



Gallium nanoparticles as novel inhibitors of A β 40 aggregation

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Alzheimer's disease (AD) has been consistently related to the formation of senile amyloid plaques mainly composed of amyloid β (A β) peptides. The toxicity of A β aggregates has been indicated to be responsible for AD pathology. One scenario to decrease A β toxicity is the development of effective inhibitors against A β amyloid formation. In this study, we investigate the effect of gallium nitride nanoparticles (GaN NPs) as inhibitors of A β 40 amyloid formation using a combination of biophysical approaches. Our results show that the lag phase of A β 40 aggregation kinetics is significantly retarded by GaN NPs in a concentration dependent manner, implying the activity of GaN NPs in interfering with the formation of the crucial nucleus during A β aggregation. Our results also show that GaN NPs can reduce the amyloid fibril elongation rate in the course of the aggregation kinetics. It is speculated that the high polarization characteristics of GaN NPs may provoke a strong interaction between the particles and A β 40 peptide and in this way decrease self-association of the peptide monomers to form amyloids.

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Introduction

Alzheimer's disease (AD) is a progressive, irreversible, and currently incurable neurodegenerative disorder characterized by neuronal loss, memory impairment, and cognitive decline.^{1,2} As the leading cause of dementia in a rapidly aging population, AD is a growing threat to global health, economies, and society in general. One possible pathway to develop AD has been linked to the formation of senile plaques due to the aggregation of a group of key amyloid- β (A β) peptides, *e.g.*, A β 40 and A β 42, that are formed from the cleavage of the transmembrane amyloid precursor protein (APP) by β - and γ -secretases.²⁻⁴ Amyloid formation of A β is typically viewed as a nucleated polymerization process, manifested by an initial lag phase in which the oligomeric nuclei are formed, followed by a rapid growth phase during which the nuclei grow by the addition of monomers to form fibrillar structures.^{5,6} Moreover, amyloid oligomers have also been shown to form hetero-oligomers by interacting with its isoforms or other proteins.^{5,7} Therefore great effort has been devoted to characterize oligomers to better understand the

amyloid formation mechanism and conceive plausible therapeutic alternatives.^{4,6}

One of the plausible therapeutic strategies for treating the disease is to inhibit the formation of toxic A β aggregate species. As such, a wide variety of amyloid inhibitors that can prevent the formation of A β aggregates, *e.g.*, small organic compounds and designed peptides, have been reported.^{2,3,8-10} Also, recent studies have demonstrated that various nanoparticles (NPs) can either completely or partially inhibit A β fibril formation, due to their interactions with A β .¹¹⁻¹³ NPs have been used as an important build block to construct new A β inhibitors using surface modification and changes in morphology of the particles.¹⁴

Different substrates have been used to establish NP cores for the development of efficient NPs for therapeutic applications. The common NP formulations employed in the diagnosis and therapeutic studies of AD include polymeric NPs, gold NPs (Au NPs), gadolinium NPs, selenium NPs, protein-based NPs, and polysaccharide-based NPs.¹⁵ For example, Liao *et al.* showed that bare Au NPs inhibited A β fibrillation, leading to the formation of fragmented fibrils and spherical oligomers.¹⁶ These authors reasoned that the negative surface potential of Au NPs is an important factor for the inhibition of A β amyloid fibril formation. Similarly, Yin *et al.* demonstrated that selenium NPs not only effectively inhibit A β aggregation but could disaggregate preformed A β fibrils into non-toxic amorphous oligomers.¹⁷ Family III nitrides, particularly gallium, aluminium, and indium nitrides, have gained considerable interest for use as transducing materials in biosensors.¹⁸ Specifically,

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gallium nitride (GaN) is a semiconductor material with wurtzite crystal structure that has a wide (3.4 eV) band gap and is widely used in ultraviolet–blue light emitting diodes, photodetectors and lasers.¹⁹ In addition, GaN demonstrates good biocompatibility and aqueous stability in conditions relevant to biological applications.^{20–22} Therefore, GaN has also been used in various biological applications. Braniste *et al.* reported the uptake of GaN NPs by porcine endothelial cells,²³ and their results suggested that the uptake of the particles by the cells is strongly dependent upon whether the GaN NPs are fixed to a substrate surface or free floating in the medium. The endothelial cells growing on surfaces functionalized with GaN NPs demonstrated excellent adhesion and proliferation, suggesting good biocompatibility of the nanoparticles. The same research group expanded their studies and demonstrated that GaN/Fe-based nanoparticles exhibiting magnetic properties can be taken up by endothelial cells and stored within vesicles.²⁴ In addition, Hofstetter *et al.* performed an investigation of the effects of differently treated GaN surfaces on mouse fibroblast cell proliferation and growth dynamics.²⁵ Their results suggest that GaN does not require specific surface treatments to be biocompatible and that cell signalling is not affected by GaN–cell interactions. Jewett *et al.* demonstrated that functionalized and etched GaN surfaces are stable in aqueous environments and leach a negligible amount of Ga in solution even in the presence of hydrogen peroxide.²⁶ Also, GaN surfaces in cell culture did not interfere with nearby cell growth, and the etched GaN promoted the adhesion of cells better than the etched silicon surfaces. On the other hand, Ga³⁺ can interfere with the iron metabolism in cells leading to apoptosis.²⁷ This was attributed to the similar ionic radius of Ga³⁺ and Fe³⁺ and Ga³⁺ being less electroactive than Fe³⁺ in the iron metabolic processes.²⁸

While GaN has emerged as a promising candidate for bio-interfacing,²⁹ to the best of our knowledge, the potential use of GaN as an inhibitor of aggregation of amyloidogenic proteins including A β , has not been explored. Since previous studies have shown that GaN can interact with various cellular components, it is possible that GaN NPs are able to interact with A β peptide and in consequence affect the aggregation behaviour. In this study, we investigate the effect of GaN NPs on the aggregation of A β 40 *in vitro* using a combination of biophysical approaches. In comparison to A β 42, A β 40 is less toxic to neurons and exerts a more moderate aggregation propensity and thus is appropriate for a sophisticated investigation of the properties of aggregation kinetics. Our results demonstrate that GaN NPs markedly affects the characteristics of A β 40 oligomerization and fibrillization. It is expected that further molecular modification of the surface characteristics such as the charge properties of GaN NPs may be an approach for designing potent NP inhibitors against protein amyloidogenesis.

Experimental

Materials

A β 40 was obtained from Life Tein Company (NJ, USA). Certified analysis of the peptide established that the purity was higher

than >95% with a formula weight of 4329.9. The peptide was used as received. Gallium nitrate hydrate was obtained from Sigma-Aldrich (USA) with >99.9% purity trace metals basis. Congo red (CR) with high purity was obtained from Calbiochem (UK). Thioflavin T (ThT) was obtained from Sigma-Aldrich (USA). Other solvents and reagents were obtained from either Sigma-Aldrich (USA) or Fisher Scientific (Cayey, PR) and used without further purification.

Synthesis of GaN NPs

The GaN NPs were synthesized according to previous reports.^{30–34} Briefly, Ga(NO₃)₃ was dissolved in nitric acid, the pH was adjusted to 8.2 with ammonium hydroxide and citric acid was added until the solution became clear. The resulting solution was dried and heated in a burnout furnace under air at 400 °C for 4 hours producing a Ga₂O₃ powder that was converted to GaN in a tube furnace at 900 °C under an NH₃ gas flow for 3 hours to ensure complete conversion.³¹

X-Ray diffraction (XRD) measurements

The phase structures of the nanoparticle samples were identified using X-ray diffraction techniques using a Bruker D2 Phaser with Cu K α radiation (wavelength of the radiation, $\lambda = 1.54 \text{ \AA}$) operating at 30 kV and 10 mA. The incident beam passed through a 2.5° Soller slit, a 0.6° (1.0 mm) divergence slit, a secondary 2.5° Soller slit, a Ni filter, and a receiving slit of 0.2 mm. After interaction with the sample, the diffracted beam was detected using a scintillation counter. The samples were analyzed utilizing a Bragg–Brentano geometry over a range of 5–85° 2θ with a step size of 0.02° 2θ , with each step measured for one second. For all XRD measurements, the samples were prepared as self-supported pressed powders over PMMA specimen holders from Bruker. Data analysis was performed using the instrument software, which includes the spectral database. In addition, XRD is a convenient method for determining the mean size of single-crystal nanoparticles or crystallites in nanocrystalline bulk materials.³⁵ The size of the synthesized nanoparticles was calculated using the Scherrer formula, $D = K\lambda/\beta\cos\theta$ where D is the average grain size, K is the shape factor (taken as 0.9), λ is the X-ray wavelength, β is the peak width measured in radians as the full width at half of the maximum intensity, and the Bragg angle is 21.485°.

A β 40 aggregation kinetics followed by thioflavin T (ThT) emission

The A β 40 peptide utilized in the kinetic aggregation assay was monomerized as described previously.^{36,37} Briefly, lyophilized A β 40 powder was dissolved in aqueous NaOH solution (2 mM) and the pH was adjusted to 11 using a 100 mM NaOH solution followed by sonicating the solution for 1 hour in an ice-cold water bath. The resulting solution was filtered through a 0.22 μm filter and kept on ice before use. The concentration of the A β 40 stock solution was determined using tyrosine UV absorbance at 280 nm [$\epsilon_{280} = 1280 \text{ (M cm)}^{-1}$] using an Agilent 8453 photodiode array spectrometer. The fluorescence kinetics were measured using a Fluoroskan Ascent microplate reader (Thermo-Nicolet, USA). Fibril formation of A β 40 was followed



using the ThT fluorescence emission using filters (bandpass of 15 nm) with an excitation of 435 and emission at 485 nm, respectively. Each well contained 20 μM A β 40, 20 μM ThT, 150 mM NaCl in 50 mM sodium phosphate buffer saline (PBS pH = 7.4) with or without GaN NPs in a total volume of 300 μL . The emission was measured through the bottom of the plate every five minutes with five seconds of gentle shaking (100 rpm) before readings at 37 $^{\circ}\text{C}$. The A β 40 aggregation kinetic parameters were obtained from the average of at least three replicates. Fitting analysis was performed according to eqn (1),³⁸

$$F(t) = F_0 + m_0t + \frac{F_f + m_f t}{(1 + \exp^{-k(t-\tau_{50})})} \quad (1)$$

where $F(t)$ is the fluorescence at time t , F_0 and m_0 are the intercept and slope of the initial baseline, F_f and m_f are the intercept and slope of the final baseline, k is the pseudo first order rate constant of aggregation, and τ_{50} is the time that is required to achieve half of the fluorescence amplitude between initial and maximum fluorescence. The lag time (τ_{lag}) is the time taken to reach the exponential phase of fibrillation and is given by $\tau_{50} - 2/k$. The fitting variables are F_0 , m_0 , F_f , m_f , k , and τ_{50} . Curve fitting was done with MS Office 2016 using the Excel Solver tool.

Congo red binding assay

Congo red (CR) was dissolved in PBS at a concentration of 100 μM and filtered through a 0.22 μm membrane. The solutions were prepared by dilution of stock solutions to obtain the final concentrations of 5.0 μM CR, 150 mM NaCl, 50 mM PBS 7.4, and A β 40 (15 μM) and incubated at 37 $^{\circ}\text{C}$ in a 10 mm pathlength optical quartz cell. The absorbance spectra were measured at certain times for the samples in triplicate. The incubation was run with gentle agitation.³⁹

Turbidity study

The aggregation kinetics of A β 40 was determined by monitoring the absorbance at 600 nm as reported previously.^{40,41} Samples were prepared as described in the CR binding studies but without the dye.

Scanning electron microscopy (SEM)

SEM images for nanoparticle characterization were acquired on a JEOL JSM-IT-100 with an Energy Dispersive Spectrometer operating at 25 or 30 kV. For amyloid fibrils, SEM images were acquired on an FEI Quanta 600 ESEM operating at 15 kV at the Singh Center for Nanotechnology at the University of Pennsylvania.⁴² The sample solution was diluted 100-fold with pure water. Subsequently, the diluted samples were placed on silicone and dried under nitrogen for 10 min before measuring the images.

Results and discussion

Characterization of the synthesized GaN NPs

The morphology of the synthesized GaN NPs was investigated using SEM. As shown in Fig. 1, the GaN NP samples dispersed

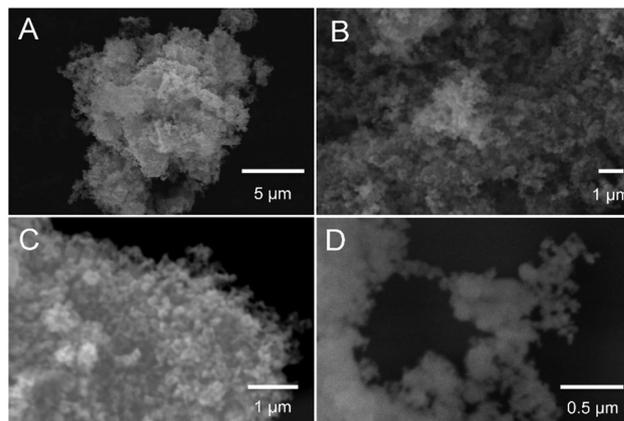


Fig. 1 SEM images of the synthesized GaN NPs. The images were measured at different kilovolts and magnifications: (A) 25 kV, 5000 \times ; (B) 30 kV, 10 000 \times ; (C) 30 kV, 20 000 \times ; and (D) 30 kV, 50 000 \times .

in water exhibit a typical crystallite morphology. The XRD spectrum of the GaN NPs resembles that previously reported by Chen *et al.*,³⁰ as shown in Fig. 2A. Specifically, the XRD pattern of the GaN NPs exhibits peaks at 2θ of 32.80, 35.20, and 37.26 and four other major peaks at 48.8, 58.2, 64.2, and 69.3 degrees, respectively. All these peaks can be indexed to a hexagonal (wurtzite) GaN structure. The GaN NP sizes determined using the Scherrer formula and the (110) peak at 58.2 degrees range from 6 to 12 nm. The energy-dispersive spectrum (Fig. 2B) reveals that the elemental composition of the GaN NPs is 88.53, 10.01, and 1.46 w/w% for gallium, nitrogen, and oxygen, respectively.

Impact of GaN NPs on A β 40 aggregation kinetics measured using ThT fluorescence

The formation of A β 40 amyloid aggregates starting from its monomeric state was studied using ThT fluorescence in the absence or presence of GaN NPs in sodium phosphate buffer (50 mM PBS buffer, 150 mM NaCl, pH 7.4). ThT is an environmentally sensitive fluorophore whose binding to amyloid fibrils dramatically increases its fluorescence quantum yield.⁴³ Fig. 3 shows the time dependent ThT fluorescence intensity changes of the A β 40 sample (20 μM) in the absence or presence of GaN NPs. The kinetic curves are fitted using eqn (1) to obtain aggregation parameters such as τ_{50} and τ_{lag} (summarized in Fig. 4). In the absence of GaN NPs, the A β 40 aggregation kinetics exhibit a typical sigmoidal shape, characteristic of a nucleation polymerization mechanism,⁴⁴ which consists of a lag phase associated with nucleation, a fast growth phase related to the propagation of fibrils, and a final stationary phase related to the aggregated state. The kinetics exhibit a τ_{50} of 1044 minutes (Fig. 4), and a growth phase with a rate constant of $5.45 \times 10^{-3} \text{ min}^{-1}$. To rule out the possibility that ThT can directly interact with the GaN NPs, causing its fluorescence intensity to change, we also performed control experiments where no A β 40 is present. The results show that the ThT emission in the presence of GaN NPs does not only change with time (Fig. 3, curve f) and that the amplitude overlaps with the



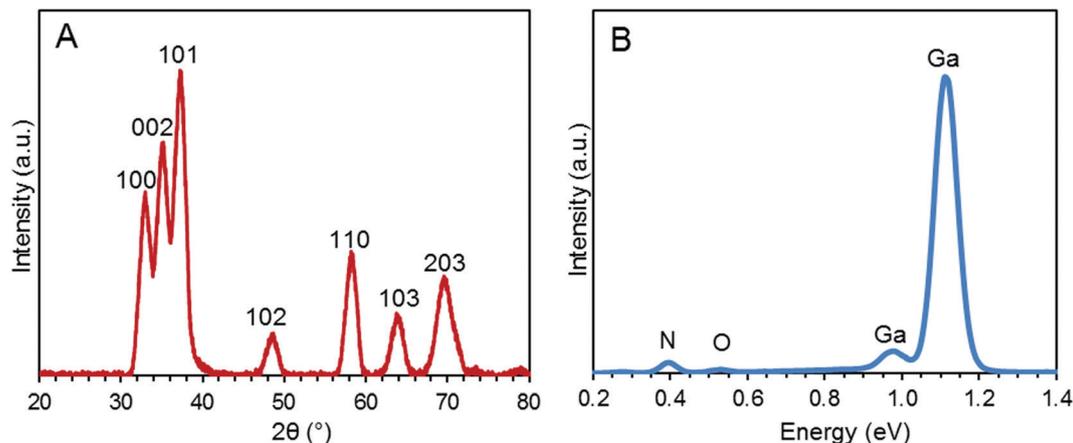


Fig. 2 (A) XRD pattern of the GaN NPs. (B) Energy-dispersive spectrum of the GaN NPs. The weight percent elemental data for nitrogen, oxygen, and gallium is 10.01, 1.46, and 88.53%, respectively.

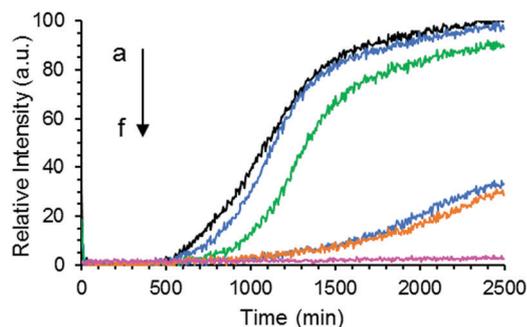


Fig. 3 Aggregation kinetics of A β 40 (20 μ M) only (a), or in the presence of 47 (b), 60 (c), 67 (d), 73 μ g mL $^{-1}$ (e) of GaN NPs, respectively followed by ThT fluorescence. Curve f is the fluorescence intensity of the sample containing 80 μ g mL $^{-1}$ GaN NPs and 20 μ M ThT only without A β 40.

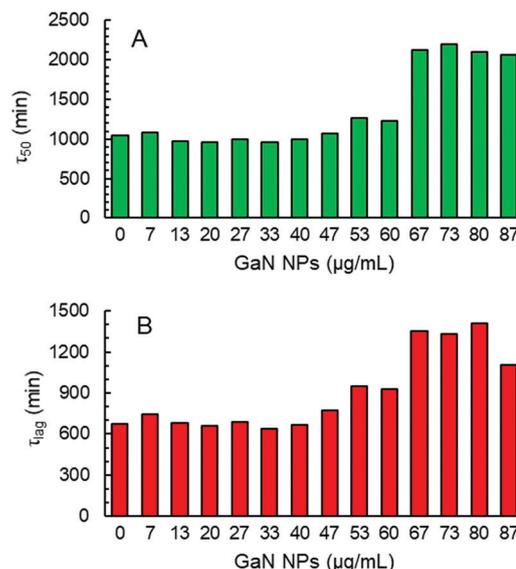


Fig. 4 (A) τ_{50} of the aggregation kinetics of A β 40 (20 μ M) in the presence of different amounts of GaN NPs. (B) τ_{lag} of the aggregation kinetics of A β 40 (20 μ M) in the presence of different amounts of GaN NPs.

initial baseline of the kinetic runs containing A β 40. This confirms that the observed ThT fluorescence intensity change in the A β 40 aggregation measurements is not due to direct interactions between ThT and GaN NPs.

The addition of GaN NPs markedly delays the aggregation kinetics of A β 40. In the presence of 60 μ g mL $^{-1}$ GaN NPs, the value of τ_{50} increases to 1227 minutes (Fig. 3 and 4), *ca.*18% longer than that of A β 40 alone. Increasing the GaN concentration to 73 μ g mL $^{-1}$ results in a two-fold increase in τ_{50} relative to that of A β 40 in the absence of GaN NPs, up to 2197 minutes. Further increase of the GaN concentration shows smaller change in τ_{50} . Furthermore, as shown in Fig. 4B, the τ_{lag} of the A β 40 aggregation kinetics is approximately 700 minutes, and the addition of 67 μ g mL $^{-1}$ GaN increases the τ_{lag} value to 1349 minutes. This result suggests that the presence of appropriate amounts of GaN can delay the rate of critical oligomeric nucleus formation of A β . It is worth noting that the highest GaN concentration achievable in the experiments is limited by its solubility. Increasing the GaN concentration also affects the apparent growth rate constant (k) of the exponential phase. For instance, the k value of the kinetics in the presence

of 67 μ g mL $^{-1}$ GaN NPs is 2.57×10^{-3} min $^{-1}$, which is less than half that of the kinetics of the peptide only. A larger k value suggests a higher order exponential phase where growth of the fibrils occurs.^{45–49} The results suggest that the addition of higher amount of GaN NPs can also significantly interfere with the growth process of the amyloid fibrils, thus inhibiting the formation of amyloids.

The maximum of ThT fluorescence amplitude in A β aggregation kinetics is influenced by the addition of GaN NPs. In general, ThT fluorescence emission intensity is proportional to the amyloid fibers present, and a decrease in the intensity can be indicative of a decrease in the overall fiber content.⁵⁰ As depicted in Fig. 5, increasing the GaN NP concentration lowers the ThT maximum fluorescence amplitude. These results



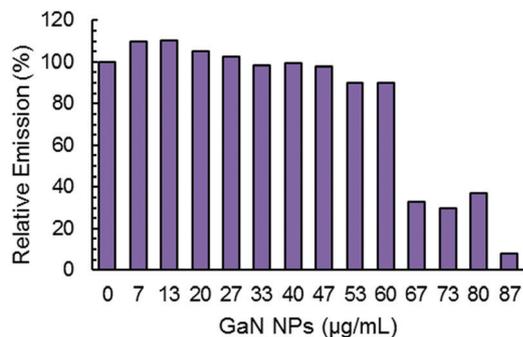


Fig. 5 Normalized maximum ThT fluorescence emission intensity during the aggregation of A β 40 (20 μ M) without or with different concentrations of GaN NPs.

suggest the inhibition of A β 40 fibril formation by GaN NPs. It is interesting that there is also an apparent threshold concentration value of approximately 60 μ g mL $^{-1}$ where the inhibition efficiency starts to increase sharply. Taken together, these results demonstrate that GaN NPs can delay the formation of the crucial nucleus in A β aggregation and slow down the rate of A β 40 amyloid fibril formation. In addition, GaN NPs can also decrease the growth rate constant of A β 40 amyloid formation.

Although a general mechanism for the underlying impact of NPs on protein fibril formation remains elusive,¹⁴ the structural characteristics of the NPs such as size, shape, and the surface properties have been demonstrated to be crucial in the

interactions of the particles and proteins and the inhibition activities of NPs on protein amyloid formation.^{49,51,52} The inhibition activity of GaN on A β 40 aggregation likely arises from interactions between the peptide and the NPs in aqueous solution, which may be driven by interactions such as electrostatic and van der Waals interactions. The non-centrosymmetric wurtzite hexagonal structure of GaN NPs has planes consisting of anions and cations that attract each other *via* electrostatic forces inducing a spontaneous polarization that is larger than other materials.¹⁸ It is conceivable that this polarization property may play an important role in the interaction of GaN and A β 40 peptide, which, as a result, would prevent or decrease peptide–peptide interactions possibly by stabilizing the A β 40 monomers.

Inhibition of GaN NPs on A β 40 aggregation measured using CR binding

CR has also been widely used in monitoring peptide aggregation and fibril formation. The absorption spectrum of CR peaks at around 490 nm (π – π^* transition of the azo group) in aqueous solution,⁵³ is shifted to *ca.* 550 nm when A β 40 fibrils are present. This bathochromic shift is easily seen visually where an orange red to rose change in colour is observed. We conducted an experiment using CR as a probe to further prove that GaN NPs can decrease the aggregation rate of A β 40 and its fibril quantity. The absorption spectra of different solutions containing CR alone, A β 40–CR, and A β 40–CR–GaN were

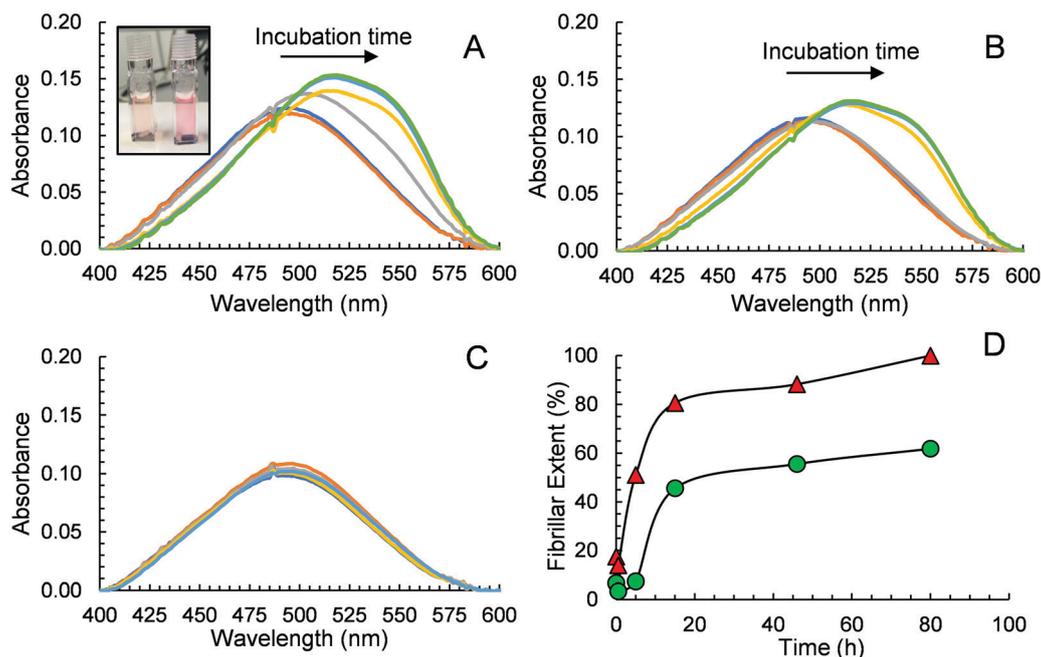


Fig. 6 Absorption spectra of A β 40 (15 μ M, containing 5 μ M CR) in the absence (A) or in the presence (B) of 87 μ g mL $^{-1}$ GaN NPs. The spectra were measured at the incubation times of 0 h (dark blue), 0.5 h (orange), 5 h (gray), 15 h (yellow), 46 h (light blue), and 80 h (green). Inset in (A): photo of samples containing GaN–A β 40 (left) and A β 40 (right) showing the difference in colour after a 5 h incubation. (C) Absorption spectra of CR and GaN NPs only measured at the incubation times of 0 h (dark blue), 0.5 h (orange), 5 h (gray), 15 h (yellow), 46 h (light blue), and 80 h (green). (D) Quantitative evaluation of fibrillar extent during the aggregation of A β 40 in the absence (red triangles) or in the presence (green circles) of GaN NPs (87 μ g mL $^{-1}$). A baseline correction due to scattering was applied to each spectrum before reading the absorbance values. The fibrillar extent values are relative to the maximum A β 40 fibrillar concentration at 80 h for an A β 40 solution in the absence of GaN NPs.



measured at different times of incubation. The solution containing only A β 40 shows an initial absorption spectrum that resembles the one of CR with a broad band peak with a maximum at 490 nm (Fig. 6A). After \sim 5 hours, the formation of A β 40 amyloids is indicated by the absorbance increase in the 520–550 nm region (Fig. 6A). The absorbance in this region increases with time reaching a plateau after \sim 20 hours. When A β 40 is incubated in the presence of GaN NPs ($87 \mu\text{g mL}^{-1}$), the intensity change of the absorbance at 541 nm, characteristic for A β 40 aggregation, is slower than that of A β 40 alone (Fig. 6B vs. 6A). This result further indicates the inhibition activity of GaN NPs on A β 40 aggregation. To rule out the possibility that CR can directly interact with GaN NPs and cause a change in the absorption, we also performed control experiments in the absence of A β 40. The results show that the CR–GaN NPs absorption spectrum does not change with incubation time (Fig. 6C), hence corroborating that the spectral changes are due to the formation of A β 40 fibrils.

Quantitative fibrillar formation was also studied using CR as a probe.⁵⁴ The concentration of A β fibrils at certain times, in $\mu\text{g mL}^{-1}$, was estimated from CR binding where $[\text{A}\beta_{40\text{fibrils}}]_t = (A_{541\text{mix}}/4780) - (A_{403\text{mix}}/6830) - (A_{403\text{CR}}/8620)$ where $A_{541\text{mix}}$ and $A_{403\text{mix}}$ are the absorbances of the solutions containing A β 40 or A β 40–GaN at 541 and 403 nm, respectively, and $A_{403\text{CR}}$ is the absorbance of the solution containing CR alone under the same experimental incubation time.⁵⁴ The results are shown in Fig. 6D. The data shows that the lag phase is around 4 hours for A β 40 in the presence of GaN ($87 \mu\text{g mL}^{-1}$), which is approximately three times longer than that in the absence of GaN. In addition, the presence of GaN NPs reduces the fibrillar content by up to 62% after 80 hours of incubation. These results are in accord with what observed in the ThT kinetics study.

Inhibition of GaN NPs on A β 40 aggregation confirmed using turbidity and imaging studies

Turbidity measurements were also performed to monitor A β 40 aggregation as it is well-known that protein aggregate formation leads to an increase in the sample absorbance at 600 nm.⁴⁰ The absorbance intensity at 600 nm was monitored as a function of time for A β 40 aggregation in the absence and presence of GaN NPs at $87 \mu\text{g mL}^{-1}$. As shown in Fig. 7, the presence of GaN NPs significantly decreases the fibril formation kinetics, consistent with both the ThT and CR results. The inhibitory activity of the particles on A β 40 fibrillogenesis was further confirmed using SEM imaging (Fig. 8). SEM images were acquired using the samples from the A β 40 aggregation kinetics study in the absence or presence of GaN NPs. In the absence of GaN NPs, A β 40 aggregated to form fibrils of a few micrometers long and produced large conglomerates. In contrast, in the presence of GaN NPs, the number of fibrils was dramatically reduced and no large clusters were observed. Taken together, these results corroborate that GaN NPs are an effective inhibitor of A β 40 amyloid fibril formation.

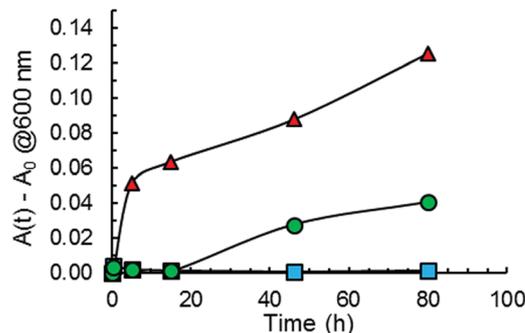


Fig. 7 Turbidity curves for A β 40 aggregation in the absence (red triangles) or in the presence of $87 \mu\text{g mL}^{-1}$ GaN NPs (green circles) measured by monitoring the absorbance at 600 nm. A curve for the solution containing GaN NP and CR alone (blue squares) incubated in parallel is also shown. Solid lines are to guide the reader.

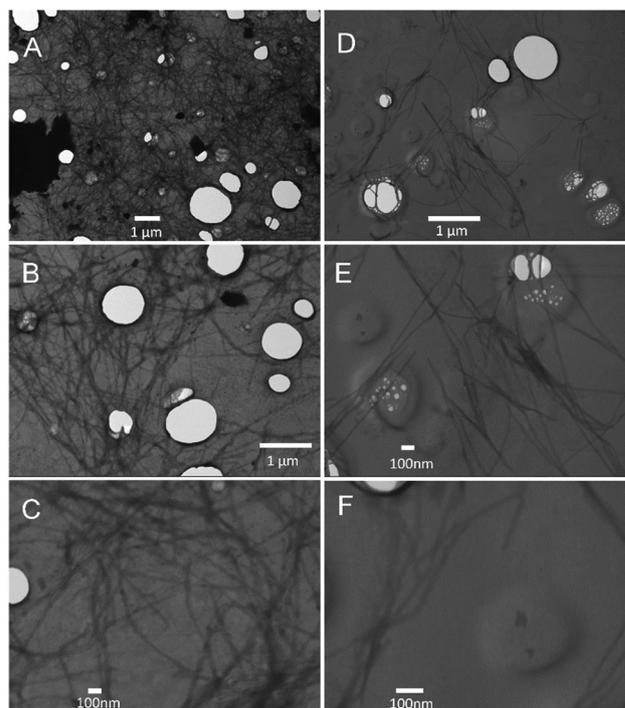


Fig. 8 SEM images of A β 40 ($20 \mu\text{M}$) acquired after the aggregation kinetics measurement using ThT fluorescence emission. Left: SEM images of A β 40 only measured at different magnifications: (A) $10\,000\times$, (B) $20\,000\times$, and (C) $50\,000\times$. Right: SEM images of A β 40 in the presence of $87 \mu\text{g mL}^{-1}$ GaN NPs measured at different magnifications: (D) $20\,000\times$, (E) $50\,000\times$, and (F) $100\,000\times$.

Conclusions

In summary, we studied the effect of GaN NPs on A β 40 amyloid fibril formation using a variety of approaches including ThT fluorescence, CR absorbance, turbidity, and SEM imaging. Our results show that the presence of GaN NPs significantly delays the lag time of A β 40 aggregation, indicating its inhibiting activity on crucial oligomeric nucleus formation of the peptide.



Also, the results show that GaN NPs can slow down the amyloid fibril elongation rate in the amyloid growth phase of the aggregation kinetics. Considering that the non-centrosymmetric wurtzite hexagonal structure of GaN NPs contains planes consisting of anions and cations that induce strong spontaneous polarization, it is plausible that this polarization characteristic of the particle can provoke a stronger interaction between the NPs and A β 40 peptide and thus decrease intermolecular interactions among the peptide monomers to form amyloids. Future studies on modification of the surface characteristics of gallium nanoparticles to enhance solubility and selectivity may lead to the discovery of new NP inhibitors with high biocompatibility and inhibition efficiency.

Author contributions

KMT, ASD, ERS, NVFC, and AM performed the experiments. IR, DD and RO conceived and wrote the paper.

Conflicts of interest

The authors declare no competing financial interests.

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References

- 1 A. K. Sahoo, J. Dandapat, U. C. Dash and S. Kanhar, *J. Ethnopharmacol.*, 2018, **215**, 42–73.
- 2 J. Cummings, G. Lee, A. Ritter and K. Zhong, *Alzheimer's Dementia*, 2018, **4**, 195–214.
- 3 S. K. Singh, S. Srivastav, A. K. Yadav, S. Srikrishna and G. Perry, *Oxid. Med. Cell. Longevity*, 2016, **2016**, 7361613.
- 4 P. H. Nguyen, A. Ramamoorthy, B. R. Sahoo, J. Zheng, P. Faller, J. E. Straub, L. Dominguez, J.-E. Shea, N. V. Dokholyan, A. De Simone, B. Ma, R. Nussinov, S. Najafi, S. T. Ngo, A. Loquet, M. Chiricotto, P. Ganguly, J. McCarty, M. S. Li, C. Hall, Y. Wang, Y. Miller, S. Melchionna, B. Habenstein, S. Timr, J. Chen, B. Hnath, B. Strodel, R. Kaye, S. Lesné, G. Wei, F. Sterpone, A. J. Doig and P. Derreumaux, *Chem. Rev.*, 2021, **121**, 2545–2647.
- 5 B. R. Sahoo, S. J. Cox and A. Ramamoorthy, *Chem. Commun.*, 2020, **56**, 4627–4639.
- 6 E. E. Cawood, T. K. Karamanos, A. J. Wilson and S. E. Radford, *Biophys. Chem.*, 2021, **268**, 106505.
- 7 M. I. Ivanova, Y. Lin, Y.-H. Lee, J. Zheng and A. Ramamoorthy, *Biophys. Chem.*, 2021, **269**, 106507.
- 8 P. Williams, A. Sorribas and M.-J. R. Howes, *Nat. Prod. Rep.*, 2011, **28**, 48–77.
- 9 C. Cabaleiro-Lago, F. Quinlan-Pluck, I. Lynch, S. Lindman, A. M. Minogue, E. Thulin, D. M. Walsh, K. A. Dawson and S. Linse, *J. Am. Chem. Soc.*, 2008, **130**, 15437–15443.
- 10 D. J. Selkoe, *Science*, 1997, **275**, 630–631.
- 11 R. Liu, J. Yang, L. Liu, Z. Lu, Z. Shi, W. Ji, J. Shen and X. Zhang, *Adv. Sci.*, 2020, **7**, 1901555.
- 12 A. Binda, C. Murano and I. Rivolta, *Int. J. Nanomed.*, 2020, **15**, 6113–6135.
- 13 G. Altinoglu and T. Adali, *Curr. Drug Targets*, 2020, **21**, 628–646.
- 14 T. John, A. Gladysz, C. Kubeil, L. L. Martin, H. J. Risselada and B. Abel, *Nanoscale*, 2018, **10**, 20894–20913.
- 15 J. Gupta, M. T. Fatima, Z. Islam, R. H. Khan, V. N. Uversky and P. Salahuddin, *Int. J. Biol. Macromol.*, 2019, **130**, 515–526.
- 16 Y.-H. Liao, Y.-J. Chang, Y. Yoshiike, Y.-C. Chang and Y.-R. Chen, *Small*, 2012, **8**, 3631–3639.
- 17 T. Yin, L. Yang, Y. Liu, X. Zhou, J. Sun and J. Liu, *Acta Biomater.*, 2015, **25**, 172–183.
- 18 X. Li and X. Liu, *Nanoscale*, 2017, **9**, 7320–7341.
- 19 D. Franke, A. Luisa da Rosa, M. Lorke and T. Frauenheim, *Phys. Status Solidi B*, 2019, **256**, 1800455.
- 20 C. M. Foster, R. Collazo, Z. Sitar and A. Ivanisevic, *Langmuir*, 2013, **29**, 8377–8384.
- 21 G. Steinhoff, O. Purrucker, M. Tanaka, M. Stutzmann and M. Eickhoff, *Adv. Funct. Mater.*, 2003, **13**, 841–846.
- 22 T. H. Young and C. R. Chen, *Biomaterials*, 2006, **27**, 3361–3367.
- 23 T. Braniste, I. Tiginyanu, T. Horvath, S. Raevschi, S. Cebotari, M. Lux, A. Haverich and A. Hilfiker, *Beilstein J. Nanotechnol.*, 2016, **7**, 1330–1337.
- 24 T. Braniste, I. Tiginyanu, T. Horvath, S. Raevschi, B. Andree, S. Cebotari, E. C. Boyle, A. Haverich and A. Hilfiker, *Nano-scale Res. Lett.*, 2017, **12**, 486.
- 25 M. Hofstetter, J. Howgate, M. Schmid, S. Schoell, M. Sachsenhauser, D. Adigüzel, M. Stutzmann, I. D. Sharp and S. Thalhammer, *Biochem. Biophys. Res. Commun.*, 2012, **424**, 348–353.
- 26 S. A. Jewett, M. S. Makowski, B. Andrews, M. J. Manfra and A. Ivanisevic, *Acta Biomater.*, 2012, **8**, 728–733.
- 27 P. Sahoo, P. S. Murthy, S. Dhara, V. P. Venugopalan, A. Das and A. K. Tyagi, *J. Nanopart. Res.*, 2013, **15**, 1841.
- 28 S. R. Choi, B. E. Britigan and P. Narayanasamy, *Antimicrob. Agents Chemother.*, 2019, **63**, e02643–02618.
- 29 L. E. Bain, M. P. Hoffmann, I. Bryan, R. Collazo and A. Ivanisevic, *Nanoscale*, 2015, **7**, 2360–2365.
- 30 Y. Chen, N. Jyoti and J. Kim, *Appl. Phys. A*, 2011, **102**, 517–519.
- 31 H.-L. Li, R.-J. Xie, N. Hirotsaki, B. Dierre, T. Sekiguchi and Y. Yajima, *J. Am. Ceram. Soc.*, 2008, **91**, 1711–1714.



- 32 J. L. Robles-Garcia, A. Melendez, D. Yates, J. J. Santiago-Aviles, I. Ramos and E. M. Campo, *Proc. SPIE*, 2011, **8036**, 80360F/80361–80360F/80367.
- 33 A. Melendez, K. Morales, I. Ramos, E. Campo and J. J. Santiago-Aviles, *Proc. SPIE*, 2009, **7402**, 740210/740211–740210/740216.
- 34 E. M. Campo, J. Poplawsky, A. Melendez, J. L. Robles-Garcia, I. Ramos, D. Yates, J. E. Fischer and J. J. Santiago-Aviles, *RSC Adv.*, 2014, **4**, 2634–2639.
- 35 U. Holzwarth and N. Gibson, *Nat. Nanotechnol.*, 2011, **6**, 534.
- 36 N. P. Cook, V. Torres, D. Jain and A. A. Martí, *J. Am. Chem. Soc.*, 2011, **133**, 11121–11123.
- 37 H. Liu, R. Lantz, P. Cosme, N. Rivera, C. Andino, W. G. Gonzalez, A. C. Terentis, E. P. Wojcikiewicz, R. Oyola, J. Miksovska and D. Du, *Chem. Commun.*, 2015, **51**, 7000–7003.
- 38 L. Nielsen, R. Khurana, A. Coats, S. Frokjaer, J. Brange, S. Vyas, V. N. Uversky and A. L. Fink, *Biochemistry*, 2001, **40**, 6036–6046.
- 39 S. Lee, E. J. Fernandez and T. A. Good, *Protein Sci.*, 2007, **16**, 723–732.
- 40 J. Jayamani, G. Shanmugam and E. R. Azhagiya Singam, *RSC Adv.*, 2014, **4**, 62326–62336.
- 41 S.-H. Wang, X.-Y. Dong and Y. Sun, *Biochem. Eng. J.*, 2012, **63**, 38–49.
- 42 M. J. Hore, X. Ye, J. Ford, Y. Gao, J. Fei, Q. Wu, S. J. Rowan, R. J. Composto, C. B. Murray and B. Hammouda, *Nano Lett.*, 2015, **15**, 5730–5738.
- 43 E. Arad, H. Green, R. Jelinek and H. Rapaport, *J. Colloid Interface Sci.*, 2020, **573**, 87–95.
- 44 J. T. Pedersen and N. H. H. Heegaard, *Anal. Chem.*, 2013, **85**, 4215–4227.
- 45 L. Hong, X. Qi and Y. Zhang, *J. Phys. Chem. B*, 2012, **116**, 6611–6617.
- 46 A. K. Buell, *Biochem. J.*, 2019, **476**, 2677–2703.
- 47 T. C. Michaels, H. W. Lazell, P. Arosio and T. P. Knowles, *J. Chem. Phys.*, 2015, **143**, 054901.
- 48 P. Arosio, T. P. Knowles and S. Linse, *Phys. Chem. Chem. Phys.*, 2015, **17**, 7606–7618.
- 49 K. Shezad, K. Zhang, M. Hussain, H. Dong, C. He, X. Gong, X. Xie, J. Zhu and L. Shen, *Langmuir*, 2016, **32**, 8238–8244.
- 50 S. J. Cox, B. Lam, A. Prasad, H. A. Marietta, N. V. Stander, J. G. Joel, B. R. Sahoo, F. Guo, A. K. Stoddard, M. I. Ivanova and A. Ramamoorthy, *Biochemistry*, 2020, **59**, 2249–2258.
- 51 G. Gao, M. Zhang, D. Gong, R. Chen, X. Hu and T. Sun, *Nanoscale*, 2017, **9**, 4107–4113.
- 52 M. Lundqvist, J. Stigler, G. Elia, I. Lynch, T. Cedervall and K. A. Dawson, *Proc. Natl. Acad. Sci. U. S. A.*, 2008, **105**, 14265–14270.
- 53 K. Yokoyama, A. Fischer, A. R. Amori, D. R. Welchons and R. E. McKnight, *J. Biophys. Chem.*, 2010, **1**, 153–163.
- 54 W. E. Klunk, R. F. Jacob and R. P. Mason, *Anal. Biochem.*, 1999, **266**, 66–76.

