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Multi-component bioresponsive nanoparticles for synchronous delivery of docetaxel and TUBB3 siRNA to lung cancer cells†

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Bioresponsive nanoparticles (NPs) are of interest for anticancer nanomedicines, owing to the possibility to 'design in' selective modulation of drug release at target sites. Here we describe the double emulsion formulation of redox-responsive NPs based on modified polyethylene glycol (PEG)-*co*-poly(lactic-*co*-glycolic acid) (PLGA) block copolymers and oligo (β -aminoesters) (OBAE), both of which contained disulfide linkages, for the co-delivery of a cytotoxic small molecule drug and a nucleic acid. In particular, we focused our attention on docetaxel (DTX) and a siRNA against *TUBB3*, a gene that encodes for β III-tubulin, in order to have a synergistic effect in the treatment of lung cancer. Spherical NPs of around 150 nm with negative zeta potential and high loading efficiencies of both drugs were obtained. Stability and release studies showed "on demand" drug release under reducing conditions. Unloaded NPs containing PEG-disulfide-PLGA and OBAE were well-tolerated by lung cancer cells, thus masking the intrinsic cytotoxicity of OBAE, while for intracellular siRNA delivery, redox responsive NPs demonstrated a higher cell internalization with a preferential cytoplasmic accumulation of siRNA, with a subsequent fast gene-silencing efficiency. The viability of cells treated with combined DTX/*TUBB3*-siRNA NPs significantly decreased as compared to NPs loaded only with DTX, thus showing an efficient combined anticancer effect, due to a substantial reduction of β -tubulin expression. Finally, in an *in vivo* feasibility study employing an orthotopic lung cancer model, NPs formulated with an anti-luciferase siRNA distributed throughout the lungs following oro-tracheal administration, and demonstrated effective gene knockdown and no apparent cytotoxicity. Taken together, these results show that the double emulsion formulated redox responsive PEG-PLGA and OBAE systems represent a promising new therapeutic approach for the local combined chemo- and gene-therapy of lung cancer.

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Introduction

Lung cancer (LC) is one of the most common cancers worldwide and non-small cell lung cancer (NSCLC) accounts for approximately 85% of all LC cases.¹ The simultaneous administration of two or more conventional anticancer drugs represents the first-line treatment for these patients, but the development of multidrug resistance (MDR) strongly limits the therapeutic effectiveness. Therefore, combinations of multiple therapeutic modalities (*e.g.* chemotherapy, gene therapy, immunotherapy, photodynamic therapy) with additive/synergistic mechanisms of cell killing are becoming standard in clinical settings for combating resistance mechanisms.^{2,3} In particular, regulation of gene expression through RNA-interference based therapeutics is currently of interest, due to the ability of these molecules to silence specific pathways involved



in MDR.^{4,5} For instance, more than 2500 short-interfering RNA (siRNA) patents for LC have been developed so far.⁶ Nevertheless, the clinical application of siRNA in combination with other therapeutics is challenging due to the difference in chemical and biopharmaceutical profiles. Moreover, major obstacles to siRNA therapy arise from extracellular and intracellular barriers, including (i) *in vivo* degradation *via* nucleases (ii) inability to cross endothelial membranes and (iii) poor efficiency of endosomal escape to reach their cytoplasmic target. As a result, better delivery of siRNA is required in order to drive these delicate molecules into the body and to modulate their fate after administration.^{7,8} In this context, nanomedicine-based approaches for RNA delivery have been explored, aiming to improve pharmacological effectiveness at disease sites and reduce off-target effects.^{9–15} However, the features of nanoparticle (NP) delivery systems need to be refined carefully in terms of composition, structure, size, surface and stability, and to take into account (i) the drugs to be delivered, (ii) the characteristics of the tumor microenvironment, (iii) the administration route and (iv) the final target.¹⁶ Current research is focused on the design of carrier materials able to respond to intrinsic or extrinsic stimuli (pH decrease, redox gradients, enzyme action, light, magnetic fields, temperature, ultrasound or electric currents) for “on demand” delivery at a diseased site.^{13,17,18} In particular, by exploiting the differences in reduction potential between normal tissues and certain tumor microenvironments, redox-responsive NPs have been shown to release drugs selectively at specific subcellular locations.¹⁹ It is well-known that the cytosol, the mitochondria and the nucleus of tumor cells are characterized by high concentrations of reductive glutathione (GSH) tripeptides (approx. 10 mM, about 100 to 1000 times higher than those in the extracellular fluids and normal tissues). For this reason, GSH-responsive NPs have been recognized as promising intracellular delivery systems for cancer and gene therapy.²⁰

Finally, for the treatment of severe lung disorders, including LC, administration by the pulmonary route is commonly preferred, since it allows fast and “needle-free” direct access to the target sites, thus reducing systemic effects. However, it is vital to consider the anatomical and physiological features of the lung that can prevent drug and siRNA accumulation. These include mucociliary clearance and enzymatic barriers in the alveolar fluid that promote siRNA degradation.²¹ For these reasons, inhalable NPs for lung delivery must meet some demanding formulation criteria.^{22,23}

In order to develop effective inhalable NPs for pulmonary delivery, we have focused on redox responsive NPs based on a biodegradable and amphiphilic diblock copolymer containing a reducible disulfide bridge between poly(lactic-co-glycolic acid) (PLGA) and polyethylene glycol (PEG) blocks intended for the treatment of LC. We have chosen as building blocks PEG and PLGA, for their well-established regulatory profile that renders our approach promising for future *in vivo* applications. PLGA and PEG are in fact both biodegradable and nontoxic polymers approved by the United States Food and Drug Administration (FDA) for medical uses.²⁴ Moreover, this

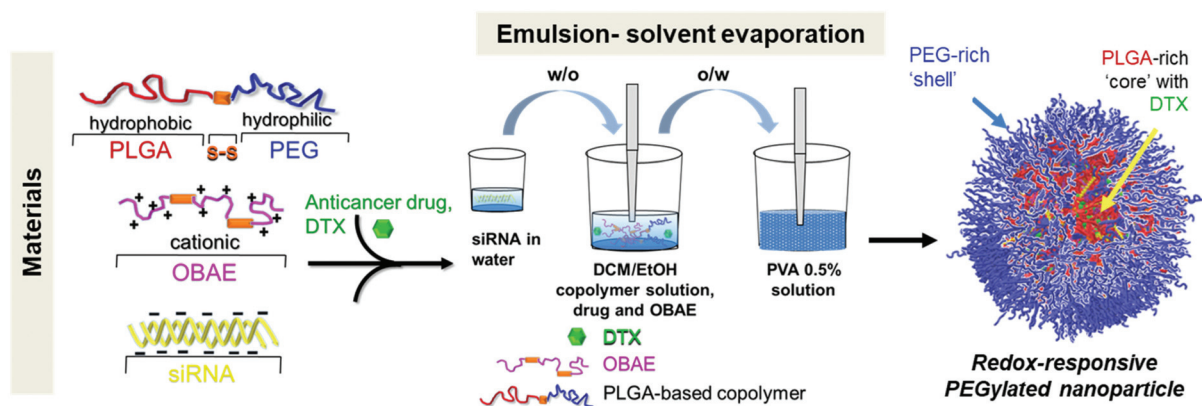
system has previously shown (i) good ability to entrap and trigger on-demand release of the conventional anticancer drug docetaxel (DTX) in conditions simulating intracellular reductive environments; (ii) high penetration properties through an artificial mucus layer due to the presence of the external PEG coating and (iii) enhanced uptake in 2D- and 3D- LC models due to a double (extra- and intra- cellular) redox responsive behavior.²⁵ For this work, the desire to co-deliver siRNA for binary-drug therapy required modification of the carrier by the introduction of cationic oligo- β -aminoesters (OBAE). We chose OBAEs for their ability to condense nucleic acids at physiological pH, protect them in their passage across cell membranes, and enable payload release into cell interiors. The latter can occur within acidic endosomal or lysosomal environments, due to the presence of reversibly protonatable amine groups and acid or base-sensitive ester bonds, which are ultimately degraded into non-toxic small molecules.²⁶ For this study, we have employed a disulfide-containing OBAE, which, as we recently reported, was able to condense siRNA into polyplexes through a fast inkjet printing method.²⁷ The specific OBAE in this study was expected to protect siRNA against biological delivery barriers but also break down intracellularly to enable complete release of the nucleic acid cargo.^{28,29} Through this chemical strategy, we combined the mucus penetration properties of PEG-coated NPs,³⁰ thus enhancing pulmonary delivery, with functionality for nucleic acid binding in the OBAE component, and bioresponsive chemistries in both components encoding for site-specific release.

We loaded NPs with DTX, clinically approved for LC treatment, and a therapeutic siRNA (*TUBB3*-siRNA) against β III-tubulin, a protein involved in microtubule formation. A growing body of preclinical and clinical data suggests that increased expression of this protein can reduce the anti-mitotic activity of DTX and taxanes in general,³¹ thus conferring cancer resistance in different tumor types, including NSCLC.^{32–34} Therefore, we considered that combining knock-down of β III-tubulin expression *via* siRNA with delivery of DTX would be a potential and promising strategy to enhance cancer cell kill with taxanes. After an in-depth evaluation of NP behavior in LC cells, proof-of-principle for the translational potential of these NPs was explored *in vivo* by assessing the tolerability, biodistribution and transfection of siRNA-loaded NPs after oro-tracheal administration in an orthotopic LC model. These studies represent the first steps in establishing the initial safety and efficacy profiles of the novel formulated redox-responsive NPs, in preparation for future full combination therapy studies, in view of a potential therapeutic alternative to the clinically used first-line lung cancer therapy.

Results and discussion

Bioresponsive NPs were effectively derived from polymers of PLGA and OBAE, each containing bioreducible disulfide





Scheme 1 Schematic illustration of materials and NP preparation technique.

bridges and with an outer PEG layer to allow good penetration through pulmonary barriers.³⁵ OBAEs are able to condense siRNA and deliver it in the same cancer cell line due to its cationic charge, redox responsiveness and high buffering capacity to allow the endosomal escape of nucleic acids. As following step, herein we have decided to combine PLGA-block copolymers with OBAEs in the same nanoplatform to: (i) ensure the tolerability of the formulation due to the well-known safety of PLGA and PEG polymers; (ii) exploit the mucus penetrating properties of PEG, critical for a pulmonary administration; (iii) load with high efficiency a siRNA for the presence of the cationic OBAE in NP core, able to induce a proton sponge effect at cell level; (iv) load a second drug in the polymeric matrix of NPs, in order to have a combined effect with siRNA.

Preparation and characterization of NPs

PLGA-PEG and PLGA-SS-PEG block copolymers were mixed with cationic disulfide containing OBAE and employed to prepare NPs which were either wholly (PLGA-SS-PEG, OBAE) or partially (PLGA-PEG, OBAE) redox-responsive, respectively. In particular, PLGA-based copolymers and OBAE were mixed in the same NP in order to have a carrier with a PEG outer layer to enhance pulmonary mucus penetration, and able to entrap simultaneously lipophilic drugs, such as DTX, and hydrophilic siRNA. The first drug is located in the PLGA core due its lipophilic profile, whereas the siRNA is entrapped through electrostatic interaction with the cationic charges of OBAE. For an

appropriate set of comparative materials, we therefore prepared unloaded NPs, DTX-loaded NPs, siRNA-loaded NPs and combined siRNA/DTX-loaded NPs.

NPs were prepared by a modified double emulsion-solvent evaporation technique²⁵ and characterized in terms of size, PDI, ζ and morphology. A schematic representation of synthesis of the materials and NP preparation is shown in Fig. S1 and Scheme 1, respectively. After a preliminary formulation study by adjusting the processing parameters of the emulsification-solvent evaporation technique, we found that the PLGA/OBAE mass ratio 10:1 was the best condition to obtain NPs with good colloidal properties and encapsulation of both DTX and siRNA. Initially, OBAE was dissolved in EtOH and then mixed with the DCM/PLGA solution at a volume ratio 1:9. DTX was co-dissolved in the organic phase with OBAE and PLGA-copolymers, and siRNA dissolved in the internal water phase before the emulsification process. Composition and properties of all the formulations are reported in Table 1. The yield for the formulations was high (~70%) indicating that no extensive polymer/drug precipitation or NP aggregation occurred during preparation. The primary properties of the PLGA-PEG and PLGA-SS-PEG formulations were similar, thus highlighting a comparable assembling behavior of the copolymers. NPs displayed hydrodynamic diameters around 150 nm, with acceptable polydispersity indices and negative ζ potential values. The spherical morphology and the absence of NP aggregation was confirmed by TEM images (Fig. 1A).

Table 1 Composition and properties of NPs. SD were calculated on three different batches

Code	OBAE (% w/w)	DTX (% w/w)	siRNA (% w/w)	Yield (%)	Mean D_H (nm \pm SD)	PDI	ζ (mV \pm SD)	DTX actual loading ^a (entr. eff% ^b)	siRNA actual loading ^a (entr. eff% ^b)
PLGA-PEG	10	—	—	75	153 \pm 7	0.1	-9.9 \pm 2.3	—	—
DTX/PLGA-PEG	10	10	—	69	174 \pm 5	0.2	-9.2 \pm 4.4	6.3 \pm 0.4 (71.2 \pm 4.8)	—
TUBB3-DTX/PLGA-PEG	10	10	0.1	65	183 \pm 9	0.2	-9.7 \pm 2.7	5.5 \pm 1.1 (57.7 \pm 8.3)	0.1 \pm 0.01 (98.1 \pm 1.9)
PLGA-SS-PEG	10	—	—	71	143 \pm 2	0.1	-11.2 \pm 3.4	—	—
DTX/PLGA-SS-PEG	10	10	—	70	173.0 \pm 7	0.2	-12.2 \pm 3.5	6.5 \pm 2.4 (74.3 \pm 7.1)	—
TUBB3-DTX/PLGA-SS-PEG	10	10	0.1	73	178 \pm 5	0.2	-8.4 \pm 3.3	6.2 \pm 5.1 (69.7 \pm 4.5)	0.1 \pm 0.03 (94.2 \pm 3.3)

^a Actual loading is expressed as the amount (mg) of drug encapsulated per 100 mg of NPs. ^b Ratio between actual and theoretical loading \times 100.



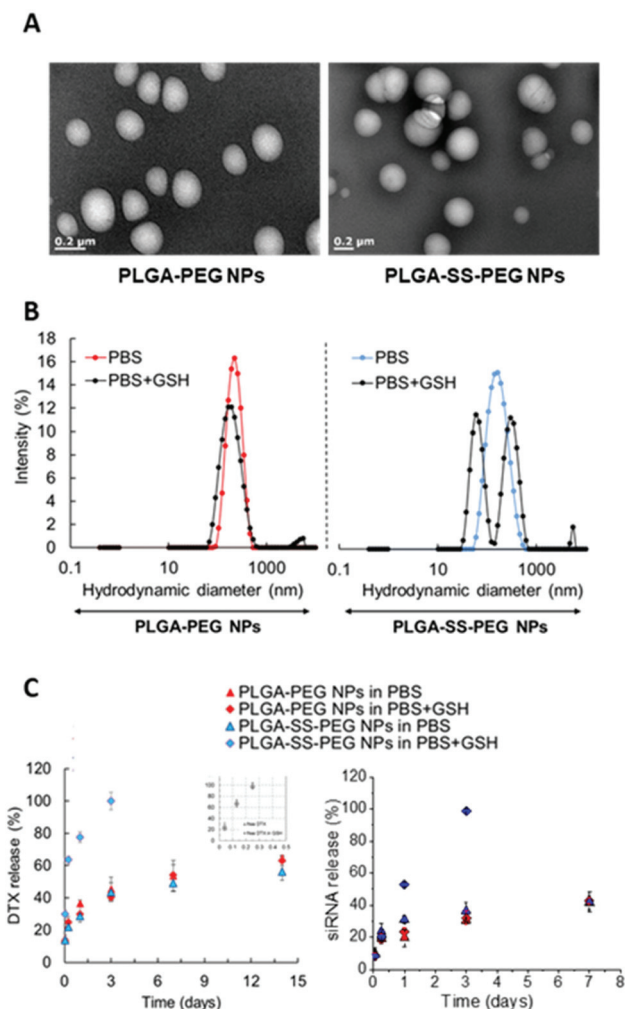


Fig. 1 NP characterization. (A) FEG-TEM images of TUBB3-DTX/NPs; (B) size distribution curves of NPs incubated in PBS 10 mM pH 7.4 with or without GSH (10 mM) at 37 °C for 30 min; (C) *in vitro* release of DTX and siRNA from TUBB3-DTX/NPs in PBS with or without GSH 10 mM. Results are expressed as release% over time \pm SD of three experiments.

Entrapment of both drugs did not influence significantly surface properties and ζ potentials of NPs, whereas a slight increase of size as well as polydispersity index for both formulations was observed when DTX was loaded.

However, the entrapment of siRNA in the systems did not influence the capability of PLGA-cores to load the lipophilic drug DTX. The complete entrapment of siRNA in all the formulations was confirmed by a quantitative assay through Nanodrop-UV measurements as well as by gel retardation assays (Fig. S2†). In order to investigate the redox responsiveness of NPs in a simulated intracellular reductive environment, different experimental techniques were performed, including stability and release studies. As clearly evidenced in Fig. 1B, PLGA-SS-PEG NPs showed low stability in GSH reducing media (10 mM), highlighted by the changing of the initial particle size and distribution curves, compared to PLGA-PEG NPs, in line with TEM images of NPs that we have already shown in

our previous paper,³⁶ where a strong deformation of PLGA-SS-PEG NP structure was clearly evidenced in GSH. At the same time, Fig. S2† evidenced a partial release of siRNA from PLGA-SS-PEG NPs after incubation in GSH. These results indicate the release of the surface-displayed PEG chains from the NPs in a simulated intracellular reductive environment through the cleavage of the disulfide bond between the PLGA and the PEG blocks, in line with prior data from similar polymers. To confirm these data, release studies of DTX from NPs retained in dialysis membranes were carried out in the presence of GSH 10 mM in the releasing medium. In Fig. 1C, it is possible to note that both PLGA-PEG NPs and PLGA-SS-PEG NPs exhibited a sustained release pattern of DTX in PBS. In particular, around 50% of the entrapped DTX was released in 14 days, compared with the free drug in the same conditions (inset). In contrast, with the addition of GSH in the releasing medium, complete drug release was observed from PLGA-SS-PEG NPs after 3 days of incubation. This trend was not significantly affected by the presence of OBAE in the formulations. The same trend was found for siRNA release.

Effects of NPs in lung cancer cells

Before moving to cell studies, we tried to predict the stability of NPs in simulated biological conditions. In fact, it is well known that on contact with physiological fluids, some NPs will interface with a wide range of biomacromolecules, including proteins that can strongly modify NP colloidal properties and identity. However, our system showed good stability in Fetal Bovin Serum (FBS), the major protein component of cell culture media as evident in Fig. S3,† which we attribute to the display of PEG-chains at the NP surfaces.

A key obstacle to clinical translation of siRNA-based nanomedicine is the extensive cytotoxicity related to many cationic materials which have been tested, a problem which is partly due to the limited biodegradability of commonly used polycations.³⁷ For this reason, the ‘bioreducible nanocarrier’ approach has been developed to resolve the efficacy–toxicity dilemma.³⁸ However, the overall molar mass and molar mass distribution can significantly affect the transfection activity and cytotoxicity even of biodegradable polymers.³⁹ We therefore compared the effects of free OBAE to unloaded NPs on cell metabolic activity, as a proxy for cell viability. These experiments were conducted in A549 and H1299 LC cells by the MTS assay as described in the Experimental section. Cells were incubated with a wide range of NP concentrations (0.05–5 mg mL⁻¹) and tested after 24 h of treatment. At the same time, the effects on metabolic activity of free OBAE in the same concentration range entrapped into NPs (0.005 mg–0.5 mg) was evaluated. Results from MTS assays (Fig. 2A and S4†) show that 24 h after treatment, A549 and H1299 cells retained about 90% viability when treated with various concentrations of NPs as compared with free OBAE. In the latter case, a higher concentration dependent cytotoxicity was apparent, with a reduction in metabolic activity of ~80%. The reduced toxicity of the PLGA-based NPs modified with cationic OBAE suggested that any cationic charges in these formulations were effectively



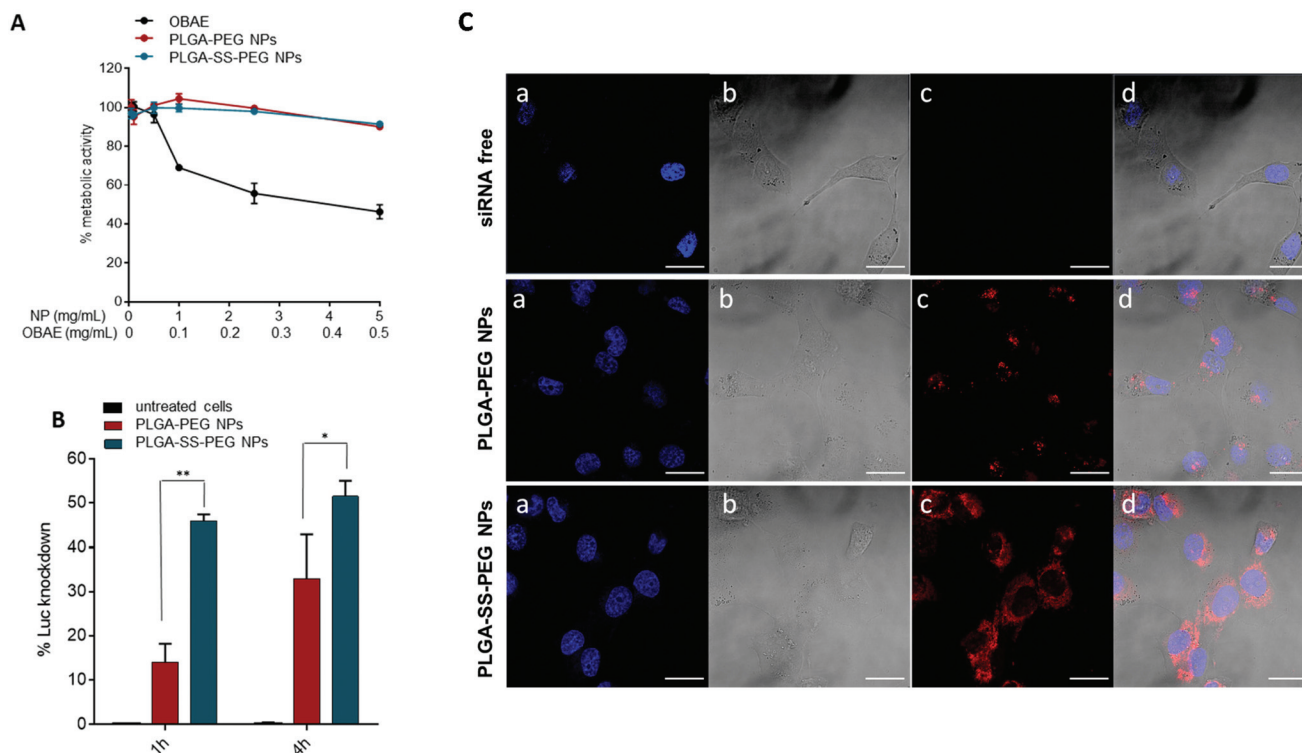


Fig. 2 (A) Cytotoxicity of unloaded NPs against A549 lung cancer cells compared to free OBAE (24 h of treatment). OBAE concentration range: 0.005–0.5 mg mL⁻¹. (B) *In vitro* luciferase siRNA transfection in A549-luciferase expressing cells after different NP/cell time contact. Results are expressed as mean \pm SD of three experiments. ** $P < 0.01$, * $P < 0.05$ two-way ANOVA test. (C) Confocal images of A549 cells after incubation with CyTM3-siRNA loaded NPs (125 μ g mL⁻¹) for 4 h: (a) nucleus stained with Hoechst 33258, (b) brightfield, (c) CyTM3-siRNA emission in red (λ_{ex} : 555 nm; λ_{em} : LP 560 nm), (d) merge. Scale bar: 20 μ m. Zen 2009 image Software was utilized for image processing.

shielded from the cells. To highlight the safety of these materials, NPs were tested on the same cell lines up to 50 mg mL⁻¹ and no toxicity was found (Fig. S5[†]). Subsequently, the transfection efficiency of NPs loaded with a siRNA-based luciferase knock-down sequence was assessed on luciferase-expressing A549 cancer cells.

The primary properties of NPs loaded with Luc-siRNA are reported in Table S1[†] and are comparable to the other formulations. The silencing efficiency was evaluated after 1 h and 4 h of NP incubation into cancer cells and compared to untreated cells. As previously stated, we have already demonstrated the ability of free OBAEs to deliver siRNA in cancer cells,²⁷ but due to the cytotoxicity of this cationic material, we decided to not include this control in the following experiments. For redox responsive PLGA-SS-PEG NPs, approximately 50% of silencing efficiency occurred after only 1 h of incubation, with a slight increase over time. In contrast, PLGA-PEG NPs showed a slower, but also time dependent, transfection efficiency, achieving 10% and 30% of Luciferase knockdown after 1 h and 4 h of contact time, respectively (Fig. 2B). In Fig. S6[†] these results are also reported as relative light units (RLU mg⁻¹ protein) of Luciferase activity. This different behavior could be attributed to the reductive triggered degradation of disulfide linkages between the PLGA

and PEGylated components, leading to subsequent NP instability and release of entrapped siRNA in the cellular environment in a shorter time.

In order to investigate the internalization properties of NPs into cells and to predict the fate of the drug cargoes, we carried out uptake experiments of labeled NPs against A549 cells with subsequent confocal microscopy analysis (Fig. 2C). For these experiments, we employed NPs loaded with a fluorescent CyTM3-siRNA (properties are reported in Table S2[†]). As evident in the images, the uptake of redox responsive NPs as well as the internalization of the siRNA (red) was higher as compared to non bioreducible NPs. This effect could be attributed to a partial extracellular responsiveness of the reducible linkage between PLGA and PEG blocks, as previously demonstrated²⁵ and in line with some other studies.^{40,41} We subsequently probed the intracellular trafficking of NPs and the eventual endosomal escape facilitated by the cationic OBAE: A549 cells were incubated for 4 h with CyTM3-siRNA-NPs and then the cell nuclei and endosome were labeled with Hoechst 33258 (blue) and LysoTracker DND-26 (green), respectively. As evident in Fig. 3, we did not observe any overlapping of the red and green colors for both formulations, thus suggesting that the majority of NPs or of the released siRNA escaped from the lysosomes to the cytoplasm.



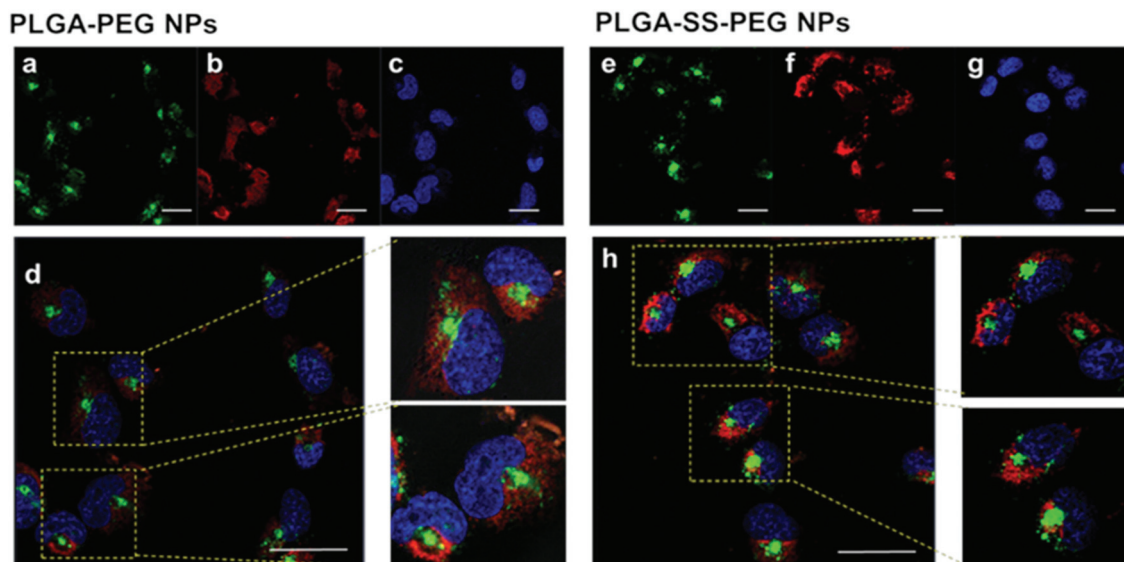


Fig. 3 Intracellular trafficking of NPs after 4 h of incubation. Plates (a–d) show PLGA-PEG NPs and plates (e–h) show PLGA-SS-PEG NPs. (a and e) Lysotracker emission in green (λ_{ex} : 488 nm; λ_{em} : SP 555), (b and f) Cy3-siRNA emission in red (λ_{ex} : 555 nm; λ_{em} : LP 560), (c and g) nucleus stained with Hoechst 33258. Images (d) and (h) show merged channels with enlarged sections contrast-enhanced for clarity. Scale bar: 20 μm . Zen 2009 image Software was utilized for image processing.

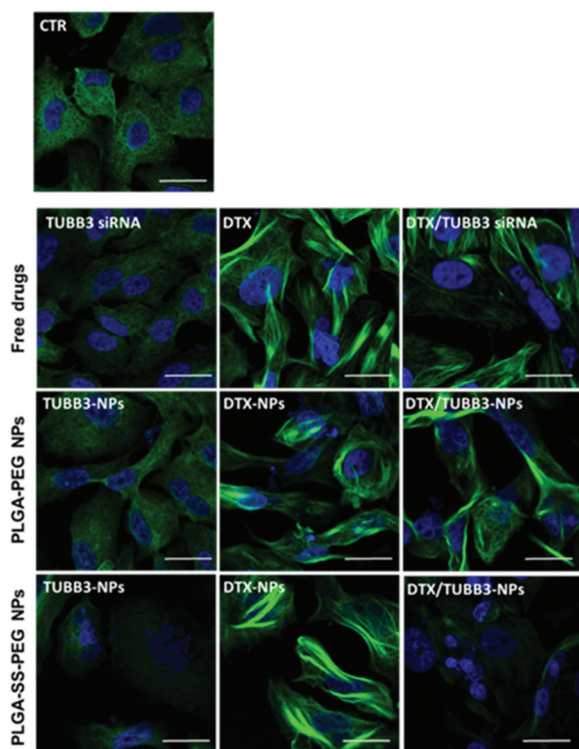


Fig. 4 Immunofluorescence microscopy of A549 cells treated with NPs for 72 h. β III-tubulin was stained in green whereas nuclei were stained with Hoechst 33258 in blue. Scale bar: 10 μm . Zen 2009 image Software was utilized for image processing.

Cell metabolic activity of TUBB3 siRNA/DTX-loaded NPs

To assess the effects of DTX and of β III-tubulin knockdown on microtubule organization, immunofluorescence staining of β III-tubulin was performed in LC cells after 72 h of NP treatment. As evident in Fig. 4 and Fig. S7,[†] untreated cells (the controls of the experiments) had an intact microtubule cytoskeleton. It is well established that microtubule bundle formation is a hallmark of taxane exposure. The dynamic reorganization of microtubules after treatment with docetaxel or paclitaxel causes the inhibition of major cellular events, and apoptosis has generally been accepted to be the predominant mechanism of cell death in response to taxane chemotherapy due to the activation of a ‘mitotic catastrophe’ by scoring of micronucleated cells and cells undergoing aberrant mitosis. In CLSM images, the treatment of cells with DTX, in free form or delivered from NPs, induced strong microtubule bundle formation in a manner not significantly different between the delivery system employed. Concerning the β III-tubulin knockdown, TUBB3 siRNA in free form or delivered from PLGA-PEG NPs did not cause any significant modification of the microtubule networks, whereas a marked decrease of β III-tubulin expression was evident from siRNA delivered from redox responsive NPs (Fig. 4). These results were in line with the metabolic activity of cells after NP treatment (Fig. 5) and the siRNA transfection properties previously discussed.

Concerning anticancer activity and effect of co-delivery, the *in vitro* cytotoxicity of NPs loaded with DTX, with or without TUBB3 siRNA, was evaluated. These MTS assays were carried out after different time exposures (24 h and 72 h), in comparison to free drugs alone or in combination.



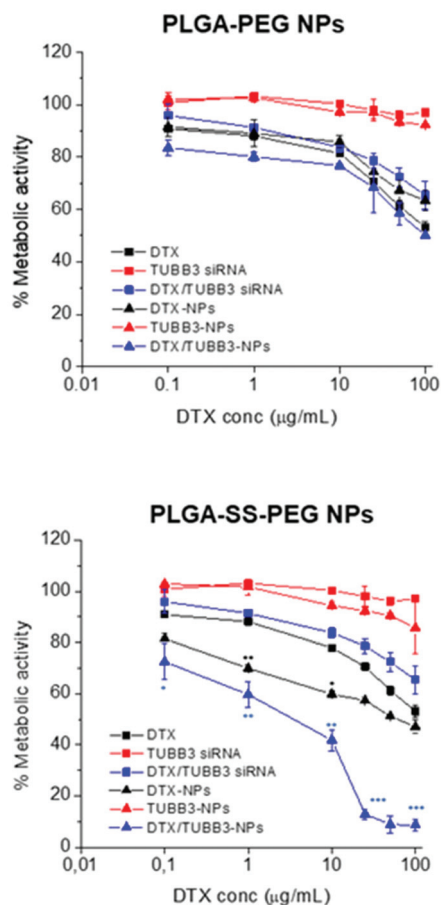


Fig. 5 Cell metabolic activity of PLGA-PEG NPs and PLGA-SS-PEG NPs toward A549 cells after 72 h of incubation. Results are expressed as mean \pm SD of three experiments. *** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$ free DTX, two-way ANOVA test.

In Fig. 5 and S8,[†] the cytotoxicity as interpreted from the loss in metabolic activity against A549 cells is reported (DTX concentration range: 0.1–100 $\mu\text{g mL}^{-1}$; TUBB3-siRNA concentration range: 1–1000 ng mL^{-1}). As expected, DTX, both in free form as well as entrapped into NPs, exhibited a time- and dose-dependent cytotoxicity due to its well-known antimetabolic mechanism of action. Furthermore, neither PLGA-PEG and PLGA-SS-PEG NPs loaded with DTX induced statistically significant differences in cell growth inhibition when compared to free drug ($P = 0.20$).

The IC_{50} values of DTX against A549 cells at 72 h were calculated to be around 102, 132 and 79 $\mu\text{g mL}^{-1}$, for free DTX, PLGA-PEG NPs and PLGA-SS-PEG NPs, respectively, with potentiating factor values (PF) lower than 1 (Table S3[†]). A similar trend was observed in H1299 cells (DTX concentration range: 0.0001–1 $\mu\text{g mL}^{-1}$; TUBB3-siRNA concentration range: 0.001–10 ng mL^{-1}) (Fig. S9[†]). The fact that DTX-NPs showed cytotoxicity similar to that of free DTX at the same dose is perhaps surprising, considering that the NPs were not expected to release all of the entrapped drug over the experimental timeframe. However, as previously reported, it can be

speculated that this effect may be related to the differing mechanisms by which DTX entered the cancer cells when encapsulated in NPs. Thus endocytic transport of NPs would be expected to reduce the interaction of DTX with extracellular proteins in the cell culture media, and also bypass any membrane-associated DTX efflux pumps.^{42–44} In addition, the encapsulation and co-delivery of TUBB3-siRNA into bioresponsive NPs appeared to potentiate the anti-proliferative activity of DTX, especially in A549 cells which were otherwise less sensitive to DTX activity. Specifically, the IC_{50} values of PLGA-SS-PEG NPs notably decreased after 72 h (6.5 $\mu\text{g mL}^{-1}$ in A549 cells), reaching PF values of 16 and 12 vs. free DTX and the corresponding bioreducible formulation loaded with only DTX, respectively. This different potentiation compared to non-responsive NPs indicated a clear DTX/TUBB3 synergistic activity, in line with the much higher efficacy of this formulation.

In vivo assays to assess potential of NPs for drug delivery after inhalation

On the basis of the above encouraging data for delivery and efficacy in cancer cell lines, we moved to preliminary *in vivo* studies. The aim of these experiments was to evaluate if the developed NPs were able to exert their silencing effect following pulmonary administration. To facilitate the study, we decided to look at an orthotopic model of lung cancer in mice established with luciferase-expressing cancer cells. siRNAs for the knock-down of luciferase activity are also readily available. These assays were expected to yield the key proof-of-principle that a biological target could be targeted by the NPs in an *in vivo* cancer setting, but without the complexities involved in evaluating the overall tumor response. In particular, since the combination of DTX and anti-TUBB3 therapies in NP formulations would require significant number of controls (single drugs NPs, free drugs, combinations) and thus of animals, it was essential for ethical reasons to carry out pilot experiments in as few mice as possible. Therefore, we applied anti-luc siRNA-containing NPs *via* intra-tracheal administration in an A549-luc bioluminescent orthotopic LC model, and assessed tolerability and biodistribution of the formulations, as well as transfection efficiency in the knock-down of luciferase.

Tumors spontaneously grew in the lungs following intravenous injection of A549-luc tumor cells in the mice. Tumor growth and the effects of treatments were monitored by whole body *in vivo* bioluminescent imaging (IVIS) and confirmed by X-ray imaging (Fig. S10[†]). Data are presented in terms of mean tumor bioluminescent intensity (BLI) and typical IVIS images from one animal representative of each treatment group at each time point. Prior to the treatment with therapeutic NPs, acute tolerability of NPs loaded with a scrambled siRNA and without DTX was assessed in tumor bearing mice (three mice per treatment under dosing as per the therapy study for 14 days, followed by cessation of dosing and observation for a further 14 days). The properties of the NPs used in these experiments are reported in Table S4,[†] and as control a free



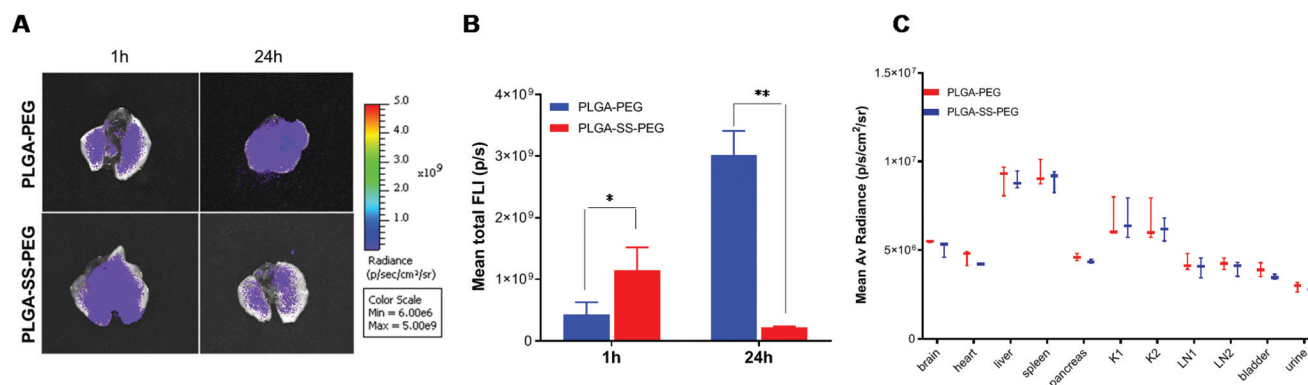


Fig. 6 (A) *Ex vivo* fluorescence images and (B) quantification of the mean fluorescent intensity of DiR in excised lungs of the A549-luc tumor-bearing mice after 1 h and 24 h of treatment with DiR-loaded NPs; (C) quantitative presentation of organ biodistribution *ex vivo* at 24 h.

scrambled siRNA at the same concentration was used. No adverse effects were noted in the mice (data not shown).

To assess NP biodistribution after intratracheal administration, tumor-bearing mice were randomized in 2 groups of 3 mice ($n = 6$). NPs loaded with the red fluorescent model drug DiR (DiR/PLGA-PEG NPs and DiR/PLGA-SS-PEG) were administered intratracheally and fluorescence images were collected live after 1 h and 24 h as well as after animal sacrifice by IVIS. The accumulation of DiR into the tumors after 1 h was higher in the case of PLGA-SS-PEG NPs with a more symmetrical distribution in the lungs (Fig. 6A and B). However, after 24 h, DiR fluorescence into the lungs strongly decreased compared to the control formulation, probably due to the redox responsiveness of the system that favours DiR release in the reductive tumor environment. These results are confirmed also by *ex vivo* analysis of the organs from sacrificed mice (Fig. 6C). After 24 h of treatment with NPs, DiR fluorescence was retained in the lungs (Fig. S11[†]), but fluorescence was found

also in other organs (*e.g.* kidney and brain) only in the case of PLGA-SS-PEG formulation, which suggests again that DiR release from redox-responsive NPs was higher than that from PLGA-PEG NPs (Fig. S12[†]), as observed in the *in vitro* study.

Finally, to evaluate the ability of the NPs to deliver siRNA into the lung, gene silencing efficacy studies were carried out. Two weeks after tumor initiation, mice were treated with NPs loaded with a siRNA against Luciferase (antiLuc-siRNA) or with a scrambled siRNA (ctr-NPs). Mice were dosed again intratracheally and after 72 h, luminescence of the tumors was monitored *in vivo* as well as *ex vivo*. A reduction of lung bioluminescence after treatment with PLGA-SS-PEG and PLGA-PEG NPs (Fig. 7A). Moreover, the effect of anti-Luc-siRNA NPs was confirmed through the analysis of excised lungs after 72 h. The data showed that bioluminescence of the lungs strongly decreased compared to those treated with NPs loaded with a scrambled siRNA as control (Fig. 7B), thus demonstrating the efficacy of this system to reduce the lung tumor.

These preliminary data established the principle of delivering functional payloads to the lungs *via* combined PLGA-SS-PEG and OBAE NPs. The use of DiR as a fluorescent payload indirectly evaluated the efficiency of the NPs to release a small hydrophobic molecule such as DTX. Fluorescence from DiR was observed throughout the lungs within 1 h from both PLGA-PEG and the PLGA-SS-PEG formulations, indicating successful transport of the formulations, while the more rapid loss of the DiR signal in the lungs from the PLGA-SS-PEG NPs compared to the non-redox responsive variant implied a faster release of the payload across the lungs and into systemic circulation. The end-fate organ accumulation data showed that DiR was present mostly in the liver and kidneys at 24 h, as expected for small molecule transport, but were also indicative that for subsequent experiments relating to DTX delivery, some formulation tuning would be required to sustain release in the lungs and delay passage through to systemic circulation. For the siRNA delivery assays, the data showed effective knockdown of luciferase from the constitutively expressing A549 luc cells after 72 h for both formulations.

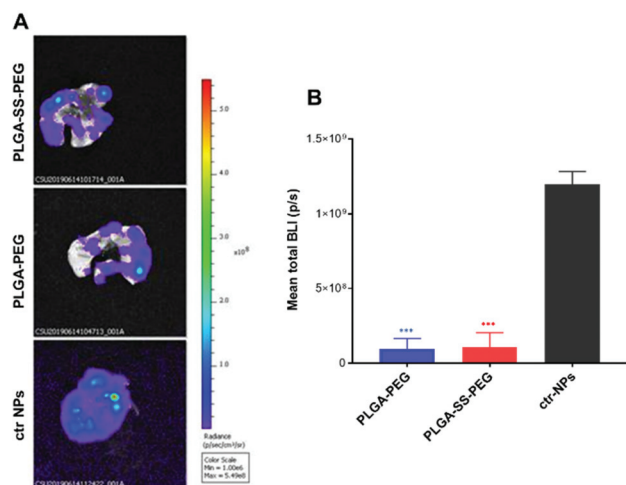


Fig. 7 (A) *Ex vivo* bioluminescence images and (B) quantitative luminescence of excised lungs of the A549-luc tumor-bearing mice after 72 h of treatment with NPs loaded with a siRNA against Luciferase. *** $P < 0.001$ vs. ctr-NPs, two-way ANOVA test.



When considered together, cell culture and preliminary *in vivo* experiments demonstrated both the promise and complexity in using combined NPs. The cell data clearly demonstrated a combined effect of DTX and TUBB3 siRNA in LC cells, and indicated that the PLGA-SS-PEG redox-responsive formulation was the most effective. For the orthotopic lung cancers in mice, successful delivery of both a small molecule dye and a siRNA was demonstrated, with effective knockdown of luciferase showing that NPs were able to cross tissue and intracellular barriers to release an active biological cargo. However, the differences in the formulations were less apparent in the *in vivo* model, which may have related to the intricacies in transport of small and large molecules in lung tissue, and the impact of redox-responsiveness on dye/drug effective dose accumulating in cancer cells. There is an imperative to evaluate differential organ accumulation of these drugs when delivered *in vivo* as well as to establish routes to control release rates for each therapeutic from combination NPs. These questions are now under evaluation and will be reported subsequently.

Conclusions

Here we have reported novel redox-responsive PEGylated NPs, with the aim to deliver simultaneously into the lung the conventional anticancer drug docetaxel and a therapeutic siRNA against β III-tubulin (TUBB3 siRNA). Both drugs were loaded inside the carrier and showed a complete and fast release at simulated intracellular levels of reductive agents. The redox-responsiveness of these systems led to higher silencing efficiency of delivered siRNA and cell uptake, in comparison to non-redox-responsive variant. Concerning anticancer activity, the co-delivery of TUBB3 siRNA through bioresponsive nanoparticles strongly increased DTX cytotoxicity in lung cancer cells, thus highlighting the effective combined anticancer activity of the compounds. *In vivo*, these systems were well tolerated after intratracheal administration in a bioluminescent A549-luc lung cancer model. The suitability of the system for inhalation delivery was also demonstrated, with data showing that NPs reached the lungs where an anti-Luc siRNA was released and exerted its silencing activity. These results indicate that combining DTX with TUBB3 siRNA in redox-responsive NPs as we have shown here could lead to promising candidates for the local and combined therapy of lung cancer with a specific final goal of reducing drug resistance related to the current clinically used therapeutics.

Experimental

Materials

All solvents were of analytical or HPLC grade and purchased from Sigma or Fisher Scientific unless otherwise stated.

Deuterated solvents, the two monomers glycolide (GL) and DL-lactide (LA) were from Sigma. Poly(ethylene glycol) methyl ether (mPEG) and α -methoxy- ω -amino poly(ethylene glycol) (mPEG-NH₂) with $M_n = 2.0$ kDa were purchased from Iris Biotech GmbH (Germany). Dialysis membrane (MWCO 3500, regenerated cellulose) was used as received from Spectrapor. Tin(II)-2-ethylhexanoate (SnOct₂), acryloyl chloride, ethylenedioxy-bis-ethylamine, triethylamine (TEA), dithiodiethanol, Tris-Acetate-EDTA (TAE) buffer, poly(vinyl alcohol) (PVA, Mowiol®40-88, $M_w \sim 31\,000$), sodium chloride, potassium phosphate dibasic and potassium phosphate monobasic, sodium azide, magnesium chloride, potassium chloride, sodium phosphate dibasic, sodium sulfate, calcium chloride dehydrated, sodium acetate, sodium bicarbonate, sodium citrate dehydrate, ethidium bromide, Hoechst 33258, BCA reagent, glutathione (GSH), were used as received from Sigma Aldrich. Docetaxel (DTX, MW = 807.88) was purchased from LC laboratories (USA). 1,1'-Dioctadecyl-3,3,3',3'-tetramethylindotricarbocyanine Iodide (DiR) was acquired by Thermo Fisher Scientific (UK). siRNA against Luciferase (Luc-siRNA) (CCGCAAGAUCGCGAGAAUU), scrambled siRNA, Silencer®CyTM3 labeled Negative Control (λ_{ex} max = 547, λ_{em} = 563 nm) and siRNA against β III-tubulin (TUBB3-siRNA) (GUACGUGCCUCGAGCCAUUUU) were provided by Eurogentec (UK). Luciferase Assay System and CellTiter 96® Aqueous One Solution Cell Proliferation Assay (MTS) were provided by Promega (UK).

Synthesis of PLGA block copolymers and OBAE

Synthesis of PLGA-PEG block copolymer. PLGA-PEG diblock copolymer was prepared as previously described.²⁵ Briefly, mPEG₂₀₀₀-OH and Sn(Oct)₂ were used as initiator and catalyst, respectively, in the bulk ring-opening polymerization (ROP) of DL-lactide (LA) and glycolide (GL) at 130 °C; LA/GL/initiator molar ratio = 25 ($M_n = 8.3$ kDa at 25 °C in chloroform; $D = 1.2$; yield = 86%).

Synthesis of PLGA-SS-PEG block copolymer. A redox responsive PLGA-SS-PEG diblock copolymer was prepared as previously described²⁵ via a three-step reaction, *i.e.* the synthesis of 2-((hydroxymethyl)disulfanyl)ethyl acrylate (SS), (2) the synthesis of a PLGA-S-S-ethyl acrylate (PLGA-SS) and (3) the Michael addition reaction between PLGA-SS and mPEG₂₀₀₀-NH₂. Briefly, the ROP initiator 2-((hydroxymethyl)disulfanyl)ethyl acrylate (2HMDEA) was prepared by reacting dithiodiethanol with acryloyl chloride through a published method with minor modifications.⁴⁵ Then, 2HMDEA and Sn(Oct)₂ were used as initiator and catalyst, respectively, in the bulk ROP of DL-lactide (LA) and glycolide (GL) at 130 °C; finally, PLGA-SS-PEG diblock copolymer was synthesized by aza-Michael Addition between the amino group of mPEG₂₀₀₀-NH₂ and the acrylic end group of PLGA-SS at 65 °C for 6 days. LA/GL/PEG molar ratio = 25 ($M_n = 9.6$ kDa at 25 °C in chloroform; $D = 1.2$; yield = 70%).

Synthesis of OBAE. The synthesis of the oligo- β -aminoester (OBAE) was *via* a two-step reaction, *i.e.* the synthesis of disulfide-based diacrylate (DSD) and (2) the Michael addition



reaction between DSD and ethylenedioxy-bis-ethylamine. Synthesis of DSD was carried out through a literature method with minor modifications.⁴⁵ Then, for the Michael Addition reaction, DSD was mixed with ethylenedioxy-bis-ethylamine (molar ratio 1:1.5) in DCM in the dark at 30 °C for 5 days under nitrogen. ($M_n = 2.1$ kDa at 25 °C in DMF; $D = 1.2$; yield = 72%).

¹H- and ¹³C NMR, FTIR spectra, size exclusion chromatography (SEC) measurements and differential scanning calorimetry (DSC) of both PLGA-based copolymers as well as OBAE were performed as previously reported.²⁵

Preparation and characterization of NPs

Preparation of NPs. PLGA-PEG and PLGA-SS-PEG were mixed with OBAE and employed to prepare NPs by a modified emulsion-solvent evaporation technique. DTX as lipophilic anti-cancer drug model and a siRNA (scrambled or against β III-tubulin) were loaded into NPs at 10% w/w and 0.1% w/w, respectively. Briefly, 1 mg of OBAE was solubilized in 0.1 mL of EtOH and then mixed to a DCM solution (0.9 mL) containing the PLGA-based copolymer (10 mg) and DTX (1 mg). Thereafter, 0.1 mL of an internal water phase containing 10 μ g of siRNA (0.8 nmol) were added to the organic phase and the mixture was sonicated for 1 min at 3 W (Ultrasonic Sonopuls HD 3200, Bandelin, UK) using a microtip probe. This first emulsion was poured in water (10 mL) in the presence of PVA 0.5% w/v, and sonicated again for 5 min at 3 W. Thereafter, the organic solvent was evaporated by mechanical stirring (300 rpm) for 3 h at r. t. and then filtered (0.45 μ m). NPs were then collected by centrifugation (14 000 rpm, 30 min, 4 °C) after washing twice with distilled water to remove the excess of PVA and the unloaded drugs. Finally, NPs were re-dispersed in water (1 mL), freeze-dried for 24 h and kept at 4 °C. Unloaded and NPs loaded with a single drug were also prepared as controls. Recovery yield of production process was evaluated by weighing the solid residue after freeze-drying. Results are expressed as the ratio of the actual NPs mass to the theoretical polymer mass $\times 100$.

Colloidal properties of NPs. Hydrodynamic diameters (D_H), polydispersity indices (PDI) and zeta potentials (ζ) of NPs were determined on a Zetasizer Nano ZS (Malvern Instruments Ltd). Results are reported as mean of three separate measurements of three different batches ($n = 9$) \pm standard deviation (SD). The morphology of the NPs was examined by transmission electron microscopy (TEM). Samples were imaged using a JEOL 2100F FEG-TEM electron microscope. A few drops were added onto a copper grid and allowed to dry in air. NPs were negatively stained with 2% phosphotungstic acid solution.

DTX and siRNA loading. DTX loading inside NPs was assessed by dissolving 1 mg of freeze-dried NPs in 1 mL of THF under stirring for 1 h. Thereafter the sample was filtered through a 0.22 μ m filter and analyzed for DTX content by HPLC on a Shimadzu apparatus equipped with a LC-10ADvp pump, a SIL-10ADvp autoinjector, a SPD-10Avp UV-Vis detector and a C-R6 integrator. The analysis was performed on a ACE 5 μ m, C18 column (250 \times 4.6 mm, Å) (Phenomenex, USA). The

mobile phase was a 40:60 (v/v) mixture of 20 mM phosphate buffer at pH 4.5 and acetonitrile pumped at a flow rate of 1 mL min⁻¹. The UV detector was set at 227 nm. A calibration curve for DTX in THF was constructed in the concentration range 1–200 μ g mL⁻¹.

The loading of siRNA was confirmed by agarose gel retardation assays. 1 mg of NPs containing 1 μ g of siRNA were loaded on 2% agarose gel in Tris-Acetate-EDTA (TAE) buffer and subjected to electrophoresis for 45 min at 70 V. siRNA bands were stained with ethidium bromide and finally visualized with an UV illuminator. The entrapment efficiency of siRNA was assessed by dissolving 1 mg of freeze dried NPs in 500 μ L of DCM under stirring for 1 h. Thereafter, 500 μ L of water were added and the sample stirred for 1 h further. After centrifugation at 5000 rpm for 10 min, the aqueous phase was collected and analysed by UV spectrophotometry (Nanodrop 2000 spectrophotometer, Thermo Scientific, UK) at 290 nm. Potential interference of the polymers and DTX on siRNA absorbance was preliminarily assessed spiking a siRNA solution in water with different amount of DTX-loaded NPs.

Redox responsiveness of NPs. To determine the stability of NPs under intracellular reductive conditions, gel retardation assay and size measurements of NPs dispersed in PBS pH 7.4 (1.5 mg mL⁻¹) enriched with GSH 10 mM were taken after incubation at 37 °C for 30 min.

In vitro release of DTX in PBS with or without GSH 10 mM was investigated by a dialysis method. 2 mg of NPs were dispersed in 0.5 mL of PBS solution and placed in a dialysis bag (MWCO = 3500 Da, Spectra/Por®). The sample was plunged in 8 mL of PBS (sink condition) and kept at 37 °C. At selected time intervals, 1 mL of release medium was withdrawn and replaced with an equal volume of fresh medium. DTX quantitative analysis was performed by HPLC as described above. Release profile of the free drug is reported for comparison. Results are expressed as release% over time \pm SD of three experiments.

For siRNA release studies, 500 μ L of NP dispersions were diluted with PBS at pH 7.4 with or without GSH 20 mM to a total volume of 1 mL and incubated at 37 °C. At different time points aliquots of 500 μ L of NP dispersions were collected and centrifuged at 16 000g for 30 min at 4 °C. The supernatants were collected and analysed at 280 nm by UV-vis spectrometry (NanoDrop™ 2000). The values were calculated as mean \pm SD of three experiments.

NP stability in protein-rich media. To determine the stability of NPs in simulated biological conditions, a known amount of NPs (0.5 mg) was dispersed in PBS 10 mM pH 7.4 with or without Fetal Bovine Serum at 10% and incubated at 37 °C for different times. Size measurements of the samples were taken by dynamic light scattering on a Zetasizer along time up to 48 h of incubation.

Cell studies

Cell culture. H1299 and A549 lung cancer cell lines were purchased from the American Type Culture Collection (USA). H1299 cells were cultured in RPMI-1640 medium supplemented with 10% v/v fetal bovine serum (FBS), 1.5 mM



L-glutamine, 100 units mL^{-1} penicillin, and 100 $\mu\text{g mL}^{-1}$ streptomycin under humidified atmosphere of 5% CO_2 at 37 °C. A549 cells were cultured in Dulbecco's Modified Eagle's Medium (DMEM), supplemented as above.

Effects of OBAAE on cell metabolic activity. The cytotoxicity of cationic OBAAE entrapped inside NPs was inferred through the MTS assay. Cells (2×10^4) were placed in 96-well plates and cultured in 200 μL of cell medium with FBS at 10%. After 24 h, cells were treated with unloaded NPs (from 0.05 to 5 mg mL^{-1}) and free OBAAE in the same concentration range theoretically entrapped into NPs (0.005–0.5 mg mL^{-1}). Cells treated with 0.1% (v/v) Triton-X 100 and fresh media were used as a positive and a negative control, respectively. After different incubation times up to 24 h, cells were washed with PBS and treated with CellTiter 96® Aqueous One Solution Cell Proliferation Assay (20 μL per well). After further incubation (3 h), the absorbance was read at 490 nm in a microplate reader (Tecan Platereader).

Transfection studies. For transfection studies, A549 luciferase expressing cells (5×10^4) were seeded into 24-well plates and cultured in 500 μL of cell medium with FBS at 10%. After 24 h, the culture medium was replaced with 0.5 mL of fresh medium and treated with NPs loaded with a siRNA against luciferase (1 μg of Luc-siRNA per well). After different incubation times (1 h and 4 h), cells were washed in order to remove NPs and incubated again at 37 °C until 48 h. Finally, after transfection, luciferase activity was measured as RLU mg^{-1} protein using a luciferase assay system and BCA reagent.

Uptake and intracellular trafficking of NPs. A549 cells were used to assess the internalization and the intracellular trafficking of NPs loaded with a fluorescent CyTM3-siRNA (0.1% w/w). Cells were seeded into 24-well/plates (2×10^5 cells per well) in 0.5 mL of DMEM with 10% FBS. After 24 h, cells were washed and incubated for 4 h with 125 $\mu\text{g mL}^{-1}$ of NPs, freshly re-suspended in milli Q water and diluted in medium without 10% of FBS. After incubation with NPs, the cells were washed and fixed in ice-cold PBS with 2% *p*-formaldehyde for 10 min followed by adding Hoechst 33258 (1 $\mu\text{g mL}^{-1}$) and LysoTracker Green (100 nM). Red fluorescence of CyTM3-siRNA, blue fluorescence of Hoechst 33258 and green fluorescence of LysoTracker Green were observed using a Zeiss LSM 700 Confocal Laser Scanning Microscope (CLSM) equipped with Argon (488 nm) and HeNe (561, 639 nm) lasers and a 40X/1.2 NA water objective. Images were acquired with λ_{ex} 555 nm and spectral filters of LP 560 nm for CyTM3-siRNA detection. Zen 2009 image Software was utilized for image processing.

Immunofluorescence. Cells (1.5×10^5) were placed in 24-well plates and cultured in 0.5 mL of cell medium with FBS at 10%. After 24 h, cells were treated with free drugs, DTX-NPs, TUBB3-NPs and TUBB3/DTX-NPs. DTX and siRNA concentrations were respectively 2.5 $\mu\text{g mL}^{-1}$ and 0.25 $\mu\text{g mL}^{-1}$ for H1299 cells, 25 $\mu\text{g mL}^{-1}$ and 2.5 $\mu\text{g mL}^{-1}$ for A549 cells. After 72 h of treatment, cells were washed and fixed in ice-cold PBS with 2% *p*-formaldehyde for 10 min. Then, cells were permeabilized with TritonX-100 (0.1%) in PBS for 20 min at r.t. and treated with an Anti-III-tubulin human antibody (Abcam, UK) followed by the secondary antibody Rhodamine Red goat anti-mouse

IgG (Invitrogen, UK). Finally, nuclei were stained with Hoechst 33258 (1 $\mu\text{g mL}^{-1}$) for an additional 15 min. Cells were observed by CLSM.

Cell metabolic activity of TUBB3 siRNA/DTX-loaded NPs. Cells (2×10^4) were placed in 96-well plates and cultured in 0.2 mL of cell medium with FBS at 10%. After 24 h, cells were treated with DTX-NPs (NPs loaded with DTX), TUBB3-NPs (NPs loaded with TUBB3-siRNA), TUBB3/DTX-NPs (NPs loaded with both drugs) and free drugs alone and in combination. DTX and siRNA concentration ranges were set respectively at 0.0001–1 $\mu\text{g mL}^{-1}$ and 0.001–10 ng mL^{-1} for H1299 cells, and at 0.1–100 $\mu\text{g mL}^{-1}$ and 1–1000 ng mL^{-1} for A549 cell line. DMSO 0.1% (v/v) was used as vehicle for free DTX. After 24 h and 72 h, cell viability was assessed through MTS assay as described above.

Statistical analysis. Experimental data were expressed as means \pm standard deviation (SD). Each value was the mean of at least three repeat experiments in each group. Normal distribution was assumed and data therefore tested using parametric tests. The statistical significance between two groups was analyzed by Student *t*-test. Differences with $P < 0.05$ (*) or $P < 0.01$ (**) were considered statistically significant. The difference between three or more groups was analyzed by one-way analysis of variance (ANOVA) multiple comparisons.

In vivo studies

Animals. Animal studies were conducted under the UK Home Office Licence (PPL P435A9CF8 protocol 11), NCRI guidelines for the welfare and use of animals in cancer research, LASA good practice guidelines and FELASA working group on pain and distress guidelines. All animal procedures were performed in accordance with the Guidelines for Care and Use of Laboratory Animals of Nottingham University and approved by the University's Animal Welfare and Ethical Review Body. 32 female CD-1 Nu mice were purchased from Charles River Laboratories (UK) and acclimatized for a week prior to study set up. Mice were maintained in IVCs (Tecniplast, UK) within a barrier unit illuminated by fluorescent lights set to give a 12 h light–dark cycle (on 07.00, off 19.00), as recommended in the UK Home Office Animals (Scientific Procedures) Act 1986. The room was air-conditioned by a system designed to maintain an air temperature range of 21 ± 2 °C and a humidity of 55% + 10%. Mice were housed in social groups during the procedure with irradiated bedding and provided with autoclaved nesting materials and environmental enrichment. Sterile irradiated 5 V5R rodent diet (IPS Ltd, UK) and autoclaved water was offered *ad libitum*.

Tumor establishment and dosing. Tumors in the mice were initiated by an approved S1 method as follows. A549-luc tumor cells were suspended in a standard formulation of Matrigel (2×10^6 cells) and the cell suspension was drawn into a 1 mL syringe. An aliquot (50 μL) of the cell suspension was then injected intravenously into the tail vein of non-anaesthetised mice, after warming the animal in a purpose-built thermostatically controlled warming unit. Tumor establishment and growth was monitored by bioluminescence imaging to assess



tumor burden prior to recruitment into the different studies through an IVIS Spectrum weekly, and at termination, to provide an optical post mortem. Tumor measurements and 3D image reconstruction were made using Living Image (4.3.1 build) software (Caliper Life Sciences Hopkinton, MA, USA). To quantify the fluorescence intensity, a fluorescence calibrated unit (termed radiant efficiency), representing the fractional ratio of emitted photons per incident excitation photon, was used to compensate for non-uniform excitation light patterns.

Dosing was initiated in established tumors (mice six to seven weeks old), those whose volume was demonstrably larger than the original implantation volume. Mice were anesthetized with 10 ml g⁻¹ ketamine/medetomidine (Ket 0.9 ml/Met 0.3 ml/NaCl 8.8 ml), restrained on a support ring and dosed intratracheally by a Microsprayer Aerosolizer (Model IA-IC, PennCentury, Philadelphia, PA, USA) (50 µL per dose). The administration was guided by a pediatric otoscope. All animals were sacrificed when tumor volume reached the maximum permissible size of 1.5 cm diameter in at least one mouse.

Tolerability study. In order to check the safety of the carrier, tolerability studies on tumor-bearing mice were performed. NPs loaded with a scrambled siRNA were intratracheally administered (50 µL of 10 mg mL⁻¹ NP dispersion) by the Microsprayer Aerosolizer. Survival and tumor growth was monitored up to 21 days after administration.

Biodistribution studies. The biodistribution of NPs in tumor-bearing mice was assessed employing NPs loaded with DiR red fluorescent dye (1% w/w). Mice were dosed intratracheally with DiR/PLGA-PEG NPs or DiR/PLGA-SS-PEG NPs (3 mice per group). Each dose contained 10 µg of DiR in 50 µL. The evaluation of biodistribution was carried out at 1 h and 24 h term. Imaging of mice on front and back was carried out in the IVIS Spectrum with parameters chosen for DiR detection and the closest red-shifted filter set. Imaging filters were selected from 710/780 (plus 430_{ex} controls). A 'blue light' control filter set was also used to provide auto-fluorescence data for un-mixing and image analysis. For the lung IVIS analysis, mice were culled by an approved S1 method post initial imaging, while still under anesthetic; all mice organs were dissected and imaged (kidney, 25 µl urine, liver, spleen/pancreas, lung, heart, bladder, brain, lymph nodes-inguinal). The *ex vivo* fluorescent images of the major organs were also detected with the same imaging system as described above.

siRNA gene knockdown efficacy studies. At a time period of 2 weeks after tumour initiation, mice were treated with fluorescent NPs loaded with DiR (1% w/w) and a Luc-siRNA (0.1% w/w). Mice were randomly allocated to different groups: (1) Luc-siRNA-DiR/PLGA-PEG NPs (2) Luc-siRNA-DiR/PLGA-SS-PEG NPs and (3) scrambled-siRNA-DiR/PLGA-SS-PEG NPs (*n* = 9). Mice were intratracheally dosed (5 µg of siRNA in 50 µl per dose) as described above. At 1, 24 and 72 h after treatment, DiR distribution was monitored *via* fluorescence imaging, whereas tumors by bioluminescent imaging. The signal intensity of tumors was quantified as the sum of all detected photon counts within the ROI. At 72 h the mice were sacrificed and the resected lungs imaged.

Statistical analysis. GraphPad Prism 7 software (La Jolla, CA, USA) was used for data analysis. Comparisons between two groups were made using independent samples Student's *t*-test and one way analysis of variance (ANOVA) was applied for the comparison of three or more groups. Tukey's multiple comparison test (*post-hoc*) was used in conjunction with ANOVA to evaluate differences in individual means in EGFR staining data. For other experiments, Bonferroni *post-hoc* test was applied for comparison of group means of three or more groups. *p* values of <.05 were considered statistically significant. *, **, ***, and **** indicate *p* < 0.05, *p* < 0.01, *p* < 0.001, and *p* < 0.0001, respectively.

Data access statement

All relevant data are available from the University of Nottingham's Research Data Management Repository or *via* email from the corresponding authors.

Conflicts of interest

There are no conflicts to declare.

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References

- 1 L. A. Torre, R. L. Siegel and A. Jemal, *Adv. Exp. Med. Biol.*, 2016, **893**, 1–19.
- 2 R. Bayat Mokhtari, T. S. Homayouni, N. Baluch, E. Morgatskaya, S. Kumar, B. Das and H. Yeger, *Oncotarget*, 2017, **8**, 38022–38043.
- 3 M. J. Smyth, S. F. Ngiow, A. Ribas and M. W. Teng, *Nat. Rev. Clin. Oncol.*, 2016, **13**, 143–158.



- 4 B. Mansoori, S. Sandoghchian Shotorbani and B. Baradaran, *Adv. Pharm. Bull.*, 2014, **4**, 313–321.
- 5 N. S. Gandhi, R. K. Tekade and M. B. Chougule, *J. Controlled Release*, 2014, **194**, 238–256.
- 6 C. Chakraborty, A. R. Sharma, G. Sharma, C. G. P. Doss and S. S. Lee, *Mol. Ther.–Nucleic Acids*, 2017, **8**, 132–143.
- 7 C. Bouillon, Y. Bessin, F. Poncet, M. Gary-Bobo, P. Dumy, M. Barboiu, N. Bettache and S. Ulrich, *J. Mater. Chem. B*, 2018, **6**, 7239–7246.
- 8 F. Freitag and E. Wagner, *Adv. Drug Delivery Rev.*, 2020, **168**, 30–54.
- 9 W. Viricel, S. Poirier, A. Mbarek, R. M. Derbali, G. Mayer and J. Leblond, *Nanoscale*, 2017, **9**, 31–36.
- 10 R. Kandil, Y. Xie, R. Heermann, L. Isert, K. Jung, A. Mehta and O. M. Merkel, *Adv. Ther.*, 2019, **2**, 1900047.
- 11 L. J. Li, W. Sun, L. Li, Y. Y. Liu, L. Wu, F. L. Wang, Z. Zhou, Z. R. Zhang and Y. Huang, *Nanoscale*, 2017, **9**, 314–325.
- 12 J. Luo, E. Wagner and Y. Wang, *J. Mater. Chem. B*, 2020, **8**, 2020–2031.
- 13 S. Baek, R. K. Singh, D. Khanal, K. D. Patel, E. J. Lee, K. W. Leong, W. Chrzanowski and H. W. Kim, *Nanoscale*, 2015, **7**, 14191–14216.
- 14 F. Saviano, T. Lovato, A. Russo, G. Russo, C. R. Bouton, R. J. Shattock, C. Alexander, F. Quaglia, A. K. Blakney, P. Gurnani and C. Conte, *J. Mater. Chem. B*, 2020, **8**, 4940–4949.
- 15 W. J. Huang, X. X. Wang, C. R. Wang, L. L. Du, J. H. Zhang, L. D. Deng, H. Q. Cao and A. J. Dong, *J. Mater. Chem. B*, 2019, **7**, 965–973.
- 16 E. Blanco, H. Shen and M. Ferrari, *Nat. Biotechnol.*, 2015, **33**, 941–951.
- 17 S. Ganta, H. Devalapally, A. Shahiwala and M. Amiji, *J. Controlled Release*, 2008, **126**, 187–204.
- 18 S. Mura, J. Nicolas and P. Couvreur, *Nat. Mater.*, 2013, **12**, 991–1003.
- 19 H. Sun, Y. Zhang and Z. Zhong, *Adv. Drug Delivery Rev.*, 2018, **132**, 16–32.
- 20 R. Cheng, F. Feng, F. H. Meng, C. Deng, J. Feijen and Z. Y. Zhong, *J. Controlled Release*, 2011, **152**, 2–12.
- 21 R. Kandil and O. M. Merkel, *Ther. Delivery*, 2019, **10**, 203–206.
- 22 I. d'Angelo, C. Conte, A. Miro, F. Quaglia and F. Ungaro, *Curr. Top. Med. Chem.*, 2015, **15**, 386–400.
- 23 F. Ungaro, I. d'Angelo, A. Miro, M. I. La Rotonda and F. Quaglia, *J. Pharm. Pharmacol.*, 2012, **64**, 1217–1235.
- 24 S. Rezvantlab, N. I. Drude, M. K. Moraveji, N. Guvener, E. K. Koons, Y. Shi, T. Lammers and F. Kiessling, *Front. Pharmacol.*, 2018, **9**, 1260.
- 25 C. Conte, F. Mastrotto, V. Taresco, A. Tchoryk, F. Quaglia, S. Stolnik and C. Alexander, *J. Controlled Release*, 2018, **277**, 126–141.
- 26 J. Karlsson, Y. Rui, K. L. Kozielski, A. L. Placone, O. Choi, S. Y. Tzeng, J. Kim, J. J. Keyes, M. I. Bogorad, K. Gabrielson, H. Guerrero-Cazares, A. Quifiones-Hinojosa, P. C. Searson and J. J. Green, *Nanoscale*, 2019, **11**, 20045–20057.
- 27 T. Lovato, V. Taresco, A. Alazzo, C. Sansone, S. Stolnik, C. Alexander and C. Conte, *J. Mater. Chem. B*, 2018, **6**, 6550–6558.
- 28 K. L. Kozielski and J. J. Green, *Methods Mol. Biol.*, 2016, **1364**, 79–87.
- 29 K. L. Kozielski, S. Y. Tzeng, B. A. De Mendoza and J. J. Green, *ACS Nano*, 2014, **8**, 3232–3241.
- 30 L. M. Ensign, C. Schneider, J. S. Suk, R. Cone and J. Hanes, *Adv. Mater.*, 2012, **24**, 3887–3894.
- 31 C. Dumontet and B. I. Sikic, *J. Clin. Oncol.*, 1999, **17**, 1061–1070.
- 32 P. P. Gan, E. Pasquier and M. Kavallaris, *Cancer Res.*, 2007, **67**, 9356–9363.
- 33 J. A. McCarroll, P. P. Gan, R. B. Erlich, M. Liu, T. Dwarde, S. S. Sagnella, M. C. Akerfeldt, L. Yang, A. L. Parker, M. H. Chang, M. S. Shum, F. L. Byrne and M. Kavallaris, *Cancer Res.*, 2015, **75**, 415–425.
- 34 J. A. McCarroll, P. P. Gan, M. Liu and M. Kavallaris, *Cancer Res.*, 2010, **70**, 4995–5003.
- 35 C. S. Schneider, Q. Xu, N. J. Boylan, J. Chisholm, B. C. Tang, B. S. Schuster, A. Henning, L. M. Ensign, E. Lee, P. Adstamongkonkul, B. W. Simons, S. S. Wang, X. Gong, T. Yu, M. P. Boyle, J. S. Suk and J. Hanes, *Sci. Adv.*, 2017, **3**, e1601556.
- 36 C. Conte, F. Mastrotto, V. Taresco, A. Tchoryk, F. Quaglia, S. Stolnik and C. Alexander, *J. Controlled Release*, 2018, **277**, 126–141.
- 37 H. Y. Xue, S. Liu and H. L. Wong, *Nanomedicine*, 2014, **9**, 295–312.
- 38 M. Breunig, U. Lungwitz, R. Liebl and A. Goepferich, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 14454–14459.
- 39 A. A. Eltoukhy, D. J. Siegwart, C. A. Alabi, J. S. Rajan, R. Langer and D. G. Anderson, *Biomaterials*, 2012, **33**, 3594–3603.
- 40 W. Sun and P. B. Davis, *J. Controlled Release*, 2010, **146**, 118–127.
- 41 S. Y. Lee, J. Y. Tyler, S. Kim, K. Park and J. X. Cheng, *Mol. Pharm.*, 2013, **10**, 3497–3506.
- 42 C. Conte, F. Ungaro, G. Maglio, P. Tirino, G. Siracusano, M. T. Sciortino, N. Leone, G. Palma, A. Barbieri, C. Arra, A. Mazzaglia and F. Quaglia, *J. Controlled Release*, 2013, **167**, 40–52.
- 43 G. Palma, C. Conte, A. Barbieri, S. Bimonte, A. Luciano, D. Rea, F. Ungaro, P. Tirino, F. Quaglia and C. Arra, *Int. J. Pharm.*, 2014, **473**, 55–63.
- 44 F. Ungaro, C. Conte, L. Ostacolo, G. Maglio, A. Barbieri, C. Arra, G. Misso, A. Abbruzzese, M. Caraglia and F. Quaglia, *Nanomedicine*, 2012, **8**, 637–646.
- 45 C. Y. Hong, Y. Z. You, D. C. Wu, Y. Liu and C. Y. Pan, *J. Am. Chem. Soc.*, 2007, **129**, 5354–5355.

