



Cite this: *Chem. Commun.*, 2021, 57, 73

Received 7th September 2020,
Accepted 20th November 2020

DOI: 10.1039/d0cc06042a

rsc.li/chemcomm

pH-Dependent peptide bond formation by the selective coupling of α -amino acids in water†

Long-Fei Wu, * Ziwei Liu  and John D. Sutherland*

A novel mechanism enabling selective peptide elongation by coupling α -amino acids over other potentially competing prebiotic amines under acidic aqueous condition is suggested. It proceeds via the generation of a carboxylic acid anhydride intermediate with subsequent intramolecular formation of the amide bond.

The widespread importance of the synthesis of amides lies not only in synthetic organic chemistry,^{1–3} but also in long-standing efforts aimed at understanding the formation of peptides composed of α -amino acids in the origin of life.^{4–7} Although the “RNA world” hypothesis is a popular working model for studying the origin of life,^{8,9} peptides could have coexisted with RNA at the outset of life. This is suggested by the fact that α -amino acids and nucleotide building blocks might have had a common chemical origin.^{10–12} We have previously shown that the chemical energy inherent in methyl isonitrile **1** – a potentially prebiotic activating reagent¹³ – can be harnessed to activate and join phosphate and carboxylate building blocks simultaneously under acidic aqueous conditions.¹⁴ We have now investigated peptide formation chemistry mediated by methyl isonitrile **1** in a systems chemistry scenario. We considered various potentially prebiotic amine nucleophiles which could have been present together in a common pool on early Earth. We found that amide bond formation shows very limited selectivity for all amines examined at pH 5, regardless of the fact that the pK_a values of their conjugate acids differ by up to 5.4 units. When the pH of the reactions was lowered to 3, however, α -amino acids had the highest reactivity amongst all the tested substrates, suggesting a unique mechanism and a potential means of selecting α -amino acids for peptide elongation.

A prebiotically plausible mixture would most likely have contained a range of different amine nucleophiles, regardless of whether it was produced endogenously^{15,16} or exogenously.^{17,18}

Thus, a model peptide, *N*-acetyl *L*-alanine **2** (Ac-Ala, 100 mM), methyl isonitrile **1** (~130 mM) and one of the different amine nucleophiles **3** (50 mM, the concentration of **2** was higher than that of the amine nucleophiles to minimise multiple elongation) were mixed in water and left to incubate at pH 3, 4 or 5 (Fig. 1A). The reaction mixture was monitored by ¹H-NMR spectroscopy at 23 °C until most of the methyl isonitrile has been consumed. After 15 days at pH 5, 62% of glycine **3a** (pK_{aH} = 9.6) was incorporated into amide products, Ac-Ala-Gly **4a** and Ac-Ala-Gly-Gly **4c** (Fig. 1B and Fig. S1, ESI†). Using β -alanine **3b** (β -Ala, pK_{aH} = 10.2), glycylglycine **3c** (Gly-Gly, pK_{aH} = 8.1), glycylglycylglycine **3d** (pK_{aH} = 8.2), glycine nitrile **3e** (pK_{aH} = 5.2) and methylamine **3f** (pK_{aH} = 10.6), we observed reaction with Ac-Ala **2** to give the elongated amide products, Ac-Ala- β -Ala **4b**, Ac-Ala-Gly-Gly **4c** (and minor amount of Ac-Ala-Gly-Gly-Gly-Gly), Ac-Ala-Gly-NH₂ **4d**, Ac-Ala-CN **4e**, and Ac-Ala-NHCH₃ **4f**, respectively, in comparable yield to the combined yield of products from **3a** (as determined by ¹H-NMR spectroscopy, Fig. 1B and Fig. S2–S6, ESI†). However, Ac-Ala-NH₂ **4g** was only formed in 17% yield when ammonia **3g** (pK_{aH} = 9.3) was used as the nucleophile (Fig. S7, ESI†), this is consistent with the lower nucleophilicity of ammonia compared to some amines with similar pK_{aH} values in water.^{19,20} At pH 4, the rate of reaction was significantly higher than it was at pH 5 with consumption of methyl isonitrile **1** in 48 hours.^{14,21,22} The yield of incorporation of glycine **3a** into peptide product **4a** was only slightly lower (58%) than the analogous reaction run at pH 5 (Fig. 1B and Fig. S8, ESI†), and the yield of **4b** decreased to 47% when using **3b** (Fig. 1B and Fig. S9, ESI†). However, the conversion of **3c** or **3d** proceeded in significantly lower yields (approximately 20%, Fig. 1B and Fig. S10, S11, ESI†), and no reaction of **3f** and **3g** with **2** was detected by ¹H-NMR spectroscopy (Fig. 1B and Fig. S12, S13, ESI†). In contrast, quantitative conversion to **4e** was obtained when glycine nitrile **3e** was used as the nucleophile (Fig. 1B and Fig. S14, ESI†). We argued that it might be related to more of the unprotonated form of glycine nitrile being available because of its low pK_{aH} .⁶ When reactions were run at pH 3, methyl isonitrile was consumed in only 6 hours. However, only glycine **3a**, β -alanine **3b**, glycine nitrile **3d** underwent coupling with **2** to give **4a**, **4b** and **4d** in yields of 46%, 5% and 30%,

MRC Laboratory of Molecular Biology, Cambridge Biomedical Campus, Cambridge, CB2 0QH, UK. E-mail: lfwu@mrc-lmb.cam.ac.uk, johns@mrc-lmb.cam.ac.uk

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0cc06042a



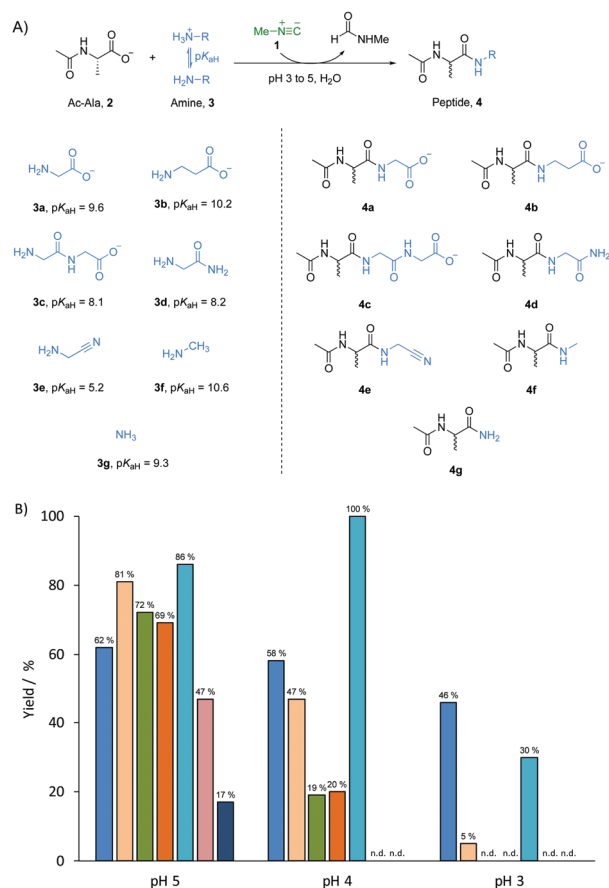


Fig. 1 (A) Formation of amide bond mediated by methyl isonitrile with different amine nucleophiles in water at acidic pH values. (B) Yields based on the incorporation of the amine nucleophiles **3** into peptide products **4** at pH 5, 4 and 3 after 15 days, 48 hours and 6 hours, respectively. Reactions were run in duplicate and average yields are given. n.d.: amide product not detected.

respectively (Fig. 1B and Fig. S15, 17, ESI[†]). The best yield when using glycine **3a** as amine nucleophile suggests there must be a unique mechanism related to glycine **3a** in amide bond formation mediated by methyl isonitrile **1** in low pH solution. To further evaluate if this behaviour also applied to other α -amino acids, we tested arginine, serine, valine and proline with Ac-Ala **2** at pH 4. Yields of incorporation of these amino acids into peptides ranging from 44% to 66% (comparable yields to glycine **3a**) were observed in 48 hours as expected (Fig. S18–S21, ESI[†]).

Accordingly, we wondered if the selectivity we had observed could play out in the sort of mixed system that might have existed on early Earth. A reaction mixture with Ac-Ala **2** (100 mM), glycine **3a** (25 mM), β -alanine **3b** (25 mM), glycylglycine **3c** (25 mM), glycylglycylglycine **3d** (25 mM), glycylglycylglycylglycine **3e** (25 mM), methylamine **3f** (25 mM), ammonia **3g** (25 mM) and methyl isonitrile **1** (~130 mM) was incubated at pH 3 and monitored by ¹H-NMR spectroscopy at 23 °C (Fig. 2A). Peptides **4a** and **4e** were the two major products with yields of 55% and 42%, respectively, based on ¹H-NMR spectroscopy (Fig. S12, ESI[†]), and this is in line with the results of the separate experiments described above. Interestingly, **4b** was not observed by ¹H-NMR spectroscopy (Fig. S18, ESI[†]), which suggests that the reactivity of **3b** was suppressed under a competing scenario.

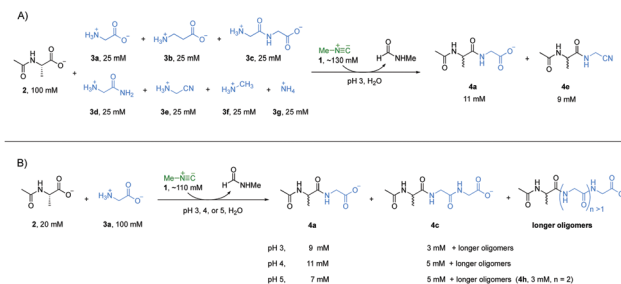


Fig. 2 (A) Model peptide elongation by coupling a mixture of various amine nucleophiles at pH 3 mediated by methyl isonitrile **1**. Concentrations of the two dominating products **4a** and **4d** were calculated based on proton signal integration on ¹H-NMR spectroscopy. (B) Multiple rounds of peptide elongation by coupling an α -amino acid with model peptide **2** mediated by methyl isonitrile at pH 3, 4 or 5. Concentrations of the products were calculated based on proton signal integration on ¹H-NMR spectroscopy.

Considering that progressive peptide elongation is needed to make potentially functional peptides, a reaction with Ac-Ala **2** (20 mM), glycine **3a** (100 mM), and methyl isonitrile **1** (~110 mM) was incubated at pH 3, 4 or 5 to evaluate multiple rounds of peptide elongation (Fig. 2B, other amine nucleophiles were excluded to simplify interpretation of NMR spectra). At pH 3, dipeptide Ac-Ala-Gly **4a** and tripeptide Ac-Ala-Gly-Gly **4c** were observed by ¹H-NMR spectroscopy in yields of 46% and 14%, respectively, after 6 hours (Fig. S23, ESI[†]), which clearly showed that progressive peptide elongation can be achieved under α -amino acid-selective conditions. At pH 4, the yields of **4a** and **4c** were 54% and 24%, respectively, after 48 hours (Fig. S24, ESI[†]). Longer peptides could be observed in minor amounts based on ¹H-NMR spectroscopy. At pH 5, after 15 days, the yields of **4a** and **4c** were 34% and 24% respectively, and tetrapeptide Ac-Ala-Gly-Gly-Gly **4h** was also observed in a yield of 13% (Fig. 2B and Fig. S25, ESI[†]). Although higher yields and longer peptides could be achieved at pH 5, other amine nucleophiles would start to compete with α -amino acids to form amide bonds as shown in Fig. 1.

Next, we investigated the underlying mechanism of the unique property of α -amino acids in the amide bond formation chemistry mediated by methyl isonitrile **1**. Considering the structural features of all the amine nucleophiles, **3a** to **3g**, we reasoned that the carboxylate group of glycine **3a** might be implicated in the special reactivity of α -amino acids. We suggest that the carboxylate group of Ac-Ala **2** was activated by methyl isonitrile **1** to give intermediate imidoyl acetyl alanine **5** via acid catalysis.¹⁴ Imidoyl acetyl alanine **5** can give oxazolone **6** via intramolecular cyclization.^{14,23,24} Both **5** and **6** can be attacked by free amines, if they are sufficiently abundant, to give the amide products intermolecularly (Fig. 3A, direct pathway). Under more acidic condition, more protonation of the amines will inhibit their reactivities as nucleophiles. However, we assumed that the carboxylate/carboxylic acid group of glycine **3a** (pK_{aH} = 2.3) also acts as a nucleophile to attack intermediates **5** and/or **6** to give mixed anhydride **7**. **7** could afford the desired peptide elongation product **4a** by an O \rightarrow N acyl transfer intramolecularly via a 5-membered ring transition state (Fig. 3A, indirect pathway). The pK_{aH} of **7** is



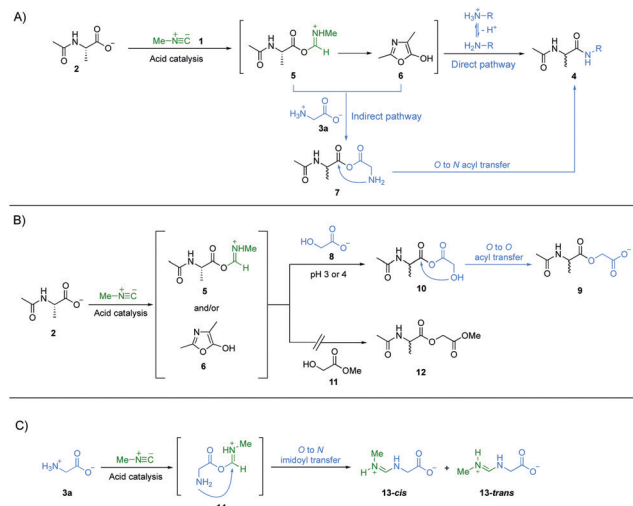


Fig. 3 (A) Proposed mechanistic scheme for methyl isonitrile mediated model peptide elongation by coupling α -amino acids under acidic conditions. The direct pathway represents the mechanism of direct amide bond formation by attacking of the amine nucleophile on intermediates **5** or **6** intermolecularly, and the indirect pathway represents the mechanism of amide bond formation intramolecularly following the mixed anhydride intermediate **7**. (B) Supporting evidence for the indirect pathway proposed in (A). (C) Possible mechanism for the formation of side product *N*-imidoyl glycine, **13-cis** and **13-trans**.

presumably lower than that of glycine **3a** and this coupled with the lower entropic penalty of an intramolecular reaction²⁵ is why this reaction can occur whilst direct intermolecular amide formation cannot (e.g. **3a** versus **3d** in amide bond formation at pH 3 in Fig. 1). To test this hypothesis, we devised an experiment to probe the putative formation of mixed anhydride intermediate **7a**. Ac-Ala **2** (100 mM), methyl isonitrile **1** (~130 mM) and glycolic acid **8** (50 mM) were mixed at pH 3 or 4, respectively, and the reactions were monitored by ¹H-NMR spectroscopy at 23 °C (Fig. 3B). The ester product **9** was observed in good yields at both pH values (Fig. S26 and S27, ESI†). In a negative control reaction, Ac-Ala **2** (100 mM), methyl isonitrile **1** (~130 mM) and methyl glycolate **11** (50 mM) were mixed at pH 4 (Fig. 3B), but ester **12** was not observed by ¹H-NMR spectroscopy after all the methyl isonitrile had been consumed (Fig. S28, ESI†). These results strongly suggest that the formation of ester **9** proceeds *via* the generation of mixed anhydride **10** followed by an O → O acyl transfer. Taken together, these results are an indication that under the most acidic conditions amide **4a** is still able to form in good yield because the reaction proceeds through the mixed anhydride intermediate **7a** followed by an O → N acyl transfer (Fig. 3A, indirect pathway). A similar mechanism would be also expected to apply to β -alanine **3b**, but the lesser amount of carboxylate anion of β -alanine **3b** relative to that of glycine **3a** because of the higher pK_a of the carboxylic acid group of **3b** (pK_a = 3.6) compared to **3a** (pK_a = 2.3), the greater degree of protonation of the amino group of β -alanine because of its higher pK_{aH} than glycine **3a**, and different ring-size of the transition states of intramolecular acyl transfer (6-membered ring *versus* 5-membered ring) might contribute to its lower yield of amide product at pH 3 (Fig. 1B). It is worth mentioning that

side products were observed (Fig. S1, S8 and S15, ESI†), and were assigned as *cis*- and *trans*-*N*-imidoyl glycine, **13-cis** and **13-trans**, based on NMR spectroscopy (Fig. 3C and Fig. S29–S33, ESI†). We suggest that the formation of **13-cis** and **13-trans** is due to the imidoylation of the amino group presumably *via* intermediate **14** followed by an O → N imidoyl transfer. **13** can hydrolyse to give *N*-formyl glycine.²⁶ And *N*-formyl alanine was also shown to elongate effectively by coupling glycine **3a** to give *N*-formylated peptides (Fig. S34, ESI†). These products are reminiscent of the formylated N-terminal residue in extant protein biosynthesis.

Prebiotic chemistry is most likely to give a mixed pool of various plausible species on early Earth.^{10–12,15–18} Exploring whether there is selectivity among those mixed species in the process of forming oligomers is of great importance to understand the transition from prebiotic chemistry to biochemistry. Poor selectivity among various amine nucleophiles was observed when amide bond formation was mediated by methyl isonitrile **1** at only slightly acidic aqueous solution (e.g. pH 5). Under conditions of pH 6 or above, peptide elongation will be very slow because of a lack of acid catalysis. But, a unique chemical property inherent to α -amino acids could enable the incorporation of α -amino acids selectively into peptides at pH 3 in the presence of other amine nucleophiles. Also, progressive peptide elongation by incorporating α -amino acids can still be achieved in the same set of conditions. A mixed carboxylic acid anhydride intermediate (e.g. **7a**, Fig. 3A) was suggested to be the reason contributing to the highest activity of α -amino acids at pH 3. As α -aminonitriles are easily hydrolysed to α -aminoamides (e.g. by organocatalysis by aldehydes²⁷), then to α -amino acids on a longer time scale (geologically plausible), one can imagine a prebiotic scenario in which α -amino acids existed alongside various other amines, but in the absence of α -aminonitriles. In such a scenario, peptides comprising only α -amino acids could have been produced at low pH by the chemistry described herein. Alternatively, if the hydrolysis of α -aminonitriles to α -amino acids was incomplete, peptides based on α -amino acids could still have been produced, but they would occasionally be terminated by α -aminonitriles. Interestingly, Powner and co-workers recently described a three-steps process in coupling each α -aminonitrile as α -amino acid equivalent in aqueous solution.⁶ The selective coupling of α -aminonitrile in their work resulted from the corresponding much lower pK_{aH} values of α -aminonitrile than other amino nucleophiles, which was also partially exemplified here. Hud and co-workers have explored the ester product **9** analogues as key intermediates in amide bond formation driven by wet-dry cycles without using activating reagent.²⁸ We anticipate that, in solution phase as described here, if the pH of the solution were brought up by environmental fluctuation, amines might react with ester **9** slowly and low selectivity would be expected.²⁹ Regarding the low pH conditions, although models predict that the pH of the ocean of early Earth was near neutral,^{30,31} regional lakes/ponds with high acidity would have been possible.^{32,33}

This work was supported by the Medical Research Council (no. MC_UP_A024_1009 to J. D. S.) and the Simons Foundation



(no. 290362 to J. D. S.). The authors thank all J. D. S. group members for fruitful discussion.

Conflicts of interest

There are no conflicts to declare.

Notes and references

- 1 A. El-Faham and F. Albericio, *Chem. Rev.*, 2011, **111**, 6557–6602.
- 2 R. M. de Figueiredo, J.-S. Supp and J.-M. Campagne, *Chem. Rev.*, 2016, **116**, 12029–12122.
- 3 M. T. Sabatini, L. T. Boulton, H. F. Sneddon and T. D. Sheppard, *Nat. Catal.*, 2019, **2**, 10–17.
- 4 G. Danger, R. Plasson and R. Pascal, *Chem. Soc. Rev.*, 2012, **41**, 5416–5429.
- 5 M. Frenkel-Pinter, M. Samanta, G. Ashkenasy and L. Leman, *Chem. Rev.*, 2020, **120**, 4707–4765.
- 6 P. Canavelli, S. Islam and M. Powner, *Nature*, 2019, **571**, 546–549.
- 7 L. Leman, L. E. Orgel and M. R. Ghadiri, *Science*, 2004, **306**, 283–286.
- 8 L. E. Orgel, *Crit. Rev. Biochem. Mol. Biol.*, 2004, **39**, 99–123.
- 9 G. F. Joyce and J. W. Szostak, *Cold Spring Harbor Perspect. Biol.*, 2018, **10**, a034801.
- 10 A. Eschenmoser, *Tetrahedron*, 2007, **63**, 12821–12844.
- 11 J. D. Sutherland, *Angew. Chem., Int. Ed.*, 2016, **55**, 104–121.
- 12 L.-F. Wu and J. D. Sutherland, *Emerging Top. Life Sci.*, 2019, **3**, 459–468.
- 13 A. Mariani, D. A. Russell, T. Javelle and J. D. Sutherland, *J. Am. Chem. Soc.*, 2018, **140**, 8657–8661.
- 14 Z. Liu, L.-F. Wu, J. Xu, C. Bonfio, D. A. Russell and J. D. Sutherland, *Nat. Chem.*, 2020, **12**, 1023–1028.
- 15 S. L. Miller and L. E. Orgel, *The Origin of Life on the Earth*, Prentice-Hall, 1974.
- 16 B. H. Patel, C. Percivalle, D. Riston, C. D. Duffy and J. D. Sutherland, *Nat. Chem.*, 2015, **7**, 301–307.
- 17 C. Chyba and C. Sagan, *Nature*, 1992, **355**, 125–132.
- 18 S. Pizzarello, *Acc. Chem. Res.*, 2006, **39**, 231–237.
- 19 F. Brotzel, Y. C. Chu and H. Mayr, *J. Org. Chem.*, 2007, **72**, 3679–3688.
- 20 F. Brotzel and H. Mayr, *Org. Biochem. Chem.*, 2007, **5**, 3814–3820.
- 21 K. Sung and C.-C. Chen, *Tetrahedron Lett.*, 2001, **42**, 4845–4848.
- 22 Y.-Y. Lim and A. R. Stein, *Can. J. Chem.*, 1971, **49**, 2455–2459.
- 23 G. Danger, A. Michaut, M. Bucchi, L. Boiteau, J. Canal, R. Plasson and R. Pascal, *Angew. Chem., Int. Ed.*, 2013, **52**, 611–614.
- 24 Z. Liu, D. Beaufils, J.-C. Rossi and R. Pascal, *Sci. Rep.*, 2014, **4**, 7440.
- 25 A. Kirby, *J. Adv. Phys. Org. Chem.*, 1980, **17**, 183.
- 26 S. Vincent, C. Mioskowski and L. Lebeau, *J. Org. Chem.*, 1999, **64**, 99–997.
- 27 J. Taillades, I. Beuzelin, G. Laurence, V. Tabacik, C. Bied and A. Commeyras, *Origins Life Evol. Biospheres*, 1998, **28**, 61–77.
- 28 J. G. Forsythe, S.-S. Yu, I. Mamajanov, M. A. Grover, R. Krishnamurthy, F. M. Fernández and N. V. Hud, *Angew. Chem., Int. Ed.*, 2015, **54**, 9871–9875.
- 29 A. C. Satterthwait and W. P. Jencks, *J. Am. Chem. Soc.*, 1974, **96**, 7018–7031.
- 30 I. Halevy and A. Bachan, *Science*, 2017, **355**, 1069–1071.
- 31 J. Krissansen-Totton, G. N. Arney and D. C. Catling, *Proc. Natl. Acad. Sci. U. S. A.*, 2018, **115**, 4015–4110.
- 32 R. W. Renaut and B. Jones, in *Encyclopedia of Geobiology*, ed. J. Reitner and V. Thiel, Springer, Berlin, Dordrecht, 2011, pp. 467–479.
- 33 R. W. Henley, in *Volcanic Lakes*, ed. D. Rouwet, B. Christenson, F. Tassi and J. Vandemeulebrouck, Springer, Berlin, Heidelberg, 2015, pp. 155–178.

