



# Chiral detection by induced surface-enhanced Raman optical activity†

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**Combination of optical activity with surface-enhanced Raman scattering has been a dream of physical chemists for a long time. We report a measurement protocol based on silver colloids and aromatic linkers where chiral acids could be detected in concentrations of about  $10^{-5}$  M. We explain the mechanism by binding and self-assembly of the linkers into chiral aggregates on the silver surface. Following the “sergeants-and-soldiers” principle, the chirality is determined by the relatively minor acidic component. Such detection of biologically relevant molecules may be useful when other methods, such as electronic circular dichroism, are not sensitive enough. In the future, variations of the chemical structure of the linker or other conditions are needed to provide a more specific signal allowing one to better discriminate among the optically active molecules.**

Since its discovery,<sup>1</sup> spectroscopy of Raman optical activity (ROA) has developed into many forms<sup>2</sup> and has been applied for various systems, such as organic molecules,<sup>3</sup> sugars,<sup>4–6</sup> peptides,<sup>7,8</sup> proteins,<sup>9–11</sup> nucleic acids, and even whole viruses.<sup>12–15</sup> The ROA measures a small difference in the scattering of right- and left-circularly polarized light, and is therefore more sensitive to enantiomers or other variations in molecular structure than unpolarized Raman scattering.

A big drawback of ROA spectroscopy is the weakness of the signal, such as the low ROA/Raman intensity ratio, the so called circular intensity difference (CID).<sup>16</sup> Typically,  $CID \sim 10^{-4}$ , which brings about the necessity of highly concentrated solutions (sometimes more than  $10 \text{ mg ml}^{-1}$ ) and long measurement times (hours/days), and causes high noise and occasional

artifacts (false signals) in the spectrum.<sup>2,17</sup> ROA analyses often become too lengthy or expensive.

One of the means of ROA sensitivity enhancement explored in recent years is a combination of optical activity with surface enhanced Raman scattering (SERS), providing surface-enhanced ROA (SEROA) spectroscopy.<sup>18,19</sup> When attached to a noble metal surface, molecules may provide astronomically enhanced Raman signals, so that even single-molecule detection seems reasonable.<sup>20</sup> It is interesting that the SERS effect was discovered at about the same time as ROA,<sup>21</sup> but the two techniques were developed rather independently. Although the first theoretical analyses that found SEROA possible appeared in the 1980s,<sup>22–26</sup> early experiments provided inconclusive results. A big problem was the intrinsic instability of the colloids and weakness of the ROA signal.<sup>27–30</sup> More stable spectra were obtained with “intelligent” colloids, stabilized either with a polymer layer,<sup>31</sup> coated with a silica oxide,<sup>32</sup> or formed by metallic nanostructures of various shapes.<sup>33</sup> Complex metal nanostructures, however, may lead to a smaller signal and a complicated optical response difficult to interpret.<sup>32</sup>

Here, we explore molecular SEROA detection through induced chirality of a linker molecule attached to a silver colloid (Fig. 1). This approach is more universal than pursuing a direct interaction since most molecules do not bind firmly to the colloid surface. Another advantage is that used linkers, such as mercaptopyridine derivatives, are prone to form spontaneous chiral aggregates.<sup>34</sup> Within the “sergeants-and-soldiers” effect<sup>35–38</sup> a small perturbation caused by the analyzed species can induce considerable chirality of the surface layer.<sup>23,34</sup> Lately, a similar effect, chirality sensing through a mercaptopyridine “mediator”, has been observed for tip-enhanced Raman scattering.<sup>39</sup> We could detect strong ROA, with a CID up to  $\sim 10^{-3}$ , universally for all the four linkers and several analytes (“chiral modifiers”) investigated. The chiral signal could be verified by “mirror image” enantiomer spectra, measurements on many combinations of the linkers and modifiers, and by reproducing the experiment in two different

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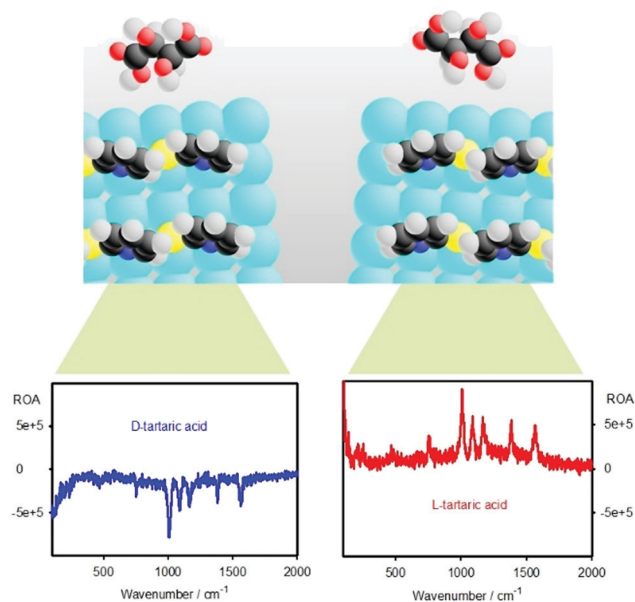


Fig. 1 Scheme of the experiment where the chirality of “linker” molecules assembled on the silver surface (blue) is controlled by chiral molecules (top) in the solution. SEROA spectra of the linker are detected.

laboratories (Academy of Sciences, Prague, and Palacký University, Olomouc).

SEROA spectra obtained for eight linker and analyte combinations are plotted in Fig. 2, and the corresponding SERS (Raman) spectra can be found in Fig. S1 in the ESI†. The SEROA spectra are dominated by the linker bands. For example, for 2-mercaptopyridine we see the ring breathing at 1005 cm<sup>-1</sup>, CH bending (1085–1124 cm<sup>-1</sup>), and C=C/C=N stretching modes around 1586 cm<sup>-1</sup> (cf. ref. 40, Table S1, ESI†). SERS bands of the chiral analytes are occasionally apparent as well; however, these are small and their SEROA is not measurable. For tartaric acid, SERS spectra with and without the linker can be seen in Fig. S2 (ESI†), and are consistent with previous literature.<sup>41</sup>

The noise signal (black in Fig. 2) is reasonably small (CID ~ 10<sup>-5</sup>).<sup>2</sup> Addition of the chiral modifiers leads to quite large CID ratios, often bigger than 10<sup>-3</sup> (Table S2, ESI†). This is up to 100 times more than typical for many organic molecules, and suggests that resonance<sup>2,42,43</sup> or surface-chirality effects<sup>23</sup> participate in the origin of the chiral signal. Also the concentration of the chiral part, ~6 × 10<sup>-6</sup> M, is much lower than in conventional ROA, where about 6 × 10<sup>-2</sup> M would be required for these acids.<sup>2</sup>

For 2-mercaptopyridine, tartaric acid provides the biggest CID, whereas the arginine and alanine values are smaller. This can be attributed to the two chiral centers and two carboxyl groups by which tartaric acid affects the chiral assembly of the linker molecules. Indeed, tartaric acid is known to be efficient in such “chirality seeding”.<sup>44,45</sup> The arginine side chain is polar and bulky, but it is not chiral, which may explain why in our experiments the arginine SEROA signal was smaller than that of alanine and tartaric acid. The signal, however, is not simply proportional to the number of chiral centers in the molecule.

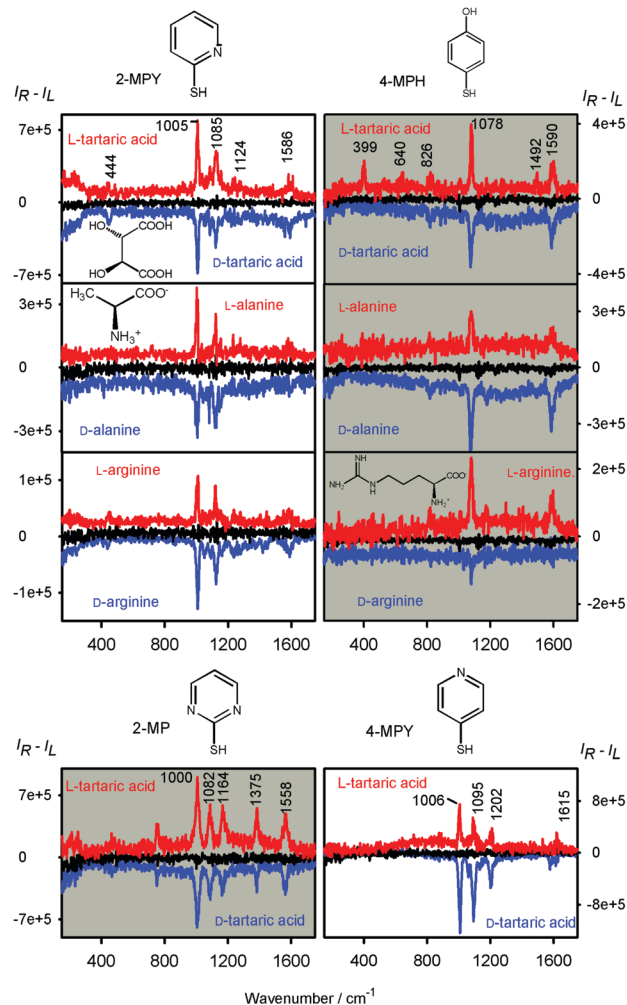


Fig. 2 SEROA spectra for eight linker and chiral acid combinations. The linkers were 2-mercaptopyridine, 2-MPY, 4-mercaptophenol, 4-MPH, 2-mercaptopyrimidine, 2-MP, and 4-mercaptopyridine, 4-MPY; noise spectra obtained without the chiral components are plotted by the black line. The concentration of the chiral acids was 6 × 10<sup>-6</sup> M.

For example, two-center L-isoleucine and D-threonine with 2-MPY (spectra are not shown) provided weak SEROA, similar to arginine.

In many cases, a large SEROA signal is apparent also around 200 cm<sup>-1</sup> and lower (Fig. 2), which can be attributed to the delocalized and inter-molecular modes in the linker aggregates and vibrations of the silver crystal lattice. A similar low-frequency ROA signal was recently also observed for a nitrile liquid.<sup>46</sup>

Variation of the linker molecules does not cause a significant increase or decrease of SEROA; however, some linkers provide more strong bands than the others and thus appear more suitable for this kind of chirality detection. The 4-mercaptopyridine and 2-mercaptopyrimidine linkers seem to give the best signal to noise SEROA ratio, perhaps also because of a positive effect on colloid stability during the measurement. The colloids did not precipitate so quickly and SEROA spectra could be accumulated for a longer time.

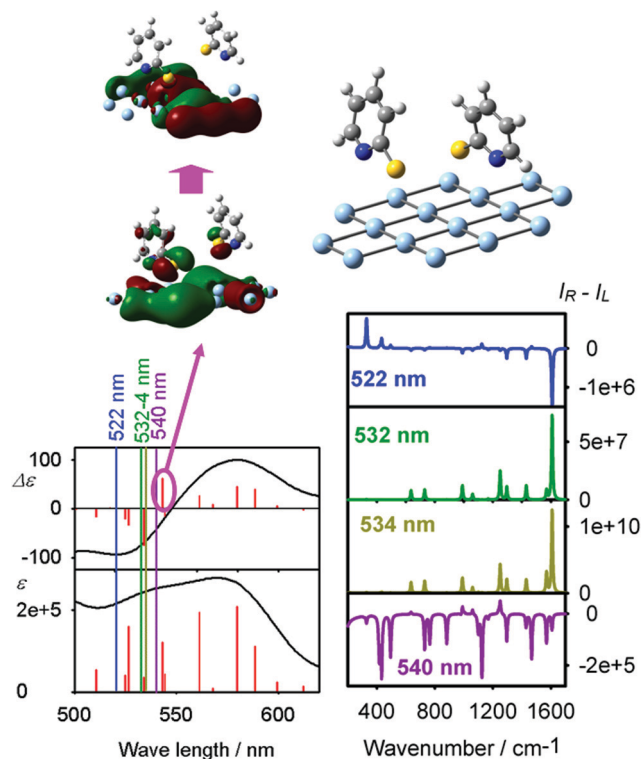


Fig. 3 A model 2-mercaptopyridine/silver cluster, CD ( $\Delta\epsilon$ ) and absorption ( $\epsilon$ ) calculated in the vicinity of the excitation frequency (532 nm), and ROA ( $I_R - I_L$ ) spectra simulated for four excitation frequencies. For the 543 nm transition (magenta ellipse), the dominant molecular orbitals are displayed.

Possible linker-analyte interactions may involve specific hydrogen bond,<sup>47</sup>  $\pi$ - $\pi$ , van der Waals or electrostatic interactions.<sup>48–51</sup> Chirality induction with charged species was also observed for gold-based clusters,<sup>52</sup> and hydrogen bond participation was also observed in SERS of chiral alcohols.<sup>53</sup>

The strong SERS and SEROA signals lasted several minutes/hours after the aggregation was triggered by HCl,<sup>54</sup> depending on the colloid quality and laser power. An example of the time dependence of the spectra for tartaric acid and 2-mercaptopyridine is given in Fig. S3 (ESI†). Normally, the limited time window for ROA accumulation would result in extensive noise. However, because of the large CIDs, SEROA could be measured relatively comfortably within a few minutes.

During the aggregation, the color of the solution changes, and a new band around 600 nm appears in the absorption spectra (Fig. S4, ESI†). Apart from the SERS/SEROA effect, interference of circular dichroism and ROA has been lately discovered as another mechanism of chirality induction in ROA experiments.<sup>55,56</sup> This mechanism can be nevertheless probably ruled out as the circular dichroism is immeasurably weak (Fig. S5, ESI†). Although plasmonic CD is sometimes observable in colloids,<sup>57,58</sup> in our case addition of a small amount of the chiral compound did not produce it.

The interpretation of the data as SEROA is consistent with the theoretical modeling. It should be noted that the computation is not accurate for the resonance,<sup>2,59</sup> but it can be used to understand the qualitative trends.

A typical outcome of the calculations is shown in Fig. 3, where a cluster consisting of 16 silver atoms and two 2-mercaptopyridine molecules provides significant chirality, manifested both in the CD and ROA spectra. For the geometry optimized at the B3LYP/6-311++G\*\*/MWB28/CPCM level, the angles between 2-mercaptopyridine and the silver planes were 70–81°, which is consistent with previous experimental and computational studies.<sup>34</sup>

In the vicinity of an electronic band the excitation light provides a single-sign ROA spectrum.<sup>43</sup> The ROA spectral shape is approximately proportional to the Raman spectrum, with  $\text{CID} = -2R/D$ , where  $R$  and  $D$  are the rotational and dipole strengths of the electronic transition. This can be demonstrated for the excitation wavelength of 540 nm, close to a 543 nm electronic band. The 543 nm electronic transition provides positive CD and the ROA spectrum is thus predominantly negative. Also other tests confirmed the propensity of the chirally attached linker on the silver surface towards one-sign ROA signal (Fig. S6, computational tests and details in S7, S8 and Table S3, ESI†). The simulated CID ratios of  $\sim 10^{-3}$ – $10^{-4}$  are reasonably close to the experimental ones given in Table S2 (ESI†) and confirm previous predictions that the surface chirality can lead to strong light polarization effects.<sup>23</sup> Therefore, the computation shows that the experimental Raman and ROA spectra (Fig. 3) can be attributed to a chiral arrangement of the linkers on the silver surface, controlled by the chiral analyte.

In conclusion, we used the sergeants-and-soldiers principle to obtain strong measurable and reproducible SEROA spectra of the linkers and several chiral analytes. The data were verified by independent measurements and comparison of enantiomers, and interpreted on the basis of model *ab initio* computations. The method leads to sensitive detection of chiral analytes in concentrations smaller than  $10^{-5}$  M. Under such conditions circular dichroism or conventional Raman optical activity spectra normally cannot be measured. The amplification of chirality through asymmetry induction, resonance and surface effects thus appears as a promising means for future biomolecular detection and analyses. It also documents the amazing flexibility and variability of chiral spectroscopy and chemical science.

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## Conflicts of interest

There are no conflicts to declare.

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