

Cite this: *Mater. Adv.*, 2021,
2, 1695

Mesoporous silica-coated gold nanorods loaded with tetrazolyl phthalocyanine as NIR light-activated nano-switches for synergistic photothermal and photodynamic inactivation of antibiotic-resistant *Escherichia coli*†

Qiu hao Ye,^a Shuang huang Xiao,^a Ting Lin,^b Yufeng Jiang,^a Yiru Peng ^{*a} and Yide Huang^{*b}

A light-controlled nano-switch was prepared by assembling mesoporous silica-coated gold nanorods with bis-(1-(4-hydroxyphenyl)-5-mercapto-tetrazolyl) silicon(iv) phthalocyanines, which exhibited excellent triple functions with controlled release of phthalocyanines and generation of reactive oxygen species (ROS) as well as temperature enhancement under laser irradiation. The nano-switch achieved an effective antimicrobial activity against a variety of antibiotic-resistant *Escherichia coli* strains through synergistic photodynamic therapy and photothermal therapy by damaging the genomic DNA and enzyme activity of bacteria.

Received 9th October 2020,
Accepted 14th January 2021

DOI: 10.1039/d0ma00782j

rsc.li/materials-advances

1. Introduction

Overuse and misuse of antibiotics by humans for a long time has led to the emergence and prevalence of antibiotic-resistant bacteria, which has reduced the therapeutic efficacy of antibiotics for human and animal pathogens.^{1–3} It is estimated that, by 2050, approximately 10 million people will die directly or indirectly from a multidrug resistant infection.⁴ Despite enormous efforts in research, the development of new antimicrobial drugs cannot catch up with the emergence of antibiotic resistant pathogens.⁵ Pursuit of alternative strategies and drugs to overcome antibiotic resistance is thus highly desirable.

In the past few decades, near-infrared (NIR) laser-induced photothermal therapy (PTT) has been used as a powerful strategy to combat cancers⁶ and bacterial infection⁷ because of its non-invasive manipulation, good controllability, and high tissue penetration. Various photothermal materials have been developed such as gold nanoparticles,⁸ gold nanorods (AuNRs),⁹ carbon nanotubes,¹⁰ two dimensional MoS₂¹¹ and

MnO₂¹² as well as graphene nanoribbons and their supermolecules.¹³ Mesoporous silica-coated gold nanorods (AuNR-SiO₂) have aroused great interest as photothermal materials because of their tenable surface plasmon resonance and excellent light-to-heat energy conversion efficiency^{14–17} as well as light controllable delivery of biomolecules into cytoplasm.¹⁸ Besides, the large specific surface area of mesoporous silica guarantees a high drug payload and optimizes the light-transparent window in the NIR region.¹⁹ Therefore, AuNR-SiO₂ seems to be a desirable candidate for highly stable and NIR laser-induced antibacterial applications. Turcheniuk *et al.* reported that AuNR-SiO₂ loaded with verteporfin could be used as an efficient near infrared nanostructure to eradicate *Escherichia coli* infection.²⁰

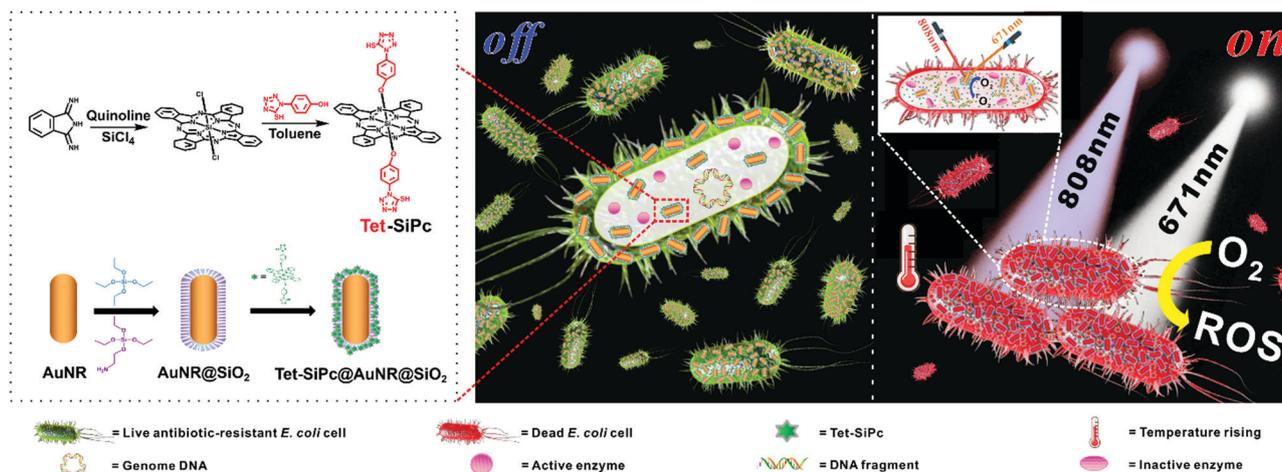
Photodynamic therapy (PDT) against microbial cells is also considered to be an alternative high-efficiency strategy to eliminate bacteria both *in vitro* and *in vivo*.^{13,21,22} The antibacterial strategy uses a specific wavelength of light to activate photosensitizers (PSS), which react with oxygen to produce reactive oxygen species (ROS) to kill bacteria. Phthalocyanines (Pcs) and metal phthalocyanines (MPcs) have been used as promising photosensitizers for PDT of cancerous and noncancerous diseases. Recently, a series of new Pcs and their nano-formulations have exhibited excellent anticancer^{23–26} and antimicrobial^{27–31} activities. However, the properties of Pcs such as easy aggregation in water, lack of ability to target specific tissue, and a limited optimal wavelength for tissue penetration hinder their applications for PDT.^{32–34} It is desired to synthesize novel Pcs to address the drawbacks of traditional Pcs.

^a College of Chemistry & Material, Fujian Provincial Key Laboratory of Advanced Materials Oriented Chemical Engineering, Fujian Province Key Laboratory of Polymer Materials, Fujian Normal University, Fuzhou, China.
E-mail: yirupeng@fjnu.edu.cn

^b Provincial University Key Laboratory of Cellular Stress Response and Metabolic Regulation, College of Life Sciences, Fujian Normal University, Fuzhou, China.
E-mail: ydhuang@fjnu.edu.cn

† Electronic supplementary information (ESI) available: Experimental details and characterization data. See DOI: 10.1039/d0ma00782j





Scheme 1 Schematic representation of the preparation of the Tet-SiPc@AuNR@SiO₂ nano-switch and the antibacterial mechanism of Tet-SiPc@AuNR@SiO₂.

In this work, we synthesized a novel silicon Pc with mercapto-tetrazolyl functional groups, named bis-(1-(4-hydroxyphenyl)-5-mercapto-tetrazolyl)silicon(IV) phthalocyanine (Tet-SiPc), and used it as a photosensitizer for antibacterial research (Scheme 1). The mercapto-tetrazolyl group, a pharmaceutical synthetic intermediate, is a main pharmacophore of antibacterial and anti-inflammatory drugs.^{35,36} Introducing mercapto-tetrazolyl functional groups to the axial position of Pc rings is expected to reduce its aggregation and improve its photodynamic antibacterial performance. To achieve the synergistic PDT and PTT, a light-controlled nano-switch was assembled through the use of mesoporous silica-coated gold nanorods with bis-(1-(4-hydroxyphenyl)-5-mercapto-tetrazolyl) silicon(IV) phthalocyanine. The effect of the nano-switch against a variety of antibiotic-resistant *E. coli* strains was evaluated.

2. Materials and methods

2.1. Materials and instruments

The organic solvents used for the preparation of the nano-switch (Tet-SiPc@AuNR@SiO₂) were of reagent grade. Sodium borohydride (NaBH₄), silver nitrate (AgNO₃), sodium hydroxide (NaOH) and hydrochloric acid (HCl) were purchased from Sino-pharm Group Chemical Reagent Co., Ltd. Hydrogen tetrachloroaurate(III) trihydrate (HAuCl₄·3H₂O) was purchased from Shanghai Bailingwei Chemical Technology Co., Ltd. Cetyltrimethylammonium bromide (CTAB) was obtained from Sigma-Aldrich (Mainland, China). 1-(4-Hydroxyphenyl)-5-mercapto-tetrazolium, tetraethyl orthosilicate (TEOS) and 3-aminopropyltriethoxysilane (APTES) were purchased from Energy Chemical. Ascorbic acid (AA) was purchased from the Xinning Chemical Plant in Shantou (Guangdong, China).

The infrared spectra (KBr pellets) were recorded on a PE-983G spectrometer. ¹H NMR spectra were recorded on a Bruker 400 MHz FT-NMR spectrometer using tetramethylsilane (TMS) as an internal standard. Mass spectra (MS) were measured

on a Bruker MALDI-TOF mass spectrometer. UV/Vis spectra were recorded on a Cary 50 UV/Vis spectrophotometer. Fluorescence emission spectra were measured on an FL900/FS920 fluorescence spectrophotometer. Raman spectra were recorded on the XploRA Plus system. The particle size distribution was analyzed using a Mastersizer 3000E laser particle size analyser. Energy dispersive X-ray spectroscopy (EDX) was performed on an FESEM-7500F scanning electron microscope (SEM). Element analysis *via* Inductively Coupled Plasma Optical Emission Spectrometry (ICP-OES) was performed on a PerkinElmer Optima 8000. A Mithras LB 940 multimode microplate reader was used to detect enzyme activity. Transmission electron microscopy (TEM) images were obtained using a JEM 1400 transmission electron microscope at an acceleration voltage of 100 kV.

2.2. Synthesis of bis-(1-(4-hydroxyphenyl)-5-mercapto-tetrazolyl)silicon(IV) phthalocyanine (Tet-SiPc)

A mixture of dichloro-phthalocyanine silicon (SiPcCl₂) (0.061 g, 0.1 mmol),³⁷ 1-(4-hydroxyphenyl)-5-mercapto-tetrazole (Tet) (0.05826 g, 0.3 mmol) and potassium carbonate (0.028 g, 0.2 mmol) in toluene (30 mL) was heated at 110 °C for 48 h. The mixture was cooled to room temperature and the solvent was removed by filtration. After evaporation under reduced pressure, the crude product was purified twice *via* alumina column chromatography using acetone and hexane (*v*:*v* = 1:5) and methanol and methylene chloride (*v*:*v* = 1:10) as eluents, respectively. The obtained product was further purified twice *via* chromatography on a silica gel column using acetone and methylene chloride (*v*:*v* = 1:10) as eluents. Tet-SiPc was obtained as a dark blue-green solid in a yield of 18%. IR ν/cm^{-1} : 736 (Ar-H), 1080 (Si-O), 1240 (C-O), 1506 (C-N); Raman ν/cm^{-1} : 681 ($\sigma_{\text{C-H}}$), 1342 (C-N), 1524 (C=N), 1612 ($\sigma_{\text{C=C}}$); ¹H NMR (400 MHz, DMSO-d₆) δ /ppm: 8.95 (2H, s, SH, D₂O exchange, H¹), 5.17 (4H, d, *J* = 8 Hz, H²), 2.28 (4H, d, *J* = 8 Hz, H³), 8.53–8.56 (8H, m, H⁴), 9.68–9.70 (8H, m, H⁵); ESI-MS calcd for *m/z* = 926.16, found: *m/z* = 925.63 [M]⁺.



2.3. Preparation of mesoporous silica-coated gold nanorods loaded with bis-(1-(4-hydroxyphenyl)-5-mercapto-tetrazolyl)silicon(IV) phthalocyanine (Tet-SiPc@AuNR@SiO₂)

AuNRs were synthesized according to the seed-mediated growth method described by Babak *et al.*³⁸ Firstly, CTAB solution (10 mL, 0.10 M) was mixed with HAuCl₄ solution (50 μL, 50 mM). Subsequently, ice-cold NaBH₄ solution (600 μL, 0.01 M) was added to the mixture, followed by stirring at 25 °C for 3 min, resulting in the formation of a brown seed solution. In order to grow the gold seeds, the seed solution was allowed to stand for 2 h. The nanorod growth solution was prepared by mixing HAuCl₄ solution (300 μL, 50 mM) and CTAB solution (30 mL, 0.10 M) with gentle stirring, then AgNO₃ solution (300 μL, 0.10 M), HCl solution (300 μL, 1.0 M) and ascorbic acid (240 μL, 0.10 M) were added in sequence to prepare the growth solution.

To grow the gold nanorods, the seed solution (75 μL) was added to the growth solution under slow stirring, and the mixture was continuously stirred at 25 °C overnight to obtain a purple-red gold nanorod mixture, then the mixture was centrifuged at 8000 rpm for 10 min and washed with ultra-pure water three times. Finally, the product, AuNRs, was dispersed in 30 mL of ultra-pure water for further use.

For preparing the mesoporous silica-coated gold nanorods,³⁹ NaOH solution (0.10 M) was added into the above prepared AuNR solution with stirring to adjust the pH of the mixed solution to 10. Next, tetraethyl orthosilicate (TEOS) (20%, 30 μL) in methanol and 10 μL of 2% 3-aminopropyltriethoxysilane (APTES) in methanol were injected into the AuNR solution three times at 30 min intervals. The mixed solution was stirred for 24 h at 25 °C to obtain AuNR@SiO₂. Finally, the synthesized AuNR@SiO₂ was collected *via* centrifugation at 8000 rpm for 5 min and washed three times with ultra-pure water to remove CTAB. The product was dispersed in 30 mL of ultra-pure water for further use. The concentrations of gold in AuNRs and AuNR@SiO₂ were determined by using element analysis *via* Inductively Coupled Plasma Optical Emission Spectrometry (ICP-OES).

To prepare Tet-SiPc@AuNR@SiO₂, Tet-SiPc DMSO solution (50 μL, 1 mM) was added to the AuNR@SiO₂ solution (10 mL), the gold concentration of which was 200 μg mL⁻¹ as determined *via* ICP-OES. The mixture was stirred at room temperature for 48 h. The obtained product solution was centrifuged at 8000 rpm for 5 min, followed by consecutive washing three times with ultra-pure water, and Tet-SiPc@AuNR@SiO₂ was then dispersed in 10 mL of ultrapure water for further use.

2.4. Fluorescence quantum yields of Tet-SiPc

Fluorescence quantum yields (Φ_F) of Tet-SiPc in DMSO were determined by using the comparative method of Eq. 1:⁴⁰

$$\Phi_F = \Phi_{F(\text{std})} \cdot F \cdot A_{\text{Std}} \cdot n^2 / (F_{\text{Std}} \cdot A \cdot n_{\text{Std}}^2) \quad (1)$$

where the integral areas of the emission curves of Tet-SiPc and unsubstituted ZnPc (*n*-ZnPc) are denoted by *F* and *F*_{Std}, respectively. The absorbance of Tet-SiPc and standard *n*-ZnPc at the

excitation wavelength are represented by *A* and *A*_{Std}, respectively. The refractive indices of Tet-SiPc and standard *n*-ZnPc solvents are represented by *n*² and *n*_{Std}², respectively. *n*-ZnPc in DMSO ($\Phi_{F(\text{std})} = 0.20$) was employed as the standard.⁴¹

2.5. Singlet oxygen quantum yields of Tet-SiPc

Singlet oxygen quantum yields (Φ_A) of Tet-SiPc were measured by the chemical trapping method based on the singlet oxygen quencher 1,3-diphenylisobenzofuran (DPBF).⁴² Tet-SiPc (3 mL, 3×10^{-6} M) and DPBF (6×10^{-5} M) were mixed using DMSO as the solvent, and the mixture was continuously irradiated with a laser (671 nm, 100 mW cm⁻²). The decrease of DPBF absorbance at 417 nm was detected from the UV-Vis spectra. Using *n*-ZnPc as the reference ($\Phi_A = 0.67$ for *n*-ZnPc in DMSO),⁴³ the Φ_A value of Tet-SiPc was calculated using Eq. 2:

$$\Phi_A = \Phi_{A\text{Std}} \cdot \frac{R}{R_{\text{Std}}} \cdot \frac{I_{\text{Abs}}^{\text{Std}}}{I_{\text{Abs}}} \quad (2)$$

2.6. Singlet oxygen quantum yields of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂

Singlet oxygen was monitored *via* chemical oxidation of 9,10-anthracenediyl-bis(methylene)dimalonic acid (ABDA) (3×10^{-4} M) in the presence of AuNRs, AuNR@SiO₂ or Tet-SiPc@AuNR@SiO₂, the gold concentration of which was found to be 100 μg mL⁻¹ *via* ICP-OES.²⁰ The decrease in ABDA absorbance at 378 nm was monitored upon laser irradiation (671 nm, 100 mW cm⁻²). The irradiation was stopped every 3 min and UV-Vis absorption spectra were recorded.

2.7. The loading rate of Tet-SiPc on AuNR@SiO₂

The loading rate of Tet-SiPc on AuNR@SiO₂ was measured according to the method described by Chen *et al.*¹⁰ The loading rate of Tet-SiPc on AuNR@SiO₂ was calculated according to eqn (3):

$$f = \frac{M_{\text{Tet-SiPc}} - M_{\text{Tet-SiPc}'}}{M_{\text{Tet-SiPc@AuNR@SiO}_2}} \times 100\% \quad (3)$$

where *f* is the loading rate of Tet-SiPc on AuNR@SiO₂, *M*_{Tet-SiPc} is the initial amount of Tet-SiPc, *M*_{Tet-SiPc'} is the amount of Tet-SiPc in solution and *M*_{Tet-SiPc@AuNR@SiO₂} is the amount of Tet-SiPc@AuNR@SiO₂ added.

2.8. The photo-induced release of Tet-SiPc from Tet-SiPc@AuNR@SiO₂

The photo-induced release of Tet-SiPc from Tet-SiPc@AuNR@SiO₂ was studied *via* fluorescence spectroscopy.¹⁰ In brief, 3 mL of Tet-SiPc@AuNR@SiO₂ solution with a 100 μg mL⁻¹ gold concentration was added to seven centrifuge tubes, respectively. Each centrifuge tube was irradiated with an infrared laser (808 nm, 0.5 W cm⁻²) and a thermocouple thermometer was used to record the temperature of the Tet-SiPc@AuNR@SiO₂ solution every one minute. AuNR@SiO₂, AuNRs and ultra-pure water were used as controls. At the same time, the Tet-SiPc released from Tet-SiPc@AuNR@SiO₂ was



monitored *via* fluorescence spectroscopy. Tet-SiPc@AuNR@SiO₂ solution was irradiated to release Tet-SiPc which was insoluble in solution at 808 nm (0.5 W cm⁻²). After the solutions were irradiated for 0, 5, 10, 15, 20 and 25 min. Tet-SiPc is insoluble in the solution, the precipitation of Tet-SiPc was centrifuged and re-dissolved in DMSO. The fluorescence of the Tet-SiPc solution obtained above was measured and the concentration of Tet-SiPc was calculated.

2.9. Construction of antibiotic-resistant *E. coli* DH5 α strains and expression of β -galactosidase in *E. coli* DH5 α cells

E. coli DH5 α cells are not resistant to Ampicillin, Kanamycin and Zeocin. The Ampicillin-, Kanamycin- or Zeocin-resistant *E. coli* DH5 α strain could be created by transforming a plasmid containing a corresponding antibiotic-resistant gene. In this study, pUC18, pET28a and pPICZa plasmids were used to transform *E. coli* DH5 α . pUC18, pET28a and pPICZa contain an Ampicillin-, a Kanamycin- and a Zeocin-resistant gene, respectively. After successful transformation, *E. coli* DH5 α will gain the ability to resist Ampicillin, Kanamycin or Zeocin. The LB plates containing Ampicillin (100 μ g mL⁻¹), Kanamycin (50 μ g mL⁻¹) or Zeocin (25 μ g mL⁻¹) were used to confirm the ability of the transformed *E. coli* DH5 α to resist the corresponding antibiotics.

β -Galactosidase is an important enzyme to metabolize the lactose in *E. coli*, but the *LacZ* gene encoding β -galactosidase is mutated in *E. coli* DH5 α , which causes the loss of the β -galactosidase activity. To regain the activity of β -galactosidase in *E. coli* DH5 α , a recombinant plasmid expressing the functional β -galactosidase was constructed by using the pAO815 plasmid as the backbone. The pAO815 was linearized *via* digestion with the *EcoR* I restriction enzyme. The *LacZ* gene with full length was amplified using the plasmid pAd/CMV/V5-GW/*lacZ* as a template and a pair of specific primers (forward primer: 5'CCGGAATTCACCATGATAGATCCCGTCG 3', reverse primer: 5'CCGGAATTCTATTTTGTACACCAGACCAACTG 3') flanked by an *EcoR* I site at both 5' and 3' ends *via* PCR. The PCR product was purified by using the Promega Wizard SV Gel and PCR Clean-up System (Promega (Beijing) Biotech Co., Ltd. Beijing, China). The purified PCR product was digested using *EcoR* I and then ligated with the linearized pAO815 plasmid using T4 DNA ligase. The recombinant plasmid was confirmed by DNA sequencing and transformed into *E. coli* DH5 α . The expression of β -galactosidase was verified on the LB plate containing 5-bromo-4-chloro-3-indolyl β -D-galactoside (X-Gal), a chromogenic substrate for β -galactosidase.

2.10. Antibacterial activity of Tet-SiPc@AuNR@SiO₂ against *E. coli* DH5 α and antibiotic-resistant *E. coli* DH5 α strains

E. coli DH5 α or antibiotic-resistant strains were cultured in the LB (Luria-Bertani) medium at 37 °C overnight until OD_{600nm} reached about 0.5. The culture was then diluted 1:10 in the LB medium. 40 μ L of Tet-SiPc@AuNR@SiO₂ (C_{Au} = 100 μ g mL⁻¹) in sterile distilled water was added into 20 μ L of bacterial dilution. The suspension was illuminated using 808 nm laser light at 0.5 W cm⁻² for 5 min and then irradiated using a 671 nm laser

for 10 min with a power density of 100 mW cm⁻². After the combined irradiation at 808 nm and 671 nm, a series of 10-fold dilutions of the bacteria were performed with the LB medium, and 5 μ L of each dilution was dropped on LB agar plates and incubated at 37 °C for 12–16 h until the colonies appeared. For the colony forming unit (CFU) assay or relative survival rate analysis, an aliquot (200 μ L) of the above diluted bacterial suspension was plated on a 10 cm LB agar plate. The plates were incubated at 37 °C for 12–16 h until the colonies appeared, and then the total number of colonies on each plate was counted. The number of CFU mL⁻¹ was calculated as the number of colonies counted on a plate/0.2 mL and the dilution factor. The cell survival rate was calculated *via* normalization with respect to the CFU value of the control group performed without the treatment of Tet-SiPc@AuNR@SiO₂. Each experiment was repeated three times.

2.11. Effect of Tet-SiPc@AuNR@SiO₂ on the genomic DNA damage of *E. coli* DH5 α

The genomic DNA of *E. coli* DH5 α with or without the treatment of Tet-SiPc@AuNR@SiO₂ under combined laser irradiation at 808 nm and 671 nm as mentioned above was extracted by using the genomic DNA extraction kit (Tiangen (Beijing) Biotech Co., Ltd Beijing, China) and quantified using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific, Wilmington, DE, USA). 500 μ g of genomic DNA was loaded into wells of a 1% agarose gel, and then electrophoresis was performed at 100 V for 30 min. The gel was stained with ethidium bromide and photographed.

2.12. Effect of Tet-SiPc@AuNR@SiO₂ on the enzyme activity of β -galactosidase

The engineered *E. coli* DH5 α cells expressing the functional β -galactosidase were treated with or without Tet-SiPc@AuNR@SiO₂ under combined laser irradiation at 808 nm and 671 nm as mentioned above. The bacterial cells were collected *via* centrifugation at 10 000 rpm for 5 min. After discarding the supernatant, the pellets were resuspended with Z-buffer (Na₂HPO₄ 60 mM, NaH₂PO₄ 40 mM, KCl 10 mM, MgSO₄ 1 mM, pH = 7.0) with β -mercaptoethanol (adding 0.27 mL β -mercaptoethanol to 100 mL of Z-buffer), and then 3 drops of chloroform and 2 drops of 0.1% sodium dodecyl sulfonate (SDS) were added, followed by vortexing at 3000 rpm for 10 s, and this procedure was repeated three times. After incubating the bacterial solution at 28 °C for 5 min, 200 μ L of 2-nitrophenyl- β -D-galactopyranoside (ONPG) (4 mg mL⁻¹ in Z-buffer) was added and incubated at 30 °C until a yellow colour developed. The elapsed time was recorded in minutes. After the yellow colour developed, 500 μ L of 1% Na₂CO₃ solution was added. The reaction tubes were centrifuged for 10 min at 14 000 rpm to pellet cell debris, and the supernatants were carefully transferred into a 96-well plate. Finally, the developed yellow colour was measured at 420 nm on a spectrophotometer. The enzyme activity of β -galactosidase was calculated according to eqn (4) and the enzyme activity of control (no treatment with



Tet-SiPc@AuNR@SiO₂) was normalized to 100%:

$$\text{Enzyme activity} = \frac{\text{OD}_{420 \text{ nm}}}{\text{OD}_{600 \text{ nm}} \times V \times t} \quad (4)$$

where V is the volume of the bacterial solution (mL) and t is the reaction time (min).

3. Results and discussion

3.1. Synthesis and characterization of Tet-SiPc

Tet-SiPc was obtained *via* a nucleophilic substitution reaction between 1-(4-hydroxyphenyl)-5-mercapto-tetrazole and dichloro-silicon phthalocyanine (SiPcCl₂) using anhydrous K₂CO₃ as the catalyst in a yield of 18% (Scheme 1). The chemical structure of Tet-SiPc was characterized by various methods including ¹H NMR, ESI-MS, FT-IR and Raman spectroscopy (Fig. S1–S4, ESI[†]). The UV/Vis spectra of Tet-SiPc in dimethyl sulfoxide (DMSO) showed typical spectra of Pc with a B band at 355 nm and a Q band at 683 nm (Fig. 1a). With the increase of the concentration of Tet-SiPc, the intensity of the Q band was enhanced, while the shape and position of the Q band did not change, indicating that the introduction of mercapto-tetrazole functional groups could reduce the aggregation of Pcs. Upon excitation at 615 nm, Tet-SiPc showed emission at 677 nm (Fig. 1b). The fluorescence lifetime of Tet-SiPc was found to be 5.23 ns, and the fluorescence quantum yield (Φ_F) was calculated to be 0.0522 (Table S1 and Fig. S5, ESI[†]). The quantum yield of singlet oxygen (¹O₂) (Φ_A) was found to be 0.355 (Table S1 and Fig. S6, ESI[†]).

3.2. Preparation and characterization of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂

In order to enhance the antibacterial efficiency of Tet-SiPc and reduce its phototoxicity, AuNR-SiO₂ was used as a nanocarrier to load Tet-SiPc to form a novel light control nano-switch (Tet-SiPc@AuNR-SiO₂). The Tet-SiPc@AuNR-SiO₂ was prepared through the adsorption of Tet-SiPc on AuNR-SiO₂. Briefly,

AuNRs were synthesized using the seed-mediated growth method.³⁸ AuNR@SiO₂, a mesoporous silica layer wrapped on the surface of AuNRs, was prepared by an improved method.³⁹ Tet-SiPc (1 mM) was added to the aqueous solution of AuNR@SiO₂ (200 μg mL⁻¹) and stirred for 48 h. After being centrifuged and dialyzed, Tet-SiPc@AuNR@SiO₂ was obtained (Scheme 1). The TEM images showed that AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ exhibited uniform size and good dispersion in solution (Fig. 2a–c). The average length and width of AuNRs were about 59 nm and 16 nm, respectively, and the aspect ratio (AR) was calculated to be about 3.70. The thickness of the initial silica coating was measured to be about 22 nm for AuNR@SiO₂. After the Tet-SiPc was successfully loaded into the mesoporous silica of AuNR@SiO₂, the colour of the silica coating layer became darker due to the contribution of the high electron density of Tet-SiPc. The dynamic light scattering (DLS) size of AuNRs was found to be 31 nm. After the mesoporous silica was coated on AuNRs, the DLS size of AuNR@SiO₂ increased from 31 nm to 67 nm. With further loading of Tet-SiPc on AuNR@SiO₂, the DLS size of Tet-SiPc@AuNR@SiO₂ was increased to 106 nm (Fig. 2d–f). The average particle sizes measured *via* DLS were larger than those measured *via* TEM, which could be related to the swelling of nanoparticles in solution.⁴⁴ Energy dispersive X-ray spectroscopy (EDX) was used to demonstrate the successful loading of Tet-SiPc on AuNR@SiO₂. The content of N element in AuNR@SiO₂ was significantly higher than that in AuNRs, and the element of Si appeared at AuNR@SiO₂, indicating the successful coating of SiO₂ on AuNRs (Fig. S7 and S8, Table S2 and S3, ESI[†]). The presence of S element in Tet-SiPc@AuNR@SiO₂ proved the successful loading of Tet-SiPc on Tet-SiPc@AuNR@SiO₂ (Fig. S9, Table S4, ESI[†]). The successful preparation of Tet-SiPc@AuNR@SiO₂ was also confirmed by the UV/Vis absorption spectra (Fig. 2g). The AuNRs exhibited two surface plasmon resonance absorption peaks. The lateral surface plasmon resonance absorption peak was located at 514 nm and the longitudinal surface plasmon resonance absorption peak was found to be at 801 nm. After the

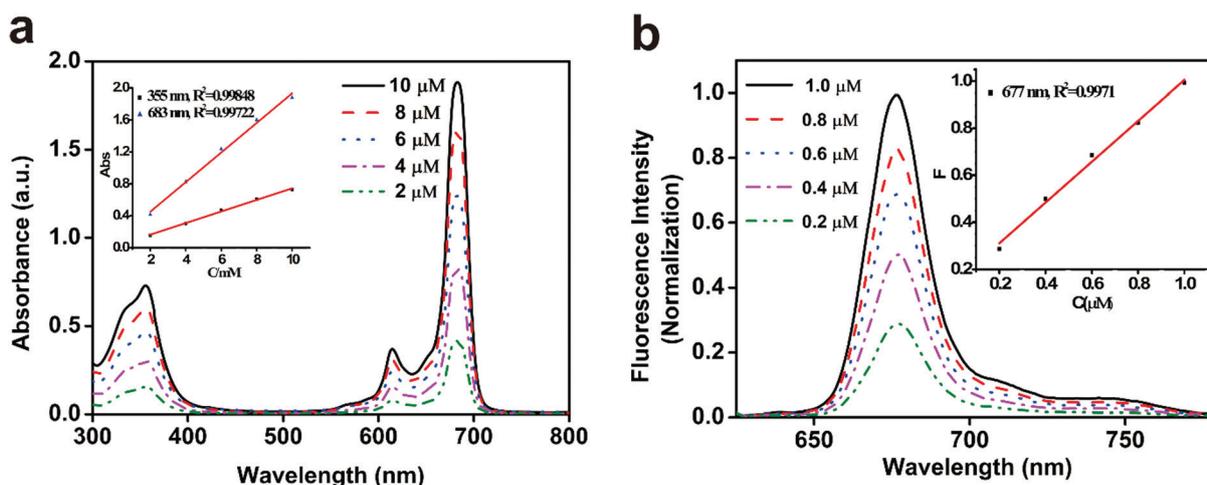


Fig. 1 (a) UV-visible spectra of Tet-SiPc in DMSO; (b) fluorescence spectra of Tet-SiPc in DMSO ($\lambda_{\text{ex}} = 615 \text{ nm}$).



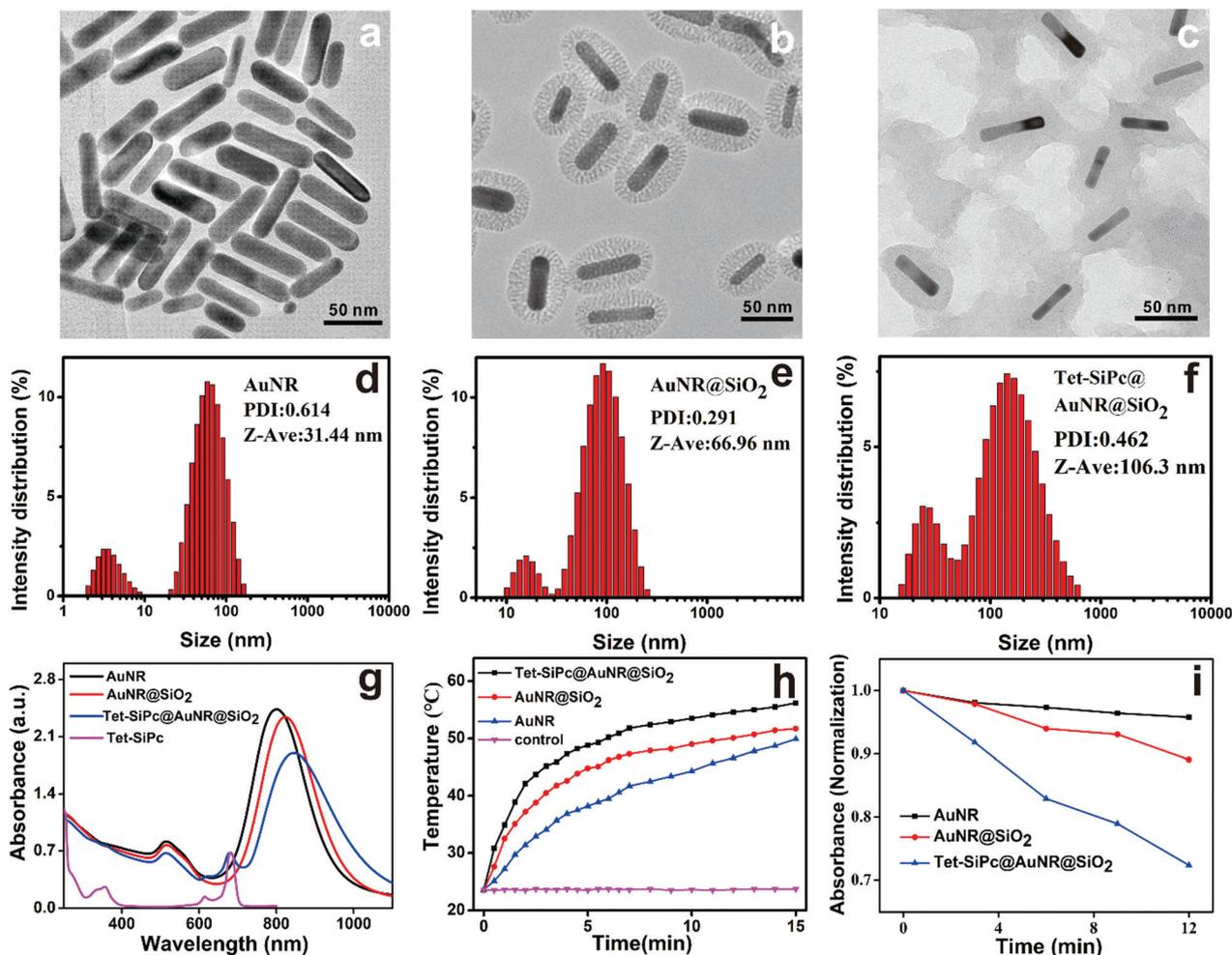


Fig. 2 TEM images of (a) AuNRs, (b) AuNR@SiO₂ and (c) Tet-SiPc@AuNR@SiO₂; particle size distribution of (d) AuNRs, (e) AuNR@SiO₂ and (f) Tet-SiPc@AuNR@SiO₂ in water; (g) UV-Vis absorption spectra of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ ($C_{Au} = 6.5 \mu\text{g mL}^{-1}$); (h) curves of temperature change of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ solutions under laser irradiation (808 nm laser with a power density of 0.5 W cm^{-2} , PBS solution as a control; the gold concentration in AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ was $100 \mu\text{g mL}^{-1}$); (i) curves of the absorption change of ABDA mixed with AuNRs, AuNR@SiO₂ or Tet-SiPc@AuNR@SiO₂ for different irradiation times (671 nm laser with a power density of 100 mW cm^{-2}). The gold concentration of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ was $100 \mu\text{g mL}^{-1}$.

AuNRs were coated with mesoporous silica, the shape and position of the lateral characteristic absorption peak at 514 nm were unchanged, while the longitudinal surface plasmon resonance was red-shifted from 801 nm to 824 nm. In the absorption spectrum of Tet-SiPc@AuNR@SiO₂, a characteristic absorption peak of Tet-SiPc was observed at 683 nm and the longitudinal surface plasmon resonance absorption peak of AuNRs continued to redshift to 845 nm, indicating that Tet-SiPc was successfully loaded in AuNR@SiO₂. The loading ratio of Tet-SiPc in Tet-SiPc@AuNR@SiO₂ was calculated to be 48% using UV-Vis absorption spectra. The photothermal effect of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ irradiated using an 808 nm laser with a power density of 0.5 W cm^{-2} for 15 min was shown in Fig. 2h. AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ exhibited excellent photothermal properties. The temperature of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ increased to 49.9, 51.7 and 56.2 °C after 15 min illumination, respectively. Tet-SiPc@AuNR@SiO₂ possessed the

highest photothermal conversion efficiency, which is probably due to the synergistic photothermal effect of Tet-SiPc loaded on the mesoporous silica and AuNR@SiO₂.⁴⁵

The ¹O₂ generation ability of AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ was evaluated by using 9,10-anthracenediyl-bis(methylene)dimalonic acid (ABDA) as a probe.²⁰ The decrease of the ABDA absorption at 378 nm as a function of irradiation time was observed upon irradiation at 671 nm (Fig. S10–S12, ESI†). The Tet-SiPc@AuNR@SiO₂ exhibited the highest ability to produce ¹O₂, followed by AuNR@SiO₂ and AuNRs (Fig. 2i). The best ¹O₂ generation ability of Tet-SiPc@AuNR@SiO₂ may be related to the synergistic generation of ¹O₂ by both Tet-SiPc and AuNR@SiO₂ in the nanosystem.⁴⁶

3.3. Light-controlled Tet-SiPc release from the nano-switch Tet-SiPc@AuNR@SiO₂

Most of the theranostic agents were “always-on” models for therapeutic intervention, leading to a low signal-to-noise ratio



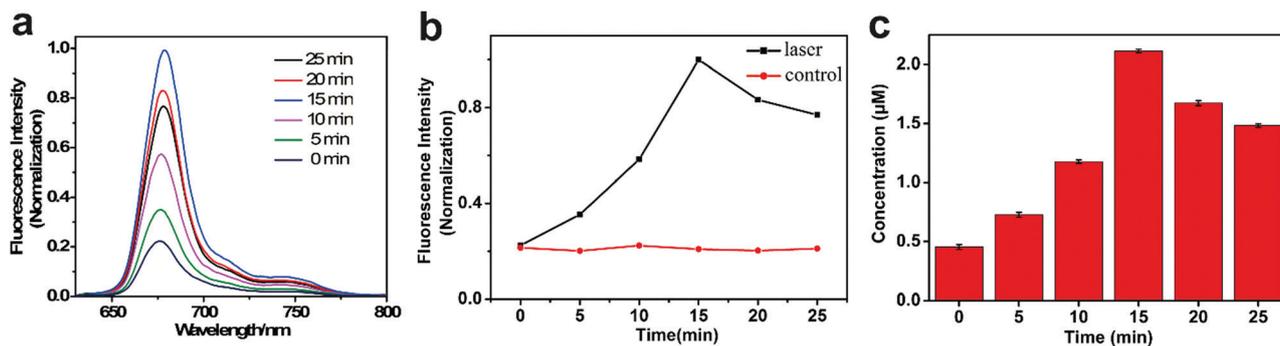


Fig. 3 (a) Fluorescence spectra of Tet-SiPc@AuNR@SiO₂ and (b) fluorescence intensity change curves of released Tet-SiPc at 677 nm as a function of illumination time when Tet-SiPc@AuNR@SiO₂ solution with a 100 μg mL⁻¹ gold concentration was irradiated using an 808 nm laser with a power density of 0.5 W cm⁻²; (c) quantitative plot of the equivalent concentration of Tet-SiPc released from Tet-SiPc@AuNR@SiO₂ under different illumination times. The error bar is shown as the standard deviation from three experiments.

and microbial drug resistance. The nanomaterials for controlled release can effectively overcome these deficiencies.

In order to confirm that Tet-SiPc@AuNR@SiO₂ is an excellent light-controlled nano-switch, a series of experiments were carried out. The fluorescence of Tet-SiPc in Tet-SiPc@AuNR@SiO₂ was quenched by AuNR@SiO₂ without irradiation. Upon irradiation with NIR light, Tet-SiPc was released from Tet-SiPc@AuNR@SiO₂

(Fig. 3a). The amount of Tet-SiPc released from Tet-SiPc@AuNR@SiO₂ was quantified using fluorescence spectra (Fig. 3b and c). The photothermal effect of Tet-SiPc@AuNR@SiO₂ could change the exothermic adsorption equilibrium and then promoted the release of Tet-SiPc. But the irradiation time was over 15 min, and a decrease in the fluorescence intensity of Tet-SiPc was observed, which may be caused by the photobleaching of

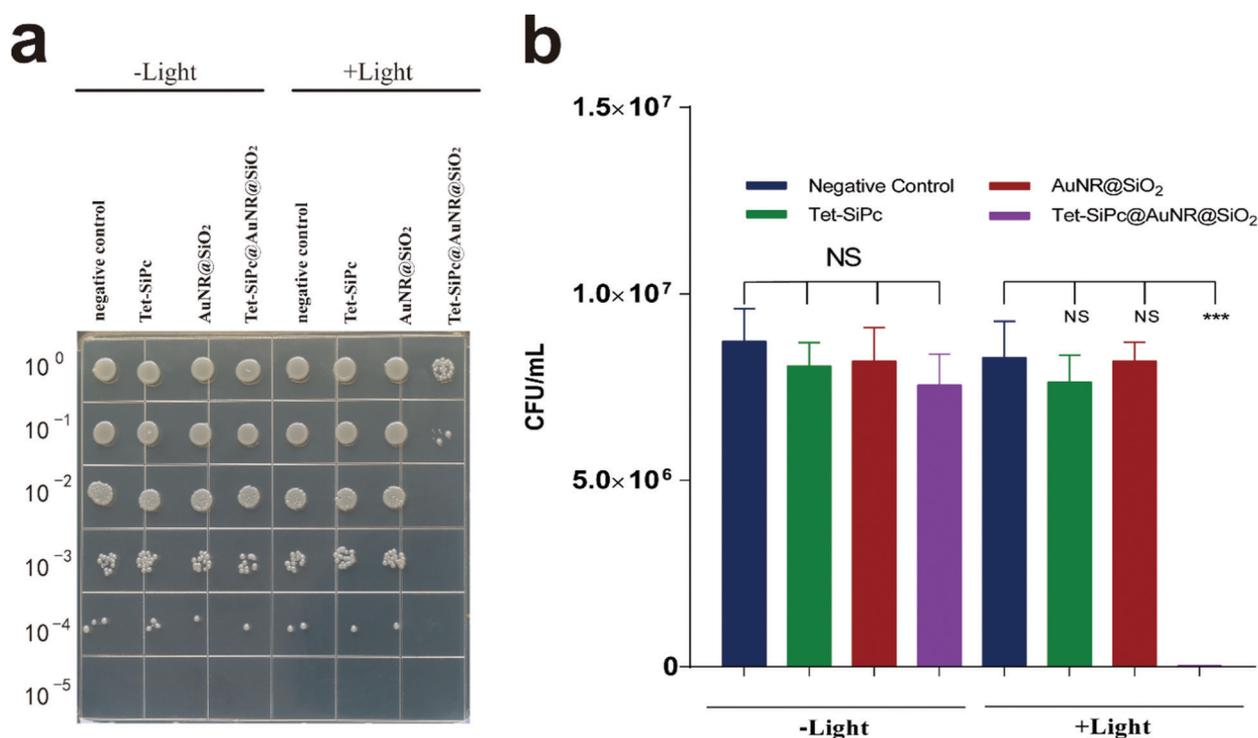


Fig. 4 Antibacterial activity of Tet-SiPc, AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ against *E. coli* DH5 α strain. (a) Colony assay was performed on *E. coli* DH5 α treated with Tet-SiPc, AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ dissolved in water or 10% DMSO aqueous solution. The concentration of Tet-SiPc was 50 μM, the Tet-SiPc concentration in Tet-SiPc@AuNR@SiO₂ was 50 μM and the gold concentration in AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ was 100 μg mL⁻¹. After laser irradiation (808 nm laser light at 0.5 W cm⁻² for 5 min, followed by 671 nm laser irradiation for 10 min with a power density of 100 mW cm⁻²), *E. coli* DH5 α solutions were diluted in 10-fold serial dilutions and 5 μL of the diluted solution was dropped on the LB plate; (b) colony forming units (CFU) of *E. coli* DH5 α treated with Tet-SiPc, AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ dissolved in water or 10% DMSO aqueous solution. The concentration of Tet-SiPc was 50 μM, the Tet-SiPc concentration in Tet-SiPc@AuNR@SiO₂ was 50 μM and the gold concentration in AuNRs, AuNR@SiO₂ and Tet-SiPc@AuNR@SiO₂ was 100 μg mL⁻¹. The data are expressed as means \pm SD of three experiments. NS means no statistical significance, ****P* < 0.001.



Tet-SiPc by irradiation or the aggregation behaviour of Tet-SiPc released into water. Light can manipulate very precisely to release the Tet-SiPc in the Tet-SiPc@AuNR@SiO₂, which provides a strategy to prevent the bacteria from developing drug resistance by long exposure to the Tet-SiPc@AuNR@SiO₂ in antibacterial application.

3.4. Antibacterial activity of the nano-switch Tet-SiPc@AuNR@SiO₂ against *E. coli* DH5 α

E. coli DH5 α was selected as a model to evaluate the antibacterial activity of Tet-SiPc@AuNR@SiO₂. Upon irradiation, both Tet-SiPc and AuNR@SiO₂ did not show obvious antibacterial efficacies against DH5 α , indicating that the photodynamic efficacy of Tet-SiPc or the photothermal efficacy of AuNR@SiO₂ alone did not present enough antibacterial activity against DH5 α .

The Tet-SiPc@AuNR@SiO₂ exhibited a significant antibacterial efficacy with a killing rate of 99.83% (Fig. 4), which can be explained by the fact that AuNR@SiO₂ served as a photothermal agent to absorb the energy of the NIR laser and convert it into heat energy, which triggered the release of Tet-SiPc from Tet-SiPc@AuNR@SiO₂ and promoted the released Tet-SiPc to produce ¹O₂ to kill the bacteria through the synergistic photodynamic and photothermal effects.⁴⁷

3.5. Antibacterial activity of the nano-switch Tet-SiPc@AuNR@SiO₂ against antibiotic-resistant *E. coli* DH5 α strains

We wondered whether the Tet-SiPc@AuNR@SiO₂ exhibited the same effect in killing the antibiotic-resistant bacteria as the non-resistant bacteria. Three antibiotic-resistant DH5 α strains,

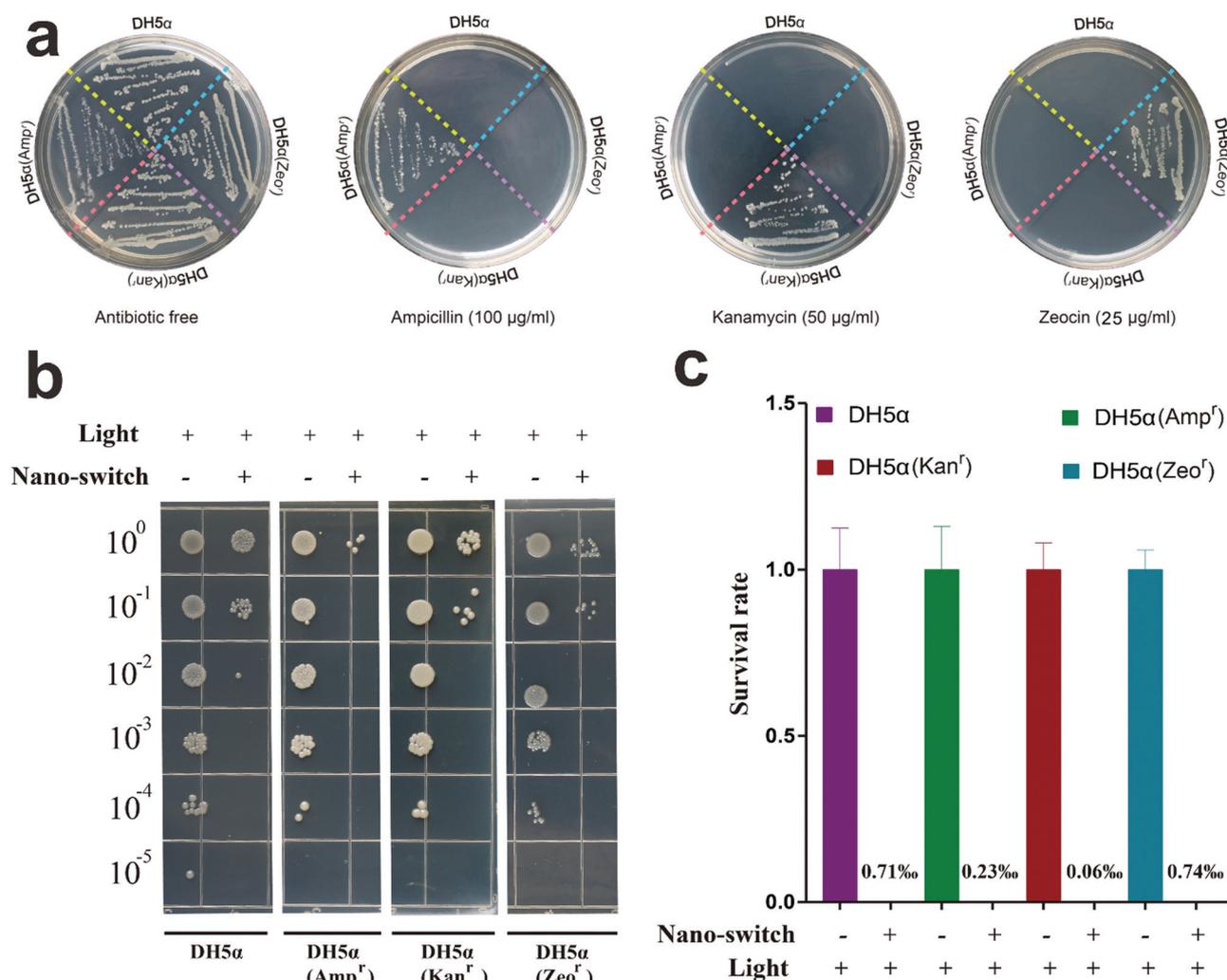


Fig. 5 Antibacterial activity of Tet-SiPc@AuNR@SiO₂ against three antibiotic-resistant *E. coli* DH5 α strains under laser irradiation (808 nm laser light at 0.5 W cm⁻² for 5 min, followed by 671 nm laser irradiation for 10 min with a power density of 100 mW cm⁻²). (a) Verification of Ampicillin-, Kanamycin- and Zeocin-resistant *E. coli* DH5 α strains on plates with the corresponding antibiotics; (b) analysis of the antibacterial activity of Tet-SiPc@AuNR@SiO₂ against three antibiotic-resistant *E. coli* DH5 α strains by the colony assay. The Tet-SiPc concentration in Tet-SiPc@AuNR@SiO₂ was 50 μ M and the gold concentration in Tet-SiPc@AuNR@SiO₂ was 100 μ g mL⁻¹; (c) analysis of the survival rates of three antibiotic-resistant *E. coli* DH5 α strains after the treatment with Tet-SiPc@AuNR@SiO₂ under laser irradiation. The Tet-SiPc concentration in Tet-SiPc@AuNR@SiO₂ was 50 μ M and the gold concentration in Tet-SiPc@AuNR@SiO₂ was 100 μ g mL⁻¹. The data are expressed as means \pm SD of three experiments.



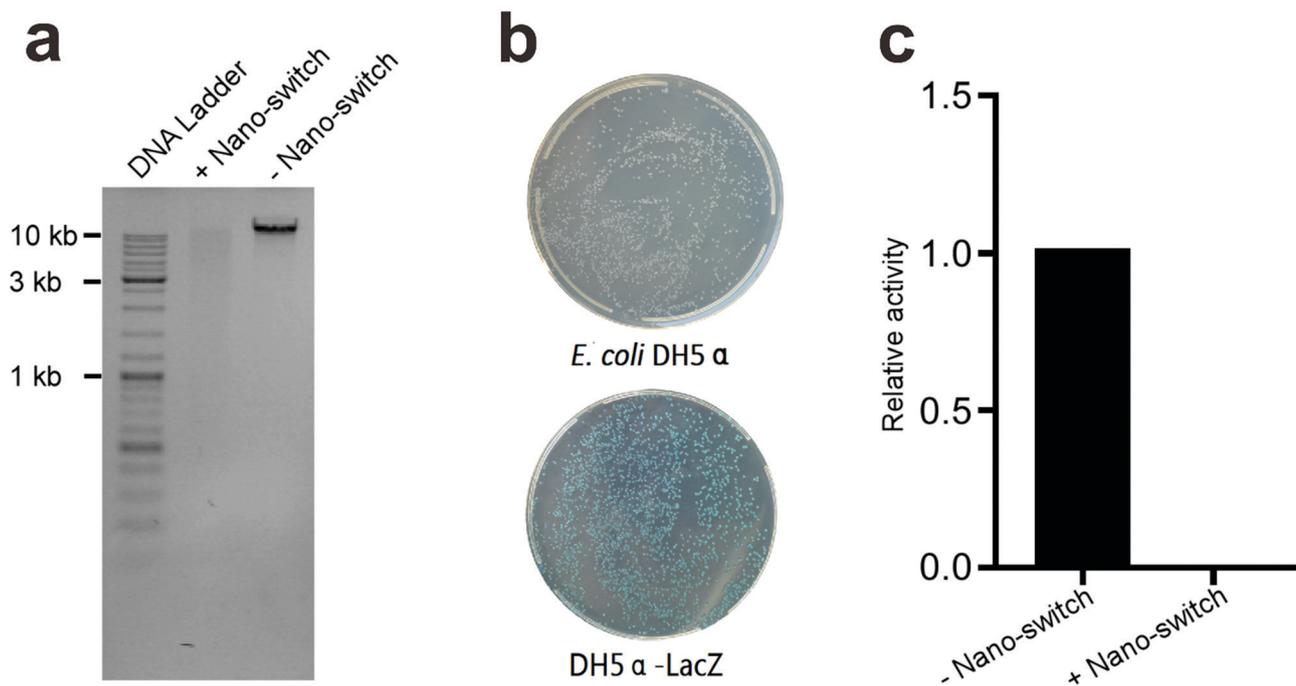


Fig. 6 Tet-SiPc@AuNR@SiO₂ impaired genomic DNA and enzymes of DH5 α cells. (a) The genomic DNA of DH5 α was degraded after the treatment with Tet-SiPc@AuNR@SiO₂ (the Tet-SiPc concentration in Tet-SiPc@AuNR@SiO₂ was 50 μ M and the gold concentration in Tet-SiPc@AuNR@SiO₂ was 100 μ g mL⁻¹) under laser irradiation (808 nm laser at 0.5 W cm⁻² for 5 min, followed by 671 nm laser irradiation for 10 min with a power density of 100 mW cm⁻²); (b) the DH5 α -LacZ strain successfully expressed the reporter β -galactosidase; (c) analysis of β -galactosidase activity with or without treatment with the Tet-SiPc@AuNR@SiO₂ (the Tet-SiPc concentration in Tet-SiPc@AuNR@SiO₂ was 50 μ M and the gold concentration in Tet-SiPc@AuNR@SiO₂ was 100 μ g mL⁻¹) under laser irradiation (808 nm laser light at 0.5 W cm⁻² for 5 min, followed by 671 nm laser irradiation for 10 min with a power density of 100 mW cm⁻²).

DH5 α (Amp^r), DH5 α (Kan^r) and DH5 α (Zeo^r), were obtained through the transformation of plasmids pUC18, pET28a and pPICZa into DH5 α cells, respectively. Their ability to resist antibiotics was confirmed on the plates containing the corresponding antibiotics (Fig. 5a). The antibacterial activity of Tet-SiPc@AuNR@SiO₂ against the antibiotic-resistant DH5 α was evaluated. The results showed that the Tet-SiPc@AuNR@SiO₂ also exhibited strong antibacterial activity against all DH5 α (Amp^r), DH5 α (Kan^r) and DH5 α (Zeo^r) strains (Fig. 5b and c).

3.6. Antibacterial mechanism of the nano-switch Tet-SiPc@AuNR@SiO₂

DNA and enzymes are the most important biological macromolecules in bacterial cells. DNA stores the genetic information of bacteria and controls the metabolism of bacteria.⁴⁸ Enzymes catalyse nearly all the chemical reactions in cells and are also essential for cell survival. We suspected that Tet-SiPc@AuNR@SiO₂ impaired DNA and enzymes in cells under laser light, which caused the death of cells. The genomic DNA of DH5 α treated with or without Tet-SiPc@AuNR@SiO₂ was isolated and analysed *via* agarose gel electrophoresis. The result was shown in Fig. 6a. The genomic DNA treated with Tet-SiPc@AuNR@SiO₂ upon irradiation showed a smear band in the agarose gel, suggesting that DNA was fragmented.

The β -galactosidase encoded by the *LacZ* gene is a favourable reporter for the quantitative analysis of enzymatic activity in microorganisms. In this study, β -galactosidase was used as a

reporter to evaluate the enzymatic damage of DH5 α treated with Tet-SiPc@AuNR@SiO₂ under laser irradiation. β -galactosidase activity is deficient in DH5 α because of the Δ M15 mutation of the *LacZ* gene. In order to restore the β -galactosidase activity, we constructed a plasmid expressing the *LacZ* gene and transformed it into DH5 α , named DH5 α -LacZ. The DH5 α -LacZ strain showed blue colonies in the plate containing X-Gal, indicating that the β -galactosidase activity was regained in DH5 α -LacZ (Fig. 6b). The β -galactosidase activity of the DH5 α -LacZ strain treated with or without Tet-SiPc@AuNR@SiO₂ was analysed. The result showed that the β -galactosidase activity of DH5 α -LacZ was totally undetectable after the treatment of Tet-SiPc@AuNR@SiO₂ under laser irradiation (Fig. 6c).

The possible antibacterial mechanism is that the Tet-SiPc@AuNR@SiO₂ more efficiently generated heat and ROS upon irradiation. The enzymes and DNA in cells are denatured at high temperatures, and these denatured macromolecules are more vulnerable to impair by ROS. The synergistic photothermal and photodynamic effect gives the Tet-SiPc@AuNR@SiO₂ a more effective ability to kill *E. coli* cells.

4. Conclusions

In this work, we successfully constructed a near-infrared light-controlled nano-switch Tet-SiPc@AuNR@SiO₂. This nano-switch was assembled through adsorption of Tet-SiPc in the mesoporous silica layer of AuNR@SiO₂. Tet-SiPc@AuNR@SiO₂



realized precisely controlled release of Tet-SiPc from AuNR@SiO₂ and the generation of ROS as well as excellent photo-thermal conversion efficiency through simple light irradiation and exhibited a synergistic photothermal and photodynamic effects in killing both *E. coli* and antibiotic-resistant *E. coli* strains. The degradation of genomic DNA and the loss of enzyme activity in *E. coli* cells after the treatment with Tet-SiPc@AuNR@SiO₂ under irradiation could be the main causes of bacteria killing.

Author contributions

Qiu hao Ye: methodology, data curation, formal analysis, writing – review & editing. Shuang huang Xiao: investigation, writing – original draft. Ting Lin: investigation. Yufeng Jiang: investigation, methodology, formal analysis. Yiru Peng: conceptualization, methodology, project administration, supervision, writing – review & editing, funding acquisition. Yide Huang: methodology, supervision, writing – review & editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by the National Key Basic Research Program of China (973 project) (2015CB35200), the National Natural Science Foundation of China (21274021), the Natural Science Foundation of Fujian (2019Y0007), the Joint Funds of Fujian Provincial Health and Education Research (2019-WJ-23), the Scientific Research Innovation Team Construction Programme of Fujian Normal University (IRTL 1702), the scientific research innovation program “Xiyuanjiang River Scholarship” of College of Life Sciences, Fujian Normal University and the Special Funds of the Central Government Guiding Local Science and Technology Development (2020L3008).

Notes and references

- M. Bassetti, G. Poulakou, E. Ruppe, E. Bouza, S. J. Van Hal and A. Brink, *Intensive Care Med.*, 2017, **43**, 1464–1475.
- J. M. Stokes, K. Yang, K. Swanson, W. Jin, A. Cubillos-Ruiz, N. M. Donghia, C. R. MacNair, S. French, L. A. Carfrae, Z. Bloom-Ackerman, V. M. Tran, A. Chiappino-Pepe, A. H. Badran, I. W. Andrews, E. J. Chory, G. M. Church, E. D. Brown, T. S. Jaakkola, R. Barzilay and J. J. Collins, *Cell*, 2020, **180**, 688–702.
- T. Wi, M. M. Lahra, F. Ndowa, M. Bala, J. R. Dillon, P. Ramon-Pardo, S. R. Eremin, G. Bolan and M. Unemo, *PLoS Med.*, 2017, **14**, e1002344.
- M. E. A. D. Kraker, A. J. Stewardson and S. Harbarth, *PLoS Med.*, 2016, **13**, e1002184.
- E. Tacconelli, E. Carrara, A. Savoldi, S. Harbarth, M. Mendelson, D. L. Monnet, C. Pulcini and G. Kahlmeter, *Lancet Infect. Dis.*, 2018, **18**, 318–327.
- T. Guo, Y. Lin, G. Jin, R. Weng, J. Song, X. Liu, G. Huang, L. Hou and H. Yang, *Chem. Commun.*, 2019, **55**, 850–853.
- N. Yang, C. Wang, X. Wang and L. Li, *Nanotechnology*, 2018, **29**, 175601.
- W.-Y. Chen, H.-Y. Chang, J.-K. Lu, Y.-C. Huang, S. G. Harroun, Y.-T. Tseng, Y.-J. Li, C.-C. Huang and H.-T. Chang, *Adv. Funct. Mater.*, 2015, **25**, 7189–7199.
- M. L. Xu, L. Y. Guan, S. K. Li, L. Chen and Z. Chen, *Chem. Commun.*, 2019, **55**, 5359–5362.
- X. Chen, S. Wu, D. Ma, J. Chen, Q. Guo, X. Han, K. Chen, H. Yang, Y. Huang and Y. Peng, *Chem. Commun.*, 2018, **54**, 13279–13282.
- X. Yang, J. Li, T. Liang, C. Ma, Y. Zhang, H. Chen, N. Hanagata, H. Su and M. Xu, *Nanoscale*, 2014, **6**, 10126–10133.
- W. Zeng, H. Zhang, Y. Deng, A. Jiang, X. Bao, M. Guo, Z. Li, M. Wu, X. Ji, X. Zeng and L. Mei, *Chem. Eng. J.*, 2020, **389**, 124494.
- Z. H. Yu, X. Li, F. Xu, X. L. Hu, J. Yan, N. Kwon, G. R. Chen, T. Tang, X. Dong, Y. Mai, D. Chen, J. Yoon, X. P. He and H. Tian, *Angew. Chem., Int. Ed.*, 2020, **59**, 3658–3664.
- X. Chen, Q. Zhang, J. Li, M. Yang, N. Zhao and F. J. Xu, *ACS Nano*, 2018, **12**, 5646–5656.
- Y. Wang, Q. Cui, X. Zhao, T. Qin, W. Wang, H. Sun, H. Zhu, H. Guo and H. Sun, *RSC Adv.*, 2018, **8**, 41454–41463.
- C. Li, Y. Zhang, Z. Li, E. Mei, J. Lin, F. Li, C. Chen, X. Qing, L. Hou, L. Xiong, H. Hao, Y. Yang and P. Huang, *Adv. Mater.*, 2018, **30**, 1706150.
- Y. Liu, P. Bhattarai, Z. Dai and X. Chen, *Chem. Soc. Rev.*, 2019, **48**, 2053–2108.
- J. Liu, C. Detrembleur, M. C. De Pauw-Gillet, S. Mornet, C. Jerome and E. Duguet, *Small*, 2015, **11**, 2323–2332.
- X. Cui, W. Cheng and X. Han, *J. Mater. Chem. B*, 2018, **6**, 8078–8084.
- K. Turcheniuk, V. Turcheniuk, C. H. Hage, T. Dumych, R. Bilyy, J. Bouckaert, L. Heliot, V. Zaitsev, R. Boukherroub and S. Szunerits, *Chem. Commun.*, 2015, **51**, 16365–16368.
- J. Sun, Y. Zhang, J. Su, T. Dai, J. Chen, L. Zhang, H. Wang, W. Liu, M. Huang and Z. Chen, *Dyes Pigm.*, 2020, **179**, 108392.
- W. Liu, Y. Zhang, W. You, J. Su, S. Yu, T. Dai, Y. Huang, X. Chen, X. Song and Z. Chen, *Nanoscale*, 2020, **12**, 13948–13957.
- E. Dube, D. O. Oluwole, N. Nwaji and T. Nyokong, *Spectrochim. Acta, Part A*, 2018, **203**, 85–95.
- P. Garcia Calavia, I. Chambrier, M. J. Cook, A. H. Haines, R. A. Field and D. A. Russell, *J. Colloid Interface Sci.*, 2018, **512**, 249–259.
- L. Lamch, J. Kulbacka, M. Dubinska-Magiera, J. Saczko and K. A. Wilk, *Photodiagn. Photodyn. Ther.*, 2019, **25**, 480–491.
- Z. Wang, T. Jia, Q. Sun, Y. Kuang, B. Liu, M. Xu, H. Zhu, F. He, S. Gai and P. Yang, *Biomaterials*, 2020, **228**, 119569.
- X. Li, D. Lee, J. D. Huang and J. Yoon, *Angew. Chem., Int. Ed.*, 2018, **57**, 9885–9890.



- 28 E. Lee, X. Li, J. Oh, N. Kwon, G. Kim, D. Kim and J. Yoon, *Chem. Sci.*, 2020, **11**, 5735–5739.
- 29 H. H. Mohamed, I. Hammami, S. Akhtar and T. E. Youssef, *Composites, Part B*, 2019, **176**, 107314.
- 30 A. Galstyan, A. Ricker, H. Nüsse, J. Klingauf and U. Dobrindt, *ACS Appl. Bio Mater.*, 2019, **3**, 400–411.
- 31 Y. Jia, J. Li, J. Chen, P. Hu, L. Jiang, X. Chen, M. Huang, Z. Chen and P. Xu, *ACS Appl. Mater. Interfaces*, 2018, **10**, 15369–15380.
- 32 K. Kuninobu, A. Tadaaki, M. Noriyuki, H. Makoto and S. Tamotsu, *Chem. Lett.*, 2002, 966–967.
- 33 N. Nishiyama, Y. Nakagishi, Y. Morimoto, P. S. Lai, K. Miyazaki, K. Urano, S. Horie, M. Kumagai, S. Fukushima, Y. Cheng, W. D. Jang, M. Kikuchi and K. Kataoka, *J. Controlled Release*, 2009, **133**, 245–251.
- 34 S. Bhana, R. O'Connor, J. Johnson, J. D. Ziebarth, L. Henderson and X. Huang, *J. Colloid Interface Sci.*, 2016, **469**, 8–16.
- 35 S. J. Wittenberger, *Org. Prep. Proc. Int.*, 1994, **26**, 499–531.
- 36 S. Ganapaty, P. Ramalingam and C. B. Rao, *Indian J. Heterocycl. Chem.*, 2007, **16**, 283–286.
- 37 T. I. Bruce, N. Diel, K. F. Schoch, T. J. Marks, J. W. Lyding and C. R. Kannewurf, *J. Am. Chem. Soc.*, 1983, **105**, 1551–1567.
- 38 N. Babak and A. E. Mostafa, *Chem. Mater.*, 2003, **15**, 1957–1962.
- 39 I. Gorelikov and N. Matsuura, *Nano Lett.*, 2008, **8**, 369–373.
- 40 X. H. Peng, S. F. Chen, B. Y. Zheng, B. D. Zheng, Q. F. Zheng, X. S. Li, M. R. Ke and J. D. Huang, *Tetrahedron*, 2017, **73**, 378–384.
- 41 X.-F. Zhang and H.-J. Xu, *J. Chem. Soc., Faraday Trans.*, 1993, **89**, 3347–3351.
- 42 N. Nwahara, J. Britton and T. Nyokong, *J. Coord. Chem.*, 2017, **70**, 1601–1616.
- 43 N. A. Kuznetsova, N. S. Gretsova and E. A. Kalmykova, *Russ. J. Gen. Chem.*, 2000, **70**, 133–140.
- 44 J. Geng, Z. Zhu, W. Qin, L. Ma, Y. Hu, G. G. Gurzadyan, B. Z. Tang and B. Liu, *Nanoscale*, 2014, **6**, 939–945.
- 45 J. Peng, L. Zhao, X. Zhu, Y. Sun, W. Feng, Y. Gao, L. Wang and F. Li, *Biomaterials*, 2013, **34**, 7905–7912.
- 46 N. T. Chen, K. C. Tang, M. F. Chung, S. H. Cheng, C. M. Huang, C. H. Chu, P. T. Chou, J. S. Souris, C. T. Chen, C. Y. Mou and L. W. Lo, *Theranostics*, 2014, **4**, 798–807.
- 47 J. Wang, Y. Liu, Y. Ma, C. Sun, W. Tao, Y. Wang, X. Yang and J. Wang, *Adv. Funct. Mater.*, 2016, **26**, 7516–7525.
- 48 N. Jimenez-Garrido, L. Perello, R. Ortiz, G. Alzuet, M. Gonzalez-Alvarez, E. Canton, M. Liu-Gonzalez, S. Garcia-Granda and M. Perez-Priede, *J. Inorg. Biochem.*, 2005, **99**, 677–689.

