. Tsai, H.-Y. Lei and P.-C. Liao, *J. Proteome Res.*, 2016, **16**(1), 264–273.
- 39 A. Bubic, N. Mrnjavac, I. Stuparevic, M. Lyczek, B. Wielgus-Kutrowska, A. Bzowska, M. Luic and I. Lescic Asler, *J. Enzyme Inhib. Med. Chem.*, 2018, **33**, 1405–1414.
- 40 L. Tao, H. Zou and Z. Huang, *Gastroent. Res. Pract.*, 2014, **2014**, 794342.
- 41 G. Wang, L. F. Lo and R. J. Maier, *DNA Repair*, 2011, **10**, 373–379.
- 42 S. Ansari and Y. Yamaoka, *Helicobacter*, 2017, **22**, DOI: 10.1111/hel.12386.
- 43 Z. Qi, S. Qi, L. Gui, L. Shen and Z. Feng, *Oncol. Lett.*, 2016, **12**, 1959–1964.
- 44 K. Xu, L. Guo, H. Bu and H. Wang, *J. Pharmacol. Sci.*, 2019, **139**, 91–97.
- 45 Y. Liu, L. Liao, Y. Chen and F. Han, *Mol. Med. Rep.*, 2019, **19**, 4673–4684.
- 46 M. Junaid, A. K. Linn, M. B. Javadi, S. Al-Gubare, N. Ali and G. Katzenmeier, *Toxicon*, 2016, **118**, 27–35.
- 47 A. Rehman, S. Akhtar, M. H. Siddiqui, U. Sayeed, S. S. Ahmad, J. M. Arif and M. K. Khan, *Bioinformation*, 2016, **12**, 400–407.
- 48 J. Nystrom and A. M. Svennerholm, *Vaccine*, 2007, **25**, 2591–2598.
- 49 F. Hongying, W. Xianbo, Y. Fang, B. Yang and L. Beiguo, *Clin. Vaccine Immunol.*, 2014, **21**, 126–132.
- 50 M. Rizwan, A. Naz, J. Ahmad, K. Naz, A. Obaid, T. Parveen, M. Ahsan and A. Ali, *BMC Bioinf.*, 2017, **18**, 106.

