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Detection of sub-atomic movement in nanostructures†

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Nanoscale objects move fast and oscillate billions of times per second. Such movements occur naturally in the form of thermal (Brownian) motion while stimulated movements underpin the functionality of nano-mechanical sensors and active nano-(electro/opto) mechanical devices. Here we introduce a methodology for detecting such movements, based on the spectral analysis of secondary electron emission from moving nanostructures, that is sensitive to displacements of sub-atomic amplitude. We demonstrate the detection of nanowire Brownian oscillations of ∼10 pm amplitude and hyper-spectral mapping of stimulated oscillations of setae on the body of a common flea. The technique opens a range of opportunities for the study of dynamic processes in materials science, nanotechnology and biology.

Introduction

Electromagnetic and quantum forces become stronger as the physical dimensions of objects decrease (*e.g.*, the Coulomb force *vs.* separation between two electrons) and at sub-micron scales they begin to compete with weakening forces of elasticity. Movements also become faster as mass decreases with size, reaching Gigahertz frequencies at the nanoscale.[1,2] These considerations present substantial technological opportunity and explain the growing interest in the fundamentals of nano-to atomic-scale dynamics[3,4] and applications of nanomechanics to electronics and photonics.[5,6] Exciting new functionalities emerge through the exploitation of nanomechanical movements in photonic metamaterials with variable optical properties achieved by nanoscale movements of artificial "meta

molecules".[1,7–9] Entropic forces, Brownian motion, cooperative dynamics,[10] quantum effects, and Casimir and Van der Waals interactions play important roles in nano- to atomic-scale dynamics.[11] Over the last decade, protein (molecular) nano-mechanics has seen tremendous progress, evolving into a burgeoning field of biochemical research, while there is considerable interest in adopting bio-inspired solutions in advanced materials, nanostructures and bionics.[12,13] However, there are no routinely available imaging technologies that can quantitatively detect and spatially map fast, complex movements with sub-atomic amplitudes in nano-objects.[14] Conventional electron and scanning probe microscopies, for example, routinely provide nano-to sub-nanometre spatial (imaging) resolution but can operate only at low frame rates, typically of not more than a few tens per second.[15] Here we report on a technique that allows detection of movements of sub-atomic amplitude, and for spatial mapping of such movements with the nanometric image-plane resolution of a scanning electron microscope.

Results and discussion

In scanning electron microscopy, a static image of an object is typically created by raster scanning a focused beam of free electrons over the object and detecting the secondary electron emission current $I(r)$, where r is positional coordinate, integrated over a certain time interval at each pixel. This routinely delivers an image with nanoscale resolution, predominantly dependent on the focal spot size of the incident electron beam. However, at every point in the image plane and at time scales shorter than the per-pixel integration interval, the secondary electron current is sensitive to small (*e.g.*, Brownian) movements $\delta r(t)$ of the object (Fig. 1), giving rise to changes in current proportional to its gradient in the displacement direction: $\delta I(r,t) \sim (\nabla I(r) \times \delta r(t))$. Since $I(r)$ is measured in the conventional mode of secondary electron imaging, $\nabla I(r)$ can readily be evaluated, and then from measurements of $\delta I(r,t)$ at any/every point, movements $\delta r(t)$ of the object can be detected

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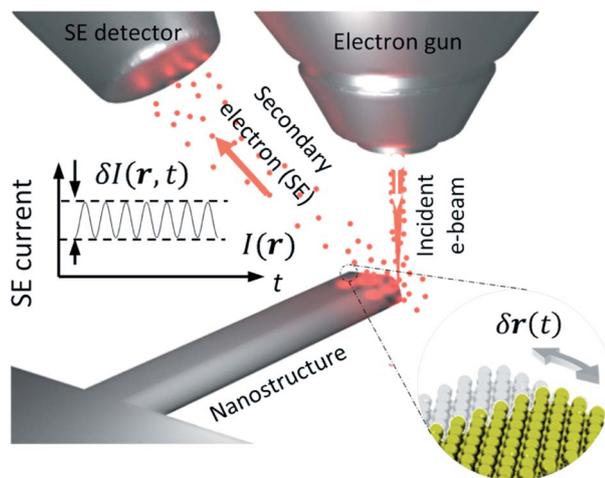
Fig. 1 Detecting movement in nanostructures. When a beam of free electrons is focused at the edge of an oscillating nanostructure, the secondary electron current $I(r,t)$ is modulated proportionally to the amplitude of movement $\delta r(t)$ and the gradient of its secondary electron emission profile at the incident electron injection point. Spectral analysis of the current as a function of spatial coordinate r over the object provides for mapping the oscillatory modes of the nanostructure.

and quantified. Displacements of picometric amplitude – smaller by orders of magnitude than the electron beam spot size – can be resolved in this way, as will be shown below.

To estimate the resolution of the technique, we assume that noise in the secondary electron current originates from the Poisson statistics of electrons incident on the detector. The signal-to-noise ratio (SNR) is then $\sqrt{N} = \sqrt{I\tau/e}$, where N is the number of detected electrons, e is electronic charge and τ is integration time. The secondary electron current gradient will be at a maximum at the sharp edges of a structure and limited by the spot size ρ of the incident electron beam. It can therefore be estimated as $\nabla I(r) \sim I/\rho$. When measuring the amplitude of displacement $a \sim |\delta r(t)|$ by detecting secondary electron current variation, unitary SNR will be achieved when $a \sim \rho/\sqrt{N} \sim \rho\sqrt{e/(I\tau)}$, giving an estimate of the resolution of the technique. For the typical values in a scanning electron microscope: $I \sim 1$ pA and $\rho \sim 1$ nm, a displacement of $a \sim 0.4$ pm could be measured with a one second integration time. By comparison, the lattice constants of solids are typically in the range from 300 to 700 pm and, by most definitions, the radii of isolated neutral atoms lie in the range between 30 and 300 pm.[16]

A point-by-point Fourier analysis of the secondary electron current provides for spatial mapping of the oscillatory modes of an object with nanoscale resolution determined by the focal spot size of the incident electron beam. This hyperspectral mode mapping can be performed within the bandwidth of the secondary electron detector and can enable mapping of Brownian motion in nanostructures. From thermodynamic considerations (the equipartition theorem), the root mean square (RMS) amplitude of Brownian motion for an oscillator with natural frequency f and effective mass m_{eff} is given by

$\langle a \rangle = 1/(2\pi f)\sqrt{k_{\mathrm{B}}T/m_{\mathrm{eff}}}$, where k_{B} is the Boltzmann constant and T is temperature. For example, a doubly clamped silicon nanowire 25 μm long and 200 nm × 200 nm in cross-section will have effective mass $\sim 9.28 \times 10^{-16}$ kg (the ratio of its effective mass and its physical mass is 0.4 in its fundamental mode[17]) and will oscillate at fundamental frequency $f \sim 3$ MHz. At room temperature ($T \sim 300$ K) it will exhibit thermal oscillations of RMS amplitude $\langle a \rangle \sim 100$ pm.

Application of the motion detection technique to the study of Brownian motion in nanostructures is illustrated in Fig. 2. The test structure consists of an array of nanowire beams of different lengths cut from a 50 nm thick silicon nitride membrane coated in a further-structured 50 nm layer of gold (as archetypal elements of, for example, a nano-mechanical photonic metamaterial). The beams range in length from 20 to 30 μm in 2 μm steps and have fundamental in-plane oscillation frequencies of between 2.11 and 1.17 MHz. With the incident electron beam (acceleration voltage 5 kV, beam current 86 pA) focused to a point at the edge of the central section of a beam, and with knowledge of the secondary electron current gradient ∇I at that point from the static image (DC component of the SE signal) of the sample $I(r)$, the frequency dispersion of calibrated in-plane Brownian displacement amplitude (Fig. 2b) can be derived from the measured frequency spectrum of secondary electron current $\delta r(\omega) \sim \delta I(\omega)/\nabla I$. The optimum injection point for each nanowire is selected on the basis of SNR (over an 18 ms integration time) from a line scan of several points traversing the high secondary electron gradient edges.

From experimental data, RMS displacement amplitude can be evaluated as $\langle a \rangle = \sqrt{\int_0^{+\infty} S_{\mathrm{th}}(f)\mathrm{d}f}$, where $S_{\mathrm{th}}(f)$ is the power spectral density (PSD) of displacement,[18] which in turn is obtained as the square of the displacement data plotted in Fig. 2b
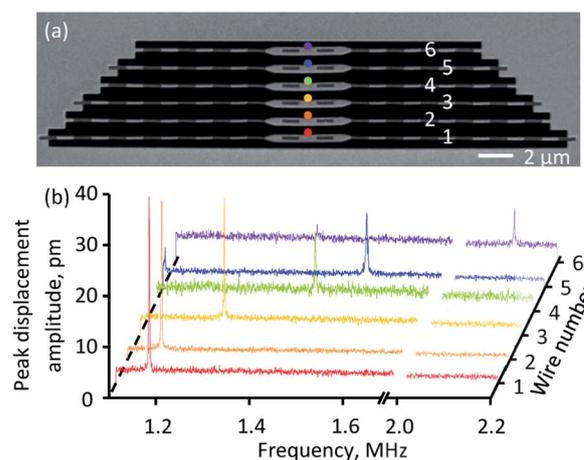

Fig. 2 Detecting thermal (Brownian) motion in nanostructures. (a) Static secondary electron image of an array of six free-standing, gold-coated silicon nitride nanowires with lengths between 20 and 30 μm. (b) Frequency dispersion of the nanowires' thermal [Brownian] motion displacement amplitude measured with the incident electron beam focused at the points denoted by colored dots in (a). Peaks correspond to the [length-dependent] fundamental in-plane oscillatory resonant frequencies.
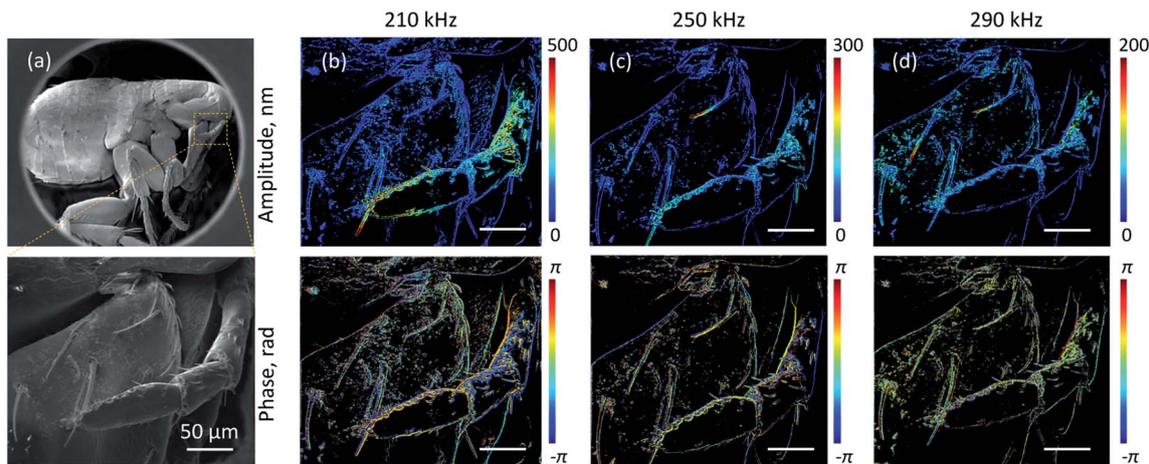
Fig. 3 Hyperspectral imaging of driven oscillations of flea setae. (a) Static secondary electron image of the entire flea and enlarged detail of the motion-imaged area. (b–d) Amplitude (upper row) and corresponding phase (lower) of secondary electron signal modulation at selected driving frequencies [as labelled].

divided by instrumental bandwidth. The area under the PSD curve gives the variance of displacement $\langle a^2 \rangle$. RMS displacement amplitudes of the six beams in the experimental sample are (in descending order of their lengths) 156, 152, 144, 132, 124 and 94 pm, *i.e.*, a small fraction of silicon–nitrides' 0.76 nm (β phase) crystal lattice parameter.[19] (We note here that the effect of electron beam-induced heating amounts to an estimated local temperature increase of only ∼1 K at the centre of a nanowire. With RMS displacement being proportional to the square root of temperature, this translates to a negligible perturbation of <0.25 pm against the measured ≥100 pm RMS nanowire displacements.)

Application of the motion detection technique to the spatial mapping of externally driven movements is illustrated in Fig. 3. For this purpose, samples are mounted on a piezoelectric actuator and the incident electron beam is raster-scanned over the sample, with the amplitude and phase of the secondary electron current oscillation detected at the piezo driving frequency by a lock-in amplifier. The point-by-point raster, with an integration time of 20 μs at each pixel (and as above a 5 kV, 86 pA electron beam), is externally triggered for consistent registration of phase over the entire image area. Fig. 3, referencing Robert Hooke's iconic 1665 optical microscopic image,[20] shows stimulated nano-motion electron microscopic images of a (deceased) flea at a selection of different driving frequencies. The imaged maxillary palpus and coxa region include setae that are a few tens of micrometres long and a few hundred nanometres in diameter, are here found to have natural oscillation frequencies in the sub-MHz range. The images (Fig. 3b–d) reveal resonant oscillatory displacement amplitudes reaching a few hundred nanometres (in response to a piezo actuator in-plane drive amplitude of ∼10 nm and against a background of the whole sample moving with said amplitude – dark blue features in the upper panels of Fig. 3b–d). The lower panels in Fig. 3b–d show the phase of the modulated secondary electron signal relative to the (piezo) driving force. This depends on both the local gradient and the motion mode shape of the structure: the

absence of phase discontinuities along the long edges of resonantly oscillating setae show that they are moving in a fundamental flexural mode, *i.e.* that all points along the length are moving in the same direction at the same time.

Conclusions

In summary, we have demonstrated that the time-domain analysis of secondary electron emission in electron microscopy can provide for mapping of fast (kHz–MHz) thermal and stimulated movement in nano/microstructures with sub-atomic displacement sensitivity, enabling the detection and hyperspectral spatial mapping of thermal (Brownian) motion, and coherent (externally driven) oscillations. The technique can be applied to a wide variety of artificial and naturally occurring objects, providing quantitative information directly on their vibrational modes, short-timescale 'ballistic' motion dynamics,[3,4,21] and indirectly on their mass, elasticity and the forces acting upon them.

Author contributions

J. Y. O. conceived the idea for the experiment. J. Y. O. and T. L. configured the instrument. T. L. carried out the measurements and manufactured the nanowire sample. All authors discussed the results, analysed the data, and contributed to writing the paper. K. F. M. and N. I. Z. supervised the work.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

References

1 N. I. Zheludev and E. Plum, *Nat. Nanotechnol.*, 2016, **11**, 16.

2 K. Ekinci and M. Roukes, *Rev. Sci. Instrum.*, 2005, **76**, 061101.

3 T. Li, S. Kheifets, D. Medellin and M. G. Raizen, *Science*, 2010, **328**, 1673–1675.

4 S. Kheifets, A. Simha, K. Melin, T. Li and M. G. Raizen, *Science*, 2014, **343**, 1493–1496.

5 M. Roukes, *Phys. World*, 2001, **14**, 25.

6 L. Midolo, A. Schliesser and A. Fiore, *Nat. Nanotechnol.*, 2018, **13**, 11–18.

7 X. Zhao, G. Duan, A. Li, C. Chen and X. Zhang, *Microsyst. Nanoeng.*, 2019, **5**, 1–17.

8 A. Karvounis, B. Gholipour, K. F. MacDonald and N. I. Zheludev, *Adv. Mater.*, 2019, **31**, 1804801.

9 A. Karvounis, N. Aspiotis, I. Zeimpekis, J. Y. Ou, C. C. Huang, D. Hewak and N. I. Zheludev, *Adv. Sci.*, 2019, **6**, 1900974.

10 A. Bernheim-Groswasser, N. S. Gov, S. A. Safran and S. Tzlil, *Adv. Mat.*, 2018, **30**, 1707028.

11 L. Woods, D. A. R. Dalvit, A. Tkatchenko, P. Rodriguez-Lopez, A. W. Rodriguez and R. Podgornik, *Rev. Mod. Phys.*, 2016, **88**, 045003.

12 M. Krieg, G. Fläschner, D. Alsteens, B. M. Gaub, W. H. Roos, G. J. Wuite, H. E. Gaub, C. Gerber, Y. F. Dufrêne and D. J. Müller, *Nat. Rev. Phys.*, 2019, **1**, 41–57.

13 F. Xia and L. Jiang, *Adv. Mater.*, 2008, **20**, 2842–2858.

14 S. Wang, Z. Shan and H. Huang, *Adv. Sci.*, 2017, **4**, 1600332.

15 J. I. Goldstein, *et al.*, *Scanning electron microscopy and X-ray microanalysis*, Springer International Publishing, 4th edn, 2017.

16 B. Averill and P. Eldredge, *General chemistry: principles, patterns, and applications*, 2011.

17 B. D. Hauer, C. Doolin, K. S. D. Beach and J. P. Davis, *Ann. Phys.*, 2013, **339**, 181–207.

18 M. Aspelmeyer, T. J. Kippenberg and F. Marquardt, *Rev. Mod. Phys.*, 2014, **86**, 1391.

19 D. Du Boulay, *et al.*, *Acta Crystallogr., Sect. B: Struct. Sci., Cryst. Eng. Mater.*, 2004, **60**, 388–405.

20 R. Hooke, *Micrographia: or some physiological descriptions of minute bodies made by magnifying glasses, with observations and inquiries thereupon*, The Royal Society, 1665.

21 Y. Shi, S. Xiong, L. K. Chin, J. Zhang, W. Ser, J. Wu, T. Chen, Z. Yang, Y. Hao, B. Liedberg, P. H. Yap, D. P. Tsai, C. W. Qiu and A. Q. Liu, *Sci. Adv.*, 2018, **4**, eaao0773.