

Cite this: *Polym. Chem.*, 2021, **12**, 4696

The effects of cononsolvents on the synthesis of responsive particles *via* polymerisation-induced thermal self-assembly†

Marissa D. Morales-Moctezuma and Sebastian G. Spain *

Nanogels have emerged as innovative platforms for numerous biomedical applications including gene and drug delivery, biosensors, imaging, and tissue engineering. Polymerisation-induced thermal self-assembly (PITSA) has been shown to be a suitable route for the synthesis of temperature-responsive nanogels, and cononsolvents have been used to improve the solubility of comonomers. Here, we show that use of cononsolvents during synthesis also has significant effects on the properties of the final nanogels. Responsive nanogels consisting of a poly(acrylic acid) corona and a crosslinked poly(*N*-isopropylacrylamide) core were synthesised *via* RAFT-mediated PITSA in water and water/EtOH mixtures. Nanogels synthesised in water were found to increase in size as the degree of polymerisation (DP_n) of the pNIPAM block was increased. For a fixed pNIPAM DP_n , particle sizes were seen to decrease with increasing EtOH content in the synthesis solvent. This is rationalised by the decreasing solubility of the growing pNIPAM in the reaction medium resulting in earlier particle nucleation. The temperature response of the particles was also found to be dependent on the synthesis conditions with increasing EtOH content in the synthesis solvent increasing the volume phase transition temperature. Model poly(acrylic acid)-*block*-poly(*N*-isopropylacrylamide) copolymers synthesised under similar conditions were seen to have differing tacticities dependent on the synthesis solvent with increasing EtOH content resulting in increased syndiotacticity and increased cloud point temperatures. We propose that the tacticity changes are due to the PITSA process, with the constrained environment of the particle resulting in increased syndiotacticity.

Received 23rd March 2021,
Accepted 19th July 2021

DOI: 10.1039/d1py00396h

rsc.li/polymers

Introduction

Nanogels/microgels are colloidal particles with a diameter in the sub-micron region and have emerged as innovative materials for numerous biomedical applications including gene and drug delivery,^{1,2} biosensors,³ imaging,⁴ and tissue engineering.⁵ Various methodologies have been reported for the preparation of nanogels. They include the crosslinking of prepolymers by non-covalent interactions (*e.g.* hydrophobic or ionic interactions)^{6–11} or the formation of covalent crosslinks between reactive functional groups of polymer chains using chemistries such as thiol-disulfide exchange,^{12–15} click

reactions,^{16,17} amidation of activated esters,^{18–20} and photoreactions.^{21–25} Moreover, covalently crosslinked nanogels are also prepared by the heterogeneous radical (co)polymerisation of vinyl monomers and a crosslinker in a one-pot synthesis. Inverse precipitation,^{26–29} (inverse) mini- or microemulsion,^{30–38} and dispersion^{39,40} processes are relevant techniques used in the synthesis of nanogels by either free radical or controlled radical polymerisation techniques. Although well-defined crosslinked nanoparticles can be prepared by these heterogeneous techniques, their main drawbacks are the use of molecular surfactants or potentially toxic solvents.⁴¹ In order to avoid these issues, nanogel formation using surfactant-free aqueous dispersion polymerisation has emerged as a versatile way to prepare colloidally stable nanoparticles from solvophilic precursors.^{42,43} For example, An *et al.* used poly(*N,N*-dimethylacrylamide) (pDMA) to both stabilise the *in situ* formation of nanoparticles and to mediate the reversible addition–fragmentation chain transfer (RAFT) polymerisation of *N*-isopropylacrylamide (NIPAM) in the presence of *N,N'*-methylenebisacrylamide (BIS) crosslinker.⁴³ This aqueous RAFT dispersion approach is based on a polymerisation-induced self-assembly (PISA) process. PISA is a poly-

Polymer and Biomaterials Chemistry Laboratories, Department of Chemistry, University of Sheffield, Sheffield S3 7HF, UK. E-mail: s.g.spain@sheffield.ac.uk

† Electronic supplementary information (ESI) available: NMR spectra, SEC chromatograms and summary data for pAA macroCTAs; synthesis and cloud point measurements of pNIPAM in different solvents; DLS (including correlation functions), TEM and additional summary data for nanogels; determination of nanogel VPTT, NMR spectra and SEC chromatograms for pNIPAM synthesised by precipitation polymerisation in different solvents; cloud points and summary data for diblock copolymers. See DOI: 10.1039/d1py00396h



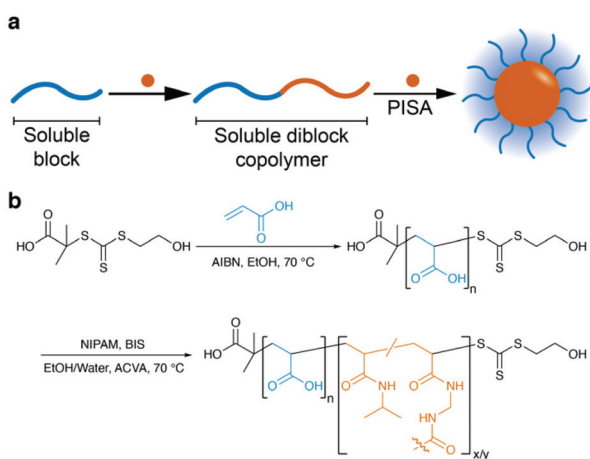
merisation technique whereby a soluble homopolymer (stabiliser) is chain extended with a second monomer that forms an insoluble polymer in the polymerisation solvent. As the second block grows and becomes insoluble, polymer chains assemble into nanoparticles stabilised with the soluble block and the insoluble block as the core (Scheme 1a). Further extension of the insoluble block allows control of particle size as well as access to higher morphologies such as cylindrical micelles (or worms) and vesicles (polymersomes).^{44–46}

Polymerisation-induced thermal self-assembly (PITSA) is a derivative of PISA whereby a temperature-responsive polymer, *e.g.* pNIPAM, is grown from the stabiliser block above its lower critical solution temperature (LCST) resulting in thermally-induced self-assembly during the polymerisation.^{43,47} Addition of a crosslinker during the PITSA process allows the synthesis of temperature-responsive pNIPAM nanogels that do not dissociate on cooling, instead they shrink and swell above and below a volume phase transition temperature (VPTT), respectively.⁴³

So far, PITSA has been used to synthesise nanogels under aqueous dispersion conditions from temperature-responsive monomers such as NIPAM,^{43,48–50} *N,N*-diethylacrylamide (DEAM),^{51,52} oligo(ethylene glycol)methacrylates (OEGMAs),⁵³ oligo(ethylene glycol)acrylates (OEGAs),^{49,54} and 2-(dimethylamino)ethyl methacrylate (DMAEMA).⁵⁵ It is noteworthy that while this technique is environmentally friendly since it is performed in water and high-yielding, its use is limited to water-soluble monomers whose polymer becomes less soluble during polymerisation. To overcome this limitation, some researchers have explored the addition of small quantities of a solvent with low toxicity such as alcohols to water as a route to include hydrophobic components (*e.g.* additional monomers) into the nanogel structure^{56,57} or to promote the immiscibility of growing chains during polymerisation.^{57–59} Evidently, water is chosen as the major solvent and an alcohol (*e.g.* ethanol,^{56–58} *tert*-butanol,⁵⁹ or 2-propanol^{60–62}) is selected as the minor solvent. For example, Xu *et al.* used a mixture of

water/ethanol (75/25, v/v) for the RAFT dispersion polymerisation of NIPAM and functional comonomers (*e.g.* hydrophobic monomers, fluorophores) with ethanol allowing solubilisation of the additional monomers.⁵⁶ This binary solvent strategy was later used for the synthesis of doubly thermoresponsive triblock nanogels that consisted of a poly(diethylene glycol ethyl acrylate) (pDEGA) outer block, a pDMA midblock, and a cross-linked pNIPAM core block using a mixture of water/ethanol (65/35, v/v).⁵⁸ In this case, ethanol allowed better solubilisation of pDEGA thus promoting the location of pDEGA in the corona and the pNIPAM block in the core of the nanogel. In a traditional aqueous PISA, the block with the lowest LCST (*i.e.* pDEGA) would otherwise be located in the core of the structure and the block with the highest LCST (*i.e.* pNIPAM) in the outer shell.⁵⁸ By using this approach, the library of nanogels synthesised by RAFT dispersion polymerisation was expanded to different compositions and functionalities that were inaccessible by using only water during synthesis. Nonetheless, it is well-established that mixtures of water and ethanol show cononsolvency for pNIPAM at certain mole fractions despite being good solvents for pNIPAM individually.⁶³ For instance, the lower critical solution temperature (LCST) of pNIPAM is around 31–35 °C in pure water,^{64,65} but in mixtures of cononsolvents (*e.g.* ethanol,^{66,67} methanol,^{68–70} THF,^{71,72} DMSO,^{73,74} and acetone^{74,75}) its LCST is lowered when small amounts of the organic solvent are added. Hence, the cononsolvent composition is likely to influence the *in situ* self-assembly process when conducting the RAFT dispersion polymerisation of NIPAM.

Herein, we aimed to investigate the effects of the synthesis cononsolvent composition on the properties of pNIPAM nanogels prepared by RAFT-mediated PITSA in water and water/ethanol mixtures. To achieve this, the RAFT dispersion polymerisation of NIPAM in the presence of BIS was conducted using poly(acrylic acid) (pAA) as the macromolecular chain transfer agent (macroCTA) to synthesise multi-responsive crosslinked particles (pAA-*b*-pNIPAM/BIS) (Scheme 1b). The nanogel sizes were found to be strongly dependent on the solvent mixture used during their synthesis. The thermal properties of the nanogels were also found to be dependent on the solvent used during the synthesis and this is demonstrated to be due to the different tacticity of the pNIPAM core.



Scheme 1 (a) Schematic of the polymerisation-induced self-assembly process. (b) Synthesis of pAA_n-*b*-pNIPAM_x/BIS_y nanogels.

Results and discussion

Synthesis of poly(acrylic acid) macro-chain transfer agents (pAA macroCTAs)

The synthesis of pAA-*b*-pNIPAM/BIS is outlined in Scheme 1b. Poly(acrylic acid) was chosen as the stabiliser block as it is soluble in all water/EtOH compositions and highly charged stabiliser blocks are known to prevent the formation of higher order morphologies (*e.g.* worm-like micelles and vesicles) during PISA,^{76–78} thereby simplifying data analysis by minimising the probability of additional phase transitions that may complicate data interpretation. A poly(acrylic acid) (pAA)



macroCTA was synthesised by RAFT polymerisation of acrylic acid in ethanol with 2-(hydroxyethylthiocarbonothioylthio)-2-methylpropanoic acid (HEMP) as the chain transfer agent. An initial kinetic experiment determined that high acrylic acid conversions (~90%) were obtained after 5 h of polymerisation. Subsequent polymerisations were stopped at approximately 70% conversion to ensure high end-group fidelity in the subsequent block copolymer and nanogel syntheses.

The number-average degree of polymerisation (DP_n) for the pAA macroCTA was 45 as determined by end-group analysis by 1H NMR spectroscopy. The pAA macroCTA was characterised by size exclusion chromatography (SEC) in THF/AcOH against pMMA standards after methylation with trimethylsilyl diazomethane and found to be mono-modal and of low dispersity ($M_n = 4.6 \text{ kg mol}^{-1}$, $M_w/M_n = 1.19$). Full characterisation (1H NMR spectra, SEC chromatograms and molar masses) of the pAA macroCTA may be found in the ESI (Fig. S1–5 and Table S1†).

Synthesis and characterisation of pAA-*b*-pNIPAM/BIS nanogels in water

Prior to investigating the effect of consolvency on nanogel synthesis a series of pAA₄₅-*b*-pNIPAM_{*x*}/BIS₃ ($x = 87, 175$ and 262) nanogels were synthesised in water. The pAA₄₅ macroCTA was chain extended with *N*-isopropylacrylamide (NIPAM) and *N,N'*-methylene bisacrylamide (BIS) *via* RAFT-mediated aqueous dispersion polymerisation at pH 6.7. This pH was chosen to ensure that the pAA was in a highly charged state (pK_a 5.8) and was reached by addition of a minimal volume of NaOH solution to minimise any changes to ionic strength that may affect the self-assembly. The resulting nanogel dispersions were purified by dialysis against deionised water and characterised with dynamic light-scattering (DLS), transmission electron microscopy (TEM) and ζ -potential measurements (Table 1, entries 1–3).

During the PITSA process, the growing pNIPAM chains will reach a critical degree of polymerisation at which they will undergo a coil-to-globule transition and aggregate into nascent particles with a pNIPAM core and a pAA shell. As pAA is highly

negatively charged at neutral pH, it was expected that the synthesised nanogels would be highly negatively charged and this is confirmed by their measured ζ -potentials (−41 mV to −29 mV). As the length of the pNIPAM block is increased, it is expected that the size of the core will increase with the increasing volume fraction of pNIPAM. From DLS measurements (Fig. 1a, Fig. S6† and Table 1, entries 1–3), it is clear that the hydrodynamic diameter, D_h , increases from 80 nm for pAA₄₅-*b*-pNIPAM₈₇/BIS₃ nanogels to 483 nm for pAA₄₅-*b*-pNIPAM₂₆₂/BIS₃ nanogels. As the volume fraction of pNIPAM is increased, it is also expected that higher order morphologies may be formed. For example, Figg *et al.* found that nano-objects formed during PITSA of NIPAM from a pDMA₃₄-*b*-p(DMA-*st*-AA)_{14/6} macroCTA were spherical when the pNIPAM DP_n was 11 and 52 but worms and vesicles were observed with DP_n of 101 and 137, respectively but required post-polymerisation crosslinking as a way to prevent dissolution of the chains during cooling.⁴⁷

Here, when a pNIPAM DP_n of 87 was targeted only spheres were evident in the TEM micrographs (Fig. 1b). For higher pNIPAM DP_n (175 and 262) particle morphologies were irregular and appear to be aggregates of smaller spherical particles. The formation of spheres rather than higher morphologies has been seen in PISA systems where a highly-charged stabiliser block is used^{76,78} and is thought to be due to charge repulsion preventing particle fusion to form worms.⁷⁷ Additionally, the presence of a crosslinker in the system may result in particles being trapped in a spherical morphology as the chains cannot reorganise. This may also explain the aggregate morphology seen in the pNIPAM samples with DP_n 175 and 262 as particle growth also occurs through coalescence of smaller particles which will be inhibited by partial crosslinking during the polymerisation and this is supported by asymmetric flow field flow fractionation studies of related nanogels.⁷⁹

Consolvents used during synthesis affect the size of pAA-*b*-pNIPAM/BIS nanogels

Despite individually being good solvents for pNIPAM, water and ethanol are known to display consolvency for pNIPAM

Table 1 Data for the synthesis of pAA₄₅-*b*-pNIPAM_{*x*}/BIS₃ ($x = 87, 175$ and 262) nanogels by RAFT dispersion polymerisation in either water or water/ethanol mixtures

Entry	Target composition [pAA ₄₅]:[NIPAM]:[BIS] ^a	X_{EtOH} ^b	Conversion ^c / %	$DP_{n,theo}$ ^d	$M_{n,theo}$ ^e / kg mol ⁻¹	$D_h \pm S.D./nm$ (PDI) ^f	ζ -Potential \pm S.D. ^g / mV
1	[1]:[87]:[3]	0	74	65	11.3	80 \pm 26 (0.106)	−40.9 \pm 0.7
2	[1]:[175]:[3]	0	90	158	21.8	231 \pm 64 (0.076)	−34.2 \pm 2.0
3	[1]:[262]:[3]	0	84	220	28.8	483 \pm 121 (0.063)	−29.9 \pm 0.7
4	[1]:[175]:[3]	0.06	98	172	23.4	159 \pm 37 (0.053)	−31.1 \pm 1.9
5	[1]:[175]:[3]	0.12	100	175	23.7	97 \pm 41 (0.175)	−34.0 \pm 2.7
6	[1]:[175]:[3]	0.14	92	161	22.2	90 \pm 42 (0.222)	−22.8 \pm 3.6

^a General reaction conditions: Targeted [pAA₄₅]/[ACVA] = 5, solids content 10% w/w, 70 °C for 18 h. ^b Mole fraction of ethanol used during nanogel synthesis. ^c Gravimetric determination by moisture analysis of solids content against predicted solids content. ^d For an equivalent diblock synthesised without crosslinker, estimated from [NIPAM]/[pAA₄₅] \times conversion. ^e Calculated using $M_{n,theo} = M_c + nM_{AA} + xM_{NIPAM} + yM_{BIS}$, where M_c is the molecular weight of the CTA end groups, n , x and y are the DP_n of pAA, pNIPAM, and BIS respectively. ^f DLS data obtained from 0.1% w/v nanogel dispersions in DI water at pH 7 and 25 °C. ^g Average zeta-potential and the standard deviation of five run measurements for 0.1% w/v nanogel dispersions in DI water with 1 mM KCl at pH 7 and 25 °C.



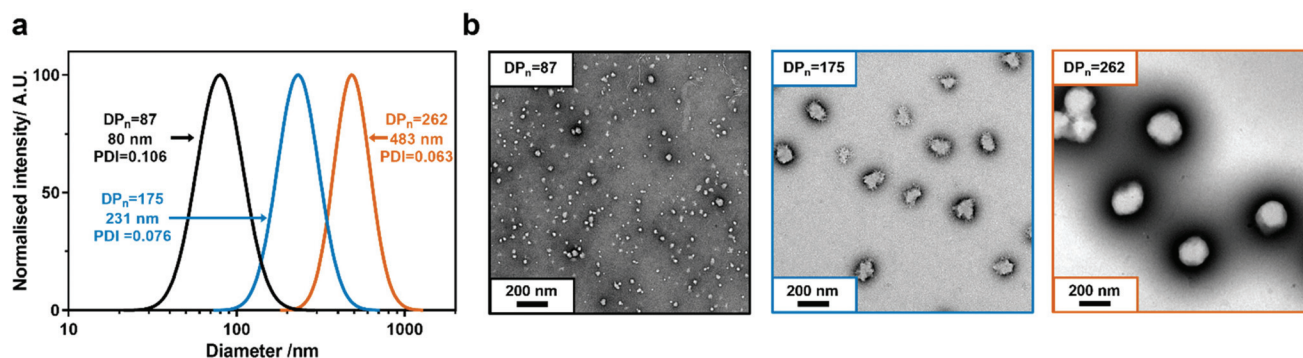


Fig. 1 Characterisation of pAA₄₅-*b*-pNIPAM_{*x*}/BIS₃ (*x* = 87, 175 and 262) nanogels synthesised in water. (a) Intensity average log normal size distributions from dynamic light-scattering. (b) Transmission electron micrographs. DP_{*n*} indicates the targeted DP of pNIPAM. DLS and TEM samples were prepared from 0.1% w/v nanogel dispersions in water at pH 7 at 25 °C. TEM samples were stained with phosphotungstic acid (0.75% w/v, pH 7).

at certain mole fractions.^{66,67} Xu *et al.* utilised this conolvency to synthesise poly(*N,N*-dimethylacrylamide)-*b*-pNIPAM/BIS nanogels containing a range of functional monomers with the addition of ethanol allowing solvation of hydrophobic monomers.⁵⁶ The composition of the con solvent is known to have a significant effect on the lower critical solution temperature (LCST) of pNIPAM⁸⁰ and can also cause an upper critical solution temperature at high mole fractions of the organic solvent.⁸⁰ Additionally, Wen *et al.* have recently demonstrated that solvent quality for the core-forming block can affect self-assembly during PISA.⁸¹ Consequently, it was expected that it would also have an effect on the nucleation and self-assembly processes during PITSA.

pAA₄₅-*b*-pNIPAM₁₇₅/BIS₃ nanogels were synthesised as before in water/EtOH mixtures at pH 6.7. The resulting nanogel dispersions were purified by dialysis against deionised water and characterised with dynamic light-scattering (DLS), transmission electron microscopy (TEM) and ζ-potential measurements (Fig. 2, Fig. S7† and Table 1, entries 4–6). For all nanogels, the ζ-potentials were strongly negative (−34 to −23 mV) as expected. Nanogel sizes measured by DLS in water were found to be strongly dependent on the composition of the solvent used during the synthesis. When synthesised in

water alone, pAA₄₅-*b*-pNIPAM₁₇₅/BIS₃ nanogels had a hydrodynamic diameter of 231 nm. For nanogels synthesised in 0.06 mol% EtOH ($X_{\text{EtOH}} = 0.06$), D_h was drastically reduced to 159 nm with no increase in the polydispersity of the particles. Further increases in EtOH concentration resulted in further reduction in nanogel hydrodynamic diameter ($D_h = 97$ nm and 90 nm for $X_{\text{EtOH}} = 0.12$ and $X_{\text{EtOH}} = 0.14$, respectively). TEM micrographs of the nanogels showed a similar reduction in size as well as a shift from the “aggregated spherical particle” morphology to individual spherical particles as the ethanol content was increased (Fig. 2 and S8†). Similar behaviour was seen for nanogels synthesised with targeted pNIPAM DP_{*n*} of 87 and 262 in con solvents (Fig. S9 and S10, Table S2†).

Wen *et al.* recently reported a similar effect in the PISA synthesis of poly(potassium 3-sulfopropyl methacrylate)-*b*-poly(benzyl methacrylate) nanoparticles where synthesis in alcohol/water mixtures resulted in increased particle size compared to the synthesis in water alone.⁸¹ This increase was rationalised by the increased solubility of the benzyl methacrylate block with increasing alcohol content allowing greater chain mobility and solubility and therefore the greater exchange of chains being growing particles. Here, the decrease in nanogel diameter with increasing ethanol content during

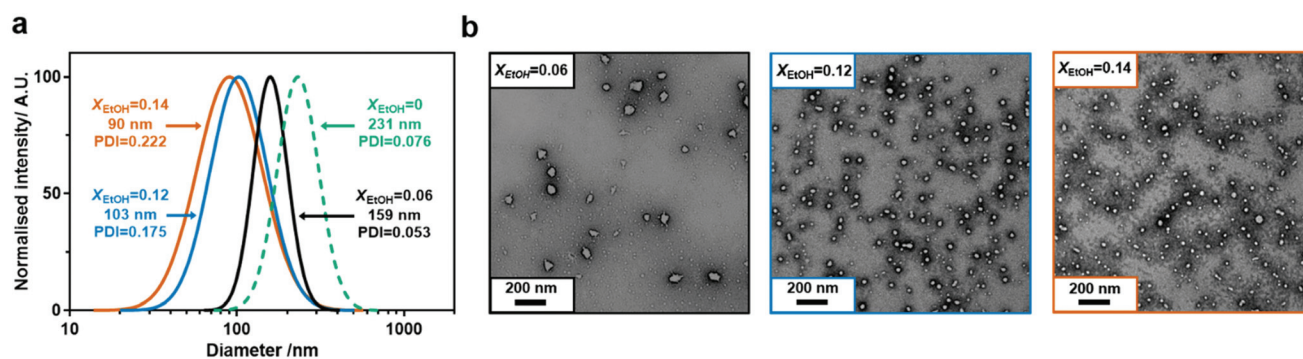


Fig. 2 Characterisation of pAA₄₅-*b*-pNIPAM₁₇₅/BIS₃ nanogels synthesised in water/EtOH mixtures. (a) Intensity average log normal size distributions from dynamic light-scattering. (b) Transmission electron micrographs. DLS and TEM samples were prepared from 0.1% w/v nanogel dispersions in water at pH 7 at 25 °C. TEM samples were stained with phosphotungstic acid (0.75% w/v, pH 7).



the PITSA process is not unexpected and can be rationalised when the effect of the cosolvent on the self-assembly process is considered. During PISA nascent particles are nucleated when the core-forming block reaches a critical DP_n where it becomes insoluble. In PITSA, this critical DP_n should be related to the LCST of the core-forming polymer in the synthesis solvent. pNIPAM is generally considered to have an LCST of 32 °C in water but cloud point measurements indicate this is reduced to below 25 °C when $X_{\text{EtOH}} = 0.05$ and below 10 °C when the $X_{\text{EtOH}} = 0.1$ (Fig. S11†), indicating that pNIPAM is significantly less soluble in the cosolvent mixtures than in water alone. Consequently, the critical DP_n at which particle nucleation occurs should be lower with increasing X_{EtOH} within the range of cosolvent compositions investigated here. To confirm this, pNIPAM homopolymers were synthesised *via* precipitation polymerisation in water and water/EtOH mixtures and it was found that the M_n of the resulting polymers was strongly dependent on the synthesis solvent (Fig. S12†) with increasing EtOH content during the synthesis reducing the M_n of the pNIPAM (*e.g.* $X_{\text{EtOH}} = 0$, $M_n = 280 \text{ kg mol}^{-1}$ versus $X_{\text{EtOH}} = 0.15$, $M_n = 35 \text{ kg mol}^{-1}$). These data indicate that pNIPAM becomes insoluble at a lower DP_n as the EtOH content increases for the molar fractions tested here, consequently the critical DP_n should be lower. As the critical DP_n is lower, then particle nucleation should occur earlier in the polymerisation. As crosslinking is more probable within particles, where the polymer (and possibly monomer) concentration is higher, than in between dissolved chains then earlier nucleation of particles should lead to smaller particles due to the spherical morphology being “trapped” earlier in the polymerisation. It should be noted that variations in pH or ionic strength could also affect the PITSA process by changing the conformation/volume of the pAA stabiliser. However, addition of ethanol changes the dielectric constant of the solvent⁸² which will also effect the pK_a of the pAA stabiliser and the apparent ionic strength and will likely have a larger effect than any small differences the quantities of reagents added.

Consolvents used during synthesis affect the thermal properties of nanogels

As the nanogels contain a pNIPAM core, they are expected to display temperature-responsive properties. These nanogels are already a dispersion and therefore do not display a cloud point or lower critical solution temperature in the same sense as a linear pNIPAM sample. Instead, these nanogels display a volume phase transition temperature (VPTT) at which the crosslinked pNIPAM core collapses, resulting in a decrease in the particle diameter at this temperature. Particle size changes with temperature were measured using variable temperature DLS studies at an aqueous pH of 7. As expected, all of the nanogels were temperature-responsive (Fig. 3a). The nanogels synthesised in X_{EtOH} 0, 0.06 and 0.12 clearly decrease in volume upon heating but this decrease is less pronounced for smaller nanogels where the core will have a smaller contribution to the overall particle volume. VPTTs of these nanogels were determined as the halfway point between the swollen and

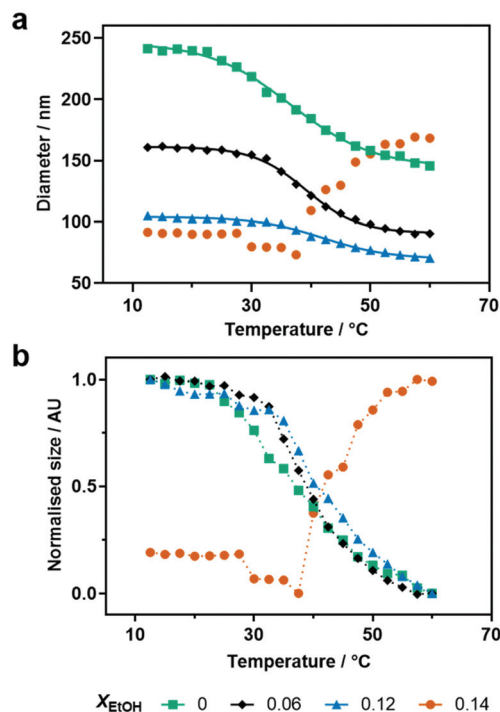


Fig. 3 (a) Variable temperature DLS data for pAA₄₅-*b*-(pNIPAM₁₇₅/BIS₃) nanogels synthesised in water and water/EtOH mixtures. (b) Normalised sizes. DLS measurements were obtained from 0.1% w/v nanogel aqueous dispersions at pH 7.

collapsed states (eqn (S1)†). Interestingly, the VPTT increased with the X_{EtOH} used during the synthesis. When synthesised in water alone the pAA₄₅-*b*-pNIPAM₁₇₅/BIS₃ nanogel had a VPTT of 37 °C, but nanogels synthesised in $X_{\text{EtOH}} = 0.06$ and 0.12 had VPTTs of 39 °C and 42 °C, respectively. VPTTs were higher than the VPTT for a pNIPAM gel (~32 °C)⁸³ due the hydrophilic characteristics of pAA at pH 7.⁸⁴ Note that, although there may be small variations in ionic strength between samples, the LCST of pNIPAM is relatively insensitive to salts at the low concentrations that may occur due to experimental variation with the LCST of pNIPAM-coated silica particles only decreasing by 1 °C in the presence of 50 mM KCl.⁸⁵ Since the nanogels had similar monomer compositions, the shift in the VPTT with increasing X_{EtOH} used during synthesis should be related to differences caused by the particle nucleation. The VPTT shift is more clear when the sizes are normalised (Fig. 3b). Notably, the onset of shrinking is also affected by the cosolvent composition used during the synthesis. The nanogels synthesised in water alone had an onset of shrinking at 21 °C, whereas the ones synthesised in X_{EtOH} 0.06 and 0.12 were at 29 °C and 34 °C, respectively. For nanogels synthesised in X_{EtOH} 0.14, there is a second transition with an increase in D_h at approximately 38 °C indicating that flocculation occurs above this temperature, which is supported by an increase in the PDI (Fig. S13†).

Synthesis and properties of pAA-*b*-pNIPAM block copolymers

The LCST of pNIPAM (and VPTT of pNIPAM gels) is known to be sensitive to end groups or, in the case of block copolymers,



the other blocks.⁸⁶ For example, Xia *et al.* demonstrated that that an *N*-phenyl amide end group lowered the cloud point of pNIPAM by as much as 9 °C compared to the unsubstituted amide.⁸⁷ Additionally, the tacticity of pNIPAM is also known to affect its LCST and the tacticity can be affected by the synthesis conditions, *e.g.* the solvent.^{88,89} For example, Biswas *et al.* demonstrated that the isotacticity of pNIPAM synthesised in presence of 0.1 M Y(OTf)₃ increased from 55% to 86% as the methanol mole fraction in water was increased from 31 to 100 mol%.⁸⁹ Additionally, the cloud point of nanoparticles coated with responsive polymers is known to be dependent on the particle size.⁹⁰

To understand how these factors affect the VPTT of pAA-*b*-pNIPAM/BIS nanogels, pAA-*b*-pNIPAM diblock copolymers were synthesised as soluble analogues that are more amenable to detailed characterisation.

A pAA₃₄ macroCTA ($M_n = 3.2 \text{ kg mol}^{-1}$, $M_w/M_n = 1.20$) was used to prepare a series of pAA₃₄-*b*-pNIPAM_x ($x = 92, 184$ and 280) diblock copolymers by RAFT dispersion polymerisation of NIPAM in either water or water/ethanol mixtures targeting nominal pNIPAM DP_n between 92 and 280 (Table S3†). The shorter pAA macroCTA was chosen to reduce the pAA contribution during later characterisation (*e.g.* in the ¹H NMR spectra). These conditions were chosen to mimic those used in the nanogel synthesis but in the absence of crosslinker. In all cases, the NIPAM conversions were at least 99% as determined by ¹H NMR spectroscopy. pAA₃₄-*b*-pNIPAM_x diblock copolymers were purified by exhaustive dialysis against deionised water for 48 h with repeated water changes before characterisation (Table 2 and Table S3†). ¹H NMR spectra of the purified copolymers (Fig. 4) were all very similar with only minor variations in the resonances for the backbone protons (δ 1.4–2.1 ppm) and the DP_n measured to be as expected. As these nano-objects scatter light, it is possible to measure the cloud point of the diblock copolymers using turbidimetry. A clear increase in the cloud point temperature, T_{cp} , from 34 °C to 39 °C was observed as X_{EtOH} was increased from 0 to 0.15

Table 2 Data for pAA₃₄-*b*-pNIPAM_x diblock copolymers synthesised in water and water/EtOH mixture

Targeted pNIPAM DP _n ^a	X_{EtOH} ^b	Conversion ^c / %	Cloud point (T _c) ^e /°C	DP _{n,theo} ^d	D _h ± S.D. ^f / nm (PDI)
184	0	>99	34	184	49 ± 12 (0.06)
184	0.06	>99	36	184	67 ± 31 (0.21)
184	0.12	>99	39	184	93 ± 44 (0.22)
184	0.15	99	39	182	140 ± 56 (0.16)

^a General reaction conditions: Targeted [pAA₃₄]/[ACVA] = 5, solids content 10% w/w, 70 °C for 18 h. ^b Mole fraction of ethanol used during nanogel synthesis. ^c Determined by ¹H NMR spectroscopy using the ratio of the sum of vinyl proton integrals of NIPAM and the polymer backbone. ^d Estimated from [NIPAM]/[pAA₃₄] × conversion. ^e Cloud points were estimated from the inflection point of the normalised absorbance curve of 1% w/v dispersions in water at 550 nm at a heating rate of 0.12 °C min⁻¹. ^f DLS data obtained from 0.1% w/v dispersions in DI water at pH 7 and 50 °C.

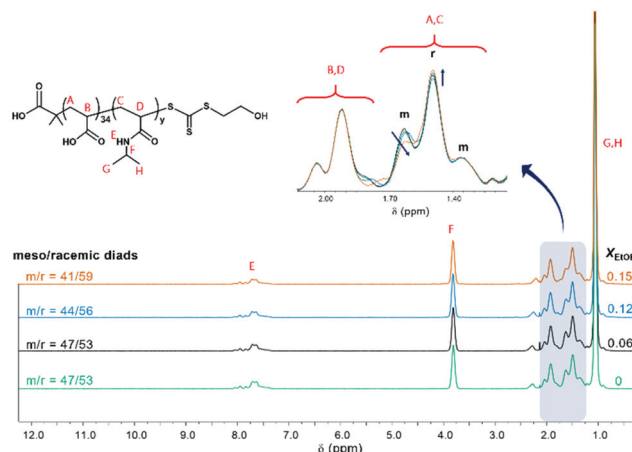


Fig. 4 ¹H NMR spectra of purified pAA₃₄-*b*-pNIPAM₁₈₄ diblock copolymers synthesised in water and water/EtOH mixtures. The r and m labels are used to denote the resonances attributed to the racemic and *meso* diads, respectively. Spectra were recorded in D₂O.

(Table 2, Table S3 and Fig. S14†). The diblock copolymers were also found to self-assemble above their T_{cp} to form defined nano-objects. For the pAA₃₄-*b*-pNIPAM₁₈₄ diblock copolymers there is a clear difference in the size of the resulting nano-objects depending on the X_{EtOH} used during the synthesis with the copolymer synthesised in water alone forming smaller dispersions ($D_h = 49$ nm) than those synthesis in water/EtOH mixtures ($D_h = 67$ – 140 nm). It should be noted that this size dependence is the reverse of that seen for the sizes of nanogels synthesised in different cononsolvent compositions. However, the diblocks were assembled in water alone and from solution which will result in different structures to those self-assembled *in situ* in varying solvent mixtures and the presence of crosslinker. As noted above, NMR spectroscopy showed very little difference between copolymers so such large differences in the self-assembly were unexpected. However, no suitable conditions for SEC analysis could be found for these diblock copolymers, therefore differences in architecture, M_w/M_n or homopolymer contamination cannot be completely eliminated. Similar trends in both nano-object size and were seen for pAA₃₄-*b*-pNIPAM₉₂ and pAA₃₄-*b*-pNIPAM₂₈₀ diblock copolymers series (Table S3†).

It is well known that solvents and additives (*e.g.* Lewis acids) used during synthesis can affect the microstructure (tacticity) of polymers.⁹¹ Biswas *et al.* reported that pNIPAM synthesised in different water/MeOH mixtures with Yb(OTf)₃ have different tacticities and that the cloud point was dependent on the tacticity of the pNIPAM, with pNIPAM with higher isotactic content having a lower cloud point.⁸⁹ Further inspection of the ¹H NMR spectra (Fig. 4) in the 1.2–1.8 ppm region resulting from the pNIPAM backbone methylene group showed small but distinct differences in the tacticity of the pNIPAM block with an increase in syndiotactic (racemic diad) from approximately 53% to 59% as X_{EtOH} is increased from 0 to 0.15.

The shift in the T_{cp} was previously attributed to the different content of *meso* diads, where diblock copolymers syn-



thesised in water were more *meso*-rich (*i.e.* less hydrophilic) than those synthesised in cononsolvents, hence these have a lower T_{cp} .⁹² An increase in racemic diads generates an alternating chirality where geometrical constraints make the formation of local hydrogen bonds between amide groups difficult and hence water molecules interact more strongly with the polymer, thus explaining the enhanced hydrophilicity and higher T_{cp} .

If the polymerisation solvent is causing the change in tacticity of pNIPAM, as has been reported in literature, then the homopolymers synthesised in different water/EtOH mixtures would also be expected to have differences in the tacticity. However, ¹H NMR spectroscopy showed no change in the 1.2–1.8 ppm region of these polymers (Fig. S15†). Changes in tacticity of poly(methyl methacrylate) have been reported in heterogeneous polymerisations compared to bulk polymerisation.^{93,94} In these cases, the volume of the polymer chain is comparable to the volume of the polymerisation loci so the increased syndiotacticity has been attributed to the confinement of the chains requiring more *gauche* conformations along the polymer backbone to enable it to fold back into the particle/loci. This restriction on the chain conformation may lead to a preferred conformation at the propagating centre and therefore the formation of different triads.

Although the particles here are unlikely to be single chain, their small volume is still likely to lead to the chains being conformationally restricted. Therefore, we suggest that the change in tacticity is due to the PITSA process with the constraints placed upon the propagating centre and accessible monomer resulting in a preference for syndiotacticity. The differences between polymers synthesised in different water/EtOH mixtures can be explained by the critical degrees of polymerisation required for nucleation. As X_{EtOH} is increased, the critical degree of polymerisation is reduced, resulting in earlier nucleation and the polymer chain growing under constrained conditions for longer, increasing the length of chain with higher syndiotacticity and the overall average syndiotacticity of the sample.

Materials and methods

Materials

2-Mercaptoethanol ($\geq 99\%$), carbon disulfide ($\geq 99\%$), (trimethylsilyl)diazomethane (2 M solution in diethyl ether) and 4,4'-azobis(4-cyanovaleric acid) (ACVA, 98%) were purchased from Sigma-Aldrich. Potassium phosphate tribasic (97%) was purchased from Alfa Aesar. 2-Bromoisobutyric acid (98%) was purchased from Acros Organics. Acrylic acid (AA, Merck, $\geq 99\%$) was passed through a column of basic Al_2O_3 prior to use. *N*-Isopropyl acrylamide (NIPAM, Fluorochem) and 2,2'-azobis(2-methylpropionitrile) (AIBN) (Sigma-Aldrich) were recrystallised from *n*-hexane and dried before use. *N,N'*-Methylene bisacrylamide (BIS) (Sigma-Aldrich) was recrystallised from hot methanol and dried before use. 2-(Hydroxyethylthiocarbonothioylthio)-2-methylpropanoic acid

(HEMP) was synthesised as previous reported.⁷⁹ All other reagents were used as received. SnakeSkin™ dialysis tubing (ThermoFisher scientific) of 3.5 kDa molecular weight cut-off (MWCO) was used to purify the macro-CTAs and nanogel solutions. For nuclear magnetic resonance (NMR) experiments, samples were prepared in either D_2O or $CDCl_3$ (Sigma Aldrich).

Synthesis of poly(acrylic acid) macroCTA via ethanolic RAFT solution polymerisation

Poly(acrylic acid) macroCTAs with different DP_n were synthesised similarly but targeting different molar ratios of [AA]:[HEMP]. Using the pAA₄₅ synthesis as an example: A mixture of HEMP (0.56 g, 2.31 mmol), AA (10 g, 138.78 mmol), AIBN (0.076 g, 0.46 mmol) and ethanol to give a 25% w/w solids solution was purged thoroughly with N_2 for 30 min. The flask was then placed onto a DrySyn® heating block preheated to 70 °C and left to react for 130 min. The reaction was quenched by removing the flask from the heat source and opening to air. AA conversion (78%) was calculated by ¹H NMR spectroscopy. The product was recovered by precipitation from diethyl ether (400 mL). Further product purification was performed by dialysis against water and freeze-drying to give a pale yellow solid. δH (400 MHz; D_2O , 25 °C) (ppm): 3.83 (2H, t, CH_2OH), 3.60 (2H, t, CH_2SC), 2.39 (1H, br. s, $-CH-$), 1.93–1.60 (2H, br. t, $-CH_2-$); δC (100 MHz; D_2O , 25 °C): 34.3 ($-CH_2-$), 41.5 ($-CH-$), 178.9 ($C(O)$); $\bar{\nu}_{max}$ (ATR) cm^{-1} : 2935 (br. s, R-COOH), 1701 (vs, $C=O$), 1449 (w, $-CH_2-$), 1411 (w, R- CH_2-S), 1214 (m, $-COOH$), 1162 (s, $C=S$), 794 (m, $-C-(CH_3)_2$). M_n , theor = 3987 $g\ mol^{-1}$; M_n , SEC = 4.7 $kg\ mol^{-1}$, M_w/M_n = 1.19.

Synthesis of poly(acrylic acid)-*b*-poly(*N*-isopropylacrylamide-*st*-*N,N'*-methylene bisacrylamide) nanogels via RAFT dispersion polymerisation in cononsolvents

RAFT dispersion polymerisation of NIPAM and BIS was performed in either water or different water–ethanol (H_2O :EtOH) mole fractions at 10% w/w solids content. The mole ratio of [NIPAM]/[pAA₄₅] was adjusted between 87 to 262 to give a range of nanogels with different sizes. An example of nanogel synthesis targeting a pAA₄₅-*b*-pNIPAM₁₇₅/BIS₃ in X_{EtOH} = 0.06 was as follows: pAA₄₅ (0.0352 g, 0.010 mmol), NIPAM (0.2003 g, 1.770 mmol), BIS (0.0041 g, 0.027 mmol), were dissolved in a mixture of H_2O :EtOH (1.8724 g of H_2O and 0.2112 g of ethanol). The pH of the solution was adjusted to pH 6.68 using an aqueous solution of NaOH (0.07 mL, 0.35 mmol). The mixture was purged with N_2 for 30 minutes, followed by the addition of the degassed ACVA solution in ethanol (0.124 mL, 14.27 mM) *via* microsyringe. All the quantities of solids and volume of solutions added were calculated in advance to give a final concentration of 10% w/w solids before reaction. The mixture was then placed into an oil bath previously set at 70 °C and left to react for 18 h. Total monomer conversion (98%) was estimated by moisture analysis. The product dispersion was purified by dialysis against DI water and was later stored at 4 °C. $\bar{\nu}_{max}$ (ATR-FTIR) cm^{-1} : 3290 (br. m, $-CONH$), 2973 (m, R-COOH), 2934 and 2873 (m,



$-\text{CH}_2$), 1640 and 1539 (s, $-\text{CONR}_2$), 1457 (m, $-\text{CH}_2-$), 1409 (w, $-\text{COOH}$), 1386 and 1367 (m, $-\text{C}(\text{CH}_3)_2$), 1172 (m, $\text{C}=\text{S}$), 1130 (m, $-\text{C}(\text{CH}_3)_2$), 879 (w, $\text{S}=\text{C}(\text{S})\text{S}$).

Synthesis of poly(acrylic acid)-*b*-poly(*N*-isopropylacrylamide) copolymers in cononsolvents

A pAA₃₄ macroCTA was chain extended with NIPAM at different [NIPAM]/[pAA₃₄] mole ratios in either water or water-ethanol (H₂O : EtOH) cononsolvents mixes at a 10% w/w solids content. An example of a diblock copolymer synthesis targeting a mole ratio of [NIPAM] : [pAA₃₄] of 184 in $X_{\text{EtOH}} = 0.06$ was as follows: the pAA₃₄ macroCTA (0.0258 g, 0.0096 mmol), and NIPAM monomer (0.2001 g, 1.77 mmol) were dissolved in a mixture of H₂O : EtOH (1.7542 g of H₂O and 0.1983 g of ethanol). The pH of the solution was then adjusted to pH 6.73 using an aqueous solution of NaOH (0.05 mL, 0.25 mmol). All the quantities of solids and volume of solutions added were calculated in advance to give a final concentration of 10% w/w solids before reaction. The solution was then degassed under nitrogen for 30 minutes, followed by the addition of the degassed ACVA solution in ethanol (0.14 mL, 14.27 mM) *via* microsyringe. The solution was immersed into an oil bath preheated to 70 °C and left to react for 18 hours. NIPAM monomer conversion (99%) was estimated by ¹H NMR spectroscopy. The crude product was purified by dialysis against DI water and freeze-dried to yield a white solid. $M_{n,\text{theo}} = 28\,644\text{ g mol}^{-1}$; δH (400 MHz, D₂O, 25 °C): 3.85 (br. d, H, $-\text{NCH}$), 2.25–1.96 (br. d, 1H, $-\text{CH}-$), 1.53 (br. t, 2H, $-\text{CH}_2-$), 1.09 (br. s, 6H, $-\text{CH}(\text{CH}_3)_2$); δC (100 MHz; D₂O, 25 °C): 21.5 ($-\text{CH}_2-$), 41.7 ($-\text{CH}-$), 175.2 (C(O)); $\tilde{\nu}_{\text{max}}$ (ATR) cm^{-1} : 3283 (br. m, $-\text{CONH}$), 2973 (m, R-COOH), 2929 and 2876 (w, $-\text{CH}_2-$), 1634 and 1538 (s, $-\text{CON-R}_2$), 1459 (m, $-\text{CH}_2-$), 1387 and 1367 (m, $-\text{C}(\text{CH}_3)_2$), 1172 (w, $\text{C}=\text{S}$).

NMR spectroscopy

Nuclear Magnetic Resonance (NMR) spectra were recorded at 25 °C on a 400 MHz Bruker Avance III HD spectrometer. Each collected spectrum was calibrated using the residual solvent peak as reference. ¹H NMR spectra were collected over 64 scans averaged per spectrum. Spectra were analysed with Bruker Topspin 3.0.

Size exclusion chromatography (SEC)

Molar masses and dispersities of pAA were determined by SEC was determined after esterification of the carboxylic groups. pAA samples were dissolved in THF/methanol followed by the dropwise addition of trimethylsilyldiazomethane. Addition of the methylation agent ended when the production of N₂ stopped, and the yellow colour remain unchanged. The solutions were left to stir allowing the solvents to evaporate overnight. Esterified samples were dissolved in the SEC eluent. Samples were analysed on an Agilent PL-GPC 50 system fitted with 2 × PLgel Mixed-C 5 μm (300 × 7.5 mm) columns. THF containing 4%v/v acetic acid and 0.025%w/v BHT was used as eluent at a flow rate of 1 mL min⁻¹ at 25 °C. The equipment was calibrated with near-monodisperse poly(methyl methacry-

late) standards (molecular weight range between 5.45×10^2 – $2.00 \times 10^6\text{ g mol}^{-1}$). Analyte samples were prepared at a concentration of 2 mg mL⁻¹ in THF (HPLC, 4%v/v acetic acid, 0.025%w/v BHT) and filtered through a 0.45 μm polytetrafluoroethylene (PTFE) filter before injection (100 μL). Molar masses and dispersities were calculated with Agilent GPC Software.

Fourier transform infrared spectroscopy (ATR-FTIR)

ATR-FTIR was used to assess the chemical composition of materials. IR spectra were recorded on a PerkinElmer Spectrum™ 100 FT-IR spectrometer using a universal diamond ATR (UATR) accessory. Data was collected over 5 scans in the 450 to 4000 cm⁻¹ region. The IR data was recorded and extracted from a Bruker software to further processing in Excel.

Gravimetric analysis

The monomer conversion for the synthesis of nanogels was estimated gravimetrically using a KERN DAB 100-3 electronic moisture analyser. Samples were weighed and heated up to 190 °C to determine their solid content.

Dynamic light scattering (DLS)

Determination of the hydrodynamic diameter of the purified nanogels (0.1% w/w) was obtained using a NanoBrook Omni particle analyser at a scattering angle of 173° using a 35 mW diode laser. Each DLS measurement consisted on 5 runs of 3 minutes at count rates between 440–550 kcps. Temperature dependent DLS experiments were performed from low to high temperatures with increments of 2.5 °C with 8 minutes in-between intervals to allow thermal stabilization. The pH of the solutions was manually adjusted by addition of NaOH or HCl solutions using a Hach H160 ISFET probe. The method of the constrained regularization method for inverting data (CONTIN) was used to obtain the size distribution from the autocorrelation function.

Cloud point measurements by UV-vis spectrometry

The absorbance of the copolymer aqueous solutions (1% w/w) at 550 nm as a function of temperature was recorded on a Cary 300 Bio UV-vis spectrometer fitted with a Peltier temperature controlled multi-cell block. The temperature was increased from 10 to 60 °C at a heating rate of 0.12 °C min⁻¹. The cloud points were calculated from the inflection point of each absorbance curve.

Aqueous electrophoresis

ζ-Potential measurements for the aqueous nanogel dispersions (0.1% w/w) containing background KCl (1 mM) were determined on a Malvern Zetasizer Nano-ZS instrument. The Smoluchowski relationship was used to determine the zeta potential from the electrophoretic mobility.



Transmission electron microscopy (TEM)

TEM images were collected using a Phillips CM100 microscope adapted with a Gatan CCD camera. Nanogel solutions (0.1% w/w, 10 μ L) were placed onto freshly glow discharged carbon-coated grids for 1 min and then blotted with filter paper. Phosphotungstic acid (0.75% w/w, 5 μ L) stain was then placed onto the sample and left for 20 s to finally blot it again with filter paper to remove the excess of stain. The grid was then dried with an adapted vacuum hose.

Conclusions

A series of responsive nanogels were prepared by RAFT-mediated PITSA of NIPAM and BIS from a PAA macroCTA in water and water/EtOH mixtures to investigate the effects of the synthesis cononsolvent composition on the properties of pNIPAM nanogels. Characterisation of the crosslinked particles *via* DLS and TEM showed that the size of the nanogels synthesised in water increased as the DP_n of the pNIPAM block was increased whereas for a fixed pNIPAM DP_n particle size was seen to decrease with increasing EtOH content. The latter was attributed to the fact that solubility of the growing pNIPAM is reduced in the reaction medium resulting in earlier particle nucleation. Furthermore, the synthesis cononsolvent composition was found to affect the thermal properties of the final nanogels. For nanogels with a fixed pNIPAM DP_n, the VPTT was found to increase to higher temperatures with increasing EtOH mole fraction in water used during synthesis. Detailed characterisation of analogue pAA-*b*-pNIPAM diblock copolymers with a fixed pNIPAM DP_n showed that the copolymers had differences in the tacticity according to the synthesis solvent composition. The copolymers were found to have higher syndiotacticity and increased cloud point temperatures in accordance with the increasing EtOH mole fraction used in their synthesis. These differences were rationalised to the PITSA process, where an earlier nucleation caused by changes in the solubility of growing pNIPAM can lead to constrained conditions resulting in increasing the average syndiotacticity of the copolymer. Our study provides new insights into the effects of using mixtures of solvents on the properties of pNIPAM nanogels prepared by a RAFT-mediated PITSA approach. This is relevant for researchers that use a cononsolvency strategy to incorporate functionalities to pNIPAM nanogels using a RAFT dispersion polymerisation. It is noteworthy that the properties of the nanogels prepared in this study were evaluated at a fixed dispersion pH and since these are composed of PAA their conformation may be sensitive to changes in pH and ionic strength. However, this will be addressed in a future publication.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors wish to thank the Consejo Nacional de Ciencia y Tecnología (CONACYT scholarship 461300, PhD studentship for MMM), the University of Sheffield and the UK Engineering and Physical Sciences Research Council [EPSRC grant EP/P027989/1] for funding. Dr Svet Tzokov is thanked for his assistance with TEM.

Notes and references

- 1 R. T. Chacko, J. Ventura, J. Zhuang and S. Thayumanavan, *Adv. Drug Delivery Rev.*, 2012, **64**, 836–851.
- 2 D. Li, C. F. van Nostrum, E. Mastrobattista, T. Vermonden and W. E. Hennink, *J. Controlled Release*, 2017, **259**, 16–28.
- 3 P. Manickam, M. Pierre, R. D. Jayant, M. Nair and S. Bhansali, in *Nanogels for Biomedical Applications*, The Royal Society of Chemistry, 2018, vol. 30, ch. 13, pp. 261–282.
- 4 S. Maya, B. Sarmento, A. Nair, N. S. Rejinold, S. V. Nair and R. Jayakumar, *Curr. Pharm. Des.*, 2013, **19**, 7203–7218.
- 5 A. Sivashanmugam, R. A. Kumar, M. V. Priya, S. V. Nair and R. Jayakumar, *Eur. Polym. J.*, 2015, **72**, 543–565.
- 6 K. Akiyoshi, S. Deguchi, N. Moriguchi, S. Yamaguchi and J. Sunamoto, *Macromolecules*, 1993, **26**, 3062–3068.
- 7 E. Akiyama, N. Morimoto, P. Kujawa, Y. Ozawa, F. M. Winnik and K. Akiyoshi, *Biomacromolecules*, 2007, **8**, 2366–2373.
- 8 S. A. Ferreira, P. J. G. Coutinho and F. M. Gama, *Langmuir*, 2010, **26**, 11413–11420.
- 9 M. Takeo, T. Mori, T. Niidome, S. Sawada, K. Akiyoshi and Y. Katayama, *J. Colloid Interface Sci.*, 2013, **390**, 78–84.
- 10 D. Y. Ko, H. J. Moon and B. Jeong, *J. Mater. Chem. B*, 2015, **3**, 3525–3530.
- 11 J. P. Quiñones, H. Peniche and C. Peniche, *Polymers*, 2018, **10**, 235.
- 12 J.-H. Ryu, R. T. Chacko, S. Jiwpanich, S. Bickerton, R. P. Babu and S. Thayumanavan, *J. Am. Chem. Soc.*, 2010, **132**, 17227–17235.
- 13 D. C. González-Toro, J.-H. Ryu, R. T. Chacko, J. Zhuang and S. Thayumanavan, *J. Am. Chem. Soc.*, 2012, **134**, 6964–6967.
- 14 L. Li, K. Raghupathi, C. Yuan and S. Thayumanavan, *Chem. Sci.*, 2013, **4**, 3654–3660.
- 15 Z. Jiang, W. Cui, J. Mager and S. Thayumanavan, *Ind. Eng. Chem. Res.*, 2019, **58**, 6982–6991.
- 16 X. T. Cao, A. M. Showkat and K. T. Lim, *Eur. Polym. J.*, 2015, **68**, 267–277.
- 17 B. Aktan, L. Chambre, R. Sanyal and A. Sanyal, *Biomacromolecules*, 2017, **18**, 490–497.
- 18 J. Zhuang, S. Jiwpanich, V. D. Deepak and S. Thayumanavan, *ACS Macro Lett.*, 2012, **1**, 175–179.
- 19 C. W. Park, H.-M. Yang, H. J. Lee and J.-D. Kim, *Soft Matter*, 2013, **9**, 1781–1788.
- 20 B. Liu and S. Thayumanavan, *J. Am. Chem. Soc.*, 2017, **139**, 2306–2317.



- 21 W. I. Choi, G. Tae and Y. H. Kim, *J. Mater. Chem.*, 2008, **18**, 2769–2774.
- 22 J. He, X. Tong and Y. Zhao, *Macromolecules*, 2009, **42**, 4845–4852.
- 23 Y.-C. Wang, J. Wu, Y. Li, J.-Z. Du, Y.-Y. Yuan and J. Wang, *Chem. Commun.*, 2010, **46**, 3520–3522.
- 24 W. Chen, Y. Hou, Z. Tu, L. Gao and R. Haag, *J. Controlled Release*, 2017, **259**, 160–167.
- 25 D. Lu, M. Zhu, S. Wu, W. Wang, Q. Lian and B. R. Saunders, *Polym. Chem.*, 2019, **10**, 2516–2526.
- 26 D. Steinhilber, M. Witting, X. Zhang, M. Staegemann, F. Paulus, W. Friess, S. Küchler and R. Haag, *J. Controlled Release*, 2013, **169**, 289–295.
- 27 M. Giubudagian, M. Asadian-Birjand, D. Steinhilber, K. Achazi, M. Molina and M. Calderón, *Polym. Chem.*, 2014, **5**, 6909–6913.
- 28 L. I. Vossen, S. Wedepohl and M. Calderón, *Polymers*, 2018, **10**, 398.
- 29 A. Oehrl, S. Schötz and R. Haag, *Colloid Polym. Sci.*, 2020, **298**, 719–733.
- 30 K. McAllister, P. Sazani, M. Adam, M. J. Cho, M. Rubinstein, R. J. Samulski and J. M. DeSimone, *J. Am. Chem. Soc.*, 2002, **124**, 15198–15207.
- 31 D. J. Bharali, S. K. Sahoo, S. Mozumdar and A. Maitra, *J. Colloid Interface Sci.*, 2003, **258**, 415–423.
- 32 J. K. Oh, D. J. Siegwart, H.-i. Lee, G. Sherwood, L. Peteanu, J. O. Hollinger, K. Kataoka and K. Matyjaszewski, *J. Am. Chem. Soc.*, 2007, **129**, 5939–5945.
- 33 P. B. Zetterlund, N. Alam and M. Okubo, *Polymer*, 2009, **50**, 5661–5667.
- 34 P. Bhardwaj, V. Singh, S. Aggarwal and U. K. Mandal, *J. Macromol. Sci., Part A: Pure Appl. Chem.*, 2009, **46**, 1083–1094.
- 35 W. Li, K. Matyjaszewski, K. Albrecht and M. Möller, *Macromolecules*, 2009, **42**, 8228–8233.
- 36 S. F. Medeiros, A. M. Santos, H. Fessi and A. Elaissari, *J. Polym. Sci., Part A: Polym. Chem.*, 2010, **48**, 3932–3941.
- 37 R. W. Graff, Y. Shi, X. Wang and H. Gao, *Macromol. Rapid Commun.*, 2015, **36**, 2076–2082.
- 38 D. S. Spencer, B. C. Luu, D. W. Beckman and N. A. Peppas, *J. Polym. Sci., Part A: Polym. Chem.*, 2018, **56**, 1536–1544.
- 39 X. Li, J. Zuo, Y. Guo and X. Yuan, *Macromolecules*, 2004, **37**, 10042–10046.
- 40 J. Moselhy, T. Vira, F. F. Liu and X. Y. Wu, *Int. J. Nanomed.*, 2009, **4**, 153–164.
- 41 D. Pamfil and C. Vasile, in *Polymer Gels: Perspectives and Applications*, ed. V. K. Thakur, M. K. Thakur and S. I. Voicu, Springer Singapore, Singapore, 2018, pp. 71–110, DOI: 10.1007/978-981-10-6080-9_4.
- 42 G. Delaittre, M. Save and B. Charleux, *Macromol. Rapid Commun.*, 2007, **28**, 1528–1533.
- 43 Z. An, Q. Shi, W. Tang, C.-K. Tsung, C. J. Hawker and G. D. Stucky, *J. Am. Chem. Soc.*, 2007, **129**, 14493–14499.
- 44 W.-M. Wan, C.-Y. Hong and C.-Y. Pan, *Chem. Commun.*, 2009, 5883–5885.
- 45 G. Delaittre, C. Dire, J. Rieger, J.-L. Putaux and B. Charleux, *Chem. Commun.*, 2009, 2887–2889.
- 46 B. Charleux, G. Delaittre, J. Rieger and F. D'Agosto, *Macromolecules*, 2012, **45**, 6753–6765.
- 47 C. A. Figg, A. Simula, K. A. Gebre, B. S. Tucker, D. M. Haddleton and B. S. Sumerlin, *Chem. Sci.*, 2015, **6**, 1230–1236.
- 48 X.-b. Liu, J.-f. Zhou and X.-d. Ye, *Chin. J. Chem. Phys.*, 2012, **25**, 463–468.
- 49 L. Hou, K. Ma, Z. An and P. Wu, *Macromolecules*, 2014, **47**, 1144–1154.
- 50 R. Rajan and K. Matsumura, *Macromol. Rapid Commun.*, 2017, **38**, 1700478.
- 51 J. Rieger, C. Grazon, B. Charleux, D. Alaimo and C. Jérôme, *J. Polym. Sci., Part A: Polym. Chem.*, 2009, **47**, 2373–2390.
- 52 C. Grazon, J. Rieger, N. Sanson and B. Charleux, *Soft Matter*, 2011, **7**, 3482–3490.
- 53 W. Shen, Y. Chang, G. Liu, H. Wang, A. Cao and Z. An, *Macromolecules*, 2011, **44**, 2524–2530.
- 54 G. Liu, Q. Qiu, W. Shen and Z. An, *Macromolecules*, 2011, **44**, 5237–5245.
- 55 L. Yan and W. Tao, *Polymer*, 2010, **51**, 2161–2167.
- 56 Y. Xu, Y. Li, X. Cao, Q. Chen and Z. An, *Polym. Chem.*, 2014, **5**, 6244–6255.
- 57 W. Fu, C. Luo, E. A. Morin, W. He, Z. Li and B. Zhao, *ACS Macro Lett.*, 2017, **6**, 127–133.
- 58 Y. Li, Z. Ye, L. Shen, Y. Xu, A. Zhu, P. Wu and Z. An, *Macromolecules*, 2016, **49**, 3038–3048.
- 59 K. Ma, Y. Xu and Z. An, *Macromol. Rapid Commun.*, 2015, **36**, 566–570.
- 60 N. Bhuchar, R. Sunasee, K. Ishihara, T. Thundat and R. Narain, *Bioconjugate Chem.*, 2012, **23**, 75–83.
- 61 Y. Kotsuchibashi and R. Narain, *Polym. Chem.*, 2014, **5**, 3061–3070.
- 62 R. Narain, Y. Wang, M. Ahmed, B. F. L. Lai and J. N. Kizhakkedathu, *Biomacromolecules*, 2015, **16**, 2990–2997.
- 63 H. M. Crowther and B. Vincent, *Colloid Polym. Sci.*, 1998, **276**, 46–51.
- 64 M. Heskins and J. E. Guillet, *J. Macromol. Sci., Part A: Pure Appl. Chem.*, 1968, **2**, 1441–1455.
- 65 H. G. Schild and D. A. Tirrell, *J. Phys. Chem.*, 1990, **94**, 4352–4356.
- 66 T. López-León, D. Bastos-González, J. L. Ortega-Vinuesa and A. Elaissari, *ChemPhysChem*, 2010, **11**, 188–194.
- 67 M. J. A. Hore, B. Hammouda, Y. Li and H. Cheng, *Macromolecules*, 2013, **46**, 7894–7901.
- 68 H. G. Schild, M. Muthukumar and D. A. Tirrell, *Macromolecules*, 1991, **24**, 948–952.
- 69 G. Zhang and C. Wu, *J. Am. Chem. Soc.*, 2001, **123**, 1376–1380.
- 70 C. Scherzinger, P. Lindner, M. Keerl and W. Richtering, *Macromolecules*, 2010, **43**, 6829–6833.
- 71 S. Zhou, S. Fan, S. C. F. Au-yeung and C. Wu, *Polymer*, 1995, **36**, 1341–1346.



- 72 J. Hao, H. Cheng, P. Butler, L. Zhang and C. C. Han, *J. Chem. Phys.*, 2010, **132**, 154902.
- 73 H. Yamauchi and Y. Maeda, *J. Phys. Chem. B*, 2007, **111**, 12964–12968.
- 74 J. Wang, N. Wang, B. Liu, J. Bai, P. Gong, G. Ru and J. Feng, *Phys. Chem. Chem. Phys.*, 2017, **19**, 30097–30106.
- 75 H. A. Pérez-Ramírez, C. Haro-Pérez, E. Vázquez-Contreras, J. Klapp, G. Bautista-Carbajal and G. Odriozola, *Phys. Chem. Chem. Phys.*, 2019, **21**, 5106–5116.
- 76 E. R. Jones, M. Semsarilar, P. Wyman, M. Boerakker and S. P. Armes, *Polym. Chem.*, 2016, **7**, 851–859.
- 77 N. J. Warren and S. P. Armes, *J. Am. Chem. Soc.*, 2014, **136**, 10174–10185.
- 78 M. Williams, N. J. W. Penfold, J. R. Lovett, N. J. Warren, C. W. I. Douglas, N. Doroshenko, P. Verstraete, J. Smets and S. P. Armes, *Polym. Chem.*, 2016, **7**, 3864–3873.
- 79 E. Niezabitowska, A. R. Town, B. Sabagh, M. D. Morales-Moctezuma, V. R. Kearns, S. G. Spain, S. P. Rannard and T. O. McDonald, *Nanoscale Adv.*, 2020, **2**, 4713–4721.
- 80 R. O. R. Costa and R. F. S. Freitas, *Polymer*, 2002, **43**, 5879–5885.
- 81 S.-P. Wen, J. G. Saunders and L. A. Fielding, *Polym. Chem.*, 2020, **11**, 3416–3426.
- 82 J. Wyman, *J. Am. Chem. Soc.*, 1931, **53**, 3292–3301.
- 83 J. Brijitta, B. V. R. Tata and T. Kaliyappan, *J. Nanosci. Nanotechnol.*, 2009, **9**, 5323–5328.
- 84 M. B. Huglin, Y. Liu and J. Velada, *Polymer*, 1997, **38**, 5785–5791.
- 85 B. A. Humphreys, E. J. Wanless and G. B. Webber, *J. Colloid Interface Sci.*, 2018, **516**, 153–161.
- 86 A. Halperin, M. Kröger and F. M. Winnik, *Angew. Chem., Int. Ed.*, 2015, **54**, 15342–15367.
- 87 Y. Xia, N. A. D. Burke and H. D. H. Stöver, *Macromolecules*, 2006, **39**, 2275–2283.
- 88 Y. Katsumoto and N. Kubosaki, *Macromolecules*, 2008, **41**, 5955–5956.
- 89 C. S. Biswas, K. Mitra, S. Singh and B. Ray, *J. Chem. Sci.*, 2016, **128**, 415–420.
- 90 M. I. Gibson, D. Paripovic and H.-A. Klok, *Adv. Mater.*, 2010, **22**, 4721–4725.
- 91 Y. Isobe, D. Fujioka, S. Habaue and Y. Okamoto, *J. Am. Chem. Soc.*, 2001, **123**, 7180–7181.
- 92 T. Hirano, A. Ono, H. Yamamoto, T. Mori, Y. Maeda, M. Oshimura and K. Ute, *Polymer*, 2013, **54**, 5601–5608.
- 93 W. Jiang, W. Yang, X. Zeng and S. Fu, *J. Polym. Sci., Part A: Polym. Chem.*, 2004, **42**, 733–741.
- 94 S. C. Pilcher and W. T. Ford, *Macromolecules*, 1998, **31**, 3454–3460.

