



Cite this: *RSC Chem. Biol.*, 2021, 2, 1291

Orthogonal coiled coils enable rapid covalent labelling of two distinct membrane proteins with peptide nucleic acid barcodes†

Georgina C. Gavins,^a Katharina Gröger,^a Marc Reimann,^a Michael D. Bartoschek,^b Sebastian Bultmann^b and Oliver Seitz^{a*}

Templated chemistry offers the prospect of addressing specificity challenges occurring in bioconjugation reactions. Here, we show two peptide-templated amide-bond forming reactions that enable the concurrent labelling of two different membrane proteins with two different peptide nucleic acid (PNA) barcodes. The reaction system is based on the mutually selective coiled coil interaction between two thioester-linked PNA-peptide conjugates and two cysteine peptides serving as genetically encoded peptide tags. Orthogonal coiled coil templated covalent labelling is highly specific, quantitative and proceeds within a minute. To demonstrate the usefulness, we evaluated receptor internalisation of two membranous receptors EGFR (epidermal growth factor) and ErbB2 (epidermal growth factor receptor 2) by first staining PNA-tagged proteins with fluorophore-DNA conjugates and then erasing signals from non-internalized receptors via toehold-mediated strand displacement.

Received 7th June 2021,
Accepted 14th July 2021

DOI: 10.1039/d1cb00126d

rsc.li/rsc-chembio

Introduction

Rapid and site-selective bioconjugation reactions are a key tool at the chemistry-biology interface to establish covalent bonds between proteins and chemically designed entities such as fluorescent labels, purification handles and small molecule drugs. A distinct challenge is to perform chemical bioconjugation reactions in or on living cells.^{1–7} Such reactions are useful for fluorescence labelling and imaging of proteins. Considering the naturally occurring functional group repertoire of biomolecules, it is difficult to site-specifically target a particular site or biomolecule without competing reactions at the multitude of other sites/biomolecules. One solution to the specificity problem is provided by using enzymes that guide the chemistry to peptide recognition tags or enable self-modification reactions at the enzyme's specific microenvironment.^{1–7} However, steric demand, the need for cofactors and carefully adjusted reaction conditions, or limited substrate scope can be reason for concern.^{8,9} By contrast, chemical bioconjugation methods potentially offer tolerance to a wider scope of reaction conditions and substrates. To enable site specific bioconjugation of a

select protein, chemists introduced templates that recognize a particular arrangement of peptide side chains and orient functional groups in a way that facilitates a bimolecular reaction. Noteworthy examples are the bisarsenic bis-thiol adducts reacting with tetracysteine motifs^{10,11} and functionalized metal complexes^{12,13} guiding alkylation reactions to a Cys side chain in the vicinity of oligohistidine or oligoaspartate tags. We, and others, have previously introduced amide and thioether bond forming bioconjugation reactions templated by the formation of coiled coils.^{14–16}

The specificity demands are particularly high for orthogonal bioconjugation chemistries enabling the simultaneous modification of two different biomolecules on live cells. While orthogonal covalent labelling has been achieved with enzyme-based chemistries^{17–19} and through biosynthetic incorporation of unnatural amino acids,²⁰ a chemical method remained elusive. Here, we report the first method to offer orthogonal, chemical bioconjugation on live cells using widely applicable peptide recognition tags. The method relies on coiled coil interactions pioneered by Matsuzaki²¹ for non-covalent labelling. Recently, Beatty used coiled coils for the imaging of two differently localized proteins *via* non-covalent labelling in fixed cells.²² For covalent modification in live cells, we designed a template system to guide an acylation reaction to N-terminal cysteine residues of genetically encoded peptide tags (Cys-Acc-POI in Scheme 1). Labelling agents (such as PNA-Don) recognize the targeted tag *via* formation of coiled coil complexes. This arranges the thioester in the immediate vicinity of the N-terminal cysteine

^a Institut für Chemie, Humboldt-Universität zu Berlin Brook-Taylor-Strasse 2, Berlin 12489, Germany. E-mail: oliver.seitz@chemie.hu-berlin.de

^b Center for Molecular Biosystems (BioSysM), Faculty of Biology, Ludwig-Maximilians-Universität München, Butenandtstr. 1, Munich 81377, Germany. E-mail: bultmann@bio.lmu.de

† Electronic supplementary information (ESI) available: Methods and supporting data. See DOI: 10.1039/d1cb00126d



We utilized *de novo* designed parallel heterodimeric coiled coils (**P1 : P2** and **P3 : P4**) developed by Gradišar and Jerala³⁰ (Fig. 1A). With self-assembly of the **P1 : P2** and **P3 : P4** complexes specificity of the acyl transfer is controlled by templation. The labeling reagents **PNA₁-P2** and **PNA₃-P4** are comprised of donor peptides **P2** and **P4** joined to a **PNA** *via* a thioester (Fig. 1B, for synthesis²⁵ see Scheme S3-1, ESI[†]). To test the feasibility of orthogonal **PNA** transfer, the donor agent **PNA₁-P2** was added in excess to the acceptor peptide **Cys-P1-TMR** (200 nM); a likely scenario in live cell applications. Fluorescence-UPLC (Fl-UPLC) analysis showed that 84% of the acceptor carried the **PNA** tag after only 10 seconds (Fig. 1B, left). Within one minute labelling was complete. The **PNA**-acceptor acyl transfer product (**P** in Fig. 1A, see also Fig. S5, S4, ESI[†]) was mainly observed in the initial 10 seconds but with excess of thioester a second acyl transfer on the free cysteine led to a doubly **PNA**-acylated thioester (**PP'**).¹⁵ As previously reported for the K3/E3 coiled coil templated reaction,²⁵ product **P** was reformed from

A)

Name	Function	<i>gabcdef</i>	<i>gabcdef</i>	<i>gabcdef</i>	<i>gabcdef</i>
Cys-P1:	Acceptor	C EIQALEE	ENAQLEQ	ENAALKE	EIAQLEY
P2:	Donor	KIAQLKE	KNAALKE	KNQQLKE	KIQALKY
Cys-P3:	Acceptor	C EIQGLEE	EIAQLEQ	KNALKE	KNQALKY
P4:	Donor	KIAQLKQ	ENAQLEQ	ENOQLEE	ENAALEY

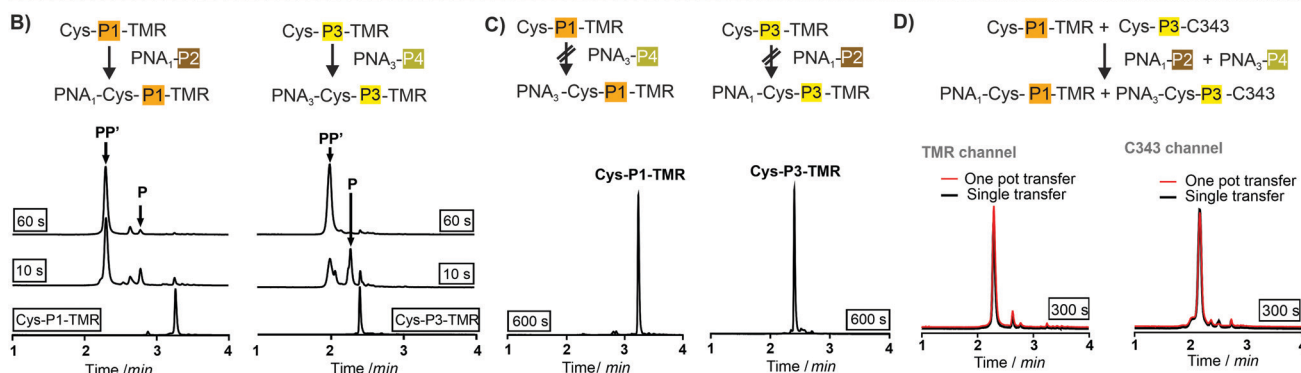
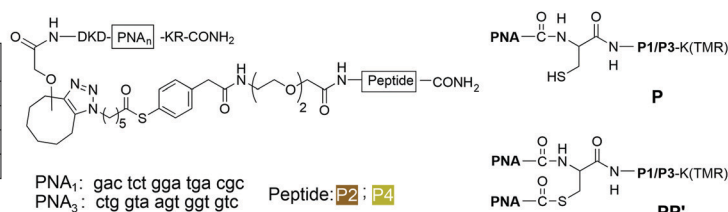


Fig. 1 (A) Orthogonal coiled coils, **PNA**-labeling reagents and products formed in coil-coil templated **PNA** transfer. FI-UPLC analysis of reactions involving (B) matched and (C) mismatched coiled coil peptides. Conditions: 200 nM **Cys-P1-TMR** or **Cys-P3-TMR**, 1200 nM **PNA₁-P2** or **PNA₃-P4** in 200 nM phosphate, 1 mM TCEP, 0.1% CHAPS, pH 7.2, 30 °C. (D) One-pot reactions of a **Cys-P1-TMR/Cys-P3-C343/PNA₁-P2/PNA₃-P4**-mixture (red traces) overlaid with FI-UPLC traces for reactions involving **Cys-P1-TMR** + **PNA₁-P2** or **Cys-P3-C343** + **PNA₃-P4** in separate pots (black traces). Detector settings: TMR, Ex 550 nm, Em 580 nm; C343, Ex 420 nm, Em 500 nm.

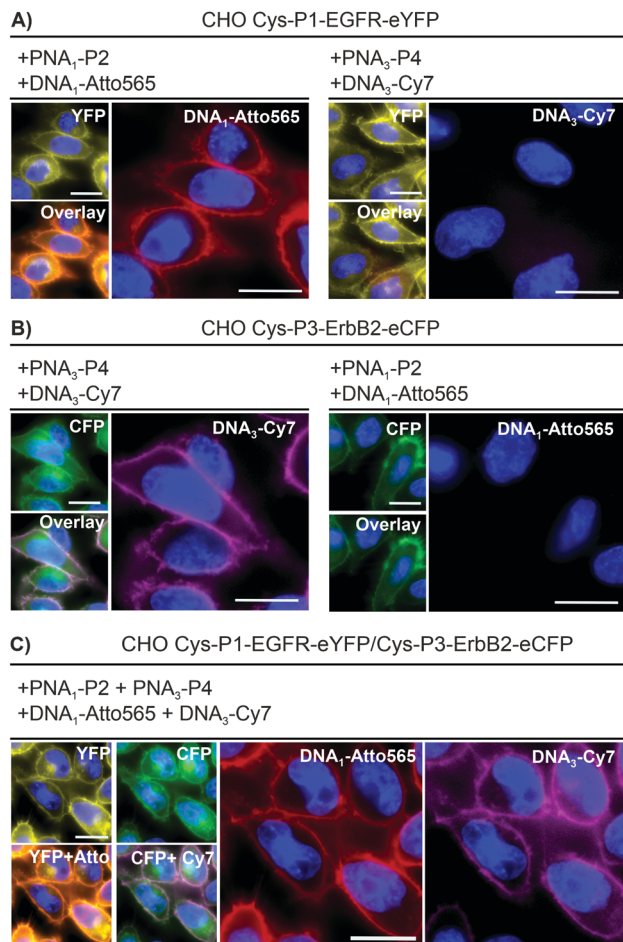


Fig. 2 Dual color live cell labeling. After nuclear staining with Hoechst33342 (blue), (A) **Cys-P1-EGFR-eYFP** or (B) **Cys-P3-ErbB2-eCFP** expressing CHO cells were treated with matched (left) or mismatched (right) **PNA** labeling reagents/fluorophore-DNA. Conditions: 4 min with 100 nM **PNA₁-P2** (EGFR) or/and **PNA₃-P4** (ErbB2) in DPBS, washing with HBSS, 4 min hybridization with 200 nM Atto565-DNA₁ (EGFR) or Cy7-DNA₃ (ErbB2). (C) Simultaneous labeling of **Cys-P1-EGFR-eYFP/Cys-P3-EGFR-eCFP** cells. Scale bar = 10 μ m.

PP' upon addition of NaOH (Fig. S5-1 and 2, ESI[†]). If desired, **PP'** can be converted to **P** in a templated reaction.²⁵ For imaging applications this conversion is not necessary. Testing the second coiled coil template reaction revealed that transfer of **PNA** from **PNA₃-P4** onto **Cys-P3-TMR** proceeded equally rapidly (Fig. 1B right).

The specificity was investigated by incubating mismatched peptides (**PNA₁-P2** with **Cys-P3-TMR** and **PNA₃-P4** with **Cys-P1-TMR**). No new peaks appeared after 10 minutes (Fig. 1C). Furthermore, all peptides were allowed to react in one pot. **Cys-P3-C343** was used in place of the TMR labelled peptide to facilitate FI-UPLC analysis. **Cys-P3-C343** and **Cys-P3-TMR** had similar reactivity (Fig. S5-3, ESI[†]). The one pot dual label transfer reaction proceeded as quickly as reactions between individual coiled coil pairs (Fig. S5-7, ESI[†]). FI-UPLC traces were superimposable suggesting that no new products were formed (Fig. 1D).

To realize live cell fluorescent labelling of the two cell surface proteins EGFR and ErbB2, we used three stable CHO

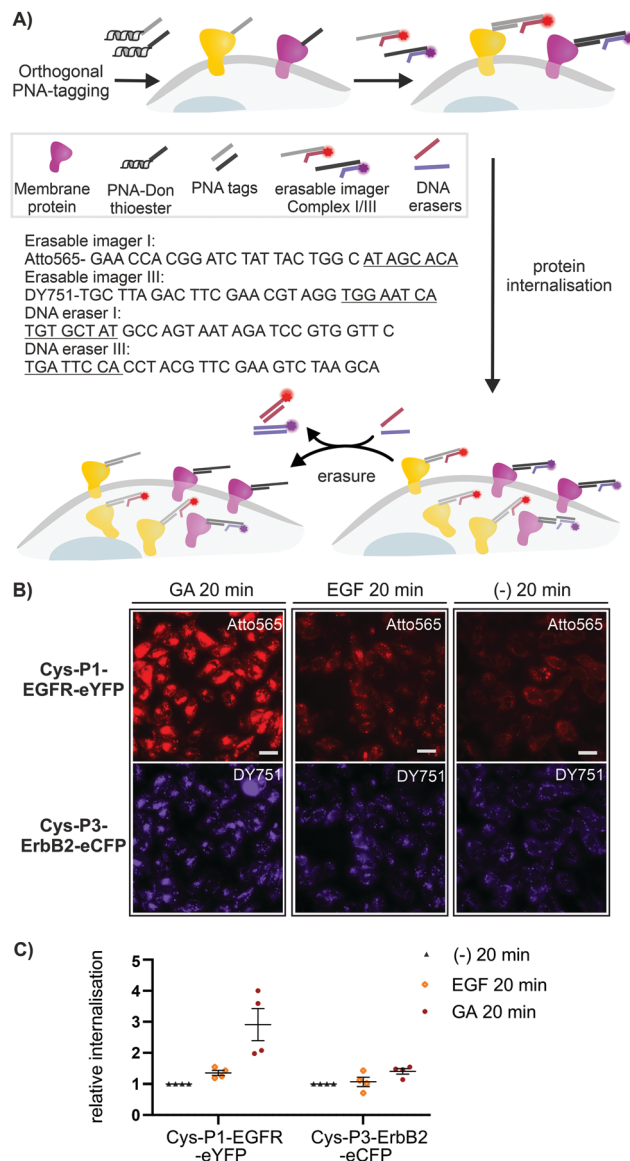


Fig. 3 (A) Dual receptor internalization assay. (B) Fluorescent microscopy imaging of internalized EGFR and ErbB2 in CHO cells. Conditions: see ESI[†]. (C) Quantitative analysis of internalization. Data is presented as internalization relative to the negative control (–) from four independent replicates ($n = 100$ cells per condition per experiment). Sequences of imager strands and eraser strands are given in 5' \rightarrow 3' direction. Toehold-related sequences are underlined. For other sequences see Table S6-1 (ESI[†]).

cells enabling doxycycline inducible expression of **Cys-P1-EGFR-eYFP**, **Cys-P3-ErbB2-eCFP**, or both (Fig. S8-2, ESI[†]). Induced **Cys-P1-EGFR-eYFP** CHO cells were treated with **PNA₁-P2** or **PNA₃-P4** (100 nM), respectively, for 4 minutes. Subsequently, cells were incubated with fluorophore conjugated complementary oligonucleotides; Atto565-DNA₁ for **PNA₁-P2** or Cy7-DNA₃ for **PNA₃-P4**. For the **Cys-P1-EGFR-eYFP** cell line, treatment with matched **PNA₁-P2** and Atto565-DNA₁ stained membrane regions (Fig. 2A). Colocalization scatter plots showed that Atto565 signals were not found in pixels lacking an eYFP signal (Fig. S9-3, ESI[†]). No staining was observed with non-matching **PNA**-donor thioester and DNA-fluorophore. A similar set



of experiments was performed with the **Cys-P3-ErbB2-eCFP**-CHO cells. Staining of membranous regions and colocalization with the eCFP signal succeeded when the dox induced cell were treated with matched **PNA₃-P4** and Cy7-DNA₃ but not with **PNA₁-P2** and Atto565-DNA₁ (compare Fig. 2B left and right). To verify that both the **PNA** conjugation and DNA hybridization steps were selective, every combination of **PNA**-Donor and fluorophore-DNA was tested on both **Cys-P1-EGFR-eYFP** and **Cys-P3-ErbB2-eCFP** cells (Fig. S9-1 and 2, ESI†). Labelling only occurred with the correct combination of peptides and complementary DNA.

For one-pot dual colour labelling CHO cells expressing both **Cys-P1-EGFR-eYFP** and **Cys-P3-ErbB2-eCFP** were submitted to the conditions used for single colour labelling, only with addition of both thioesters, **PNA₁-P2** and **PNA₃-P4**, together followed by hybridization of both oligonucleotides together (Fig. 2C). As before, Atto565 and Cy7 signals marked membrane regions.

To assess the usefulness of our dual colour labelling methodology, we analysed the concurrent internalisation of EGFR and ErbB2 (Fig. 3). These two members of the epidermal growth factor receptor family are critically involved in cancer genesis, and their trafficking is well studied.^{31,32} Of note, there are no known ligands for ErbB2, which renders classical ligand-based assays unsuitable. Cells were incubated with the **PNA₁-P2/PNA₃-P4** mixture. For increasing brightness and enabling erasure of staining, **PNA**-labelled cells were treated with DNA complexes comprised of a 57/59 nt long DNA adaptor carrying two imaging strands (Atto565- or Dy751-DNA conjugate) with a 21/22mer portion complementary to the adaptor strand, and an 8mer overhang (Fig. 3A). This serves as a toehold which enables removal of the imaging strand by treatment with a fully complementary 'eraser DNA' utilizing a process known as toehold-mediated strand displacement. After staining with the DNA complexes, cells were incubated with either EGF (100 nM), geldanamycin (GA, 3 μ M) or just media. Prior to analysis, eraser DNA was added for 8 minutes. Endocytosed receptors are protected from the eraser DNA and any remaining fluorescent signal must be due to internalisation. Therefore, quantification of the fluorescence provides a direct read-out of internalisation. Control experiments confirmed that imager DNA did not stain cells unselectively (Fig. S9-5, ESI†). Cells that were incubated for 20 min with cell medium only showed weak signals within intracellular locations (Fig. 3B, right) indicative of basal internalisation after serum starvation.³³ Treatments with EGF and GA revealed the different internalisation behaviour. After 20 min EGF stimulation, EGFR internalisation increased moderately (\approx 53%) over basal levels, whilst ErbB2 internalisation remained unchanged (Fig. 3B middle and Fig. 3C). Previous measurements had shown that **PNA**-labelled EGFR undergoes autophosphorylation and internalizes upon EGF activation.²⁵ The rather weak internalisation of EGFR observed here is probably due to coexpression with ErbB2, which has been reported to impair EGF stimulated endocytosis of EGFR.³⁴ Both receptors responded more strongly to 20 min treatment with GA. Of note, GA increased EGFR internalisation by a 2.9-fold compared to 1.4-fold for ErbB2. The modest

internalisation of ErbB2 is in agreement with a previous study, which showed that the surface levels of ErbB2 expressed on HEK293 cells were only moderately affected during 1 h of GA stimulation.³⁵

Conclusions

Our data on simultaneous dual-colour labelling of EGFR and ErbB2 on CHO cells shows that coiled coil templation of two parallel acyl transfer reactions is rapid (4 min), bioorthogonal and compatible with cell culture conditions. We used parallel dual labelling to install two different **PNA** barcodes on EGFR and ErbB2, which enable selective fluorescence staining *via* hybridization with commercially available oligonucleotide-fluorophore conjugates. This barcoding method also unlocks the potential for interfacing DNA nanotechnologies, such as toehold-mediated strand displacement, which we used to facilitate the analysis of receptor internalisation by removing signals from non-internalized receptors. As a result, the signal obtained by a direct read-out positively correlates with the amount of internalized receptor. The internalisation assay proceeds with live cells and requires only 20 min. Here, we present orthogonal labelling of two proteins. Moreover, six coiled coil pairs from the same *de novo* designed set have already been shown to be orthogonally functional in mammalian cells by Lebar *et al.*,³⁶ where their interactions were used to simultaneously control protein localization. We believe that using orthogonal coiled coils to template covalent bond forming reactions on live cells is a useful new application and suggests intriguing possibilities for rapid oligonucleotide barcoding of multiple surface proteins on live cells.

Author contributions

G. G. planned and performed experiments and drafted the manuscript. K. G. and M. R. did orienting studies on orthogonal coiled coil-templated reactions. M. D. B. and S. B. constructed the stable CHO cell lines. O. S. conceived the study and wrote the manuscript. All authors have given approval to the final version of the manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We acknowledge financial support from the Deutsche Forschungsgemeinschaft (SPP1623, Se 819/17-2). M. D. B. is a fellow of the International Max Planck Research School for Molecular Life Sciences (IMPRS-LS).



References

- 1 B. N. G. Giepmans, S. R. Adams, M. H. Ellisman and R. Y. Tsien, *Science*, 2006, **312**, 217–224.
- 2 C. Jing and V. W. Cornish, *Acc. Chem. Res.*, 2011, **44**, 784–792.
- 3 L. Xue, I. A. Karpenko, J. Hiblot and K. Johnsson, *Nat. Chem. Biol.*, 2015, **11**, 917–923.
- 4 X. Chen and Y.-W. Wu, *Org. Biomol. Chem.*, 2016, **14**, 5417–5439.
- 5 J. Lotze, U. Reinhardt, O. Seitz and A. G. Beck-Sickinger, *Mol. BioSyst.*, 2016, **12**, 1731–1745.
- 6 A. F. L. Schneider and C. P. R. Hackenberger, *Curr. Opin. Biotechnol.*, 2017, **48**, 61–68.
- 7 P. Wolf, G. Gavins, A. G. Beck-Sickinger and O. Seitz, *ChemBioChem*, 2021, **22**, 1717–1732.
- 8 I. C. Vreja, I. Nikić, F. Göttfert, M. Bates, K. Kröhnert, T. F. Outeiro, S. W. Hell, E. A. Lemke and S. O. Rizzoli, *ACS Nano*, 2015, **9**, 11034–11041.
- 9 H. Gelman, A. J. Wirth and M. Gruebele, *Biochemistry*, 2016, **55**, 1968–1976.
- 10 B. A. Griffin, S. R. Adams and R. Y. Tsien, *Science*, 1998, **281**, 269–272.
- 11 S. R. Adams, R. E. Campbell, L. A. Gross, B. R. Martin, G. K. Walkup, Y. Yao, J. Llopis and R. Y. Tsien, *J. Am. Chem. Soc.*, 2002, **124**, 6063–6076.
- 12 H. Nonaka, S.-h. Fujishima, S.-h. Uchinomiya, A. Ojida and I. Hamachi, *J. Am. Chem. Soc.*, 2010, **132**, 9301–9309.
- 13 I. Takahira, H. Fuchida, S. Tabata, N. Shindo, S. Uchinomiya, I. Hamachi and A. Ojida, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 2855–2858.
- 14 J. Wang, Y. Yu and J. Xia, *Bioconjugate Chem.*, 2014, **25**, 178–187.
- 15 U. Reinhardt, J. Lotze, S. Zernia, K. Mörl, A. G. Beck-Sickinger and O. Seitz, *Angew. Chem., Int. Ed.*, 2014, **53**, 10237–10241.
- 16 U. Reinhardt, J. Lotze, K. Morl, A. G. Beck-Sickinger and O. Seitz, *Bioconjugate Chem.*, 2015, **26**, 2106–2117.
- 17 A. Keppler, S. Gendreizig, T. Gronemeyer, H. Pick, H. Vogel and K. Johnsson, *Nat. Biotechnol.*, 2003, **21**, 86–89.
- 18 A. Gautier, A. Juillerat, C. Heinis, I. R. Corrêa, M. Kindermann, F. Beaufils and K. Johnsson, *Chem. Biol.*, 2008, **15**, 128–136.
- 19 G. V. Los, L. P. Encell, M. G. McDougall, D. D. Hartzell, N. Karassina, C. Zimprich, M. G. Wood, R. Learish, R. F. Ohana, M. Urh, D. Simpson, J. Mendez, K. Zimmerman, P. Otto, G. Vidugiris, J. Zhu, A. Darzins, D. H. Klaubert, R. F. Bulleit and K. V. Wood, *ACS Chem. Biol.*, 2008, **3**, 373–382.
- 20 I. Nikić, T. Plass, O. Schraidt, J. Szymański, J. A. G. Briggs, C. Schultz and E. A. Lemke, *Angew. Chem., Int. Ed.*, 2014, **53**, 2245–2249.
- 21 Y. Yano, A. Yano, S. Oishi, Y. Sugimoto, G. Tsujimoto, N. Fujii and K. Matsuzaki, *ACS Chem. Biol.*, 2008, **3**, 341–345.
- 22 J. K. Doh, S. J. Tobin and K. E. Beatty, *Biochemistry*, 2020, **59**, 3051–3059.
- 23 U. Reinhardt, J. Lotze, K. Mörl, A. G. Beck-Sickinger and O. Seitz, *Bioconjugate Chem.*, 2015, **26**, 2106–2117.
- 24 J. Lotze, P. Wolf, U. Reinhardt, O. Seitz, K. Mörl and A. G. Beck-Sickinger, *ACS Chem. Biol.*, 2018, **13**, 618–627.
- 25 G. C. Gavins, K. Gröger, M. D. Bartoschek, P. Wolf, A. G. Beck-Sickinger, S. Bultmann and O. Seitz, *Nat. Chem.*, 2021, **13**, 15–23.
- 26 A. Zieba, F. Ponten, M. Uhlén and U. Landegren, *Mod. Pathol.*, 2018, **31**, 253–263.
- 27 S. B. Yeldell and O. Seitz, *Chem. Soc. Rev.*, 2020, **49**, 6848–6865.
- 28 R. Jungmann, M. S. Avendaño, J. B. Woehrstein, M. Dai, W. M. Shih and P. Yin, *Nat. Methods*, 2014, **11**, 313–318.
- 29 G. Liszczak and T. W. Muir, *Angew. Chem., Int. Ed.*, 2019, **58**, 4144–4162.
- 30 H. Gradišar and R. Jerala, *J. Pept. Sci.*, 2011, **17**, 100–106.
- 31 V. Bertelsen and E. Stang, *Membranes*, 2014, **4**, 424–446.
- 32 A. Tomas, C. E. Futter and E. R. Eden, *Trends Cell Biol.*, 2014, **24**, 26–34.
- 33 X. Tan, P. F. Lambert, A. C. Rapraeger and R. A. Anderson, *Trends Cell Biol.*, 2016, **26**, 352–366.
- 34 B. S. Hendriks, L. K. Opresko, H. S. Wiley and D. Lauffenburger, *J. Biol. Chem.*, 2003, **278**, 23343–23351.
- 35 J. C. Stüber, F. Kast and A. Plückthun, *ACS Chem. Biol.*, 2019, **14**, 1154–1163.
- 36 T. Lebar, D. Lainšček, E. Merljak, J. Aupič and R. Jerala, *Nat. Chem. Biol.*, 2020, **16**, 513–519.

