A multifunctional Fenton nanoagent for microenvironment-selective anti-biofilm and anti-inflammatory therapy†

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Bacterial biofilm infections are intractable to traditional antibiotic treatment and usually cause persistent inflammation. Chemodynamic therapy (CDT) based on the Fenton reaction has recently emerged as a promising anti-biofilm strategy. However, the therapeutic efficacy of current Fenton agents often suffers from inefficient Fenton activity and lacks anti-inflammatory capability. Herein, FePS3 nanosheets (NSs) are explored for the first time as novel microenvironment-selective therapeutic nanoagents for bacterial biofilm infections with both self-enhanced Fenton activity for an anti-biofilm effect and reactive oxygen species (ROS) scavenging properties for an anti-inflammatory effect. In biofilms with acidic microenvironments, FePS3 NSs release Fe3+ to generate toxic ROS by Fenton reaction and reductive [P2S6]4− to enhance the Fenton activity by reducing Fe3+ to Fe2+. In the surrounding normal tissues with neutral pH, FePS3 NSs scavenge ROS by reductive [P2S6]4− with an anti-inflammatory effect. This work demonstrates multifunctional Fenton nanoagents with microenvironment-selective ROS generation and elimination properties for effective treatment of bacterial biofilm infections with both anti-biofilm and anti-inflammatory effects.

Bacterial infections have emerged as ever-growing threats to human health, and many recalcitrant bacterial infections have been proven to correlate with the formation of biofilms, such as lung infections in cystic fibrosis, implant-related infections, chronic wounds, dental caries, and so on.1–3 A bacterial biofilm is an aggregate of bacterial populations encapsulated in the extracellular polymeric substance (EPS) matrix, which can protect bacteria from the attack of host immune defenses and cause high resistance to antibiotics.4–6 Furthermore, persistent inflammation is often caused by bacterial biofilm infection, which can damage the surrounding normal tissues and hamper the recovery of infected tissues.7–9 To date, the effective treatment of bacterial biofilm infections is still highly desired, especially a method with both anti-biofilm and anti-inflammatory effects.

CDT based on Fenton/Fenton-like reactions has recently been considered a promising therapeutic strategy for the treatment of cancer and bacterial infections.10–14 Fenton agents catalytically convert hydrogen peroxide (H2O2) to highly oxidative hydroxyl radicals (•OH), which irreversibly damage various
biomolecules (proteins, lipids, DNA, etc.) and effectively kill drug-resistant cancer cells or bacteria. Various Fenton agents have been used to kill bacteria both in planktonic form and in biofilms. However, the anti-biofilm effect of Fenton agents is limited by the inefficient iron redox cycling and *OH production, because the reduction of the as-formed Fe$^{3+}$ to Fe$^{2+}$ by H$_2$O$_2$ is much slower than the oxidation of Fe$^{2+}$. Although chelating agents or reductive agents can improve the iron redox cycling during Fenton reaction, they are complicated and unstable for biological use. On the other hand, common Fenton agents lack the anti-inflammatory capability to scavenge excessive ROS accumulated in the normal tissues around biofilms. Hence, Fenton agents with enhanced iron redox cycling and ROS scavenging activity are highly desired to treat bacterial biofilm infections.

In bacterial biofilm infected tissues, interactions between the bacteria and the host result in unique biofilm microenvironments. The respiratory burst of abundant phagocytes around bacterial biofilms results in high levels of ROS (*e.g., H$_2$O$_2$) in biofilm infected tissues. Besides, bacteria in oxygen-limited regions of the biofilm conduct anaerobic fermentation along with the accumulation of organic acids, which causes a more acidic microenvironment in biofilm infected tissues compared with neutral normal tissues. By utilizing the biofilm microenvironment as a stimulus, responsive nanoagents have shown selective anti-biofilm outcomes with diminished side effects to normal tissues. Inspired by these reports, we expect that FePS$_3$ NSs have potential as microenvironment-selective therapeutic agents for biofilm infections based on the existence of both Fenton-active Fe cations (Fe$^{2+}$/Fe$^{3+}$) for bacterial killing and reductive anions ([P$_2$S$_6$]$^{4-}$) for ROS scavenging.

FePS$_3$ NSs were prepared from bulk FePS$_3$ by ball-milling and subsequent ultrasonic exfoliation (Scheme 1). FePS$_3$ NSs exhibit acid-responsive dissociation with the release of Fe$^{2+}$ and [P$_2$S$_6$]$^{4-}$ while being relatively stable under neutral conditions. In bacterial biofilm infected tissues with acidic pH, the released Fe$^{2+}$ further converts H$_2$O$_2$ to *OH through Fenton reaction, and the [P$_2$S$_6$]$^{4-}$ can reduce the as-formed Fe$^{3+}$ to Fe$^{2+}$ with accelerated iron redox cycling and enhance the Fenton activity. In contrast, in normal tissues with neutral pH, FePS$_3$ NSs show antioxidative activity and can effectively scavenge H$_2$O$_2$ or *OH by the reductive [P$_2$S$_6$]$^{4-}$ through redox reaction. Experimental results show that the microenvironment-selective ROS regulation properties of FePS$_3$ NSs enable simultaneous anti-biofilm and anti-inflammatory therapy for bacterial biofilm infections.

Bulk FePS$_3$ was synthesized by high temperature solid state reaction using Fe, P, and S powders. The scanning electron microscopy (SEM) image (Fig. S1a, ESIF) shows clear layered morphology of bulk FePS$_3$. The X-ray diffraction (XRD) pattern (Fig. S1b, ESIF) further confirms the successful synthesis of bulk FePS$_3$. FePS$_3$ NSs were obtained by ball-milling and subsequent liquid ultrasonication of the bulk FePS$_3$ with the assistance of poly(vinyl pyrrolidone) (PVP). The transmission electron microscopy (TEM) image (Fig. 1a) and the statistical analysis (Fig. S2a, ESIF) illustrate that FePS$_3$ NSs have uniform morphology with an average size of ~15 nm. Dynamic light scattering (DLS) analysis shows that the hydrodynamic diameter of FePS$_3$ NSs is around 24 nm (Fig. S2b, ESIF). As the high-resolution TEM (HRTEM) image shows in Fig. 1b, the crystal lattice space of 1.86 Å can be ascribed to the (310) plane of FePS$_3$. High-angle annular dark field scanning TEM (HAADF-STEM) image and energy dispersive spectroscopy (EDS) elemental mapping images (Fig. 1c) show the homogeneous element distribution of Fe, P, and S in FePS$_3$ NSs. Atomic force microscopy (AFM) measurement (Fig. 1d and Fig. S2c, ESIF) indicates the average thickness of FePS$_3$ NSs to be ~2.1 nm, suggesting that FePS$_3$ NSs have 2-3 single layers. Besides, FePS$_3$ NSs have negative charges on the surface with a zeta potential of about ~10 mV (Fig. S2d, ESIF).

![Scheme 1](attachment:image1.png)

**Scheme 1** (a) Preparation of FePS$_3$ NSs by ball-milling and ultrasonication method. (b) FePS$_3$ NSs exhibit both self-enhanced Fenton activity in biofilm infected tissues with acidic pH for the eradication of biofilms and ROS scavenging properties in normal tissues with neutral pH for the remission of inflammation.
As the XRD pattern shows in Fig. 1e, the diffraction peaks of FePS3 NSs located at 13.8° and 27.8° can be ascribed to (001) and (002) planes of FePS3, respectively. Fig. S3a (ESI†) shows that FePS3 layers are formed by \([\text{P}_2\text{S}_6]^{4-}\) units and Fe ions, and then weakly bond with each other through van der Waals forces.\(^4\) The Raman spectrum of FePS3 NSs (Fig. S3b, ESI†) exhibits a peak at 385 cm\(^{-1}\) (\(A_2(2)\)), which represents P–P bond stretching and the symmetric stretching vibration of the P–S bond in the \([\text{P}_2\text{S}_6]^{4-}\) unit. Raman peaks at 215 cm\(^{-1}\) (\(E_g(1)\)), 275 cm\(^{-1}\) (\(E_g(2)\)), and 588 cm\(^{-1}\) (\(E_g(3)\)) originate from the tangential vibration of the P–P bond, which suggests the inplane vibrations of the \([\text{P}_2\text{S}_6]^{4-}\) unit.\(^4\) Fourier transform infrared (FT-IR) spectra in Fig. S3c (ESI†) show the asymmetric stretching vibration of the P–S bond (572 cm\(^{-1}\)) and the P–P vibration (442 cm\(^{-1}\)) of FePS3.\(^5\)\(^6\)\(^1\)\(^4\) IR bands at 1642 cm\(^{-1}\) and 1428 cm\(^{-1}\) belong to the stretching vibration of amide C=O and the C–H bending vibration, respectively, suggesting the presence of PVP on FePS3 NSs.

FePS3 NSs were further characterized by using X-ray photoelectron spectroscopy (XPS). XPS survey spectra show that FePS3 NSs and bulk FePS3 have similar binding energy peaks of the main elements, while the N 1s peak for FePS3 NSs can be ascribed to the surface adsorbed PVP (Fig. 1f). As the high-resolution XPS spectra of Fe 2p show in Fig. S4a (ESI†), the binding energies of 2p\(3/2\) and 2p\(1/2\) orbitals of Fe(II) are 709.3 eV and 722.9 eV, respectively. And the peaks near 712.0 eV and 725.6 eV belong to the 2p\(3/2\) and 2p\(1/2\) orbitals of Fe(III), indicating the coexistence of Fe(II) and Fe(III) on the surface of FePS3 NSs. Moreover, the generation of Fe(II) in FePS3 NSs indicates their potential Fenton activity, whereas the \([\text{P}_2\text{S}_6]^{4-}\) anions in FePS3 may influence their dissociation at different pH conditions.\(^4\)^\(^9\)^\(^5\)\(^3\) Besides, no obvious difference of S 2p binding energy peaks can be observed between FePS3 NSs and bulk FePS3 (Fig. S4c, ESI†). Moreover, the peaks at 188.7 eV and 225.6 eV represent the P 2s and S 2s orbitals of FePS3, respectively; whereas the C 1s peak (284.6 eV) and O 1s peak (531.6 eV) may originate from the test environment (Fig. 1f).

The existence of Fe(II) in FePS3 NSs indicates their potential Fenton activity, whereas the \([\text{P}_2\text{S}_6]^{4-}\) may possess reducibility due to the low valence states of P and S (+4 and −2, respectively). First of all, 3,3',5,5'-tetramethylbenzidine (TMB) was used as the substrate to carry on the colorimetric assay of Fenton activity.\(^2\)\(^4\) As shown in Fig. 2a, the characteristic absorbance at 652 nm increases significantly with the decrease of pH, while no obvious change is observed at pH 7.4, indicating that FePS3 NSs exhibit Fenton activity only under acidic conditions. Moreover, the generation of *OH in these reactions was further detected by electron paramagnetic resonance (EPR) by using 5,5-dimethyl-1-pyridine N-oxide (DMPO) as the spin trap.\(^5\)\(^5\) Fig. 2b shows obvious DMPO/OH signals for FePS3 NSs + H\(_2\)O\(_2\) mixture at pH 5.0, while no characteristic signal appears at pH 7.4, which further verifies the acid-responsive Fenton activity of FePS3 NSs.

The time-dependent absorbance of FePS3 NSs (at 375 nm) was monitored under different pH conditions, and the corresponding dissolution percentage was calculated based on the decrease of absorbance over time (Fig. S5 and S6, ESI†). As shown in Fig. 2c, FePS3 NSs show little dissolution under neutral conditions, while they can completely dissolve under acidic conditions. As previously reported, FePS3 is formed based on the ionic bonds between Fe cations and \([\text{P}_2\text{S}_6]^{4-}\), which may influence their dissociation at different pH conditions.\(^4\)^\(^9\)^\(^5\)\(^3\) Besides, XPS analysis (Fig. S7, ESI†) demonstrates no obvious change on the valence states of the elements in FePS3 NSs whether dispersed in aqueous solutions with different pH conditions (5.0 or 7.4), suggesting that the decreased absorbance of FePS3 NSs is related to the dissociation of FePS3 rather than the oxidative decomposition by the dissolved oxygen. These results reveal that FePS3 NSs may dissolve and release Fe\(^{2+}\) cations and \([\text{P}_2\text{S}_6]^{4-}\) anions in response to the acidic environment while keeping stable under neutral conditions.

The acid-responsive Fe\(^{2+}\) release can account for the acid-responsive Fenton activity of FePS3 NSs (eqn (1) and (2)), while the \([\text{P}_2\text{S}_6]^{4-}\) also exerts an essential function in Fenton reaction.
Under neutral condition, FePS₃ NSs have negligible Fenton activity due to the limited release of Fe²⁺ and acid-dependent Fenton activity, while the reductive [P₂S₆]₄⁻ potentially endows FePS₃ NSs with ROS scavenging ability. Thus, 2-phenyl-4,4,5,5-tetramethylimidazolinium-3-oxo-1-oxo (PTIO) radicals (PTIO*) and 2,2′-biazinobis(3-ethylbenzothiazoline-6-sulfonic acid) (ABTS) radicals (ABTS*) were used to test the antioxidative activity of FePS₃ NSs.⁶⁶ As Fig. 2e shows, the PTIO* scavenging capability of FePS₃ NSs is about 3.2 times as high as that of a typical antioxidant (6-hydroxy-2,5,7,8-tetramethychroman-2-carboxylic acid, Trolox). Moreover, calcein acetoxymethyl ester (calcein-AM) was used to stain live bacteria in biofilms. As shown in Fig. 3a and Fig. S14b, ESIF, indicating the excellent antioxidative activity of FePS₃ NSs (Fig. S13c and d, ESIF). The ROS level in biofilms was measured using 2′,7′-dichlorodihydrofluorescein diacetate (DCFH-DA).⁶⁴,⁶⁶,⁶⁷ As shown in the 3D confocal laser scanning microscopy (CLSM) images, the fluorescence intensity of the S. aureus biofilms treated by FePS₃ NSs + H₂O₂ is much higher than those treated by H₂O₂ or FePS₃ NSs (Fig. 3a and Fig. S14a, ESIF), indicating the generation of *OH caused by Fenton-active FePS₃ NSs. Moreover, calcium acetoxyethyl ester (calcine-AM) was used to stain live bacteria in S. aureus biofilms. Among them, biofilms treated by FePS₃ NSs + H₂O₂ show the lowest fluorescence intensity (Fig. 3a and Fig. S14b, ESIF), demonstrating their excellent bacterial killing effect in biofilms. As shown in Fig. 3b, the bacterial inactivation efficiency of H₂O₂ (100 µM) alone is only 0.13 log (−25.89%), while that of FePS₃ NSs with lower concentration (27 µM, 5 µg mL⁻¹) is 0.28 log (−47.64%), indicating the better anti-biofilm effect of FePS₃ NSs compared with H₂O₂. Besides, the colony-forming units (CFU) number of bacteria in biofilms treated by FePS₃ NSs (50 µg mL⁻¹) with H₂O₂ (100 µM) decreased by about 4 log (−99.99%) and the CFU number of the group treated by only FePS₃ NSs (50 µg mL⁻¹) decreased by less than 2 log (−98.62%), which revealed the excellent sterilization of *OH generated via FePS₃ NSs through Fenton reaction. In addition, crystal violet staining was conducted after different treatments to evaluate the total biofilm biomass including bacteria and EPS. Compared with the sole treatment of H₂O₂ or FePS₃ NSs, the S. aureus biofilms treated by H₂O₂ + FePS₃ NSs show much less biomass (Fig. S15, ESIF). As the quantitative analysis shows in Fig. 3c, the relative biofilm biomass is reduced to about 24% after the treatment of FePS₃ NSs (50 µg mL⁻¹) with the presence of H₂O₂ (100 µM), much lower than that of H₂O₂ (−95%) or FePS₃ NSs (−50%). The morphology of the biofilms after different treatments was studied by SEM images (Fig. 3d). S. aureus inside biofilms without treatment presents sphere-like morphology with smooth and intact cell walls, while the bacterial surface becomes distorted and wrinkled in the FePS₃ NSs + H₂O₂ group. These results demonstrate that FePS₃ NSs have excellent *OH scavenging properties (Fig. S12, ESIF). Hence, FePS₃ NSs possess antioxidative activity to effectively scavenge H₂O₂ and *OH in neutral conditions.

The acid-responsive self-enhanced Fenton activity of FePS₃ NSs was further examined for the treatment of both planktonic bacteria and bacterial biofilms. The bacterial growth curves of Staphylococcus aureus (S. aureus) treated with FePS₃ NSs show a significant bacteriostatic effect either without or with H₂O₂ (Fig. S13a and b, ESIF). Also, the bacterial growth inhibition effect of FePS₃ NSs + H₂O₂ is much higher than that of FePS₃ NSs, indicating enhanced antibacterial activity of FePS₃ NSs via Fenton reaction. On the other hand, the inactivation efficiency of FePS₃ NSs (50 µg mL⁻¹) against planktonic S. aureus reached 4.2 log (−99.99%) and 6.2 log (−99.999%) without and with H₂O₂ (100 µM), respectively, which reveals the good antibacterial activity of FePS₃ NSs (Fig. S13c and d, ESIF). Moreover, S. aureus biofilms were further challenged by FePS₃ NSs to investigate the in vitro anti-biofilm effect. Extra H₂O₂ (100 µM) was introduced to mimic the in vivo biofilm microenvironment.⁶⁴,⁶⁵ The ROS level in biofilms was measured using 2′,7′-dichlorodihydrofluorescein diacetate (DCFH-DA).⁶⁴,⁶⁶,⁶⁷ As shown in Fig. 4c, the relative antibacterial activity of FePS₃ NSs (50 µg mL⁻¹) with H₂O₂ (100 µM) increased by about 4 log (−99.99%) and the CFU number of the group treated by only FePS₃ NSs (50 µg mL⁻¹) decreased by less than 2 log (−98.62%), which revealed the excellent sterilization of *OH generated via FePS₃ NSs through Fenton reaction. In addition, crystal violet staining was conducted after different treatments to evaluate the total biofilm biomass including bacteria and EPS. Compared with the sole treatment of H₂O₂ or FePS₃ NSs, the S. aureus biofilms treated by H₂O₂ + FePS₃ NSs show much less biomass (Fig. S15, ESIF). As the quantitative analysis shows in Fig. 4c, the relative biofilm biomass is reduced to about 24% after the treatment of FePS₃ NSs (50 µg mL⁻¹) with the presence of H₂O₂ (100 µM), much lower than that of H₂O₂ (−95%) or FePS₃ NSs (−50%). The morphology of the biofilms after different treatments was studied by SEM images (Fig. 4d). S. aureus inside biofilms without treatment presents sphere-like morphology with smooth and intact cell walls, while the bacterial surface becomes distorted and wrinkled in the FePS₃ NSs + H₂O₂ group. These results demonstrate that FePS₃ NSs have excellent...
anti-biofilm effect in response to the acid and H$_2$O$_2$ in the biofilm microenvironment by self-enhanced Fenton activity.

The cytotoxicity of FePS$_3$ NSs was first studied using murine fibroblast (NIH-3T3) cells. After incubation with FePS$_3$ NSs at the concentration up to 80 $\mu$g mL$^{-1}$, no obvious decrease in the viability of NIH-3T3 cells can be observed (Fig. S16a, ESI†). Besides, FePS$_3$ NSs exhibit low hemolysis effect, indicating their good blood compatibility (Fig. S16b, ESI†). Furthermore, the in vitro toxicity of FePS$_3$ NSs was assessed in healthy mice by tail vein injection (~1 mg kg$^{-1}$). The body weights of the mice injected with FePS$_3$ NSs have no noticeable difference from those injected with saline (0.85% NaCl) (Fig. S16c, ESI†). Hematoxylin and eosin (H&E) staining images show that the major organs from these mice have no observable damage post-injection for 21 days (Fig. S16d, ESI†). Therefore, FePS$_3$ NSs have negligible toxicity to mice at the dose used in this work.

To evaluate the in vitro ROS scavenging properties of FePS$_3$ NSs, NIH-3T3 cells were pretreated with Rosup to induce intracellular ROS generation, and then treated with FePS$_3$ NSs$^{59,68}$ The fluorescence intensity of NIH-3T3 cells stained with DCFH-DA decreases significantly upon the addition of FePS$_3$ NSs (Fig. 4a and Fig. S18, ESI†), suggesting the lowered intracellular ROS level.$^{69}$ The cell viability of NIH-3T3 cells treated by Rosup decreased to 55% due to the oxidative stress (Fig. 4b). In comparison, the viability of NIH-3T3 cells stimulated by Rosup and followed by FePS$_3$ NS treatment remained up to 95%, which indicates the cell protection effect of FePS$_3$ NSs from excessive ROS. Besides, when H$_2$O$_2$ was used to induce oxidative damage to NIH-3T3 cells, the pretreatment by FePS$_3$ NSs can also efficiently protect them from oxidative stress (Fig. 4c). These results suggest that FePS$_3$ NSs are an effective ROS scavenger for the relief of oxidative stress. On the other hand, since ROS can act as a secondary cellular messenger for inflammatory cytokine signalling, the anti-inflammatory effect of FePS$_3$ NSs by ROS scavenging was further studied.$^{70,71}$ Lipoteichoic acid (LTA) was employed to stimulate murine macrophage (RAW264.7) cells to induce an in vitro inflammatory response. After the treatment, tumour necrosis factor-$\alpha$ (TNF-$\alpha$) and interleukin-6 (IL-6) were tested as inflammation markers. Pretreatment with FePS$_3$ NSs significantly reduced both the TNF-$\alpha$ level and IL-6 level (Fig. 4d and e), suggesting reduced inflammation. Hence, FePS$_3$ NSs may mitigate the oxidative stress and inflammation in normal tissues by scavenging excessive ROS.

Considering the difference of pH conditions between biofilm infected tissues and normal tissues, FePS$_3$ NSs are expected to have microenvironment-selective anti-biofilm and anti-inflammatory effects. The in vivo therapeutic efficacy of FePS$_3$ NSs was studied on S. aureus biofilm infected mice models (Fig. 5a). One day after infection, FePS$_3$ NSs (~0.5 mg kg$^{-1}$) and saline (control) were in situ injected into the abscesses of infected mice, respectively.
indicates that FePS3 NSs can effectively facilitate the recovery of the abscesses and wound beds of mice almost disappear, which treated by saline. At the 16th-day post-treatment with FePS3 NSs, observed in the infected tissues of the saline group, while that effect of FePS3 NSs. Moreover, neutrophils in the infected tissues characterized by immunofluorescence staining of MPO (MPO: green; nuclei: blue). The scale bar is 200 μm. (f) Immunofluorescence staining images of TNF-α and IL-6 in the infected tissues (nuclei: blue; TNF-α: red; IL-6: green). The scale bar is 50 μm. (g) H&E and (h) Masson’s trichrome staining images of the infected tissues. The scale bar is 500 μm.

As illustrated in Fig. 5b and c, the infected areas of the mice treated with FePS3 NSs are much smaller than that of the mice treated by saline. At the 16th-day post-treatment with FePS3 NSs, the abscesses and wound beds of mice almost disappear, which indicates that FePS3 NSs can effectively facilitate the recovery of S. aureus biofilm infected tissues. As shown in Fig. 5d, the number of live S. aureus in the infected tissues was quantified, and the bacterial inactivation efficiency of FePS3 NSs is about 3.7 log (99.98%), suggesting an excellent in vivo anti-biofilm effect of FePS3 NSs. Moreover, neutrophils in the infected tissues were detected by immunofluorescence staining of myeloperoxidase (MPO). Strong fluorescence can be clearly observed in the infected tissues of the saline group, while that of the FePS3 NS group is remarkably reduced, indicating fewer neutrophils infiltration and reduced inflammation (Fig. 5e). Meanwhile, the expressions of typical pro-inflammatory cytokines (TNF-α and IL-6) in the infected tissues treated by FePS3 NSs show a significant reduction, which further confirms the in vivo anti-inflammatory effect of FePS3 NSs (Fig. 5f). The therapeutic efficiency of FePS3 NSs towards S. aureus biofilm infected mice was also evaluated by histological analysis. As the H&E staining images (Fig. 5g) show, the infiltration of plenty of inflammatory cells indicates that the intensive inflammation still remained in the infected tissues of the saline group. In contrast, fewer inflammatory cells and a more intact epidermis layer can be observed in the FePS3 NS group. Besides, Masson’s trichrome staining images (Fig. 5h) for the FePS3 NS group show better collagen fiber formation in the infected tissues. These results demonstrate that FePS3 NSs exhibit excellent anti-biofilm and anti-inflammatory efficacy in the S. aureus biofilm infected mice.

Conclusions

In summary, FePS3 NSs possess both self-enhanced Fenton activity under acidic conditions and ROS scavenging properties under neutral conditions. As a pH-responsive multifunctional Fenton nanoagent, FePS3 NSs exhibit microenvironment-selective anti-biofilm and anti-inflammatory effects against bacterial biofilm infections. In bacterial biofilms with an acidic microenvironment, FePS3 NSs can release Fe²⁺ and react with H₂O₂ by Fenton reaction to generate Fe³⁺ and highly oxidative •OH for bacteria-killing in biofilms; while the [P₂S₆]⁴⁻ can reduce the as-formed Fe³⁺ to Fe²⁺, which significantly enhances the therapeutic effect of the Fenton reaction by promoting iron redox cycling. In normal tissues with neutral pH, FePS3 NSs exhibit effective ROS scavenging properties via the reductive [P₂S₆]⁴⁻, which can relieve oxidative stress and exert an anti-inflammatory effect. Both in vitro and in vivo experimental results demonstrate that FePS3 NSs can simultaneously eradicate bacterial biofilms and mitigate tissue inflammation, and achieve excellent therapeutic efficacy for bacterial biofilm infections. Besides, FePS3 NSs show good biocompatibility with neglectable toxicity to mice. Overall, this work demonstrates that the integration of Fe cations and reductive anions in one Fenton agent makes FePS3 NSs a kind of novel pH-responsive therapeutic nanoagent with self-enhanced Fenton activity and ROS scavenging properties, facilitating the microenvironment-selective anti-biofilm and anti-inflammatory therapy for bacterial biofilm infections.

Authors’ contributions

L. Wang and L. Yuwen conceived the idea and supervised the study. Y. Li carried out the experiments, analysed the results, and wrote the manuscript. W. Xiu, K. Yang, Q. Wen and Z. Luo helped with the experimental characterization and the results discussion. D. Yang supported the operation of animal...
experiments. X. Liu and X. Xie contributed to the scientific discussion of the manuscript. All authors discussed, commented and agreed on the manuscript.

Conflicts of interest
The authors declare no conflict of interest.

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