



Cite this: *Org. Biomol. Chem.*, 2021, **19**, 6916

Received 22nd June 2021,
Accepted 16th July 2021

DOI: 10.1039/d1ob01202a

rsc.li/obc

Silicon compounds in carbon-11 radiochemistry: present use and future perspectives

Federico Luzi,  Antony D. Gee * and Salvatore Bongarzone *

Positron emission tomography (PET) is a powerful functional imaging technique that requires the use of positron emitting nuclides. Carbon-11 (^{11}C) radionuclide has several advantages related to the ubiquity of carbon atoms in biomolecules and the conservation of pharmacological properties of the molecule upon isotopic exchange of carbon-12 with carbon-11. However, due to the short half-life of ^{11}C (20.4 minutes) and the low scale with which it is produced by the cyclotron (sub-nanomolar concentrations), quick, robust and chemospecific radiolabelling strategies are required to minimise activity loss during incorporation of the ^{11}C nuclide into the final product. To address some of the constraints of working with ^{11}C , the use of silicon-based chemistry for ^{11}C -labelling was proposed as a rapid and effective route for radiopharmaceutical production due to the broad applicability and high efficiency showed in organic chemistry. In the past years several organic chemistry methodologies have been successfully applied to ^{11}C -chemistry. In this short review, we examine silicon-based ^{11}C -chemistry, with a particular emphasis on the radiotracers that have been successfully produced and potential improvements to further expand the applicability of silicon in radiochemistry.

1. Introduction

Positron emission tomography (PET) is a powerful functional imaging technique that allows the *in vivo* detection of normo- and patho-physiological changes in humans by using mole-

School of Biomedical Engineering and Imaging Sciences, 4th floor Lambeth Wing, St Thomas' Hospital, King's College London, London SE1 7EH, UK.
E-mail: antony.gee@kcl.ac.uk, salvatore.bongarzone@kcl.ac.uk



Federico Luzi

Federico Luzi is currently pursuing a PhD degree in Medical Imaging at King's College London under the supervision of Prof. Antony Gee. His research is focused on the development of novel methodologies for carbon-11 labelling and the application to neurodegeneration.

Federico previously obtained a Master's degree in Medicinal Chemistry and Pharmaceutical Technology from Alma Mater Studiorum, University of

Bologna (Italy) and a MRes degree in Medical Imaging from King's College, London.



Antony D. Gee

Antony Gee is Professor of PET and Radiochemistry in the School of Biomedical Engineering and Imaging Sciences at King's College London and President of the Society of Radiopharmaceutical Sciences. He obtained a BSc (Hons) in Chemistry at the University of Sussex (1985), and his PhD in Radiopharmaceutical Organic Chemistry at Uppsala University, Sweden (1991). Since then he has worked as the Director of PET Chemistry at the

Guy's and St Thomas' Hospitals Clinical PET centre, UMDS, London, the Aarhus University Hospital PET Centre in Aarhus Denmark, and SmithKlineBeecham/GlaxosmithKline, before returning to academia and current position at King's College in 2010.



cules radiolabelled with positron (β^+) emitting nuclides (radiotracers).¹ To achieve radiopharmaceutical targeting, radiotracers are often derived from biologically-active compounds with a known pharmacological profile, possessing high selectivity for a molecular target or physiological process.² The inclusion of a positron-emitting nuclide in the molecule of interest enables the *in vivo* visualization of the molecules biodistribution and kinetics metabolism.^{2,3} Of all the available PET nuclides, carbon-11 (^{11}C) is of particular interest due to the ubiquity of carbon atoms in biomolecules and because isotopic substitution of carbon-12 for carbon-11 preserves the biological properties of the non-radioactive isotope.^{2,4,5} However, due to the rapid radioactive decay of carbon-11 (radioactive half-life $t_{1/2} = 20.4$ minutes), the radio-synthesis, purification, formulation and quality control of carbon-11 radiopharmaceuticals must be accomplished in short times (the whole process should not exceed 60 minutes), hence quick and robust chemistry are needed to avoid substantial activity loss.^{2,6} The sub-nanomolar scale with which the radioisotope is produced from the cyclotron also represents a burden when performing ^{11}C -labelling, with the non-radioactive reactants being in large stoichiometric excess. Each minor impurity in the solvents and the reagents may generate side-products or degradation of reagents resulting in unwanted intermediates, so a high degree of chemoselectivity is required, as well.²

To meet criteria suitable for ^{11}C -labelling, silicon-containing compounds have received increased interest in the field. Organosilicon compounds have already demonstrated wide applicability in traditional organic chemistry enabling the development of well-known methodologies such as the Hiyama cross-coupling reactions for the formation of carbon-carbon bonds between silylated compounds and aryl halides,^{7,8} tosylates,⁹ mesylates,¹⁰ sulfinates¹¹ and phosphates¹² *via* palladium catalysis and fluoride or base activation.⁷⁻¹² Moreover, silyl compounds act as effective protecting groups due to the large number of functional groups

that can be protected (*e.g.* alcohols, alkynes, amines, carboxylic acids...) and the ease of the protecting/deprotecting steps.^{13,14} Another interesting application involves the use of hydrosilicon compounds, possessing one or more Si-H bonds, for the reduction of carbon dioxide (CO_2) to more reactive species.¹⁵⁻¹⁸ In the hydrosilylation reaction, CO_2 is used as a building block for the synthesis of a variety of functional groups such as formamides,¹⁵ methylamines,^{15,16} aldehydes¹⁷ and amins.¹⁸ Moreover, organosilicates have shown to be optimal substrates for the electrophilic fluorination of aryl and alkenyl substrates under mild conditions (*e.g.* room temperature, 18 hours) whilst having regio- and enantio-selectivity on the final product.¹⁹ Besides the large number of reactions available, silicon chemistry is cost-effective (the silylated reagents are easily synthesized or commercially available)²⁰ and eco-friendly (organosilicon compounds are ultimately catabolised into silica gel in the environment).^{7,8}

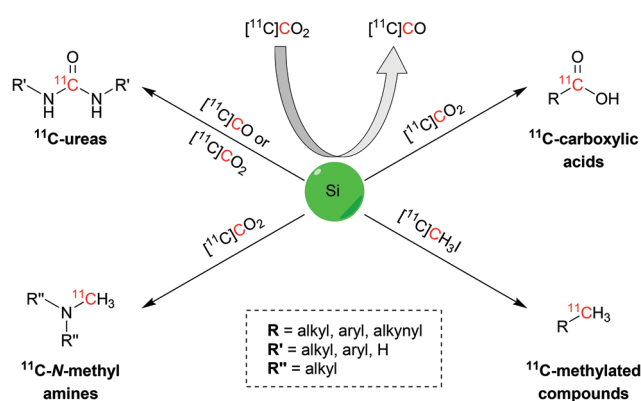
Given the high versatility and ease of handling of organosilane compounds, several methodologies have been successfully translated into carbon-11 and fluorine-18 chemistry in the past years. In particular, silicon-based compounds were applied in the production of a variety of fluorine-18 labelled small molecules and peptides as prosthetic groups, where the radionuclide was attached on *via* isotopic exchange (obtaining silicon-fluoride-acceptors – SiFAs),²¹ or as substrates for electrophilic fluorination.¹⁹ The application of silicon in fluorine-18 chemistry was recently reviewed by Bernard-Gauthier *et al.* and Tredwell *et al.*^{19,21}

Herein we report the latest advances in the ^{11}C -chemistry field based on silicon starting materials or reagents (Scheme 1). This review will initially disclose the atomic properties of silicon to then discuss the main uses of organosilane compounds in carbon-11 chemistry: (i) conversion from $[^{11}\text{C}]\text{CO}_2$ to $[^{11}\text{C}]\text{CO}$; (ii) trapping and activating agents for $[^{11}\text{C}]\text{CO}_2$ and (iii) precursors of the target radiopharmaceutical. This review will also report the biologically-active molecules that were successfully radiolabelled with the discussed reactions.



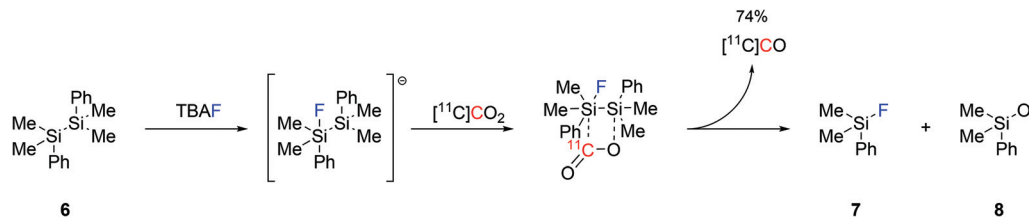
Salvatore Bongarzone

Dr. Bongarzone attained his PhD at the International School for Advanced Studies (SISSA, Trieste, Italy, 2007). Subsequent postdoctoral positions were held at the Institute for Research in Biomedicine, Barcelona (IRB) and the School of Biomedical Engineering and Imaging Sciences at King's College London (KCL). At KCL, Dr. Bongarzone conceived novel radiochemical reactions for developing PET imaging probes ($[^{11}\text{C}]\text{Niacin}$, $[^{11}\text{C}]\text{Biotin}$, $[^{18}\text{F}]\text{FAMTO}$, and $[^{11}\text{C}]\text{FPSZM1}$) and their preclinical characterization.



Scheme 1 Schematic representation of the ^{11}C functional groups and synthons obtainable *via* silicon chemistry.





Scheme 3 Proposed mechanism for the fluoride-promoted $[^{11}\text{C}]\text{CO}$ production from disilane species.⁴⁰

midate, the dopamine D2 selective antagonist $[^{11}\text{C}]\text{raclopride}$, $[^{11}\text{C}]\text{olaparib}$ for cancer imaging and the receptor for advanced glycation end-products (RAGE) radiotracer $[^{11}\text{C}]\text{FPS-ZM1}$ in radiochemical yields (RCYs) ranging between 29% and 91% (estimated by radioHPLC).^{37–39} Follow-up studies on the ^{11}C -silacarboxylate production of $[^{11}\text{C}]\text{CO}$ and subsequent ^{11}C -carbonylation showed that the whole process can be fully automated using a commercially available radiochemistry synthesis module.³⁸ With an initial delivery of 10 GBq of $[^{11}\text{C}]\text{CO}_2$, the radiotracer $[^{11}\text{C}]\text{FPS-ZM1}$ (Scheme 2) was achieved with a RCY of 34% (isolated, decay-corrected to end of $[^{11}\text{C}]\text{CO}_2$ delivery (EOD)) and molar activity (A_m) 28 GBq per μmol within 25 minutes from end of bombardment (EOB).³⁸

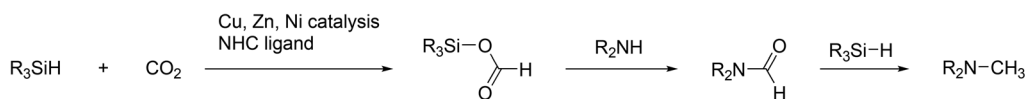
Another approach to produce $[^{11}\text{C}]\text{CO}$ was reported by reacting disilanes with $[^{11}\text{C}]\text{CO}_2$. Four different disilane species were tested for their ability of trapping $[^{11}\text{C}]\text{CO}_2$ and converting it into $[^{11}\text{C}]\text{CO}$, with 1,2-diphenyl-1,1,2,2-tetramethyldisilane ($(\text{Me}_2\text{PhSi})_2$, **6**, Scheme 3) being the most effective.⁴⁰ With the aid of catalytic amounts of TBAF (0.1 equiv.) as a fluoride source, the production of $[^{11}\text{C}]\text{CO}$ reached 74% using mild reaction conditions and within 3 minutes from EOB.⁴⁰ The reaction mechanism is believed to proceed by the initial formation of a pentavalent fluorosilyl anion which rapidly interacts with $[^{11}\text{C}]\text{CO}_2$ forming an unstable intermediate that spontaneously rearranges into a silyl fluoride derivative (**7**), a silyloxy derivative (**8**) and $[^{11}\text{C}]\text{CO}$ (Scheme 3). As a proof-of-concept, the applicability of the produced $[^{11}\text{C}]\text{CO}$ in ^{11}C -carbonylation reactions was tested *via* the radiosynthesis of

$[^{11}\text{C}]\text{benzylbenzamide}$ which was obtained with a good RCY (74%, estimated by radioHPLC).⁴⁰

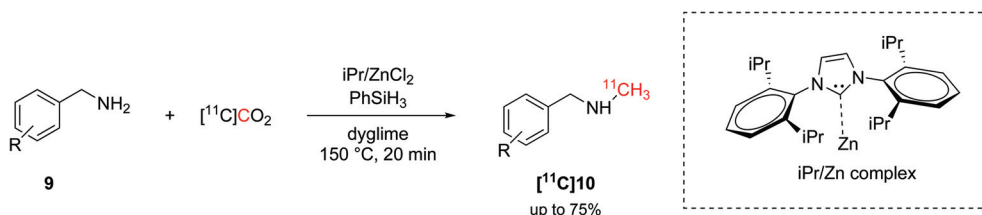
2.3. Reductive functionalisation of $[^{11}\text{C}]\text{CO}_2$ using organosilanes

The high reactivity that organosilanes have towards CO_2 enabled other interesting applications, such as the conversion of the cyclotron-produced $[^{11}\text{C}]\text{CO}_2$ to a ^{11}C -*N*-methyl group without the need of using $[^{11}\text{C}]\text{CH}_3\text{I}$. This approach was vastly exploited in synthetic chemistry to use CO_2 as a C_1 building block in hydrosilylation reactions of CO_2 .^{16,41,42} The gas is initially trapped by the organosilicon compound in the form of silyl formate (Scheme 4)⁴¹ with the aid of transition-metal catalysis (*e.g.* Cu, Zn, Ni) and bulky ligands (*e.g.* σ -donor *N*-heterocyclic carbenes, NHCs).^{16,41,42} Then, the carbonyl group reacts with the target amino compound to form formamides (Scheme 4).⁴² The presence of an excess of silane provokes the reduction of the formamide to *N*-methylamine (Scheme 4).¹⁶

The hydrosilylation of CO_2 was later translated into carbon-11 chemistry by Liger *et al.*⁴³ Similarly to the aforementioned non-radioactive reactions,¹⁶ this methodology exploited a NHC (1,3-bis(2,6-diisopropylphenyl)-1,3-dihydro-2*H*-imidazol-2-ylidene, *iPr*, Scheme 5) as ligand, zinc catalysis and high temperature (150 °C) to yield ^{11}C -*N*-methylamines within 20 minutes from EOD (Scheme 5). The radiolabelling of the amyloid- β plaque imaging agent $[^{11}\text{C}]\text{PiB}$ was achieved, as well, with an overall time of 50 minutes from EOD and in good

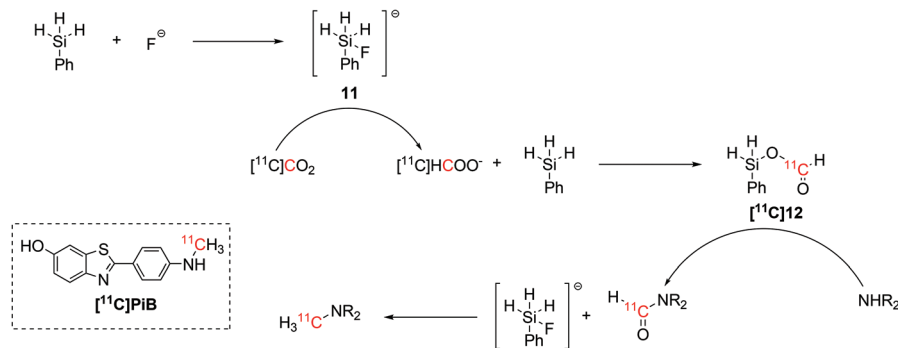


Scheme 4 Hydrosilylation of CO_2 forming a silyl formate. The reaction of an amine with a silyl formate produces a formamide which can be reduced by an excess of hydrosilane to yield an *N*-methylamine.¹⁶



Scheme 5 Hydrosilylation of $[^{11}\text{C}]\text{CO}_2$ and application on the ^{11}C -*N*-methylation.⁴³





Scheme 6 Fluoride-activated hydrosilylation of $[^{11}\text{C}]\text{CO}_2$ and application in the radiosynthesis of ^{11}C -*N*-methylamines, including the amyloid- β plaque imaging agent $[^{11}\text{C}]\text{PiB}$.⁴⁴

RCY (38%, isolated, decay-corrected) but low A_m (15 GBq per μmol).⁴³

A simpler set up that would not require the use of unstable NHCs was lately proposed for the synthesis of ^{11}C -*N*-methylamines.⁴⁴ Inspired by non-radioactive experiments,⁴⁵ the radiolabelling step required the use of only hydrosilane and TBAF as fluoride source.⁴⁴ The hydrosilane and the fluoride source were initially mixed in order to form an activated pentavalent fluorosilyl anion (**11**, Scheme 6) which instantly reduced the delivered $[^{11}\text{C}]\text{CO}_2$ to $[^{11}\text{C}]\text{formate}$, allowing the formation of a ^{11}C -silylformate ($[^{11}\text{C}]\text{12}$, Scheme 6). The ^{11}C -formyl group was then attached to the aminic precursor, yielding a ^{11}C -formamide that was reduced to ^{11}C -*N*-methylamine by the excess of fluorosilyl anion previously formed (Scheme 6).⁴⁴

This fluoride-activated radiosynthesis of ^{11}C -*N*-methylamines was also fully automated and exploited for the synthesis of $[^{11}\text{C}]\text{PiB}$ with RCY of 15% (isolated, decay-corrected) and A_m 61 GBq per μmol within 32 min from EOB.⁴⁴

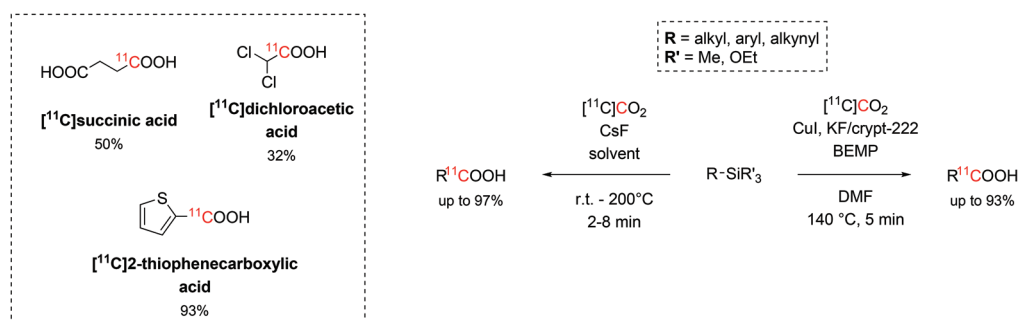
2.4. Organosilicon as precursors for carbon-11 radiolabelling

Recent findings revealed that organosilicon compounds are also viable radiolabelling precursors, especially in the form of trialkylsilyl and trialkoxysilyl arenes. The synthesis of these versatile organosilicon precursors is readily achieved *via* transition metal catalysis, such as Ni, Rh, Cu, allowing the silylation of a large number of aromatic compounds (*e.g.* Grignards,

amides, cyanides, esters and acyl fluorides) with a wide functional group tolerance.^{20,46–49}

Trialkylsilyl and trialkoxysilyl arenes showed high reactivity towards copper-catalysed desilylative carboxylation reactions.⁵⁰ With the aid of a fluoride source, the precursor was initially converted into a pentavalent silyl fluoride anion which then undergoes oxidative addition onto the Cu catalyst and react with the cyclotron-produced $[^{11}\text{C}]\text{CO}_2$. The highest reactivity was achieved when using DMF as solvent, KF/Kryptofix® 222 (crypt-222, 0.25 equiv.) as fluoride source, 2-tert-butylimino-2-diethylamino-1,3-dimethylperhydro-1,3,2-diazaphosphorine (BEMP, 0.6 equiv.) as CO_2 -trapping agent and a temperature of 140 °C for 5 minutes (Scheme 7). This method resulted equally efficient in the radiolabelling of alkynyl, aryl and heteroaryl precursors in short times (12 min) and with RCYs ranging between 19% and 93% (estimated by radioHPLC).⁵⁰

A similar methodology that did not require copper catalysis was developed, as well. When CsF (1 equiv.) was employed as fluoride source and the reaction proceeded for 2–8 minutes at temperatures between r.t. and 200 °C, a large number of alkynyl, aryl and heteroaryl precursors (26 examples, Scheme 7) were successfully ^{11}C -carboxylated with RCYs from 11% to 99%.⁵¹ A variety of solvents (DMF, DMA, DMSO/THF) were also tested returning similar trapping efficiency and reactivity. Using this strategy, eleven alkyl silyl precursors were successfully ^{11}C -carboxylated (RCYs = 19%–97%), as well, including the mitochondrial kinase inhibitor dichloroacetic acid



Scheme 7 The use of trialkylsilyl and trialkoxysilyl as precursors in the ^{11}C -carboxylation with $[^{11}\text{C}]\text{CO}_2$.

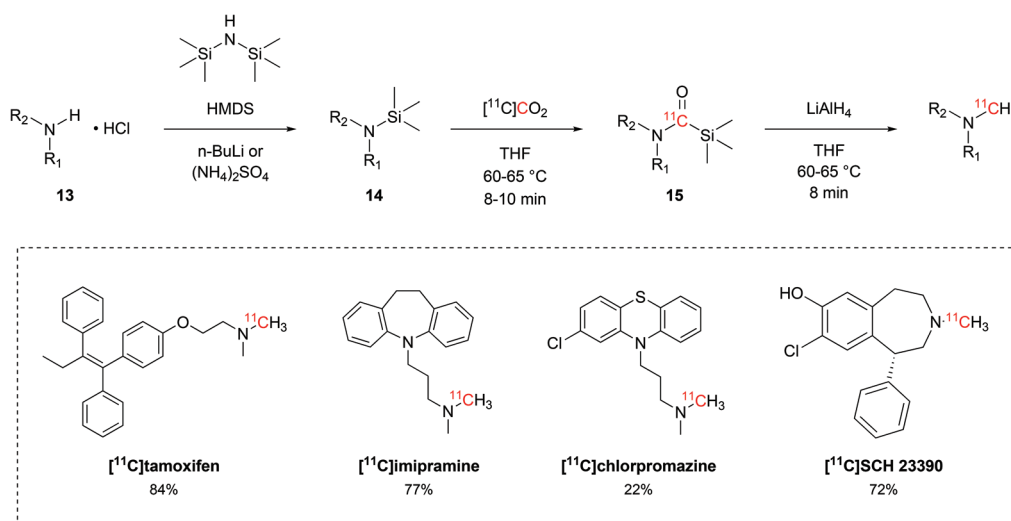


(^{11}C)dichloroacetic acid) and ^{11}C succinic acid (isolated decay-corrected RCYs = 32% and 50%, respectively, Scheme 7).⁵¹

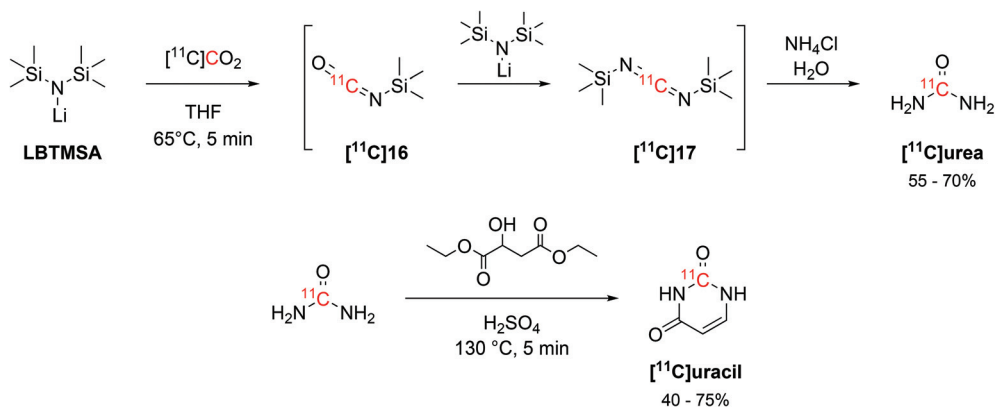
Besides the production of ^{11}C -carboxylic acids, the interaction between alkylsilyl precursors and ^{11}C CO₂ showed to effectively yield other moieties, such as ^{11}C -*N*-methylamines. Following a protocol that was initially developed with non-radioactive CO₂,⁵² Ram *et al.* established a reliable method for ^{11}C -*N*-methylation of secondary amines to methyl- ^{11}C -tertiary amines *via* direct use of ^{11}C CO₂ and silyl amine precursors. The hydrochloride salt of the amine precursor (**13**, Scheme 8) was initially treated with hexamethyldisilazane (HMDS, Scheme 8) in the presence of either *n*-butyl lithium or ammonium sulphate to yield the respective silyl amine (**14**, Scheme 8). Upon delivery, ^{11}C CO₂ would then be incorporated in the precursor as ^{11}C -silyl carbamate (**15**, Scheme 8) when reacted for 8–10 minutes at 60–65 °C. **15** is then reduced by lithium aluminium hydride to the desired tertiary ^{11}C -*N*-

methylamine with RCYs ranging between 22% and 84% (isolated, decay-corrected at EOB) and A_m of 1.5–15 GBq per μmol (calculated at EOB) within 36–50 minutes.^{52–56} This method was applied in the radiolabelling of a variety of tertiary alkyl amines, including the biologically-active molecules ^{11}C tamoxifen,⁵³ ^{11}C imipramine,⁵⁴ ^{11}C chlorpromazine⁵⁵ and ^{11}C SCH 23390 (Scheme 8).⁵⁶

The interaction of alkylsilyl precursors with ^{11}C CO₂ showed to be effective in the radiosynthesis of ^{11}C -ureas, as well.⁵⁷ The reaction uses lithium bis(trimethylsilyl)amide (LBTMSA, Scheme 9) as a precursor which reacts with ^{11}C CO₂ for 5 minutes at 65 °C to yield the corresponding ^{11}C -carbodiimide (^{11}C 17) species *via* a ^{11}C -isocyanate intermediate (^{11}C 16, Scheme 9). The hydrolysis of the ^{11}C -carbodiimide with an aqueous solution of ammonium chloride then produced ^{11}C urea (Scheme 9) with RCY of 55–70% (estimated by radioHPLC) in 16 minutes from EOB. The obtained ^{11}C urea was then employed in the synthesis of the nucleotide ^{11}C

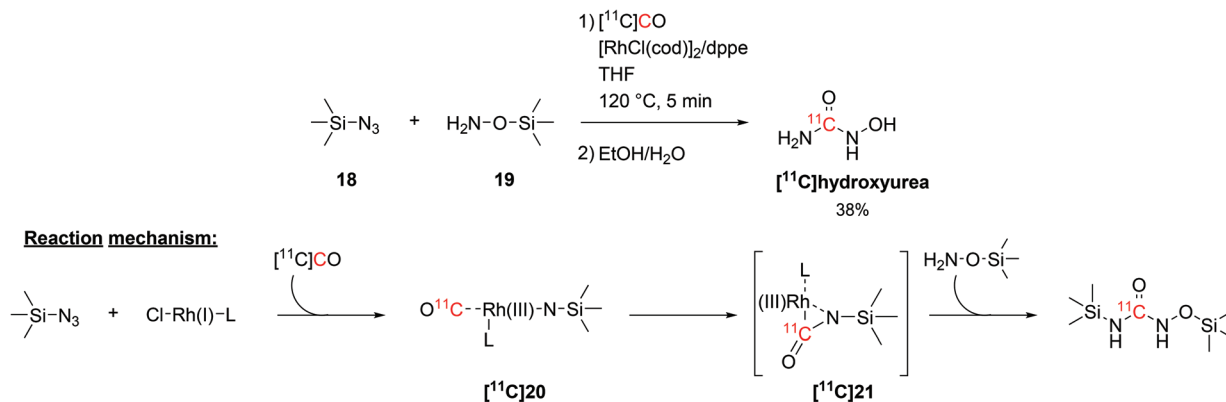


Scheme 8 ^{11}C -methylation of secondary amines from ^{11}C CO₂ and trimethylsilyl amines.^{52–56}



Scheme 9 Synthesis of ^{11}C urea *via* the coupling of a silylamine precursor and ^{11}C CO₂ and subsequent condensation with diethyl malate for ^{11}C uracil production.⁵⁷





Scheme 10 Coupling of $[^{11}\text{C}]\text{CO}$ with trimethylsilyl precursors to yield $[^{11}\text{C}]\text{hydroxyurea}$.⁵⁸

uracil by condensation with diethyl malate in presence of fuming sulphuric acid (Scheme 9) at 130 °C for 5 minutes. $[^{11}\text{C}]\text{Uracil}$ was obtained with RCY of 40–75% (estimated by radioHPLC).⁵⁷

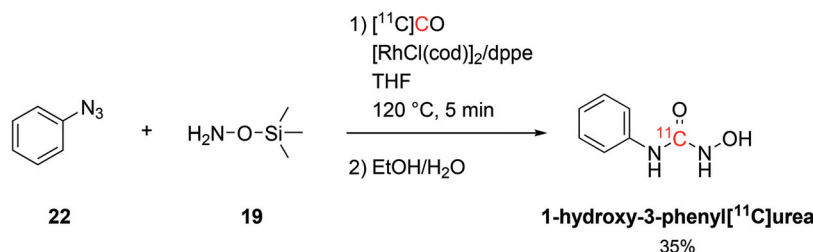
The radiolabelling of ^{11}C -ureas could also be achieved by coupling organosilicon compounds with $[^{11}\text{C}]\text{CO}$. This reaction requires the presence of a silylazide (**18**) and a (silyl) hydroxylamine (**19**, Scheme 10) and proceeds *via* transition metal catalysis in short times.⁵⁸ In particular, the reaction was initially developed using trimethylsilyl azide and *O*-(trimethylsilyl)hydroxylamine in THF and in the presence of chloro(1,5-cyclooctadiene)rhodium(i) dimer ($[\text{RhCl}(\text{cod})]_2$) and 1,2-bis(diphenylphosphino)ethane (dppe) as catalyst and ligand, respectively (Scheme 10). The reaction proceeded at a temperature of 120 °C for 5 minutes and the resulting $[^{11}\text{C}]\text{hydroxyurea}$ was obtained with a RCY of 38% (isolated, decay-corrected, based on delivered $[^{11}\text{C}]\text{CO}$).⁵⁸ The suggested reaction mechanism proceeds with the formation of a Rh(i)-bound nitrene intermediate ($[^{11}\text{C}]\text{20}$) which then reacts with the delivered $[^{11}\text{C}]\text{CO}$ to yield a Rh(III)-coordinated ^{11}C -isocyanate ($[^{11}\text{C}]\text{21}$, Scheme 10). Then, the hydroxylamine acting as nucleophile reacts with the ^{11}C -isocyanate forming a silyl-protected ^{11}C -urea (Scheme 10). The cleavage of the silyl groups was then easily achieved with a mixture of water and ethanol.⁵⁸

The synthesised $[^{11}\text{C}]\text{hydroxyurea}$ (Scheme 10), which in the body acts as a ribonucleoside reductase inhibitor, was then uti-

lised to study its pharmacokinetics across the blood brain barrier *in vivo* and explore the interaction with multidrug resistance transporters.⁵⁹ This was achieved by measuring the brain activity in rats with and without the simultaneous administration of multidrug resistance protein inhibitors (such as cyclosporin A and probenecid). The influx of $[^{11}\text{C}]\text{hydroxyurea}$ in the rat brain, however, was not significantly modified by the used intervention drugs, suggesting that hydroxyurea is not a substrate for active efflux transporters at the blood brain barrier.⁵⁹

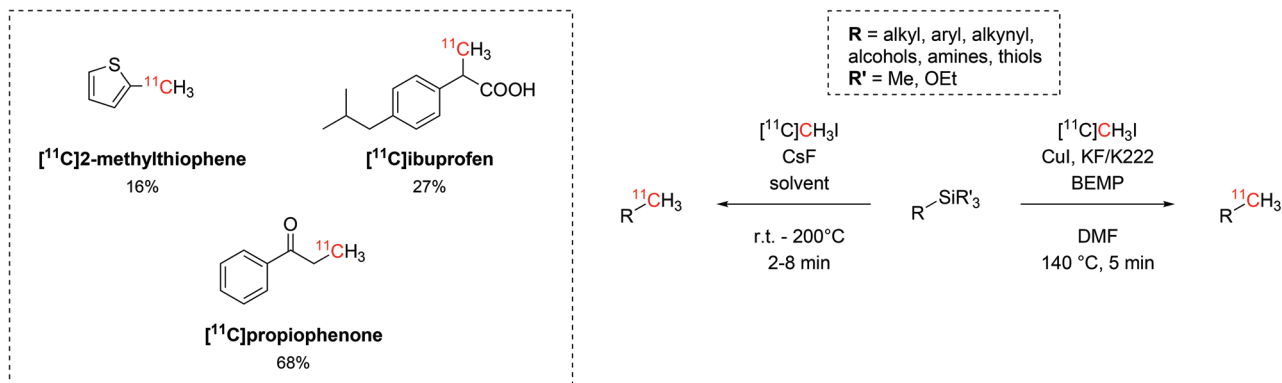
The coupling of organosilicon compounds with $[^{11}\text{C}]\text{CO}$ was also employed in the synthesis of 1-hydroxy-3-phenyl $[^{11}\text{C}]\text{urea}$ by using phenylazide (**19**, Scheme 11) and *O*-(trimethylsilyl)hydroxylamine as reagents (**22**, Scheme 11) whilst keeping the same conditions.⁵⁸ 1-Hydroxy-3-phenyl $[^{11}\text{C}]\text{urea}$ was successfully radiolabelled with RCY of 35% (isolated, decay-corrected, based on delivered $[^{11}\text{C}]\text{CO}$) and A_m of 686 GBq per μmol at 21 minutes from EOB.⁵⁸

The use of trialkylsilyl and trialkoxysilyl precursors was then tested for ^{11}C -methylation with $[^{11}\text{C}]\text{CH}_3\text{I}$ (Scheme 12). The radiolabelling on heteroatoms (O, N, S), aryl and alkyl precursors was readily accomplished by either copper-catalysed or fluoride-activated desilylative reactions and two biologically-active molecules such as $[^{11}\text{C}]\text{propiophenone}$ and $[^{11}\text{C}]\text{jibuprofen}$ were also produced (isolated decay-corrected RCYs = 68% and 27%, respectively, Scheme 12).^{50,51}



Scheme 11 Synthesis of 1-hydroxy-3-phenyl $[^{11}\text{C}]\text{urea}$.⁵⁸





Scheme 12 The use of trialkylsilyl and trialkoxysilyl as precursors in the ^{11}C -methylation with $[^{11}\text{C}]\text{CH}_3\text{I}$.

3. Conclusion

In the past years, the interesting chemistry of silicon-containing compounds has been embraced for radiolabelling purposes, meeting the need for simpler, cost-effective and more efficient radiolabelling methodologies. The versatility of the silicon atom unlocked a plethora of different applications, from $[^{11}\text{C}]\text{CO}_2$ converting agents to radiolabelling precursors, and enabled the production of a large number of biologically-active radiopharmaceuticals. Nonetheless, the applicability of compounds bearing silicon is yet to be fully explored. Taking inspiration from conventional organic chemistry, where silicon-based chemistry is already widely utilised, many more applications could be developed for ^{11}C -labelling. For example, the hydrosilylation of CO_2 have been used for the synthesis of a larger number of chemical structures (formamides, aldehydes, aminals)^{10–13} whereas in ^{11}C -chemistry it's been used only for the production of ^{11}C -*N*-methylamines.^{43,44}

Another potential route for widening the use of ^{11}C -Si chemistry would be taking inspiration from the more established ^{11}C -boron chemistry.⁶⁰ Boron and silicon share many similarities with respect to bond energy and general chemical properties,^{61,62} suggesting that they may also have similar applicability in radiochemistry. Whilst some of these methodologies showed to be suitable for both boron- and silicon-containing molecules (e.g. ^{11}C -carboxylation with $[^{11}\text{C}]\text{CO}_2$, ^{11}C -methylation with $[^{11}\text{C}]\text{CH}_3\text{I}$), organosilicates have not yet been tested in many other radiolabelling strategies, such as the ^{11}C -carbonylation with $[^{11}\text{C}]\text{CO}$ or the ^{11}C -cyanation with $[^{11}\text{C}]\text{HCN}$, which instead are well-established with organoboronates.⁶⁰ Regarding the potential coupling with $[^{11}\text{C}]\text{CO}$, evidence from non-radioactive studies already show its feasibility: the synthesis of unsymmetrical diaryl ketones was achieved by combining a silyl arene, a iodoarene and CO in the presence of KF as activator.⁶³ Likewise, non-radioactive experiments showed that the synthesis of thioesters is achievable by coupling CO with organosilicon precursors.⁶⁴

The development of novel organosilicon radiolabelling strategies could also take inspiration from the well-established

radiochemistry of organostannic compounds.⁶⁵ Exploiting the Stille coupling reaction, the use of organostannanes as radiolabelling precursors found several applications and was coupled with a variety of carbon-11 synthons, such as $[^{11}\text{C}]\text{CH}_3\text{I}$, $[^{11}\text{C}]\text{CO}$ and $[^{11}\text{C}]\text{acyl chloride}$, for the production of a variety of functional groups (e.g. aryl ^{11}C -methylated compounds and ^{11}C -ketones).⁶⁵ The translation of these methodologies into silicon-based ^{11}C -chemistry would represent a big step forward.

Besides the development of novel methodologies, organosilicon radiochemistry may also benefit from broadening the pool of radiolabel-able substrates. The previously discussed techniques are indeed still limited to certain classes of precursors, like the reductive functionalisation of $[^{11}\text{C}]\text{CO}_2$ (Scheme 6) which was poorly tested on alkyl precursors. The development of a “universal” radiolabelling tool, applicable to a larger number of radiotracer regardless of their chemical nature, should then also be taken into consideration.

The use of the radiochemistry of organosilicon compounds should also be enhanced in regular radiopharmaceutical production. The aforementioned methods are still restricted to the research world and not have impacted the broader clinical production of radiotracers yet. Although some clinically-used radiotracers were successfully produced with the discussed methods (e.g. $[^{11}\text{C}]\text{PiB}$ via reductive functionalisation of $[^{11}\text{C}]\text{CO}_2$),^{43,44} traditional radiolabelling processes (e.g. ^{11}C -methylation with $[^{11}\text{C}]\text{CH}_3\text{I}$) are still preferred for routine clinical production in spite of the advantages that novel methods would bring to routine production such as the much shorter radiolabelling time. When considering $[^{11}\text{C}]\text{PiB}$ labelling, for example, the traditional production via ^{11}C -methylation with $[^{11}\text{C}]\text{CH}_3\text{I}$ requires the prior conversion of $[^{11}\text{C}]\text{CO}_2$ to $[^{11}\text{C}]\text{CH}_3\text{I}$, a time-consuming step (around 5 minutes)^{66,67} that instead is not needed in $[^{11}\text{C}]\text{CO}_2$ -based silicon chemistry. Moreover, the presence of a free hydroxyl group on $[^{11}\text{C}]\text{PiB}$ demands for protection/deprotection⁶⁸ to avoid side reactions whilst silicon-based methods do not have these limitations as specific towards amines.⁴⁴ All these factors significantly lower the radiolabelling time from 15 to 1 minute (from end of $[^{11}\text{C}]\text{CO}_2$).



CO₂ delivery to end of synthesis (EOS)),^{44,68} a critical improvement considering the short half-life of ¹¹C. The use of organosilicate methodologies would also lower the costs as not requiring the purchase and maintenance of costly infrastructure such as a standalone methyl iodide production unit – potentially increasing the availability of carbon-11 labelled compounds in laboratories. The RCY of the final radiopharmaceutical would also benefit from the use of the aforementioned methods (12% with [¹¹C]CH₃I versus 26% with silicon chemistry).^{44,68} Hence, the translation of silicon radiochemistry into PET laboratory routine production is another important step that is required.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

King's College London and UCL Comprehensive Cancer Imaging Centre is funded by the CRUK and EPSRC in association with the MRC and DoH (England). The research was supported by the Wellcome's Multi-User Equipment 'A multiuser radioanalytical facility for molecular imaging and radionuclide therapy research' [212885/Z/18/Z]. It was additionally supported by the Wellcome/EPSRC Centre for Medical Engineering at King's College London [WT 203148/Z/16/Z] and the EPSRC Centre for Doctoral Training in Medical Imaging [EP/L015226/1]. The authors also acknowledge PMB Alcen for the financial support.

References

- W. W. Moses, *Nucl. Instrum. Methods Phys. Res., Sect. A*, 2011, **648**(Supplement 1), S236–S240.
- P. W. Miller, N. J. Long, R. Vilar and A. D. Gee, *Angew. Chem., Int. Ed.*, 2008, **47**, 8998–9033.
- M. Conti and L. Eriksson, *EJNMMI Phys.*, 2016, **3**, 8.
- B. H. Rotstein, S. H. Liang, J. P. Holland, T. L. Collier, J. M. Hooker, A. A. Wilson and N. Vasdev, *Chem. Commun.*, 2013, **49**, 5621–5629.
- M. E. Phelps, *Proc. Natl. Acad. Sci. U. S. A.*, 2000, **97**, 9226–9233.
- K. Dahl, C. Halldin and M. Schou, *Clin. Transl. Imaging*, 2017, **5**, 275–289.
- T. Hiyama, *J. Organomet. Chem.*, 2002, **653**, 58–61.
- T. Hiyama and E. Shirakawa, *Cross-Coupling React.*, 2002, 61–85.
- L. Zhang and J. Wu, *J. Am. Chem. Soc.*, 2008, **130**, 12250–12251.
- C. M. So, H. W. Lee, C. P. Lau and F. Y. Kwong, *Org. Lett.*, 2009, **11**, 317–320.
- K. Cheng, S. Hu, B. Zhao, X.-M. Zhang and C. Qi, *J. Org. Chem.*, 2013, **78**, 5022–5025.
- P. Zhang, J. Xu, Y. Gao, X. Li, G. Tang and Y. Zhao, *Synlett*, 2014, **25**, 2928–2932.
- K. Toshima, S. Mukaiyama, M. Kinoshita and K. Tatsuta, *Tetrahedron Lett.*, 1989, **30**, 6413–6416.
- R. D. Crouch, M. Stieff, J. L. Frie, A. B. Cadwallader and D. C. Bevis, *Tetrahedron Lett.*, 1999, **40**, 3133–3136.
- X. Frogneux, O. Jacquet and T. Cantat, *Catal. Sci. Technol.*, 2014, **4**, 1529–1533.
- O. Jacquet, X. Frogneux, C. Das Neves Gomes and T. Cantat, *Chem. Sci.*, 2013, **4**, 2127–2131.
- T. Murata, M. Hiyoshi, M. Ratanasak, J.-y. Hasegawa and T. Ema, *Chem. Commun.*, 2020, **56**, 5783–5786.
- X. Frogneux, E. Blondiaux, P. Thuéry and T. Cantat, *ACS Catal.*, 2015, **5**, 3983–3987.
- M. Tredwell and V. Gouverneur, *Org. Biomol. Chem.*, 2006, **4**, 26–32.
- L. Guo, A. Chatupheeraphat and M. Rueping, *Angew. Chem., Int. Ed.*, 2016, **55**, 11810–11813.
- V. Bernard-Gauthier, C. Wängler, E. Schirrmacher, A. Kostikov, K. Jurkschat, B. Wängler and R. Schirrmacher, *BioMed Res. Int.*, 2014, **2014**, 454503.
- R. Walsh, *Acc. Chem. Res.*, 1981, **14**, 246–252.
- J. Wagler, U. Böhme and G. Roewer, *Organometallics*, 2004, **23**, 6066–6069.
- E. Magnusson, *J. Am. Chem. Soc.*, 1990, **112**, 7940–7951.
- G. L. Larson and J. L. Fry, in *Organic Reactions*, 2009, pp. 1–737. DOI: 10.1002/0471264180.or071.01.
- K. Junold, J. A. Baus, C. Burschka, T. Vent-Schmidt, S. Riedel and R. Tacke, *Inorg. Chem.*, 2013, **52**, 11593–11599.
- H. Kwart and K. King, *d-Orbitals in the Chemistry of Silicon, Phosphorus and Sulfur*, Springer Science & Business Media, 2012.
- R. Motterlini, J. E. Clark, R. Foresti, P. Sarathchandra, B. E. Mann and C. J. Green, *Circ. Res.*, 2002, **90**, e17–e24.
- S. D. Friis, R. H. Taaning, A. T. Lindhardt and T. Skrydstrup, *J. Am. Chem. Soc.*, 2011, **133**, 18114–18117.
- C. Lescot, D. U. Nielsen, I. S. Makarov, A. T. Lindhardt, K. Daasbjerg and T. Skrydstrup, *J. Am. Chem. Soc.*, 2014, **136**, 6142–6147.
- A. G. Brook, *Acc. Chem. Res.*, 1974, **7**, 77–84.
- A. G. Brook, *J. Am. Chem. Soc.*, 1955, **77**, 4827–4829.
- A. G. Brook and H. Gilman, *J. Am. Chem. Soc.*, 1955, **77**, 2322–2325.
- S. K. Zeisler, M. Nader, A. Theobald and F. Oberdorfer, *Appl. Radiat. Isot.*, 1997, **48**, 1091–1095.
- J. M. Lehn and R. Ziessel, *Proc. Natl. Acad. Sci. U. S. A.*, 1982, **79**, 701–704.
- J. Medina-Ramos, R. C. Pupillo, T. P. Keane, J. L. DiMeglio and J. Rosenthal, *J. Am. Chem. Soc.*, 2015, **137**, 5021–5027.
- C. Taddei, S. Bongarzone, A. K. Haji Dheere and A. D. Gee, *Chem. Commun.*, 2015, **51**, 11795–11797.
- F. Luzi, V. Savickas, C. Taddei, S. Hader, N. Singh, A. D. Gee and S. Bongarzone, *Future Med. Chem.*, 2020, **12**, 511–521.



- 39 P. Nordeman, S. D. Friis, T. L. Andersen, H. Audrain, M. Larhed, T. Skrydstrup and G. Antoni, *Chem. – Eur. J.*, 2015, **21**, 17601–17604.
- 40 C. Taddei, S. Bongarzone and A. D. Gee, *Chemistry*, 2017, **23**, 7682–7685.
- 41 L. Zhang, J. Cheng and Z. Hou, *Chem. Commun.*, 2013, **49**, 4782–4784.
- 42 L. González-Sebastián, M. Flores-Alamo and J. J. García, *Organometallics*, 2013, **32**, 7186–7194.
- 43 F. Liger, T. Eijsbouts, F. Cadarossanesaib, C. Tourvieille, D. Le Bars and T. Billard, *Eur. J. Org. Chem.*, 2015, **2015**, 6434–6438.
- 44 P. Buccino, E. Savio and W. Porcal, *EJNMMI Radiopharm. Chem.*, 2019, **4**, 14.
- 45 X.-F. Liu, R. Ma, C. Qiao, H. Cao and L.-N. He, *Chem. – Eur. J.*, 2016, **22**, 16489–16493.
- 46 A. S. Manoso, C. Ahn, A. Soheili, C. J. Handy, R. Correia, W. M. Seganish and P. Deshong, *J. Org. Chem.*, 2004, **69**, 8305–8314.
- 47 S.-C. Lee, L. Guo, H. Yue, H.-H. Liao and M. Rueping, *Synlett*, 2017, **28**, 2594–2598.
- 48 M. Tobisu, Y. Kita and N. Chatani, *J. Am. Chem. Soc.*, 2006, **128**, 8152–8153.
- 49 X. Wang, Z. Wang and Y. Nishihara, *Chem. Commun.*, 2019, **55**, 10507–10510.
- 50 S. Bongarzone, N. Raucci, I. C. Fontana, F. Luzi and A. D. Gee, *Chem. Commun.*, 2020, **56**, 4668–4671.
- 51 W. Qu, B. Hu, J. W. Babich, N. Waterhouse, M. Dooley, S. Ponnala and J. Urgiles, *Nat. Commun.*, 2020, **11**, 1736.
- 52 S. Ram and R. E. Ehrenkaufner, *Tetrahedron Lett.*, 1985, **26**, 5367–5370.
- 53 S. Ram and L. D. Spicer, *J. Labelled Compd. Radiopharm.*, 1989, **27**, 661–668.
- 54 S. Ram, R. E. Ehrenkaufner and D. M. Jewett, *Int. J. Radiat. Appl. Instrum., Part A*, 1986, **37**, 391–395.
- 55 S. Ram and L. D. Spicer, *Int. J. Radiat. Appl. Instrum., Part A*, 1989, **40**, 413–416.
- 56 S. Ram, R. E. Ehrenkaufner and L. D. Spicer, *Int. J. Radiat. Appl. Instrum., Part A*, 1989, **40**, 425–427.
- 57 P. K. Chakraborty, T. J. Mangner and H. T. Chugani, *Appl. Radiat. Isot.*, 1997, **48**, 619–621.
- 58 J. Barletta, F. Karimi and B. Långström, *J. Labelled Compd. Radiopharm.*, 2006, **49**, 429–436.
- 59 S. Syvänen, J. Barletta, G. Blomquist, B. Långström and M. Bergström, *Drug Metab. Lett.*, 2007, **1**, 189–194.
- 60 T. C. Wilson, T. Cailly and V. Gouverneur, *Chem. Soc. Rev.*, 2018, **47**, 6990–7005.
- 61 P. R. Rablen and J. F. Hartwig, *J. Am. Chem. Soc.*, 1996, **118**, 4648–4653.
- 62 G. Rayner-Canham, *Found. Chem.*, 2011, **13**, 121–129.
- 63 T. Hiyama and Y. Hatanaka, *Pure Appl. Chem.*, 1994, **66**, 1471–1478.
- 64 Z. Qiao and X. Jiang, *Org. Lett.*, 2016, **18**, 1550–1553.
- 65 M. Pretze, P. Grosse-Gehling and C. Mamat, *Molecules*, 2011, **16**, 1129–1165.
- 66 J. M. Link, K. A. Krohn and J. C. Clark, *Nucl. Med. Biol.*, 1997, **24**, 93–97.
- 67 B. Långström, G. Antoni, P. Gullberg, C. Halldin, P. Malmberg, K. Nägren, A. Rimland and H. Svärd, *J. Nucl. Med.*, 1987, **28**, 1037–1040.
- 68 C. A. Mathis, Y. Wang, D. P. Holt, G.-F. Huang, M. L. Debnath and W. E. Klunk, *J. Med. Chem.*, 2003, **46**, 2740–2754.

