



Cite this: *New J. Chem.*, 2021, 45, 22639

Received 31st August 2021,  
Accepted 1st November 2021

DOI: 10.1039/d1nj04173h

rsc.li/njc

## When crown ethers finally click: novel, click-assembled, fluorescent enantiopure pyridino-crown ether-based chemosensors – and an *N*-2-aryl-1,2,3-triazole containing one†

Balázs Szemenyei, <sup>a</sup> Mira Malmosi, <sup>a</sup> Dávid Pál, <sup>a</sup> Péter Baranyai, <sup>b</sup> László Drahos, <sup>c</sup> Ildikó Móczár <sup>\*a</sup> and Péter Huszthy <sup>\*a</sup>

Three novel, click-assembled fluorescent pyridino-18-crown-6 ethers have been synthesized for enantiomeric sensing. We also prepared new azide- and ethynyl-substituted pyridino-18-crown-6 ethers as their precursors, which open the way for further interesting applications using click chemistry. An optically active pyridino-18-crown-6 ether containing an *N*-2-aryl-1,2,3-triazole type fluorophore unit was also synthesized *via* post-triazole arylation. These four fluorescent sensor molecules were studied in terms of their optical properties as well as their enantiomeric recognition abilities toward the hydrogen perchlorate salts of 1-phenylethylamine, 1-(1-naphthyl)ethylamine, phenylglycine methyl ester and phenylalanine methyl ester in acetonitrile.

## Introduction

Enantiomeric differentiation of chiral primary ammonium salts is of great importance,<sup>1,2</sup> because organic molecules with similar structures are often drug molecules. Many times only one enantiomer of these molecules gives the biologically desired effect, and the other one might be responsible for dangerous side effects.<sup>3</sup> As purity requirements in the pharmaceutical and agrochemical industries are becoming stricter,<sup>4</sup> sensor molecules which enable selective and sensitive recognition of enantiomers have gained much interest.<sup>5</sup> Fluorescence spectroscopy is a highly useful tool for sensing enantiomers, because of its selectivity and high sensitivity;<sup>6</sup> therefore, fluorophore units are widely used in chiral chemosensors,<sup>5,7–12</sup> including fluorescent optically active crown ethers.<sup>13–17</sup> Chiral crown ethers containing a pyridine moiety were also prepared by Izatt and Bradshaw *et al.*, and these macrocycles showed excellent selectivity toward the enantiomers of protonated primary amines, amino acid esters and amino alcohols.<sup>18–20</sup> Therefore, they are widely used as sensor molecules<sup>21</sup> as well as selector molecules in chiral chromatography.<sup>22,23</sup>

We set the goal to synthesize fluorescent enantiopure pyridino-crown ether-based sensor molecules. We have already

prepared and examined optically active pyridino-crown ethers having anthracene,<sup>24</sup> and more recently, benzothiazole<sup>14</sup> fluorophore units connected directly or through a linker, respectively, to position 4 of the pyridine ring. Herein, we report the introduction of azide and ethynyl groups to position 4 of the pyridine ring, as a preparative novelty. These results opened the way for click chemistry, which is not only useful for incorporating new kinds of fluorophore moieties, but also enables other interesting applications in the future.

There is a wide range of reported fluorophores which can be assembled by a click reaction.<sup>25–30</sup> Fluorescence can be induced *via* a click reaction from two non-fluorescent precursors.<sup>27</sup> Attaching an electron-donating group to the electron-deficient pyridine unit through a triazole moiety creates push-pull fluorophores,<sup>30</sup> which can, in some cases, even be utilized as ratiometric sensors.<sup>25</sup> Furthermore, these fluorophores are more easily synthesized than 5-aryl-2-(4-pyridyl)oxazoles (*e.g.*, PYMPO), which bear similar photophysical properties.<sup>25</sup> In this work, we synthesized pyridino-crown ethers (*S,S*)-1 and (*S,S*)-2 (Fig. 1) having an *N*-1-aryl-1,2,3-triazole unit, where fluorescence was installed by click chemistry itself. Also, there have been reported examples, where azides or alkynes bearing already fluorescent moieties were connected to macrocycles by a click reaction to obtain chemosensors having various fluorophores,<sup>29</sup> *e.g.*, 1,8-naphthalimide among others.<sup>31,32</sup> Our interest turned to 1,8-naphthalimides for their advantageous properties for sensing, easy synthesis and derivatization,<sup>33</sup> thus we prepared pyridino-crown ether (*S,S*)-3 (Fig. 1) into which a naphthalimide unit was introduced *via* a click reaction. It is known, however, that *N*-2-aryl-1,2,3-triazoles have much stronger fluorescence than

<sup>a</sup> Department of Organic Chemistry and Technology, Budapest University of Technology and Economics, PO Box 91, H-1521 Budapest, Hungary.

E-mail: [moczar.ildiko@vbk.bme.hu](mailto:moczar.ildiko@vbk.bme.hu), [huszthy.peter@vbk.bme.hu](mailto:huszthy.peter@vbk.bme.hu)

<sup>b</sup> Wigner Research Centre for Physics, PO Box 49, H-1525 Budapest, Hungary

<sup>c</sup> Institute of Organic Chemistry, Research Centre for Natural Sciences, PO Box 286, H-1519 Budapest, Hungary

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1nj04173h



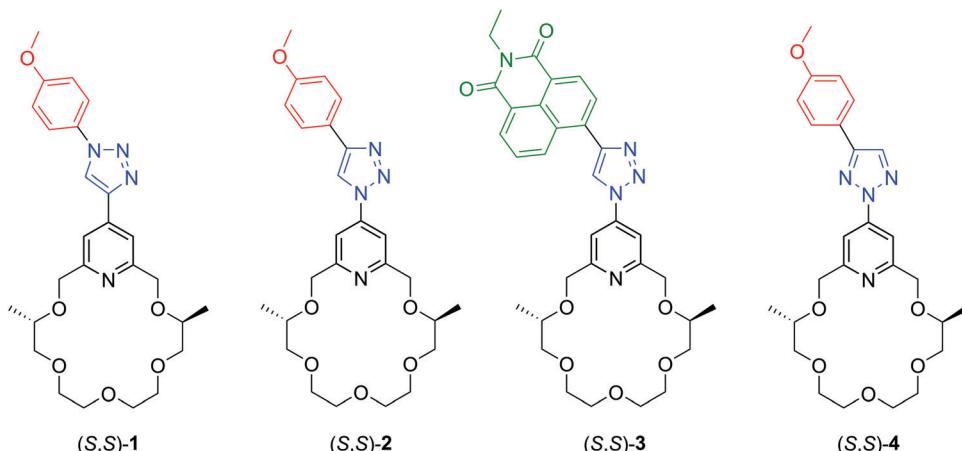


Fig. 1 Schematics of novel fluorescent pyridino-crown ether type chemosensors containing a triazole unit.

*N*-1-aryl-1,2,3-triazoles,<sup>34–38</sup> making the incorporation of the former type fluorophores into chemosensors advantageous.<sup>36</sup> Motivated by this, we also synthesized fluorescent sensor molecule (S,S)-4 (Fig. 1), which contains an *N*-2-aryl-1,2,3-triazole type fluorophore.

We studied the optical properties, especially fluorescence changes, of these four ligands [(S,S)-1–(S,S)-4] upon complexation with the enantiomers of protonated chiral primary amines and amino acid esters, focusing on their ability to discriminate between the enantiomers of these ammonium guests.

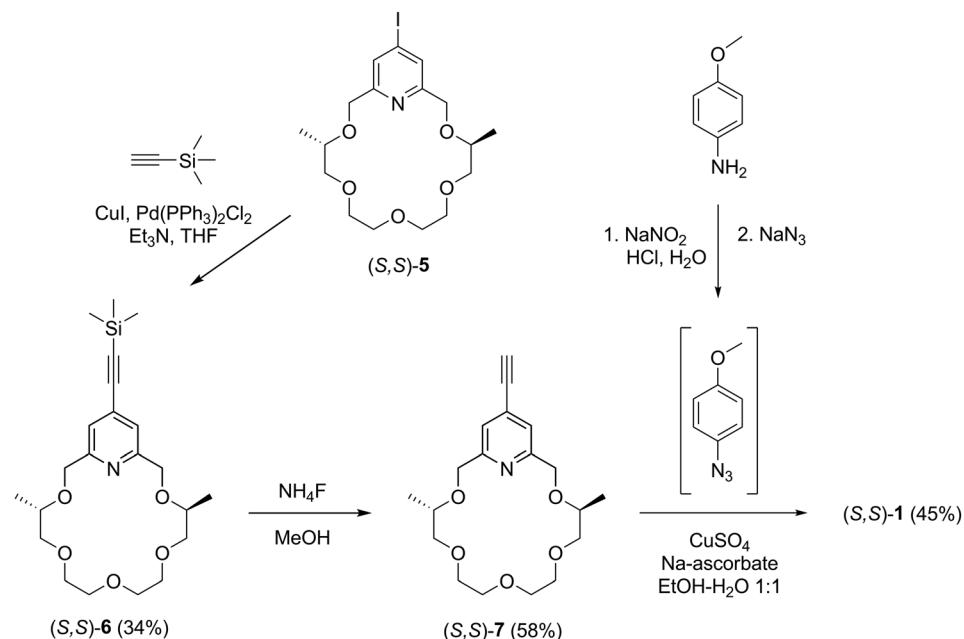
## Results and discussion

### Synthesis

The synthesis of pyridino-crown ether type sensor molecules (S,S)-1–(S,S)-4 containing a triazole unit is outlined in Schemes 1–3.

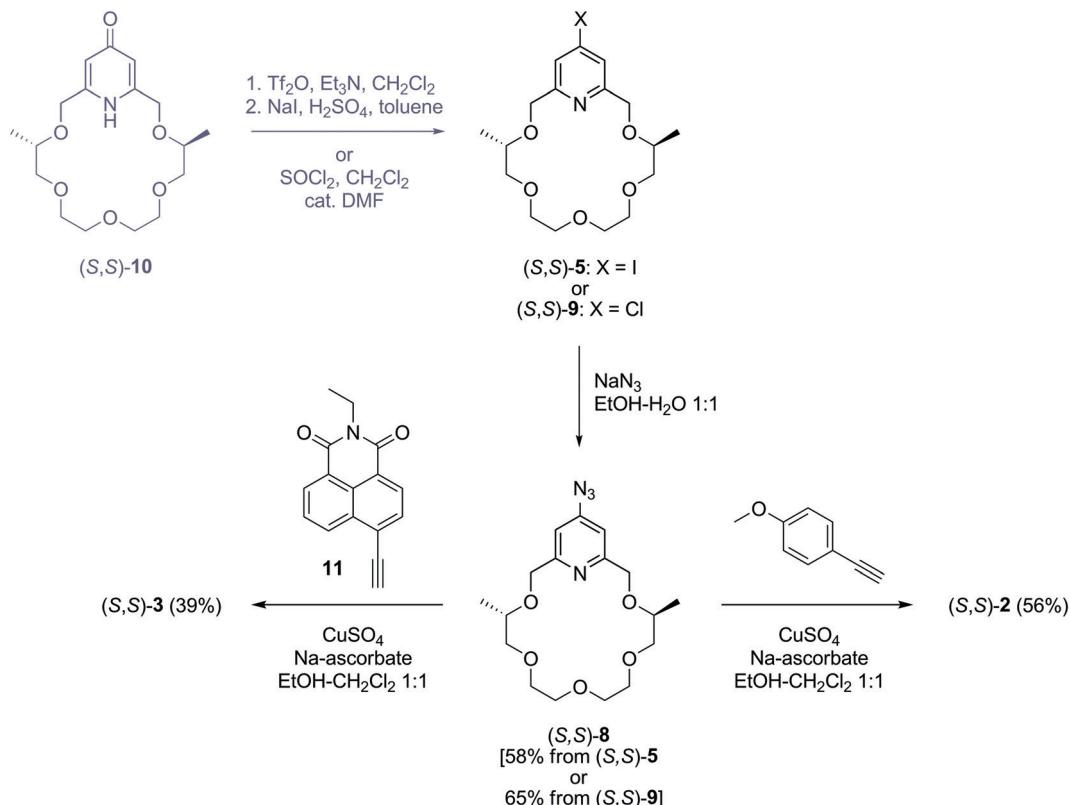
Iodopyridino-crown ether (S,S)-5<sup>22</sup> was reacted with ethynyltrimethylsilane by Sonogashira coupling according to the reported methods<sup>31,39</sup> to give crown ether derivative (S,S)-6 (Scheme 1). The trimethylsilyl protecting group was removed by ammonium fluoride in methanol to yield ethynyl-substituted pyridino-crown ether (S,S)-7. Chemosensor (S,S)-1 was synthesized from ethynyl derivative (S,S)-7 by a click reaction in a mixture of ethanol–water (1:1)<sup>40</sup> using a copper(II) sulfate–sodium ascorbate system as a catalyst and 4-methoxyphenyl azide as a reagent, which was prepared<sup>41</sup> *in situ* from 4-methoxyaniline by diazotization.

Azidopyridino-crown ether (S,S)-8 was prepared from iodopyridino-crown ether (S,S)-5<sup>22</sup> or, alternatively, from chloropyridino-crown ether (S,S)-9<sup>22</sup> with sodium azide in a mixture of ethanol–water (1:1) (Scheme 2) according to the method reported<sup>40</sup> for the synthesis of 4-azidopyridine.

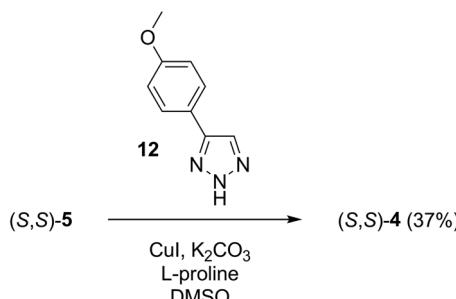


Scheme 1 Synthesis of sensor molecule (S,S)-1.





Scheme 2 Synthesis of sensor molecules (S,S)-2 and (S,S)-3.



Scheme 3 Synthesis of sensor molecule (S,S)-4.

We obtained azide (S,S)-8 with a slightly better yield from chloro compound (S,S)-9 than from iodo compound (S,S)-5. This is in accordance with the well-known fact that chlorine is displaced easier by an aromatic nucleophilic substitution than iodine. Moreover, if we take into account that chloro compound (S,S)-9 can be prepared from pyridono-crown ether (S,S)-10 with a better yield (74%) than using two steps to get iodo compound (S,S)-5 with 56% yield,<sup>22</sup> the overall yield of azide (S,S)-8 from macrocycle (S,S)-10 is much better for the “chloro compound” route (48% in two steps) than for the “iodo compound” one (32% in three steps).

Chemosensors (S,S)-2 and (S,S)-3 were synthesized in click reactions by reacting azide derivative (S,S)-8 with 4-ethynylanisole and naphthalimide derivative 11, respectively, in a mixture of

ethanol–dichloromethane (1 : 1)<sup>28</sup> using a copper(II) sulfate–sodium ascorbate system as a catalyst (Scheme 2). Naphthalimide derivative 11 was obtained as described in the literature starting from 4-bromo-1,8-naphthalic anhydride.<sup>31</sup>

The synthesis of triazole 12 was performed according to the reported procedures starting from 4-hydroxybenzaldehyde.<sup>42–44</sup> The direct triazole arylation reaction<sup>35</sup> of the former compound by iodopyridino-crown ether (S,S)-5<sup>22</sup> in DMSO in the presence of potassium carbonate as a base afforded sensor molecule (S,S)-4 (Scheme 3). The formation of some chemosensor (S,S)-2 as a byproduct was also observed in this reaction, of which separation by chromatography was possible.

### Structure elucidation

Sensor molecules (S,S)-1–(S,S)-4 were characterized by <sup>1</sup>H NMR, <sup>13</sup>C NMR, IR and HRMS methods (see the Experimental section and ESI<sup>†</sup>). The connectivity of the triazole nitrogen atom for sensor molecule (S,S)-4 was proved by a <sup>1</sup>H–<sup>1</sup>H ROESY experiment. In this case, one of the three possible isomers is ligand (S,S)-2, which formed as a byproduct, and its physical and spectroscopic properties are different from those of ligand (S,S)-4. The absence of a ROE cross-peak between the pyridine proton (7.99 ppm) and phenyl proton (7.84 ppm) excludes the third isomer (Fig. 2). It is well-known that Cu(I)-catalysed azide–alkyne cycloaddition reactions yield 1,4-disubstituted 1,2,3-triazoles with exquisite selectivity.<sup>45–47</sup> The 2D NMR spectra taken for click-derived ligands (S,S)-1–(S,S)-3 gave the expected results (see the ESI<sup>†</sup>).



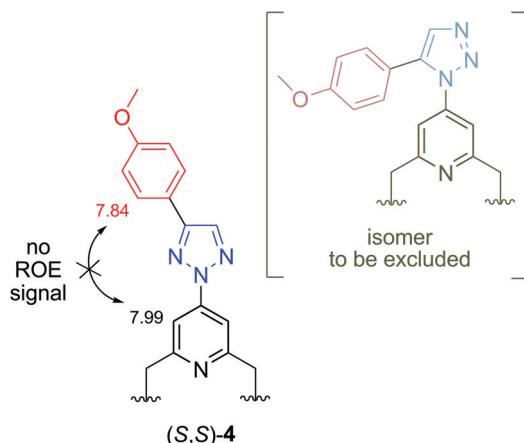


Fig. 2 Part of the structure of sensor molecule (S,S)-4 with proton signal values (ppm) and an alternative structure excluded by a 2D ROESY experiment.

### Optical properties

First, we studied the optical (absorption and fluorescence) properties of the sensor molecules, which are summarized in Table 1.

*N*-1-Aryl triazoles (S,S)-1 and (S,S)-2 have absorption maxima at shorter wavelengths than *N*-2-aryl triazole (S,S)-4. Shi *et al.* found that while 1,4-disubstituted *N*-1-aryl-1,2,3-triazoles prefer twisted conformations around the C–N bond, 2,4-disubstituted *N*-2-aryl-1,2,3-triazoles favor a co-planar conformation with the possibility for a higher degree of conjugation,<sup>34,36</sup> which can be an explanation for the latter statement. The most red-shifted absorption band was recorded in the case of ligand (S,S)-3 having a naphthalimide moiety with a more extended  $\pi$  system. Regarding the position of the absorption bands, reported fluorescent pyridino-crown ethers having a benzothiazole unit ( $\lambda_{\text{abs,max}} = 300\text{--}320\text{ nm}$ )<sup>14</sup> are similar to ligand (S,S)-4, and those bearing an anthracene moiety ( $\lambda_{\text{abs,max}}$  around 365 nm)<sup>24</sup> resemble sensor molecule (S,S)-3. Among 1,2,3-triazole-, benzothiazole-<sup>14</sup> and anthracene-containing<sup>24</sup> pyridino-crown ethers, the latter have by far the smallest molar absorption coefficients at the lowest energy absorption maxima.

It can be observed that while the fluorescence quantum yields of *N*-1-aryl triazoles (S,S)-1 and (S,S)-2 are quite low, that of *N*-2-aryl triazole (S,S)-4 is significantly higher, in accordance with a series of reported examples.<sup>26,34–37</sup> It can also be stated that the difference in the connection of aryl groups to N-1 and C-4 atoms in triazoles (S,S)-1 and (S,S)-2 hardly influenced the fluorescence quantum yield. For comparison, benzothiazole-

containing pyridino-crown ethers have fluorescence quantum yields of 0.007–0.135 (in acetonitrile),<sup>14</sup> and those with an anthracene unit showed 0.58 and 0.60 (in acetonitrile).<sup>24</sup> Therefore, the strongest fluorescence (and brightness) was achieved with ligands (S,S)-4 and (S,S)-3 among the new and reported<sup>14,24</sup> fluorescent pyridino-crown ether-based enantioselective sensors.

### Enantiomeric recognition studies

The chiral recognition abilities of pyridino-crown ethers (S,S)-1–(S,S)-4 toward the enantiomers of 1-phenylethylamine hydrogen perchlorate (PEA), 1-(1-naphthyl)ethylamine hydrogen perchlorate (NEA), phenylglycine methyl ester hydrogen perchlorate (PGME) and phenylalanine methyl ester hydrogen perchlorate (PAME) (Fig. 3) were studied in acetonitrile by UV-vis and fluorescence spectroscopies.

The absorption spectra showed no [in the case of (S,S)-3] or little to moderate [in the cases of (S,S)-1, (S,S)-2 and (S,S)-4] changes upon the addition of the chiral ammonium guests (Fig. 4), apart from the partial to total overlap of the longest wavelength bands of macrocycles (S,S)-1, (S,S)-2 and (S,S)-4 with that of NEA. Based on the absorption changes, crown ethers (S,S)-1 and (S,S)-4 also revealed slight protonation (besides the complexation) with PGME and PAME, which could be neglected during the evaluation of fluorescence titration data (see below).

Compared to the above spectral changes, the fluorescence ones were much more pronounced in most cases, which were used to determine the stability constants of the complexes by global nonlinear regression analysis. The titration series of emission spectra could be fitted satisfactorily using a 1:1 complex formation model.

In the case of sensor molecule (S,S)-2, significant fluorescence quenching was observed upon the addition of the optically active salts (Fig. 5A), rendering appreciable enantioselectivity toward NEA and PEA, but only slight recognition to the enantiomers of PGME and practically no for PAME (Table 2).

Fluorescence spectral changes did not occur when ligand (S,S)-3 was titrated with PEA, PGME and PAME, but interestingly about 20% quenching could be detected upon the addition of the enantiomers of NEA (Fig. 5B), which enabled the determination of the complex stability constants (Table 2). Probably, the electronic signal transmission between the electron-deficient pyridine and similarly electron-deficient naphthalimide units is hindered, thereby resulting in no fluorescence changes by the addition of PEA, PGME and PAME (presumably complexation with these salts also took place similarly to other pyridino-crown ethers<sup>14,24</sup>). However, the extended aromatic moiety of NEA may show a little overlap with the triazole ring, so a limited emission change can be caused by a direct  $\pi$ – $\pi$  interaction between the fluorophore unit and the guest molecule. Appreciable selectivity between the enantiomers was also experienced in this case (Table 2).

Titration of pyridino-crown ether (S,S)-4 with all the four chiral salts induced significant fluorescence quenching (Fig. 5C) similarly to crown ether (S,S)-2. The extent of enantiomeric differentiation was appreciable for NEA and PEA, moderate toward PGME, and practically zero for PAME (Table 2).

Table 1 UV-vis and fluorescence spectroscopic data of the free sensor molecules in MeCN

	(S,S)-1	(S,S)-2	(S,S)-3	(S,S)-4
$\lambda_{\text{abs,max}}$ (nm)	271	257	240, 355	289, 312
$\epsilon_{\text{max}} (\times 10^4 \text{ M}^{-1} \text{ cm}^{-1})$	2.7	2.4	4.4, 1.8	2.0, 2.5
$\lambda_{\text{em,max}}$ (nm)	384	405	428	378
$\Phi_f$	0.018	0.022	0.81	0.93



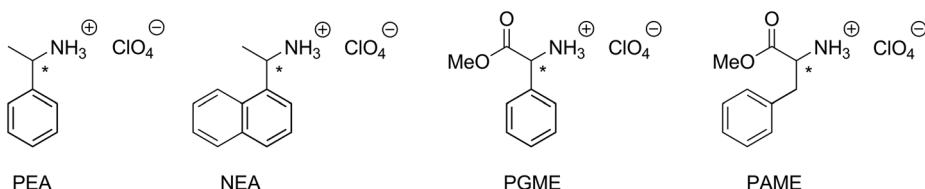


Fig. 3 Chiral primary ammonium salts used in the enantiomeric recognition studies.

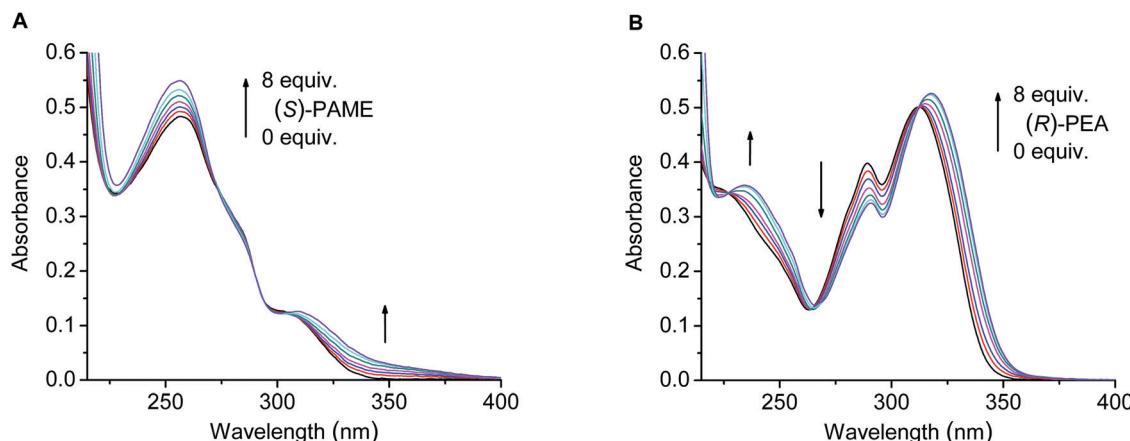


Fig. 4 Series of absorption spectra upon the titration of (S,S)-2 (20  $\mu$ M) with (S)-PAME (0, 0.2, 0.5, 1, 2, 4, 8 equiv.) (A) and (S,S)-4 (20  $\mu$ M) with (R)-PEA (0, 0.2, 0.5, 1, 2, 4, 8 equiv.) (B) in MeCN.

Free sensor molecule (S,S)-1 showed an emission peak at 384 nm, which decreased to a great extent when the ammonium salts were added; however, simultaneously, a new, much more intense band emerged with a maximum of around 580 nm (Fig. 5D). The latter spectral shape corresponds to the emission spectrum of protonated ligand (S,S)-1 (Fig. 6A). Since these guests caused no (PEA and NEA) or negligible (PGME and PAME) protonation in the ground state (Fig. 6B), the protonation of ligand (S,S)-1 took place only in the excited state resulting in emission from the protonated species (Fig. 6A). Presumably, the protonation process occurred from the hydrogen-bonded complexes, because the series of spectra for fluorescence titration could be fitted satisfactorily using a 1:1 complexation model (Table 2). It should be noted here that this phenomenon (*i.e.*, proton transfer in the complex upon excitation) has also been observed in the cases of pyridino-crown ethers having a benzothiazole unit.<sup>14</sup> Additionally, it is also worth mentioning that host (S,S)-1 has a similar behavior to its reported<sup>25</sup> analogous pyridine derivative having no crown ether macroring, regarding the appearance of a new, red-shifted, high-intensity emission band upon protonation.

A problem which was encountered with chemosensor (S,S)-1 is that it does not absorb light above 325 nm, which means that it could not be selectively excited when titrated with NEA, which absorbs all the way up to 325 nm. As a consequence of this, distorted titration curves were obtained, which did not allow the determination of stability constants. Macrocycle (S,S)-1 showed moderate and no enantioselectivity toward PGME and

PAME, respectively, but it exhibited considerable recognition toward the enantiomers of PEA (Table 2).

Considering the results for chemosensors (S,S)-1–(S,S)-4 in general, the highest enantioselectivities were found toward NEA (Table 2), which can be attributed to the extended  $\pi$  system of the guest conferring stronger  $\pi$ – $\pi$  interactions.<sup>21</sup> In the cases of crown ethers (S,S)-3 and (S,S)-4, these  $\Delta \log K$  values (0.45 and 0.44, Table 2) are slightly or significantly higher than those for similar dimethyl-substituted pyridino-crown ethers having an anthracene (0.38)<sup>24</sup> or a benzothiazole unit (0.30, 0.25).<sup>14</sup>

Regarding analogous ligands (S,S)-1, (S,S)-2 and (S,S)-4, it can be seen that the connectivity of the triazole moiety to the pyridine ring had an influence on the stability of complexes (Table 2). Namely, the equilibrium constants for each optically active guest increase in the order of macrocycles (S,S)-2, (S,S)-4 and (S,S)-1. This phenomenon may be explained by different conformational and electronic effects in the aromatic parts of the chemosensors, *e.g.*, different dihedral angles between the pyridine and triazole rings<sup>34,36</sup> or different electron densities in the pyridine rings. However, the enantioselectivities were quite similar for a given chiral salt [where it could be determined and including (S,S)-3 also], and only ligand (S,S)-2 showed slightly smaller recognition for the enantiomers of NEA and PGME (Table 2).

A pair of titration curves of sensor molecule (S,S)-4 with the enantiomers of NEA is shown in Fig. 7A. Larger fluorescence quenching when adding (R)-NEA is clearly visible, indicating a heterochiral preference. Similarly, the complex formation of



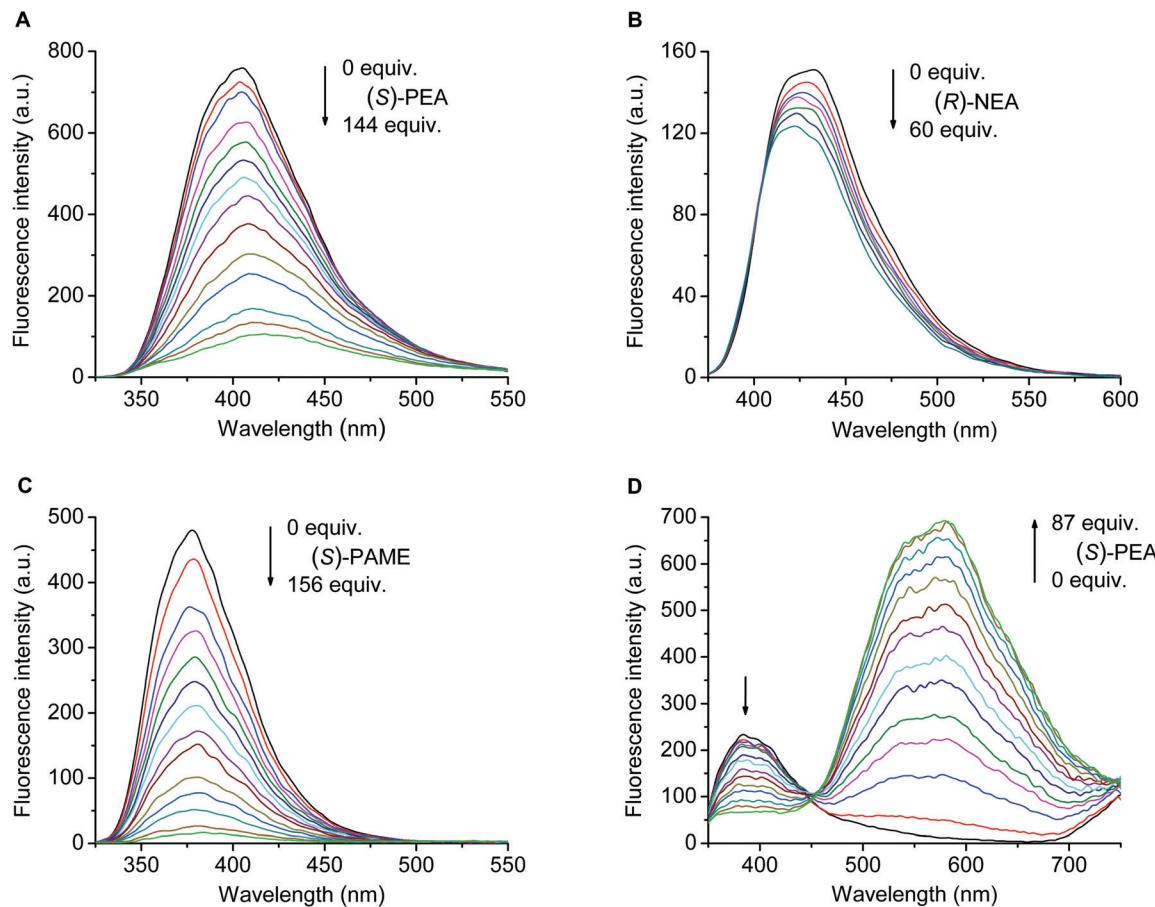


Fig. 5 Series of fluorescence emission spectra upon the titration of (S,S)-2 (5  $\mu$ M) with (S)-PEA (0, 0.4, 0.8, 1.6, 2.4, 3.2, 4.4, 6.4, 9.6, 16, 24, 48, 80, 144 equiv.),  $\lambda_{\text{ex}} = 300$  nm (A), (S,S)-3 (2  $\mu$ M) with (R)-NEA (0, 1.5, 4, 6, 10, 20, 60 equiv.),  $\lambda_{\text{ex}} = 355$  nm (B), (S,S)-4 (5  $\mu$ M) with (S)-PAME (0, 0.5, 1, 1.5, 2, 3, 4, 6, 8, 12, 20, 36, 76, 156 equiv.),  $\lambda_{\text{ex}} = 300$  nm (C), and (S,S)-1 (2  $\mu$ M) with (S)-PEA (0, 0.5, 1, 1.5, 2, 3, 4, 6, 8, 12, 17, 27, 47, 87 equiv.),  $\lambda_{\text{ex}} = 265$  nm (D) in MeCN.

Table 2 Stability constants for complexes of (S,S)-1–(S,S)-4 with the enantiomers of chiral primary ammonium salts and the degrees of enantiomeric differentiation in MeCN<sup>a</sup>

	(S,S)-1		(S,S)-2		(S,S)-3		(S,S)-4	
	$\log K$	$\Delta \log K$						
(R)-PEA	5.48 $\pm$ 0.03	0.28	4.83 $\pm$ 0.02	0.28	c	c	5.25 $\pm$ 0.02	0.28
(S)-PEA	5.20 $\pm$ 0.02	b	4.55 $\pm$ 0.03	b	c	c	4.97 $\pm$ 0.02	0.44
(R)-NEA	b	b	5.01 $\pm$ 0.03	0.37	5.13 $\pm$ 0.03	0.45	5.37 $\pm$ 0.02	0.44
(S)-NEA	b	b	4.64 $\pm$ 0.03	b	4.68 $\pm$ 0.04	c	4.93 $\pm$ 0.02	b
(R)-PGME	5.67 $\pm$ 0.04	-0.13	4.74 $\pm$ 0.03	-0.07	c	c	5.30 $\pm$ 0.03	-0.11
(S)-PGME	5.80 $\pm$ 0.04	b	4.81 $\pm$ 0.03	b	c	c	5.41 $\pm$ 0.04	b
(R)-PAME	5.48 $\pm$ 0.03	-0.03	4.23 $\pm$ 0.05	-0.05	c	c	4.90 $\pm$ 0.02	0.01
(S)-PAME	5.51 $\pm$ 0.03	b	4.28 $\pm$ 0.06	b	c	c	4.89 $\pm$ 0.03	b

<sup>a</sup> Complex stability constants ( $K$ ) are given in  $M^{-1}$ . <sup>b</sup> Complex stability constants ( $K$ ) could not be determined because of the total spectral overlap of the host and the guest. <sup>c</sup> No spectral changes occurred.

host (S,S)-1 with (R)-PEA caused more significant fluorescence enhancement (on the basis of the excited state proton transfer in the complex) than with the (S)-enantiomer (Fig. 7B). In the latter case, the quenching processes at 384 nm were similar to those in Fig. 7A, but exhibited a smaller difference.

Toward the enantiomers of PGME, apparently inverse discrimination was observed than for NEA or PEA. However,

this is because the absolute configuration of PGME is reversed according to the CIP rules. Thus, (S)-PGME has the same spatial arrangement of substituents (aromatic, amino and the third groups) around the chiral center as (R)-NEA or (R)-PEA.<sup>14,24</sup> The enantiomers of PAME were not differentiated by the triazolo macrocycles, similar to the earlier reported pyridino-crown ethers.<sup>14,24</sup> It may be explained by the presence of a methylene



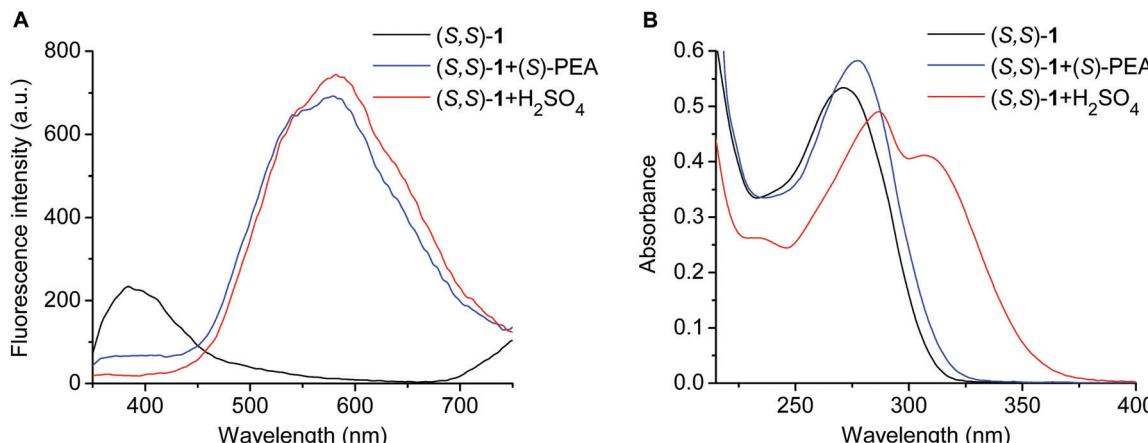


Fig. 6 Fluorescence emission spectra of (S,S)-1 alone (5  $\mu$ M) and the end-points of its titrations with (S)-PEA and H<sub>2</sub>SO<sub>4</sub> in MeCN,  $\lambda_{\text{ex}} = 265$  nm (A). Absorption spectra of (S,S)-1 alone (20  $\mu$ M) and the end-points of its titrations with (S)-PEA and H<sub>2</sub>SO<sub>4</sub> in MeCN (B).

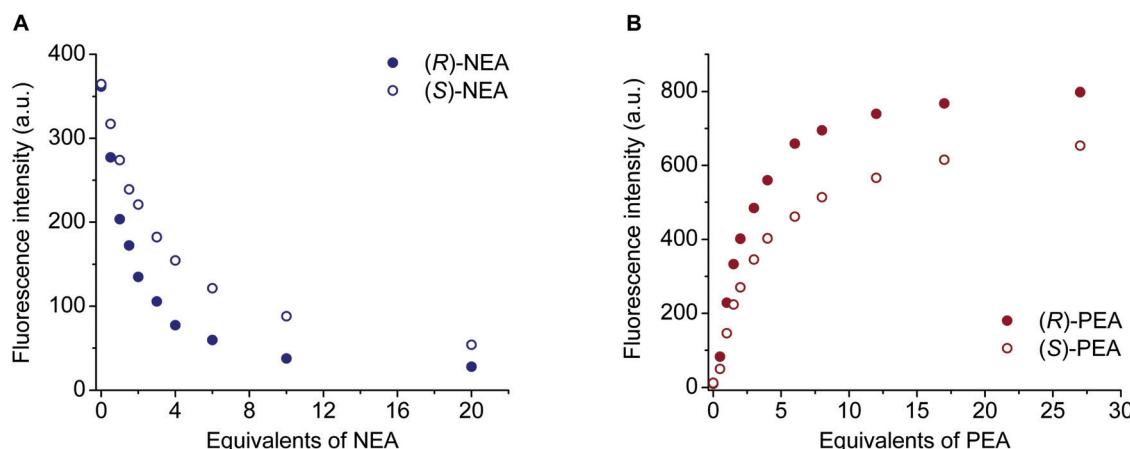


Fig. 7 Titration curves of (S,S)-4 (5  $\mu$ M) with the enantiomers of NEA (0, 0.5, 1, 2, 4, 5, 10, 20 equiv.) at 378 nm,  $\lambda_{\text{ex}} = 330$  nm (A) and (S,S)-1 (2  $\mu$ M) with the enantiomers of PEA (0, 0.5, 1, 1.5, 2, 3, 4, 6, 8, 12, 17, 27 equiv.) at 580 nm,  $\lambda_{\text{ex}} = 265$  nm (B) in MeCN.

spacer between the aromatic group and the stereogenic center.<sup>14,24</sup>

## Conclusions

The synthesis of crown ether-based fluorescent sensors (S,S)-1–(S,S)-4 containing a triazole moiety has been achieved. Macro-cycles (S,S)-1–(S,S)-3 were obtained by a click reaction, enabled by the synthesis of new pyridino-crown ether derivatives having an ethynyl or an azido group [(S,S)-7 and (S,S)-8]. Pyridino-crown ether (S,S)-4 was prepared by direct *N*-arylation.

Enantiomeric recognition of crown ethers (S,S)-1–(S,S)-4 toward the enantiomers of chiral protonated primary aralkylamines and amino acid esters was studied in acetonitrile by fluorescence spectroscopy. Sensor molecules (S,S)-2–(S,S)-4 exhibited fluorescence quenching upon complexation, but in the case of ligand (S,S)-3, just a moderate decrease in emission was experienced and only with NEA. Titrations of crown ether (S,S)-1 were accompanied by the appearance of a second

emission band, due to the excited state proton transfer in the complexes. Appreciable enantioselectivities were observed for NEA and PEA; however, PGME was differentiated only moderately.

## Experimental

### General

The starting materials were purchased from Sigma-Aldrich, Merck and Alfa Aesar. Aluminum oxide 60 F<sub>254</sub> neutral type E plates (Merck) were used for thin-layer chromatography (TLC). Aluminum oxide (neutral, activated, Brockmann I, 40–160  $\mu$ m) was used for column chromatography. The ratios of solvents for the eluents are given in volumes (mL/mL). Solvents were dried and purified according to well-established methods.<sup>48</sup> Evaporations were carried out under reduced pressure.

Melting points were taken on a Boetius micro-melting point apparatus and are uncorrected. Optical rotations were taken on a PerkinElmer 241 polarimeter that was calibrated by measuring



the optical rotations of both enantiomers of menthol. IR spectra were recorded using a Bruker Alpha-T Fourier transform infrared (FT-IR) spectrometer.  $^1\text{H}$  (500 MHz) and  $^{13}\text{C}$  (125 MHz) NMR spectra were obtained using a Bruker DRX-500 Avance spectrometer.  $^1\text{H}$  (300 MHz) and  $^{13}\text{C}$  (75.5 MHz) NMR spectra were recorded using a Bruker 300 Avance spectrometer. 2D NMR spectra were obtained using a Bruker DRX-500 Avance spectrometer. HRMS analyses were performed using a Waters Q-TOF Premier or a Thermo Velos Pro Orbitrap Elite system mass spectrometer in positive ESI mode.

UV-vis spectra were recorded using a Unicam UV4-100 spectrophotometer. Quartz cuvettes with a path length of 1 cm were used. Fluorescence emission spectra were recorded using a PerkinElmer LS 50B luminescent spectrometer and were corrected by the spectrometer software. Quartz cuvettes with a path length of 1 cm were used. Fluorescence quantum yields were determined relative to quinine sulfate ( $\Phi_f = 0.53$  in 0.1 M  $\text{H}_2\text{SO}_4$ ).<sup>6</sup> Enantiomers of PEA, NEA, PGME and PAME were prepared as reported<sup>49</sup> in our laboratory. For fluorescence titrations, the concentrations were 2  $\mu\text{M}$  for *(S,S)*-1 and *(S,S)*-3, and 5  $\mu\text{M}$  for *(S,S)*-2 and *(S,S)*-4. In order to determine the stability constants of complexes by global nonlinear regression analysis, SPECFIT/32<sup>TM</sup> software was used.

### Syntheses and characterizations of new compounds

**(4S,14S)-19-[1-(4-Methoxyphenyl)-1H-1,2,3-triazol-4-yl]-4,14-dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene [(S,S)-1].** To a suspension of 4-methoxyaniline (550 mg, 4.47 mmol) in water (4.5 mL), 37% aqueous HCl solution (2.3 mL, 27 mmol) was added. The mixture was cooled to 0 °C, and a solution of  $\text{NaNO}_2$  (463 mg, 6.71 mmol) in water (11 mL) was added dropwise. After 0.5 h of stirring at 0 °C, a solution of  $\text{NaN}_3$  (1.16 g, 17.9 mmol) in water (22 mL) was added dropwise. The reaction mixture was stirred for 3 h at 0 °C, and then it was extracted with ethyl acetate (3 × 50 mL). The combined organic phase was washed with water (30 mL), dried over anhydrous  $\text{MgSO}_4$ , filtered, and the solvent was removed.<sup>41</sup>

To the resulting crude 4-methoxyphenyl azide, ethynylpyridino-crown ether *(S,S)*-7 (156 mg, 0.447 mmol) was added, followed by  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$  (1.0 mg, 0.0040 mmol), sodium ascorbate (5.3 mg, 0.027 mmol), ethanol (1.25 mL) and water (1.25 mL), and the mixture was stirred at rt for 20 h. The solvent was removed at rt, and the residue was dissolved in a mixture of ethyl acetate (50 mL) and water (50 mL). The phases were shaken well and separated. The aqueous phase was further extracted with ethyl acetate (2 × 50 mL). The combined organic phase was dried over anhydrous  $\text{MgSO}_4$ , filtered, and evaporated. The crude product was purified by column chromatography on alumina using ethanol-toluene 1:80 mixture as an eluent, followed by recrystallization from toluene-hexane to give *(S,S)*-1 (76 mg, 34%) as an off-white powder.

$R_f$ : 0.30 (alumina TLC, EtOH-toluene 1:40); m.p.: 101–104 °C;  $[\alpha]_D^{25} = +24.2$  ( $c = 1.15$ ,  $\text{CHCl}_3$ ); IR (KBr)  $\tilde{\nu}_{\text{max}}$  (cm<sup>-1</sup>) 3137, 3089, 2967, 2894, 2862, 1618, 1595, 1567, 1521, 1471, 1446, 1392, 1369, 1353, 1331, 1306, 1259, 1195, 1165, 1115,

1098, 1068, 1042, 1029, 1007, 922, 882, 865, 836, 817, 798, 697, 624, 541, 524, 496, 465, 407;  $^1\text{H}$  NMR (500 MHz, acetone- $d_6$ )  $\delta$  (ppm) 1.16 (d,  $J = 6$  Hz, 6H), 3.39–3.58 (m, 12H), 3.79–3.87 (m, 2H), 3.89 (s, 3H), the diastereotopic benzylic type  $-\text{CH}_2-$  protons give an AB quartet:  $\delta_A$  4.79 and  $\delta_B$  4.82 ( $J_{\text{AB}} = 13$  Hz, 4H), 7.16 (d,  $J = 9$  Hz, 2H), 7.87 (s, 2H), 7.88 (d,  $J = 9$  Hz, 2H), 9.09 (s, 1H);  $^{13}\text{C}$  NMR (75.5 MHz, acetone- $d_6$ )  $\delta$  (ppm) 17.75, 56.06, 71.17, 71.31, 72.54, 74.43, 76.44, 115.69, 117.13, 121.41, 122.80, 131.34, 139.87, 146.63, 160.50, 160.93; HRMS  $m/z$  (M + H)<sup>+</sup> found: 499.2541,  $\text{C}_{26}\text{H}_{35}\text{N}_4\text{O}_6^+$  requires 499.2551.

**(4S,14S)-19-[4-(4-Methoxyphenyl)-1H-1,2,3-triazol-4-yl]-4,14-dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene [(S,S)-2].** To a solution of azidopyridino-crown ether *(S,S)*-8 (80 mg, 0.23 mmol) in a mixture of dichloromethane (0.5 mL) and ethanol (0.5 mL), 4-ethynylanisole (46 mg, 0.34 mmol), sodium ascorbate (2.7 mg, 0.014 mmol) and freshly dried  $\text{CuSO}_4$  (0.4 mg, 0.0025 mmol) were added, and the resulting mixture was stirred at rt for 2 days. The solvent was removed, and the residue was washed into a separatory funnel with dichloromethane (10 mL) and water (10 mL). The phases were shaken well and separated. The aqueous phase was extracted with dichloromethane (2 × 10 mL). The combined organic phase was dried over anhydrous  $\text{MgSO}_4$ , filtered, and evaporated. The crude product was purified by column chromatography on alumina using ethanol-toluene 1:80 mixture as an eluent, followed by recrystallization from toluene-hexane to give *(S,S)*-2 (64 mg, 56%) as a white powder.

$R_f$ : 0.34 (alumina TLC, EtOH-toluene 1:40); m.p.: 99–103 °C;  $[\alpha]_D^{25} = +19.0$  ( $c = 1.00$ ,  $\text{CHCl}_3$ ); IR (KBr)  $\tilde{\nu}_{\text{max}}$  (cm<sup>-1</sup>) 3143, 3124, 3083, 3044, 2965, 2907, 2868, 1619, 1600, 1567, 1505, 1476, 1448, 1416, 1368, 1353, 1330, 1309, 1287, 1260, 1100, 1068, 1055, 1020, 944, 922, 870, 838, 818, 796, 736, 696, 636, 617, 553, 541, 528, 495, 404;  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm) 1.19 (d,  $J = 6$  Hz, 6H), 3.43–3.65 (m, 12H), 3.82–3.90 (m, 2H), 3.83 (s, 3H), the diastereotopic benzylic type  $-\text{CH}_2-$  protons give an AB quartet:  $\delta_A$  4.89 and  $\delta_B$  4.93 ( $J_{\text{AB}} = 14$  Hz, 4H), 6.97 (d,  $J = 8$  Hz, 2H), 7.71 (s, 2H), 7.82 (d,  $J = 8$  Hz, 2H), 8.25 (s, 1H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm) 17.16, 55.48, 70.79, 71.02, 71.54, 74.31, 76.33, 110.10, 114.52, 116.28, 122.50, 127.44, 144.18, 148.85, 160.17, 161.50; HRMS  $m/z$  (M + H)<sup>+</sup> found: 499.2572,  $\text{C}_{26}\text{H}_{35}\text{N}_4\text{O}_6^+$  requires: 499.2551.

**6-[1-[(4S,14S)-4,14-Dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene-19-yl]-1H-1,2,3-triazol-4-yl]-2-ethyl-1H-benzo[de]isoquinoline-1,3(2H)-dione [(S,S)-3].** To a solution of azidopyridino-crown ether *(S,S)*-8 (92 mg, 0.25 mmol) in a mixture of dichloromethane (0.75 mL) and ethanol (0.75 mL), naphthalimide derivative **11**<sup>31</sup> (62 mg, 0.25 mmol) was added, followed by sodium ascorbate (3.0 mg, 0.015 mmol) and freshly dried  $\text{CuSO}_4$  (0.5 mg, 0.003 mmol). The reaction mixture was stirred at rt for 2 days. The solvent was removed, and the residue was washed into a separatory funnel with dichloromethane (10 mL) and water (10 mL). The phases were shaken well and separated. The aqueous phase was extracted with dichloromethane (2 × 10 mL). The combined organic phase was dried over anhydrous  $\text{MgSO}_4$ , filtered, and evaporated. The crude product was purified by column chromatography on



alumina using ethanol–toluene 1:80 mixture as an eluent, followed by recrystallization from toluene–hexane to give (*S,S*)-3 (60 mg, 39%) as yellowish crystals.

*R*<sub>f</sub>: 0.34 (alumina TLC, EtOH–toluene 1:40); m.p.: 158–159 °C;  $[\alpha]_D^{25} = +15.0$  (*c* = 0.91, CHCl<sub>3</sub>); IR (KBr)  $\tilde{\nu}_{\text{max}}$  (cm<sup>−1</sup>) 3344, 3308, 3170, 3088, 2973, 2926, 2874, 1700, 1665, 1588, 1543, 1510, 1475, 1462, 1445, 1383, 1371, 1342, 1244, 1222, 1110, 1065, 1048, 1025, 893, 867, 819, 785, 693, 659, 584, 490, 446, 415; <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>CN)  $\delta$  (ppm) 1.18 (d, *J* = 6 Hz, 6H), 1.25 (t, *J* = 7 Hz, 3H), 3.39–3.52 (m, 12H), 3.76–3.86 (m, 2H), 4.08 (q, *J* = 7 Hz, 2H), 4.73–4.82 (m, 4H), 7.70 (t, *J* = 8 Hz, 1H), 7.77 (s, 2H), 7.93 (d, *J* = 7 Hz, 1H), 8.38 (d, *J* = 7 Hz, 1H), 8.41 (d, *J* = 7 Hz, 1H), 8.82 (s, 1H), 8.88 (d, *J* = 8 Hz, 1H); <sup>13</sup>C NMR (125 MHz, CD<sub>3</sub>CN)  $\delta$  (ppm) 13.52, 17.14, 36.11, 71.09, 71.29, 71.80, 74.91, 76.29, 111.64, 123.66, 123.88, 128.35, 128.41, 129.36, 129.61, 130.99, 131.69, 133.11, 134.04, 134.21, 145.01, 147.15, 161.92, 164.19, 164.50; HRMS *m/z* (M + H)<sup>+</sup> found: 616.2752, C<sub>33</sub>H<sub>38</sub>N<sub>5</sub>O<sub>7</sub><sup>+</sup> requires 616.2766.

**(4S,14S)-19-[4-(4-Methoxyphenyl)-2*H*-1,2,3-triazol-2-yl]-4,14-dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene** [(*S,S*)-4]. To a solution of iodopyridino-crown ether (*S,S*)-5<sup>22</sup> (200 mg, 0.443 mmol) in dry DMSO (2.5 mL) were added triazole 12<sup>43,44</sup> (124 mg, 0.708 mmol), CuI (5.6 mg, 0.029 mmol), 1-proline (6.8 mg, 0.059 mmol) and K<sub>2</sub>CO<sub>3</sub> (81 mg, 0.59 mmol). The resulting mixture was heated at 110 °C for 20 h. The mixture was washed into a separatory funnel with dichloromethane (50 mL) and water (50 mL). The phases were shaken well and separated. The aqueous phase was extracted with dichloromethane (2 × 50 mL). The combined organic phase was dried over anhydrous MgSO<sub>4</sub>, filtered, and evaporated. The crude product was purified by column chromatography on alumina using first dioxane–hexane 1:6 and then dimethoxyethane–toluene 1:4 mixtures as eluents to give (*S,S*)-4 (84 mg, 37%) as a pale brown oil.

*R*<sub>f</sub>: 0.38 (alumina TLC, EtOH–toluene 1:40);  $[\alpha]_D^{25} = +6.4$  (*c* = 1.00, CHCl<sub>3</sub>); IR (neat)  $\tilde{\nu}_{\text{max}}$  (cm<sup>−1</sup>) 3115, 2967, 2865, 1613, 1595, 1547, 1490, 1460, 1436, 1349, 1292, 1250, 1176, 1107, 1084, 1030, 991, 976, 967, 915, 878, 832, 797, 729, 653, 616, 529, 490; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) 1.22 (d, *J* = 6 Hz, 6H), 3.45–3.66 (m, 12H), 3.85–3.92 (m, 2H), 3.86 (s, 3H), 4.92 (s, 4H), 7.00 (d, *J* = 9 Hz, 2H), 7.84 (d, *J* = 9 Hz, 2H), 7.99 (s, 2H), 8.05 (s, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) 17.23, 55.52, 70.81, 71.06, 71.77, 74.26, 76.30, 109.33, 114.59, 122.09, 127.86, 133.68, 146.83, 149.96, 160.67 (two <sup>13</sup>C signals together); HRMS *m/z* (M + H)<sup>+</sup> found: 499.2542, C<sub>26</sub>H<sub>35</sub>N<sub>4</sub>O<sub>6</sub><sup>+</sup> requires 499.2551.

**(4S,14S)-19-[(Trimethylsilyl)ethynyl]-4,14-dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene** [(*S,S*)-6]. To a solution of iodopyridino-crown ether (*S,S*)-5<sup>22</sup> (280 mg, 0.620 mmol) in dry and peroxide-free THF (7.5 mL) was added ethynyltrimethylsilane (0.42 mL, 295 mg, 3.11 mmol), followed by Pd(PPh<sub>3</sub>)<sub>2</sub>Cl<sub>2</sub> (42 mg, 0.059 mmol), CuI (23 mg, 0.12 mmol) and triethylamine (0.29 mL, 224 mg, 2.31 mmol). The mixture was stirred under Ar at rt for 40 h. The volatile compounds were removed, and the residue was washed into a separatory funnel with ethyl acetate (30 mL) and water (30 mL). The phases were shaken well and separated. The

aqueous phase was extracted with ethyl acetate (2 × 30 mL). The combined organic phase was dried over anhydrous MgSO<sub>4</sub>, filtered, and the solvent was removed. The crude product was purified by column chromatography on alumina using first dioxane–hexane 1:7 and then ethanol–toluene 1:140 mixtures as eluents to give (*S,S*)-6 (90 mg, 34%) as a brown oil.

*R*<sub>f</sub>: 0.39 (alumina TLC, EtOH–toluene 1:40);  $[\alpha]_D^{25} = +12.3$  (*c* = 1.00, CHCl<sub>3</sub>); IR (neat)  $\tilde{\nu}_{\text{max}}$  (cm<sup>−1</sup>) 2966, 2931, 2866, 1597, 1551, 1452, 1411, 1372, 1350, 1339, 1250, 1110, 954, 923, 843, 759, 699, 663, 588; <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) 0.24 (s, 9H), 1.16 (d, *J* = 6 Hz, 6H), 3.36–3.67 (m, 12H), 3.71–3.86 (m, 2H), 4.77 (s, 4H), 7.26 (s, 2H and solvent signal together); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) −0.23, 17.05, 70.68, 70.89, 71.54, 73.89, 76.10, 99.18, 102.70, 122.40, 131.87, 158.67; HRMS *m/z* (M + H)<sup>+</sup> found: 422.2347, C<sub>22</sub>H<sub>36</sub>NO<sub>5</sub>Si<sup>+</sup> requires 422.2357.

**(4S,14S)-19-Ethynyl-4,14-dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene** [(*S,S*)-7]. To a solution of [(trimethylsilyl)ethynyl]pyridino-crown ether (*S,S*)-6 (79.4 mg, 0.188 mmol) in dry methanol (9 mL) was added NH<sub>4</sub>F (25.5 mg, 0.689 mmol), and the mixture was stirred at rt for a day. The solvent was removed, and the residue was washed into a separatory funnel with ethyl acetate (10 mL) and water (10 mL). The phases were shaken well and separated. The aqueous phase was extracted with ethyl acetate (2 × 10 mL). The combined organic phase was dried over anhydrous MgSO<sub>4</sub>, filtered, and evaporated. The crude product was purified by column chromatography on alumina using ethanol–toluene 1:120 mixture as an eluent to give (*S,S*)-7 (38 mg, 58%) as an off-white solid.

*R*<sub>f</sub>: 0.39 (alumina TLC, EtOH–toluene 1:40); m.p.: 74–77 °C;  $[\alpha]_D^{25} = +27.7$  (*c* = 1.00, CHCl<sub>3</sub>); IR (KBr)  $\tilde{\nu}_{\text{max}}$  (cm<sup>−1</sup>) 3418, 3243, 2971, 2901, 2871, 2782, 2108, 1759, 1598, 1551, 1490, 1460, 1411, 1376, 1352, 1337, 1305, 1283, 1251, 1214, 1167, 1110, 1042, 1004, 976, 958, 932, 915, 898, 856, 806, 758, 683, 637, 585, 579, 545, 532, 509, 463; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) 1.18 (d, *J* = 6 Hz, 6H), 3.25 (s, 1H), 3.41–3.68 (m, 12H), 3.74–3.89 (m, 2H), 4.81 (s, 4H), 7.32 (s, 2H); <sup>13</sup>C NMR (75.5 MHz, CDCl<sub>3</sub>)  $\delta$  (ppm) 17.08, 70.70, 70.92, 71.60, 73.97, 76.18, 81.06, 81.65, 122.53, 130.82, 158.97; HRMS *m/z* (M + H)<sup>+</sup> found: 350.1956, C<sub>19</sub>H<sub>28</sub>NO<sub>5</sub><sup>+</sup> requires 350.1962.

**(4S,14S)-19-Azido-4,14-dimethyl-3,6,9,12,15-pentaoxa-21-azabicyclo[15.3.1]heneicos-1(21),17,19-triene** [(*S,S*)-8]

Starting from chloropyridino-crown ether (*S,S*)-9. Chloro compound (*S,S*)-9<sup>22</sup> (97.5 mg, 0.271 mmol), NaN<sub>3</sub> (1.0 g, 15 mmol), ethanol (1 mL) and water (1 mL) were mixed in a round-bottom flask, and stirred at 80 °C for a week. The reaction mixture was washed into a separatory funnel with ethyl acetate (50 mL) and water (50 mL). The phases were shaken well and separated. The aqueous phase was extracted with ethyl acetate (2 × 50 mL). The combined organic phase was dried over anhydrous MgSO<sub>4</sub>, filtered, and the solvent was removed. The crude product was purified by column chromatography on alumina using first ethanol–toluene 1:60 and then dioxane–hexane 1:6 mixtures as eluents to give (*S,S*)-8 (64 mg, 65%) as a yellow oil.

*R*<sub>f</sub>: 0.38 (alumina TLC, EtOH–toluene 1:40);  $[\alpha]_D^{25} = +30.7$  (*c* = 1.00, CHCl<sub>3</sub>); IR (neat)  $\tilde{\nu}_{\text{max}}$  (cm<sup>−1</sup>) 2968, 2930, 2864, 2107,



1592, 1572, 1448, 1371, 1339, 1319, 1225, 1104, 921, 854, 763, 719, 670, 572, 538;  $^1\text{H}$  NMR (300 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm) 1.16 (d,  $J$  = 6 Hz, 6H), 3.38–3.68 (m, 12H), 3.72–3.89 (m, 2H), 4.79 (s, 4H), 6.90 (s, 2H);  $^{13}\text{C}$  NMR (75.5 MHz,  $\text{CDCl}_3$ )  $\delta$  (ppm) 17.24, 70.84, 71.02, 71.68, 74.13, 76.29, 110.58, 149.71, 160.82; HRMS  $m/z$  (M + H) $^+$  found: 367.1980,  $\text{C}_{17}\text{H}_{27}\text{N}_4\text{O}_5^+$  requires: 367.1976.

Starting from iodopyridino-crown ether (*S,S*)-5. Iodo compound (*S,S*)-5<sup>22</sup> (69 mg, 0.15 mmol) and  $\text{NaN}_3$  (1.0 g, 15 mmol) were reacted in a mixture of ethanol (0.5 mL) and water (0.5 mL), in the same way as described above for chloro compound (*S,S*)-9, to give azide (*S,S*)-8 (32 mg, 58%).

All the properties of (*S,S*)-8 prepared this way were identical to those of the one prepared from (*S,S*)-9.

## Author contributions

Conceptualization: B. S., I. M. and P. H.; methodology: B. S. and M. M.; formal analysis: P. B.; investigation: B. S., M. M., D. P. and L. D.; writing – original draft: B. S.; writing – review and editing: P. B., I. M. and P. H.; visualization: B. S. and I. M.; supervision: I. M. and P. H.; project administration: I. M. and P. H.; funding acquisition: I. M. and P. H.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

Financial support from the National Research, Development and Innovation Office (NKFIH No. K128473, K112289 and PD104618) is gratefully acknowledged. B. Szemenyei is grateful for the fellowship provided by the Gedeon Richter's Talentum Foundation (1103 Budapest, Gyömrői út 19–21). We acknowledge the Governmental Agency for IT Development (KIFÜ) for awarding us access to resource based in Hungary. The authors thank Dr. György Tibor Balogh for recording some of the high-resolution mass spectra and Áron Szigetvári for elucidation of the 2D NMR spectra.

## References

- X. X. Zhang, J. S. Bradshaw and R. M. Izatt, *Chem. Rev.*, 1997, **97**, 3313–3361.
- V. N. Khose, M. E. John, A. D. Pandey, V. Borovkov and A. V. Karnik, *Symmetry*, 2018, **10**, 34.
- L. A. Nguyen, H. He and C. Pham-Huy, *Int. J. Biomed. Sci.*, 2006, **2**, 85–100.
- M. C. Núñez, M. E. García-Rubiño, A. Conejo-García, O. Cruz-López, M. Kimatrai, M. A. Gallo, A. Espinosa and J. M. Campos, *Curr. Med. Chem.*, 2009, **16**, 2064–2074.
- L. Pu, *Acc. Chem. Res.*, 2017, **50**, 1032–1040.
- J. R. Lakowicz, *Principles of fluorescence spectroscopy*, Springer Science + Business Media, New York, NY, 3rd edn, 2006.
- L. Pu, *Chem. Rev.*, 2004, **104**, 1687–1716.
- A. Accetta, R. Corradini and R. Marchelli, *Top. Curr. Chem.*, 2011, **300**, 175–216.
- L. Pu, *Acc. Chem. Res.*, 2012, **45**, 150–163.
- X. Zhang, J. Yin and J. Yoon, *Chem. Rev.*, 2014, **114**, 4918–4959.
- S. Yu and L. Pu, *Tetrahedron*, 2015, **71**, 745–772.
- O. K. Grigorova, D. I. Gusev, A. D. Averin, O. A. Maloshitskaya and I. P. Beletskaya, *Russ. Chem. Bull. Int. Ed.*, 2019, **68**, 848–854.
- I. Móczár and P. Huszthy, *Chirality*, 2019, **31**, 97–109.
- D. Pál, I. Móczár, B. Szemenyei, D. Marczona, I. Kocsis, G. Prikler, P. Vezse, P. Baranyai and P. Huszthy, *Tetrahedron*, 2019, **75**, 2900–2909.
- D. Pál, M. Gede, I. Móczár, P. Baranyai, P. Bagi and P. Huszthy, *Period. Polytech., Chem. Eng.*, 2020, **64**, 20–36.
- Á. Golcs, B. Á. Ádám, V. Horváth, T. Tóth and P. Huszthy, *Molecules*, 2020, **25**, 2571.
- H. Szabó-Szentjóbi, A. Márton, D. Pál, G. Dargó, Á. Szigetvári, C. Szántay, G. T. Balogh, T. Tóth and P. Huszthy, *Period. Polytech., Chem. Eng.*, 2020, **64**, 37–45.
- B. A. Jones, J. S. Bradshaw and R. M. Izatt, *J. Heterocycl. Chem.*, 1982, **19**, 551–556.
- R. B. Davidson, J. S. Bradshaw, B. A. Jones, N. K. Dalley, J. J. Christensen, R. M. Izatt, F. G. Morin and D. M. Grant, *J. Org. Chem.*, 1984, **49**, 353–357.
- J. S. Bradshaw, P. K. Thompson, R. M. Izatt, F. G. Morin and D. M. Grant, *J. Heterocycl. Chem.*, 1984, **21**, 897–901.
- R. M. Izatt, T. Wang, J. K. Hathaway, X. X. Zhang, J. C. Curtis, J. S. Bradshaw, C. Y. Zhu and P. Huszthy, *J. Inclusion Phenom. Mol. Recognit. Chem.*, 1994, **17**, 157–175.
- J. Kupai, S. Lévai, K. Antal, G. T. Balogh, T. Tóth and P. Huszthy, *Tetrahedron: Asymmetry*, 2012, **23**, 415–427.
- T. Németh, S. Lévai, A. Kormos, J. Kupai, T. Tóth, G. T. Balogh and P. Huszthy, *Chirality*, 2014, **26**, 651–654.
- B. Szemenyei, I. Móczár, D. Pál, I. Kocsis, P. Baranyai and P. Huszthy, *Chirality*, 2016, **28**, 562–568.
- J. Shi, L. Liu, J. He, X. Meng and Q. Guo, *Chem. Lett.*, 2007, **36**, 1142–1143.
- D. Schweinfurth, K. I. Hardcastle and U. H. F. Bunz, *Chem. Commun.*, 2008, 2203–2205.
- C. Le Droumaguet, C. Wang and Q. Wang, *Chem. Soc. Rev.*, 2010, **39**, 1233–1239.
- S. S. Bag and R. Kundu, *J. Org. Chem.*, 2011, **76**, 3348–3356.
- Y. H. Lau, P. J. Rutledge, M. Watkinson and M. H. Todd, *Chem. Soc. Rev.*, 2011, **40**, 2848–2866.
- P. Kautny, D. Bader, B. Stöger, G. A. Reider, J. Fröhlich and D. Lumpi, *Chem. – Eur. J.*, 2016, **22**, 18887–18898.
- S. Ast, P. J. Rutledge and M. H. Todd, *Eur. J. Inorg. Chem.*, 2012, 5611–5615.
- J. K.-H. Wong, M. H. Todd and P. J. Rutledge, *Molecules*, 2017, **22**, 200.
- R. M. Duke, E. B. Veale, F. M. Pfeffer, P. E. Kruger and T. Gunnlaugsson, *Chem. Soc. Rev.*, 2010, **39**, 3936–3953.
- W. Yan, Q. Wang, Q. Lin, M. Li, J. L. Petersen and X. Shi, *Chem. – Eur. J.*, 2011, **17**, 5011–5018.
- Y. Liu, W. Yan, Y. Chen, J. L. Petersen and X. Shi, *Org. Lett.*, 2008, **10**, 5389–5392.

36 Q. Lai, Q. Liu, Y. He, K. Zhao, C. Wei, L. Wojtas, X. Shi and Z. Song, *Org. Biomol. Chem.*, 2018, **16**, 7801–7805.

37 Q. Lai, Q. Liu, K. Zhao, C. Shan, L. Wojtas, Q. Zheng, X. Shi and Z. Song, *Chem. Commun.*, 2019, **55**, 4603–4606.

38 Y. Zhang, X. Ye, J. L. Petersen, M. Li and X. Shi, *J. Org. Chem.*, 2015, **80**, 3664–3669.

39 A. Petrosyan, P. Ehlers, A.-E. Surkus, T. V. Ghochikyan, A. S. Saghyan, S. Lochbrunner and P. Langer, *Org. Biomol. Chem.*, 2016, **14**, 1442–1449.

40 Z. Jia and Q. Zhu, *Bioorg. Med. Chem. Lett.*, 2010, **20**, 6222–6225.

41 O. Berger, A. Kaniti, C. T. van Ba, H. Vial, S. A. Ward, G. A. Biagini, P. G. Bray and P. M. O'Neill, *ChemMedChem*, 2011, **6**, 2094–2108.

42 S. P. Chavan, S. Garai and K. P. Pawar, *Tetrahedron Lett.*, 2013, **54**, 2137–2139.

43 X.-J. Quan, Z.-H. Ren, Y.-Y. Wang and Z.-H. Guan, *Org. Lett.*, 2014, **16**, 5728–5731.

44 H. Wang, P. Mi, W. Zao, R. Kumar and X. Bi, *Org. Lett.*, 2017, **19**, 5613–5616.

45 F. Himo, T. Lovell, R. Hilgraf, V. V. Rostovtsev, L. Noodleman, K. B. Sharpless and V. V. Fokin, *J. Am. Chem. Soc.*, 2005, **127**, 210–216.

46 A. Dondoni and A. Marra, *J. Org. Chem.*, 2006, **71**, 7546–7557.

47 V. K. Tiwari, B. B. Mishra, K. B. Mishra, N. Mishra, A. S. Singh and X. Chen, *Chem. Rev.*, 2016, **116**, 3086–3240.

48 J. A. Riddick, W. B. Bunker and T. K. Sakano, in *Techniques of Chemistry*, ed. A. Weissberger, Wiley-Interscience, New York, NY, 4th edn, 1986, vol. 2.

49 Z. Köntös, P. Huszthy, J. S. Bradshaw and R. M. Izatt, *Tetrahedron: Asymmetry*, 1999, **10**, 2087–2099.

