



Cite this: *Environ. Sci.: Atmos.*, 2021, **1**, 498

## Chemical functionality at the liquid surface of pure unsaturated fatty acids†

Aisling C. Stewart, Martin J. Paterson and Stuart J. Greaves \*

The C=C double bonds in unsaturated fatty acids found within atmospheric aerosols play an important role in tropospheric pollution cycles. An understanding of the affinity of such double bonds for the surface of these aerosols is thus vital to the accurate modelling of reactions occurring at these interfaces. To simulate the structures of the liquid surfaces of a series of C18 fatty acids with increasing degrees of unsaturation molecular dynamics calculations have been carried out using the GROMACS suite of programs, employing the general Amber force field. The relative surface coverage of HC=CH, carboxylic acid, methyl and CH<sub>2</sub> groups has been probed and found to be significantly different from that predicted based on a purely statistical distribution of groups or assumed based on X-ray diffraction data. HC=CH, methyl, and CH<sub>2</sub> groups dominate the surface, with the methyl groups being found disproportionately at the surface relative to their bulk concentration. The HC=CH surface exposure does not increase linearly as the acids become more unsaturated. The distinct structure found at the interface extends only a few nanometers into the sample, suggesting that surface order only affects the first layer of molecules at the interface, but is key to the exposure of highly reactive chemical functional groups to atmospheric oxidants.

Received 2nd June 2021  
Accepted 23rd August 2021

DOI: 10.1039/d1ea00043h  
rsc.li/esatmospheres

### Environmental significance

Unsaturated fatty acids are key components of organic atmospheric aerosols, with oleic acid also being used as a mimic for aerosol droplets in both experimental and computational work. The presence of alkene groups within aerosols plays an important role in the chemistry of these aerosols; the oxidation of unsaturated groups leads to the formation of secondary aerosols. Models of aerosol structure and behaviour often assume that the distribution of functional groups at surfaces will be stoichiometric. This molecular dynamics study of the liquid interfaces of fatty acids highlights the enhanced surface presence of particular functional groups as compared to bulk liquid samples. The implications of this surface preference on the reactivity of atmospheric aerosols are discussed.

### Introduction

The gas–liquid interface is of central importance to atmospheric chemistry as well as a wide range of other fields, both within industry and the natural world. Studies of reactions at, and the transfer of species across, boundaries between gases and liquids form the basis of our understanding of the reactivity of atmospheric aerosols.<sup>1–8</sup> Reactions at the gas–liquid interface have, however, been less extensively studied than those occurring in either the bulk gas or bulk liquid phases.<sup>7–9</sup>

The importance of aerosols as systems of high chemical reactivity within the atmosphere has been well documented.<sup>6,8,10–12</sup> The surfaces of aerosols provide sites for the adsorption and reaction of gaseous species, which can drive atmospheric processes such as those involved in the evolution of greenhouse gases and other pollutants<sup>13–15</sup> and in the destruction of Antarctic ozone.<sup>16–18</sup> In addition to this aerosols

are in themselves important atmospheric pollutants, with enhanced concentrations known to have a detrimental effect on human health.<sup>15,19,20</sup> Aerosols also have an important role to play as cloud condensation nuclei and for direct and indirect radiative forcing.<sup>12,18,21–23</sup> The exact effect of aerosols within atmospheric systems depends on their composition and structure. This is in turn affected by the reactions of these aerosols with atmospheric radicals, which can oxidise the organic components of primary aerosols, emitted from both natural and anthropogenic sources, and secondary organic aerosols (SOAs).<sup>18,24</sup>

Long chain unsaturated fatty acids, such as oleic acid, are chemically significant components of atmospheric aerosols,<sup>25–31</sup> and have been used in laboratory studies of proxies for aerosols.<sup>32–35</sup> The presence of carbon–carbon double bonds within such molecules is of particular importance to the chemistry that occurs at the surface of these aerosols. Alkene groups can be oxidised by radicals, such as odd oxygen and odd hydrogen species, chemically ageing the aerosols and generating SOAs.<sup>27,30,33,35–38</sup> This altering of the chemical nature of aerosols naturally brings with it a change in their activities in chemical processes within the atmosphere, and being able to model the

Institute of Chemical Sciences, Heriot-Watt University, Edinburgh EH14 4AS, UK.  
E-mail: s.j.greaves@hw.ac.uk

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d1ea00043h



nature and rates of formation of secondary aerosols is thus of great interest to atmospheric chemists.

As a component in marine, arboreal and anthropogenic aerosols, the oleic acid family has also previously been used in experiments on model systems for aerosols.<sup>30,33,35,39–41</sup> Given the highly reactive nature of C=C groups,<sup>39,42–44</sup> it is of particular interest to carry out comparative studies using oleic acid and other unbranched C18 fatty acids, as such studies allow the investigation of the importance of degree of unsaturation on the rate of reactions without this being significantly affected by differences in mass or steric bulk. Such experiments have shown that the uptake of OH and Cl<sub>2</sub> by a wetted surface increases linearly (up to  $n = 3$ ) with the number of double bonds in the hydrocarbon chain of the fatty acid present.<sup>42,43</sup> The importance of carbon–carbon double bonds at the surface of a liquid is thus central to determining the rate at which oxidation can occur. It is therefore of particular relevance to atmospheric chemists to gain an insight into the preference of alkene groups for the surface, as compared to other groups within fatty acids, such as the carboxylic acid group and alkane groups. Studies of model systems for aerosols, however, sometimes make assumptions about fatty acids on the surfaces of these based on work on bulk liquid samples<sup>45,46</sup> or solid salts of these acids,<sup>47</sup> without explicit knowledge about the distribution and orientation of fatty acid functional groups at the gas–liquid interface. Interfaces are, however, known to be energetically, and as result often chemically, distinct from the bulk of a system<sup>7,8,48</sup> and an understanding of the surface preferences of different molecules and functional groups is vital in order to improve the modelling of surfaces, including those of atmospheric relevance.

While interactions at the gas–liquid interface show many similar features to those at the gas–solid interface,<sup>7,49,50</sup> the disordered nature of liquids often makes their surface structure complex, meaning predictions of the nature and distribution of functional groups at these interfaces can be hard to predict using simple models.<sup>7,51</sup> Molecular dynamics (MD) simulations are a popular way of modelling the interactions of systems containing large numbers of atoms. Indeed, a range of hydrocarbon and ionic liquid surfaces have already been studied using this method.<sup>52–56</sup>

This paper presents the findings of MD simulations using the general Amber force field (GAFF),<sup>57</sup> an all-atom force field, to simulate the structure of surfaces of four members of the oleic

acid family with increasing degrees of saturation: oleic acid, linoleic acid, linolenic acid and stearidonic acid (Fig. 1). The concentration of different functional groups at the surface of the liquid, as compared to the bulk, is discussed. Simulations have been carried out at 273 K, 298 K and 333 K, to investigate the effect of increasing energy on the surface preferences of the functional groups within these molecules.

## Methods

Simulations were carried out using the GROMACS software package (Version 2018.2)<sup>58–63</sup> and employing GAFF for all calculations. This force field had been chosen as it had all the necessary parameters to describe the systems studied and was better at recreating experimentally measurable parameters for the bulk systems (further information on force field selection is available in the ESI S1†). All-atom structure files for oleic, linoleic, linolenic and stearidonic acids were obtained from the Automatic Topology Builder database (ATB, Version 3.0)<sup>64–66</sup> and converted to input files for GROMACS using the Antechamber program of AmberTools.<sup>67</sup> For each of the target systems a box of dimensions 6.5 by 6.5 by 13.0 nm was filled with molecules, by inserting them in a random fashion, up to a total of 600. The procedure for determining the optimal size of the simulation box is described in Section S2 of the ESI.†

The resulting samples of pure fatty acids underwent 100–200 cycles of energy minimization using the steepest-descent algorithm, until the energy of the system had converged. A constant volume (NVT) equilibration stage was then carried out for 5 ns at 298 K using a velocity Verlet algorithm, with a time step of 1 fs. Following on from this constant pressure (NPT) equilibration was carried out in two stages. The first stage was carried out at 298 K and employed a leap-frog algorithm combined with a modified Berendsen thermostat with a time constant of 0.1 ps for the temperature coupling.<sup>68</sup> This was carried out at a time step of 1 fs until the total density of the system had stabilised, with this typically requiring a total length of 5–10 ns. The next pressure equilibration stage was carried out at the temperature of interest using the velocity Verlet algorithm<sup>69</sup> and Martyna–Tuckerman–Tobias–Klein pressure coupling.<sup>70</sup> This equilibration stage was also carried out for between 5 and 10 ns, until the total density of the system had stabilised (for final bulk density values see Table S1 of the ESI S1†).

Having equilibrated a box of a bulk sample of the mixture this was converted into a ‘slab’, by extending the simulation by a factor of 3 along the direction of the longest side of the box (the ‘z’ direction). This generated interfaces between the liquid and the regions of vacuum above and below it. The slab of molecules was twice as thick as it was wide with volumes of vacuum above and below the slab that each had the same dimensions as the slab itself (see ESI Section S3†). The number of molecules used within the simulation was selected based on prior tests of the interface *versus* bulk properties of slabs of different thicknesses (see ESI Section S2†). The size of the slab was chosen so as to ensure that this was thick enough for there to be a clear distinction between the surface and the bulk while

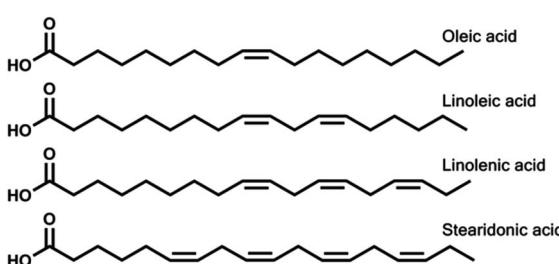


Fig. 1 The structures of the four all-*cis* fatty acids investigated within this study.



taking into account the higher computational cost of simulating larger numbers of molecules.

The slab was then subjected to a series of temperature equilibration and annealing cycles, which proceeded as follows. Temperature equilibration was carried out at the temperature of interest. As with the previous temperature equilibration process a velocity Verlet algorithm was used, with a time step of 1 fs. This equilibration process was carried out in 5 ns steps until the distribution of functional groups throughout the slab was found to be the same for two consecutive steps. Typically it took a total of 15–20 ns for this to be the case. The slab was then annealed in order to destroy any preferential positioning of the molecules with respect to either each other or to the surface. This was achieved by carrying out an annealing cycle at a higher temperature, typically around 600 K for a short length of time ( $\approx 20$  ps). The exact temperature and length of time were chosen on a slab-to-slab basis, as longer times and higher temperatures had a greater effectiveness at destroying the internal structure of the slab, but also risked causing large numbers of molecules to evaporate off the surface of the slab. The distribution of molecules throughout the slab was checked at the end of the annealing step in order to ensure that this had been successful. Following annealing the slab was cooled back to the temperature of interest and then underwent a further temperature equilibration cycle. This was carried out as before, until two of the consecutive 5 ns equilibration steps gave the same distribution of functional groups throughout the slab. Annealing and equilibration cycles repeated until the distributions reached by the slabs at the end of two equilibration stages were the same, thus indicating that an equilibrium distribution had been reached.

Following this equilibration process two ‘production runs’ were carried out, with the slab undergoing two temperature equilibration runs for 20 ns at the temperature of interest, with a final annealing step, as described above, in between the two runs. All calculations employed 3D periodic boundary conditions and particle-mesh Ewald long-range electrostatics. A timeline summarizing the stages in the generation, equilibration and production runs of the slabs is shown in Fig. S4 of the ESI.†

## Results and discussion

The structures of the four members of the oleic acid family studied are shown in Fig. 1, with the number of double bonds increasing in the order oleic acid, linoleic acid, linolenic acid, stearidonic acid. The oleic acid family was chosen as oleic acid has commonly been used as a model compound for organic aerosol component, and each of the other acids chosen has the same number of carbon atoms and differs only by the number of double bonds within it. Each of these acids has only *cis*-C=C double bonds, as is this is more common for naturally occurring fatty acids.<sup>71</sup> However it should be noted that increasing numbers of *cis*-double bonds leads to molecules that are more bent in nature, and how this affects the simulations is discussed later in the paper and in the ESI S5.† Simulations of slabs made from pure samples of each of these molecules were carried out

at 273 K, 298 K and 333 K. Each of the slabs consisted of 600 molecules of the acid of interest, the interface had a square cross section and the slab was twice as thick as it was wide. On either side of the slab were regions of vacuum, which each had the same dimensions as the slab itself.

Fig. 2 presents MD snapshots looking down onto the final surfaces of slabs of the four acids at 298 K, as shown using the Visual Molecular Dynamics (VMD, Version 1.9.3) software.<sup>72</sup> These were taken at the end of the final 20 ns production runs and show typical equilibrated positions of the molecules with relation to the surface. Colour-coding has been used to highlight the distributions of the functional groups present at the surface. It can be seen that the surface coverage of carboxylic acid groups (blue) is very low in all of these slabs. Visual comparisons of the HC=CH and CH<sub>2</sub> distributions across the different species are, however, made more complicated, as on moving from oleic acid to stearidonic acid the degree of unsaturation, and therefore the ratio of alkene groups to CH<sub>2</sub> groups, increases. As a result the surface of oleic acid seems dominated by CH<sub>2</sub> groups (green) and that of stearidonic acid by HC=CH groups (black), but it is not clear by inspection whether this is purely based on a statistical distribution of these differing numbers of groups, or if there is an underlying preference for a particular moiety to be found at the interface.

Information about the surface area coverage of different groups can be obtained in a more quantitative manner by the use of solvent accessible surface area (SASA) analysis,<sup>73,74</sup> included as gmx sasa in the GROMACS software package. This calculates the total surface area of the sample that would be accessible to a solvent molecule of a given size, with this being simulated as a hard sphere probe of a chosen radius approaching from the vacuum side of the interface. This probe records the atoms that it comes into contact with and the total surface area contact that it has with each of these. This then

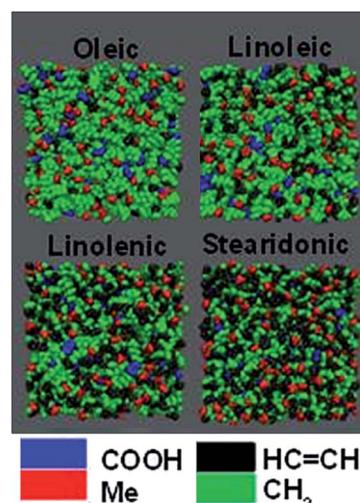


Fig. 2 Top-down view of the surfaces of slabs of each of the fatty acids studied, at 298 K. Different functional groups are colour-coded, with methyl groups in red, CH<sub>2</sub> groups in green, alkene groups in black and carboxylic acid groups in blue. The same colouring scheme is used throughout (see main text for details).



allows information to be gained about both the number of atoms of each functional group at the surface and the total surface area coverage by this functional group.

Fig. 3 shows the results of carrying out SASA analysis on the four samples, as measured by a probe of radius 0.15 nm. This corresponds to the van der Waals radius of an OH radical,<sup>75</sup> one of the most common atmospherically relevant species that are likely to react with aerosols<sup>75–77</sup> (for a discussion of how changing the probe size affects the outcome of the SASA analysis see ESI S6†). For each slab, analysis was carried out at 100 ps intervals over the course of the two 20 ns production runs, following the methodology of McKendrick and coworkers.<sup>54,78</sup> The graphs shown in Fig. 3 represent the averaged values across

each of these intervals and production runs. Additional details of the analysis procedure are given in ESI Sections S6–S8.† Carrying out analysis at 100 ps intervals allowed the surface to be observed over time, giving evidence that the slabs were well equilibrated during the production runs, as the distributions of the groups did not vary significantly over the course of these. The time interval was chosen so as to ensure that enough frames had been taken so as to be representative of the entire run, without sampling the same configuration multiple times (see ESI, Section S7† for details). Analysis was only carried out at times  $>2.5$  ns, to ensure that the atoms had fully relaxed to their equilibrium positions at the temperature of interest prior to probing. When counting those atoms that were considered to be present at the surface a minimum surface area of contact was required in order for the atoms of interest to be counted, with this threshold being set in order to avoid spurious signals from the bulk (ESI S8† describes how this threshold was chosen). The absolute values for Fig. 3 are shown in Tables S3–S5 of the ESI (Section S9†) along with their corresponding error values, which are typically around 3%.

The graphs in Fig. 3 show the SASA coverage of different functional groups within each of the acid samples. The data is ordered so that the degree of unsaturation of the acids increases on moving from left to right across each of the plots. These plots represent the surface area of each of the groups that can come into contact with probe that is similar in size to an OH radical, and thus are related to the chance of those groups reacting in the atmosphere. It should be noted that the probe used in SASA calculations is inert, and thus the analysis here cannot account for any changes to the surface structure caused the polarising abilities of the incoming species, however such interactions are beyond the scope of MD simulations and as such we present here a description of the native structure of the interface.

It can be seen from these graphs that there is only a very small surface area coverage from the COOH groups, with significantly larger amounts coming from the methyl and CH<sub>2</sub> groups. The comparison between the COOH and methyl group coverages is particularly interesting, as in each of the molecules there are the same number of methyl and acid groups (one) and these are present at the termini of the molecules, so if the chains were orientated at random, one would expect a similar surface area coverage for the two groups. Indeed, as the van der Waals radius of the COOH group is greater than that of a methyl group, a higher surface area coverage of COOH would be predicted based purely on statistical coverage.

This observed non-statistical distribution of groups at the surface is likely to be based on the energetic preferences of different groups for the surface or the bulk. Positioning the non-polar methyl groups at the surface, with the polar acid groups oriented inwards increases the strength of the interactions within the sample as a whole. Polar groups are able to form stronger intermolecular interactions both with each other and with other non-polar molecules *via* induction effects. Orienting COOH groups so that they are directed outwards towards an area of vacuum would mean that these could be involved in fewer interactions with other molecules and it is thus favourable to have the methyl groups at the surface, with

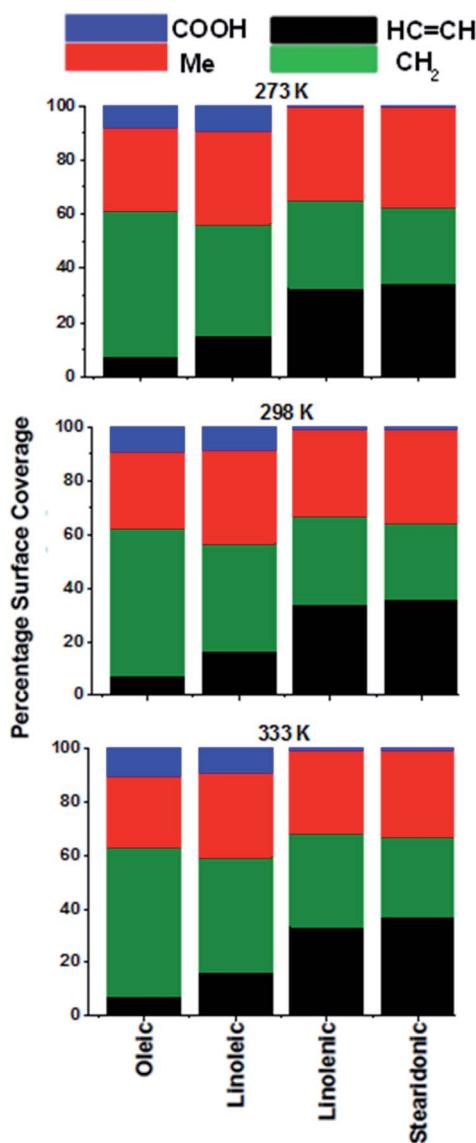


Fig. 3 SASA results showing the fractional surface area coverage of each of the functional groups studied for the oleic acids samples at each of the simulated temperatures, as measured using a probe of radius 0.15 nm. Each bar represents the averaged results from two consecutive 20 ns production runs, separated by an annealing step. Within each run only data from times  $>2.5$  ns was used.



a corresponding below-statistical presence of each of the other groups at the surface. This effect would be expected to be more prominent at lower temperatures, as higher slab energies would mean that intermolecular interactions within the slab would become less important, and surfaces with more equal coverages of different groups, and thus higher entropies, more favourable. The surface coverage of groups presented in Fig. 3, however, seems to be temperature independent, and thus it appears that this effect is not significant over the temperature range studied.

The surface area coverage of  $\text{HC}=\text{CH}$  groups within an aerosol is of interest to atmospheric chemists, as reactions involving these groups play a particularly significant role in the conversion of primary to secondary aerosols. It can be seen in Fig. 3 that the surface presence of the alkene groups varies significantly from acid to acid; this is to be expected, as the degree of unsaturation increases on moving across the series of the oleic acids. Dividing the data for each functional group by the number of those functional groups in the molecule allows for a comparison of trends between the different acids. Fig. 4 shows the results of this; here the percentage coverages from Fig. 3 have been divided by the number of functional group units present within the acid of interest, before the results were renormalized to 100.

It can be seen from Fig. 4 that the surface preference of  $\text{COOH}$  is higher in oleic and linoleic acid than in their more saturated counterparts: there is hardly any  $\text{COOH}$  present at the surface of the more unsaturated acids. The bars for the alkene group coverage are a similar height for each of the acids, suggesting that the coverage of these groups at the surface increases roughly in proportion with their number in the samples as a whole, although surface  $\text{HC}=\text{CH}$  coverage is always below statistical, as a result of the enhanced  $\text{CH}_3$  presence there.

For each of the acids there is a much lower than expected proportional  $\text{CH}_2$  presence at the surface. This can already be seen in Fig. 3 – although there are more  $\text{CH}_2$  than any other group in each of the samples, the surface area coverage of these is not overwhelming, with the surface area coverage of  $\text{CH}_2$  and  $\text{CH}_3$  often similar, although each of the acid molecules has only one  $\text{CH}_3$  and between 8 and 14  $\text{CH}_2$  groups depending on the species. Fig. 4 shows this contrast more starkly, by dividing the surface area of the  $\text{CH}_2$  groups by the presence of these in each molecule. It can be seen that there is a much lower than expected  $\text{CH}_2$  presence at the surface. This suggests that there is a preference for the termini of the molecules to be present at, or even protruding from, the interfaces, a result previously seen in united atom MD studies of alkanes.<sup>53,79</sup>

An additional method to investigate ordering of molecules relative to the interface is to calculate the concentrations of functional groups at different distances from the centre of the slab ( $z = 0$ ). The gmx density function of GROMACS was used for this. This splits the slab along the  $z$  direction into equal slices of a given thickness and calculates the densities of the functional groups of interest within these. Plots of distance from the centre of the slab *versus* concentration for each of the functional groups are shown in Fig. 5. Here each slab was split into 100 slices, with this number being a compromise between

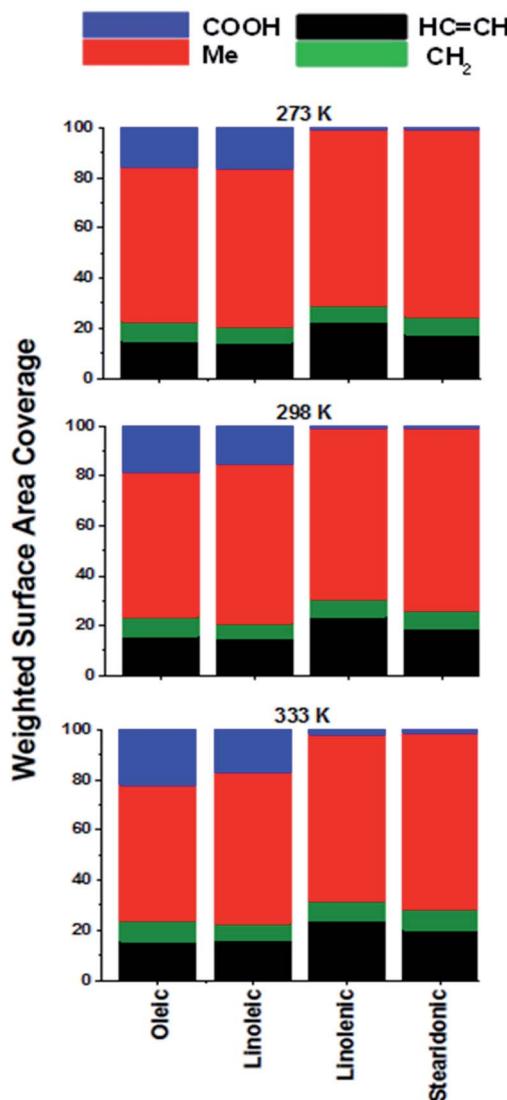


Fig. 4 Weighted surface area coverages of different groups, generated by dividing the SASA areas (Fig. 3) by the number of groups present in the molecule of interest and normalizing the results to 100.

the greater resolution afforded by smaller slice sizes and the need to have enough atoms within each slice for reliable statistical analysis. The graphs shown have been averaged across slices an equal distance from the centre of the slab in the  $+z$  and  $-z$  directions in order to gain a clearer picture of the preferences of different functional groups for the interface of the slab. Just as for the SASA analysis the results have been symmetrised as the upper and lower surfaces of the slab have identical energetic properties. For each sample the analysis was carried out on the two production runs, with the results shown as the average across these two runs. The error bars represent one standard error of the mean. As for the SASA analysis only times  $> 2.5$  ns were included.

On examination of the graphs in Fig. 5 it is clear that the surfaces of the slabs (*i.e.*  $z \gtrsim 3.5$ ) have a distinct distribution of functional groups which differs from that of the bulk ( $z = 0 \rightarrow 3.5$ ). This distribution can be understood by starting at the



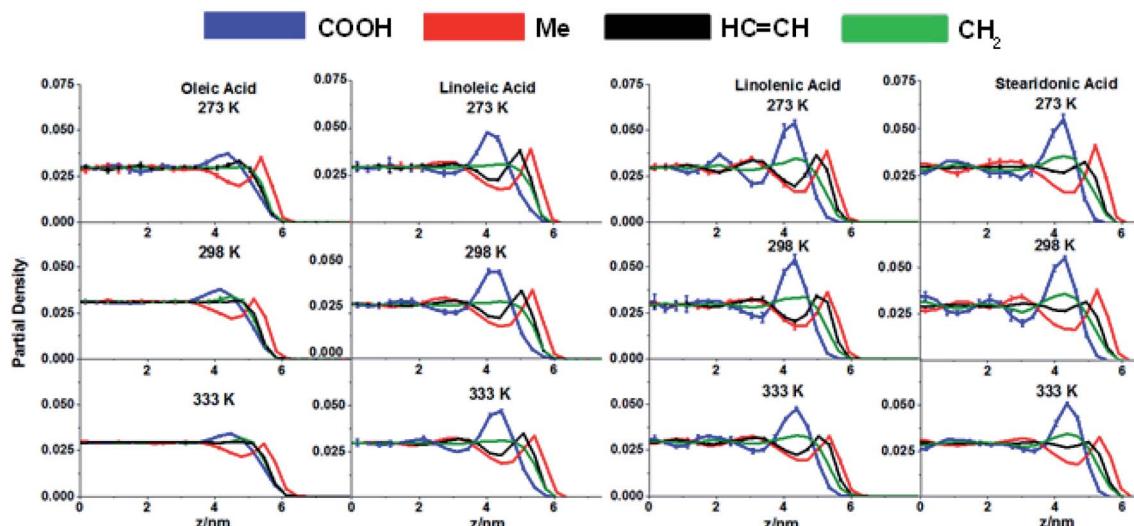


Fig. 5 Partial density analysis for each of the slabs studied, showing the distributions of each of the functional groups studied at different distances from the centre of the slab ( $z = 0$ ). Each curve is the average of results from two 20 ns production runs, separated by an annealing step, with the error bars corresponding to one standard error about this mean. Each of the curves has been area normalised to 0.5, to take into account the different numbers of each of the functional groups present within the samples. The curves represent the averages over the positive and negative  $z$  directions. Colour scheme matches Fig. 2–4.

extreme positive  $z$  values and moving in towards the centre of the slab; at the extreme outermost parts of the slabs it is almost exclusively  $\text{CH}_3$  groups present, with regions of enhanced  $\text{HC}=\text{CH}$  and  $\text{CH}_2$  concentration appearing consistently slightly closer to the centre of the slab, and then the  $\text{COOH}$  inside this. This supports the conclusions drawn from the SASA analysis (highlighted in Fig. 3 and 4), in which the greatly enhanced presence of methyl groups, and diminished presence of acid groups, at the surface was observed. However, whereas SASA analysis only probes those atoms that are directly present at the interface, partial density analysis allows for information to be gained on the ordering of molecules further into the slab. From Fig. 5 it can be seen that while there is a lower presence of  $\text{COOH}$  groups at the surface, there is a region of enhanced concentration in the sub-interfacial region at  $z \approx 4.3$  nm. This corresponds to approximately a single molecule length below the surface. The enhanced sub-surface concentration of  $\text{COOH}$  coincides with a diminished concentration of methyl in all molecules, and of  $\text{HC}=\text{CH}$  in linoleic, linolenic and stearidonic acids. This gives further evidence of the preferential configuration of those molecules directly at the surface, so that the  $\text{CH}_3$  groups are positioned pointing outwards, as discussed with regards to the SASA analysis. There is, however, only a single peak for each of the functional groups in the positive and negative  $z$  directions with the distributions settling down to their bulk values in the centre of the slab. It seems that this preferential ordering of groups is only present in those molecules directly at the interface, a long-distance order extending inwards towards the centre of the slab is not observed. Other than the preference of methyl groups to be at the surface there was no observed ordering of the chains with relation to each other, as is discussed in the ESI S10.† There was also no ordering observed in any directions parallel to the surfaces,

showing that the effects discussed above were purely a result of the presence of the interfaces (see ESI S11†).

## Conclusions

The chemical functionality at the interface of a series of pure fatty acids has been investigated using molecular dynamics simulations. Calculations have been carried out at three different temperatures on slabs of four different members of the oleic acid family, with increasing degrees of saturation. The distribution of different functional groups with relation to the vacuum–liquid interface has been analysed and compared to that which would be expected if the relative proportions of groups at the surface were defined purely by the concentrations of these within the bulk samples.<sup>45,46</sup>

Solvent accessible surface area analysis was used to probe the coverage of different functional groups present directly at the interface. We find that there was a much higher than statistical number of methyl groups observed at the surface for all acids and at all temperatures studied compared to that predicted based on the stoichiometry of the sample. Conversely the proportions of  $\text{CH}_2$ ,  $\text{HC}=\text{CH}$  and  $\text{COOH}$  groups present at the interface was surprisingly low (and essentially zero for  $\text{COOH}$  in linolenic and stearidonic acids) for all samples. This was postulated to be due to the favourable energetics of having the non-polar  $\text{CH}_3$  at the interface, with the more polar groups directed towards the centre of the slab. The exposure of  $\text{HC}=\text{CH}$  at the interface was seen to increase linearly with the increase in the number of double bonds present in the molecules up to linolenic acid ( $n = 3$ ) as found experimentally by Li *et al.*<sup>42</sup> and Nah *et al.*<sup>43</sup> However, this trend did not continue to stearidonic acid ( $n = 4$ ), which displayed a similar level of exposure to linolenic acid.



Partial density analysis has been used in order to probe the concentrations of each of the functional groups at different distances from the interface. This confirms the higher concentration of methyl groups directly at the interface, with enhanced concentrations of each of the other groups in the regions immediately inwards of this. For each of the functional groups only a single peak is observed in the partial density plot, with values for all groups returning to their bulk values a single molecule length in from the interface. This suggests that any non-statistical distribution of functional groups as a result of the presence of the surface is only present in the first layer of molecules at this interface, with no evidence of long-range ordering in the direction perpendicular to the interface.

The presence of non-statistical coverages of groups at the interface is relevant when modelling the reactions that take place at the surface of atmospheric aerosols. The greater affinity of less reactive methyl groups for the surface could lead to slower rates of reaction for these processes than would be calculated using models which assume that the surface functional group concentrations are the same as in the bulk. The effect of the presence of the surface on the orientation of molecules at an interface must therefore be explicitly taken into account when modelling the rates of reactions that occur there. Whilst the presence of water and other species within an aerosol could alter the orientations of fatty acid molecules present within these, as compared to in the pure samples simulated here, the presence of such groups is likely to only enhance this surface preference for methyl groups, as further species that can form hydrogen bonds within the slab would increasingly favour COOH groups being directed inwards. The use of pure fatty acids as model systems for aerosols is indeed common,<sup>40,45,80,81</sup> and the functional group surface preferences found here using MD simulations could be confirmed using highly surface sensitive experimental techniques such as reactive atom scattering.<sup>7,52,78,82</sup> A technique that can provide surface atom (as opposed to surface layer) resolution using reactive OH probes. This would allow this MD methodology to be used to make predictions about the surfaces of more complex systems, such as those that would be present on the surface of atmospheric aerosols, which are currently not experimentally tractable.

## Conflicts of interest

There are no conflicts to declare.

## Acknowledgements

This work used the Cirrus UK National Tier-2 HPC Service at EPCC (<https://www.cirrus.ac.uk>) funded by the University of Edinburgh and EPSRC (EP/P020267/1), with access via the EPSRC Tier 2 Open Access RAP. S. J. G and M. J. P. thank the EPSRC for funding via the EP/T021675/1 Programme and EP/P001459/1 Platform grants. A. C. S. acknowledges Heriot-Watt University for a James Watt PhD scholarship.

## References

- W. C. Keene, R. Sander, A. A. P. Pszenny, R. Vogt, P. J. Crutzen and J. N. Galloway, *Aerosol PH in the Marine Boundary Layer: A Review and Model Evaluation*, *J. Aerosol Sci.*, 1998, **29**(3), 339–356, DOI: 10.1016/S0021-8502(97)10011-8.
- M. Kanakidou, J. H. Seinfeld, S. N. Pandis, I. Barnes, F. J. Dentener, M. C. Facchini, R. Van Dingenen, B. Ervens, A. Nenes, C. J. Nielsen, E. Swietlicki, J. P. Putaud, Y. Balkanski, S. Fuzzi, J. Horth, G. K. Moortgat, R. Winterhalter, C. E. L. Myhre, K. Tsigaridis, E. Vignati, E. G. Stephanou and J. Wilson, *Organic Aerosol and Global Climate Modelling: A Review*, *Atmos. Chem. Phys.*, 2005, **5**(4), 1053–1123, DOI: 10.5194/acp-5-1053-2005.
- D. Warburton, M. Schwarz, D. Tefft, G. Flores-Delgado, K. D. Anderson and W. V. Cardoso, *The Molecular Basis of Lung Morphogenesis*, *Mech. Dev.*, 2000, **92**(1), 55–81, DOI: 10.1016/S0925-4773(99)00325-1.
- J. A. Whitsett, S. E. Wert and T. E. Weaver, *Alveolar Surfactant Homeostasis and the Pathogenesis of Pulmonary Disease*, *Annu. Rev. Med.*, 2010, **61**, 105–119, DOI: 10.1146/annurev.med.60.041807.123500.
- G. M. Nathanson, P. Davidovits, D. R. Worsnop and C. E. Kolb, *Dynamics and Kinetics at the Gas-Liquid Interface*, *J. Phys. Chem.*, 1996, **100**(31), 13007–13020, DOI: 10.1021/jp953548e.
- C. P. Mehnert, R. A. Cook, N. C. Dispenziere and M. Afeworki, *Supported Ionic Liquid Catalysis - A New Concept for Homogeneous Hydroformylation Catalysis*, *J. Am. Chem. Soc.*, 2002, **124**(44), 12932–12933, DOI: 10.1021/ja0279242.
- M. A. Tesa-Serrate, E. J. Smoll, T. K. Minton and K. G. McKendrick, *Atomic and Molecular Collisions at Liquid Surfaces*, *Annu. Rev. Phys. Chem.*, 2016, **67**(1), 515–540, DOI: 10.1146/annurev-physchem-040215-112355.
- P. Davidovits, C. E. Kolb, L. R. Williams, J. T. Jayne and D. R. Worsnop, *Mass Accommodation and Chemical Reactions at Gas-Liquid Interfaces*, *Chem. Rev.*, 2006, **106**(4), 1323–1354, DOI: 10.1021/cr040366k.
- A. J. Kenyon, A. J. McCaffery, C. M. Quintella and M. D. Zidan, *Dynamics of the Gas/Liquid Interface from Laser Molecular Beam Scattering*, *Faraday Discuss.*, 1993, **96**, 245–254, DOI: 10.1039/FD9939600245.
- A. D. Estillore, J. V. Trueblood and V. H. Grassian, *Atmospheric Chemistry of Bioaerosols: Heterogeneous and Multiphase Reactions with Atmospheric Oxidants and Other Trace Gases*, *Chem. Sci.*, 2016, **7**(11), 6604–6616, DOI: 10.1039/c6sc02353c.
- J. Zhong, M. Kumar, J. S. Francisco and X. C. Zeng, *Insight into Chemistry on Cloud/Aerosol Water Surfaces*, *Acc. Chem. Res.*, 2018, **51**(5), 1229–1237, DOI: 10.1021/acs.accounts.8b00051.
- U. Pöschl and M. Shiraiwa, *Multiphase Chemistry at the Atmosphere-Biosphere Interface Influencing Climate and Public Health in the Anthropocene*, *Chem. Rev.*, 2015, 4440–4475, DOI: 10.1021/cr500487s.



13 A. L. Steiner, Role of the Terrestrial Biosphere in Atmospheric Chemistry and Climate, *Acc. Chem. Res.*, 2020, **53**(7), 1260–1268, DOI: 10.1021/acs.accounts.0c00116.

14 M. O. Andreae and P. J. Crutzen, Atmospheric Aerosols: Biogeochemical Sources and Role in Atmospheric Chemistry, *Science*, 1997, **276**(5315), 1052–1058, DOI: 10.1126/science.276.5315.1052.

15 M. C. Jacobson, H. Hansson, K. J. Noone and R. J. Charlson, Organic Atmospheric Aerosols: Review and State of the Science, *Rev. Geophys.*, 2000, **25**(9), 267–294, DOI: 10.1029/1998RG000045.

16 S. Solomon, Stratospheric Ozone Depletion: A Review of Concepts and History, *Rev. Geophys.*, 1999, **37**(3), 275–316, DOI: 10.1029/1999RG900008.

17 I. J. George and J. P. D. Abbatt, Heterogeneous Oxidation of Atmospheric Aerosol Particles by Gas-Phase Radicals, *Nat. Chem.*, 2010, **2**, 713–722, DOI: 10.1038/nchem.806.

18 C. E. Kolb and D. R. Worsnop, Chemistry and Composition of Atmospheric Aerosol Particles, *Annu. Rev. Phys. Chem.*, 2012, **63**, 471–491, DOI: 10.1146/annurev-physchem-032511-143706.

19 Y. Ajili, K. Hammami, N. E. Jaidane, M. Lanza, Y. N. Kalugina, F. Lique and M. Hochlaf, On the Accuracy of Explicitly Correlated Methods to Generate Potential Energy Surfaces for Scattering Calculations and Clustering: Application to the HCl-He Complex, *Phys. Chem. Chem. Phys.*, 2013, **15**, 10062–10070, DOI: 10.1039/c3cp44708a.

20 J. A. S. Lighty, J. M. Veranth and A. F. Sarofim, Combustion Aerosols: Factors Governing Their Size and Composition and Implications to Human Health, *J. Air Waste Manag. Assoc.*, 2000, **50**(9), 1565–1618, DOI: 10.1080/10473289.2000.10464197.

21 O. Tkáč, A. G. Sage, S. J. Greaves, A. J. Orr-Ewing, P. J. Dagdigian, Q. Ma and M. H. Alexander, Rotationally Inelastic Scattering of  $\text{CD}_3$  and  $\text{CH}_3$  with He: Comparison of Velocity Map-Imaging Data with Quantum Scattering Calculations, *Chem. Sci.*, 2013, **4**, 4199–4211, DOI: 10.1039/c3sc52002a.

22 R. J. Charlson, S. E. Schwartz, J. M. Hales, R. D. Cess, J. A. Coakley, J. E. Hansen and D. J. Hofmann, *Information Processing: from Neurons to Climate Forcing by Anthropogenic Aerosols*, MIT Press, 1981, vol. 289, DOI: 10.1029/JC084iC09p05707.

23 J. Haywood and O. Boucher, Estimates of the Direct and Indirect Radiative Forcing Due to Tropospheric Aerosols: A Review, *Rev. Geophys.*, 2000, **38**(4), 513–543, DOI: 10.1029/1999RG000078.

24 J. H. Kroll and J. H. Seinfeld, Chemistry of Secondary Organic Aerosol: Formation and Evolution of Low-Volatility Organics in the Atmosphere, *Atmos. Environ.*, 2008, **42**(16), 3593–3624, DOI: 10.1016/j.atmosenv.2008.01.003.

25 J. C. Marty, A. Saliot, P. Buat-Ménard, R. Chesselet and K. A. Hunter, Relationship between the Lipid Compositions of Marine Aerosols, the Sea Surface Microlayer, and Subsurface Water, *J. Geophys. Res.*, 1979, **84**, 5707–5716, DOI: 10.1029/JC084iC09p05707.

26 Y. Cheng, S.-M. Li, A. Leithead, P. C. Brickell and W. R. Leitch, Characterizations of Cis-Pinonic Acid and n-Fatty Acids on Fine Aerosols in the Lower Fraser Valley during Pacific 2001 Air Quality Study, *Atmos. Environ.*, 2004, **38**(34), 5789–5800, DOI: 10.1016/j.atmosenv.2004.01.051.

27 J. Zahardis and G. A. Petrucci, The Oleic Acid-Ozone Heterogeneous Reaction System: Products, Kinetics, Secondary Chemistry, and Atmospheric Implications of a Model System - A Review, *Atmos. Chem. Phys.*, 2007, **7**(5), 1237–1274, DOI: 10.5194/acp-7-1237-2007.

28 M. D. King, A. R. Rennie, K. C. Thompson, F. N. Fisher, C. C. Dong, R. K. Thomas, C. Pfrang and A. V. Hughes, Oxidation of Oleic Acid at the Air-Water Interface and Its Potential Effects on Cloud Critical Supersaturations, *Phys. Chem. Chem. Phys.*, 2009, **11**(35), 7699–7707, DOI: 10.1039/b906517b.

29 P. S. Gill, T. E. Graedel and C. S. Weschler, Organic Film on Atmospheric Aerosol-Particles, Fog Droplets, Cloud Droplets, Raindrops, and Snowflakes, *Rev. Geophys.*, 1983, **21**, 903–920, DOI: 10.1029/RG021i004p00903.

30 T. Moise and Y. Rudich, Reactive Uptake of Ozone by Aerosol-Associated Unsaturated Fatty Acids: Kinetics, Mechanism, and Products, *J. Phys. Chem. A*, 2002, **106**(27), 6469–6476, DOI: 10.1021/jp025597e.

31 W. F. Rogge, M. A. Mazurek, L. M. Hildemann, G. R. Cass and B. R. T. Simoneit, Quantification of Urban Organic Aerosols at a Molecular Level: Identification, Abundance and Seasonal Variation, *Atmos. Environ. Part A, Gen. Top.*, 1993, **27**(8), 1309–1330, DOI: 10.1016/0960-1686(93)90257-Y.

32 C. Pfrang, K. Rastogi, E. R. Cabrera-Martinez, A. M. Seddon, C. Dicko, A. Labrador, T. S. Plivelic, N. Cowles and A. M. Squires, Complex Three-Dimensional Self-Assembly in Proxies for Atmospheric Aerosols, *Nat. Commun.*, 2017, **8**(1), DOI: 10.1038/s41467-017-01918-1.

33 M. D. King, K. C. Thompson and A. D. Ward, Laser Tweezers Raman Study of Optically Trapped Aerosol Droplets of Seawater and Oleic Acid Reacting with Ozone: Implications for Cloud-Droplet Properties, *J. Am. Chem. Soc.*, 2004, **126**(51), 16710–16711, DOI: 10.1021/ja0447170.

34 U. K. Krieger, C. Marcolli and J. P. Reid, Exploring the Complexity of Aerosol Particle Properties and Processes Using Single Particle Techniques, *Chem. Soc. Rev.*, 2012, **41**(19), 6631–6662, DOI: 10.1039/c2cs35082c.

35 H. M. Hung and P. Ariya, Oxidation of Oleic Acid and Oleic Acid/Sodium Chloride(Aq) Mixture Droplets with Ozone: Changes of Hygroscopicity and Role of Secondary Reactions, *J. Phys. Chem. A*, 2007, **111**(4), 620–632, DOI: 10.1021/jp0654563.

36 M. D. King, K. C. Thompson, A. D. Ward, C. Pfrang and B. R. Hughes, Oxidation of Biogenic and Water-Soluble Compounds in Aqueous and Organic Aerosol Droplets by Ozone: A Kinetic and Product Analysis Approach Using Laser Raman Tweezers, *Faraday Discuss.*, 2007, **137**, 173–192, DOI: 10.1039/b702199b.

37 H. J. L. Forstner, R. C. Flagan and J. H. Seinfeld, Secondary Organic Aerosol from the Photooxidation of Aromatic



Hydrocarbons: Molecular Composition, *Environmental Sci. Technol.*, 1997, **31**(5), 1345–1358, DOI: 10.1021/es9605376.

38 G. B. Ellison, A. F. Tuck and V. Vaida, Atmospheric Processing of Organic Aerosols, *J. Geophys. Res.*, 1999, **104**, 11633–11641, DOI: 10.1029/1999JD900073.

39 C. Waring, K. L. King, P. A. J. Bagot, M. L. Costen and K. G. McKendrick, Collision Dynamics and Reactive Uptake of OH Radicals at Liquid Surfaces of Atmospheric Interest, *Phys. Chem. Chem. Phys.*, 2011, **13**(18), 8457–8469, DOI: 10.1039/c0cp02734k.

40 J. W. Morris, P. Davidovits, J. T. Jayne, J. L. Jimenez, Q. Shi, C. E. Kolb, D. R. Worsnop, W. S. Barney and G. Cass, Kinetics of Submicron Oleic Acid Aerosols with Ozone: A Novel Aerosol Mass Spectrometric Technique, *Geophys. Res. Lett.*, 2002, **29**(9), 71–74, DOI: 10.1029/2002gl014692.

41 T. Thornberry and J. P. D. Abbatt, Heterogeneous Reaction of Ozone with Liquid Unsaturated Fatty Acids: Detailed Kinetics and Gas-Phase Product Studies, *Phys. Chem. Chem. Phys.*, 2004, **6**(1), 84–93, DOI: 10.1039/b310149e.

42 J. Li, S. M. Forrester and D. A. Knopf, Heterogeneous Oxidation of Amorphous Organic Aerosol Surrogates by O<sub>3</sub>, NO<sub>3</sub>, and OH at Typical Tropospheric Temperatures, *Atmos. Chem. Phys.*, 2020, **20**(10), 6055–6080, DOI: 10.5194/acp-20-6055-2020.

43 T. Nah, S. H. Kessler, K. E. Daumit, J. H. Kroll, S. R. Leone and K. R. Wilson, OH-Initiated Oxidation of Sub-Micron Unsaturated Fatty Acid Particles, *Phys. Chem. Chem. Phys.*, 2013, **15**(42), 18649–18663, DOI: 10.1039/c3cp52655k.

44 D. M. Popolan-Vaida, C. L. Liu, T. Nah, K. R. Wilson and S. R. Leone, Reaction of Chlorine Molecules with Unsaturated Submicron Organic Particles, *Zeitschrift für Phys. Chemie*, 2015, **229**(10–12), 1521–1540, DOI: 10.1515/zpch-2015-0662.

45 J. D. Hearn, A. J. Lovett and G. D. Smith, Ozonolysis of Oleic Acid Particles: Evidence for a Surface Reaction and Secondary Reactions Involving Criegee Intermediates, *Phys. Chem. Chem. Phys.*, 2005, **7**(3), 501–511, DOI: 10.1039/b414472d.

46 M. Iwahashi, Y. Kasahara, H. Matsuzawa, K. Yagi, K. Nomura, H. Terauchi, Y. Ozaki and M. Suzuki, Self-Diffusion, Dynamical Molecular Conformation, and Liquid Structures of n-Saturated and Unsaturated Fatty Acids, *J. Phys. Chem. B*, 2000, **104**(26), 6186–6194, DOI: 10.1021/jp000610l.

47 J. J. Nájera, C. J. Percival and A. B. Horn, Infrared Spectroscopic Evidence for a Heterogeneous Reaction between Ozone and Sodium Oleate at the Gas-Aerosol Interface: Effect of Relative Humidity, *Int. J. Chem. Kinet.*, 2015, 277–288, DOI: 10.1002/kin.20907.

48 G. M. Nathanson, Molecular Beam Studies of Gas-Liquid Interfaces, *Annu. Rev. Phys. Chem.*, 2004, **55**, 231–255, DOI: 10.1146/annurev.physchem.55.091602.094357.

49 C. T. Rettner, J. A. Barker and D. S. Bethune, Angular and Velocity Distributions Characteristic of the Transition between the Thermal and Structure Regimes of Gas-Surface Scattering, *Phys. Rev. Lett.*, 1991, **67**, 2183–2186, DOI: 10.1103/PhysRevLett.67.2183.

50 C. Steinbruechel, Gas-Surface Scattering Distributions According to the Hard-Spheroid Model, *Surf. Sci.*, 1982, **115**, 247–258, DOI: 10.1016/0039-6028(82)90405-8.

51 I. Benjamin, Reaction Dynamics at Liquid Interfaces, *Annu. Rev. Phys. Chem.*, 2015, **66**(1), 165–188, DOI: 10.1146/annurev-physchem-040214-121428.

52 E. J. Smoll, M. A. Tesa-Serrate, S. M. Purcell, L. D'Andrea, D. W. Bruce, J. M. Slattery, M. L. Costen, T. K. Minton and K. G. McKendrick, Determining the Composition of the Vacuum-Liquid Interface in Ionic-Liquid Mixtures, *Faraday Discuss.*, 2018, **206**, 497–522, DOI: 10.1039/c7fd00175d.

53 S. P. K. Köhler, S. K. Reed, R. E. Westacott and K. G. McKendrick, Molecular Dynamics Study to Identify the Reactive Sites of a Liquid Squalane Surface, *J. Phys. Chem. B*, 2006, **110**(24), 11717–11724, DOI: 10.1021/jp060431p.

54 M. A. Tesa-Serrate, E. J. Smoll, L. D'Andrea, S. M. Purcell, M. L. Costen, D. W. Bruce, J. M. Slattery, T. K. Minton and K. G. McKendrick, Hiding the Headgroup? Remarkable Similarity in Alkyl Coverage of the Surfaces of Pyrrolidinium- and Imidazolium-Based Ionic Liquids, *J. Phys. Chem. C*, 2016, **120**, 27369–27379, DOI: 10.1021/acs.jpcc.6b09315.

55 B. K. Radak, S. Yockel, D. Kim and G. C. Schatz, Modeling Reactive Scattering of F(2P) at a Liquid Squalane Interface: A Hybrid QM/MM Molecular Dynamics Study, *J. Phys. Chem. A*, 2009, **113**(26), 7218–7226, DOI: 10.1021/jp809546r.

56 Y. Peng, L. Liu, Z. Cao, S. Li, O. A. Mazyar, W. L. Hase and T. Yan, Chemical Dynamics Simulation of Ne Atom Scattering off a Squalane Surface, *J. Phys. Chem. C*, 2008, **112**(51), 20340–20346, DOI: 10.1021/jp804854m.

57 J. Wang, R. M. Wolf, J. W. Caldwell, P. A. Kollman and D. A. Case, Development and Testing of a General Amber Force Field, *J. Comput. Chem.*, 2004, **25**, 1157–1174, DOI: 10.1002/jcc.20035.

58 D. Van Der Spoel, E. Lindahl, B. Hess, G. Groenhof, A. E. Mark and H. J. C. Berendsen, GROMACS: Fast, Flexible, and Free, *J. Comput. Chem.*, 2005, **26**(16), 1701–1718, DOI: 10.1002/jcc.20291.

59 H. J. C. Berendsen, D. Van der Spoel and R. Van Drunen, GROMACS: A Message-Passing Parallel Molecular Dynamics Implementation, *Comput. Phys. Commun.*, 1995, **91**(1–3), 43–56, DOI: 10.1016/0010-4655(95)00042-E.

60 E. Lindahl, B. Hess and D. van der Spoel, GROMACS 3.0: A Package for Molecular Simulation and Trajectory Analysis, *J. Mol. Mod.*, 2001, **7**, 306–317, DOI: 10.1007/s008940100045.

61 B. Hess, C. Kutzner, D. van der Spoel and E. Lindahl, GROMACS 4: Algorithms for Highly Efficient, Load-Balanced, and Scalable Molecular Simulation, *J. Chem. Theory Comput.*, 2008, **4**(3), 435–447, DOI: 10.1021/ct700301q.

62 S. Pronk, S. Páll, R. Schulz, P. Larsson, P. Bjelkmar, R. Apostolov, M. R. Shirts, J. C. Smith, P. M. Kasson, D. Van Der Spoel, B. Hess and E. Lindahl, GROMACS 4.5: A High-Throughput and Highly Parallel Open Source Molecular Simulation Toolkit, *Bioinformatics*, 2013, **29**(7), 845–854, DOI: 10.1093/bioinformatics/btt055.

63 S. Páll, M. J. Abraham, C. Kutzner, B. Hess and E. Lindahl, Tackling Exascale Software Challenges in Molecular Dynamics Simulations with GROMACS, in *Solving Software Challenges for Exascale, Lecture Notes in Computer Science*, ed. S. Markidis and E. Laure, E., vol. 8759, 2015, pp. 3–27.

64 K. A. Malde, L. Zuo, M. Breeze, M. Stroet, D. Poger, P. C. Nair, C. Oostenbrin and A. E. Mark, An Automated Force Field Topology Builder (ATB) and Repository: Version 1.0, *J. Chem. Theory Comput.*, 2011, **7**(12), 4026–4037, DOI: 10.1021/ct200196m.

65 M. Stroet, B. Caron, K. M. Visscher, D. P. Geerke, A. K. Malde and A. E. Mark, Automated Topology Builder Version 3.0: Prediction of Solvation Free Enthalpies in Water and Hexane, *Chem. Theory Comput.*, 2018, **14**(11), 5834–5845, DOI: 10.1021/acs.jctc.8b00768.

66 K. B. Koziara, M. Stroet, A. K. Malde and A. E. Mark, Testing and Validation of the Automated Topology Builder (ATB) Version 2.0: Prediction of Hydration Free Enthalpies, *J. Comput. Aided. Mol. Des.*, 2014, **28**(3), 221–233, DOI: 10.1007/s10822-014-9713-7.

67 J. Wang, W. Wang, P. A. Kollman and D. A. Case, Automatic Atom Type and Bond Type Perception in Molecular Mechanical Calculations, *J. Mol. Graph. Model.*, 2006, **25**(2), 247–260, DOI: 10.1016/j.jmgm.2005.12.005.

68 H. J. C. Berendsen, J. P. M. Postma, W. F. Van Gunsteren, A. DiNola and J. R. Haak, Molecular Dynamics with Coupling to an External Bath, *J. Chem. Phys.*, 1984, **81**(8), 3684–3690, DOI: 10.1063/1.448118.

69 W. C. Swope, H. C. Andersen, P. H. Berens and K. R. Wilson, A Computer-Simulation Method for the Calculation of Equilibrium-Constants for the Formation of Physical Clusters of Molecules: Application to Small Water Clusters, *J. Chem. Phys.*, 1982, **76**(1), 637–649, DOI: 10.1063/1.442716.

70 G. J. Martyna, D. J. Tobias and M. L. Klein, Constant Pressure Molecular Dynamics Algorithms, *J. Chem. Phys.*, 1994, **101**(5), 4177–4189, DOI: 10.1063/1.467468.

71 J. A. Kenar, B. R. Moser and G. R. Lis, Naturally Occurring Fatty Acids, in *Fatty Acids: Chemistry, Synthesis, and Applications*, ed. M. U. Ahmad, Elsevier Inc., Libertyville, 2017, pp. 39–43.

72 W. Humphrey, A. Dalke and K. Schulten, VMD - Visual Molecular Dynamics, *J. Molec. Graph.*, 1996, **14**, 33–38, DOI: 10.1016/0263-7855(96)00018-5.

73 F. Eisenhaber, P. Lijnzaad, P. Argos, C. Sander and M. Scharf, The Double Cubic Lattice Method: Integration of Surface Area and Volume Efficient Approaches to Numerical Molecular Assemblies and to Dot Surface Contouring of Molecular Assemblies, *J. Comput. Chem.*, 1995, **16**, 273–284, DOI: 10.1177/026565909100700111.

74 B. Lee and F. M. Richards, The Interpretation of Protein Structures: Estimation of Static Accessibility, *J. Mol. Biol.*, 1995, **55**(3), 379–400, DOI: 10.1016/0022-2836(71)90324-X.

75 S. S. Batsanov, Radii of Elements from Structural Inorganic Chemistry Data, *Russ. Chem. Bull.*, 1995, **1**, 24–29, DOI: 10.1007/BF00696950.

76 L. B. Algrim and P. J. Ziemann, Effect of the Hydroxyl Group on Yields and Composition of Organic Aerosol Formed from OH Radical-Initiated Reactions of Alcohols in the Presence of NO<sub>x</sub>, *ACS Earth Sp. Chem.*, 2019, **3**(3), 413–423, DOI: 10.1021/acsearthspacechem.9b00015.

77 F. A. Houle, W. D. Hinsberg and K. R. Wilson, Oxidation of a Model Alkane Aerosol by OH Radical: The Emergent Nature of Reactive Uptake, *Phys. Chem. Chem. Phys.*, 2015, **17**(6), 4412–4423, DOI: 10.1039/c4cp05093b.

78 M. A. Tesa-Serrate, B. C. Marshall, E. J. Smoll, S. M. Purcell, M. L. Costen, J. M. Slattery, T. K. Minton and K. G. McKendrick, Ionic Liquid-Vacuum Interfaces Probed by Reactive Atom Scattering: Influence of Alkyl Chain Length and Anion Volume, *J. Phys. Chem. C*, 2015, **119**(10), 5491–5505, DOI: 10.1021/jp5126238.

79 J. G. Harris, Liquid-Vapor Interfaces of Alkane Oligomers. Structure and Thermodynamics from Molecular Dynamics Simulations of Chemically Realistic Models, *J. Phys. Chem.*, 1992, **96**(12), 5077–5086, DOI: 10.1021/j100191a062.

80 G. D. Smith, E. Woods, C. L. DeForest, T. Baer and R. E. Miller, Reactive Uptake of Ozone by Oleic Acid Aerosol Particles: Application of Single-Particle Mass Spectrometry to Heterogeneous Reaction Kinetics, *J. Phys. Chem. A*, 2002, **106**(35), 8085–8095, DOI: 10.1021/jp020527t.

81 A. Asad, B. T. Mmereki and D. J. Donaldson, Enhanced Uptake of Water by Oxidatively Processed Oleic Acid, *Atmos. Chem. Phys.*, 2004, **4**(8), 2083–2089, DOI: 10.5194/acp-4-2083-2004.

82 J. Zhang, D. J. Garton and T. K. Minton, Reactive and Inelastic Scattering Dynamics of Hyperthermal Oxygen Atoms on a Saturated Hydrocarbon Surface, *J. Chem. Phys.*, 2002, **117**(13), 6239–6251, DOI: 10.1063/1.1460858.

