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# A novel aggregation-induced emission poly(phosphoramidate): fluorescence properties and visual detection for Cu<sup>2+</sup>†

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Non-aromatic aggregation-induced emission (AIE) polymers have become a hot spot in the materials fields due to their low toxicity and excellent environmental friendliness. However, this kind of polymer usually suffers from insufficient fluorescence properties and limited varieties. Herein, we have synthesized a novel kind of non-aromatic AIE polymer—poly(phosphoramidate) (PPA) for the first time in consideration of the electron-rich properties of P atoms and excellent biocompatibility of phosphate esters. The PPA is synthesized through a green and facile one-pot ester ammonolysis of triethyl phosphate with ethylenediamine (P1–P3) or 1,6-diaminohexane (P4). The synthesized PPA shows typical AIE characteristics and can emit bright blue fluorescence under UV light. We found that the molecular weight, terminal groups and chain length all have remarkable influence on its fluorescence properties. Transmission electron microscope (TEM) images and density functional theory (DFT) calculations reveal that the fluorescence of PPA can be attributed to the formation of a luminogen cluster and the electronic delocalization caused by through-space conjugation. Notably, PPA is sensitive to Cu<sup>2+</sup> and it can change the colour of CuCl<sub>2</sub> solution from colourless to purple. Thus, this novel PPA not only enriches the species of non-aromatic AIE polymers, but also provides a convenient visual detection agent for Cu<sup>2+</sup>.

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## 1. Introduction

Compared with conventional aggregation-induced emission (AIE) luminogens,<sup>1,2</sup> non-aromatic AIE polymers<sup>3,4</sup> are emerging in biological imaging, metal ion detection and other fields<sup>1,4,5</sup> due to their low toxicity and excellent environmental friendliness. However, the species of non-aromatic AIE polymers are mainly limited to N-containing polymers such as poly(amino ester)s,<sup>6–8</sup> poly(amidoamine)s,<sup>9</sup> and polyethylenimines.<sup>10</sup> Recently, our group synthesized a series of Si-containing non-aromatic AIE polymers—hyperbranched polysiloxane (HBPSi)<sup>11–14</sup>—and found that they can emit bright fluorescence and show a strong AIE effect. Similarly, in consideration of the electron-rich properties of P atoms and excellent biocompatibility of P-containing polymers, the poly(phosphoramidate) (PPA) can be expected as a novel non-aromatic AIE polymer; however, there has been insufficient study of this kind of polymer until now.

Phosphorous polymers mainly include polyphosphoesters, polyphosphoric acids and poly(phosphonamidites) (PPA). Among these polymers, PPA shows good degradability, low toxicity and good designability and has been widely used as a flame retardant.<sup>15–18</sup> Up to now, PPAs have mostly been synthesized *via* polycondensation by phosphorus oxychloride and diamine,<sup>19–22</sup> accompanied by hydrogen chloride as a by-product. However, the reactant phosphorus oxychloride is dangerous due to its strong corrosiveness and the removal of hydrogen chloride complicates the synthesis process. Besides, the fluorescence properties of PPA are seldom studied. Such drawbacks greatly limit the applications of PPA. Thus, the development of an innovative strategy to prepare PPA and a systematic study of its fluorescence property is highly desirable.

Herein, we creatively develop a green and facile one-pot strategy to synthesize PPA through an ester ammonolysis reaction for the first time. By using triethyl phosphate (TEP), ethylenediamine (EDM) and 1,6-diaminohexane (DAH) as reactants, the synthesis process is simple, safe and without complicated separation procedures. Most importantly, the synthesized PPA can emit bright blue fluorescence under UV light and show an AIE effect. We systematically investigated the influence of molecular weight, terminal groups and chain

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length on the fluorescence property of PPA. Transmission electron microscopy (TEM) and density functional theory (DFT) were also applied to explore the fluorescence mechanism of PPA. Moreover, the synthesized PPA could change the colour of  $\text{CuCl}_2$  solution from colourless to purple. Thus, it can be used as an on-site analysing chromogenic reagent for  $\text{Cu}^{2+}$  instead of time-consuming and expensive detection methods, such as electroanalytical chemistry,<sup>23</sup> atomic absorption spectrometry<sup>24</sup> or inductively coupled plasma mass spectrometry.<sup>25</sup> This work provides a green, facile and safe approach to synthesize PPA, and develop a new kind of P-containing non-aromatic AIE polymer. The synthesized PPA is sensitive to  $\text{Cu}^{2+}$  and can be used as a chromogenic reagent for the on-site visual detection of  $\text{Cu}^{2+}$ .

## 2. Experimental

### 2.1 Materials

Triethyl phosphate (TEP, AR) was purchased from Shanghai Macklin Biochemical Co., Ltd. Ethylenediamine (EDM, AR) and 1,6-diaminohexane (DAH, AR) were obtained from Tianjin Tianli Chemical Reagents Co., Ltd. Absolute ethanol (AR) was supplied by Guangdong Guanghua Sci-Tech Co., Ltd. Sodium hydroxide was purchased from Tianjin Jinbei Fine Chemical Co., Ltd. Regenerated cellulose (RC) dialysis membranes with a molecular weight cut off (MWOC) of 1000 were purchased from Xi'an Yobios Biotechnology Co., Ltd. The RC dialysis membranes were cut off and boiled in distilled water for 15 minutes. Other materials were used without further purification.

### 2.2 Methods

Nuclear magnetic resonance spectra ( $^1\text{H}$ ,  $^{13}\text{C}$ ,  $^{31}\text{P}$  NMR) were detected on a Bruker Avance 400 MHz superconducting Fourier digital NMR spectrometer using dimethyl sulfoxide ( $\text{DMSO}-d_6$ ) as the solvent. Fourier Transform infrared spectroscopy (FT-IR) was recorded on a Bruker infrared spectrometer with a wavenumber ranging from 4000 to  $400\text{ cm}^{-1}$ . Ultraviolet-visible (UV-vis) absorption spectroscopy was measured using a HITACHI U-3900H UV-visible absorption spectrometer. Fluorescent excitation/emission spectra of PPA solution were measured on a F-30 fluorescence spectrophotometer with a scanning speed of  $240\text{ nm min}^{-1}$ . Fluorescent excitation/emission spectra, fluorescence lifetime and absolute quantum yield (QY) of the pure PPA were measured on a UK Edinburgh FLS980 full-featured steady-state/transient fluorescence spectrometer at room temperature. A 340W xenon lamp was used as the light source. Gel permeation chromatography (GPC) was used to measure the molecular weight and distribution of the PPA, with 0.1M  $\text{NaNO}_3$  aqueous solution as eluent with a flow rate of  $1\text{ mL min}^{-1}$  at room temperature. The density functional theory (DFT) method at the B3LYP/6-31 G (d) level was used to calculate the energy level of the PPA. Transmission electron microscopy (TEM) was applied to observe the morphology of PPA clusters on a FEI Tecnai G2 F20 microscope. The

measurement was performed at 200 kV and room temperature. The cell biocompatibility of PPA was assessed by CCK-8 assays.

### 2.3 Synthesis of PPA

The specific synthesis method is as follows. First, 27.32 g (0.15 mol) of TEP and 14.42 g (0.24 mol) of EDM (the ratio of  $-\text{OCH}_2\text{CH}_3$  to  $-\text{NH}_2$  is 3:3.2) were added to a three-necked 100 mL flask equipped with a nitrogen gas inlet, a condenser and a mechanical stirrer. The mixture was stirred at an appropriate rate. Subsequently, the system was slowly heated to  $85^\circ\text{C}$  to reach the reaction temperature. This temperature was kept for 8 hours. Then, the temperature was increased by  $10^\circ\text{C}$ , lasting for 4 hours. Finally, the light-yellow viscous liquid was collected into the vial as the product cooled.

The product was dissolved in an appropriate amount of ethanol and loaded into the RC dialysis membranes. After 24 hours of dialysis, steaming was carried out to remove the solvent. After the steaming, the sample was placed in a vacuum oven overnight. The product PPA (named P1) was then obtained.

$^1\text{H}$  NMR (400 Hz,  $\text{DMSO}-d_6$ ):  $\delta$  3.69 (p, 16H), 3.44 (q, 1H), 2.77 (m, 14H), 2.63 (dq, 1H), 1.11 (m, 30H).

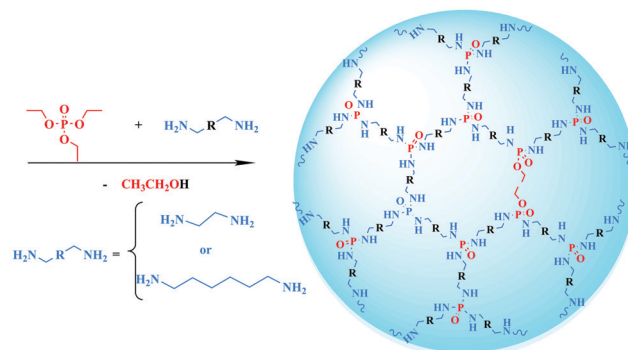
P2 was synthesized using 27.32 g (0.15 mol) of TEP and 12.60 g (0.21 mol) of EDM (the ratio of  $-\text{OCH}_2\text{CH}_3$  to  $-\text{NH}_2$  is 3:2.8) in a similar synthesis process to P1.

P3 was synthesized using 27.32 g (0.15 mol) of TEP and 16.20 g (0.27 mol) of EDM (the ratio of  $-\text{OCH}_2\text{CH}_3$  to  $-\text{NH}_2$  is 3:3.6) based on the synthesis process for P1.

P4 was synthesized using 27.32 g (0.15 mol) of TEP and 31.38 g (0.27 mol) of 1,6-diaminohexane (the ratio of  $-\text{OCH}_2\text{CH}_3$  to  $-\text{NH}_2$  is 3:3.6) based on the synthesis process for P1. The reaction temperature was  $120^\circ\text{C}$ .

$^1\text{H}$  NMR (400Hz,  $\text{DMSO}-d_6$ ):  $\delta$  3.68 (p, 1H),  $\delta$  2.57 (m, 1H),  $\delta$  1.50 (tq, 1H),  $\delta$  1.28 (q, 1H),  $\delta$  1.08 (dt, 2H).

The synthesis route and specific synthesis data of PPA are shown in Scheme 1 and Table S1 (ESI<sup>†</sup>). The schematic structures of P1–P4 are shown in Fig. S1 (ESI<sup>†</sup>). Moreover, the possible reaction mechanism is exhibited in Scheme S1 (ESI<sup>†</sup>). It is notable that the reactivity of EDM is higher than that of DAH, so we selected P1 as the main research object.



Scheme 1 The synthetic route of PPA.



### 3. Results and discussion

Distinct from the current method using phosphorus oxychloride for the synthesis of PPA, in this work, P1–P4 were obtained using TEP and EDM or DAH via one-pot ester aminolysis. In addition, no solvent was applied during the process and the by-product ethanol can be easily removed by distillation. Thus, the proposed synthesis method is simple, safe and has no complicated purification steps.

#### 3.1 FT-IR study

FT-IR was carried out to determine the chemical structure of the obtained PPA, and the spectra of TEP, EDM, DAH, P1 and P4 are shown in Fig. 1. In Fig. 1A, the band at about  $3390\text{ cm}^{-1}$  is related to the telescopic vibration of the amino groups ( $-\text{NH}_2$ ). The telescopic vibration of  $-\text{CH}_2-$  is observed at around  $2965\text{ cm}^{-1}$ . The absorption peak of the bending vibration of N–H appears at about  $1377\text{ cm}^{-1}$ . The bands at  $1215\text{ cm}^{-1}$  and  $1058\text{ cm}^{-1}$  belong to  $\text{P}=\text{O}$  and  $\text{C}-\text{N}$ , respectively. In Fig. 1B, the adsorption peak at around  $3325\text{ cm}^{-1}$  is classified as the telescopic vibration of the amino groups. The adsorption peaks of  $\text{P}=\text{O}$  and  $\text{C}-\text{N}$  appear at about  $1225\text{ cm}^{-1}$  and  $1051\text{ cm}^{-1}$ , respectively, while the bands at  $1035\text{ cm}^{-1}$  ( $\text{C}-\text{O}$ ) and  $929\text{ cm}^{-1}$  ( $\text{P}-\text{O}$ ) disappear. The obtained FT-IR spectra offer obvious evidence for the synthesis of the designed PPA.

#### 3.2 NMR study and degree of branching of PPA

To further confirm the structure of the synthesized PPA, we obtained  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{31}\text{P}$  NMR spectra of TEP, EDM, DAH, P1–P4 (Fig. 2 and Fig. S2–S4, ESI†). In the spectrum of TEP in Fig. 2A, the proton peaks marked by 1 and 2 are associated with  $-\text{OCH}_2\text{CH}_3$  and  $-\text{OCH}_2\text{CH}_3$ , respectively. While the peaks at about 2.50 and 3.30 are consistent with DMSO- $d_6$  and water. In the EDM spectra (Fig. 2A), the proton peaks marked by 1 and 2 are associated with  $-\text{NH}_2$  and  $-\text{CH}_2-$ . In the  $^1\text{H}$  NMR spectra of P1, the signal marked by 4 at  $\delta = 1.11\text{ ppm}$  belongs to  $-\text{NH}-$ . The signal marked by 2 at  $\delta = 2.63\text{ ppm}$  is related to  $-\text{CH}_2-\text{NH}_2$ . The signal of  $-\text{CH}_2-\text{NH}-$  appears at  $\delta = 2.77\text{ ppm}$  is marked by 3. The peak marked by 1 at 3.69 ppm comes from  $-\text{NH}_2$ . The detailed and analytical  $^1\text{H}$  NMR spectra of P1 are displayed in Fig. 2B. In the DAH spectra (Fig. 2C), the proton peaks marked by 2, 3 and 4 are associated with  $\text{H}_2\text{N}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{NH}_2$  (2.49–2.52 ppm),  $\text{H}_2\text{N}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{NH}_2$  (1.23–1.28 ppm), and  $\text{H}_2\text{N}-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{NH}_2$

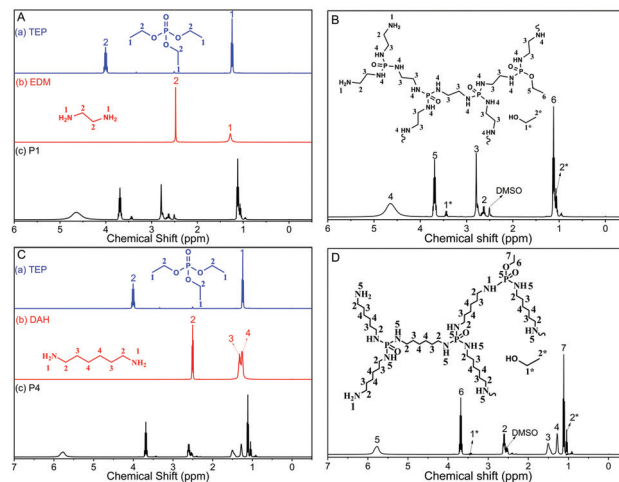


Fig. 2 (A)  $^1\text{H}$  NMR spectra of TEP, EDM and P1; (B)  $^1\text{H}$  NMR spectra of P1; (C)  $^1\text{H}$  NMR spectra of TEP, DAH and P4; (D)  $^1\text{H}$  NMR spectra of P4.

(1.32–1.37 ppm) respectively. In Fig. 2D, it can be seen that the proton peak associated with  $\text{CH}_2-\text{NH}_2$  arises at 3.65–3.72 ppm, while the proton peaks at 2.51–2.61 ppm are associated with  $\text{CH}_2-\text{NH}-\text{P}$  and  $\text{CH}_2-\text{NH}_2$ . The proton peaks associated with  $\text{CH}_2-\text{CH}_2-\text{NH}-\text{P}$  (or  $\text{CH}_2-\text{CH}_2-\text{NH}_2$ ),  $\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{NH}-\text{P}$  (or  $\text{CH}_2-\text{CH}_2-\text{CH}_2-\text{NH}_2$ ) and  $\text{CH}_2-\text{NH}-\text{P}$  appear at 1.44–1.53 ppm, 1.26–1.30 ppm and 1.03–1.13 ppm, respectively. The  $^1\text{H}$  NMR analysis of P2–P3 as well as the  $^{13}\text{C}$  and  $^{31}\text{P}$  NMR analysis of reactants and P1–P4 are shown in Fig. S2–S4 (ESI†). Furthermore, the degree of branching (DB) is an important parameter of hyperbranched polymers. We calculated the DB of P1–P4 according to the  $^1\text{H}$  NMR data, and the results are shown in Table S2 (ESI†).

The NMR spectra and DB value further confirm that PPA (P1–P4) have been successfully synthesized through the proposed one-pot ester aminolysis method.

#### 3.3 GPC Study

The GPC data including the weight average molecular weight ( $M_w$ ), number average molecular weight ( $M_n$ ) and  $\text{Đ}$  ( $M_w/M_n$ ) of P1–P4 are shown in Fig. S5 (ESI†). For P1, the  $M_w$ ,  $M_n$  and  $\text{Đ}$  are 16 500, 10 600 and 1.56, respectively, which verified that the synthesized P1 is a polymer with a high molecular weight.

The FT-IR spectra, NMR spectra and GPC results sufficiently prove that PPA has been synthesized successfully.

#### 3.4 Optical properties

The obtained PPA is a light-yellow viscous liquid in sunlight, but it can emit intense fluorescence under a 365 nm UV lamp, hence, we systematically studied the optical properties of PPA for the first time. The results showed that the fluorescence of PPA is closely related to the concentration of PPA: that is, a dilute solution of PPA shows weak fluorescence but the pure state of PPA can emit strong blue fluorescence, which indicates that PPA is a novel AIE polymer.

Subsequently, we employ ethanol and tetrahydrofuran (THF) as a good solvent and a poor solvent to further verify the AIE

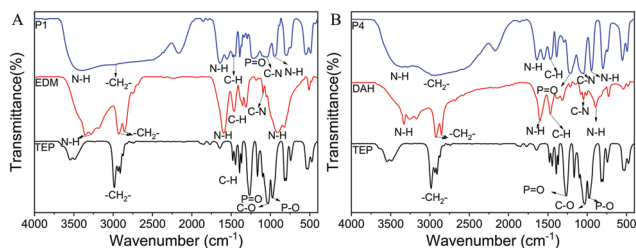
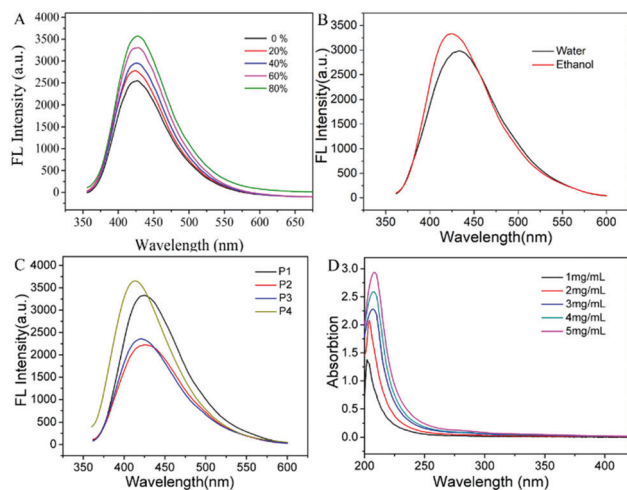


Fig. 1 (A) FT-IR spectra of TEP, EDM and P1; (B) FT-IR spectra of TEP, DAH and P4.





**Fig. 3** (A) Fluorescence spectra of P1 at 20 mg mL<sup>-1</sup> in THF/ethanol mixtures with different fractions of THF ( $\lambda_{\text{ex}} = 345$  nm); (B) fluorescence spectra of P1 in different solvents ( $\lambda_{\text{ex}} = 345$  nm); (C) fluorescence spectra of P1, P2, P3 and P4 ( $\lambda_{\text{ex}} = 345$  nm); (D) UV-vis absorption spectra of P1 solution at various concentrations.

character of PPA. In ethanol solution, P1 emits relatively weak blue fluorescence with a wavelength of 415 nm. With an increase in THF fraction ( $f_{\text{T}}$ ), the emission intensity gradually increases. The fluorescence intensity increased by 41% in the ethanol/THF mixture ( $f_{\text{T}} = 80\%$ ) compared to that in pure ethanol solution. It should be noted that, according to classical AIE theory, the ethanol solution of PPA should show barely any fluorescence. But in this experiment, due to the physicochemical properties of the polymer, it will form a partial aggregation driven by hydrogen bonds even in good solvents (Fig. S6, ESI<sup>†</sup>), resulting in relatively strong fluorescence. With the addition of a poor solvent, the degree of aggregation of the polymer will further be enhanced, and thus its fluorescence intensity will also increase to a certain extent (Fig. 3A). Such a phenomenon is consistent with the AIE effect.

To our surprise, PPA is soluble both in water and in ethanol. This property is undoubtedly beneficial to the application of PPA. Then we investigated the fluorescence properties of P1 solution in water and in ethanol, respectively. As can be seen in Fig. 3B, P1 can emit strong fluorescence both in water and ethanol, but its fluorescence intensity is stronger in ethanol. This could be caused by the different polarities of water and ethanol. It is well known that the polarity of water is stronger than that of amino groups, which makes it more soluble in water. So P1 tends to form tighter aggregates in ethanol than in water. The formation of polymer aggregates reduces the non-radiative loss of energy, so the fluorescence intensity of P1 in ethanol is stronger than that in water. The strong fluorescence intensity in aqueous solution and good biocompatibility of PPA indicate great potential in chemical sensing and bio-imaging.

In order to explore the relationship between the fluorescence properties of PPA and its molecular structure, four kinds of PPA were prepared. P1–P3 are synthesized with the TEP to EDM ratios at 1:1.6, 1:1.4, 1:1.8, respectively, and P4 is

prepared with the TEP to DAH ratio at 1:1.6. The photoluminescence properties of P1–P4 in ethanol solution are shown in Fig. 3C. As can be seen in Fig. 3C, P4 shows the strongest fluorescence intensity and P2 with ethoxy termination shows the lowest. By comparing the fluorescence intensity of ethoxy terminated P2 and primary amine terminated P3, we can draw the conclusion that a primary amine group is not the key factor for the fluorescence of the PPA, but it can increase the fluorescence intensity. Apart from the functional group, molecular weight is another factor that can influence the fluorescence intensity of PPA. P1 has a higher molecular weight than P3 (data shown in Fig. S5, ESI<sup>†</sup>) and exhibits stronger fluorescence intensity than P3, indicating that the fluorescence intensity of PPA is positively correlated with its molecular weight. Compared with P1, P4 has a smaller molecular weight and a longer chain. As shown in Fig. 3C, the fluorescence intensity of P4 is stronger than that of P1, indicating that the chain length plays a more important role than molecular weight in affecting the fluorescence of PPA.

The UV absorption spectra of P1 in ethanol solution were acquired at diverse concentrations. The detailed spectra are shown in Fig. 3D. The UV absorption band of PPA is mainly concentrated around 205 nm, which is caused by the  $n-\pi^*$  electronic transition of  $\text{C}-\text{NR}_2$  and  $\text{O}=\text{P}-\text{NH}$ . Along with the increase in the PPA concentration, UV absorption intensity is gradually enhanced. The enhancement is attributed to the clusterization of  $-\text{NH}_2$ ,  $\text{P}-\text{O}$  and  $\text{P}=\text{O}$  with the growing concentration of PPA. We speculate that such a phenomenon is caused by the formation of an “electronic delocalization system”<sup>26</sup> as the concentration increases, which decreases the energy gap between the HOMO and LUMO orbitals and makes it easier to trigger an electronic transition, resulting in an enhancement in the absorption intensity. The formed electronic delocalization system also leads to a red-shift of the absorption wavelength.

Subsequently, we investigated the fluorescence properties of P1 at different concentrations. As the concentration of P1 increases, the fluorescence intensity gradually increases, showing a significant AIE property. The specific fluorescence excitation/emission spectra of P1 at different concentrations are shown in Fig. 4A. Apparently, both the excitation and emission intensity increase with the increase in P1 concentration. In Fig. 4A, the excitation bands of P1 appear at around 262 and 345 nm, while the emission band appears at about 425 nm. The excitation band at around 262 nm red-shifts to 272 nm as the concentration increases, and the shapes of the excitation band at 345 nm and the emission band at 425 nm are symmetric as mirror images. It is obvious that the peak intensity at around 262 nm increases more slowly than that at 345 nm as the concentration increases. It is believed that the band at 262 nm belongs to the molecular skeleton of P1, while the band at about 345 nm is attributed to the formed electronic delocalization system. As the concentration increases, the fluorescence is mainly caused by the electronic delocalization system rather than the molecular skeleton. In contrast to P1, there is only one band in the excitation spectra of P4 (Fig. 4B). Such results indicate that the molecular skeleton of P4 can hardly emit





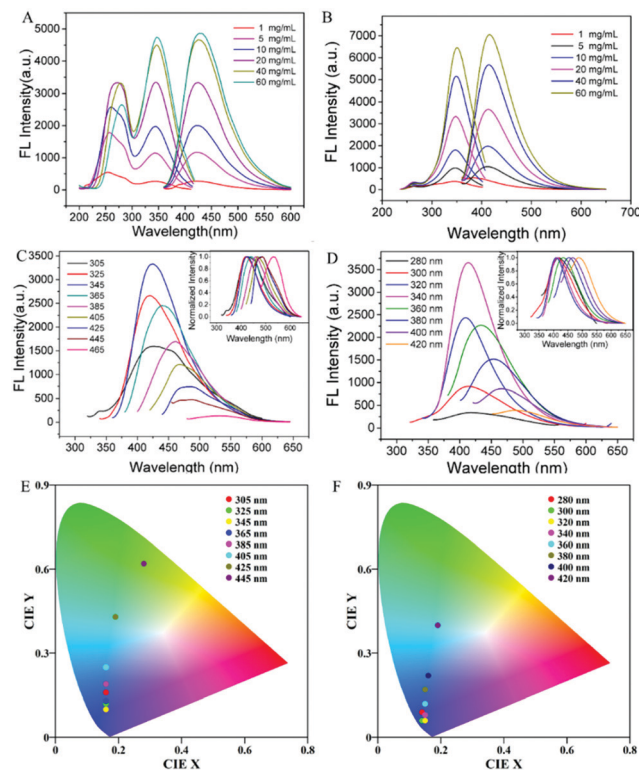


Fig. 4 (A) Emission spectra of P1 under different concentrations ( $\text{mg mL}^{-1}$ ,  $\lambda_{\text{ex}} = 345 \text{ nm}$ ); (B) emission spectra of P4 under different concentrations ( $\text{mg mL}^{-1}$ ,  $\lambda_{\text{ex}} = 340 \text{ nm}$ ); (C) excitation-dependent fluorescence of P1 at  $20 \text{ mg mL}^{-1}$  (Inset: Normalized emission); (D) excitation-dependent fluorescence of P4 with at  $20 \text{ mg mL}^{-1}$  (Inset: Normalized emission); (E) CIE chromaticity coordinates obtained from the emission spectra of P1 at  $20 \text{ mg mL}^{-1}$ ; (F) CIE chromaticity coordinates obtained from the emission spectra from P4 at  $20 \text{ mg mL}^{-1}$ .

fluorescence under UV light, while the fluorescence mainly originates from the electronic delocalization system.

Then we further studied the effect of excitation wavelength on PPA's fluorescence performance. As shown in Fig. 4C, the highest fluorescence intensity exists at the excitation wavelength of  $345 \text{ nm}$ . When the wavelength is longer than  $345 \text{ nm}$ , the fluorescence intensity decreases with the increase in wavelength, along with an obvious red-shift (Fig. 4C, inset). Afterward, Commission Internationale d'Éclairage (CIE) chromaticity coordinates of P1 solution at  $20 \text{ mg mL}^{-1}$  were calculated from the emission spectra under various wavelengths and are shown in Fig. 4E. From Fig. 4E, we can see that the emission colours vary from blue to green, which is consistent with the clustering-triggered emission mechanism.<sup>27–29</sup> Compared with P1, the red-shift of P4 is not that obvious (Fig. 4F). Moreover, with the longer chain, P4 can only form local through-space conjugation, while P1 can form a through-space conjugation ring (confirmed by the subsequent DFT calculation results). Therefore, the absorption range of P4 narrows, resulting in a less obvious red-shift.<sup>30</sup>

Moreover, we obtained the fluorescence spectra, fluorescence lifetime and absolute quantum yield (QY) of pure PPA. The fluorescence spectra of pure P1 are shown in Fig. 5A. The

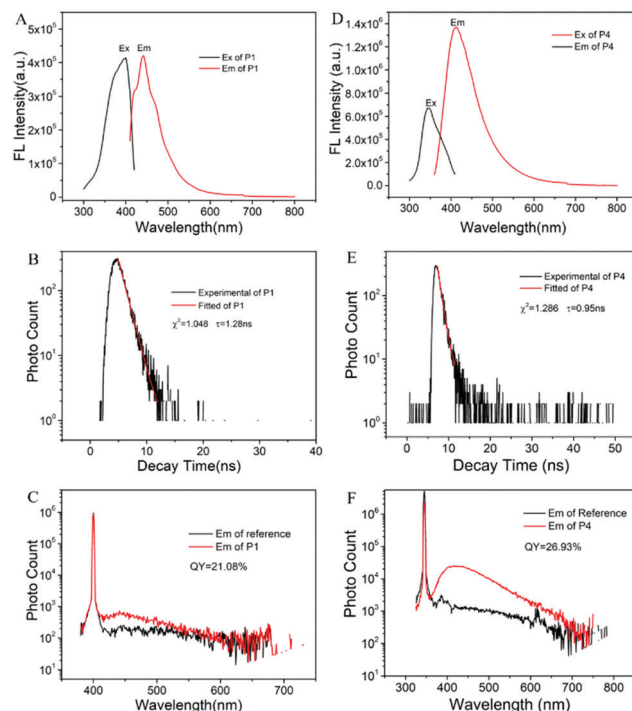


Fig. 5 (A) Excitation spectrum (Ex,  $\lambda_{\text{em}} = 443 \text{ nm}$ ) and emission spectrum (Em,  $\lambda_{\text{ex}} = 399 \text{ nm}$ ) of P1; (B) transient photoluminescence decay curve of P1 at  $443 \text{ nm}$  after excitation at  $399 \text{ nm}$ ; (C) absolute fluorescence quantum yield of P1 excited at  $399 \text{ nm}$ ; (D) excitation spectrum (Ex,  $\lambda_{\text{em}} = 412 \text{ nm}$ ) and emission spectrum (Em,  $\lambda_{\text{ex}} = 345 \text{ nm}$ ) of P4; (E) Transient photoluminescence decay curve of P4 at  $412 \text{ nm}$  after excitation at  $345 \text{ nm}$ ; (F) absolute fluorescence quantum yield of P4 excited at  $345 \text{ nm}$ .

excitation band appears at  $399 \text{ nm}$ , and the emission band appears at  $443 \text{ nm}$ . Compared with the excitation and emission spectra of P1 solution in ethanol, the excitation and emission bands red-shift for about  $50 \text{ nm}$  and  $20 \text{ nm}$ , respectively. This phenomenon is caused by the electron delocalization system formed in different chemical environments. Fig. 5B displays the fluorescence decay curve of P1 at  $443 \text{ nm}$  after the excitation at  $399 \text{ nm}$ . The measured fluorescence lifetime of P1 is  $1.28 \text{ ns}$ . Fig. 5C shows that the absolute QY of pure P1 is  $21.08\%$ , which is relatively high among the non-aromatic AIE polymers.<sup>3,4</sup>

We also tested the fluorescence properties of P4 to systematically study the influence of chain length. By comparing Fig. 5A and D, we can see that the fluorescence intensity of P4 is stronger than that of P1. We believe that the longer chain in P4 increases the flexibility of the P4 molecules, resulting in the formation of denser luminogen clusters and leading to stronger fluorescence. The fluorescence lifetime of P4 is  $0.95 \text{ ns}$ , which is shorter than that of P1, owing to the smaller energy gap (confirmed by subsequent DFT calculation). Besides, the QY of P4 is  $26.93\%$ , slightly higher than that of P1 (Fig. 5C and F), which provides a possible method to improve the QY of non-aromatic AIE polymers.

Compared with other non-aromatic AIE polymers, especially poly(amidoamine)s,<sup>31–33</sup> P incorporated PPA shows outstanding fluorescence properties, including strong fluorescence

intensity and high QY.<sup>21,34,35</sup> The P incorporated PPA provides a possible method for the design of non-aromatic AIE polymers with high fluorescence intensity and QY.

### 3.5 The fluorescence mechanism study

According to the fluorescence properties of PPA, there is no doubt that the fluorescence intensity of PPA is related to its aggregate state. Hence, we explore the fluorescence mechanism of PPA through theoretical calculation and experiment investigation. Firstly, we employ density functional theory (DFT) at the level of B3LYP/6-31G (d) to obtain the optimized conformations and energy level of PPA (P1 and P4) with the Gaussian 09 program package. To simplify the calculation, we calculated the energy levels of the first-generation P1 and P4 with the molecule numbers increased from 1 to 4 (Table 1), and their optimized structures are displayed in Fig. S7 (ESI<sup>†</sup>). With the increase in the molecule numbers of P1, the compact structure of P1 is formed due to the intermolecular O $\cdots$ H and N $\cdots$ H interactions (1.949 Å, 2.059 Å, 2.050 Å, 2.113 Å, 2.035 Å, 2.149 Å, 2.202 Å) (Fig. 6A). The abundant hydrogen bonds diminish the distance between P1 molecules and rigidify the conformation of P1, which is beneficial to the formation of through-space conjugation. Similarly, intermolecular O $\cdots$ H and N $\cdots$ H interactions also promote the self-assembly of P4 (2.060 Å, 2.217 Å, 2.134 Å, 2.031 Å, 2.270 Å) (Fig. 6C). These interactions could lead to the overlap of the electron clouds of N and O, resulting in the formation of an electron delocalization system. At the same time, the HOMO–LUMO energy gap decreases as the number of P1/P4 molecules increases, which demonstrates that the molecules are more likely to be excited.

To further explore the fluorescence mechanism of PPA, we investigated the morphology of P1 at different concentrations (10 mg mL<sup>−1</sup> and 50 mg mL<sup>−1</sup>) through TEM. As shown in Fig. 7A, P1 assembles into loose irregular structures at a concentration of 10 mg mL<sup>−1</sup>. Nevertheless, at a concentration of 50 mg mL<sup>−1</sup>, P1 assembles into spherical structures with uneven size (Fig. 7B and C). The compact conformation at high concentrations is conducive to forming clusters of P1, resulting in the stronger fluorescence intensity.

On the basis of the above theoretical calculations and investigation, we presume that the clusteroluminescence affected by the electron delocalization and molecular self-assembly is the origin of PPA's fluorescence. The fluorescence

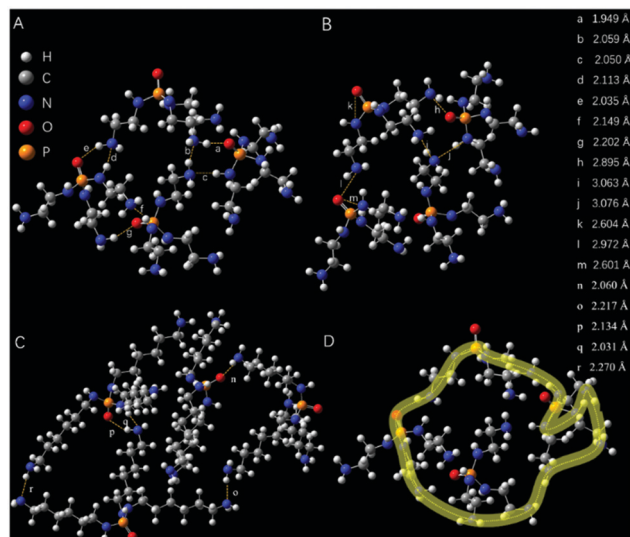


Fig. 6 (A) The accumulation of four P1 first-generation molecules induced by H $\cdots$ O and H $\cdots$ N strong intermolecular interactions; (B) intramolecular and intermolecular O $\cdots$ N and N $\cdots$ N interactions between four P1 molecules; (C) the accumulation of four P4 first-generation molecules induced by H $\cdots$ O and H $\cdots$ N strong intermolecular interactions; (D) schematic diagram of the through-space conjugation for P1.

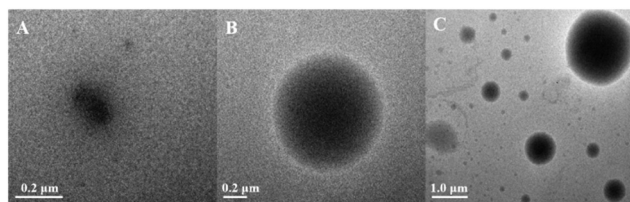


Fig. 7 Micrograph of P1 self-assembly morphology in ethanol with concentrations of 10 mg mL<sup>−1</sup> (A) and 50 mg mL<sup>−1</sup> (B–C).

mechanism schematic diagram of PPA is shown in Fig. 8. As the concentration of PPA or ratio of poor solvent increases, the PPA tends to form a tighter cluster owing to the intermolecular hydrogen bonds, leading to stronger fluorescence-emission.

### 3.6 pH dependency

Considering that PPA contains a large amount of O and N atoms, we further explored the pH dependency property of P1. The pH responsiveness of P1 was investigated at different

Table 1 DFT calculation results of HOMO–LUMO energy levels of the conformations with 1–4 first-generation P1 and P4 molecules

Molecules number		E(HOMO) (a.u.)	E(LUMO) (a.u.)	Energy gap (a.u.)	Energy gap (eV)
P1	1	−0.226	0.066	0.292	7.946
	2	−0.210	0.051	0.261	7.102
	3	−0.219	0.045	0.264	7.184
	4	−0.207	0.043	0.250	6.803
P4	1	−0.226	0.065	0.291	7.919
	2	−0.208	0.062	0.270	7.347
	3	−0.205	0.047	0.252	6.857
	4	−0.204	0.045	0.249	6.776

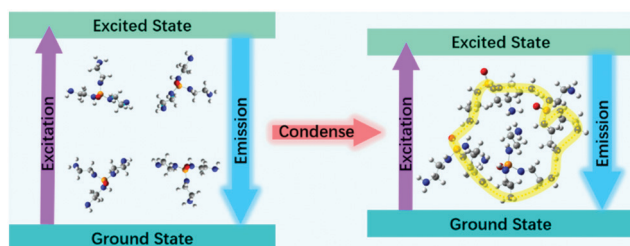


Fig. 8 Fluorescence mechanism schematic diagram of PPA.



pH ranging from 2 to 12 with a P1 concentration of 5 mg mL<sup>-1</sup>. The original pH of the P1 aqueous solution is 8.48 (Fig. S8, ESI<sup>†</sup>), and the fluorescence intensity is strongest when pH = 4. When the pH value is in the range of 4–12, as the pH value increases, the intramolecular and intermolecular hydrogen bonds between P1 molecules are affected by increasing OH<sup>-</sup>, and this will lead to the loose structure of P1 with weak fluorescence. When the pH value is below 4, the amino groups in P1 will combine with the hydrogen ions to form ammonium ions, resulting in a repulsive force between P1 molecules. With the formation of the looser conformation of P1, a decrease in the fluorescence intensity of P1 is observed. Due to the pH dependency property, PPA is expected to be used as a pH probe in the chemical sensing field.

### 3.7 Ion probe

To our surprise, the addition of PPA can change the colour of copper chloride solution from colourless to purple without any effect on the fluorescence of PPA (Fig. 9A and Fig. S9, ESI<sup>†</sup>). This is attributed to the large amount of phosphoryl bonds contained in the synthesized PPA. After adding PPA to copper chloride solution, the secondary amines in the phosphoryl bonds are complexed with the copper ions. While the coordination complex shows a purple colour, thereby making the solution turn from colourless to purple (Scheme S2, ESI<sup>†</sup>). The secondary amine groups have no effect on the fluorescence of PPA, so its fluorescence intensity does not change. Then we analyzed the relationship between the absorbance of coordination complex solution (with a P1 concentration of 20 mg mL<sup>-1</sup>) and the concentration of copper ions (Fig. 9B). It is obvious that the absorbance increased linearly with the increase in the concentration of copper ions ( $R^2 = 0.9996$ ). The detection limit is 0.15 mmol L<sup>-1</sup> in theory. Besides, the other metal ions (Na<sup>+</sup>, Cd<sup>2+</sup>, Fe<sup>3+</sup>, Hg<sup>2+</sup>, Co<sup>2+</sup>, Al<sup>3+</sup>, Zn<sup>2+</sup>) have no such effect, for they cannot complex with PPA and form coloured coordination complexes (Fig. S10, ESI<sup>†</sup>). We also found out that the solution colour will change to wathet blue (Fig. S11, ESI<sup>†</sup>) after removing Cu<sup>2+</sup> by ethylenediaminetetraacetic acid disodium salt (EDTA-2Na). Furthermore, we explored the biocompatibility of PPA by CCK-8 assays,<sup>36</sup> and the results are shown in Fig. S12 (ESI<sup>†</sup>). We can see that the cell viability is more than 84% after incubation for 24 h with a concentration of PPA as high as 1.0 mg mL<sup>-1</sup>, indicating the excellent biocompatibility of PPA. This discovery

provides an effective and environmentally friendly means for the qualitative and quantitative detection of copper ions.

## 4. Conclusions

In summary, a green and convenient method is proposed for the synthesis of PPA through an ester ammonolysis reaction by TEP and EDM or DAH instead of polycondensation by phosphorus oxychloride and diamine. The by-products are facile to separate and harmless to the environment. What is more, PPA could emit strong blue fluorescence under UV light and shows an obvious AIE character. This is attributed to the fact that PPA could assemble into aggregates and form a through-space conjugation due to intermolecular and intramolecular hydrogen bonds and the interactions between phosphoryl bonds and amino groups. Such aggregation is helpful to the formation of luminogen clusters, resulting in the fluorescence of PPA. The molecular weight, amino groups and chain length are all important factors that effectively affect the PPA's fluorescence intensity, HOMO–LUMO energy gap and quantum yield. What is more, PPA is sensitive to Cu<sup>2+</sup> and can change the colour of copper chloride solution from colourless to purple, realizing the qualitative and quantitative visual detection of Cu<sup>2+</sup> instead of time-consuming and expensive Cu<sup>2+</sup> detection methods. This work provides an innovative method for PPA synthesis, as well as an enrichment of P incorporated AIE polymers and an *in situ* visual detection agent for Cu<sup>2+</sup>.

## Author contributions

L. G.: synthesis, structural analysis, writing – original draft. H. Y.: supervision of conceptualization and methodology. L. W.: synthesis, data plotting, writing – original draft. P. Y.: DFT calculation, structural analysis. L. Y.: TEM experiment. Y. Z.: writing – polishing.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- 1 J. Mei, N. Leung, R. Kwok, J. Lam and B. Z. Tang, *Chem. Rev.*, 2015, **115**, 11718–11940.

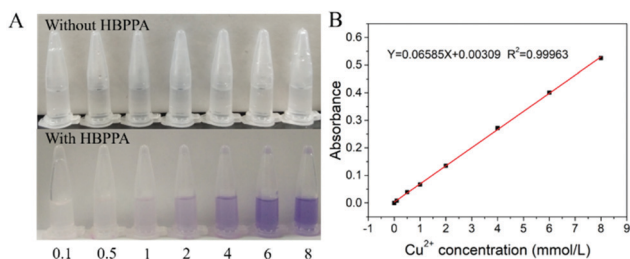


Fig. 9 (A) The colour change phenomenon of PPA aqueous solution (20 mg mL<sup>-1</sup>) with different concentrations of Cu<sup>2+</sup> (mmol L<sup>-1</sup>); (B) The relationship between absorbance and the concentration of Cu<sup>2+</sup>.





- 2 J. Yang, Z. Chi, W. Zhu, B. Z. Tang and Z. Li, *Sci. China: Chem.*, 2019, **62**, 1090–1098.
- 3 W. Yuan and Y. Zhang, *J. Polym. Sci., Part A: Polym. Chem.*, 2017, **55**, 560–574.
- 4 D. Tomalia, B. Klajnert-Maculewicz, K. Johnson, H. Brinkman, A. Janaszewska and D. Hedstrand, *Prog. Polym. Sci.*, 2019, **90**, 35–117.
- 5 Y. Feng, H. Yan, F. Ding, T. Bai, Y. Nie, Y. Zhao, W. Feng and B. Z. Tang, *Mater. Chem. Front.*, 2020, **4**, 1375–1382.
- 6 D. Wu, Y. Liu, C. He and S. Goh, *Macromolecules*, 2005, **38**, 9906–9909.
- 7 L. Yuan, H. Yan, L. Bai, T. Bai, Y. Zhao, L. Wang and Y. Feng, *Macromol. Rapid Commun.*, 2019, **40**, 1800658.
- 8 L. Bai, H. Yan, L. Wang, T. Bai, L. Yuan, Y. Zhao and W. Feng, *Macromol. Mater. Eng.*, 2020, **305**, 2000126.
- 9 R. Wang, W. Yuan and X. Zhu, *Chin. J. Polym. Sci.*, 2015, **33**, 680–687.
- 10 R. Restani, P. Morgado, M. Ribeiro, I. Correia, A. Aguiar-Ricardo and V. Bonifácio, *Angew. Chem., Int. Ed.*, 2012, **51**, 5162–5165.
- 11 S. Niu, H. Yan, S. Li, P. Xu, X. Zhi and T. Li, *Macromol. Chem. Phys.*, 2016, **217**, 1185–1190.
- 12 Y. Feng, T. Bai, H. Yan, F. Ding, L. Bai and W. Feng, *Macromolecules*, 2019, **52**, 3075–3082.
- 13 L. Bai, H. Yan, T. Bai, Y. Feng, Y. Zhao, Y. Ji, W. Feng, T. Lu and Y. Nie, *Biomacromolecules*, 2019, **20**, 4230–4240.
- 14 S. Niu, H. Yan, Z. Chen, L. Yuan, T. Liu and C. Liu, *Macromol. Rapid Commun.*, 2016, **37**, 136–142.
- 15 F. Liao, Y. Ju, X. Dai, Y. Cao, J. Li and X. Wang, *Polym. Degrad. Stab.*, 2015, **120**, 251–261.
- 16 X. Hu, Y. Guo, L. Chen, X. Wang, L. Li and Y. Wang, *Polym. Degrad. Stab.*, 2012, **97**, 1772–1778.
- 17 Q. Lv, J. Huang, M. Chen, J. Zhao, Y. Tan, L. Chen and Y. Wang, *Ind. Eng. Chem. Res.*, 2013, **52**, 9397–9404.
- 18 F. Liao, L. Zhou, Y. Ju, Y. Yang and X. Wang, *Ind. Eng. Chem. Res.*, 2014, **53**, 10015–10023.
- 19 W. Xu, Y. Wang, S. Li, Z. Ke, Y. Yan, S. Li, Z. Xing, C. Wang, F. Zeng, R. Liu and F. Deng, *RSC Adv.*, 2015, **5**, 5425–5432.
- 20 J. Köhling, V. Kozel, V. Jovanov, R. Pajkert, S. Tverdomed, O. Gridenco, M. Fugel, S. Grabowsky, G. Rösenthaller and V. Wagner, *ChemPhysChem*, 2019, **20**, 665–671.
- 21 T. Baumgartner, *Acc. Chem. Res.*, 2014, **47**, 1613–1622.
- 22 S. Padghan, R. Bhosale, S. Bhosale, F. Antolasic, M. Kobaisi and S. Bhosale, *Molecules*, 2017, **22**, 1417.
- 23 J. Zhang, S. Devaramani, D. Shan and X. Lu, *Anal. Bioanal. Chem.*, 2016, **408**, 7155–7163.
- 24 C. Karadaş and D. Kara, *Food Chem.*, 2017, **232**, 242–248.
- 25 I. Rapp, C. Schlosser, D. Rusiecka, M. Gledhill and E. Achterberg, *Anal. Chim. Acta*, 2017, **976**, 1–13.
- 26 V. Bekiari, P. Lianos, U. Stangar, B. Orel and P. Judeinstein, *Chem. Mater.*, 2000, **12**, 3095–3099.
- 27 Q. Zhou, Z. Wang, X. Dou, Y. Wang, S. Liu, Y. Zhang and W. Yuan, *Mater. Chem. Front.*, 2019, **3**, 257–264.
- 28 X. Dou, Q. Zhou, X. Chen, Y. Tan, X. He, P. Lu, K. Sui, B. Tang, Y. Zhang and W. Yuan, *Biomacromolecules*, 2018, **19**, 2014–2022.
- 29 S. Zheng, T. Zhu, Y. Wang, T. Yang and W. Yuan, *Angew. Chem., Int. Ed.*, 2020, **59**, 1–6.
- 30 Y. Feng, H. Yan, F. Ding, T. Bai, Y. Nie, Y. Zhao, W. Feng and B. Tang, *Mater. Chem. Front.*, 2020, **4**, 1375–1382.
- 31 S. Lin, T. Wu, Y. Jao, C. Liu, H. Lin, L. Lo and C. Yang, *Chem. – Eur. J.*, 2011, **17**, 7158–7161.
- 32 S. Koley and S. Ghosh, *Phys. Chem. Chem. Phys.*, 2016, **18**, 24830–24834.
- 33 W. Yang, X. Wu, F. Liu, Y. Dou, Z. Hu and W. Hao, *RSC Adv.*, 2016, **41**, 34252–34260.
- 34 I. Manners, *J. Polym. Sci., Part A: Polym. Chem.*, 2002, **40**, 179–191.
- 35 T. Baumgartner and R. Réau, *Chem. Rev.*, 2006, **106**, 4681–4727.
- 36 C. Yan, N. Zhang, P. Guan, P. Chen, S. Ding, T. Hou, X. Hu, J. Wang and C. Wang, *Int. J. Biol. Macromol.*, 2020, **153**, 723–735.

