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and nuclear lipid droplet dynamic study**

A near-infrared AIE probe was developed for super-resolution imaging of lipid droplets (LDs) and dynamic study of nuclear LDs (nLDs) in physiological process. This work provides a promising tool for LDs studies and contributes to a better understanding of nLDs.

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See Sijie Chen *et al.*,
Mater. Chem. Front., 2021, 5, 3043.



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RESEARCH ARTICLE

View Article Online
View Journal | View IssueCite this: *Mater. Chem. Front.*,
2021, 5, 3043Received 8th November 2020,
Accepted 10th February 2021

DOI: 10.1039/d0qm00914h

rsc.li/frontiers-materials

A near-infrared AIE probe for super-resolution imaging and nuclear lipid droplet dynamic study†

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In contrast to the lipid droplets (LDs) in the cytoplasm, nuclear lipid droplets (nLDs), generated from the metabolically active inner nuclear membrane, contribute to the nuclear lipid homeostasis, locally regulate the availability of signaling lipids, and exchange the proteins between LDs and nuclei. Since the currently available fluorescence tools for imaging and tracking nLDs are limited, exploring a new nLD probe would be highly beneficial for studying the biogenesis and functional significance of nLDs. Herein, we developed a near-infrared probe, namely **DTZ-TPA-DCN**, which possessed the aggregation-induced emission (AIE) property with good biocompatibility for cytoplasm LD and nLD imaging. **DTZ-TPA-DCN** could be used for the super-resolution imaging of LDs with the structured illumination microscopy. By labelling nLDs, we could monitor the nLD formation under an ER stress, and for the first time, we observed that the signaling lipid, namely diacylglycerol (DAG), enhanced the formation of the nLDs. Together, **DTZ-TPA-DCN** provided a promising imaging tool for nLD studies, which contributed to a better understanding of the physiological significance of nLDs.

Introduction

Lipid droplets (LDs) are indispensable in lipid metabolism and energy storage.¹ Although they are primarily responsible for the lipid storage, LDs are also crucial in numerous biological processes, including the regulation of the protein degradation,² the homeostasis of the plasma membrane,³ and cell signaling.⁴ In addition, LDs play a protective role against lipotoxicity and endoplasmic reticulum (ER) stress.⁵ Moreover, researches have also revealed that LDs are implicated in the progression of numerous diseases, such as cancer,⁶ diabetes,⁷ fatty liver disease,⁸ atherosclerosis,⁹ hyperlipidemia,¹⁰ and neurodegeneration.¹¹

In general, LDs are thought to be originated from ER and further translocated to cytoplasm.¹² However, a sub-population of LDs are found inside the nucleus of hepatocytes.¹³ These nuclear lipid droplets (nLDs) are derived from apolipoprotein B-free luminal LDs, which highly accumulate inside the inner nuclear membrane (INM) upon an ER stress.¹⁴ In yeasts, on the

other hand, nLDs are generated in the metabolically active territory of INM, which contributes to the nuclear lipid homeostasis.¹⁵ However, due to the limitation of research tools, the origin and physiological significance of nLDs are still poorly understood.

Fluorescence imaging has been widely used for visualizing biological molecules and structures, tracking physiological and pathological processes, and diagnosing diseases in biological systems.¹⁶ Conventional techniques, such as immunofluorescence and ectopically expressed genes fused with fluorescent protein, are commonly used to monitor the LDs in nucleus, which involve time-consuming experimental procedures.¹⁷ Although a considerable number of fluorescent probes have been developed recently for imaging cytoplasmic LDs, BODIPY 493/503 is the only one used for imaging nLDs.¹⁸ Nevertheless, most of the conventional dyes suffer from the inherent aggregation-caused quenching (ACQ) property of being easily self-quenched when the luminophores are aggregated. Probes with aggregation-induced emission (AIE) properties have received wide attention since the first reporting of the phenomenon in 2001.¹⁹ Compared with the ACQ fluorophores, AIE luminophores show weak or undetectable emissions in dilute solutions but emit strong fluorescence in the aggregated or solid states. These characteristics of AIE luminophores allow the fluorescence imaging with low background, high signal-to-noise ratio, and good photostability, making them ideal tools for the fluorescence imaging of biological targets and even the ultrastructural characterization of biological targets *via* super-resolution imaging.²⁰

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† Electronic supplementary information (ESI) available: Synthesis processes and characterization, photophysical properties, cell viability, and cell imaging results. See DOI: 10.1039/d0qm00914h

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In this contribution, a near-infrared (NIR) fluorescent probe, namely **DTZ-TPA-DCN**, for nLD imaging was developed based on a D- π -A framework, in which non-planar triphenylamine (TPA) was the donor to evade the ACQ effect and the lipophilic cyano groups were the acceptors. Furthermore, the bithiophene and carbon-carbon double bonds were conjugation groups that could extend the absorption or emission wavelength.^{19e}

DTZ-TPA-DCN exhibited AIE properties and could be used for imaging LDs in both live and fixed cells with high selectivity. By using **DTZ-TPA-DCN**, we could clearly observe the LDs in the nucleus of cells in two-dimensional (2D) and three-dimensional (3D) images. Thanks to its good photostability, **DTZ-TPA-DCN** could also be used for structured-illumination microscopy (SIM)-based super-resolution imaging of LDs. Furthermore, the dynamics of LDs inside the nucleus was also highlighted and investigated using **DTZ-TPA-DCN**.

Results and discussion

Synthesis and photophysical properties

DTZ-TPA-DCN was synthesized *via* a simple Knoevenagel condensation reaction of aldehyde and malononitrile (Scheme 1) with 87.5% yield. It was characterized by ¹H NMR, ¹³C NMR and high-resolution mass spectrometry (ESI⁺). The optical properties of **DTZ-TPA-DCN** are illustrated in Fig. 1. From photoluminescence spectrometry results, it was evident that **DTZ-TPA-DCN** showed its maximum absorption peak at 495 nm and near-infrared emission peak at 715 nm with a large Stokes shift of 220 nm (Fig. 1A). The emission spectra displayed a strong solvent effect from toluene (605 nm) to acetonitrile (700 nm), representing a 105 nm bathochromic shift (Fig. S1, ESI[†]). To further investigate the AIE features of **DTZ-TPA-DCN**, acetonitrile/water (MeCN/H₂O) solvent mixture with different H₂O fractions was utilized as the solvent system. As depicted in Fig. 1B and C, **DTZ-TPA-DCN** showed a weak fluorescence emission at 700 nm with a 0.04% quantum yield (QY) in a pure MeCN solution. When adding H₂O to the MeCN solution, the maximum emission gradually red-shifted from 700 nm to 715 nm, and the fluorescence intensity slowly decreased at first as the water fractions were increased from 10% to 40%. This result could be attributed to the twisted intramolecular charge transfer (TICT) effect. When the ratio of water in the mixed solvent was further increased from 40% to 90%, the fluorescence intensity of **DTZ-TPA-DCN** rebounded dramatically due to the formation of the probe aggregates, and the fluorescence intensity of **DTZ-TPA-DCN** in the 90% H₂O solution (QY: 0.28%) was 8.1-fold of that observed in pure MeCN solution. The aggregate formed with an average size of approximately 113.5 nm in diameter was confirmed *via* the dynamic light scattering technique (Fig. 1D). QY in the solid state

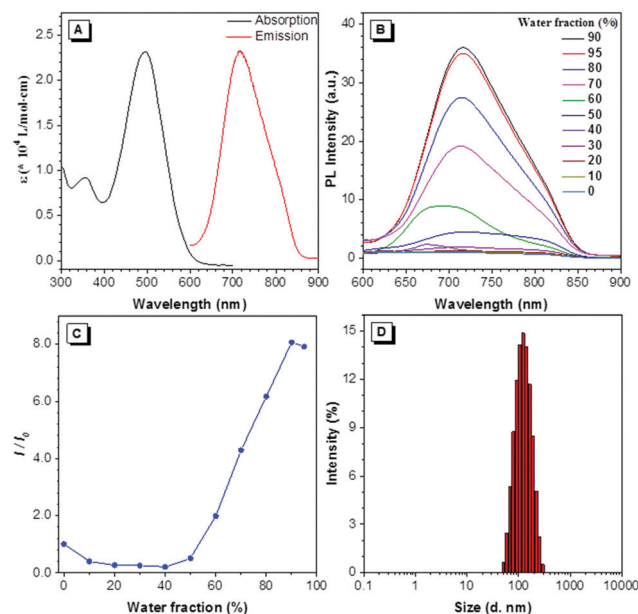


Fig. 1 General optical properties of **DTZ-TPA-DCN**. (A) Molar absorption and emission spectra of **DTZ-TPA-DCN**. (B) Photoluminescence (PL) intensity of **DTZ-TPA-DCN** (10 μ M) in mixtures of MeCN and water with different water contents. (C) Plots of the relative emission intensity of **DTZ-TPA-DCN** versus water fraction. I_0 and I are the peak values of the PL intensities of **DTZ-TPA-DCN** (10 μ M) in MeCN and MeCN/water mixtures, respectively. (D) Size distribution of **DTZ-TPA-DCN** in the mixture of MeCN/water with 90% water content.

is 2.3%, which is much higher than that observed in the aggregated and solution states. As a result, these data affirmatively revealed the AIE characteristics of **DTZ-TPA-DCN**.

Cytotoxicity and co-localization experiment

The cytotoxicity of **DTZ-TPA-DCN** in live cells was evaluated using a standard Cell Counting Kit-8 (CCK-8) assay, and HepG2 cells were treated with different concentrations of **DTZ-TPA-DCN** (from 2.5 to 20 μ M). As illustrated in Fig. S2 (ESI[†]), the cells were still highly viable even after incubated with 20 μ M **DTZ-TPA-DCN** for 24 h, which demonstrated an excellent biocompatibility of **DTZ-TPA-DCN**. We further examined the cellular uptake and the subcellular pattern of **DTZ-TPA-DCN** in HepG2 cells. After incubating the cells with 10 μ M **DTZ-TPA-DCN** for 60 min, a number of punctate structures in the cytoplasm were observed under the confocal laser scanning microscopy. These puncta highly colocalized with the commercial LD probe BODIPY 493/503 (Fig. 2A–C) with a Pearson's correlation coefficient (R_r) of 0.937 (Fig. S3, ESI[†]), which indicated the high labelling specificity of **DTZ-TPA-DCN** to LDs. Besides live-cell imaging, **DTZ-TPA-DCN** also demonstrated a good performance in fixed cell imaging (Fig. S4, ESI[†]). Moreover, as depicted in Fig. 2, 85% of the fluorescence intensity of **DTZ-TPA-DCN**-stained LDs remained unchanged after 100 repetitive scanning under confocal microscopy, while the fluorescence intensity of BODIPY 493/503-stained LDs dropped to only 30% of its original value when using the same scanning parameter (Fig. 2D). Compared with BODIPY 493/503,



Scheme 1 The synthetic route and structure of **DTZ-TPA-DCN**.



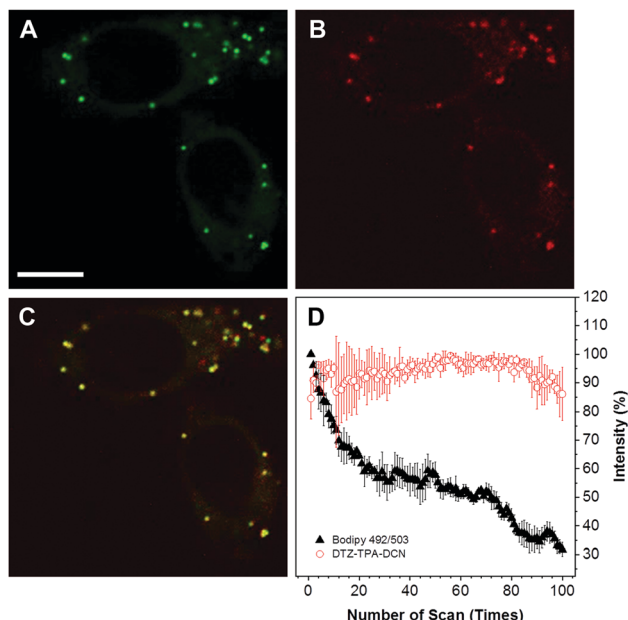


Fig. 2 Fluorescence imaging of HepG2 cells stained with BODIPY 493/503 and **DTZ-TPA-DCN**. (A and B) LDs were labelled by (A) BODIPY 493/503 and (B) **DTZ-TPA-DCN**. (C) Merged fluorescence and brightfield images. Scale bar: 10 μm . BODIPY: 493/503, λ_{ex} = 488 nm, λ_{em} = 510–540 nm; **DTZ-TPA-DCN**: λ_{ex} = 514 nm, λ_{em} = 550–650 nm. (D) Fluorescence intensity of BODIPY 493/503 (solid triangle) and **DTZ-TPA-DCN** (open circle) after 100 times of scanning by a confocal laser scanning microscope.

DTZ-TPA-DCN showed a superior photostability, and this prompted further investigation of its performance under super-resolution fluorescence imaging, which could only be achieved using the fluorophores with an outstanding photostability.²¹

Super-resolution imaging of LDs with SIM

There have been few reports using AIE probes for SIM imaging till now.²² To test the feasibility of using **DTZ-TPA-DCN** for super-resolution imaging, SIM was used in the imaging of **DTZ-TPA-DCN**-stained LDs in HepG2 cells pre-treated with oleic acid (OA). OA is a fatty acid that exists naturally in numerous animal fats and vegetable oils, which can stimulate LD formation in both nucleus and cytoplasm by activating the long-chain fatty acid receptor.²³ After treating HepG2 cells with 0.4 mM OA for 24 h, not only the quantity but also the diameters of LDs in HepG2 cells were increased (Fig. S5, ESI[†]). As shown in Fig. 3, the LD image acquired by the SIM exhibited an enhanced spatial resolution and an improved image quality compared with the images obtained from the confocal laser scanning microscopes (Nikon and ZEISS LSM 880, Fig. 3A, C and Fig. S6, ESI[†]). With respect to SIM, LDs with sharper signals were observed, making them easier to be distinguished from each other in clusters, while the confocal images showed fuzzy LD signals. The line-scan plot of individual LD also demonstrated an enhanced spatial resolution (Fig. 3B and D), where the lateral full-width at half maximum (FWHM) of LDs in the SIM imaging was narrower than those observed in confocal



Fig. 3 **DTZ-TPA-DCN**-stained HepG2 cells captured by the structured illumination microscope (SIM) and confocal microscope. (A and C) The image on the left half is captured by the SIM, where the nucleus is highlighted in blue and the lipid droplets in yellow. The image on the right half is captured by the confocal microscope, where the nucleus is labelled in green and lipid droplets in red. Scale bar: 5 μm . (B) The super-resolution image of the lipid droplets showed more specific fluorescent signals with an FWHM of 450 nm, which was narrower compared to the signals from the confocal microscope (672 nm). (D) Compared to the image obtained from CLSM, the super-resolution image showed a sharper signal that aided in the discrimination of the two adjacent lipid droplets from each other easily.

imaging (Fig. 3B). These results suggested that **DTZ-TPA-DCN** was applicable for imaging LDs by SIM.

Fluorescence imaging of nLDs with DTZ-TPA-DCN

Aside from the cytoplasmic LDs that originated from ER, the nLDs generated in the INM have drawn considerable attention because of its non-canonical role in the nuclear lipid homeostasis.^{14,15} However, the physiological functions, cellular regulatory machinery, and membrane contact site of nLDs remain to be explored. Taking advantage of **DTZ-TPA-DCN** possessing near-infrared emission, good LD selectivity, and practical application to super-resolution imaging, we applied this fluorophore to the study of nLDs. After treating HepG2 cells with OA, the cells were counterstained with 10 μM **DTZ-TPA-DCN** and 20 μM Hoechst 33342. As shown in Fig. 4A, most near-infrared **DTZ-TPA-DCN** puncta localized around the Hoechst-labelled nuclei. However, there were several **DTZ-TPA-DCN**-labelled LDs localized inside the nuclei, which were possibly the nLDs (Fig. 4A, white arrowheads). Through the 3D reconstructed images obtained by the Z-stack acquisition, we further verified that these red puncta were localized inside the nuclei instead of in close proximity to the nuclear surface (Fig. 4B and Fig. S7, Video S1, ESI[†]).





Fig. 4 Analysis of nLDs in HepG2 cells treated with OA. The fluorescence images were acquired by confocal laser scanning microscopy. The nuclei of the HepG2 cells were stained by Hoechst 33342 (blue), and the nLDs were stained by **DTZ-TPA-DCN** (red). (A) The image on the edge represents the orthogonal maximum intensity projection of the cells. (B) The rotated 3D reconstruction of the cell. The white arrowheads in both the images indicate the location of the nLD. Scale bar: 5 μm .

As indicated by the white arrowheads in Fig. 4, the red signal is suspended in the cell nuclei. These results demonstrated the ability of **DTZ-TPA-DCN** to label nLDs and prompted us to further track the dynamics of nLDs by **DTZ-TPA-DCN**.

Monitor the nLD dynamics

To further examine the capability of **DTZ-TPA-DCN** in the monitoring of nLD dynamics, drugs which induced or suppressed the formation of nLDs were applied to the OA-treated HepG2 cells. It was reported that ER stress led to the formation of the nLDs,^{14,24} and tunicamycin (TM) induced the ER stress, which in turn enhanced the nLD formation in OA-treated hepatic cell lines.²⁵ In our experiment, after treating the HepG2 cells with 0.4 mM OA and 5 $\mu\text{g mL}^{-1}$ TM for 12 h, the cells were stained with **DTZ-TPA-DCN**, followed by imaging. We then counted and analyzed the percentage of cells enriched with nLDs in each population with different treatments. For the statistical analysis, around 100 cells were sampled for each of the population in triplicate. As shown in Fig. 5 and Fig. S8 (ESI[†]), the OA-treated group shows a drastic increase in the nLDs compared with that of the control group. When TM was further added into the OA-treated group, the percentage of cells with nLDs also increased from 42.2% to 57.9% (Fig. 5A–C and E).

On the other hand, CP-346086, a microsomal triglyceride transfer protein (MTP) activity inhibitor, was used to suppress the nLD formation.²⁶ When the OA/TM-treated cells were further treated with CP-346086, the percentage of cells with nLDs dropped and restored to a comparable level as compared to the group with the OA treatment only (Fig. 5D and E). Moreover, **DTZ-TPA-DCN** staining also revealed that CP-346086 could suppress the nLD formation with the OA treatment (Fig. S9 and S10, ESI[†]). These results showed that **DTZ-TPA-DCN** could be used for the monitoring of the nLD formation. By using **DTZ-TPA-DCN**, the nLD dynamics in the hepatic cell lines was successfully tracked.

In addition, researches have shown that the active lipid metabolism and the distinct lipid composition of the INM affect the ability of the cells to generate nLDs.¹⁵ For instance, triacylglycerols (TAGs), which are the major neutral lipids stored in LDs, are essential factors in the biogenesis of LDs.^{1b,27} To promote the formation of LDs, the TAG in cells needs to reach a critical concentration.²⁸ Moreover, the synthesis of TAGs from DAGs was catalysed by diacylglycerol acyltransferase (DGAT). Interestingly, DAGs were also highly related to the formation of nLDs.^{1b,27,29} Therefore, we would like to investigate whether the addition of DAG could promote the hepatocellular nLD formation. As shown in Fig. 6 and Fig. S11 (ESI[†]), by using **DTZ-TPA-DCN** staining, we can observe that the nLDs were unevenly dispersed in cells with 0.4 mM OA treatment. Interestingly, the percentage of cells containing nLDs increased from around 44.5% to more than 61.4% when the cells were treated with 0.4 mM OA and DAG (10 μM), suggesting that DAG enhanced the formation of nLDs in the hepatic cells (Fig. 6C). Some researchers pointed out that the seipin-deficient yeasts might form more nLDs when compared to the wild-type yeasts, and DAG is one of the factors involved in this phenomenon. The lack of seipin is believed to cause an increase in phosphatidic acid (PA), a precursor of DAG, in yeasts. When the free PA level inside the cells increased, excess PA would diffuse into the INM. These PAs would then be converted to DAG by PA phosphatases, which further upregulated the TAG synthesis and eventually contributed to more nLD formation.^{15,27} Despite this pathway was demonstrated using yeasts as a model organism only, it was postulated that the connection of lipid metabolism and nLD biogenesis were evolutionarily conserved in higher eukaryotes. This experiment suggested that **DTZ-TPA-DCN** could be a useful



Fig. 5 Monitoring the nLD formation using **DTZ-TPA-DCN**. (A–C) HepG2 cells treated with 0.4 mM of OA showed an increase in nLDs; nearly 60% of the cells had nLDs in the OA/TM (0.4 mM OA and 5 $\mu\text{g mL}^{-1}$ TM) treatment. (D) Addition of CP-346086 (100 nM) to OA/TM-treated cells inhibited the nLD formation, and the proportion of cells having the nLDs was comparable to that of the OA-treated group. Scale bar: 10 μm . (E) Percentage of cells with nLDs in each group. (Data were from the corresponding images of A–D.) Data are represented as mean \pm SEM from at least three independent experiments. $n = 29/30/25/27$, $*P < 0.05$.



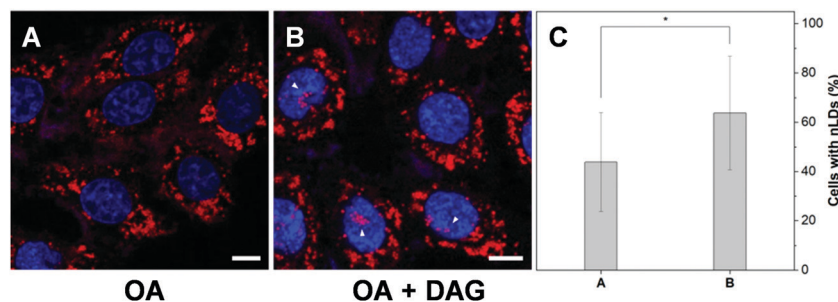


Fig. 6 Monitoring the nLD change using **DTZ-TPA-DCN** during the DAG treatment. (A and B) Fluorescence images of the OA (0.4 mM) and OA (0.4 mM) plus DAG (10 μ M)-treated group acquired by confocal laser scanning microscopy. Scale bar: 10 μ m. (C) The percentage of the HepG2 cells with nLDs increased from around 44.5% to 61.4% when the cells were further treated with DAG. Data were from the corresponding images of (A) and (B). Data are represented as mean \pm SEM from at least three independent experiments. $n = 31/29$, $*P < 0.05$.

tool to explore the unravelled mechanism of nLD biogenesis and regulation.

Conclusions

In summary, a lipophilic near-infrared fluorescent probe, namely **DTZ-TPA-DCN**, was developed based on a common D- π -A framework *via* a simple Knoevenagel condensation reaction. **DTZ-TPA-DCN** exhibited AIE properties with large Stokes shift and a distinct solvatochromic effect with high sensitivity towards environmental polarity. It was highly biocompatible and was able to label LDs specifically in both live and fixed cells. Owing to its excellent photostability, **DTZ-TPA-DCN** was successfully used in the SIM imaging of LDs. Furthermore, the nLDs were clearly observed in 2D or 3D imaging using **DTZ-TPA-DCN** in the labelling. The favourable optical properties of **DTZ-TPA-DCN** enabled further monitoring and tracking of the nLD dynamics under ER stress or by a lipid metabolism of the INM. It was shown for the first time that the important signaling lipid, DAG, induced nLD formation in mammalian cells. Conclusively, this study provided a versatile and reliable tool, **DTZ-TPA-DCN**, in the studies of nLDs, and its great potential for exploring the functional significance of nLDs was envisioned.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

M.-Y. W. thanks the National Natural Science Foundation of China (21708030) and the Applied Basic Research of Sichuan Province (2021YJ0397). S. C. acknowledges the start-up funding from Ming Wai Lau Centre for Reparative Medicine, Karolinska Institutet.

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