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Biocatalytic enantioselective hydroaminations enabling synthesis of *N*-arylalkyl-substituted L-aspartic acids†

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***N*-Substituted L-aspartic acids are important chiral building blocks for pharmaceuticals and food additives. Here we report the asymmetric synthesis of various *N*-arylalkyl-substituted L-aspartic acids using ethylenediamine-*N,N'*-disuccinic acid lyase (EDDS lyase) as a biocatalyst. This C–N lyase shows a broad non-natural amine substrate scope and outstanding enantioselectivity, allowing the efficient addition of structurally diverse arylalkylamines to fumarate to afford the corresponding *N*-arylalkyl-substituted L-aspartic acids in good isolated yield (up to 79%) and with excellent enantiopurity (>99% ee). These results further demonstrate that C–N lyases working in reverse constitute an extremely powerful synthetic tool to prepare difficult noncanonical amino acids.**

Introduction

N-Substituted L-aspartic acids and their derivatives are key intermediates in developing promising pharmaceuticals and nutraceuticals with potent biological activities.^{1–5} Therefore, there is a need to prepare these L-aspartic acid derivatives expediently. The conventional chemical synthesis of *N*-substituted L-aspartic acids usually relies on reductive amination or asymmetric hydroamination strategies, which often involves multiple steps and harsh reaction conditions.^{6–9} In this respect, alternative methods need to be pursued to achieve environmentally friendlier and more-step economic synthesis of optically pure *N*-substituted L-aspartic acids. Hence, an enantio-

selective biocatalytic hydroamination approach would be an interesting option.

Two well-studied carbon–nitrogen (C–N) lyases, aspartate ammonia lyase (DAL)^{10–12} and 3-methylaspartate ammonia lyase (MAL),^{10,11,13} have been employed as biocatalysts in the enantioselective hydroamination of fumaric acid to afford *N*-substituted L-aspartic acids. In addition, we started to investigate another interesting C–N lyase, ethylenediamine-*N,N'*-disuccinic acid (EDDS) lyase from *Chelativorans sp.* BNC1, which utilizes a characteristic serine residue (Ser-280) as the catalytic base to facilitate the deamination of (*S,S*)-EDDS to give ethylenediamine and two fumarate molecules.¹⁴ We showed that EDDS lyase accepts various non-native amine substrates in the hydroamination of fumaric acid, including various linear mono- and diamines, homo- and heterocycloalkyl amines, arylamines, and arylhydrazines, enabling the enantioselective synthesis of a wide variety of *N*-substituted L-aspartic acid derivatives.^{14–16} The broad amine scope of EDDS lyase also supported the chemo-enzymatic synthesis of the fungal natural products aspergillomarasmine A, aspergillomarasmine B, and toxin A, as well as related aminocarboxylic acids.¹⁷ EDDS lyase was recently optimized by structure-inspired engineering for the enantioselective synthesis of several challenging *N*-substituted aspartic acids, which are important chiral precursors to artificial dipeptide sweeteners such as neotame and advantame.¹⁸

The absence of cofactors and its capacity to catalyze enantioselective hydroaminations with a variety of amines, make EDDS lyase an attractive enzyme for biocatalytic applications. In this study, we further explored the non-native amine substrate scope of EDDS lyase using a panel of structurally diverse arylalkylamines in the hydroamination of fumaric acid to yield various enantiopure *N*-arylalkyl-substituted L-aspartic acids.

Results and discussion

We previously reported that both aniline (**2x**, *n* = 0) and benzylamine (**2y**, *n* = 1) are accepted as non-native substrates in the hydroamination of fumaric acid (**1**) by EDDS lyase

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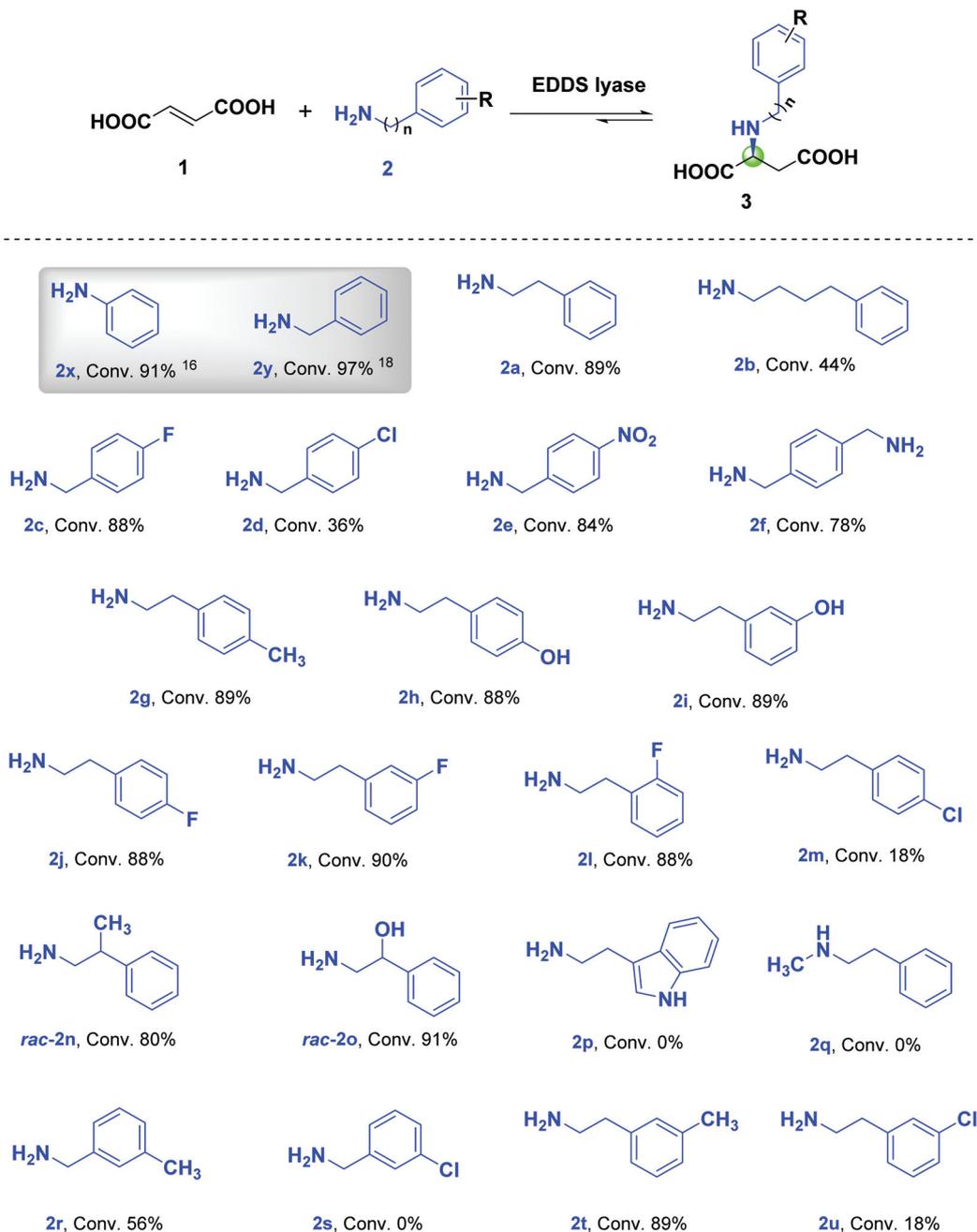
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(Scheme 1).^{16,18} This observation prompted us to test a wide variety of arylalkylamines (**2a–2u**) with different aliphatic linkers between the amino and aryl groups, as well as having different substituents, as potential substrates in the hydroamination reaction catalyzed by EDDS lyase. The progress of the enzymatic hydroaminations was monitored by ¹H NMR spectroscopy (Fig. S1–S21[†]). Pleasingly, amines **2a–2o**, **2r**, **2t** and

2u were accepted as non-native substrates by EDDS lyase (up to 91% conversion), while amines **2p**, **2q** and **2s** were not processed (Scheme 1).

The EDDS lyase catalyzed hydroamination of **1** with 2-phenylethylamine (**2a**) and 4-phenylbutylamine (**2b**) resulted in 89% and 44% conversions, respectively (Scheme 1, Fig. S1 and S2[†]). The lower conversion observed with **2b** compared to



Scheme 1 The broad arylalkylamine substrate scope of EDDS lyase enables the asymmetric synthesis of difficult *N*-substituted L-aspartic acids. Reaction mixtures contained EDDS lyase (15 μM), **1** (10 mM) and **2a–2u** (50 mM) in 3 mL NaH_2PO_4 buffer (20 mM, pH 8.5). Reaction mixtures were incubated at room temperature for 24 h; except for those containing **2c**, **2e**, **2f**, and **2n**, which were incubated for 48 h. Conversion was determined by ¹H NMR spectroscopy by comparing signals of the amine substrates and corresponding products.



2a (as well as **2x** and **2y**) clearly shows that EDDS lyase prefers amine substrates with short (≤ 2) aliphatic linkers between the amino and aryl groups. Interestingly, the enzyme accepts distinct *para*-substituted benzylamines (**2c–2e**) with reasonable to good conversions (Fig. S3–S5†). Under the conditions used, the EDDS lyase catalyzed hydroamination reaction between diamine **2f** (1,4-phenylenedimethanamine) and **1** resulted in the single-addition product with 78% conversion (Fig. S6†).

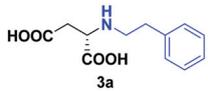
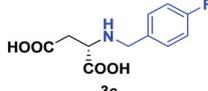
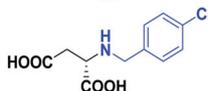
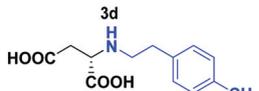
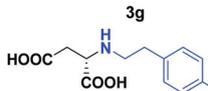
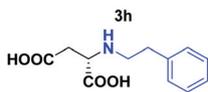
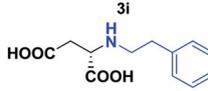
To further examine the versatility of EDDS lyase, a variety of phenethylamines with different substitutions on the aryl ring such as *p*-methyl-phenethylamine (**2g**), *p*- and *m*-hydroxy-phenethylamine (**2h** and **2i**), *p*-, *m*- and *o*-fluoro-phenethylamine (**2j–2l**) and *p*-chloro-phenethylamine (**2m**) were tested as non-native amine substrates. EDDS lyase accepted these phenethylamines as substrates in the hydroamination of **1**, giving excellent conversion (88–90%) for all amines with the exception of **2m** (18%) (Fig. S7–S13†). Overall, the electronic nature and position of the substituents on the aryl ring did not show a significant effect on conversion. However, the *p*-chloro substituent

on phenethylamine (**2m**) had a negative influence on conversion, which resembles the negative consequence of the *p*-chloro substituent on the transformation of benzylamine (**2d**). The crystal structure of EDDS lyase does not provide an immediate explanation for the difficulties in accommodating the *p*-chloro-phenyl group.¹⁴

Notably, prolonged incubation of EDDS lyase with amines **2d** and **2m** resulted in limited enzyme precipitation, which may also at least partly explain their lower conversion. This effect was not observed upon incubation of the enzyme with the other amines. Further investigation indeed shows that amines **2r** and **2t** (carrying a *m*-methyl group) are well converted, whereas the corresponding amines **2s** and **2u** (carrying a *m*-chlorine group) are not or poorly processed (Fig. S18–S21†), showing limited enzyme precipitation upon extended incubation.

We further tested racemic 2-substituted phenethylamines (*rac*-**2n** and *rac*-**2o**) as non-native amine substrates to study the enantioselectivity of EDDS lyase. The enzyme accepted both

Table 1 Semi-preparative-scale synthesis of *N*-arylkyl-substituted L-aspartic acids via EDDS lyase catalyzed enantioselective hydroamination^a

Entry	Amine	Product	Reaction time (h)	Yield ^b (%) (mg)	ee ^c (%)	Abs. conf. ^d
1	2a		24	70 (25)	>99	S
2	2c		48	75 (27)	>99	S
3	2d		24	28 (11)	>99	S
4	2g		24	66 (25)	>99	S
5	2h		24	61 (23)	>99	S
6	2i		24	63 (24)	>99	S
7	2j		24	76 (29)	>99	S

^a Reaction mixtures contained **1** (10 mM, 0.15 mmol), **2a**, **2c**, **2d** or **2g–j** (50 mM), and EDDS lyase (15 μ M) in 15 ml NaH₂PO₄ buffer (20 mM, pH 8.5, room temperature). ^b Isolated product yield after ion-exchange chromatography. ^c The ee was determined by high-performance liquid chromatography (HPLC) using a chiral stationary phase and chemically synthesized authentic standards. ^d Determined by HPLC using a chiral stationary phase and chemically synthesized authentic standards.



enantiomers of *rac*-**2n** and *rac*-**2o**, resulting in 80% and 91% conversion, respectively (Fig. S14 and S15†). Hence, the relatively small methyl and hydroxyl substituents are not able to induce enantioselective enzymatic transformation. Lastly, tryptamine (**2p**) and *N*-methyl-phenethylamine (**2q**) were tested as potential amine substrates for EDDS lyase. These two compounds were not accepted by EDDS lyase (Fig. S16 and S17†), indicating that the enzyme does not accept very bulky amines nor secondary amines as substrates for hydroamination of **1**.

Despite the use of non-native amine substrates, respectable specific activities are observed, illustrated by the addition of **2a** (140 mU mg⁻¹), **2c** (21 mU mg⁻¹) and **2j** (119 mU mg⁻¹) to **1** (Fig. S71†). For comparison, EDDS lyase exhibits a specific activity of 543 mU mg⁻¹ for the addition of ethylenediamine to **1**. To further demonstrate the synthetic usefulness of the EDDS lyase catalyzed hydroamination reactions, we performed semi-preparative scale synthesis of a few selected *N*-substituted aspartic acids (**3a**, **3c**, **3d**, and **3g–3j**). High conversions, excellent enantiocontrol (>99% ee), and good isolated product yields (up to 76%) were achieved (Table 1, Fig. S22–S28 and S43–S49†). Conveniently, EDDS lyase exclusively produced the desired *S* enantiomer of the *N*-substituted-aspartic acids (Fig. S64–S70†), consistent with the previously reported enantioselectivity of this enzyme, and in accordance with the experimentally determined binding mode of (*S,S*)-EDDS in the crystal structure of EDDS lyase (Fig. S72†).^{14–18} With fumarate being firmly bound at the back of the active site pocket, the amino group of the second substrate can only perform a pro-*S* attack (*re*-face of C α) because of steric reasons (*si*-face is not accessible) and because of a favourable hydrogen bond formed with the side chain of Asn113. Thus, despite the use of structurally diverse amines, ranging from small alkylamines to large amino acids, the stereochemical outcome of the EDDS lyase catalyzed hydroamination reaction is not affected.

Next, we investigated whether the stereochemical outcome of the enzymatic hydroamination reaction could be altered by using maleic acid (*i.e.*, *cis*-butenedioic acid) as electrophile instead of fumarate (*i.e.*, *trans*-butenedioic acid). However, the enzyme was found to be stereospecific for fumarate, with maleic acid not accepted as alternative electrophile (Fig. S73†). Given that EDDS lyase can also not accept fumaric acid monomethyl ester, crotonic acid, mesaconic acid, itaconic acid, 2-pentenoic acid, and glutaconic acid as alternative substrates,¹⁴ the redesign of EDDS lyase for hydroamination of alternative electrophiles remains an important challenge.

In summary, we have shown that EDDS lyase accepts a wide variety of arylalkylamines, with different aliphatic linkers between the amino and aryl groups, as well as carrying different substituents, in the asymmetric hydroamination of fumarate yielding the desired *N*-substituted *L*-aspartic acids with high optical purity and in good isolated yield. Together with previously reported work from our laboratory,^{14–18} this study demonstrates that EDDS lyase has an exceptionally

broad amine scope, making it a very attractive biocatalyst for enantioselective production of various hard to synthesize *L*-aspartate derivatives. Examples of important *L*-aspartate derivatives include biodegradable metal chelators, metallo-beta-lactamase and glutamate transporter inhibitors, photo-switchable and photocaged drug-like molecules, as well as various *N*-heterocycles and other chiral synthons that are widely used in pharmaceutical synthesis.^{19–21} We have recently initiated structure-based and computer-aided protein engineering studies with the aim to enlarge the electrophile substrate scope of EDDS lyase to further increase its synthetic usefulness.

Conflicts of interest

There are no conflicts to declare.

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References

- M. A. T. Blaskovich, *J. Med. Chem.*, 2016, **59**, 10807–10836.
- M. Maiti, M. Maiti, J. Rozenski, S. De Jonghe and P. Herdewijn, *Org. Biomol. Chem.*, 2015, **13**, 5158–5174.
- A. M. King, S. A. Reid-Yu, W. Wang, D. T. King, G. De Pascale, N. C. Strynadka, T. R. Walsh, B. K. Coombes and G. D. Wright, *Nature*, 2014, **510**, 503.
- B. C. McIlwain, R. J. Vandenberg and R. M. Ryan, *Biochemistry*, 2016, **55**, 6801–6810.
- S. Chattopadhyay, U. Raychaudhuri and R. Chakraborty, *J. Food Sci. Technol.*, 2014, **51**, 611–621.
- A. Zilkha and M. D. Bachi, *J. Org. Chem.*, 1959, **24**, 1096–1098.
- D. M. Roundhill, *Chem. Rev.*, 1992, **92**, 1–27.
- P. S. Piispanen and P. M. Pihko, *Tetrahedron Lett.*, 2005, **46**, 2751–2755.
- M. Boros, J. Kökösi, J. Vámos, I. Kövesdi and B. Noszál, *Amino Acids*, 2007, **33**, 709–717.
- M. de Villiers, V. Puthan Veetil, H. Raj, J. de Villiers and G. J. Poelarends, *ACS Chem. Biol.*, 2012, **7**, 1618–1628.
- F. Parmeggiani, N. J. Weise, S. T. Ahmed and N. J. Turner, *Chem. Rev.*, 2018, **118**, 73–118.
- R. Li, H. J. Wijma, L. Song, Y. Cui, M. Otzen, Y. Tian, J. Du, T. Li, D. Niu, Y. Chen, J. Feng, J. Han, H. Chen, Y. Tao, D. B. Janssen and B. Wu, *Nat. Chem. Biol.*, 2018, **14**, 664–670.
- H. Raj, W. Szymański, J. De Villiers, H. J. Rozeboom, V. Puthan Veetil, C. R. Reis, M. De Villiers, F. J. Dekker,



- S. De Wildeman, W. J. Quax, A.-M. W. H. Thunnissen, B. L. Feringa, D. B. Janssen and G. J. Poelarends, *Nat. Chem.*, 2012, **4**, 478–484.
- 14 H. Poddar, J. de Villiers, J. Zhang, V. Puthan Veetil, H. Raj, A.-M. W. H. Thunnissen and G. J. Poelarends, *Biochemistry*, 2018, **57**, 3752–3763.
- 15 J. Zhang, H. Fu, P. G. Tepper and G. J. Poelarends, *Adv. Synth. Catal.*, 2019, **361**, 2433–2437.
- 16 H. Fu, A. Prats Luján, L. Bothof, J. Zhang, P. G. Tepper and G. J. Poelarends, *ACS Catal.*, 2019, **9**, 7292–7299.
- 17 H. Fu, J. Zhang, M. Saifuddin, G. Cruiming, P. G. Tepper and G. J. Poelarends, *Nat. Catal.*, 2018, **1**, 186–191.
- 18 J. Zhang, E. Grandi, H. Fu, T. Saravanan, L. Bothof, P. G. Tepper, A.-M. W. H. Thunnissen and G. J. Poelarends, *Angew. Chem.*, 2020, **59**, 429–435.
- 19 K. H. M. E. Tehrani, H. Fu, N. C. Brüche, V. Mashayekhi, A. Prats Luján, M. J. van Haren, G. J. Poelarends and N. I. Martin, *Chem. Commun.*, 2020, **56**, 3047–3049.
- 20 J. Zhang, M. Z. Abidin, T. Saravanan and G. J. Poelarends, *ChemBioChem*, 2020, **21**, 2733–2742.
- 21 V. Arkhipova, H. Fu, M. W. H. Hoorens, G. Trinco, L. N. Lameijer, E. Marin, B. L. Feringa, G. J. Poelarends, W. Szymanski, D. J. Slotboom and A. Guskov, *J. Am. Chem. Soc.*, 2021, **143**, 1513–1520.

