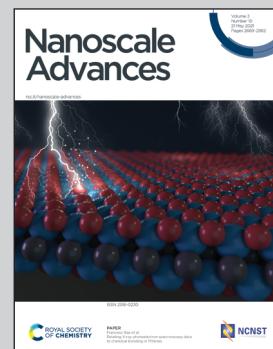


Showcasing research from Professor Mina Hoorfar's laboratory, School of Engineering, University of British Columbia, Kelowna, Canada.

Nanomaterial-based encapsulation for controlled gastrointestinal delivery of viable probiotic bacteria

Probiotics lose their viability during intestinal passage resulting in poor gastrointestinal delivery. Microencapsulation technology has been proposed as a successful strategy to address this problem by maintaining the viability of probiotics, thereby improving their efficacy following oral administration. Nanomaterials have demonstrated a significant promise as encapsulation materials to improve probiotics encapsulation. The integration of nanotechnology with microencapsulation techniques can improve the controlled delivery of viable probiotic bacteria to the gut.

### As featured in:



See Mina Hoorfar *et al.*,  
*Nanoscale Adv.*, 2021, **3**, 2699.

## REVIEW

[View Article Online](#)  
[View Journal](#) | [View Issue](#)



Cite this: *Nanoscale Adv.*, 2021, **3**, 2699

## Nanomaterial-based encapsulation for controlled gastrointestinal delivery of viable probiotic bacteria

Seyedehhamideh Razavi,<sup>†,a</sup> Sajjad Janfaza,<sup>†,a</sup> Nishat Tasnim,<sup>†,a</sup> Deanna L. Gibson<sup>bc</sup> and Mina Hoofar<sup>†,a\*</sup>

Probiotics are microorganisms that have beneficial health effects when administered in adequate dosages. The oral administration of probiotic bacteria is widely considered beneficial for both intestinal as well as systemic health but its clinical efficacy is conflicted in the literature. This may at least in part be due to the loss of viability during gastrointestinal passage resulting in poor intestinal delivery. Microencapsulation technology has been proposed as a successful strategy to address this problem by maintaining the viability of probiotics, thereby improving their efficacy following oral administration. More recently, nanomaterials have demonstrated significant promise as encapsulation materials to improve probiotic encapsulation. The integration of nanotechnology with microencapsulation techniques can improve the controlled delivery of viable probiotic bacteria to the gut. The current review aims at summarizing the types of nanomaterials used for the microencapsulation of probiotics and showing how they can achieve the delivery and controlled release of probiotics at the site of action.

Received 13th November 2020

Accepted 25th March 2021

DOI: 10.1039/d0na00952k

[rsc.li/nanoscale-advances](http://rsc.li/nanoscale-advances)

## 1. Introduction

The World Health Organization defines probiotics as live microorganisms that confer various health benefits to the host when administered in adequate amounts at the desired target site in the digestive tract.<sup>1</sup> Probiotics have been reported to offer protection from enteric infectious disease,<sup>2</sup> enhance metabolism (including lactose intolerance,<sup>3</sup> absorption of calcium,<sup>4</sup> synthesis of vitamins and pre-digestion of proteins<sup>5</sup>), have anti-carcinogenic and anti-mutagenic activities,<sup>6</sup> and reduce symptoms associated with anxiety and depression.<sup>7,8</sup> Probiotic bacteria provide positive health responses based on three mechanisms: production of nutrients and cofactors, competition with pathogens for nutrients or adhesion sites, and stimulation of the host immune response.<sup>9</sup> Given their immunomodulatory potential, probiotics have potential in the treatment of diseases such as inflammatory bowel disease (IBD) and cancer,<sup>10–12</sup> and have been proposed to treat a multitude of immune and metabolic-driven diseases.<sup>13,14</sup>

While the potential of the use of probiotics in the treatment or even prevention of gastrointestinal diseases is high, their clinical efficacy remains low due to conflicting clinical trial

results for many diseases.<sup>15,16</sup> This is in part due to the lack of viability of probiotics using traditional manufacturing and packaging methods.<sup>17</sup> Several factors (such as pH, oxygen, and temperature) can significantly affect the viability and survival rate of probiotics during processing, storage, and passage through the gastrointestinal tract.<sup>18</sup> Thus, probiotics must be able to survive the harsh gastric environment, remain metabolically active, and be released (in a controlled manner) in large enough quantities at the site of action in the lower gastrointestinal (GI) tract to confer beneficial health effects.<sup>19,20</sup> Microencapsulation of probiotics has been proposed as an effective solution to improve the survival, resistance, and targeted release of sensitive microorganisms in the GI tract.<sup>21</sup> It is a technology for entrapping small quantities of bioactive compounds and/or microorganisms in small polymer capsules. In essence, the goal of microencapsulation is to create a microenvironment that protects bacteria from exposure to exterior factors (such as low gastric pH) during digestion, and subsequently to reduce cell injury or cell death before their release at the target site.<sup>22</sup> There are many reports on micro-encapsulation of probiotic bacteria resulting in highly enhanced viability of probiotics during storage and administration.<sup>23</sup> The majority of these reports are based on the immobilization of probiotics into a polymer matrix, which retains its structure in the acidic environment of the stomach before swelling or dissolving in the intestine as a result of the pH change.<sup>24</sup> With the advent of nanotechnology, innovative encapsulating materials (EMs) have recently been developed based on nanostructured compounds. These nanomaterials have shown improved properties for the encapsulation of

<sup>a</sup>School of Engineering, University of British Columbia, Kelowna, BC, Canada. E-mail: [mina.hoofar@ubc.ca](mailto:mina.hoofar@ubc.ca)

<sup>b</sup>Department of Biology, Faculty of Science, University of British Columbia, Kelowna, Canada

<sup>c</sup>Department of Medicine, Faculty of Medicine, University of British Columbia, Vancouver, Canada

\* These authors contributed equally to this work and should be considered co-first authors.



probiotics. Due to their unique physical and chemical properties, nanostructured EMs demonstrate great promise for protection of microorganisms from the acidic conditions of the stomach and hence allow for the successful release of entrapped probiotic cells in the intestinal lumen with natural pH.

In this paper, we provide a critical review of nanomaterials in light of their suitability for probiotic microencapsulation. We provide an overview of nanostructured capsules used to protect probiotics from environmental and physiological factors (e.g., pH, oxygen, and temperature) and their advantages and limitations.

## 2. Microencapsulation of probiotics

Microencapsulation of probiotics has been widely utilized in the food and pharmaceutical industries. Microencapsulation results in the protection of probiotics by encapsulating them in small capsules. By building a functional barrier between the cells and environment, the capsule prevents probiotic bacteria from damage, leading to improved viability of probiotics. Encapsulated cells can reach the site of action without being adversely affected by environmental factors such as oxygen and pH.

Exposure to a high oxygen environment is harmful to microaerophilic and anaerobic probiotics such as *L. acidophilus* (microaerophilic) and bifidobacteria (obligate anaerobic). It can result in cell lysis due to the formation of reactive oxygen species such as superoxides that can cause oxidative DNA damage to bacterial cells. It has been shown that the viability of free anaerobic probiotic cells (e.g., *Bifidobacterium*) upon exposure to oxygen decreases,<sup>25</sup> while encapsulated cells survive for longer.<sup>26</sup>

Microencapsulation can also improve the survival of probiotic cells during gastrointestinal digestion by protecting them from the acidic environment of the stomach. It has been shown that a considerable number of probiotics (such as *Lactobacillus rhamnosus* and *Bifidobacterium longum*) are severely injured by the low pH of the stomach and high bile salt conditions of the intestine before entry into the colon.<sup>27</sup>

A key requirement for microencapsulation is the ability of the encapsulating material to release loaded probiotic cells from the microcapsules using environmental stimuli at the site of delivery. Encapsulated cells can be released by various mechanisms such as biodegradation, pH variation, mechanical rupture, and diffusion.<sup>9</sup> In recent years, promising encapsulating materials have been developed for the targeted delivery of probiotics in the gastrointestinal tract with improved mucoadhesive properties. Mucoadhesion is one of the most important factors determining the ability of encapsulating materials to adhere to mucosal membranes and provide temporary retention in the GI tract, resulting in an increased residence time of probiotics, reduced administration frequency, improved bioavailability, and improved targeting of particular sites.<sup>28</sup> The improvement of microcapsule mucoadhesion by the use of mucoadhesive encapsulating materials is, therefore, a great strategy for the delivery of probiotics to the gut. The low survival rate of probiotics due to undesirable reactions with external

factors such as oxygen, temperature, and light during handling and storage is another challenge to be addressed. In this regard, microencapsulation has shown great potential in increasing the stability of probiotics.<sup>29,30</sup>

Despite the remarkable improvement in viability and delivery of probiotic cells by encapsulation using traditional materials and methods, there is still a need to develop new and innovative microcapsules for better protection of probiotics and improved function. Using nanostructures as EMs has been proposed as a promising approach to design an improved probiotic delivery system.

## 3. Nanostructured encapsulating materials

Nanostructured materials for the encapsulation of probiotics can be selected from a wide variety of natural polymers (e.g. carbohydrates, gums, and proteins) and synthetic materials.

The composition of the EM affects the functional properties of the microcapsule,<sup>31</sup> and each group of encapsulating materials has its pros and cons in terms of mechanical, chemical, physical, and biological properties.

Recently, many nanomaterials with different sizes, shapes, textures, and compositions have been investigated for the microencapsulation of probiotics. Because of their unique physical and chemical properties, nanostructured microcapsules have shown promising improvement in protecting probiotics from harsh environments. The employed nanomaterials (e.g., nanocellulose) are nontoxic and compatible with both probiotics and the body while providing maximum protection to probiotic bacteria. Some of them are capable of releasing loaded probiotics under certain conditions (e.g. pH).<sup>19</sup> In the following subsections, we review these nanomaterials along with their applications in the microencapsulation of probiotics.

### 3.1. Nanocellulose

Cellulose is the most ubiquitous and abundant natural biomacromolecule.<sup>32</sup> The cellulose macromolecule consists of disordered (amorphous) regions and highly ordered (crystalline) regions.<sup>33</sup> Nano-sized cellulose, also known as nanocellulose, has recently received much attention because of its remarkable features including low risk of toxicity, biocompatibility, and tunable surface properties.<sup>34-36</sup> Nanocellulose is available in two forms: (1) cellulose nanocrystals (CNCs) and (2) cellulose nanofibers (CNFs), as shown in Fig. 1.<sup>37</sup> The former is extracted from fibers after complete dissolution of the amorphous fractions of cellulose in acid media, while the latter results from mechanical treatment leading to a high degree of fibrillation, which yields highly interconnected fibrils. CNCs and CNFs with high surface areas can be modified by a wide variety of additives.<sup>36</sup> Because of its physical and chemical properties, nanocellulose has attracted significant attention for applications in biomedical science and engineering.<sup>38</sup> Diverse nanocellulose-based drug delivery systems for the controlled release of drugs have been developed recently using nanocellulose.<sup>34</sup> More recently, it has been found that nanocellulose

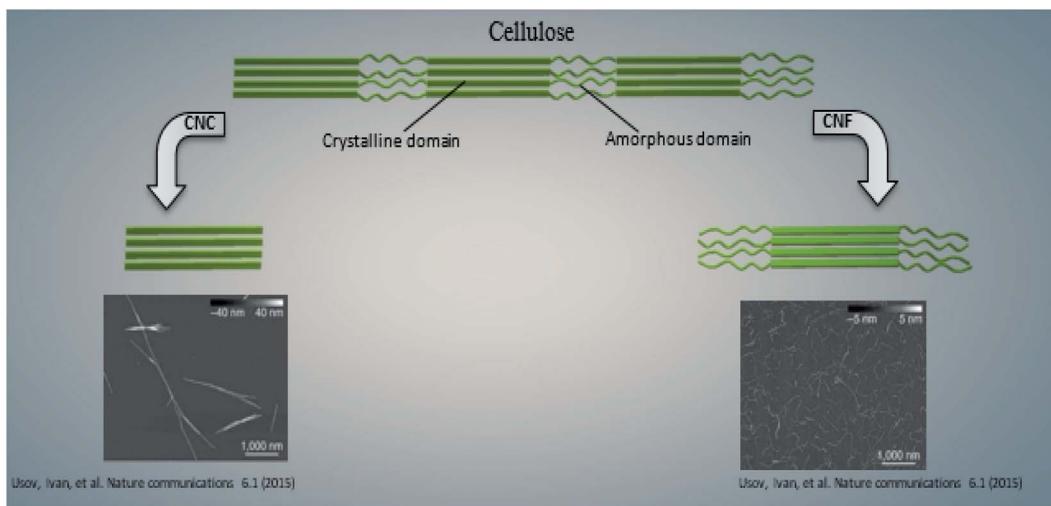


Fig. 1 The structure of cellulose nanocrystals (CNCs) and cellulose nanofibers (CNFs).

can improve the properties of probiotic delivery systems when used as the encapsulating material.<sup>39–41</sup> CNC is a promising cellulosic nanomaterial for applications involving non-refrigerated long shelf-life pharmaceutical products.<sup>39</sup> CNC improves the mechanical properties of the carrier. It has been shown that the presence of CNC in the formulation improves the compression strength of freeze-dried microbeads.<sup>39</sup> Also, CNC as an encapsulating material decreases gastric fluid absorption<sup>39</sup> and significantly improves the survival of probiotic bacteria in the gastrointestinal system.<sup>42</sup>

CNF can be used as a filler for various biomaterials and has strong intermolecular bonds.<sup>43</sup> It can form aqueous hydrogels for cell encapsulation, and the carboxyl groups in CNFs can be used to change the pore size of cellulose macrogels to improve their loading capacity.<sup>44,45</sup> For instance, macrogels made with a high content of CNFs showed sustained release of *Lactobacillus plantarum* cells and delivered adequate viable cells to the desired region in the intestinal tract.<sup>45</sup>

Nanocellulose can also act as an agent of cryoprotection.<sup>40</sup> Nanocellulose can be adsorbed on the surface of microorganisms and form a viscous layer that prevents the growth of ice crystals by increasing the viscosity of the solution and keeping the glassy structure of ice near the cells.<sup>40</sup> For instance, the combination of nanocellulose produced from cotton lint with conventional cryoprotective agents has been shown to improve the cell viability of *Lactobacillus plantarum* during freeze-drying.<sup>40</sup> Cellulose nanostructures can also be effective for the development of controlled release systems. Zhang *et al.*<sup>41</sup> developed microspheres with pH-responsive properties based on the sodium alginate (SA)/cellulose nanofiber. The probiotic *L. plantarum* was encapsulated by extruding a mixture of sodium alginate and TEMPO-oxidized CNFs. The acidic environment causes the formation of hydrogen bonds between SA and CNFs, stabilizing the gel microspheres and providing better protection for the encapsulated bacteria. Their results presented successful protection of cells in the simulated gastric fluid (SGF) and targeted release in the neutral simulated

intestinal fluid (SIF). In essence, the pH-responsive property of nanocellulose-based carriers is very important for intestine-targeted delivery of probiotics.

Nanocellulose can be obtained from different sources including plants and microorganisms.<sup>46,47</sup> Among naturally occurring nanocellulose sources, bacterial nanocellulose (BNC) has recently received great interest in the field of biomedical materials. BNC is biosynthesized as a primary extracellular metabolite by several bacterial species, such as *Komagataeibacter xylinus* (*K. xylinus*), during a fermentation process that results in the secretion of high-quality cellulose ribbons of microfibrillar bundles.<sup>48</sup> BNC is non-toxic<sup>49</sup> and can be produced in various shapes, sizes, and surface structures depending on the manufacturing process.<sup>50,51</sup> Compared to plant cellulose, BNC possesses higher purity, is devoid of other compounds present in plant pulp such as lignin and hemicellulose, and does not contain components of animal origin.<sup>52</sup> BNC also has more stable mechanical properties compared to plant cellulose.<sup>53</sup> BNC as a nanostructured biopolymer can improve probiotic protection due to its high crystallinity and available surface area.<sup>54</sup> Khorasani *et al.*<sup>54</sup> improved the survivability of *Bacillus coagulans* IBRC-M 10807 by encapsulating it into bionanocomposites composed of BNC, pectin, and *Schizophyllum commune* extract. Using the optimal bionanocomposite formulation (20% pectin with 80% BNC), the authors obtained a survival rate of 99.43% after microwave drying and 94.76% following sequential digestion under simulated gastrointestinal fluids. Moreover, BNC bionanocomposite encapsulation improved the viability of probiotic bacteria during long time storage at various temperatures.<sup>54</sup>

### 3.2. Chitosan nanoparticles

Chitosan (CS) is a polycationic natural polysaccharide derived from the alkaline deacetylation of chitin, a natural macromolecule forming the major constituent of arthropod exoskeletons and fungal cell walls. CS contains alternating units of (1 → 4) linked N-acetyl glucosamine and glucosamine units (Fig. 2a).<sup>55</sup>



Fig. 2 (a) Chemical structure of chitosan, and (b) the field emission scanning electron microscopy image of chitosan nanoparticles (the image was obtained with permission from Elsevier, copyright 2020).<sup>64</sup>

The cationic properties of CS, resulting from its primary amino groups, present great potential (due to its biocompatibility, non-toxicity, and low cost) for a wide range of biomedical applications (including tissue engineering, drug delivery, and wound dressing<sup>56–58</sup>), as well as applications in the food and pharmaceutical industries.<sup>59</sup> A variety of cells and molecules (including hydrophobic and hydrophilic molecules) can be encapsulated using CS.<sup>60</sup>

CS-based encapsulation of probiotic cells, in particular, has attracted a lot of attention recently, as CS protects encapsulated probiotics from the harsh conditions of the gastrointestinal tract and enhances its mucoadhesive properties.<sup>61</sup> The mucoadhesive property of CS makes it an ideal candidate nanomaterial for developing controlled release drug-delivery systems. In essence, intestinal mucus is composed of negatively charged mucins. The structure of intestinal mucus offers many opportunities for the development of mucoadhesive delivery systems. Chitosan with a positive charge can form an electrostatic attraction with the sticky layer of mucus. Also, physical entanglements between chitosan and mucus components can enhance the adhesion of chitosan-based capsules to the gastric mucosa.<sup>62,63</sup>

More recently, chitosan nanoparticles (CSNPs) have been proposed for the micro- and nano-encapsulation of cells and molecules,<sup>64</sup> respectively, including probiotic bacteria (Fig. 2b). Delivery systems based on CS-nanoparticles (CSNPs) have shown great mucoadhesive strength compared to bulk CS due to their significantly higher surface-to-volume ratio.<sup>28,58,65</sup> There are various methods for the synthesis of CSNPs including ionotropic gelation, microemulsion, polyelectrolyte complexation, emulsification solvent diffusion, and the reverse micellar method.<sup>66</sup> CSNPs also have excellent physicochemical, antimicrobial, and biological properties that make them a promising biopolymer for drug delivery applications.<sup>67</sup> Several studies have shown the ability of CSNPs to improve the bioavailability and

resistance of encapsulated drugs for effective GI delivery.<sup>68,69</sup> CSNPs along with alginate have also been used to encapsulate probiotic *Escherichia coli* Nissle 1917 (EcN). Microencapsulated EcN was able to resist environmental stresses like low pH, high bile salt concentration, and high temperature. The survivability of encapsulated probiotic cells was significantly increased in an acidic environment (pH 1.5), high bile salt concentration (4%), and high temperature (70 °C) compared to non-encapsulated cells. Moreover, encapsulated EcN showed antimicrobial properties against *Campylobacter jejuni*, *i.e.*, a leading bacterial pathogen responsible for gastroenteritis in humans. A reduction in the *Campylobacter jejuni* growth (by 2 log CFU) after exposure to microencapsulated EcN was observed.<sup>70</sup> It has also been shown that the released bacteria have better contact with intestinal mucosa, which facilitates the successful colonization of probiotic cells in the target region. This is vital for pharmaceutical applications of probiotics and causes reduction of the required dose or frequency of administration.<sup>28,65</sup> For instance, it has been reported that EcN encapsulated in CSNPs and alginate efficiently adhered to intestinal HT-29 cells reduce the invasion of the pathogenic microorganism *C. jejuni*.<sup>70</sup>

### 3.3. Eudragit S100 nanoparticles

Eudragit®S100 [Eudragit S100] is a non-toxic anionic polymer synthesized from methacrylic acid and methacrylic acid methyl ester.<sup>71,72</sup> The solubility of Eudragit®S100 is pH-dependent: it is insoluble in strongly acidic solutions, while it is slightly soluble in the region of the digestive tract with neutral to weakly alkaline pH.<sup>73,74</sup>

Multi-walled microcapsules based on Eudragit S100 nanoparticles, alginate, and chitosan have been developed for improving the viability of the probiotic bacteria *Lactobacillus acidophilus* and *Lactobacillus rhamnosus* in simulated gastrointestinal conditions.<sup>75</sup> In essence, the negatively charged Eudragit S100 nanoparticles attract positively charged chitosan



Table 1 Nanostructured capsules used for the delivery of probiotics to the gastrointestinal tract

Microencapsulating material	Probiotic	Storage conditions	Functionality	Freeze drying	Size	Microencapsulation technique	Ref.
Sodium alginate (SA)/cellulose nanofiber (CNF)	<i>Lactobacillus plantarum</i> CICC 6240	—	—	Yes	A width of less than 50 nm and a length of less than 500 nm	Extrusion	41
Bacterial nanocellulose (BNC), pectin, and schizophyllum commune extract	<i>Bacillus coagulans</i> IBRC-M 10807	At 25 °C, 4 °C, and –20 °C for 7, 15 and 30 days	Survival in the GI tract model	No	50 nm	—	54
Whey protein isolate-crystalline nanocellulose-inulin	<i>Lactobacillus rhamnosus</i> ATCC 7469	—	Survival in the GI tract model	Yes	—	—	56
Alginate-cellulose nanocrystal (CNC)-lecithin	<i>Lactobacillus rhamnosus</i> ATCC 9595	At 4 °C and 25 °C up to 42 days	Survival in the GI tract model	Yes	—	Extrusion	39
Alginate and chitosan nanoparticles (CSNPs)	<i>Escherichia coli</i> Nissle 1917 (EcN)	At 4 °C	Survival in the GI tract model	Yes	—	Extrusion	70
Eudragit S100 nanoparticles, alginate, and chitosan	<i>Lactobacillus acidophilus</i> and <i>Lactobacillus rhamnosus</i>	—	Survival in the GI tract model	No	100 nm	Extrusion	75
Alginate-gelatin-MgO NPs	<i>Pediococcus pentosaceus</i> Li05	At 4 °C for 4 weeks	Survival in the GI tract model	Yes	—	Extrusion	80
Native starch and starch nanoparticles (SNPs)	<i>Pediococcus acidolactici</i>	—	Survival in the GI tract model	Yes	271 nm	Extrusion	90
Starch nanocrystal-alginate	<i>Lactobacillus brevis</i> ST-69	At 4 °C for 10 weeks	Survival in the GI tract model	NO	—	Emulsion technique	92

in the first coating layer with ionic bonds to form the second layer. Since the chitosan layer is porous, coating it with Eudragit S100 nanoparticles covers the porous beads and forms an airtight bubble that protects the encapsulated core from adverse environments.<sup>75</sup> Studies have shown that with Eudragit nanoparticles as the outer coating layer there is an increase in the strength of the encapsulated beads and an improvement in the viability of probiotic bacteria in comparison to single-coated beads.<sup>75</sup>

### 3.4. Magnesium oxide nanoparticles

Magnesium oxide (MgO) is a functional semiconductor with many applications ranging from pharmaceuticals to optoelectronics.<sup>76</sup> MgO is administered medically for the relief of cardiovascular diseases and stomach problems.<sup>77</sup> Recently nanoscale MgO has attracted significant attention due to its high surface area, nontoxicity, high mechanical strength, thermal stability, and low cost.<sup>78</sup> Magnesium oxide nanoparticles (MgO NPs) have been used in the industry for pharmaceuticals, toxic waste remediation, and toxic gas removal.<sup>79</sup> More recently, MgO NPs have been used for the

microencapsulation of probiotics. For instance, Yao *et al.*<sup>80</sup> have encapsulated a model probiotic (*Pediococcus pentosaceus* Li05) in alginate-gelatin microbeads in the presence of MgO NPs. Probiotics encapsulated in MgO-loaded microcapsules have shown better viability (less than 2 log CFU reduction after 40 min incubation) than free bacterial cells (5 log CFU reduction in 10 min) in gastric fluids.<sup>80</sup> There are two potential reasons for this cell survival improvement: MgO NPs fill pores inside the alginate-gelatin microgels, which may inhibit the oxygen exposure of probiotics, and MgO NPs neutralize hydrogen ions in the acidic environment, protecting probiotics in gastric fluids.

### 3.5. Starch nanoparticles

Starch is one of the most abundant biopolymers in nature produced by many plants and crops such as cereals.<sup>81,82</sup> Starch consists of glucose residues linked by two types of bonds:  $\alpha$ -1,4, and  $\alpha$ -1,6 glucosidic linkages. Despite its simple chemistry, the starch granule is a complex semicrystalline structure (up to 100  $\mu$ m long), containing linear amylose and highly branched amylopectin.<sup>83</sup> Amylose has an amorphous structure



comprising linear chains of  $\alpha$ -1,4-linked glucose units.<sup>83</sup> Amylopectin is a semicrystalline polysaccharide and consists solely of D-glucose residues covalently interconnected mostly by  $\alpha$ -(1  $\rightarrow$  4) glucosidic bonds, but also 4–5%  $\alpha$ -(1  $\rightarrow$  6) glucosidic bonds.<sup>84</sup>

In recent years, the preparation and application of starch nanostructures have attracted significant research interest. Starch nanoparticles (SNPs) and nanocrystals (SNCs) are two nanostructures that have been widely used for biomedical applications, especially drug delivery applications.<sup>85,86</sup> SNPs and SNCs are both nano-sized. SNCs refer to the crystalline portion of starch granules remaining after acid or enzymatic hydrolysis of their amorphous structure.<sup>87,88</sup> SNCs have a crystal structure, while SNPs are amorphous. SNPs are generated from congealed starch, while starch crystallites develop as a result of the disruption of non-crystalline domains of semi-crystalline granules.<sup>89</sup>

Despite showing promising results for the delivery of bioactive compounds, SNPs may not be a great candidate for the microencapsulation of probiotics. *Pediococcus acidilactici* encapsulated using SNPs showed lower viability (1.47 log CFU g<sup>-1</sup>) as compared to that encapsulated in native starch (2.65 log CFU g<sup>-1</sup>). In essence, SNPs cannot contain probiotic cells inside nanostructured microcapsules, and hence cells are released immediately in an environment with harsh conditions leading to cell death. Besides, cells are not protected against higher temperatures by SNPs; however, significantly better viability can be achieved at lower temperatures.<sup>90</sup>

Chemical modification of SNPs has been widely used to improve their properties *via* three important strategies namely

chemical reactions with small molecules, the 'grafting onto' strategy (*i.e.*, grafting of polymer chains in the presence of coupling agents), and the 'grafting from' strategy (*i.e.*, polymer chain grafting by polymerization of monomers).<sup>91</sup> The chemical modification of SNPs in combination with another encapsulating material may address their instability and improve their application in the microencapsulation of probiotics.

In contrast to SNPs, SNCs are reported<sup>92</sup> to protect probiotics by encapsulation in alginate–starch nanocrystals under simulated gastrointestinal conditions and during long-life storage at 4 °C. This proves that alginate and starch nanocrystals are appropriate materials for probiotic encapsulation. As an example, *Lactobacillus brevis* encapsulated in alginate–nanocrystalline starch gel capsules showed a high survival rate of 94.97% in simulated gastrointestinal fluids. They also improved storage at 25 °C by up to two weeks. As a result, alginate–nanocrystalline starch is a promising material for probiotic preservation at room temperature (Table 1).

#### 4. Methods of nanostructured encapsulation

Several methods have been used for microencapsulation of probiotics, including chemical methods (such as interfacial polymerization), physical methods (such as spray drying), and physicochemical methods (such as coacervation and ionic gelation).<sup>93,94</sup> Many of these microencapsulation methods can result in a significant decrease in probiotic cell viability because of the high temperature range or organic agents used in the encapsulation process.<sup>95</sup> The principles of these methods have

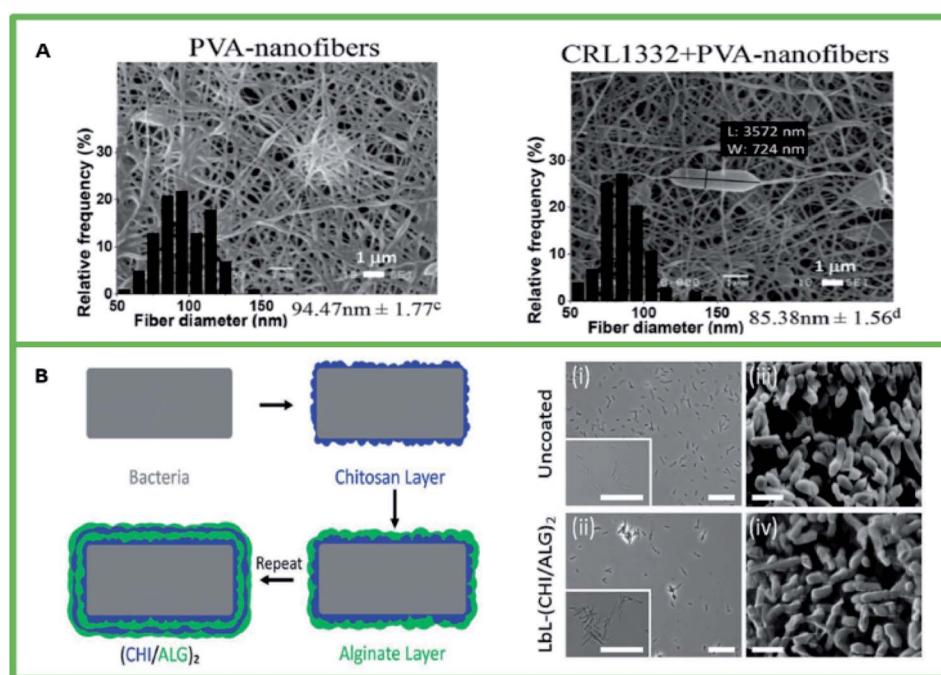


Fig. 3 (A) Scanning electron microscopy of PVA nanofibers (left) and PVA nanofibers containing *L. rhamnosus* CRL1332 (right) obtained by electrospinning. Reproduced with permission from Elsevier, copyright 2021.<sup>100</sup> (B) Layer-by-layer encapsulation of probiotic cells. (Left) Schematic LbL templating of chitosan and alginate on bacteria. (Right) Optical microscopy images of (i) uncoated bacteria and (ii) bacteria coated with CHI/ALG. SEM images of (iii) uncoated bacteria and (iv) bacteria coated with CHI/ALG. Reproduced with permission from Wiley, copyright 2021.<sup>99</sup>



been comprehensively reviewed elsewhere.<sup>96,97</sup> Here, we highlight two emerging and promising strategies for nanostructured encapsulation of probiotic cells, namely electrospinning<sup>98</sup> and the layer-by-layer (LbL) method.<sup>99</sup>

#### 4.1. Electrospinning

Electrospinning has demonstrated great potential for the encapsulation of bacterial cells by nanofibers that protect probiotic cells from harsh environmental conditions in the gut and enhance their survival (Fig. 3A). Electrospun nanofibers based on several polymers (such as polyvinyl alcohol (PVA),<sup>100</sup> fructooligosaccharide (FOS)/PVA,<sup>98</sup> gum arabic/pullulan,<sup>101</sup> and alginate<sup>102</sup>) have demonstrated the ability to protect probiotic cells from harsh environments. As an example, the encapsulation of *Lactobacillus plantarum* by fructooligosaccharides developed by electrospinning was studied by Feng *et al.*<sup>98</sup> Prebiotics are indigestible fibers that serve as substrates for probiotics (such as *Bifidobacteria* spp. and *Lactobacilli*) and selectively stimulate their growth and activity. The co-administration of probiotics and prebiotics (such as oligosaccharides and inulin) is enabled by nanostructured encapsulation techniques and has been proposed as a new strategy to improve probiotic survival, for example, by protecting them from acidity and bile salts,<sup>103</sup> and promoting probiotic cell proliferation. It has also been reported that electrospun FOS/PVA nanofibers can significantly improve the survivability of encapsulated cells under moist heat treatment.<sup>98</sup> Electrospun nanofibers may be composed of a single polymer or a combination of two or more polymers. Encapsulation by alginate-based electrospun nanofiber mats has been shown to protect probiotic bacteria from the harsh gastrointestinal environment. A viability rate of 64.1 to 70.8 log CFU mL<sup>-1</sup> has been achieved for *Lactobacillus paracasei* KS-199 encapsulated by alginate-based nanofibers in simulated gastrointestinal fluids.<sup>102</sup> Electrospun nanofibers based on gum arabic/pullulan are another new type of material that can be used for the protection of probiotic cells. For instance, *Lactobacillus*-loaded electrospun nanofibres showed a probiotic survivability of 85.38–97.83% and retained viability during 28 day storage at 4 °C.<sup>101</sup>

#### 4.2. Layer-by-layer method

The layer-by-layer (LbL) method is another promising approach to encapsulate and introduce specific probiotic species into the GI tract (Fig. 3B). This strategy is based on coating alternating layers of cationic (e.g., chitosan) and anionic (e.g., alginate) polymers on bacteria *via* electrostatic interaction.<sup>99</sup> The LbL method enhances bacterial protection against acidic environments and bile salts, resulting in the proliferation of encapsulated probiotics on intestinal tissues. This method requires the minimum amount of polymer for complete encapsulation. The LbL technique can be utilized to encapsulate various types of probiotic cells and produce homogeneous nanocoatings with precise control of the structure.<sup>104</sup>

Encapsulating probiotic cells using the LbL technique has been shown to improve bacterial protection in the

gastrointestinal tract and promote microbial adhesion and growth at the targeted sites.<sup>99,105,106</sup> For instance, Periya *et al.*<sup>105</sup> have shown an enhancement in the survival of probiotic *Lactobacillus acidophilus* in adverse conditions encountered in the GI tract by encapsulation through LbL self-assembly of polyelectrolytes, chitosan, and carboxymethyl cellulose. The results showed that about 106 CFU/500 mg (33 log%) of bacterial cells with nanocoatings survived when exposed to simulated gastric and intestinal fluids for 120 min; however, almost complete death of free cells is observed.<sup>105</sup>

## 5. Summary and future perspectives

Probiotics as pharmaceutical and nutraceutical products are becoming more and more popular in the market. Microencapsulation of probiotics offers several advantages including (1) improving the survival of probiotic bacteria by protecting them against the harsh gastrointestinal environment and undesirable reactions with external factors (such as oxygen, temperature, and light during handling and storage); (2) promoting the effective attachment of probiotics to intestinal mucosal tissues; (3) ensuring the controlled and targeted release of the encapsulated materials with desired concentrations in the gastrointestinal tract; and (4) increasing the ability to incorporate desirable concentrations of probiotics (ranging from low to high levels).

Various types of synthetic and natural materials have been traditionally used as EMs; however, there is still room for improvement. The use of nanomaterials in microencapsulation is an innovative approach that enables the manipulation of the physical and chemical properties of microcapsules to improve the delivery of active probiotics to the site of action.

Various types of nanostructures including cellulose, chitosan, MgO, starch, and PLGA nanoparticles have been employed as encapsulating materials to address two main challenges: (1) the viability of probiotics during processing, long-term storage, and passage through the gastrointestinal tract, and (2) controlled release of probiotics in the desired region of the intestinal tract.

The coating of microcapsules with certain materials, such as Eudragit S100 and MgO nanoparticles, has been shown to fill pores in the hydrogel structure of the beads and increase protection from harsh environments for probiotic bacteria. Other nanomaterials such as nanocellulose can be adsorbed on the surface of microorganisms to form a viscous layer that acts as a barrier against the growth of ice crystals near the cells, thereby protecting them during freeze-drying. Also, different types of nanofibers and nanocoatings (e.g., alginate electrospun nanofibers) have been used to improve the viability of probiotic cells during storage and gastrointestinal digestion. EMs can protect probiotic cells from the acidic pH of the stomach and release them in the gut. Moreover, the major challenge faced during the oral administration of probiotic bacteria is the rapid transit from the intestine in feces; this can be addressed by the use of some nanomaterials such as chitosan nanoparticles or nanocoatings, which have been shown to enhance mucoadhesion to the gut wall.

Future advances in nanotechnology and nanostructured microencapsulation techniques can lead to the development of



improved probiotic products that can further our understanding of their health benefits.

## Conflicts of interest

There are no conflicts to declare.

## Notes and references

- 1 A. Sharma, *Importance of Probiotics in Cancer Prevention and Treatment, recent developments in applied microbiology and biochemistry*, Elsevier, 2019, pp. 33–45.
- 2 M. S. R. Rajoka, H. Zhao, H. M. Mehwish, N. Li, Y. Lu, Z. Lian, D. Shao, M. Jin, Q. Li and L. Zhao, Anti-tumor potential of cell free culture supernatant of *Lactobacillus rhamnosus* strains isolated from human breast milk, *Food Res. Int.*, 2019, **123**, 286–297.
- 3 T. Ayicew, A. Belete, T. Alebachew, H. Tsehay, H. Berhanu and A. Minwuyelet, Bacterial probiotics their importances and limitations: a review, *J. Nutr. Health Sci.*, 2017, **4**, 202.
- 4 M. R. Dubey and V. P. Patel, Probiotics: A Promising Tool for Calcium Absorption, *Open Nutr. J.*, 2018, **12**(1), 59–69.
- 5 L. Lazo-Pérez, Q. Ruiz, A. Elías, F. Herrera and I. Z. Rodríguez, Effect of a VITAFERT microbial additive on some bioprotective and health indicators in growing pigs, *Cuban J. Agric. Sci.*, 2018, **51**(3), 321–328.
- 6 F. Aboulfazli, A. B. Shori and A. S. Baba, Effects of the replacement of cow milk with vegetable milk on probiotics and nutritional profile of fermented ice cream, *LWT-Food Sci. Technol.*, 2016, **70**, 261–270.
- 7 M. Pirbaglou, J. Katz, R. J. de Souza, J. C. Stearns, M. Motamed and P. Ritvo, Probiotic supplementation can positively affect anxiety and depressive symptoms: a systematic review of randomized controlled trials, *Nutr. Res.*, 2016, **36**, 889–898.
- 8 S. Kamiya, H. Yonezawa and T. Osaki, Role of Probiotics in Eradication Therapy for Helicobacter pylori Infection, *Helicobacter pylori in Human Diseases: Advances in Microbiology, Infectious Diseases and Public Health*, 2019, **11**, 243–255.
- 9 M. T. Cook, G. Tzortzis, D. Charalampopoulos and V. V. Khutoryanskiy, Microencapsulation of probiotics for gastrointestinal delivery, *J. Controlled Release*, 2012, **162**, 56–67.
- 10 A. Tarasiuk and G. Eibl, Nutritional support and probiotics as potential treatment in IBD, *Curr. Drug Targets*, 2020, **21**(14), 1417–1427.
- 11 S. Guandalini and N. Sansotta, Probiotics in the treatment of inflammatory bowel disease, *Probiotics and Child Gastrointestinal Health*, Springer, 2019, pp. 101–107.
- 12 D. Gibson, A. Godovannyi and S. Gill, Probiotic compositions and uses thereof, *US pat.*, US20200069747A1, 2020, <https://patents.google.com/patent/US20200069747A1/en>.
- 13 J. Aggarwal, G. Swami and M. Kumar, Probiotics and their effects on metabolic diseases: an update, *J. Clin. Diagn. Res.*, 2013, **7**, 173.
- 14 K. J. Chua, W. C. Kwok, N. Aggarwal, T. Sun and M. W. Chang, Designer probiotics for the prevention and treatment of human diseases, *Curr. Opin. Chem. Biol.*, 2017, **40**, 8–16.
- 15 K. Jia, X. Tong, R. Wang and X. Song, The clinical effects of probiotics for inflammatory bowel disease: A meta-analysis, *Medicine*, 2018, **97**(51), e13792.
- 16 Z. Akram, S. S. Shafqat, S. Aati, O. Kujan and A. Fawzy, Clinical efficacy of probiotics in the treatment of gingivitis: A systematic review and meta-analysis, *Aust. Dent. J.*, 2020, **65**, 12–20.
- 17 J. Suez, N. Zmora, E. Segal and E. Elinav, The pros, cons, and many unknowns of probiotics, *Nat. Med.*, 2019, **25**, 716–729.
- 18 N. P. Shah, W. E. Lankaputhra, M. L. Britz and W. S. Kyle, Survival of *Lactobacillus acidophilus* and *Bifidobacterium bifidum* in commercial yoghurt during refrigerated storage, *Int. Dairy J.*, 1995, **5**, 515–521.
- 19 A. Geirnaert, A. Steyaert, V. Eeckhaut, B. Debruyne, J. B. Arends, F. Van Immerseel, N. Boon and T. Van de Wiele, *Butyricicoccus pullicaeorum*, a butyrate producer with probiotic potential, is intrinsically tolerant to stomach and small intestine conditions, *Anaerobe*, 2014, **30**, 70–74.
- 20 S. Raghuwanshi, S. Misra, R. Sharma and P. Bisen, Probiotics: nutritional therapeutic tool, *J. Probiotics Health*, 2018, **6**, 2.
- 21 D. Schell and C. Beermann, Fluidized bed microencapsulation of *Lactobacillus reuteri* with sweet whey and shellac for improved acid resistance and in-vitro gastro-intestinal survival, *Food Res. Int.*, 2014, **62**, 308–314.
- 22 M. Jantzen, A. Göpel and C. Beermann, Direct spray drying and microencapsulation of probiotic *Lactobacillus reuteri* from slurry fermentation with whey, *J. Appl. Microbiol.*, 2013, **115**, 1029–1036.
- 23 T. Petrović, V. Nedović, S. Dimitrijević-Branković, B. Bugarski and C. Lacroix, Protection of probiotic microorganisms by microencapsulation, *Chem. Ind. Chem. Eng. Q.*, 2007, **13**, 169–174.
- 24 S. Chen, Y. Cao, L. R. Ferguson, Q. Shu and S. Garg, The effect of immobilization of probiotic *Lactobacillus reuteri* DPC16 in sub-100  $\mu\text{m}$  microcapsule on food-borne pathogens, *World J. Microbiol. Biotechnol.*, 2012, **28**, 2447–2452.
- 25 J.-B. Ahn, H.-J. Hwang and J.-H. Park, Physiological responses of oxygen-tolerant anaerobic *Bifidobacterium longum* under oxygen, *J. Microbiol. Biotechnol.*, 2001, **11**, 443–451.
- 26 A. Talwalkar and K. Kailasapathy, Effect of microencapsulation on oxygen toxicity in probiotic bacteria, *Aust. J. Dairy Technol.*, 2003, **58**, 36.
- 27 W. Ding and N. P. Shah, An improved method of microencapsulation of probiotic bacteria for their stability in acidic and bile conditions during storage, *J. Food Sci.*, 2009, **74**, M53–M61.



28 S. Chen, Y. Cao, L. R. Ferguson, Q. Shu and S. Garg, Evaluation of mucoadhesive coatings of chitosan and thiolated chitosan for the colonic delivery of microencapsulated probiotic bacteria, *J. Microencapsulation*, 2013, **30**, 103–115.

29 L. A. A. Lopes, R. d. S. F. Carvalho, N. S. S. Magalhães, M. S. Madruga, A. J. A. A. Athayde, I. A. Portela, C. E. Barão, T. C. Pimentel, M. Magnani and T. C. M. Stamford, Microencapsulation of *Lactobacillus acidophilus* La-05 and incorporation in vegan milks: Physicochemical characteristics and survival during storage, exposure to stress conditions, and simulated gastrointestinal digestion, *Food Res. Int.*, 2020, **135**, 109295.

30 H. Liu, S. W. Cui, M. Chen, Y. Li, R. Liang, F. Xu and F. Zhong, Protective approaches and mechanisms of microencapsulation to the survival of probiotic bacteria during processing, storage and gastrointestinal digestion: a review, *Crit. Rev. Food Sci. Nutr.*, 2019, **59**, 2863–2878.

31 Y. Byun, Y. T. Kim, K. G. H. Desai and H. J. Park, Microencapsulation techniques for food flavour, *Chem. Biol. Volatiles*, 2010, 307–332.

32 Q. Shi, D. Liu, Y. Wang, Y. Zhao, X. Yang and J. Huang, High-Performance Sodium-Ion Battery Anode via Rapid Microwave Carbonization of Natural Cellulose Nanofibers with Graphene Initiator, *Small*, 2019, **15**, 1901724.

33 N.-M. Park, S. Choi, J. E. Oh and D. Y. Hwang, Facile extraction of cellulose nanocrystals, *Carbohydr. Polym.*, 2019, **223**, 115114.

34 S. Salimi, R. Sotudeh-Gharebagh, R. Zarghami, S. Y. Chan and K. H. Yuen, Production of nanocellulose and its applications in drug delivery: A critical review, *ACS Sustainable Chem. Eng.*, 2019, **7**, 15800–15827.

35 H. Liu, B. Geng, Y. Chen and H. Wang, Review on the aerogel-type oil sorbents derived from nanocellulose, *ACS Sustainable Chem. Eng.*, 2017, **5**, 49–66.

36 L. Kian, N. Saba, M. Jawaid and M. Sultan, A review on processing techniques of bast fibers nanocellulose and its polylactic acid (PLA) nanocomposites, *Int. J. Biol. Macromol.*, 2019, **121**, 1314–1328.

37 F. Hoeng, A. Denneulin and J. Bras, Use of nanocellulose in printed electronics: a review, *Nanoscale*, 2016, **8**, 13131–13154.

38 S. Mondal, Preparation, properties and applications of nanocellulosic materials, *Carbohydr. Polym.*, 2017, **163**, 301–316.

39 T. Huq, C. Fraschini, A. Khan, B. Riedl, J. Bouchard and M. Lacroix, Alginate based nanocomposite for microencapsulation of probiotic: Effect of cellulose nanocrystal (CNC) and lecithin, *Carbohydr. Polym.*, 2017, **168**, 61–69.

40 F. K. Nahr, R. R. Mokarram, M. A. Hejazi, B. Ghanbarzadeh, M. S. Khiyabani and K. Z. Benis, Optimization of the nanocellulose based cryoprotective medium to enhance the viability of freeze dried *Lactobacillus plantarum* using response surface methodology, *LWT-Food Sci. Technol.*, 2015, **64**, 326–332.

41 H. Zhang, C. Yang, W. Zhou, Q. Luan, W. Li, Q. Deng, X. Dong, H. Tang and F. Huang, A pH-responsive gel macrosphere based on sodium alginate and cellulose nanofiber for potential intestinal delivery of probiotics, *ACS Sustainable Chem. Eng.*, 2018, **6**, 13924–13931.

42 O. Maleki, M. A. Khaledabad, S. Amiri, A. K. Asl and S. Makouie, Microencapsulation of *Lactobacillus rhamnosus* ATCC 7469 in whey protein isolate-crystalline nanocellulose-inulin composite enhanced gastrointestinal survivability, *LWT-Food Sci. Technol.*, 2020, 109224.

43 A. Amalraj, S. Gopi, S. Thomas and J. T. Haponiuk, *Cellulose Nanomaterials in Biomedical, Food, and Nutraceutical Applications: A Review*, *Macromolecular Symposia*, Wiley Online Library, 2018, p. 1800115.

44 R. Das and J. G. Fernandez, Cellulose nanofibers for encapsulation and pluripotency preservation in the early development of embryonic stem cells, *Biomacromolecules*, 2020, **21**(12), 4814–4822.

45 Q. Luan, W. Zhou, H. Zhang, Y. Bao, M. Zheng, J. Shi, H. Tang and F. Huang, Cellulose-based composite macrogels from cellulose fiber and cellulose nanofiber as intestine delivery vehicles for probiotics, *J. Agric. Food Chem.*, 2018, **66**, 339–345.

46 M. Osorio, A. Cañas, R. Zuluaga, P. Gañán, I. Ortiz and C. Castro, Nanocellulose-Based Composites in Biomedical Applications, *Adv. Green Compos.*, 2018, 369.

47 M. M. Haafiz, A. Hassan, Z. Zakaria, I. Inuwa and M. Islam, Physicochemical characterization of cellulose nanowhiskers extracted from oil palm biomass microcrystalline cellulose, *Mater. Lett.*, 2013, **113**, 87–89.

48 D. Klemm, E. D. Cranston, D. Fischer, M. Gama, S. A. Kedzior, D. Kralisch, F. Kramer, T. Kondo, T. Lindström and S. Nietzsche, Nanocellulose as a natural source for groundbreaking applications in materials science: Today's state, *Mater. Today*, 2018, **21**, 720–748.

49 D. Klemm, D. Schumann, U. Udhhardt and S. Marsch, Bacterial synthesized cellulose—artificial blood vessels for microsurgery, *Prog. Polym. Sci.*, 2001, **26**, 1561–1603.

50 M. Zaborowska, A. Bodin, H. Bäckdahl, J. Popp, A. Goldstein and P. Gatenholm, Microporous bacterial cellulose as a potential scaffold for bone regeneration, *Acta Biomater.*, 2010, **6**, 2540–2547.

51 S. Bottan, F. Robotti, P. Jayathissa, A. Hegglin, N. Bahamonde, J. A. Heredia-Guerrero, I. S. Bayer, A. Scarpellini, H. Merker and N. Lindenblatt, Surface-structured bacterial cellulose with guided assembly-based biolithography (GAB), *ACS Nano*, 2015, **9**, 206–219.

52 A. F. Jozala, L. C. de Lencastre-Novaes, A. M. Lopes, V. de Carvalho Santos-Ebinuma, P. G. Mazzola, A. Pessoa Jr, D. Grotto, M. Gerenucci and M. V. Chaud, Bacterial nanocellulose production and application: a 10-year overview, *Appl. Microbiol. Biotechnol.*, 2016, **100**, 2063–2072.

53 L. M. Sampaio, J. Padrão, J. Faria, J. P. Silva, C. J. Silva, F. Dourado and A. Zille, Laccase immobilization on bacterial nanocellulose membranes: Antimicrobial, kinetic and stability properties, *Carbohydr. Polym.*, 2016, **145**, 1–12.



54 A. C. Khorasani and S. A. Shojaosadati, Bacterial nanocellulose-pectin bionanocomposites as prebiotics against drying and gastrointestinal condition, *Int. J. Biol. Macromol.*, 2016, **83**, 9–18.

55 M. E. Badawy and E. I. Rabea, A biopolymer chitosan and its derivatives as promising antimicrobial agents against plant pathogens and their applications in crop protection, *Int. J. Carbohydr. Chem.*, 2011, **2011**, 460381.

56 J. Liang, H. Yan, P. Puligundla, X. Gao, Y. Zhou and X. Wan, Applications of chitosan nanoparticles to enhance absorption and bioavailability of tea polyphenols: A review, *Food Hydrocolloids*, 2017, **69**, 286–292.

57 F. Ahmed, F. M. Soliman, M. A. Adly, H. A. Soliman, M. El-Matbouli and M. Saleh, Recent Progress in Biomedical Applications of Chitosan and its Nanocomposites in Aquaculture: A Review, *Res. Vet. Sci.*, 2019, **126**, 68–82.

58 M. A. Mohammed, J. Syeda, K. M. Wasan and E. K. Wasan, An overview of chitosan nanoparticles and its application in non-parenteral drug delivery, *Pharmaceutics*, 2017, **9**, 53.

59 L.-F. Călinou, B. E. Ștefănescu, I. D. Pop, L. Muntean and D. C. Vodnar, Chitosan coating applications in probiotic microencapsulation, *Coatings*, 2019, **9**, 194.

60 L.-M. Zhao, L.-E. Shi, Z.-L. Zhang, J.-M. Chen, D.-D. Shi, J. Yang and Z.-X. Tang, Preparation and application of chitosan nanoparticles and nanofibers, *Braz. J. Chem. Eng.*, 2011, **28**, 353–362.

61 W. Krasaekoopt, B. Bhandari and H. C. Deeth, Survival of probiotics encapsulated in chitosan-coated alginate beads in yoghurt from UHT-and conventionally treated milk during storage, *LWT-Food Sci. Technol.*, 2006, **39**, 177–183.

62 I. Bravo-Osuna, C. Vauthier, A. Farabollini, G. F. Palmieri and G. Ponchel, Mucoadhesion mechanism of chitosan and thiolated chitosan-poly (isobutyl cyanoacrylate) core-shell nanoparticles, *Biomaterials*, 2007, **28**, 2233–2243.

63 Y. Kawashima, H. Yamamoto, H. Takeuchi and Y. Kuno, Mucoadhesive DL-lactide/glycolide copolymer nanospheres coated with chitosan to improve oral delivery of elcatonin, *Pharm. Dev. Technol.*, 2000, **5**, 77–85.

64 A. Shajahan, S. Shankar, A. Sathiyaseelan, K. S. Narayan, V. Narayanan, V. Kaviyarasan and S. Ignacimuthu, Comparative studies of chitosan and its nanoparticles for the adsorption efficiency of various dyes, *Int. J. Biol. Macromol.*, 2017, **104**, 1449–1458.

65 N. A. Peppas and Y. Huang, Nanoscale technology of mucoadhesive interactions, *Adv. Drug Delivery Rev.*, 2004, **56**, 1675–1687.

66 S. Raghuvanshi, R. Agarwal, R. Raval and R. K. Gutti, *Chitosan Nanoparticles and Their Applications in Drug Delivery, Hemostasis, and Stem Cell Research, Functional Bionanomaterials*, Springer, 2020, pp. 129–143.

67 F. Assa, H. Jafarizadeh-Malmiri, H. Ajamein, H. Vaghari, N. Anarjan, O. Ahmadi and A. Berenjian, Chitosan magnetic nanoparticles for drug delivery systems, *Crit. Rev. Biotechnol.*, 2017, **37**, 492–509.

68 K. A. Janes, M. P. Fresneau, A. Marazuela, A. Fabra and M. A. J. Alonso, Chitosan nanoparticles as delivery systems for doxorubicin, *J. Controlled Release*, 2001, **73**, 255–267.

69 B. Shi, Z. Shen, H. Zhang, J. Bi and S. Dai, Exploring N-imidazolyl-O-carboxymethyl chitosan for high performance gene delivery, *Biomacromolecules*, 2012, **13**, 146–153.

70 A. Mawad, Y. A. Helmy, A.-G. Shalkami, D. Kathayat, G. Rajashekara and E. coli, Nissle microencapsulation in alginate-chitosan nanoparticles and its effect on *Campylobacter jejuni* in vitro, *Appl. Microbiol. Biotechnol.*, 2018, **102**, 10675–10690.

71 R. C. Rowe, P. Sheskey and M. Quinn, *Handbook of Pharmaceutical Excipients*, Libros Digitales-Pharmaceutical Press, 2009.

72 D. Jain, D. Majumdar and A. Panda, Insulin loaded eudragit L100 microspheres for oral delivery: preliminary *in vitro* studies, *J. Biomater. Appl.*, 2006, **21**, 195–211.

73 L. A. Soares and E. Crema, Study of a delayed-release system for hard and soft capsules coated with eudragit® s100 acrylic polymers, *Acta Sci., Health Sci.*, 2020, **42**, e48422.

74 D. Jain, A. K. Panda and D. K. Majumdar, Eudragit S100 entrapped insulin microspheres for oral delivery, *AAPS PharmSciTech*, 2005, **6**, E100–E107.

75 F. Ansari, H. Pourjafar, V. Jodat, J. Sahebi and A. Ataei, Effect of Eudragit S100 nanoparticles and alginate chitosan encapsulation on the viability of *Lactobacillus acidophilus* and *Lactobacillus rhamnosus*, *AMB Express*, 2017, **7**, 144.

76 Z. Camtakan, S. Erenturk and S. Yusun, Magnesium oxide nanoparticles: preparation, characterization, and uranium sorption properties, *Environ. Prog. Sustainable Energy*, 2012, **31**, 536–543.

77 S. Ge, G. Wang, Y. Shen, Q. Zhang, D. Jia, H. Wang, Q. Dong and T. Yin, Cytotoxic effects of MgO nanoparticles on human umbilical vein endothelial cells in vitro, *IET Nanobiotechnol.*, 2011, **5**, 36–40.

78 S. Shukla, G. Parashar, A. Mishra, P. Misra, B. Yadav, R. Shukla, L. Bali and G. Dubey, Nano-like magnesium oxide films and its significance in optical fiber humidity sensor, *Sens. Actuators, B*, 2004, **98**, 5–11.

79 K. Imada, S. Sakai, H. Kajihara, S. Tanaka and S. Ito, Magnesium oxide nanoparticles induce systemic resistance in tomato against bacterial wilt disease, *Plant Pathol.*, 2016, **65**, 551–560.

80 M. Yao, B. Li, H. Ye, W. Huang, Q. Luo, H. Xiao, D. J. McClements and L. Li, Enhanced viability of probiotics (*Pediococcus pentosaceus* Li05) by encapsulation in microgels doped with inorganic nanoparticles, *Food Hydrocolloids*, 2018, **83**, 246–252.

81 J. Kaur, G. Kaur, S. Sharma and K. Jeet, Cereal starch nanoparticles—A prospective food additive: A review, *Crit. Rev. Food Sci. Nutr.*, 2018, **58**, 1097–1107.

82 Á. L. Santana and M. A. A. Meireles, New starches are the trend for industry applications: a review, *Food Publ. Health*, 2014, **4**, 229–241.



83 A. Blennow, T. H. Nielsen, L. Baunsgaard, R. Mikkelsen and S. B. Engelsen, Starch phosphorylation: a new front line in starch research, *Trends Plant Sci.*, 2002, **7**, 445–450.

84 F. Zhu, H. Corke and E. Bertoft, Amylopectin internal molecular structure in relation to physical properties of sweetpotato starch, *Carbohydr. Polym.*, 2011, **84**, 907–918.

85 H.-Y. Kim, S. S. Park and S.-T. Lim, Preparation, characterization and utilization of starch nanoparticles, *Colloids Surf., B*, 2015, **126**, 607–620.

86 M. A. OdeniyiA, O. A. OmotesoB, A. O. AdepojuB and K. T. JaiyeobaE, Starch nanoparticles in drug delivery: A review, *Polim. Med.*, 2018, **48**, 41–45.

87 P. H. Campelo, A. S. Sant'Ana and M. T. P. S. Clerici, Starch nanoparticles: production methods, structure, and properties for food applications, *Current Opinion in Food Science*, 2020, 136–140.

88 A. Dufresne, Processing of polymer nanocomposites reinforced with polysaccharide nanocrystals, *Molecules*, 2010, **15**, 4111–4128.

89 D. Le Corre, J. Bras and A. Dufresne, Starch nanoparticles: a review, *Biomacromolecules*, 2010, **11**, 1139–1153.

90 M. Ahmad, A. Gani, F. Hamed and S. Maqsood, Comparative study on utilization of micro and nano sized starch particles for encapsulation of camel milk derived probiotics (*Pediococcus acidolactici*), *LWT-Food Sci. Technol.*, 2019, **110**, 231–238.

91 S. Kumari, B. S. Yadav and R. B. Yadav, Synthesis and modification approaches for starch nanoparticles for their emerging food industrial applications: A review, *Food Res. Int.*, 2020, **128**, 108765.

92 S. Thangrongthong, N. Puttarat, B. Ladda, T. Itthisoponkul, W. Pinket, K. Kasemwong and M. Taweechotipatr, Microencapsulation of probiotic *Lactobacillus brevis* ST-69 producing GABA using alginate supplemented with nanocrystalline starch, *Food Sci. Biotechnol.*, 2020, **29**, 1475–1482.

93 G. Ozkan, P. Franco, I. De Marco, J. Xiao and E. Capanoglu, A review of microencapsulation methods for food antioxidants: Principles, advantages, drawbacks and applications, *Food Chem.*, 2019, **272**, 494–506.

94 K. S. Prakash, R. Chavan and V. Mishra, *Microencapsulation of Probiotics and its Applications, Frontier Discoveries and Innovations in Interdisciplinary Microbiology*, Springer, 2016, pp. 33–44.

95 A. Šipailienė and S. Petraitytė, Encapsulation of probiotics: proper selection of the probiotic strain and the influence of encapsulation technology and materials on the viability of encapsulated microorganisms, *Probiotics Antimicrob. Proteins*, 2018, **10**, 1–10.

96 S. Benita, *Microencapsulation: Methods and Industrial Applications*, Crc Press, 2005.

97 A. Mortazavian, S. H. Razavi, M. R. Ehsani and S. Sohrabvandi, *Principles and Methods of Microencapsulation of Probiotic Microorganisms*, 2007.

98 K. Feng, M.-Y. Zhai, Y. Zhang, R. J. Linhardt, M.-H. Zong, L. Li and H. Wu, Improved viability and thermal stability of the probiotics encapsulated in a novel electrospun fiber mat, *J. Agric. Food Chem.*, 2018, **66**, 10890–10897.

99 A. C. Anselmo, K. J. McHugh, J. Webster, R. Langer and A. Jaklenec, Layer-by-layer encapsulation of probiotics for delivery to the microbiome, *Adv. Mater.*, 2016, **28**, 9486–9490.

100 J. A. Silva, P. R. De Gregorio, G. Rivero, G. A. Abraham and M. E. F. Nader-Macías, Immobilization of vaginal *Lactobacillus* in polymeric nanofibers for its incorporation in vaginal probiotic products, *Eur. J. Pharm. Sci.*, 2021, **156**, 105563.

101 J. Ma, C. Xu, H. Yu, Z. Feng, W. Yu, L. Gu, Z. Liu, L. Chen, Z. Jiang and J. Hou, Electro-encapsulation of probiotics in gum Arabic-pullulan blend nanofibres using electrospinning technology, *Food Hydrocolloids*, 2021, **111**, 106381.

102 M. T. Yilmaz, O. Taylan, C. Y. Karakas and E. Dertli, An alternative way to encapsulate probiotics within electrospun alginate nanofibers as monitored under simulated gastrointestinal conditions and in kefir, *Carbohydr. Polym.*, 2020, **244**, 116447.

103 O. O. Adebola, O. Corcoran and W. A. Morgan, Synbiotics: the impact of potential prebiotics inulin, lactulose and lactobionic acid on the survival and growth of lactobacilli probiotics, *J. Funct. Foods*, 2014, **10**, 75–84.

104 T. Liu, Y. Wang, W. Zhong, B. Li, K. Mequanint, G. Luo and M. Xing, Biomedical Applications of Layer-by-Layer Self-Assembly for Cell Encapsulation: Current Status and Future Perspectives, *Adv. Healthcare Mater.*, 2019, **8**, 1800939.

105 A. J. Priya, S. Vijayalakshmi and A. M. Raichur, Enhanced survival of probiotic *Lactobacillus acidophilus* by encapsulation with nanostructured polyelectrolyte layers through layer-by-layer approach, *J. Agric. Food Chem.*, 2011, **59**, 11838–11845.

106 M. Wang, J. Yang, M. Li, Y. Wang, H. Wu, L. Xiong and Q. Sun, Enhanced viability of layer-by-layer encapsulated *Lactobacillus pentosus* using chitosan and sodium phytate, *Food Chem.*, 2019, **285**, 260–265.

