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Nano-evolution and protein-based enzymatic evolution predicts novel types of natural product nanozymes of traditional Chinese medicine: cases of herbzymes of Taishan-Huangjing (*Rhizoma polygonati*) and Goji (*Lycium chinense*)†

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Nanozymes and natural product-derived herbzymes have been identified in different types of enzymes simulating the natural protein-based enzyme function. How to explore and predict enzyme types of novel nanozymes when synthesized remains elusive. An informed analysis might be useful for the prediction. Here, we applied a protein-evolution analysis method to predict novel types of enzymes with experimental validation. First, reported nanozymes were analyzed by chemical classification and nano-evolution. We found that nanozymes are predominantly classified as protein-based EC1 oxidoreductase. In comparison, we analyzed the evolution of protein-based natural enzymes by a phylogenetic tree and the most conserved enzymes were found to be peroxidase and lyase. Therefore, the natural products of *Rhizoma polygonati* and Goji herbs were analyzed to explore and test the potent new types of natural nanozymes/herbzymes using the simplicity simulation of natural protein enzyme evolution as they contain these conserved enzyme types. The experimental validation showed that the natural products from the total extract of nanoscale traditional Chinese medicine Huangjing (RP, *Rhizoma polygonati*) from Mount-Tai (Taishan) exhibit fructose-bisphosphate aldolase of lyase while nanoscale Goji (*Lycium chinense*) extract exhibits peroxidase activities. Thus, the bioinformatics analysis would provide an additional tool for the virtual discovery of natural product nanozymes.

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Introduction

Nanozymes are functional nanomaterials with enzyme-like activity and biocatalytic function that mimic natural enzymes (“learning from nature”) while retaining

nanomaterial properties.^{1,2} Since the discovery of iron nanoparticles with intrinsic enzyme activity, more than 540 novel nanozymes have been reported with modifications since the first generation of classical Fe₃O₄ nanozyme with peroxidase-like activity.³ Recently, Meng *et al.* reported an improved peroxidase activity of a pyrite peroxidase nanozyme with super strong binding ability towards substrate hydrogen peroxide (H₂O₂) compared to the traditional nanozyme Fe₃O₄ and protein-based horseradish peroxidase, and additional glutathione oxidase-like activity, following the catalysis of H₂O₂ generation.⁴ The dual nanozyme finally targeted KRAS mutation with tumor catalytic therapy.⁴

Currently, there are numerous types of nanozymes reported but most are oxidoreductase, hydrolases and hydrolases.¹ In particular, peroxidase POD is widely studied and reported as a type of oxidoreductase.² There are basically 6 or 7 large types of protein-based enzymes, namely natural enzymes, which are classified according to the Enzyme Commission (EC) classification as EC1, 2, 3, 4, 5, 6, 7 representing oxidoreductases, transferases, hydrolases, lyases, isomerases, ligases, and

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translocases, respectively.⁵ Frances H. Arnold proposed the concept of the directed evolution of enzymes by introducing random mutation of genes encoding target protein enzymes and specific chemical reaction-based screening of target mutants, with repeated cycles to obtain the final expected enzymes.⁶ The directed evolution of enzymes is artificially created to simulate natural evolution mechanisms with natural selection.⁷ For example, cytochrome P450 can be evolved by directed evolution with synthetic carbene and nitrene to improve the catalysis activity *via* genetic coding changes or mutation.⁷ Thus, engineered enzyme evolution can be realized by modifications.

Nanozymes can combine the advantages of natural enzymes and traditional artificial mimicking enzymes with high-efficiency due to adjustable catalytic activity and nano-structures.² In addition, nanozymes can function under extreme pH and high temperature conditions.¹ The catalytic function of a nanozyme is related to its own nano effects such as size, surface area, and structure.² These effects also provide an operational way for the optimization of simulated enzymes by manually controlling the nano effects to regulate the catalytic activity and substrate selectivity, which are simulating the natural enzyme evolution and selection. For example, Gao *et al.* synthesized Fe₃O₄ nanoparticles of different sizes and the relative catalytic activity was enhanced 5 times using 30 nm rather than 300 nm.⁸

A type of natural product-based nanozyme and nano-assembly has been arising as a new star in nano-natural product research. These include the natural compound-based self-assembly and nanozyme, namely herbzymes.^{9,10} Recently, we reported natural product-based nanozymes of protease-like, phosphatase-like, and α -amylases (EC 3.2.1.1), which are mainly hydrolases.⁹ However, whether natural products nanoparticles exert other types of enzyme activity and the evolution of nanozymes remains elusive.

Although numerous nanozymes were discovered and applied in biomedicine, their evolution is still unclear. Here, this study aims to analyze the nano-evolution of nanozymes and natural enzymatic phylogenetic trees to predict and discover novel types of nanozymes. We compared and analyzed the association between nanozyme types using six types of natural enzymes, and then identified evolutionary links to predict new potential nanozyme types with experimental validations and found a novel type of natural product nanozyme.

Methods

Evolutionary analysis of the protein-based natural enzyme

Sequences collected for this study were aligned using MEGAX. Maximum likelihood (ML) trees were generated using the RAxML-HPC2 on XSEDE (8.2.10) in the CIPRES Science Gateway platform by the GTR model of evolution, and fast bootstrap analyses of 1000 pseudoreplicates were completed. The

combined maximum likelihood analyses were run using amino acid sequences that were downloaded from Genbank.

Chemical nanozyme evolution analysis

First, the collected data¹¹ showing that the most current reported nanozymes were analyzed from the list of recent review and searched papers in Pubmed, and then, the evolution of the same type of element-based families were divided according to the enzyme types and the elements of nanozymes. In detail, before making the self-made chemical evolutionary tree, all the data were classified into six categories according to the types of catalytic reactions of traditional classifications of EC1,2,3,4,5,6 representing oxidoreductase, transferase, hydrolase, lyase, isomerase, and ligase, respectively and an alternative type EC7 for translocases. Then, the chemical element forms of each enzyme were found successively and the branches were classified. Homology was calculated according to the element composition percentage, and the semi-quantitative evolution maps were drawn according to the homology.

In the process of making a chemical evolutionary tree, the data of elements used were sorted into the same group, and the element frequency was counted and plotted according to the number of occurrences. The frequency of seven types of enzymes was plotted and analyzed according to the number of elements involved.

Preparation of herbal nanoparticles, fluorescence measurement and scanning electron microscopy (SEM)

Herbal nanoparticles were prepared from Mount Tai area-derived traditional Chinese medicine *Rhizoma polygonat* (RP) also named as Huangjing, and confirmed as described previously.⁹ For the Goji (*Lycium chinense*) nanoparticles prepared by a baking process,⁹ extract fluorescence was measured by a fluorometer described previously.⁹ The SEM analysis of baked Goji nanoparticles was performed by dropping Goji extract on a carrier and air-drying it before putting it into a scanning electron microscope.⁹

Prediction of novel enzyme types of herbzyme/nanozyme

Based on the above evolutionary analysis of protein-based maximum likelihood (ML) trees and nanozyme-based natural enzyme type analysis, the most conserved types of enzyme based on EC1,2,3,4,5,6 representing oxidoreductase, transferase, hydrolase, lyase, isomerase, and ligase, respectively, were searched and analyzed.¹¹ The most predominant chemical-based nanozymes were classified as EC1 oxidoreductase, while that of the protein-based enzymes was found to be fructose-bisphosphate aldolase. Thus, the hypothesis is that the nanozyme might have evolved from similarity to simple conserved protein enzymes to a complicated enzyme type. The predicted enzyme types of oxidoreductase and fructose-bisphosphate aldolase were further used for



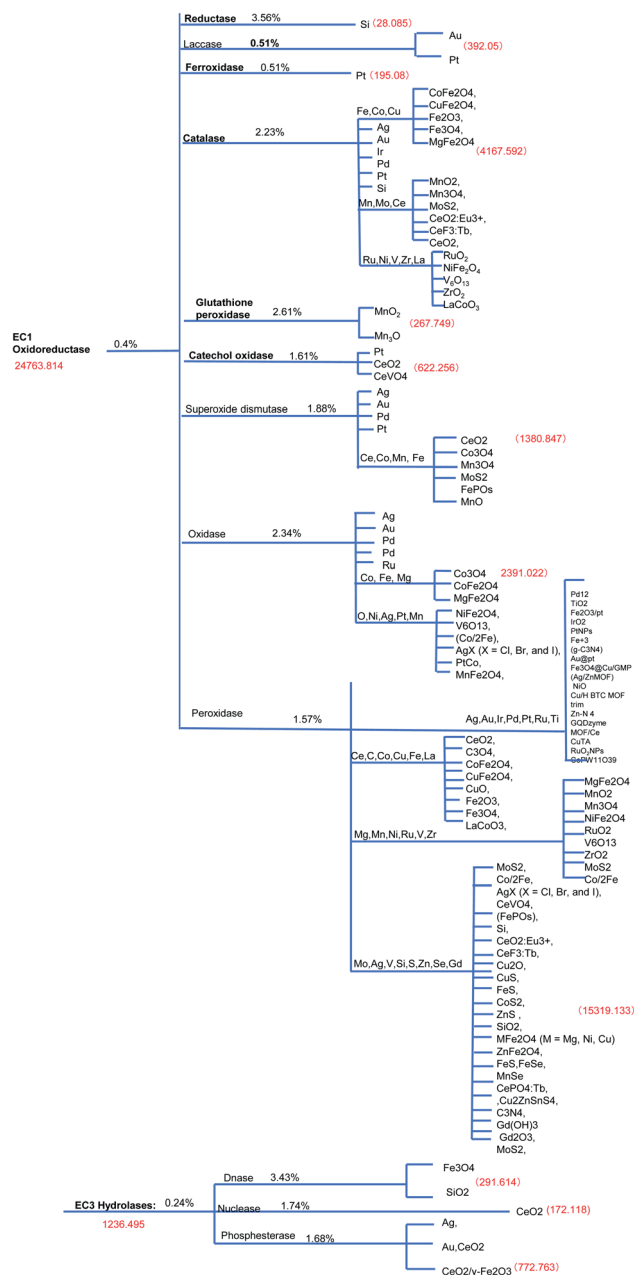


Fig. 2 Details of nanozyme distance based on element and enzyme types.

(1.61%), superoxide dismutase (1.88%) and catechol oxidase (1.61%) and EC2 hydrolase (0.24%) consists of nuclease (1.74%), DNase (3.43%), and phosphatase (1.68%) (Figure 1B and Figure 2).¹¹ The evolutionary tree of nanozyme elements also branched according to the distance of the element family, which collected most data of the latest literature from 2018 review to 2020 new publications reviewed (ESI references[†]).¹¹ Our data suggest that peroxidase, catalase, and oxidase were the most conserved types among tested, while non-conservative types are reductases and ferroxidase.

Phylogenetic tree analysis of natural enzyme evolution to predict novel types of nanozyme

To better explore and understand the different types of nanozymes, we further tested and compared each type of natural enzyme evolutionary conservancy.

Peroxidase. Natural peroxidase (EC 1.1.1.1) is a protein enzyme widely distributed in organisms, which acts as an electron acceptor to catalyze the redox reaction between H_2O_2 and a variety of electron donors.¹² Mainly iron porphyrin as the auxiliary group has the dual role of catalyzing the decomposition of numerous phenolic and amine compounds to eliminate the toxic free radicals in cells.¹² According to the protein sequence and structure, peroxidase can be divided into non-heme peroxidase and heme peroxidase.¹³ A phylogenetic tree of peroxidase was established with the maximum likelihood tree. Peroxidase exists in the bodies of various animals, plants and microorganisms. When establishing the evolutionary tree, different species were selected and the three types of peroxidase were used to establish the evolutionary tree: myeloperoxidase, lactoperoxidase and thyroid peroxidase with peroxidase as the outgroup. From the topological structure (Fig. 3), the phylogenetic tree is divided into two large branches both having a higher support rate of 100%. In one of the branches, the relationship between myeloperoxidase and lactoperoxidase is closer than that of thyroid peroxidase. All the species in peroxidase have a higher support rate of genetic relationship among each species (all 100%).

Transferases of hexokinase and glutathione S-transferase (GST). A class of transferases are GSTs (EC 2.5.1.18), which are phase II metabolizing enzymes and play an important role in cell detoxification and stress response.¹⁴ The number of genes in each subclass is different in the phylogenetic tree. The phylogenetic tree of GST displays only species from the animal kingdom making it uniquely specific to animals. Its major 2 branches have a support rate of 89% and 77%, respectively, indicating the high rate of mutations and evolutionary changes within these groups (Fig. 4).

Hexokinase (HK) (EC 2.7.1.1) is also a class of transferase, which can catalyze the phosphorylation of hexose in sugar catabolism and has a dual role of catalysis and regulation. In mammals, of which four subtypes of HK-I, -II, -III and -IV, the first 3 have higher affinity towards glucose than HK-IV.¹⁵ To find the evolutionary relationship among different species of hexokinase, we used the maximum likelihood method to create the relationship with galactokinase as the outgroup. The topology of the maximum likelihood tree is that all hexokinase species form large branches have a support rate of 100%. The first hexokinase branch is the animal branch, which includes humans, mice, and fish, with a bootstrap value of 87%, followed by the plant branch, which has a bootstrap value of 88% (Fig. 4). There is another bacterium branch with a self-expanding value of 72%. Animal branch and plant branch constitute a large branch with a low support rate of bootstrap support (BS) at 24%.

Hydrolase of ALP. One example of a class of hydrolase is alkaline phosphatase (ALP) (EC 3.1.3.1), which functions by



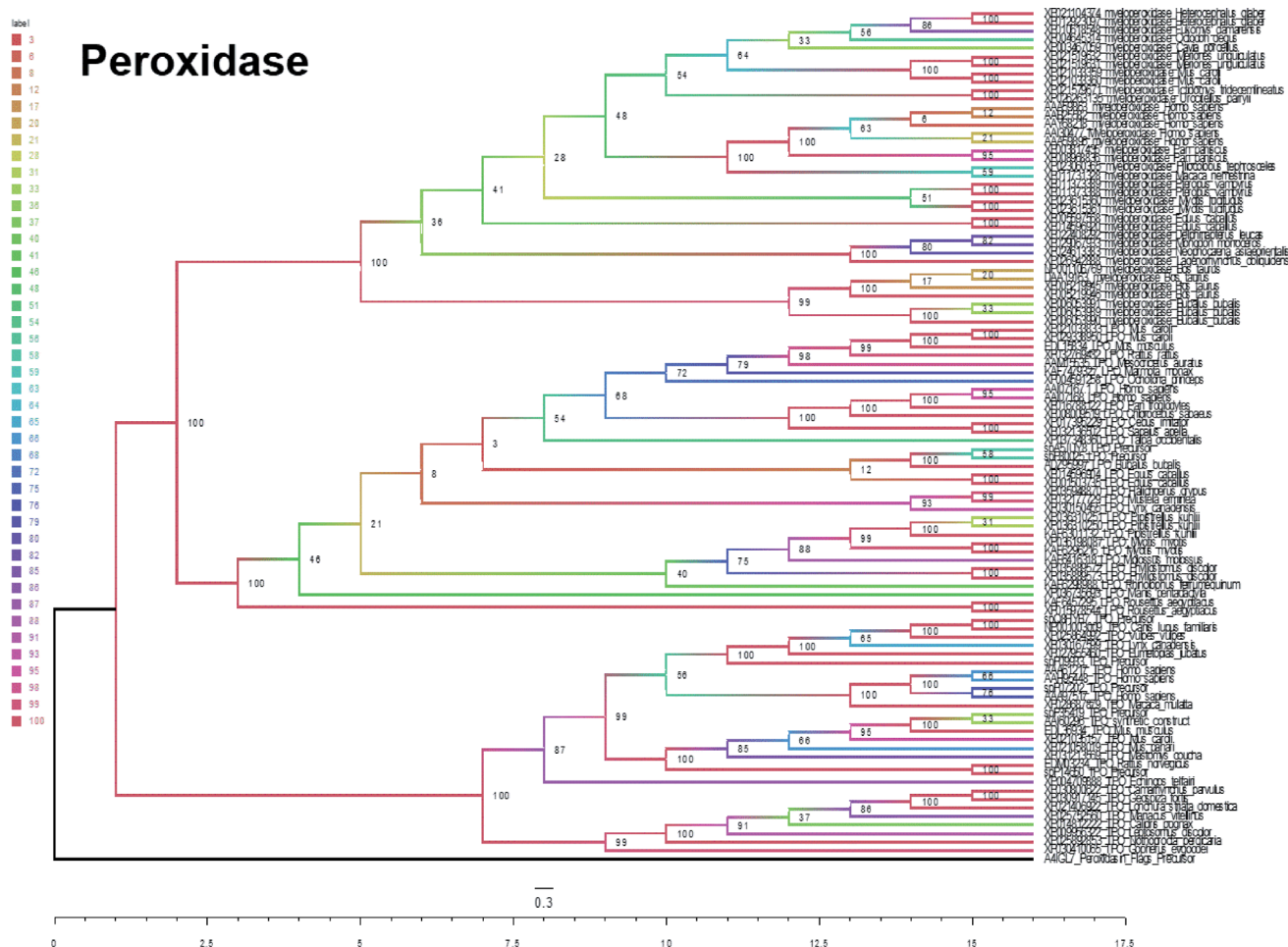


Fig. 3 Maximum likelihood tree of peroxidase.

reversing phosphorylation; some types of nanoparticles have been reported to have intrinsic phosphatase activity.^{11,16} Recently, we reported that natural product-derived carbon nanodots and nanoflowers, can dephosphorylate substrates as ALP-like nanozymes in addition to the chemical synthesized nanozymes.^{9,11,17}

The phylogenetic tree of ALP was established with the maximum likelihood tree, in which ALPI (alkaline phosphatase intestinal) was selected as the outer group. Based on the topological structure of the maximum likelihood tree, the evolutionary tree is divided into two branches: bacteria and animal (Fig. 5). The support rate of the bacterium branch is 100%, and the support rate of the animal branch is 78%. The bacterium and animal branches form large branches with an approval rate of 88% (Fig. 5).

Fructose-bisphosphate aldolase. Fructose-bisphosphate aldolase (EC4.1.2.13) is an example of lyases. For instance, fructose-1,6-bisphosphate aldolase plays one of the key roles in cellular metabolism, *i.e.*, it catalyzes the reversible aldol condensation of glyceraldehyde 3-phosphate and dihydroxyacetone phosphate to form fructose 1,6-bisphosphate.¹⁸ Fructose-

bisphosphate aldolase has been divided into archaeal or bacteria and eukaryotic classes.¹⁹

The evolutionary relations of fructose-bisphosphate aldolase were established using the maximum likelihood tree, and sedoheptulose-bisphosphatase was selected as the outer group. The maximum likelihood tree is divided into two major branches as animal (BS: 100%) and bacteria branches (BS: 100%) (Fig. 6).

Isomerase. Isomerase transfers a chemical subgroup to create a new isomer in the reaction. Isomerase evolution are much complicated due to different substrates and their specific function in different organisms, and there is no convergent evolution between different types of isomerases, such as α -arabinose isomerase (EC 5.3.1.4) and xylose isomerase.²⁰ The isomerase evolution is much complex with unique characteristics of gaining new functions with different EC classes, particularly by interchange with lyases (EC 4) with an advantage in evolution.²¹ Thus, the isomerase evolution shows much diversity, and the potential of nanozyme simulating would not be superior.

DNA ligase. DNA ligase (EC6.5.1.1) is an example of a type of ligase, which is widely applied in biotechnology, in *in vivo* DNA



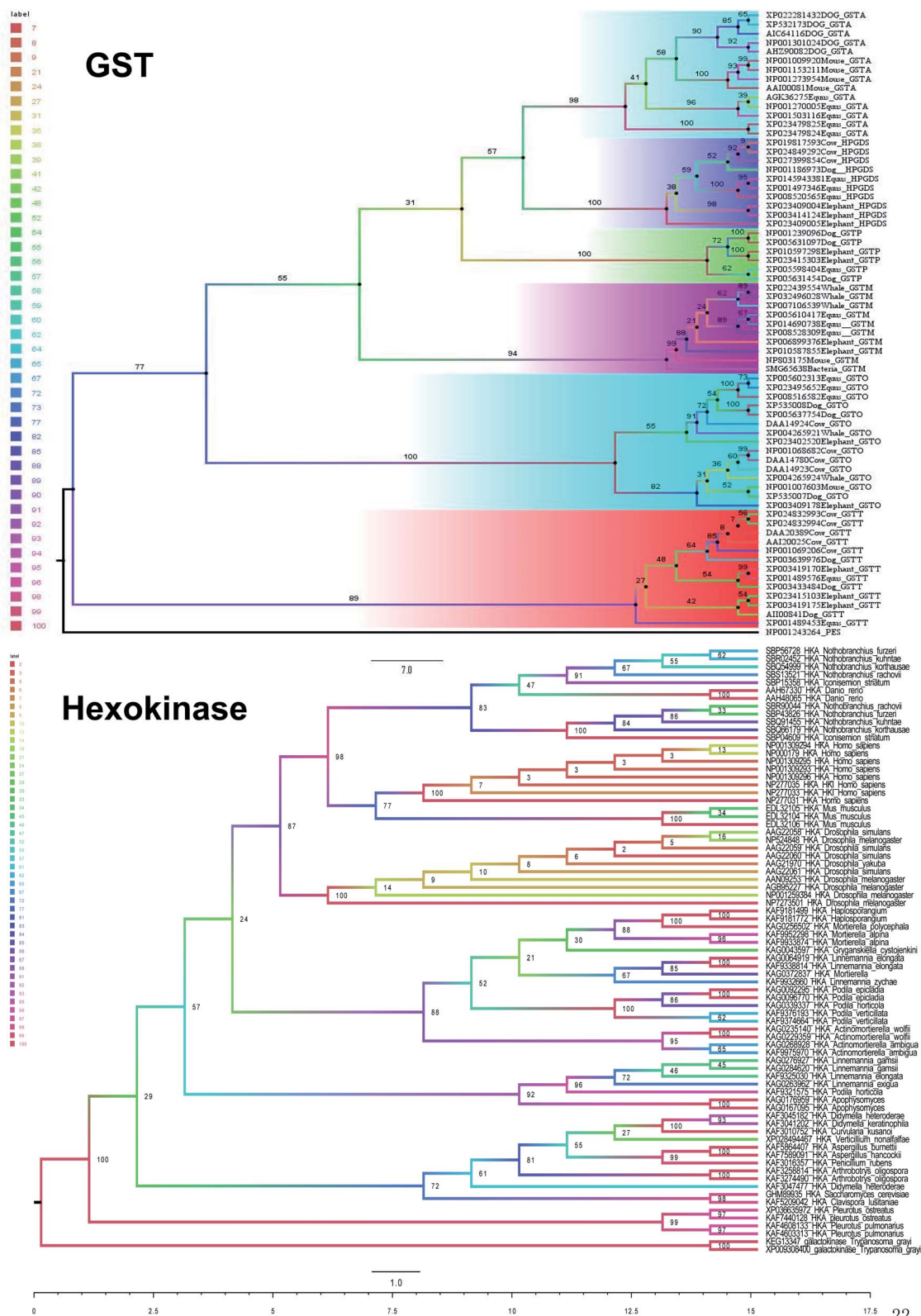


Fig. 4 Maximum likelihood tree of hexokinase and GST.



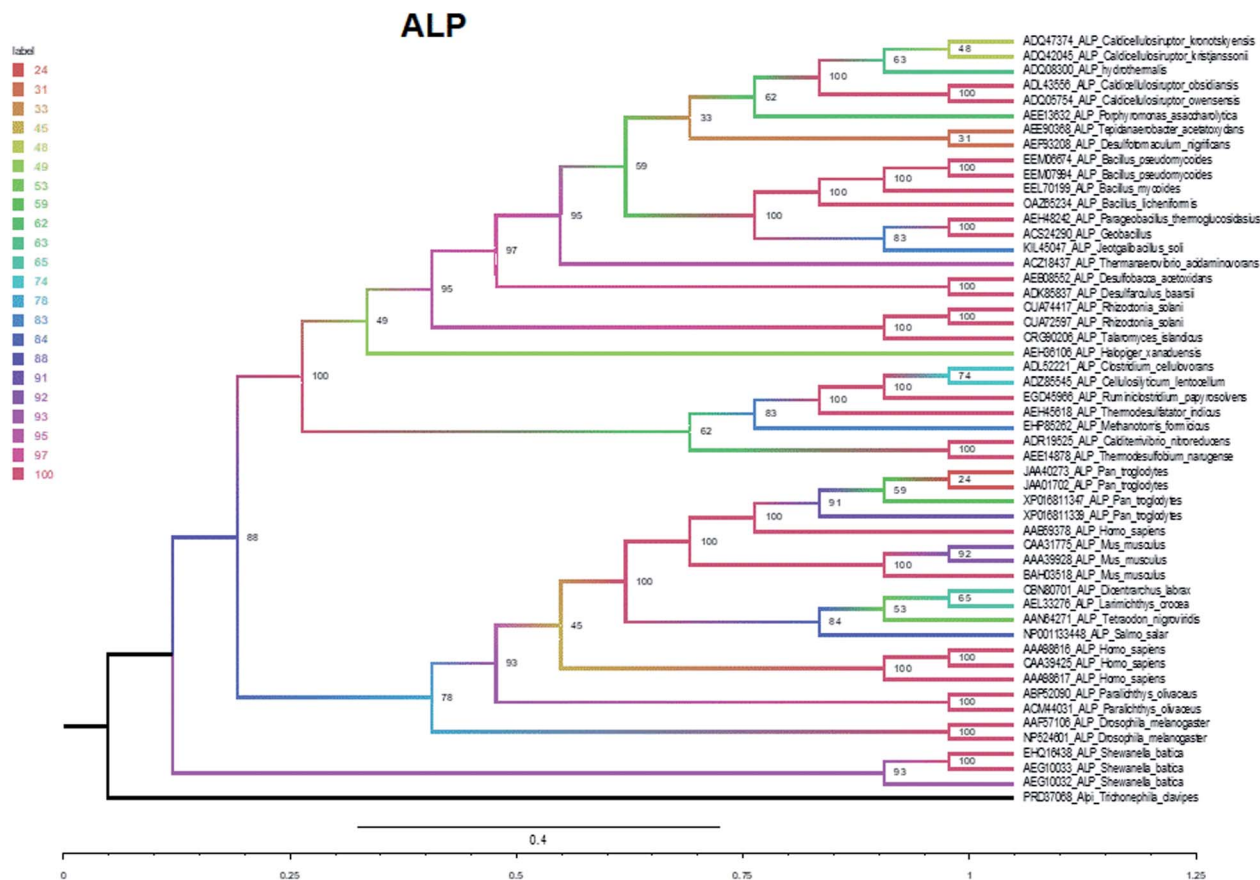


Fig. 5 Maximum likelihood tree of ALP.

repair, catalyzes the formation of the phosphodiester bond between 3'-OH and 5'-phosphate ends of adjacent DNA. DNA ligase has been found in viruses, bacteria, and eukaryotes.²² The mechanisms of catalysis of different types of DNA ligases differ. In general, DNA ligases can be divided into adenosine triphosphate (ATP)-dependent and nicotinamide adenine dinucleotide (NAD)-dependent types according to the energy sources required for the catalytic reaction of DNA ligases, which are widely distributed in eukaryotes, archaea, eubacteria and viruses.²³ The evolutionary analysis of DNA ligases suggests that ATP-dependent ligases acquired from prokaryotic and eukaryotic species evolved compared to the previously used NAD-dependent ligases obtained from bacteriophages in eukaryotic nucleocytoplasmic large DNA viruses during their complex evolutionary stages.²⁴

In summary, five evolutionary trees were established using the maximum likelihood tree. By observing the topological structure of the six phylogenetic trees here, it was found that the fructose-bisphosphate aldolase and peroxidase enzymes are the most conserved among the various species, which suggests that their potent bio-inspired nanoparticles would be prevalent to easily evolve and exert the functions of the two enzyme types.

Experimental validation of novel types of natural product nanoparticle-based-nanozymes: herbzymes

Given fructose-bisphosphate aldolase and peroxidase are the most conserved nanozyme types among those tested, we further explored the possibility of the prediction of natural product-nanoparticle based-nanozyme of herbzymes in these two new types as there is a report on the ALP activity of herbzyme. We first found that *Rhizoma polygonati* (RP) has the fructose-bisphosphate aldolase activity compared to the control (Fig. 7A). Moreover, the Goji extract of the nanoscale natural product showed quantum fluorescence at 420 nm and peroxidase activity when using TMB as the substrate with the highest activity at pH 10 and concentration-dependent kinetics when less than V_{max} (Fig. 7B-E). Thus, in addition to the herbzyme with phosphatase activity of RP, as we reported before,⁹ fructose-bisphosphate aldolase of lyases by RP and peroxidase of Goji were discovered from prediction and validation.

Discussion

Here, we report a bioinformatics-based analysis of protein enzyme evolution and comparisons among 6 types of classified enzymes with the nano-evolution of material-based



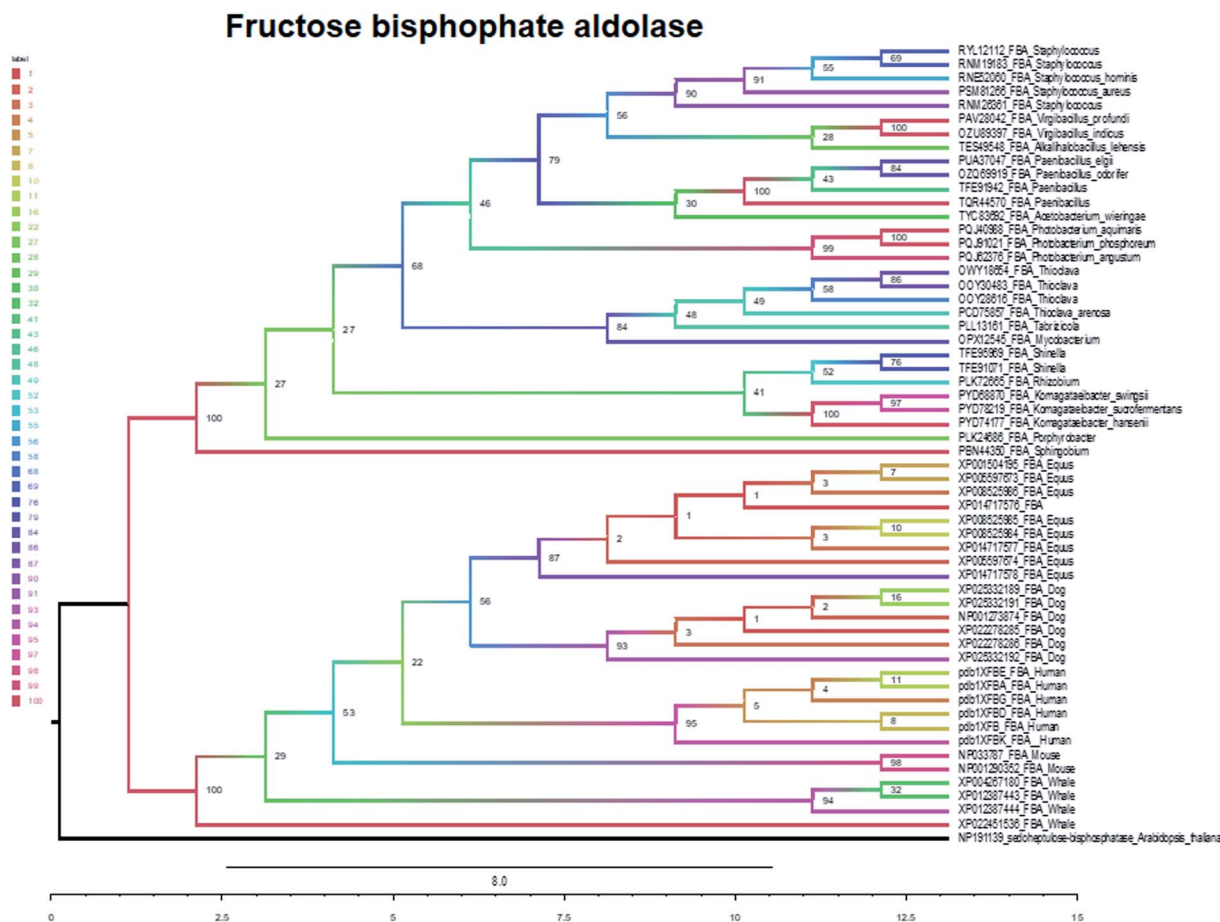


Fig. 6 Maximum likelihood tree of fructose biphosphate aldolase.

nanozymes, and from the results, we further predicted the bio-inspired nanozyme of herbzyme's novel types corresponding to the protein enzyme types. Our results suggest that self-assembly of natural product of herbal nanoparticles may contribute to the fructose-bisphosphate aldolase and peroxidase activities of the natural products which is consistent with our previous report.⁹

From protein-based enzyme evolution, we found that fructose-bisphosphate aldolase is much conserved among different species. According to Say *et al.*,²⁵ fructose-bisphosphate aldolase is a conserved bifunctional enzyme with aldolase catalytic activity in archaea and bacteria. Similarly, Shams *et al.*²⁶ indicate that fructose-bisphosphate aldolase housekeeping has a metabolic role as it is a moonlighting protein meaning that it can perform several key functions. It is due to its involvement in the Calvin cycle and glycolysis in addition to gluconeogenesis, which are believed to be acquired additional functions as ancestral organisms evolved. Furthermore, the deletion experiment of genes responsible for fructose-bisphosphate aldolase in *C. albicans*²⁷ suggests that fructose-bisphosphate aldolase also plays a critical role in the organism's growth. These studies clearly demonstrate the significance of fructose-bisphosphate aldolase for the survival of the organisms, hence indicating why it is highly conserved.

Peroxidase has been largely reported in metal material-based nanozymes. We here found the highly conserved possibility in protein-based enzymes, which correlated with the material-based nanozymes as the highly discovered enzyme of the EC family type. We then tested the herb natural product and we found some natural herbzymes have peroxidase activity, but not all herbs (data not shown) as we found RP did not have this type of enzyme function at this condition *in vitro*. The RP contains many polysaccharides, and the herbal processing induced chemical reaction results in the loss or generation of different sugars as reported from other herbs too.^{28,29} Whether it might be repressed or promoted by surface chemical groups, the novel types of nanoscale herbzymes may be predicted by the protein-based enzyme evolutionary conservation analysis.

Conclusion

Nanomaterial-evolution and protein-based enzyme evolutionary analysis showed relatively most conserved types of protein enzymes. The natural product-derived herbzyme was predicted to be a novel type of enzyme by analysis through bio-inspired simulation and experimental validation for the



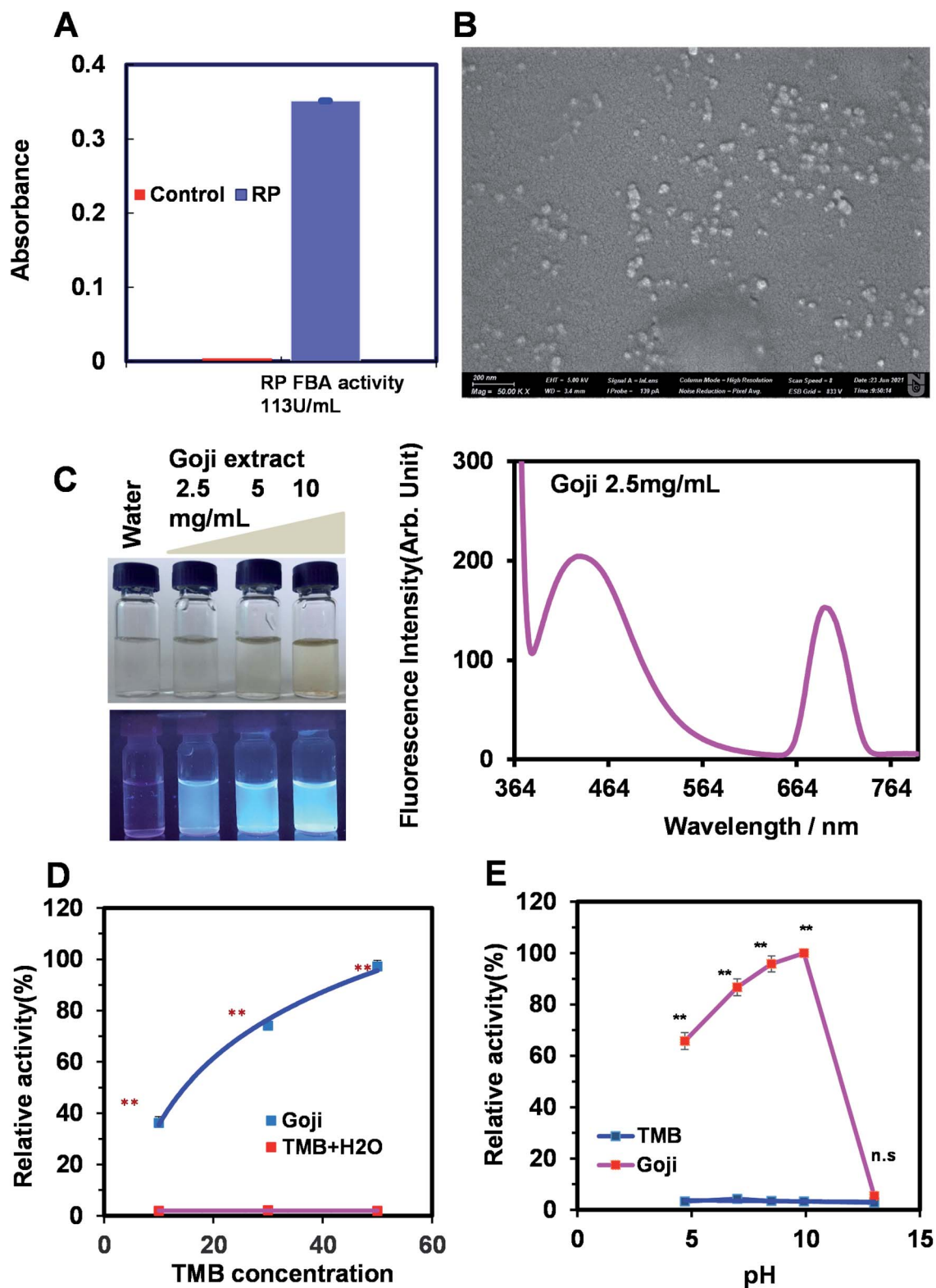


Fig. 7 Experimental validation of predicted novel type of herbzymes. (A) Fructose bisphosphate aldolase (FBA) of RP nanoparticles using the RP extract with a FBA assay kit. Nanoscale characteristics of the RP extract were shown by us previously.⁹ (B) Nanoparticle characteristics of the Goji extract by fluorescence. (C) Nanoparticle characteristics of the Goji extract by size investigated by SEM. (D) Nanoscale Goji extract exhibits the nanozyme activity of peroxidase using TMB as the substrate showing the enzyme kinetics by different concentrations of the substrate. (E) pH effect on the nanoscale Goji extract on peroxidase activity.



discovery of new types of nanozymes/herbzymes such as fructose-bisphosphate aldolase of lyases and peroxidase.

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Author contributions

Yingqiu Xie developed the novel concept, planned experiments, and wrote paper. Guldan Nazarbek, Lazzat Nurtay, performed enzyme and nanozyme evolutionary analysis and wrote paper. Aidana Kutzhanova, Chenglin Mu performed herbzyme experiments, analyzed data, and wrote some methods. Cuiping Ma and Amr Amin revised paper, and provided consulting. Xugang Li, and Bexultan Kazybay analyzed the results, discussed the data, and wrote part of the paper.

Conflicts of interest

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References

- M. Liang and X. Yan, Nanozymes: From New Concepts, Mechanisms, and Standards to Applications, *Acc. Chem. Res.*, 2019, **52**(8), 2190–2200, DOI: 10.1021/acs.accounts.9b00140.
- R. Zhang, K. Fan and X. Yan, Nanozymes: created by learning from nature, *Sci. China Life Sci.*, 2020, **63**(8), 1183–1200, DOI: 10.1007/s11427-019-1570-7.
- B. Jiang, L. Fang, K. Wu, X. Yan and K. Fan, Ferritins as natural and artificial nanozymes for theranostics, *Theranostics*, 2020, **10**(2), 687–706, DOI: 10.7150/thno.39827.
- X. Meng, D. Li, L. Chen, H. He, Q. Wang, C. Hong, J. He, X. Gao, Y. Yang, B. Jiang, G. Nie, X. Yan, L. Gao and K. Fan, High-Performance Self-Cascade Pyrite Nanozymes for Apoptosis-Ferroptosis Synergistic Tumor Therapy, *ACS Nano*, 2021, **15**(3), 5735–5751, DOI: 10.1021/acsnano.1c01248.
- P. K. Robinson, Enzymes: principles and biotechnological applications, *Essays Biochem.*, 2015, **59**, 1–41, DOI: 10.1042/bse0590001. Erratum in: *Essays Biochem.*, 2015, **59**, 75.
- F. H. Arnold, Innovation by Evolution: Bringing New Chemistry to Life (Nobel Lecture), *Angew. Chem., Int. Ed.*, 2019, **58**(41), 14420–14426, DOI: 10.1002/anie.201907729.
- Y. Yang and F. H. Arnold, Navigating the Unnatural Reaction Space: Directed Evolution of Heme Proteins for Selective Carbene and Nitrene Transfer, *Acc. Chem. Res.*, 2021, **54**(5), 1209–1225, DOI: 10.1021/acs.accounts.0c00591.
- L. Gao, J. Zhuang, L. Nie, J. Zhang, Y. Zhang, N. Gu, T. Wang, J. Feng, D. Yang, S. Perrett and X. Yan, Intrinsic peroxidase-like activity of ferromagnetic nanoparticles, *Nat. Nanotechnol.*, 2007, **2**(9), 577–583, DOI: 10.1038/nnano.2007.260.
- E. Benassi, H. Fan, Q. Sun, K. Dukenbayev, Q. Wang, A. Shaimoldina, A. Tassanbiyeva, L. Nurtay, A. Nurkesh, A. Kutzhanova, C. Mu, A. Dautov, M. Razbekova, A. Kabylda, Q. Yang, Z. Li, A. Amin, X. Li and Y. Xie, Generation of particle assemblies mimicking enzymatic activity by processing of herbal food: the case of rhizoma polygonati and other natural ingredients in traditional Chinese medicine, *Nanoscale Adv.*, 2021, **3**, 2222–2235, DOI: 10.1039/D0NA00958J.
- T. Li, P. Wang, W. Guo, X. Huang, X. Tian, G. Wu, B. Xu, F. Li, C. Yan, X. J. Liang and H. Lei, Natural Berberine-Based Chinese Herb Medicine Assembled Nanostructures with Modified Antibacterial Application, *ACS Nano*, 2019, **13**(6), 6770–6781, DOI: 10.1021/acsnano.9b01346.
- J. Wu, X. Wang, Q. Wang, Z. Lou, S. Li, Y. Zhu, L. Qin and H. Wei, Nanomaterials with enzyme-like characteristics (nanozymes): next-generation artificial enzymes (II), *Chem. Soc. Rev.*, 2019, **48**(4), 1004–1076, DOI: 10.1039/C8CS00457A.
- A. A. Khan, A. H. Rahmani, Y. H. Aldehbi and S. M. Aly, Biochemical and pathological studies on peroxidases—an updated review, *Glob. J. Health Sci.*, 2014, **6**(5), 87–98, DOI: 10.5539/gjhs.v6n5p87.
- Y. Sugano, DyP-type peroxidases comprise a novel heme peroxidase family, *Cell. Mol. Life Sci.*, 2009, **66**(8), 1387–1403, DOI: 10.1007/s00018-008-8651-8.
- R. R. Singh and K. M. Reindl, Glutathione S-Transferases in Cancer, *Antioxidants*, 2021, **10**(5), 701, DOI: 10.3390/antiox10050701.
- F. Ciscato, L. Ferrone, I. Masgras, C. Laquatra and A. Rasola, Hexokinase 2 in Cancer: A Prima Donna Playing Multiple Characters, *Int. J. Mol. Sci.*, 2021, **22**(9), 4716, DOI: 10.3390/ijms22094716.
- X. Xie, Y. Wang, X. Zhou, J. Chen, M. Wang and X. Su, Fe-N-C single-atom nanozymes with peroxidase-like activity for the detection of alkaline phosphatase, *Analyst*, 2021, **146**(3), 896–903, DOI: 10.1039/D0AN01846E.



- 17 Y. Xie, H. Fan, W. Lu, Q. Yang, A. Nurkesh, T. Yeleussizov, A. Maipas, J. Lu, L. Manarbek, Z. Chen and E. Benassi, Nuclear MET requires ARF and is inhibited by carbon nanodots through binding to phospho-tyrosine in prostate cancer, *Oncogene*, 2019, **38**(16), 2967–2983, DOI: 10.1038/s41388-018-0608-2.
- 18 E. Lorentzen, B. Siebers, R. Hensel and E. Pohl, Structure, function and evolution of the Archaeal class I fructose-1,6-bisphosphate aldolase, *Biochem. Soc. Trans.*, 2004, **32**(Pt 2), 259–263, DOI: 10.1042/bst0320259.
- 19 G. J. Thomson, G. J. Howlett, A. E. Ashcroft and A. Berry, The dhna gene of Escherichia coli encodes a Class I fructose bisphosphate aldolase, *Biochem. J.*, 1998, **331**(2), 437–445, DOI: 10.1042/bj3310437.
- 20 S. Banerjee, F. Anderson and G. K. Farber, The evolution of sugar isomerases, *Protein Eng.*, 1995, **8**(12), 1189–1195, DOI: 10.1093/protein/8.12.1189.
- 21 S. Martinez Cuesta, N. Furnham, S. A. Rahman, I. Sillitoe and J. M. Thornton, The evolution of enzyme function in the isomerases, *Curr. Opin. Struct. Biol.*, 2014, **26**(100), 121–130, DOI: 10.1016/j.sbi.2014.06.002.
- 22 S. Shuman, DNA ligases: progress and prospects, *J. Biol. Chem.*, 2009, **284**(26), 17365–17369, DOI: 10.1074/jbc.R900017200.
- 23 A. Sallmyr, I. Rashid, S. K. Bhandari, T. Naila and A. E. Tomkinson, Human DNA ligases in replication and repair, *DNA Repair*, 2020, **93**, 102908, DOI: 10.1016/j.dnarep.2020.102908.
- 24 N. Yutin and E. V. Koonin, Evolution of DNA ligases of nucleocytoplasmic large DNA viruses of eukaryotes: a case of hidden complexity, *Biol. Direct*, 2009, **4**, 51, DOI: 10.1186/1745-6150-4-51.
- 25 R. F. Say and G. Fuchs, Fructose 1,6-bisphosphate aldolase/phosphatase may be an ancestral gluconeogenic enzyme, *Nature*, 2010, **464**(7291), 1077–1081, DOI: 10.1038/nature08884.
- 26 F. Shams, N. J. Oldfield, K. G. Wooldridge and D. P. Turner, Fructose-1,6-bisphosphate aldolase (FBA)-a conserved glycolytic enzyme with virulence functions in bacteria: ‘ill met by moonlight’, *Biochem. Soc. Trans.*, 2014, **42**(6), 1792–1795, DOI: 10.1042/BST20140203.
- 27 A. Rodaki, T. Young and A. J. Brown, Effects of depleting the essential central metabolic enzyme fructose-1,6-bisphosphate aldolase on the growth and viability of *Candida albicans*: implications for antifungal drug target discovery, *Eukaryot. Cell*, 2006, **5**(8), 1371–1377, DOI: 10.1128/EC.00115-06.
- 28 I. Pawlaczyk-Graja, S. Balicki and K. A. Wilk, Effect of various extraction methods on the structure of polyphenolic-polysaccharide conjugates from *Fragaria vesca* L. leaf, *Int. J. Biol. Macromol.*, 2019, **130**, 664–674, DOI: 10.1016/j.ijbiomac.2019.03.013.
- 29 Y. Xie, L. Nurtay, Q. Sun, Q. Wang, C. Mu, Z. Liang, A. Amin and X. Li, Rhizoma polygonati from Mount Tai: nutritional value and usefulness as a traditional Chinese medicine, source of herbzyme, and potential remediating agent for COVID-19 and food shortage, *Acupuncture and Herbal Medicine*, 2021, **1**, 36–44.

