Natural Product Reports

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Needles in haystacks: reevaluating old paradigms for the discovery of bacterial secondary metabolites

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Natural products research is in the midst of a renaissance ushered in by a modern understanding of microbiology and the technological explosions of genomics and metabolomics. As the exploration of uncharted chemical space expands into high-throughput discovery campaigns, it has become increasingly clear how design elements influence success: (bio)geography, habitat, community dynamics, culturing/induction methods, screening methods, dereplication, and more. We explore critical considerations and assumptions in natural products discovery. We revisit previous estimates of chemical rediscovery and discuss their relatedness to study design and producer taxonomy. Through frequency analyses of biosynthetic gene clusters in publicly available genomic data, we highlight phylogenetic biases that influence rediscovery rates. Through selected examples of how study design at each level determines discovery outcomes, we discuss the challenges and opportunities for the future of high-throughput natural product discovery.

Contents

1. Introduction

Bacteria represent an astounding proportion of life's diversity, occupying nearly every terrestrial, marine, and aerial niche yet

investigated. Some grow at extremes of heat and cold, some thrive in concentrations of salts and metals that are toxic to more temperate organisms, and others survive nutrient deprivation and desiccating conditions that thwart most other forms of life. Bacteria seldom live in isolation. Most depend upon other species to meet their mosaic of needs to acquire nutrients, detoxify waste, and be transported to new locations. These co-dependencies develop in complex communities containing a few to thousands of other species (and, in some environments, perhaps orders of magnitude more), which can present microbes with useful collaborations and hostile opposition. Consequently, bacterial fitness is determined in part by success in the elaborate networks that connect microorganisms, macroorganisms, and the environment.

All communities are defined by their members' interactions ranging across a continuum of cooperation to competition. In bacterial communities, secondary metabolites are the currency of many of these interactions: bacteria assemble and deploy these molecules to mediate inter- and intra-species interactions, both cooperative and competitive. Secondary metabolites serve as weapons, regulatory signals, community stabilizers, and resource acquisition tools. Research has begun to dissect the impact of bacterial secondary metabolites on organismal fitness and community dynamics.[1-13] The understanding of eco-evolutionary pressures that influence the distribution and activities of secondary metabolites has

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informed new strategies for discovery of antibiotics and other useful molecules.[2,14–16] Indeed, bacterial secondary metabolites and their derivatives are the source of many valuable, bioactive molecules, with many applications across medicine, agriculture, and biotechnology.

Given their importance in natural systems and for human benefit, many large-scale efforts to discover new bacterial metabolites have been launched over the last 100 years to much success. As early as 1876, Louis Pasteur noted that harnessing antagonism between microbes would offer "perhaps the greatest hopes for therapeutics."[17] Systematic screening efforts were implemented by Waksman and others in the mid-twentieth century to identify soil bacteria that produced antibiotics. These early efforts ushered in the "golden age of antibiotic discovery," yielding many new classes of antibiotics including the aminoglycosides, glycopeptides, macrolides, ansamycins, cephalosporins, tetracyclines, and many others that became central to combatting infectious diseases.[18] Similar efforts to screen for immunosuppressive, anticancer, and other therapeutically relevant molecules followed and have continually fueled both the industrial and academic drug-discovery pipelines: over 68% of the nearly 3000 FDA approved drugs from 1981–2019 are either bacterial natural products or are inspired by their chemistry.[19]

Despite these successes, discovery has begun to stagnate, in part due to the frequent "rediscovery" of already known secondary metabolites,[20–23] leading some to claim that we have exhausted bacterial natural product diversity. This is especially true of antibiotics—no new major classes were discovered between 1962 and 2000.[24] Often cited estimates from the late 1950s predict that in a random sampling of Actinomycetes, 90% will produce streptothricin, 5% will produce streptomycin, and 1.6% will produce tetracycline antibiotics, which could lead to wasteful efforts culminating in rediscovery of known compounds.[23–25] Some estimates

claim rediscovery rates for particular antibiotics can be higher than 99%,[26] while rare natural products may be produced by one in 10 million strains,[23] implying large-scale efforts are required to overcome rediscovery and reveal true novelty. Most of large pharma abandoned natural product discovery in the 1990s, citing rediscovery and lack of technological innovation as reasons for unfavorable returns on investment.[18,27]

Discovery has been limited by focusing screening efforts on a narrow band of habitats and few phenotypes. Only one trillionth of bacteria in Earth's soils have been screened for antibiotic activity.[25,28] Many other environments and the myriad of other phenotypes beyond inhibition of other microorganisms await exploration. The confined view of bacterial diversity imposed by focus on the natural products produced by a few select taxa has persisted for decades. Current day researchers can do better—here, we discuss design strategies to tap the undiscovered cornucopia of molecules and optimize discovery rates. We invoke the "search for needles in a haystack" metaphor to analyze the features that will enable future large-scale screening to deprioritize known compounds and focus on novel bacterial secondary metabolites. We offer examples of recent large-scale sampling initiatives and prioritization strategies that enhance discovery rates for natural products (*i.e.*, needles) with new sampling approaches in different ecosystems (*i.e.*, haystacks).

Technical advances in genomic sequencing and metabolomic profiling have rekindled interest in probing nature's medicine cabinet, and Big Data resources are becoming increasingly available for natural product discovery efforts. Analyses of these growing datasets suggest that we are nowhere near exhausting nature's extreme chemical and biosynthetic diversity. This leads to the challenge of exploiting this biosynthetic potential more effectively—as our catalog of the microbial world continues to grow rapidly, it is essential to reexamine the biases that have constrained discovery in the past. In the

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following sections, we explore how taxonomy, habitat, methods of microbial capture, and screening can be reformulated to expand discovery.

2. Overcoming rediscovery: examining the relationship of taxonomy and chemical diversity

Historically, natural product discovery has focused on a narrow group of taxa, largely in soil. This resulted, in part, from biases developed from early discoveries among the *Streptomyces*. Only a tiny portion of the vast taxonomic (and thus chemical) diversity in soil has been captured by past searches. The amount of soil sampled, for example, is a miniscule portion of the world's soil. Typical sampling methods are often insufficient to describe the 10^3 to 10^7 species of bacteria estimated to be present in many soil samples.[29–35] Given our recently expanded view of the tree of life,[36] true species richness may even be higher. Since many secondary metabolites are observed in only one species, it stands to reason that taxonomic diversity begets chemical diversity, and therefore monumental chemical variety likely awaits discovery among the thousands or millions of as yet undiscovered species. Indeed, genomic surveys of biosynthetic diversity substantiate this assertion.[22,37–41] Surveys of metagenomic and 16S rRNA gene sequences have begun to describe a multitude of microbiomes across the planet (*e.g.*, the JGI Genomes from Earth's Microbiomes [GEM] Catalog[41] and the Earth Microbiome Project[42]) and provide the first step in estimating the extant biological and genetic diversity available for exploration. However, surveys based on phylogenetic markers rather than entire genomes will necessarily underestimate the potential for natural product discovery as many bacteria carry numerous biosynthetic pathways and the relationships between taxonomic and metabolic diversity remain obscure.[43] These relationships have emerged as worthy of exploration based on analyses of data from genomic databases.

Repositories such as MIBiG (Minimum Information about a Biosynthetic Gene cluster[44]) that curate previously described biosynthetic gene clusters are skewed by historical focus on a small number of genera. The genus *Streptomyces*, for example, has been the dominant focus of the academic and industry screening efforts since the 1940s. *Streptomyces* are indeed gifted producers of secondary metabolites, with BGCs often representing large proportions of their genomes (median ∼ 15%),[14] but chemical diversity abounds in many other taxa. Searching public genomes (NCBI) for each BGC found in MIBiG reveals that 88.8% of BGCs are found only in a single phylum (*e.g.*, streptomycin is found only within Actinobacteria; Fig. 1A). Each genome was used as a DIAMOND blastx query against a protein sequence database containing each gene in MIBiG BGCs (percent identity ≥ 70; query coverage ≥ 70). A BGC was denoted present if at least 50% of the MIBiG BGC's genes were present in the genome. We note that this gene-centric approach an indirect measure of BGC content (genes hits may not necessarily be at the same genomic locus), but has the advantage of scaling to large genomic datasets of variable quality, from very fragmented to complete assemblies. Although we impose high coverage and identity requirements for gene matches, hits may exist for related, yet distinct, BGCs or other gene clusters. This may be the case for some oligosaccharides where many BGC genes are found throughout primary and secondary metabolism alike. For example, we note that identification of at least 50% of the kanamycin MIBiG BGC's genes should be interpreted as indicating that a genome has kanamycin-like genes present and such hits do not necessarily represent a gene-by-gene rediscovery of the kanamycin BGC.

Shannon entropy, a measure of diversity, at the level of phylum ranges from between 0 (no diversity; a BGC is only found within one particular phylum) and 0.47, with 11.2% of BGCs occurring in more than one phylum (Fig. 1A). BGCs are found across a wider diversity of orders—30.2% of BGCs are found in more than one order (Fig. 1B) and the range of Shannon entropy of each BGC with respect to order also expands to a range of 0 to
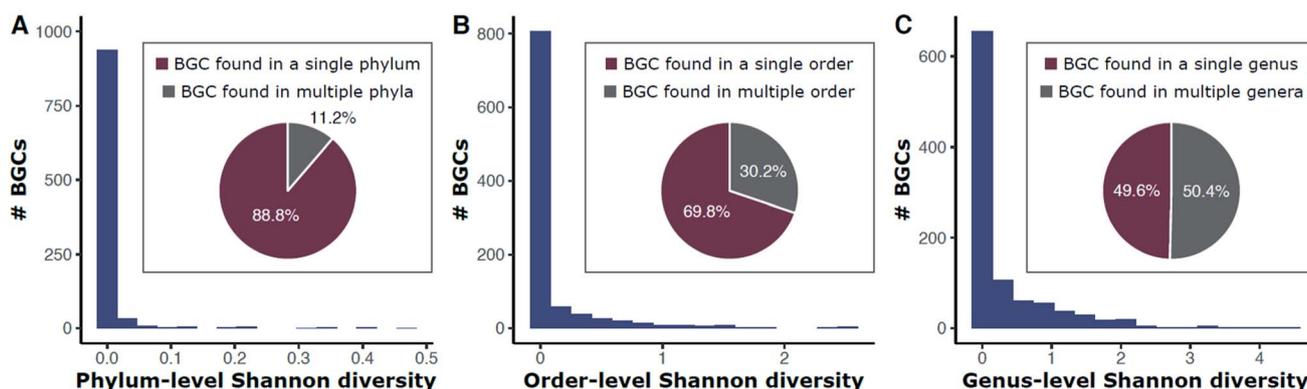
Fig. 1 Diversity of phyla (A), orders (B), and genera (C) for BGCs in public bacterial genomes. Bars are a histogram of Shannon entropy for each MIBiG v2.0 BGC in 352 275 bacterial genomes (NCBI). Each genome was used as a DIAMOND[183] blastx query against a protein sequence database containing each gene in MIBiG BGCs (percent identity ≥ 70; query coverage ≥ 70). A BGC was denoted present if at least 50% of the MIBiG BGC's genes were present in the genome. Shannon entropy (H) was calculated for each BGC at the phylum (A), order (B), and genus (C) levels. Proportions of BGCs with no diversity ($H = 0$) *versus* some diversity ($H > 0$) are shown in the inset pie charts. For example, 88.8% of BGCs are found in only a single phylum and 11.2% are found in more than one phylum (A).

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1.36. Still, most BGCs are found only within one order. This trend continues down to the genus level with half (49.6%) of BGCs found only within one genus and the other half present in more than one. The Shannon entropy with respect to genus of each BGC is much broader (range 0 to 4.44; Fig. 1C).

Arguments about the relationship between taxonomy and chemical diversity are intrinsically flawed because taxonomic designations are a human, rather than biological, construct. Although often informative and useful, taxonomy is an imperfect measure of diversity. As a consequence, the genetic diversity (and accompanying chemical diversity) within these arbitrary groups varies wildly across the tree of life. Compare, for example, the extreme genetic diversity among *Escherichia coli* strains to the nearly identical genomes within *Mycobacterium tuberculosis*. Yet these groups are both designated "species". Therefore, are observations about frequency of discovery of natural products in a certain taxon an artifact of how much biological diversity has been arbitrarily crammed into the group with that name? Or rather, do these observations reflect an intrinsic propensity of the group? Often these questions are difficult to disambiguate. In addition to taxonomic anomalies affecting conclusions about which groups are prolific producers, past searches influence today's perceptions of productive taxa. For example, discovery frequencies established decades ago with a small sampling of *Streptomyces* from selected soils can hardly be extrapolated to an entire genus that has had ~380 million years of evolution in which to diversify metabolically.[43,45]

The frequency of a given BGC across members of a genus can provide information about when the pathway evolved, about the likelihood of finding it again in that group, and about how likely it is that members of a habitat may have encountered it. Distribution frequencies vary by pathway and taxon. Within publicly available genome sequences of *Bacillus*, 51.6% contain the BGC for the siderophore petrobactin and 11.4% have the BGC for the antibiotic zwittermicin A (Fig. 2A). The BGC for the multifunctional signal, quinolone (or PQS) is in 50.2% of *Pseudomonas* genomes whereas the antibiotic koreenceine appears less frequently (2.3%; Fig. 2B). Often cited estimates

that 90% and 5% of *Streptomyces* produce streptothricin and streptomycin, respectively, are questionable based on genomic presence of the BGCs, which are found in 3.6% and 1.9% of public *Streptomyces* genomes, respectively (Fig. 2C). This discrepancy can be reconciled by the phylogenetic distribution of streptomycin producers: *Streptomyces* capable of producing streptomycin are relegated to a very small portion of the *Streptomyces* phylogenetic tree comprised of *S. griseus* and close relatives,[14] suggesting that the abundance of these BGCs in soil estimated from screening initiatives is representative of neither the natural abundance of these organisms in soil nor the phylogenetic breadth of *Streptomyces* as a genus. Inflated rediscovery rates of streptomycin may be due to the relative ease of isolating *S. griseus* (and its immediate sister taxa) compared to other lineages that evolved over nearly 380M years of divergence within *Streptomyces*.[43,45] This could be influenced by culture methods, biases in environmental sampling, or even the biology of *S. griseus* itself (*e.g.*, growth rates, niche-protecting compounds to discourage the growth of other *Streptomyces*, *etc.*).

The lenses through which we view the biological world determine what we see. Culturing provided insight into the taxa that produce natural products. The advent of genomics and increased computational power provide a view of the world of natural products and their distribution among microorganisms at higher resolution. This insight demands that we let go of some of the biases generated by decades of culturing a narrow spectrum of taxa from a narrow spectrum of environments. Although it remains the most powerful tool in microbiology, culturing must be treated as only one lens among many, and its limitations must be critically assessed if we are to make inferences about features of the microbiological world that are not measured directly.

2.1. Considering environment

The environment from which microbes are sampled is an extremely important consideration in the search for new natural
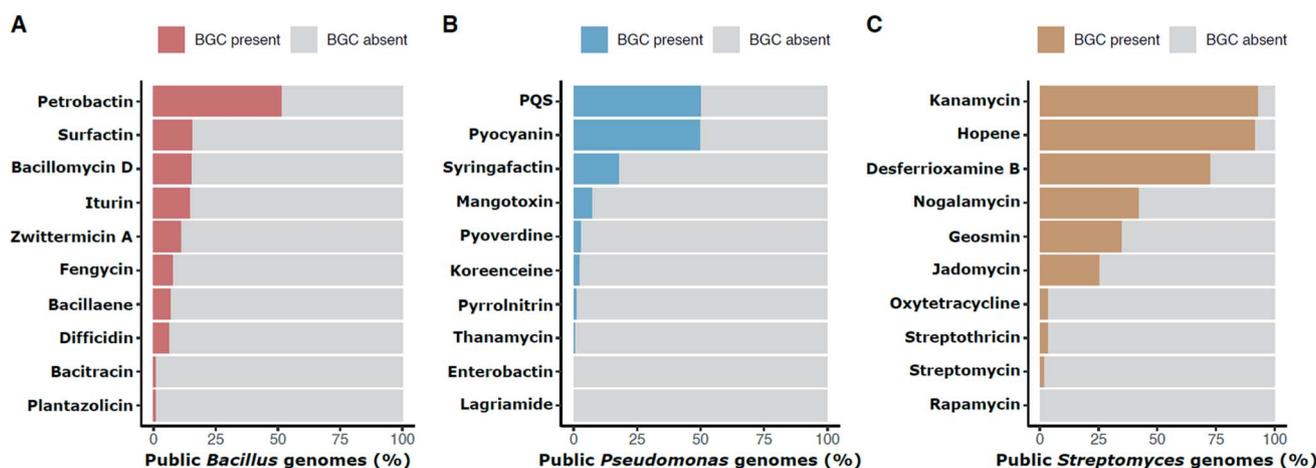

Fig. 2 Percentage presence of selected BGCs in public genomes of individual genera: *Bacillus* (A), *Pseudomonas* (B), and *Streptomyces* (C). Presence is defined as in Fig. 1. For example, the petrobactin BGC is found in approximately 51.6% of public *Bacillus* genomes.

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products. There has been some success by sampling previously unexplored microbial communities, such as the walls and deposits of secluded cave systems,[46,47] ocean sediments,[48–51] and even built environments like the New York City subway system.[52] Expanded focus on relatively undescribed taxa, such as *Salinispora*,[48–50] *Myxobacteria*,[53] *Pseudomonas*,[54] and *Phodorhabdus*55 has also yielded new and exciting molecular discoveries. Here, we focus on two major sources for natural product-producing microbes: soil, a traditional source with subtle complexity that has been largely ignored, and host-associated microbiomes, an emerging source of bioactive compounds.

2.1.1. Soil. Soil is probably the most species-rich environment on Earth. It is a matrix of minerals mixed with live and decomposing organisms that is continually changing. Plants are the primary source of carbon in soil and alter the soil as they nourish it with their nutrient-rich root exudates that foster bacterial growth. Soils have their origins in their underlying geologic material, which varies in composition and rates of change. Some soils are ancient—some as old as 3.7 billion years—and some formed during recent geologic events, such as glacier retreat, in recent centuries.[56,57] Geologic origins and age are significant aspects of soil variation, but they are certainly not the only ones. The weathering of soil minerals, plant and animal diversity, climate, and land use all contribute to the evolution of soil. Different outcomes shape the 22 000 different soil series (series are the lowest taxonomic level, akin to species in biological taxonomy).[58] Soil census data suggest that each soil series might contain a unique bacterial species. There are certainly strains unique to any environment. How many of these strains and species carry a pathway for synthesis of a unique natural product? Historically, the natural products field has largely equated tapping soil diversity with sampling multiple countries or continents at one time point—hardly enough to capture the mosaic of soil types, land use, and temporal events (*e.g.* ecological succession, seasonal changes, weather-driven disturbances, *etc.*) that vary widely across geographic scales. It is doubtful that all 22 000 types of soil have been sampled, and similarly the diversity within each series, is largely unknown and untapped. It remains unclear what the proper (and practical) sampling scale should be to capture meaningful interactions between microbes in soil for natural product discovery. Disease suppressiveness[59] and sampling depth[60] appear to influence the biosynthetic potential of some soils. Likely many other parameters do as well. Some natural product surveys compare samples that are on opposite sides of continents to infer biome-level variation in biosynthetic potential, whereas others observe metabolic differences in communities associated with a single grain of sand[61,62] and in experimental systems at the micron scale.[63]

New natural products have been found even in ordinary soils that are readily accessible to people.[26,64–68] It is likely that these novel compounds are the tip of the proverbial iceberg that represents all of Earth's soil. The 22 000 soil types in locations that have not been sampled previously could be subjected to systematic sampling that accounts for soil type and land use. Soil sampling by chemical hunters has instead been dictated by proximity to the researcher's lab, travel destinations, or the

habitat's macroorganism diversity. For example, in the 1990s, tropical rainforests became popular habitats for seeking new chemistry in soil bacteria because of the species richness of the aboveground plants and animals. The relationship between soil location and taxonomy in chemical discovery remains murky, and therefore it might be just as productive to seek novelty in the drained lake basins in the arctic[69] or the drylands of the Tabernas Desert in Spain.[70] Biosynthetic surveys that focus on historically neglected microbial taxa suggest that there is value in exploring taxonomic diversity as well.[71] For those who focus efforts on the rain forests, a fruitful source are the symbionts of the staggering diversity of plants and animals that are likely a prolific source of novel bacterial species, and consequently, chemical diversity.[2,14,72–74]

2.1.2. Host-associated microbiomes. The microbiomes of eukaryotic hosts have received increased attention in recent years as a promising source of bioactive molecules, leveraging the unique chemistry that bacteria use to interact with other members of the resident microbiome, would-be microbial invaders, and the host itself.[14] In human health, bacterial metabolites have been implicated in numerous disease states and clinical outcomes.[75–78] Growing appreciation for microbiome composition and metabolite function in non-human hosts, including marine invertebrates, insects, and plants, has also led to fruitful discovery efforts.[2,74,79–83]

Invertebrates and plants lack the antibody-based adaptive immunity that is typical of higher animals, instead relying on innate immunity and chemical protection from pathogens. Many macroorganisms enlist bacterial partners in providing chemical barriers to invasion by pathogens.[1,5,84–87] Some of the most prolific sources of natural product-producing bacteria are insects such as the fungus-growing ant,[1,84–86] solitary wasp,[87,88] and southern pine beetle[5] systems, in which Actinobacteria (typically *Streptomyces*) are deployed to suppress insect infectious disease in an intriguing parallel with human use of the same phylum of bacteria for antibiotics. For example, the *Streptomyces* symbiont of the southern pine beetle (*Dendroctonus frontalis*) produces the secondary metabolites frontalamide A, frontalamide B, and mycangimycin, each targeted to a different fungus that is pathogenic to the insect.[5,89] Mycangimycin also has potent activity against the malaria-causing *Plasmodium*, and the frontalamides have general antifungal activity.[5,89] The natalamycin derivatives produced by *Streptomyces* from the fungus-growing termite system provide similar antifungal defense.[90] *Streptomyces* provide chemical protection to the larvae of solitary wasps through production of streptochlorin, piericidin analogs, and sceliphrolactam.[87,88,91] Beewolves (genus *Philanthus*) have associated with antimicrobial producing *Streptomyces* symbionts for many millions of years, with evidence that biogeography has in part shaped their unique chemistry over evolutionary time.[88] Natural products from insect-borne bacteria are not limited to Actinobacteria. The invasive beetle, *Lagria villosa*, for example, harbors *Burkholderia gladioli*, the source of the gladiofungins, novel compounds that provide the beetle with protection from the entomopathogen, *Purpureocillium lilacinum*.[73] Similarly, the distribution of the BGCs and antibiotic molecules of

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Pseudonocardia symbionts of fungus-growing ants (tribe Attini) suggests that biosynthetic potential is influenced by biogeography, even at very fine geographic scales of less than a few kilometers.[1,92] In the ant-*Pseudonocardia* symbiosis, these bacteria protect the ant's food source from coevolved parasitic fungi[85,93] and the ants themselves from entomopathogens.[94,95] Systematic genomic and metabolomic surveys across insect-associated *Streptomyces* reveal that the diversity of insect symbiont chemistry is both diverse and unique, and thus is a source of previously undescribed bioactive molecules.[2] As a group, insects occupy almost every terrestrial niche, so a variety of environmental influences are reflected in a wide diversity of insect-microbe symbiotic systems. A recent example resulting in molecular discovery is cyphomycin, an antimicrobial polyene from an ant-associated *Streptomyces* with activity against the fungal pathogen (genus *Escovopsis*) that invades the ant's farms. Cyphomycin also has activity against multidrug-resistant clinical isolates of *Candida* spp., including *C. auris*, and leishmanial human parasites (genus *Leishmania*).[2,96]

Other host-microbe systems have provided sources for molecular discovery as well. Mining the microbiomes of tunicates and other marine filter-feeding animals has yielded new molecules with potential uses in human medicine.[97–99] In particular, symbiont Actinobacteria (genus *Micromonospora*) from sea squirt microbiomes have emerged as a promising source of antimicrobial metabolites active against multidrug resistant bacteria and fungi.[100,101] This propensity towards the production of bioactive molecules may in part be due to the host's filter-feeding, sessile lifestyle, compensating for lack of mobility with chemical defences. *Bacillus* spp. from sourced from tunicate microbiomes have also shown promise,[102] highlighting that discoveries await across bacterial phylogenetic space, not just in the Actinomycetes. Bacteria in the human microbiome also produce antimicrobial compounds and may serve as a new source for discovery. The human body carries quite distinct communities in various sites such as the nose, skin, gut, vagina, and inner elbow. Microbial members of these communities produce metabolites specific to interactions with other microorganisms that reside in that location.[103,104] Members of the vaginal microbiome, for example, deploy antibiotics such as the lactocillins for defense against pathogens.[105] Likewise, metabolites from a *Staphylococcus* sp. from the nasal microbiome inhibit growth of multi-drug resistant pathogens.[106] Other natural products shape competitive landscapes other body sites.[106–110]

In a remarkable project at Yale University, Strobel and Bascom-Slack engaged undergraduate students in scientific research through a course that involved a field trip to the Equadorian rainforest to isolate fungi from plants and screen them for biological activities upon return to the lab.[74] The project explored several novelties—fungi, which have been less well combed than bacteria for natural products; Equador's rainforest, which is inaccessible to most researchers; and endophytes, which reside inside plant stems. The yield of novel compounds was outstanding. One endophytic fungus produced a novel alkaloid, irrepairzepine, that is toxic to glioblastoma tumor cells through it ability to inhibit DNA repair (which is implied in the compound's name).[111] A screen for volatile organic compounds found 140 unique compounds from 113 fungal endophytes.[112] Another screen identified a fungal endophyte of the tropical tree, *Duroia hirsute*, which produces a new sesequiterpene-polyol, pyrrolocin, with inhibitory activity against *Staphylococcus aureus*, *Enterococcus faecalis*, and *Candida albicans*.[113,114] Another student found xyolide, a novel nonenolide containing a 10-membered lactone ring. Xyolide inhibits growth of *Pythium*, a member of the destructive plant pathogens, the oomycetes.[115] The novelty of compounds discovered by students in tropical plant endophytes provides evidence of the untapped potential of unusual bacterial habitats such as the inner sanctum of plants residing in the Equadorian rain forest.

Bacteria from inside and outside of plant roots have also been the source of novel compounds. Isolates from the rhizosphere—the region on and around the root that is affected by the root—are prolific producers of natural products. Strains of *Bacillus* and *Pseudomonas* spp. produce cyclic lipopeptides, some of which are antimicrobial and other influence bacterial motility and biofilm formation.[116] Genomic analysis of root endophytes led to the inference that they were products of a new nonribosomal peptide and polyketide hybrid pathway of a *Flavobacterium* sp. that suppresses infection by the root pathogen, *Rhizoctonia solani*. *Flavobacterium* induces expression of the antibiotic in *R. solani*.[117] The producing organism was isolated from a sugar beet root collected by a field in The Netherlands. Similarly, abundant isolates that produce zwittermicin have been found in the rhizospheres of alfalfa, soybean, and other plants, and have allowed for dissection of zwittermicin's biosynthetic pathway.[118,119] These examples of discovery of novel compounds from plant-associated microorganisms in the Equadorian rain forest and agricultural sites in North America and the Netherlands indicates that new chemistry is waiting to be discovered in environments that many scientists consider both exotic and ordinary.

2.2. Considering microbial capture

2.2.1. Relationships between isolation media and metabolic phenotypes.
Decades of experimental evidence suggests that most bacteria in soil and many other natural habitats are not readily culturable on standard laboratory media. The "Great Plate Anomoly" notes that the number of bacteria seen by culturing methods vastly underestimates the numbers predicted by microscopy[120] or sequence-based surveys (*e.g.*, 16S amplicon sequencing, shotgun metagenomic sequencing, *etc.*[121]). Just as bacteria vary in their metabolic potential (*i.e.*, their ability to produce different molecules), so too do they vary in their metabolic requirements (*i.e.*, what nutrients are required for growth). Culture conditions dictate which organisms grow in culture, enriching for some while excluding others, and therefore they affect the profile of natural product-producing potential. For example, a pilot study from Tiny Earth, a student-driven antibiotic discovery effort based on the Yale Equadorian rainforest course, showed that soil bacteria isolated on tryptic soy agar (TSA), in which glucose is the major

carbohydrate source, enriched for isolates with different inhibitory phenotypes than did potato dextrose agar (PDA) which has both simple dextrose and more complex starchy carbohydrates.[122] Tiny Earth students isolated bacteria from four United States soil samples on PDA and TSA. On average, more bacteria isolated on PDA displayed antimicrobial activity. The spectrum of activity of the isolates also varied with isolation medium: PDA enriched for strains that inhibited *Acinetobacter baylyi* and *Pseudomonas putida*, whereas TSA enriched for those that inhibited *Erwinia carotovora*. Selective media can enrich for specific taxa whose isolation might otherwise be overwhelmed by more abundant and/or fast-growing organisms. For example, the media described by the International *Streptomyces* Project (ISP) have been used for many decades to enrich for various Actinobacteria.[123]

2.2.2. Enrichment for resistant and/or hard-to-culture strains. Similarly, antibiotics added to the isolation medium select for organisms with resistance and exclude those without, in an effort to skew sampling towards taxa of interest. Exploiting molecular resistance mechanisms can also yield promising new molecules within a chemical class. For example, adding vancomycin to culture medium has been used to select for organisms with glycopeptide resistance who themselves may have the biosynthetic capacity to produce glycopeptides.[124,125] Vancomycin-infused media enriched for glycopeptide producers, some of which may be capable of making previously undescribed glycopeptides. A necessary assumption of this approach is that resistance to the known and new glycopeptides works by the same molecular mechanisms. Although this is not a perfect selection, because evolutionarily related compounds with different resistance mechanism(s) will be excluded, and organisms that have resistance genes without accompanying biosynthetic pathways will be included. Nevertheless, this enrichment strategy has proven quite useful, enhancing the frequency of finding producers of target pathways by many orders of magnitude.[124,125] This method works because resistance genes such as *vanY* appeared in glycopeptide pathways between 400 and 150 million years ago, contemporaneously with some biosynthetic genes, suggesting that a shared mechanism of action of glycopeptides allowed for pathway diversification (while retaining activity).[126] In these cases resistance and mechanism of action are either intrinsically linked (*e.g.*, the resistance mechanism chemically interferes with the interaction of the antibiotic with the molecular target), or the resistance determinant recognizes structurally similar compounds (*e.g.*, efflux pumps). Thus, exploiting resistance is yet another way to increase the frequency of active compounds.

Rare community members are often present in their habitats at such low cell densities that they are unable to compete under typical isolation conditions. The discovery of teixobactin, a broad-spectrum antibiotic, has demonstrated the power of the *in situ* culture of microorganisms within a complex ecosystem to give a boost to rare organisms.[127] In this case, a multi-well device called iChip enabled incubation of bacteria in the presence of a microorganisms within their natural soil environment.[113] The intent is to culture bacteria proximal to abiotic and biotic cues until cell counts are high enough that they can be subcultured readily in the lab. This *in situ* culture enabled the growth of bacteria that are usually "non-cultivable," including the producer of the novel antibiotic teixobactin.

2.2.3. Elicitation of silent secondary metabolism. The search for needles in haystacks continues after a microbe is cultured in the laboratory. The microbe must then produce a molecule that elicits a desired phenotype (*e.g.*, binding to a molecular target or killing a pathogen of interest). The BGCs of many natural products are under tight regulatory control, causing these pathways to be expressed under some conditions and repressed under others. Thus, exploring different laboratory conditions that introduce a myriad of biotic and abiotic cues to stimulate the production of otherwise "silent" BGCs has been successful. Co-culture of microorganisms can activate the expression the biosynthetic genes responsible for production of certain antimicrobial compounds, which may have evolved to mediate cooperative or competitive interactions between the organisms in their natural habitat. Coculture with the target organism (*e.g.*, a pathogen for antibiotic screening) or with sympatric organism(s) (*e.g.*, coisolates from the same community as the producer strain) has been a useful tool to stimulate expression.[100,128–133] High-throughput screening of elicitor compounds has also been useful. This has included subinhibitory levels of antibiotics and metals[134–137] and inhibitors of epigenetic machinery[138,139] that have been used to increase BGC expression.[140–142] Both targeted (*e.g.*, promotor engineering) and untargeted (*e.g.*, forward genetic screens[143]) engineering strategies can be employed to increase production in the native producer or in heterologous expression strategies. Molecular capture methods have enabled the recovery of entire BGCs directly from metagenomic DNA, skipping the need to isolate and culture the native producer itself, instead expressing the BGC in an amenable chassis organism.[26,64,144,145]

2.3. Considering targets

The serendipitous discovery of penicillin in 1929 shaped the search for antibiotics for the rest of the 20th century. The striking zone of inhibition of *Staphylococcus aureus* around the *Penicillium* mold was quickly recognized as a means to detect antibiotic-producing microorganisms and became the basis of the systematic screening of soil bacteria that led to discovery of streptothricin, streptomycin,[146] and the plethora of compounds that followed. The screen's power lies in its simplicity, visual potency, and lack of assumption of underlying mechanism of action. It is limited by the ability of the producing organism to accumulate sufficient quantities of the active molecule to produce a zone of inhibition under the conditions selected. In addition, the choice of target organism determines the universe of discovery.

The first decades of antibiotic discovery were defined by the dominant bacterial pathogens of the time. In 1900, pneumonia, tuberculosis, and enteritis were the leading causes of death in the United States. By the end of the century only pneumonia remained on the list of 10 leading causes of death, and many deaths due to pneumonia were the result of viral rather than bacterial infections.[147] In the latter half of the 20th century the

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profile of infectious disease began to change. The increase in immunosuppression as a side effect of organ transplants, chemotherapy, and HIV infections resulted in many more cases of sepsis, often caused by Gram-negative bacteria.[148–150] Increasing incidence of infections by *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, and *Escherichia coli* was followed by antibiotic resistance, creating a public health crisis even in countries with ready access to antibiotics. Screening expanded to include organisms such as *Acinetobacter baumannii*, a Gram-negative pathogen that emerged in the 1990s during military conflict in the Iraq desert, and *Candida auris*, a fungal pathogen that emerged in 2018; both are now causes of opportunistic infections in healthcare settings.[151,152] Antibiotics that selectively inhibit growth of these pathogens would not have been detected by the screens used in the 20th century and thus, would have been missed by the vast screening efforts of that era.

Likewise, screening against plant pathogens has yielded antibiotics that were not detected in traditional searches against human pathogens. A screen for suppression of damping off of alfalfa,[153] a disease caused by *Phytophthora medicaginis*, led to discovery of zwittermicin, produced by *Bacillus cereus*.[154] An antibiotic of many surprising features, zwittermicin remains the only member of its class—an aminopolyol containing a D-amino acid, ethanolamine, and an unusual terminal amide—even today, 25 years after its discovery. It is biosynthesized by a hybrid modular type 1 polyketide synthase (PKS) and non-ribosomal synthetase biosynthetic scheme[155] that requires two previously unknown PKS extender units, hydroxymalonyl- and aminomalonyl-acyl carrier proteins and incorporates both *cis*- and *trans*-acyltransferase PKS biosynthetic logic.[119,156] A worldwide screen of soil samples generated the puzzling finding that zwittermicin-producing strains of *B. cereus* are ubiquitous and abundant in soil, estimated to represent 10^4 culturable cells per gram of soil in diverse samples from four continents.[157] Why did an antibiotic produced by a readily culturable, widely distributed bacterium elude discovery throughout 60 years of intensive screening of soil bacteria for antibiotics? The answer reveals several biases in mainstream screening of the 20th century that likely led to omission of many interesting natural products.

First, the bias toward Actinobacteria led to relatively less attention to *Bacillus* spp., to date the only genus shown to produce zwittermicin. Second, the labor-intensive screen for suppression of alfalfa damping off was new to the antibiotic-discovery effort. The causal agent, *Phytophthora medicaginis*, is a protist most closely related to the golden brown algae, certainly not a likely target of previous screens. Although zwittermicin was later found to have a broad target range, including several Gram-negative bacteria, the detection of antibacterial activity required optimizing culture conditions, resulting in parameters that were not standard in industry screening.[158] Finally, its polar and water-soluble structure made zwittermicin challenging to purify and likely to have been discarded in programs that focused on organic-soluble compounds.[154] Zwittermicin provides tantalizing evidence that by selecting new targets, broadening the taxonomic scope of potential producers, and including polar compounds among those pursued, there is

a plethora of natural products lurking in soil waiting to be discovered.

The discovery of ivermectin illustrates the importance of target, assay, and luck in finding new natural products.[159] Bill Campbell's group at Merck screened 40 000 Actinobacteria for the ability to cure mice of nematode infections. Only one, now known as a strain of *Streptomyces avermitilis*, produced the avermectins, a mixture of macrocyclic lactones that had strong anthelmitic activity. The 22,23-dihydro derivative of avermectin B became the highly successful drug ivermectin, which is used widely for filarial diseases, leading to a $2 billion annual market for animal health applications, such as canine heartworm, and the projected eradication of river blindness in people by 2025.[160–162] Initial detection of avermectin was enabled by its strong antihelmintic activity in the mouse model. The isolate likely would have been overlooked in typical antibacterial screens (although avermectins was eventually shown to have activity against *Mycobacterium tuberculosis*, the causal agent of TB).[163] The discovery also relied on an assay that was filled with so many unknowns that it almost defied the tenets of science. As Bill Campbell noted in his Nobel Prize acceptance speech, "It was bizarre!—but simple…. You line up a series of individual infected mice. You treat each mouse with an unknown amount of an unknown substance that might not be there. Then you check to see if the treatment worked."[164]

The discovery of the avermectins was also the product of a serendipitous collaboration between Satoshi Omura at the Kitasato Institute in Japan and Bill Campbell at Merck in the United States.[165] Omura collected the key soil sample on a golf course in Japan and recognized the unique morphology of the *S. avermitilis* isolate, which was among a collection of Actinobacteria that he sent to Campbell, whose group screened the isolates for activity against mammalian diseases caused by filarial worms.[163] If Omura had not sampled that particular golf course, did not have a keen focus on bacterial morphology, had not struck up a collaboration with Campbell, and if Campbell had worked on a different class of pathogens, the avermectins might have been missed. And the rarity of avermectin production among the Actinobacteria should be all the evidence we need to compel the continued hunt for natural products among cultured bacteria. The discovery process can get more efficient using both laboratory methods and genomic information to hunt smarter.

2.4. Increasingly digital needles and haystacks

Whether we look outwards to the microbes of extreme environments or inwards to our own microbiomes, genomic and metabolomic advances have enabled an increasingly digital search for new chemistry and natural product-producing organisms. The sequences of biosynthetic genes and domains are often used to query digital haystacks to identify biosynthetic pathways and predict their chemical products. Reference catalogs of BGCs with varying levels of experimental validation can be used to estimate the distribution of BGCs and assess their novelty.[44,166–168] Spectra generated by tandem mass spectroscopy (MSMS) can be networked across many different datasets to aid

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in molecular characterization and dereplication.[169] Untargeted metagenomic approaches in the human and other microbiomes have unlocked hidden taxonomic and biosynthetic insight of these environments at ever-increasing rates.[4,170–172] Increasingly, integration of many or all of these "digital" technologies is at the heart of molecular discovery efforts.[2,122,173]

For example, Warp Drive Bio[174] assembled a collection of over 1.35×10^5 Actinomycetes, primarily from legacy big pharma and public collections of *Streptomyces* and related organisms, to search for novel examples of rapamycin-family T1PKS-NRPS hybrid natural products. Rapamycin, FK506, and other important molecules in this class engage their molecular targets only when in complex with a "presenting" FK506 binding protein 12 (FKBP); neither FKBP12 nor the metabolite alone is sufficient for target engagement. The surface of this binary metabolite + FKBP12 complex engages completely different targets: calcineurin and mTOR for FK506 and rapamycin, respectively. The hemisphere of the metabolite that initially binds FKBP12 is conserved, while the hemisphere that engages the target is variable. Using this knowledge of evolutionary conservation and diversification, DNA from the collection was pooled and sequenced at low depth. Enzyme sequence for key biosynthetic steps in rapamycin and FK506's conserved FKBP12-binding region were used as queries to identify pools containing hits. Individual members of positive pools were deconvoluted *via* PCR and follow up deep sequencing was used to identify novel members of the rapamycin/FK class of BGCs. Overall, this effort yielded seven novel, distinct pathway architectures, notably the BGC X1 which assembles the CEP250-binding molecule WDB002, yet another example of a completely different target for a member of this class. Estimated frequencies of X1, rapamycin, FK506/520, and antascomycin were 1 in 3068, 1 in 5870, 1 in 6136, and 1 in 45 000, respectively. Other novel rapamycin-class pathway architectures were found only once within the ~135 000-organism dataset, suggesting even this large sampling had not reached saturation of rapamycin-family pathways. As large-scale sequencing studies become increasingly feasible, economically and technically, digitized haystacks are now typically a combination of public and newly generated data and have been used to search for particular chemical or biosynthetic needles, including phosphonates,[175,176] lasso peptides,[177] betalactones,[178] *trans*-acetyltransferase polyketides,[179] enediynes,[180,181] and many others.
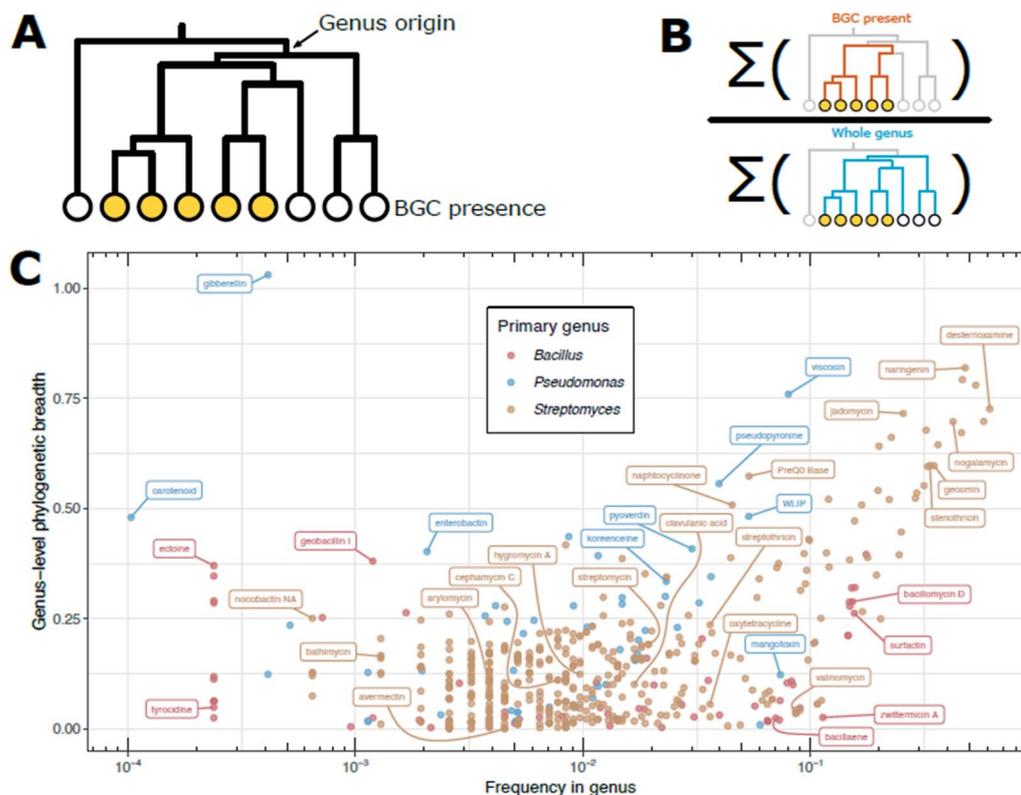
Sampling biases determine that the frequencies reported in the literature are not necessarily representative of the distribution and frequency of certain natural products (and their BGCs) across taxonomy. We can see many examples of this if we adjust for phylogenetic biases in sampling. For example, a search of all publicly available *Bacillus* genomes for the zwittermicin BGC, shows that it is found at a frequency of approximately 1 in every 10 *Bacillus* genomes. It may be tempting to then surmise that screening 10 *Bacillus* isolates would turn up zwittermicin, but this is not necessarily the case. On further inspection, it appears that zwittermicin BGC is found in a very narrow region of the overall *Bacillus* phylogeny, only in the *B. cereus* group and closely related species. We can quantify the

fraction of the overall *Bacillus* phylogeny that contains a given BGC as "phylogenetic breadth." If we construct a phylogenetic tree of an entire genus and label members that have a BGC (Fig. 3A), we can calculate phylogenetic breadth as the ratio of the sums of the branch lengths for strains with the BGC present to strains in the entire genus (Fig. 3B). Intuitively, a genus-level phylogenetic breadth of 0 signifies a given BGC is not present in the genus, and a phylogenetic breadth of 1.0 signifies that the total branch length spanned by producers is equal to that of all members of the genus. Phylogenetic breadth may be greater than 1 if a BGC is found in outgroups from the focal genus. For all publicly available *Bacillus*, *Pseudomonas*, and *Streptomyces* genomes, we can calculate genus-level phylogenetic breadth for BGCs in the MIBiG v2.0 BGC repository and compare that to the unadjusted frequency of those BGCs to visualize this taxonomic sampling bias (Fig. 3C).

Zwittermicin, for example, is found in the lower right quadrant of Fig. 3C, signifying it is frequently found in *Bacillus* genomes, yet these genome are relegated to a very narrow segment of the overall *Bacillus* phylogeny. There may be many explanations for this discrepancy; perhaps *B. cereus* and close relatives are extremely abundant in the environments sampled and/or are readily and easily culturable compared to other *Bacillus* spp. The frequency of the streptomycin BGC in the public genomes of *Streptomyces* spp. is approximately 1 in 100, which is consistent with often-cited frequency estimates. However, the *Streptomyces* spp. that produce streptomycin span only 20% of the phylogeny of the genus (*i.e.*, a phylogenetic breadth score of 0.2). This may indicate that isolation media and/or selection strategies for *Streptomyces* spp. tend to enrich for *S. griseus* and near relatives that produce streptomycin, or that these organisms are abundant in the environments that have been heavily sampled for natural product screening thus far. Strains carrying BGCs for desferrioxamine and geosmin (upper right quadrant of Fig. 3C) cover greater phylogenetic breadth within the *Streptomyces*, suggesting that since these molecules are found throughout the phylogenetic group, perhaps because their metabolic products provide important functional roles for *Streptomyces* spp., generally. Avermectin provides an extreme example: frequency within genomes is between 1 in 100 and 1 in 1000, yet those strains have near-zero genus-level phylogenic breadth.

These examples caution that frequencies of discovery alone can be misleading if only a small proportion of phylogenetic space is sampled or [unintentionally] selected. The relegation of BGCs to specific (often narrow) evolutionary lineages also supports the growing appreciation that vertical evolution is a major mechanism of BGC evolution and horizontal transfer events, while observed, are relatively rare.[14,45,182] The phylogenetic observations discussed here are based only in genomic information in contrast to previous screening estimates based primarily on observed bioactivity. Similarly, Warp Drive Bio's discovery of the X1 BGC further illustrates the power of genomic information. All members of Warp Drive Bio's isolate collection were part of private or public collections, some of which were decades old. It was only through sequencing that the rapamycin-like X1 BGC was revealed, and within that sample set

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Fig. 3 Phylogenetic breadth of BGCs in public genomes. (A) Example dendrogram for a given BGC noting the genus origin. Filled circles at the leaves indicate the BGC is present in those strains. (B) Cartoon representation of the calculation of phylogenetic breadth. Phylogenetic breadth is expressed as ratio of (i) the sum of the branch lengths for the subset of the tree that has a BGC present (orange branches) to (ii) the sum of the branch lengths for the entire genus (blue branches). In other words, how much of the tree is populated by organisms with the BGC compared to the entire genus (scale 0 to 1). (C) Measure of phylogenetic breath *versus* frequency for BGCs in *Bacillus*, *Pseudomonas*, and *Streptomyces* genomes. Presence is defined as in Fig. 1. Frequency of a BGC in all public genomes of a genus (either *Bacillus*, *Pseudomonas*, or *Streptomyces*) is shown on the *x*-axis. Since there is inherent sampling bias in public databases, we calculate a measure of phylogenetic breadth to describe how phylogenetically distinct strains are at the genome level. For each BGC, dashing was used to compute pairwise MASH distances for all genomes with the BGC. The MASH distance matrix was used to create an UPGMA dendrogram. We call "genus-level phylogenetic breadth" the ratio of the UPGMA dendrogram's total branch length for those genomes with a BGC divided by the total branch length for the UPGMA dendrogram for all members of the genus. For example, if all members of a genus have a BGC, the phylogenetic breadth would be 1. If a BGC is relegated to only 50% of the total phylogenetic space of a genus, then the breadth would be 0.5.

it appeared at higher frequency than the BGCs for rapamycin itself and those for FK506/520 and related, previously described molecules.

Many in the field have come to call the products of BGCs "specialized metabolites." If these are indeed specialized through evolutionary processes, then the producer's (and the BGC's) evolutionary history is shaped by the biotic and abiotic environment. Therefore, neither lineage nor environment can be ignored when designing sampling strategies to tap chemical diversity most effectively. In almost all cases it is difficult (if not impossible) to know how these molecules function in their natural environments. Further, it is often impossible discern which environments and/or lineages will enrich for the sought afer needle *a priori*. Which environments will yield antibiotics? Which lineages should be targeted to find immunosuppressants? Slowly, in particular systems under prescribed experimental conditions, understanding of such relationships is emerging, but how these principles apply across life's vast diversity will likely forever remain unknown. Thus, we must

integrate what we have learned from these few well-studied systems and simultaneously recognize and guard against sampling biases, whenever possible, to maximize discovery.

3. Concluding remarks and perspectives: lessons for building a systematic discovery engine

Strong evidence indicates that a plethora of natural products await discovery. Strategic selection of new habitats will provide novel microbes and their attendant chemistry. Well combed habitats such as soil will be a source for discovery if attention is paid to the subtle differences among soil types. New culture techniques will enable growth of microorganisms never before grown in captivity and elicit expression of pathways that have previously been silent. Molecular insights from genomics is driving discovery based on novelty of the BGC followed by attempts to induce expression of the intriguing candidates.

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Metabolomics offers comprehensive profiles that can be scanned for novelty to ensure judicious investment of effort in compound purification and structural analysis, reducing the rate of rediscovery.

These strategies to enhance the discovery rate are combined in Tiny Earth,[122] a global coalition of instructors and students dedicated to antibiotic discovery in a research course. Taking advantage of the breadth of soils that the students access, Tiny Earthlings are sampling soils on five continents that have never before been screened for antibiotic-producing bacteria. The students' ingenuity leads them to culture conditions that have not been used for isolation or antibiotic production. The sheer size of the Tiny Earth network (as many as 10 000 students take the course per year) should also enhance the rate of discovery. Chemical analysis of the isolates from the Tiny Earth classrooms around the world has just begun, but it is expected to provide a test of the strategies we have discussed here. Many other researchers around the world are evaluating methods for increasing the rate of discovery. Productive combinations of factors will no doubt lead to great discoveries, which will then rekindle interest in small molecules from natural environments.

4. Conflicts of interest

J. H. holds equity in Wacasa, Inc.

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