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# Detection and mapping of haemoglobin variants in blood fingerprints by MALDI MS for suspect "profiling"†

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Over the past seven years Matrix Assisted Laser Desorption Ionisation Mass Spectrometry Profiling (MALDI MSP) and Imaging (MALDI MSI) have proven to be feasible tools for the detection of blood and its provenance in stains and fingerprints. However, whilst this capability as a confirmatory test addresses the primary questions at the scene of a violent crime, additional intelligence recoverable from blood can also prove important for investigations. A DNA profile is the most obvious and important example of such intelligence; however, it is not always suitable for identification purposes, depending on quantity, age and environmental conditions. Proteins are much more stable and determining the presence of haemoglobin variants in blood recovered at a crime scene may provide associative and possibly corroborating evidence on the presence of an individual at a particular location. This evidence gains more incriminatory value, the lower the incidence of the variant in a certain geographical area or population and may contribute to narrowing down the pool of suspects. In this study, a MALDI based mass spectrometric method has been developed and tested on six haemoglobin variants for their fast and reliable identification and mapping in blood fingerprints.

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## 1. Introduction

Over the past seven years, Matrix Assisted Laser Desorption Ionisation has proven its capability to detect and visualise blood in stains and fingerprints.<sup>1–6</sup> The compatibility with a range of blood enhancement techniques as well as its feasibility of application in aged blood (as old as 37 years<sup>5</sup>) confer the technique operational significance. Recently, it has been possible to provide additional information from this evidence by establishing its human or animal provenance (down to animal species) as well as discriminating this from other biological matrices<sup>4,6–9</sup> and in a pre-validation study.<sup>6</sup>

Whilst reliable detection of blood and the additional information on blood provenance address the first lines of a forensic inquiry, other intelligence may also be useful and, in some cases, crucial in informing investigations. A prime example of vital intelligence is offered by DNA profiling, used to identify

the perpetrator or to establish foul play in a murder inquiry without a body. However, DNA amplification, profiling and matching may not always be possible depending on initial DNA quantity, degradation, possible presence in a mixture and the existence of a record in DNA databases. Although only of an associative type, blood typing or detection of drugs or medications in a bloodstain or a blood fingerprint may be important to an investigation. Whilst a MALDI based method is available for the former,<sup>10,11</sup> there is no report in the literature covering the latter.

Additional intelligence on the presence of Hb variants (Hb var) could also be beneficial to narrow down the pool of suspects, depending on their incidence and occurrence in certain geographical areas or linked to the inhabiting multiethnic population (although migration alters the geographical occurrence over time). In 2001 Weatherall and Clegg<sup>12</sup> reported that mutations in genes encoding for haemoglobin occurred in the 7% of the global population. However, in some parts of the world, the sickle cell gene (HbS) can be present in up to 40% of the population, and the HbE variant can be present in excess of 60% of the population.<sup>12</sup> Both instances suggest that the detection of haemoglobin variants may be considered as useful intelligence to narrow down the pool of suspects. A genetic modification affecting the rate of synthesis gives rise to haemoglobin conferring a thalassemic trait; a genetic modification

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fication yielding a structural change in the form of amino acidic substitutions in the protein chain, generates Hb var which can also occur alongside 'normal' haemoglobin tetramers within some individuals. Most Hb var are the result of a single amino acid substitution on either the  $\alpha$  or  $\beta$  chain, although multiple substitutions can and do occur as well as insertion or deletion of amino acids. When variants occur as traits, an individual has one copy of an abnormal allele of the mutated haemoglobin gene and one copy of the 'normal' haemoglobin gene (heterozygous), or two copies of the abnormal allele (homozygous). For example, in the case of sickle cell haemoglobin, these are referred to as HbAS and HbSS respectively, where someone with the trait may not display the severe symptoms of sickle cell disease. Haemoglobin A<sup>0</sup> (HbA<sup>0</sup> or "normal" haemoglobin), is a tetramer containing two alpha subunits and two beta subunits ( $\alpha_2\beta_2$ ) and accounts for 97–98% of the total red blood cell Hb in a "normal" individual.<sup>13</sup>

According to the HbVar database (<https://globin.bx.psu.edu/globin/hbvar/>), regularly updated and part of the Globin Gene Server database [<http://globin.cse.psu.edu/>], there are a total of 1833 known Hb var of which 1393 are haemoglobin variants and 534 are thalassemias. Nine hundred and forty one entries arise from a modification to the  $\beta$  gene. Swedan *et al.* reported that more than 50 HbJ variants were known in 2008<sup>14</sup> but the Hb var database currently reports only 46 HbJ variants, possibly indicating that the overall number of Hb variants of 1831 may be a considerable under-estimation. However, incidence is what makes their detection interesting from a forensic perspective. Brunel *et al.* reported that in their laboratory, only 30 variants are detected in over 12 000 tests per annum.<sup>15</sup> In another example, as part of the North American "biometric screening programme" for the reduction and management of the healthcare costs and health risks of their employees, the Houston Methodist Hospital tested 13 913 individuals in 2.5 months.<sup>16</sup> Of this number, only 3.77% exhibited an Hb variant for a total of 11 different variants. Amongst these 524 employees, the HbS trait was present in 2.85% of the individuals, the HbSS disease in 0.03%, the HbC trait in 0.61%, the HbCC disease in 0.01% and the HbE trait in 0.13% of the individuals. Due to the potential contribution in forensics, this study has selected and investigated the six most common and clinically significant Hb variants, namely HbS, HbJ-Baltimore, HbC, HbE, HbD-Iran and HbD-Punjab/Los Angeles, all bearing modifications to the  $\beta$  globin chain. Prevalence in the UK varies by region due to the ethnic component of the distribution dependant on migration. As a reference, Sheffield, where the authors operate, has performed 10 196 Haemoglobinopathy screens last year, of which 464 (4.6%) presented a variant. According to Public Health England, Sheffield is considered high prevalence (>2.0% screen positive), although this percentage may be skewed upwards by the fact that Sheffield is a referral lab for many surrounding areas and beyond (Jason Eyre, personal communication).

Of these variants, HbS gives rise to the most important of the haemoglobinopathies (sickle cell anemia), where types C,

E and D Punjab/Los Angeles might lead to sickle cell anemia when associated with HbS.<sup>17,18</sup>

Sickle cell anemia is a life-threatening haemoglobinopathy characterised by low solubility of Hb which determines the rod-like shape of the red cells (sickle).<sup>19,20</sup> The low solubility of this variant arises from a substitution of valine for glutamic acid at the 6th amino acid position in the beta globin gene ( $\beta_6$  Glu  $\rightarrow$  Val). In 2008 Swedan *et al.* observed that in the US, the sickle trait has a prevalence of 8–10% among African Americans, and it is calculated that  $\sim$ 1/500 African Americans and 1/1000 Hispanics have sickle cell anemia.<sup>14</sup> HbJ-Baltimore was first reported in 1963 by Baglioni and Weatherall in a black American family<sup>21</sup> and it arises from a substitution of glycine for an aspartic acid at the 16th amino acid position ( $\beta_{16}$  Gly  $\rightarrow$  Asp). HbC and HbE are also frequently observed in some populations; the former has a high occurrence in Africa, while the latter is common in South East Asia.<sup>18,22</sup> The HbC variant is caused by the substitution of a glutamic acid for a lysine at the 6th position in the beta globin gene ( $\beta_6$  Glu  $\rightarrow$  Lys), whereas HbE arises from a substitution of a glutamic acid for a lysine at the 26th amino acid position in the beta globin gene ( $\beta_{26}$  Glu  $\rightarrow$  Lys). HbD Los Angeles was the 4th Hb var to be discovered and was described by Itano in 1951.<sup>23</sup> It arises from a substitution of a glutamic acid for a glutamine at the 121st position in the  $\beta$  globin chain ( $\beta_{121}$  Glu  $\rightarrow$  Gln). In 1962 Baglioni discovered that other HbD variants that had emerged had the same molecular composition as HbD-Los Angeles, one of these being HbD-Punjab.<sup>24</sup> Since then, this variant is denominated HbD-Punjab/Los Angeles. Whilst it is reported as one of the most common Hb var worldwide after HbS and HbC,<sup>25</sup> its prevalence varies according to geographical location; in the Northwest India (the Punjab region) it has a prevalence of 2%, dropping to 1% in Western India. This variant has also been detected in some European countries such as Italy, Belgium and Austria as well as Turkey.<sup>26–29</sup> For Italy, the findings cannot exclude either an independent origin or a spread from India due to migration.<sup>26</sup> For Belgium the latter is instead considered unlikely,<sup>27</sup> whereas for Austria a spread from Turkey is considered possible.<sup>28</sup> Whilst these variants are described as common in these four countries,<sup>25</sup> it does not mean that they have a high incidence. For example in the Belgium study, in 1986 it was reported that over the course of 9 years, 19 000 individuals had been screened for haemoglobin abnormalities and only nine unrelated carriers of HbD-Punjab/Los Angeles were discovered;<sup>26</sup> in Turkey, it occurs in 0.2% of the population.<sup>25</sup> Dass *et al.*<sup>30</sup> reported HbD-Iran as a rare variant which was first discovered in a family in central Iran in 1973.<sup>31</sup> This variant arises from a substitution of a glutamic acid for a glutamine at the 22<sup>nd</sup> amino acid position in the  $\beta$  globin chain ( $\beta_{22}$  Glu  $\rightarrow$  Lys). Table 1 summarises the modifications on the  $\beta$  globin chain for each of the variants of interest.

In healthcare, there are a number of techniques that are currently used to screen for Hb variants including cation exchange HPLC, tandem mass spectrometry, acid and alkaline gel electrophoresis (or isoelectric focusing electrophoresis),



**Table 1** Hb variants investigated in this study, nature and location of the amino acid mutation

Variant	HbA <sup>0</sup>	HbC	HbD-Punjab/Los Angeles	HbD-Iran	HbE	HbJ-Baltimore	HbS
Chain	$\alpha 2\beta 2$	$\alpha 2\beta^C 2$	$\alpha 2\beta^D 2$	$\alpha 2\beta^D 2$	$\alpha 2\beta^E 2$	$\alpha 2\beta^J 2$	$\alpha 2\beta^{S2}$
Amino acid modification	NA	Glu > Lys	Glu > Gln	Glu > Gln	Glu > Lys	Gly > Asp	Glu > Val
Modification position	NA	6 $\beta$ chain	121 $\beta$ chain	22 $\beta$ chain	26 $\beta$ chain	16 $\beta$ chain	6 $\beta$ chain

capillary zone electrophoresis and molecular methods, depending on the target of the screening,<sup>32,33</sup> with HPLC being the more commonly used technique across the UK.

Depending on the laboratories, a combination of the methods above is used to detect as many variants as possible. However, to the authors' knowledge no forensic screening for Hb variants has ever been reported in the literature so far. The techniques are not only generally time consuming but are also not applicable to blood fingermarks as they require destruction of the evidence (due to swabbing and extraction). In the work presented here, MALDI MSP and MSI have been investigated to evaluate the possibility to detect the six selected haemoglobin variants in blood and to visualise them in blood fingermarks. The use of diverse mass spectrometry instrumentation has shown that, whilst the highest sensitivity is required to detect (and map) the Hb var under investigation, MALDI MS is, within the system investigated, a suitable technique to recover this type of intelligence from blood. With further optimisation it could also be a valid alternative/addition to the current clinical diagnostics methods for Hb var screening.

## 2. Experimental

### 2.1 Materials

Trifluoroacetic acid (TFA),  $\alpha$ -cyano-4-hydroxycinnamic acid ( $\alpha$ -CHCA) phosphorous red and TLC sheets were purchased from Sigma Aldrich (Poole, UK). Acetonitrile, (ACN) was purchased from Fisher Scientific (Loughborough, UK). Lyophilised sequencing grade modified Trypsin was obtained from Promega in 20  $\mu$ g vials (Southampton, UK). RapiGest was obtained in 1 mg vials from Waters (Waters Corp, Manchester, UK). Double sided conductive tape was purchased from TAAB (Aldemastor, UK), Intellislide (glass slides) were made available by Bruker Daltonik (GmbH, Bremen, Germany). Indium tin oxide (ITO) slides were purchased from Sigma Aldrich (Poole, UK). Six Haemoglobin variants were investigated by MALDI MS, MS/MS and MSI namely, HbC, HbD-Iran, HbD-Punjab/Los Angeles, HbE, HbJ Baltimore and HbS. HbA<sup>0</sup> ("normal" Hb) was also included as a reference. Samples were available as standards (HbA<sup>0</sup>, and HbS were purchased from Sigma Aldrich (Dorset, UK) and a mixed Hb AFSC standard was purchased from Sebia (Lisses, France), as they were not commercially available individually, or as patients' blood. Patients' blood was provided by Sheffield Teaching Hospitals under the IRAS and the Sheffield Hallam University Ethics approvals 160418 and ER6558932 respectively. These samples were analysed using three types of MALDI Mass Spectrometers.

### 2.2 Instruments and instrumental conditions

Three sets of data were acquired from three MALDI mass spectrometers, namely the MALDI QToF Synapt G2 HDMS (Waters Corp. Manchester, UK), MALDI TOF/TOF rapifleX and MALDI qTOF timsTOF fleX (ttfleX, Bruker Daltonik GmbH, Bremen, Germany).

**Data acquisition: MALDI MSP.** The Synapt G2 HDMS is supplied with a neodymium-doped yttrium aluminium garnet (Nd:YAG) laser which was set at a repetition rate of 1 kHz. Data acquisition was performed over the  $m/z$  range 600–2500 in positive mode. A 0.5  $\mu$ L spot of saturated red phosphorus solution in ACN was used as the internal calibrant for each sample by acquiring a spectrum in the same acquisition instance as the sample. MALDI MS/MS spectra were obtained setting the resolution of the quadrupole at 4.4 and 11 for low mass (LM) and high mass (HM) respectively. Argon was used as the collision gas; with the cooling gas set at 10.0 a.u. and the trap gas at 7.5 a.u. The trap collision energy was ramped between 80 and 100 eV. The MALDI rapifleX is equipped with a Nd:YAG 355 nm SmartBeamTM 3D laser operating at a repetition rate of 10 kHz. The instrument was calibrated using a Bruker peptide calibration standard mono mix between  $m/z$  1046–2455. MS spectra were obtained in reflectron positive mode in the mass range  $m/z$  600–3200, with laser power of 35 a.u. The MALDI ttffeX mass spectrometer is equipped with a Smartbeam 3D laser operated at a repetition rate of 10 kHz. When using the ttffeX for MALDI MSP, the instrument was calibrated in positive mode over the  $m/z$  range 100–3000 using red phosphorus. In MS mode, five hundred laser shots per spectrum were fired and typically 10 acquired single spectra were summed. MS/MS spectra were obtained by ramping the collision energy between 80–130 eV. Five thousand laser shot per single spectra were acquired and typically 10 single spectra were summed.

**Data acquisition: MALDI MSI.** Imaging data were acquired on the MALDI Synapt G2 HDMS and on a MALDI ttffeX mass spectrometer.

In the former case, side by side blood stain images were acquired in the mass range  $m/z$  600–2000, setting the laser power at 250 a.u. When using the MALDI ttffeX mass spectrometer, blood fingermarks (including overlapping) were acquired at a lateral resolution of 50  $\mu$ m, a laser power of 65% and accumulating 600 laser shots per pixel. The instrument was previously calibrated in ESI mode using Agilent low concentration calibration mix (G1969-85000) and operated in positive mode and in full scan over the  $m/z$  range 100–3000.

**Data processing.** *In silico* determination of the proteotypic peptide sequences (across the six variants and the HbA<sup>0</sup>) was



performed by preliminarily identifying the amino acid mutation in the  $\beta$ -globin chain using a database of haemoglobin variants accessible at <https://globin.bx.psu.edu/globin/Hbvar/>. Subsequently, an *in silico* digestion with trypsin was performed using Uniprot KB (<https://www.uniprot.org/>). Here the  $\beta$  haemoglobin chain sequence was *in silico* digested setting the maximum number of missed cleavages to 2 and to yield  $MH^+$  monoisotopic peptides up to  $m/z$  3000. The results were navigated by identifying the peptides with up to 3 cleavages sites containing the amino acid modification for each variant. Finally proteotypic peptides were identified through filtering out peptides with the same theoretical  $m/z$  and those differing by less than 15 ppm (Table 2).

MALDI MS and MS/MS spectra acquired from all three MALDI Mass Spectrometers were exported to txt files and processed using mMass, an open source mass spectrometry tool.<sup>34,35</sup> For the Synapt G2 HDMS and rapifleX data, spectra were batch processed by smoothing using the Savitzky–Golay algorithm, a  $m/z$  window size of 0.2 and 1 smoothing cycle; peak picking was also performed by setting the S/N threshold at 4.0, the absolute and relative intensity threshold at 0.0 a.u. and 0.5 respectively and the picking height at 100. MALDI MS and MS/MS spectra acquired using the ttfleX instrument were preliminarily processed using DataAnalysis software (Bruker Daltonik GmbH, Bremen, Germany) and then converted into txt file to be further processed in mMass where no further smoothing was applied; the peak picking was performed for all ttfleX spectra by setting the S/N threshold at 20.0, the absolute and relative intensity threshold at 0.0 a.u. and 0.5 respectively and the picking height at 98 a.u. Haemoglobin variant proteotypic peptides were used to build a database in mMass for the automatic assignment of Hb var in MS mode. Depending on the instrument employed and calibration achieved, a different mass tolerance for assignment was employed:  $\pm 13$  ppm for Synapt,  $\pm 11$  pm for rapifleX and  $\pm 4$  ppm for ttfleX MS spectra.

ttfleX MALDI MS images were generated using SCiSLab 2021a software (Bruker Daltonik, Bremen, Germany) with a mass accuracy of  $\pm 15$  ppm, using no de-noising (to avoid loss of ridge detail) and normalising against RMS. Brightness and contrast were adjusted to optimal values to improve clarity of ridge detail. Synapt MALDI MS images were generated using the HDI software (Waters Corp Manchester) and normalised against the total ion current.

### 2.3 Trypsin digestion

For MALDI MS Profiling experiments, all in-solution trypsin digestions were performed as follows: 5  $\mu$ L of blood sample were dissolved in 1 mL of  $H_2O$ . Ten  $\mu$ L of diluted blood sample was added to 40  $\mu$ L 40 mM AmBic. Nine  $\mu$ L of trypsin at 20  $\mu$ g  $mL^{-1}$  (containing 0.1% RapiGest) was added and the mixture was incubated at 37  $^{\circ}C$  for 1 h. Two  $\mu$ L 5% TFA were then added to stop the proteolysis.

For MALDI MSI analysis on the Synapt G2 HDMS instrument, 9 layers of trypsin gold at 100  $\mu$ g  $mL^{-1}$  (containing 0.1% RapiGest) were sprayed over the blood stains using a SunCollect matrix sprayer (SunChrom, Germany), at a flow rate of 2  $\mu$ L  $min^{-1}$ . The stains were incubated in a sealed container containing 50 : 50 MeOH :  $H_2O$  at 37  $^{\circ}C$  for 3 h. The samples were then sprayed with 5 layers of  $\alpha$ -CHCA matrix at 5 mg  $mL^{-1}$  in 70 : 30 ACN:0.5% TFA at 4  $\mu$ L  $min^{-1}$ .

Blood contaminated fingerprints were prepared by depositing 10  $\mu$ L of patient blood samples (containing either one of the Hb variants investigated) on the fingertip of a in house 3D printed silicon fingertip and quickly dragging the blood around with the pipette tip until covering the mark. The fingertip was subsequently pressed on an Intellislide to generate a Hb var contaminated blood fingerprint. An additional sample was prepared by pressing a silicon fingertip contaminated with HbC blood and a silicon fingertip contaminated by HbJ-Baltimore blood on an Intellislide to generate overlapping marks. These marks were subsequently enhanced using Acid

**Table 2** Expected proteotypic peptides for the six haemoglobin variants selected in this study used to build the mMass database for automatic peptide assignment. The bold font indicates the amino acid mutation

$\beta$ Hb variant and proteotypic peptide	Proteotypic peptide sequence	Expected $m/z$
HbC (2–18)	VHLTP <b>KE</b> SAVTALW GK	1865.0646
HbC (2–9)	VHLTP <b>KE</b> K	951.5626
HbC (2–7)	VHLTP <b>K</b>	694.4251
HbS (2–9)	VHLTP <b>VE</b> K	922.5361
HbS(2–18)	VHLTP <b>VE</b> SAVTALW GK	1836.0381
HbE (19–41)	VNVDEVGGKALGRLLVVYPW TQR	2569.4253
HbE (10–31)	SAVTALW GKVNVD <b>EV</b> GGKAL GR	2227.2197
HbE (10–27)	SAVTALW GKVNVD <b>EV</b> GGK	1829.9760
HbE (19–31)	VNVDEVGGKALGR	1313.7175
HbE (19–27)	VNVDEVGG <b>K</b>	916.4739
HbD-Punjab/Los Angeles (122–145)	QFTPPVQAAYQKV <b>V</b> AGVANA LAHK	2508.3726
HbD-Punjab/Los Angeles (122–147)	QFTPPVQAAYQKV <b>V</b> AGVANA LAHKYH	2826.5051
HbD-Punjab/Los Angeles (122–133)	QFTPPVQAAYQ <b>K</b>	1377.7164
HbD-Iran (19–41)	VNV <b>D</b> QVGGEALGRLLVVYPW TQR	2569.3889
HbD-Iran (10–31)	SAVTALW GKVNVD <b>Q</b> VGGGEAL GR	2227.1831
HbD-Iran (19–31)	VNV <b>D</b> QVGGEALGR	1313.6812
HbJ-Baltimore (10–31)	SAVTALW <b>D</b> KVNVD <b>EV</b> GGGEAL GR	2286.1726
HbJ-Baltimore (1–18)	MVHLTP <b>EE</b> SAVTALW <b>D</b> K	2055.0583
HbJ-Baltimore (10–18)	SAVTALW <b>D</b> K	990.5259





Black 1, a commonly used crime lab technique for blood enhancement in fingerprints, according to the methanol-based protocol described by the Home Office edited fingerprint visualisation manual.<sup>36</sup> Sequencing grade Trypsin was reconstituted in 20 mM of ammonium bicarbonate to a final concentration of 25 ng  $\mu\text{L}^{-1}$ . Trypsin was deposited using the HTX M3 sprayer (HTX Technologies LLC, US) at a temperature of 30 °C and a pressure of 10 psi. The flow rate and the nozzle height were set at 30  $\mu\text{L min}^{-1}$  and 40 mm respectively. A total of 8 layers of trypsin were deposited at a velocity of 750 mm  $\text{min}^{-1}$  and using a criss-cross pattern with 2 mm track spacing. Following trypsin deposition, the glass slides containing the fingerprints were placed on top of two glass tubes or 15 mL Sarstedt tubes in a lunch box. Ten  $\mu\text{L}$  of water were pipetted on top of the glass slide but far away from the fingerprints. The lunch box contained 40 mL of a saturated  $\text{K}_2\text{SO}_4$  solution (75 g in 500 mL water) and a further spoonful of  $\text{K}_2\text{SO}_4$  was added before sealing the lunch box. The slides were incubated for 2 h at 50 °C. Overlapping fingerprints were submitted to MALDI MSI on a ttfleX mass spectrometer.

## 2.4 Matrix deposition

For MALDI MS and MS/MS data acquisition on the Synapt G2 HDMS,  $\alpha$ -cyano 4 hydroxycinnamic acid ( $\alpha$ -CHCA) was prepared at a concentration of 5 mg  $\text{mL}^{-1}$  in 70:30 ACN:0.5% TFA, and 0.5  $\mu\text{L}$  was pipette mixed with the sample on a MALDI target plate.

For MALDI MS on the rapifleX,  $\alpha$ -CHCA was prepared at 5 mg  $\text{mL}^{-1}$  in 70:30 ACN:0.5% TFA and 0.5  $\mu\text{L}$  was spotted on top of the sample spots that had been allowed to dry in a vacuum desiccator. For MALDI MS and MS/MS on the ttfleX mass spectrometer, haemoglobin standard and patient blood digests were mixed 1:1 with  $\alpha$ -CHCA solution saturated in 30/70 ACN/0.1%TFA. One  $\mu\text{L}$  of this solution was applied on an Anchor 800 target plate. After drying (crystallization) the MALDI spot was washed once with 3  $\mu\text{L}$  water containing 0.1% TFA as a clean-up procedure.

For MALDI MSI on the Synapt, 5 layers of  $\alpha$ -CHCA at 5 mg  $\text{mL}^{-1}$  in 70:30 ACN:0.5% TFA were sprayed at 2  $\mu\text{L min}^{-1}$ .

For MALDI MSI on the ttfleX, a solution of 10 mg  $\text{mL}^{-1}$  of  $\alpha$ -CHCA, in 70:30 ACN:TFAaq 0.2%, was sprayed using a HTX M3 sprayer (HTX Technologies LLC, US) at 75 °C, 10 psi pressure and at a flow rate of 120  $\mu\text{L min}^{-1}$ . The nozzle height was set at 40 mm. Four layers of matrix were sprayed with 2 mm track spacing and a velocity of 1200 mm  $\text{min}^{-1}$ .

## 3. Results and discussion

In this study, the first of its kind, a combined proteomics-MALDI MSP/MSI based approach was developed to quickly identify (in bloodstains) and map (in blood fingerprints) six most common haemoglobin variants (Hb var), HbS, HbC, HbD-Punjab/Los Angeles, HbD-Iran, HbE and HbJ-Baltimore. Hb var screening belongs to clinical diagnostic settings.

However, it has been pursued here through a forensic perspective as a means to provide additional intelligence from examination of blood evidence. A relatively large sample set of patients' blood, containing different Hb var was used for method development and investigated on the MALDI Synapt G2 system, for a total of 44 samples which were spotted and analysed in triplicate. The samples were received in different batches at different times over the course of 2 years in which instrumental conditions may have varied. Although here the 44 samples are discussed as a whole, dates of shipment have been highlighted. The identity of the samples was blind to the analyst processing the data until the Hb var assignment was completed. Table S1† reports a summary of the putative detection of the Hb variants during method development on the Synapt G2 HDMS instrument.

The HbD-Punjab/Los Angeles variant was detected only in one out of the seven supplied patients' samples and the HbJ-Baltimore variant was detected in the only patient's sample supplied of this kind. The three normal haemoglobin samples ( $\text{HbA}^0$ ) and the two HbD-Iran samples were always correctly labelled as "no variant detection" and HbD-Iran respectively.

Nine out of ten patients' samples were correctly labelled with the HbS variant reflecting the high ionisability of the proteotypic peptide 2–9. The next best result was achieved for the HbC variant which was correctly identified in 8/11 patients' samples. Finally, of the ten HbE patients' samples, six samples were correctly labelled as HbE. It is important to note that as quantification is not performed, it is not possible to establish whether the variant detected indicates that a patient is heterozygous or homozygous for the variant identified. For example, in the case of sickle cell anaemia heterozygous HbS, the variant is indicated as HbAS whereas the pathology as HbSS. Our results would report HbS, regardless of whether the patients have a trait (HbAS) or the actual pathology (HbSS).

Table S2† reports the accurate list of proteotypic peptides  $m/z$  assignments (using the custom built mMass library shown in Fig. 2) relating to 1 replicate per samples only, for readability.

An additional batch of nine patients' blood samples was received and submitted to the MALDI Synapt (3 replicates per sample) as well as to additional two state of the art MALDI mass Spectrometers, namely rapifleX (3 replicates per sample) and timsTOF fleX (ttfleX) (2 replicates per sample) to ascertain whether correct putative identification frequency could be improved as well as confirming identity using tandem MS. Although it is difficult to directly compare on paper the sensitivities of the three MALDI instruments based on the manufacturer specifications, these are all very sensitive instruments; the sensitivity specifications are all based on the measurement of the same peak at  $m/z$  1570.7 relating to [Glu1]-Fibrinopeptide B (GluFib) as follows: (i) Synapt: 10 fmol GluFib at S/N > 90:1; (ii) rapifleX: 250 amol GluFib at S/N  $\geq$  200:1 shown on Bruker AnchorChip™ target with 2000 laser shots; (iii) ttfleX: S/N > 100:1 for 250 amol GluFib on target S/N with 2000 laser shots. MALDI MS analyses were conducted



**Table 3** MALDI MS of patients' blood samples containing different variants. The same set of samples was analysed on three different instruments, namely the timsTOF fleX (ttfleX), the rapifleX and the Synapt G2 HDMS. Cells are coloured in green if the correct variant was identified; cells are coloured in red if the incorrect Hb var or no Hb var was identified. Spectra were run in duplicates on the ttfleX and in triplicates on the rapifleX and on the Synapt. In the table only one replicate *m/z* per proteotypic peptide detected is reported having the lowest relative error

ttfleX				rapifleX			Synapt			
Sample no.	Experim. ID	proteotypic peptide <i>m/z</i>	Mass Accuracy (ppm)	Experim. ID	proteotypic peptide <i>m/z</i>	Mass Accuracy (ppm)	Experim. ID	proteotypic peptide <i>m/z</i>	Mass Accuracy (ppm)	True ID
843	HbJ-Baltimore	2286.172	-0.1	HbJ-Baltimore	2286.171	-0.4	HbJ-Baltimore	2286.165	3.6	HbJ-Baltimore
844	HbC	694.425 951.559	0 3.1	HbC	694.426	1.2	HbC	951.57	8.2	HbC
845	HbC	1865.057	-3.9	HbC	694.426 951.565	10.5 2.6	HbC	694.422 951.561	-4.2 -1.1	HbC
846	HbE	1829.976 2227.217	0.3 -1.1	HbD-Iran	1313.68	-0.6	HbE	916.468 1829.956 1313.706	-4.3 -5.6 -8.6	HbE
847	HbE	1829.976	0.3	HbE	1313.708 2227.221	7.1 0.7	HbE	916.471 1829.971	-2.2 -2.4	HbE
848	HbD-Punjab	1377.716	-1.3	no variant	NA	NA	no variant	NA	NA	HbD-Punjab
849	no variant	NA	NA	no variant	NA	NA	no variant	NA	NA	HbD-Punjab
850	HbJ-Baltimore	2286.172	-3.2	HbJ-Baltimore	2286.166	-3.5	HbJ-Baltimore	2286.145	12.1	HbJ-Baltimore
851	HbD-Iran	1313.681 2227.183	-0.2 -0.1	HbD-Iran	1313.676 2227.18	-3.7 -1.1	HbD-Iran	1313.674 2227.190	-4.9 3.2	HbD-Iran

once again in a blind fashion and using the custom built mMass database of proteotypic peptides (Table 2). Isopro software, based on the Yergey algorithm,<sup>37</sup> was used to calculate the molecular formula of each peptide, given the amino acidic sequence, to build the mMass library. Table 3 reports the Hb var identification for each patient sample on the three instruments (only one replicate per instrument is shown). As it can be seen from Table 3, in terms of correct identifications, the ttfleX performed better than the rapifleX and the Synapt with 8/9 correct identifications and mass accuracies ranging between  $-3.2$  and  $3.1$  ppm, versus 6/9 and 7/9 correct identifications using the rapifleX and the Synapt respectively; taking into account one replicate per sample with the highest mass accuracy of Hb var identifying signals, mass accuracies on the rapifleX and Synapt ranged between  $-3.7$  and  $10.5$  ppm and  $-8.6$  and  $12.1$  ppm respectively. The Synapt performed better than the rapifleX in terms of number of correct putative IDs. Sample 846 was correctly identified as HbE both on the ttfleX and the Synapt, whereas the rapifleX incorrectly indicated the presence of HbD-Iran. Sample 848 was correctly identified as HbD-Punjab on the ttfleX, whereas the other two instruments were unable to detect any variant. However, when sample 849, containing again the HbD-Punjab variant, was analysed, neither of the mass spectrometers were able to detect it and a result of "no Hb variant" was yielded; all the assignments are reported in Table 3.

The two variants were quantified in patients 848 and 849 at 41.3% and 41.6%, therefore there was no appreciable difference that can explain lack of detection of this variant in patient 849. However, when preliminary Synapt experiments were performed, HbD-Punjab was the least successfully detected variant with only 1/7 samples correctly identified and 5/7 samples showing no HbD-Punjab/Los Angeles related signals (Table S1†).

It is possible that the HbD-Punjab/Los Angeles proteotypic peptides are generated below the limits of detection due to a combination of proteolytic concentration and ionisation efficiency.

*In situ* MALDI MS/MS analyses are generally and notoriously challenging for non-single analyte samples; this is why shotgun proteomics of complex samples (such as blood) is more frequently conducted by LC MS/MS, where the separation dimension improves on the selection of the precursor ion. Given the observed performances, *in situ* MALDI MS/MS analyses were nevertheless attempted in a comparison between the Bruker Daltonik higher end instrument, the ttfleX, and the Waters Synapt G2. The ttfleX enabled confirmation *via* MS/MS of all the variants except for HbE in patient 845 as the precursor ion could not be isolated and for patient 849, yielding no variants signals. For the reminder of the samples, fragment ions were assigned in high mass accuracy, ranging between  $-5$  and  $4.9$  ppm (across samples and ion fragments) (Table S3†).



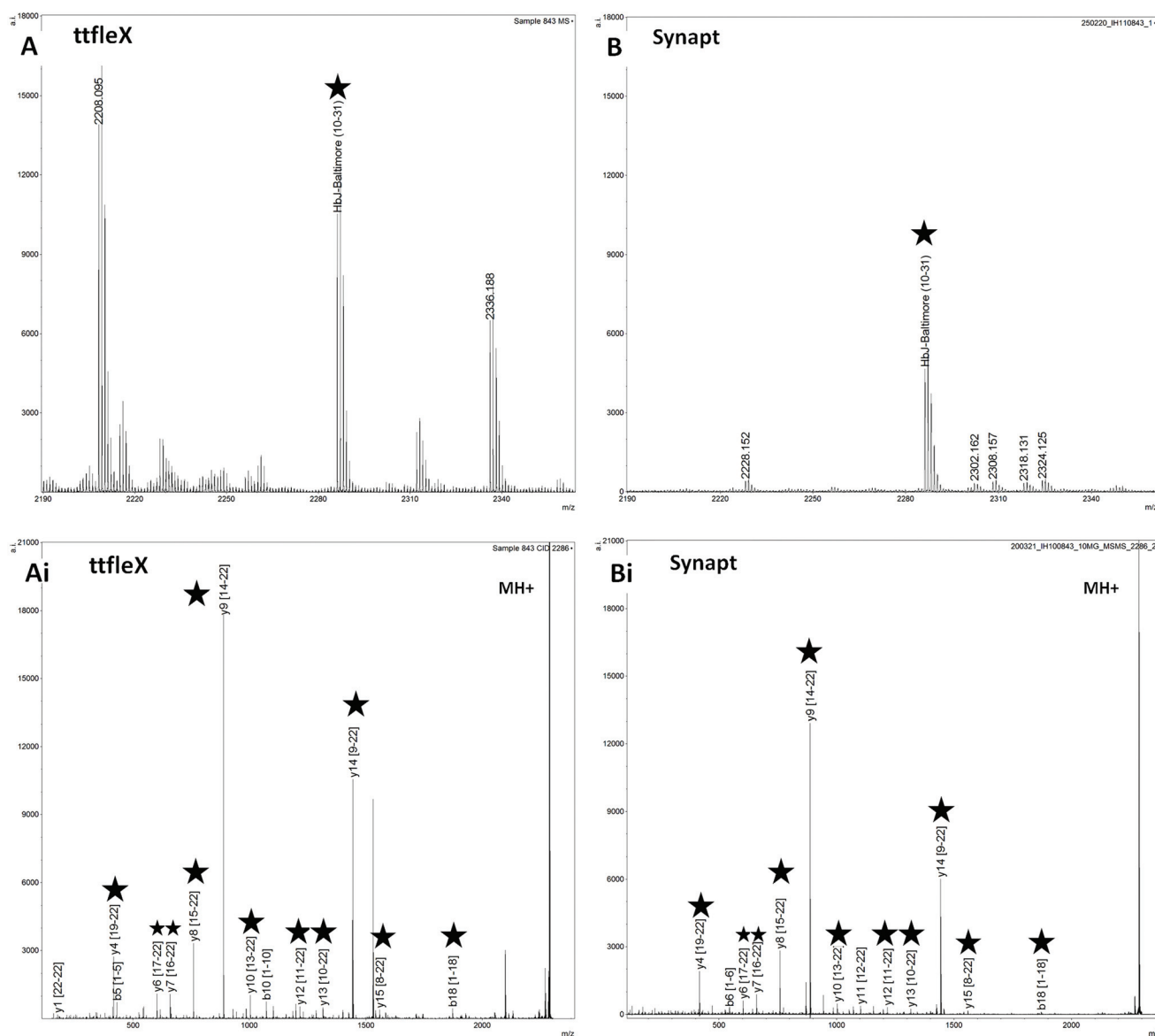
The Synapt G2 yielded reliable confirmatory MS/MS spectra for samples 843, 850 and 851 with a mass accuracy ranging between  $-18.5$  and  $7.8$  ppm (across samples and ion fragments) (Table S3†).

As an example, Fig. 1 shows a comparison of MS and MS/MS spectra obtained on the ttflEX and on the Synapt for patient sample 843, with this being the sample for which the Synapt MS/MS analyses yielded the highest number of ion fragments. Both in MS mode (Fig. 1A and B) and in MS/MS mode (Fig. 1Ai, Bi), the ttflEX exhibited superior sensitivity.

In MS mode, the HbJ-Baltimore proteotypic peptide labelled with a star was detected as more than twice as intense on the ttflEX; in MS/MS mode, 14 ion fragments could be

assigned using ttflEX *versus* 13 on the Synapt, with 11 ion fragments in common. Although this difference is not meaningful in terms of performance, the ion fragments in common were generally detected with higher intensity on the ttflEX. A final observation is noteworthy in that the ttflEX detected the peptide at nominal  $m/z$  2227 for two different samples (846 and 851) and was able to easily discriminate HbE (10–31) (sample 846) from HbD Iran (10–31) (sample 851) at a theoretical  $m/z$  of 2227.2197 and 2227.1831 respectively. Table S3† reports all of the ion fragments identified within the MS/MS spectra acquired on the ttflEX and on the Synapt.

*In silico* methods were employed to establish both protein and species specificity. In other words, it was assessed



**Fig. 1** MALDI MS and MS/MS analysis of sample 843 on ttflEX (A, Ai) and Synapt (B, Bi); Ai and Bi show the MALDI MS/MS spectra of the ion at nominal  $m/z$  2286 (HbJ-Baltimore (10–31) SAVTALWDKVNVDVGEALGR) leading to the confirmation of the HbJ-Baltimore variant. The ttflEX instrument exhibited a superior sensitivity both in MS mode (A, B – star symbols indicate the proteotypic peptide (HbJ-Baltimore (10–31)) and in MS/MS mode (Ai, Bi – star symbol indicates the fragments detected in common).



whether or not the proteotypic peptides employed to detect each of the Hb var: (1) could only derive from those human  $\beta$  Haemoglobin variants and (2) are specific to human. For this purpose, the amino acid sequences of all proteotypic peptides were screened against the UniProtKB/Swiss-Prot protein knowledgebase release 2021\_01 containing 564 277 manually annotated and reviewed proteins from 14 014 species. Using the peptide search tool against the entirety of the taxonomy included in the database, searches were run only against the manually annotated and reviewed proteins. Table S4† reports the summary of these searches.

In brief, using the “peptide search” tool, only 8/19 proteotypic peptide sequences yielded a hit and all hits were against human haemoglobin except for HbC (2–7) peptide. This sequence was matched to proteins in Fungi, metazoa, bacteria and human; interestingly, this sequence in human matches to the thyroid adenoma-associated protein. This protein is expressed in pancreas, adrenal medulla, thyroid, adrenal cortex, testis, thymus, small intestine and stomach and as such it seems unlikely to be detected in blood. However, to dissipate any doubt on the identity of the detected peptide, the additional detection of the HbC (2–9) could be checked to confirm the presence of HbC. The Blast tool was also used in parallel to check homologies and HbC (2–7) yielded no hits having a 100% homology. This was the case also for all the proteotypic peptides except for HbE (10–27) and HbJ-Baltimore (10–31). In the former case, a 100% homology with  $\beta$  haemoglobin was found for both human and primates and in the latter case a 100% homology with  $\beta$  haemoglobin was found for the species *Gorilla gorilla*.

Whilst detection of Hb var in blood stains and/or blood fingermarks may contribute to narrowing down the pool of suspects, visualisation of these variants onto the fingerprint ridge pattern provides important and direct associative evidence. A first MALDI MS imaging experiment was performed aiming at

the visualisation of Hb var and differentiation in the presence of blood of different origin. Initially, two bloodstains from HbS and normal HbA<sup>0</sup> patients' blood were deposited side by side and imaged on a MALDI Synapt G2 as a proof of concept. Fig. 2 shows the visualisation of the aspecific  $\alpha$ -Hb peptide at  $m/z$  1529.68 which was present in both stains as expected.

In contrast, the HbS proteotypic peptide at  $m/z$  922.48 was only visible in the HbS bloodstain and well discriminated from the stain containing the normal HbA<sup>0</sup> variant, as both the MALDI images and the spectra extracted from the regions of interest of patients 1 and 2 show (Fig. 2B). A subsequent attempt to image overlapping HbS/HbA<sup>0</sup> fingermarks failed on this mass spectrometer. For this reason, blood fingermarks were produced using patient blood samples containing the six variants under investigation and imaged using the ttflex MALDI system. MALDI MSI data reflected at large the data obtained through MALDI MSP using the same instrument; generally, Hb var proteotypic peptides that were detected in MS mode and confirmed by MS/MS, were also mapped onto the ridge pattern of the mark and contributed to some degree of ridge detail and *minutiae* visualisation. In particular Fig. 3 reports the distribution maps of the proteotypic peptides HbC (2–7) at  $m/z$  694.420 (i), HbC (2–9) at  $m/z$  951.563 (ii), HbE (10–27) at  $m/z$  1829.976 (iii), HbE (10–31) at  $m/z$  2227.220 (iv), HbD-Punjab/Los Angeles (121–133) at  $m/z$  1377.719 (v), HbD-Iran (19–31) at  $m/z$  1313.687 (vi), HbD-Iran (10–31) at  $m/z$  2227.183 (vii), HbJ-Baltimore (10–31) at  $m/z$  2286.173 (viii) and HbS (2–9) at  $m/z$  922.536 (ix), within a mass accuracy of  $\pm 10$  ppm.

The HbS variant proteotypic peptide provided the most ridge flow and *minutiae* visualisation, in line with the better ionisation efficiency amongst all other Hb variant proteotypic peptides observed in MALDI MS mode. Other Hb var proteotypic peptides were visualised on the fingerprint samples (largely in a “dotted” fashion) but they are not reported in

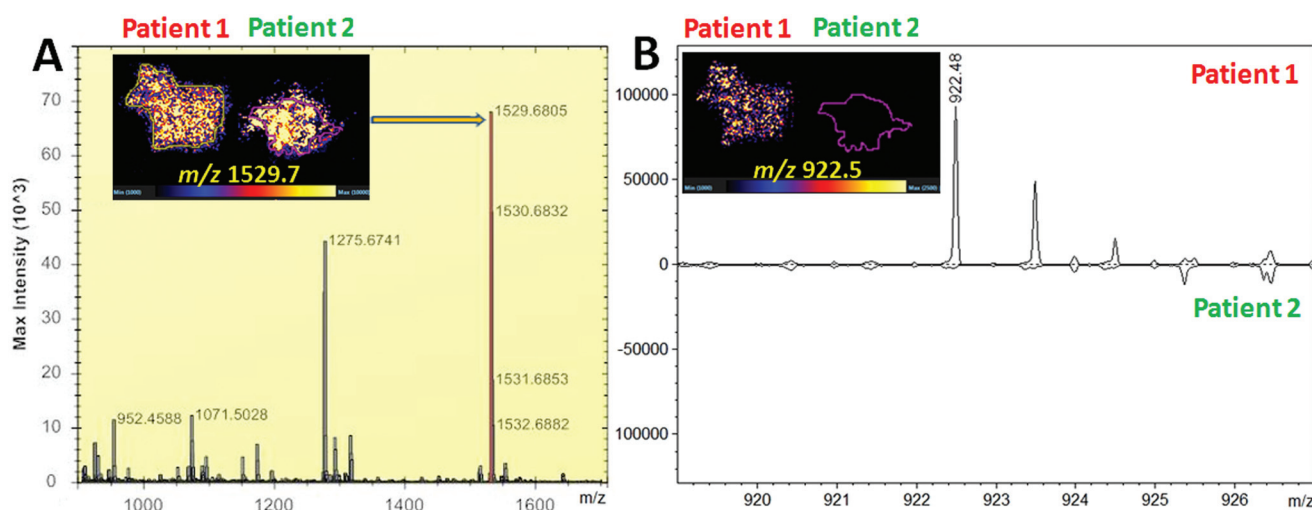
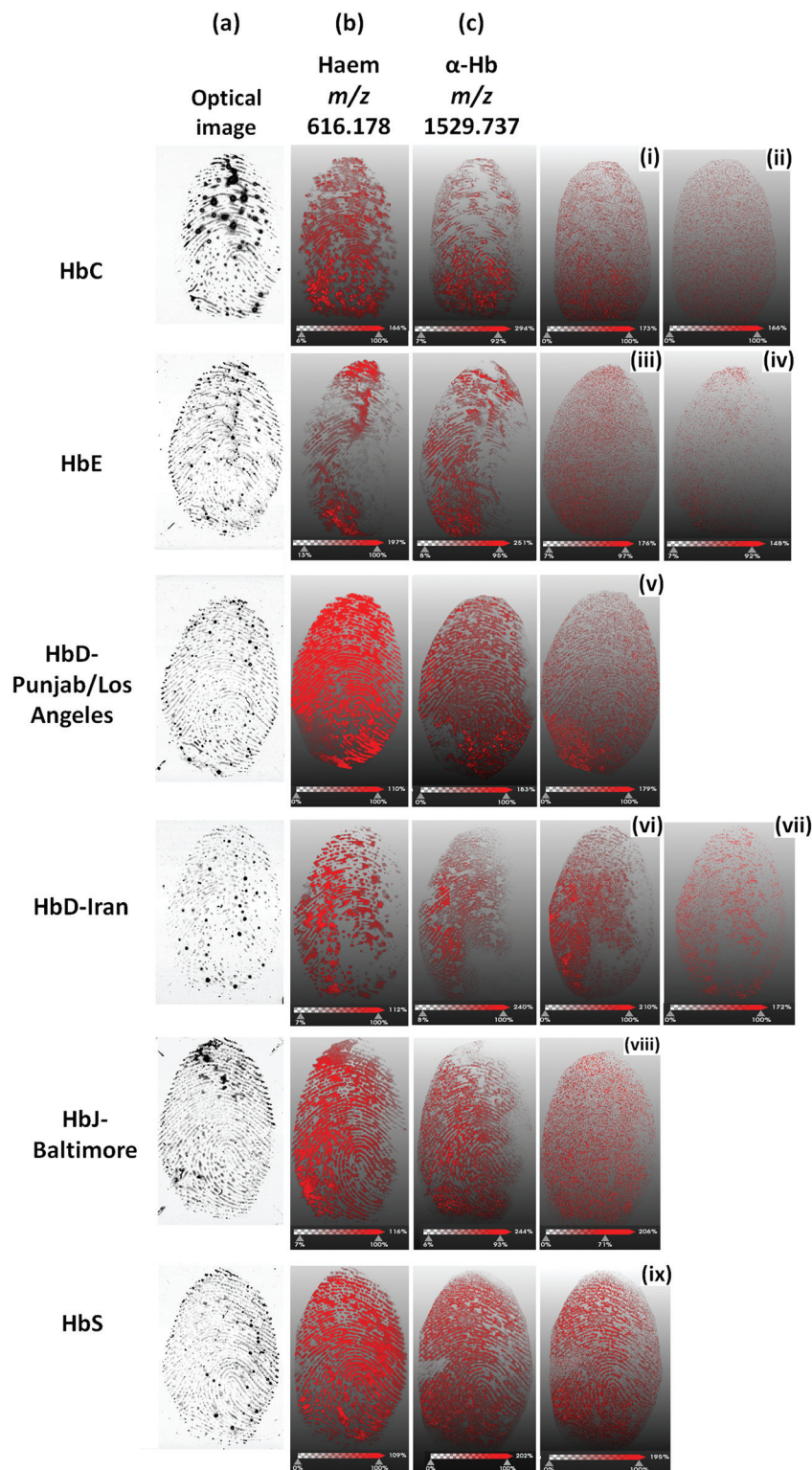


Fig. 2 MALDI MS imaging of two bloodstains from HbS and HbA<sup>0</sup> patients (patients 1 and 2 respectively) on the Synapt G2 HDMS system. Panel A shows visualisation of the aspecific  $\alpha$ -Hb peptide at  $m/z$  1529.680 in both stains as expected. Panel B shows the presence of the HbS proteotypic peptide at  $m/z$  922.48 only in the bloodstain of the HbS patient as also confirmed in the ROIs extracted spectra.







**Fig. 3** MALDI MS images of Hb var proteotypic peptides from silicon fingertips contaminated with patients' blood. (a) Optical image of the blood mark; (b) MALDI MS images of haem at  $m/z$  616.178; (c) MALDI MS images of  $\alpha$ -haemoglobin peptides at nominal  $m/z$  1530; ((i)–(ix)) MALDI MS images of proteotypic Hb var peptides detected and confirmed in MALDI MS and MS/MS mode. Proteotypic peptides are shown for HbC (2–7) at  $m/z$  694.420 (i), HbC (2–9) at  $m/z$  951.563 (ii), HbE (10–27) at  $m/z$  1829.976 (iii), HbE (10–31) at  $m/z$  2227.220 (iv), HbD-Punjab/Los Angeles (121–133) at  $m/z$  1377.719 (v), HbD-Iran (19–31) at  $m/z$  1313.687 (vi), HbD-Iran (10–31) at  $m/z$  2227.183 (vii), HbJ-Baltimore (10–31) at  $m/z$  2286.173 (viii) and HbS (2–9) at  $m/z$  922.536 (ix), within a mass accuracy of  $\pm 10$  ppm.



Fig. 3 as they were undetected in Profiling mode. It is interesting to note that the highest quality/quantity of ridge detail deriving from Hb var proteotypic peptides, was provided by HbS and HbD-Punjab/Los Angeles blood generated fingerprints. These prints correspond to the optical images showing the faintest presence of blood. In other instances, where optical images clearly and strongly showed the presence of blood, the corresponding MALDI images of Hb var peptides were fainter and exhibited a poorer ridge coverage.

Indeed, there is an optimal trypsin/substrate ratio and evidently in the poorer quality MS images, the substrate (Hb) is in excess and falling out of the optimal ratio with the enzyme, as opposed to the ratio for HbS and HbD-Punjab/Los Angeles.

The Hb-Baltimore MS image does not follow this trend; although the corresponding optical image also shows a very weak presence of blood, the MALDI MS image also generates poor quality and clarity of ridge detail. However, this instance can be explained by the generally lower ionisation efficiency of the HbJ-Baltimore proteotypic peptide, as also observed in the profiling analyses; an additional hypothesis is that the amount of HbJ-Baltimore sits outside the optimal trypsin/substrate ratio where trypsin is, in this instance, present in excess (leading to more auto-proteolysis events).

Fig. 3 also reports for each Hb var image, the corresponding haem at nominal  $m/z$  616 and the aspecific  $\alpha$ Hb peptide at nominal  $m/z$  1530. These additional images show that, in terms of fingerprint images, the haem group performs better than Hb deriving peptides yielding the most complete and continuous ridge pattern with the most *minutiae*. In practice, this occurrence suggests that when examining a blood mark by MALDI MSI, the ridge pattern should be preferably reconstructed by visualising haem whereas the presence of the variants could be visualised and superimposed on the haem mass image to provide additional intelligence on the suspect. Interestingly, the HbD-Punjab Los Angeles (121–133) peptide localised with a

higher intensity in the left bottom corner area, exactly where the signals of haem and  $\alpha$ -Hb peptide at  $m/z$  616.178 and 1529.737 respectively are almost absent. It is speculated that this proteotypic peptide experiences ion suppression exerted by haem and the  $\alpha$ -Hb peptide and, therefore, ionises better in those areas where these signals are much less intense or absent.

It is also important to note that, although better than the Hb peptides, the quality of the haem image reflects that of a non-ideal fingerprint ridge pattern ("non-textbook"), with smudged areas and interrupted ridge flow. This instance ultimately reflects (i) the imperfections of the fingertip surface of 3D printed silicon finger, (ii) the inhomogeneity of the coating of the silicon fingertip with blood and (iii) the difficulty of controlling the pressure and angle when using this artefact to generate blood fingerprints. Notwithstanding the challenges, silicon fingertips have to be employed instead of human fingertips due to blood being a biohazard. However, in real crime scenes, blood fingerprints are also less than ideal, with blood pooling and partial or discontinuous ridge detail due to the careless and unintentional deposition. Altogether, the imaging data presented here demonstrate the ability to map the six Hb variants investigated.

A subsequent experiment was conducted in which two blood fingerprints were overlapped during deposition; in particular one fingerprint was generated using patient blood containing the HbC variant and the other by using patient blood containing the HbJ-Baltimore variant (Fig. 4).

These overlapping prints were pre-enhanced using a blood enhancement technique, namely Acid Black 1, to simulate operational conditions.

A preliminary observation to make is that overlapping blood marks at real crime scenes may be deposited in several ways. In this instance, the blood in the first print was still wet when the second wet blood fingerprint was overlapped. This occurrence would have generated a certain degree of ridge

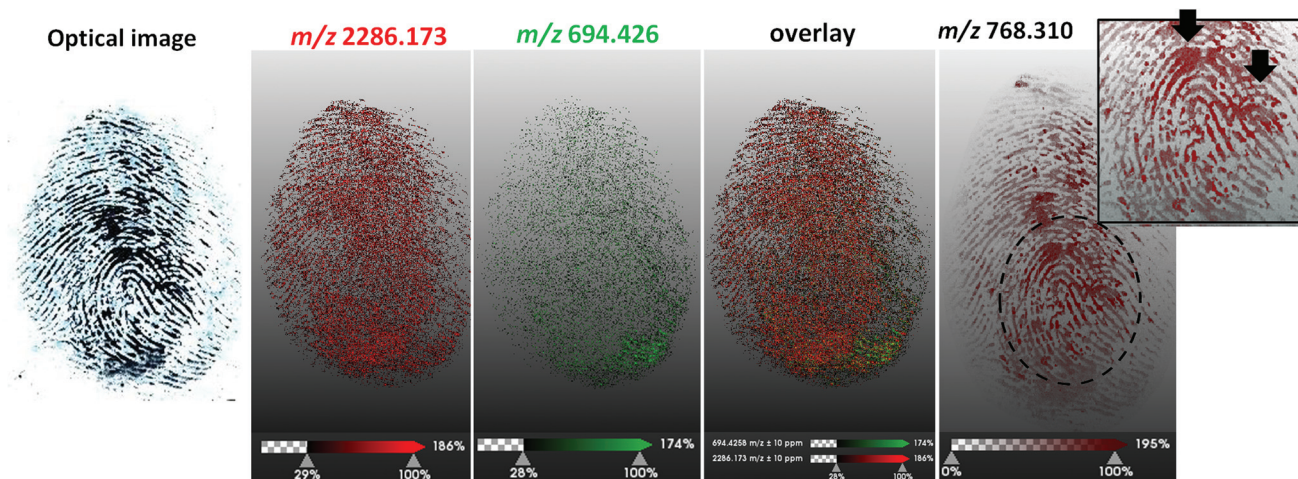


Fig. 4 MALDI MS images of overlapping fingerprints contaminated with blood containing HbC and HbJ-Baltimore variants respectively and pre-enhanced with acid black 1. HbJ-Baltimore blood fingerprint ( $m/z$  2286.173) has been separated from HbC blood fingerprint ( $m/z$  694.426) as the overlay of the two ion signals demonstrates; a fourth ion has been imaged at  $m/z$  768.310 showing clearly two ulnar loops (labelled with the black arrows in the zoom in inset), indicating the presence of two fingerprints.



smudging and analyte migration. Additionally, the marks were generated using 3D printed silicon fingertips to mitigate the biohazard, which have not allowed generation of clear ridge detail, due to the lack of sebaceous material that helps 'fix' the blood marks on surfaces and the inflexibility of the moulds compared to real fingertips. As a result, these overlapping fingerprints were not "textbook models" either. The MALDI Imaging data reflected the quality of the mark deposition and it was not possible to obtain good quality ridge detail. In terms of the separation of the fingerprints, although it was not possible to obtain two distinct ridge patterns, Fig. 4 very clearly shows that it is possible to indicate the presence of two prints and two donors, compatibly with the prior application of a typical crime lab blood enhancement technique, through to the detection and visualisation of the two proteotypic peptides HbJ-Baltimore at  $m/z$  2286.173 (Fig. 4b) and HbC (Fig. 4c) at  $m/z$  694.426. The image of an unidentified ion at  $m/z$  768.310 shows additional ridge flow but most importantly corroborates the presence of 2 marks due to the visualisation of two ulnar loops indicated by the black arrows in the zoomed inset.

This initial work, limited to the six most common Hb variants, demonstrates in principle the possibility to inform investigation on the presence of any of these variants in blood evidence. This intelligence could be important associative evidence to narrow down the pool of suspects. An interesting prospect of this research involves the hypothesis to attempt ethnic and/or geographic provenance. The former hypothesis is formulated on the knowledge that, for example, sickle cell anaemia is prevalent in the African American and Hispanic populations.<sup>12</sup> The second hypothesis has stemmed from the observation reported by one of the co-authors routinely screening Hb variants in the Sheffield laboratory serving many surrounding areas and beyond. The detection of HbD-Punjab is often an indication of the possible provenance of the donor of the geographical location; Sheffield/Rotherham area has a large South Asian population (predominantly of Pakistani heritage where the HbD Punjab variant is present in approx. 0.8% of the population) therefore the detection of this variant often rightly indicates a geographical location of the donor in the Sheffield/Rotherham (Jason Eyre, personal communication). However, a thorough study statistically investigating the correlation between Hb var prevalence and ethnicity/geographic provenance is needed to prove both hypotheses. Furthermore it is also important to factor in demographics when assessing the prevalence of variants as large populations from a certain ethnic background may go unscreened (for example University students from abroad), and a better indication of prevalence may be gained from looking at the incidence in the country of ethnic origin.

The use of a combined proteomic/MALDI approach is also interesting as a possible alternative or as a preliminary screening in clinical settings to reduce analytical time. In fact, although CE-HPLC or CE allow to easily discriminate for example HbD-Punjab/Los Angeles from HbS or Hb O-Arab from HbE and HbC,<sup>15</sup> these methods are still time consuming, whereas the MALDI screening is faster and can easily discriminate between HbD Punjab/Los Angeles and HbS and HbC

from HbE as shown in this study. However, further investigation and improvements are needed to demonstrate both robustness and quantitative capabilities; studies are needed to assess the possibility to distinguish between traits and pathology (for example between HbAS and HbSS) as well as the association of other variants with HbS leading to pathology (meaning the detection of both variants in the blood sample).

## 4. Conclusions

In this work, detection and mapping of haemoglobin variants (Hb var) in bloodstains and blood fingermarks have been conducted through a combined proteomics-MALDI based approach. Different MALDI instrumentation with different sensitivity specifications has been employed, highlighting the need for high end equipment to reliably detect all of the six Hb var investigated (HbS, HbC HbD-Punjab/Los Angeles, HbD-Iran, HbE, HbJ-Baltimore). The opportunity to detect and map Hb var could provide associative evidence to narrow down the pool of suspects during an investigation. An examination of the relevant literature has shown that there are many screening programmes across the world at birth and/or in workplaces including detection of Hb var. Whilst these programmes are aimed at prevention at reducing healthcare costs, the Hb var medical records could in principle be accessed during an investigation of a person of interest and compared with the Hb var findings at a crime scene. Additional research is required to investigate detectability of additional variants, compatibly with blood enhancement techniques, as well as robustness of the method against different environmental conditions and evidence age. Additional work is also required to fully understand the potential for Hb var to provide a meaningful indication of ethnicity as well as geographical provenance based on data on their prevalence. Finally, this work highlights the potential to provide an alternative or first screening method for Hb var that could be used in clinical settings. However, this hypothesis also needs further research.

## Author contributions

Cameron Heaton: investigation, methodology, formal analysis, reviewing and editing; Matthias Witt: investigation, methodology, formal analysis, reviewing and editing; Laura Cole: supervision, reviewing and editing; Simon Tazzyman and Jason Eyre: resources, reviewing and editing; Richard McColm: supervision, funding acquisition, reviewing; Simona Francese: funding acquisition, conceptualisation, project administration, supervision, writing original draft, formal analysis.

## Conflicts of interest

There are no conflicts to declare.





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## References

- 1 R. Bradshaw, S. Bleay, M. R. Clench and S. Francese, Direct detection of blood in fingerprints by MALDI MS profiling and Imaging, *Sci. Justice*, 2014, **54**, 110–117.
- 2 E. Patel, P. Cicatiello, L. Deininger, M. R. Clench, G. Marino, P. Giardina, G. Langenburg, A. West, P. Marshall, V. Sears and S. Francese, A proteomic approach for the rapid, multi-informative and reliable identification of blood, *Analyst*, 2015, **141**, 191–198.
- 3 L. Deininger, E. Patel, M. R. Clench, V. Sears, C. Sammon and S. Francese, Investigation of infinite focus microscopy for the determination of the association of blood with fingerprints, *Proteomics*, 2016, **16**, 1707–1717.
- 4 S. Kamanna, J. Henry, N. Voelcker, A. Linacre and K. P. Kirkbride, Bottom-up" in situ proteomic differentiation of human and non-human haemoglobins for forensic purposes by matrix-assisted laser desorption/ionization time of flight tandem mass spectrometry, *Rapid Commun. Mass Spectrom.*, 2017, **31**, 1927–1937.
- 5 S. Francese, Criminal profiling through MALDI MS based technologies – breaking barriers towards border free forensic science, *Aust. J. Forensic Sci.*, 2019, **51**, 623–635.
- 6 K. Kennedy, C. Heaton, G. Langenburg, L. Cole, T. Clark, M. R. Clench, V. Sears, M. Sealey, R. McColm and S. Francese, Pre-validation of a MALDI MS proteomics-based method for the reliable detection of blood and blood provenance, *Sci. Rep.*, 2020, **10**, 17087–17104.
- 7 M. Witt, K. Kennedy, C. Heaton, G. Langenburg and S. Francese, Forensic visualization of blood and blood provenance in old fingerprints by MALDI MS Imaging. Bruker Daltonik Application Note MSI-22, 2021, [https://www.bruker.com/en/products-and-solutions/mass-spectrometry/ms-software/scils-lab/\\_jcr\\_content/root/sections/more\\_information/sectionpar/search\\_copy\\_copy\\_cop.download-asset.pdf/c74307d7-facb-48a8-b702-76d89d408c2a/1885564-msi-22-imaging-fingerprints-ebook.pdf](https://www.bruker.com/en/products-and-solutions/mass-spectrometry/ms-software/scils-lab/_jcr_content/root/sections/more_information/sectionpar/search_copy_copy_cop.download-asset.pdf/c74307d7-facb-48a8-b702-76d89d408c2a/1885564-msi-22-imaging-fingerprints-ebook.pdf) (accessed March 22, 2021).
- 8 S. Kamanna, J. H. Nicholas, H. Voelcker, A. Linacre and K. P. Kirkbride, Direct identification of forensic body fluids using matrix-assisted laser desorption/ionization time-of-flight mass spectrometry, *Int. J. Mass Spectrom.*, 2016, **397–398**, 18–26.
- 9 S. Kamanna, J. Henry, N. H. Voelcker, A. Linacre and K. P. Kirkbride, A mass spectrometry-based forensic toolbox for imaging and detecting biological fluid evidence in fingerprints and fingernail scrapings, *Int. J. Legal Med.*, 2017, **131**, 1413–1422.
- 10 S. Meyer, N. Trost, B. M. Frey and C. Gassner, Parallel donor genotyping for 46 selected blood group and 4 human platelet antigens using high-throughput MALDI-TOF mass spectrometry, *Methods Mol. Biol.*, 2015, **1310**, 51–70.
- 11 B. K. Flesch, V. Scherer, B. Just, A. Opitz, O. Ochmann, A. Janson, M. Steitz and T. Zeiler, Molecular Blood Group Screening in Donors from Arabian Countries and Iran Using High-Throughput MALDI-TOF Mass Spectrometry and PCR-SSP, *Transfus. Med. Hemother.*, 2020, **47**, 396–408.
- 12 D. J. Weatherall and J. B. Clegg, Inherited haemoglobin disorders: an increasing global health problem, *Bull. World Health Organ.*, 2001, **79**, 704–712.
- 13 V. Viprakasit, R. Origa and S. Fucharoen, in *Guidelines for the Management of Transfusion Dependent Thalassaemia (TDT)*, ed. M. D. Cappellini, A. Cohen, J. Porter, A. Taher and V. Viprakasit, Thalassaemia International Federation Cyprus, Nicosia, 2014.
- 14 N. Swedan, K. Nicol, P. Moder and S. Kahawash, Haemoglobin J-Baltimore can be detected by HbA1c electropherogram but with underestimated HbA1c value, *Ther. Clin. Risk Manag.*, 2008, **4**, 649–652.
- 15 V. Brunel, A. Lahary, A. Chagraoui and C. Thuillez, Haemoglobin J-Baltimore can be detected by HbA1c electropherogram but with underestimated HbA1c value, *Biochem. Med.*, 2016, **26**, 240–242.
- 16 C. R. Wilburn, D. W. Bernard, A. W. Zieske, J. Andrieni, T. Miller and P. Wang, The Prevalence and Role of Hemoglobin Variants in Biometric Screening of a Multiethnic Population: One Large Health System's Experience, *Am. J. Clin. Pathol.*, 2017, **147**, 589–595.
- 17 H. Wajcman and K. Moradkhani, Abnormal haemoglobins: Detection & characterization, *Indian J. Med. Res.*, 2011, **4**, 538–546.
- 18 S. L. Thein, in *Blood and Bone Marrow Pathology*, ed. S. N. Wickramasinghe and J. McCullough, Elsevier, UK, Edinburgh, 2011, pp. 131–155.
- 19 A. Suarez, J. Polski and B. Grossman, Blood transfusion-acquired hemoglobin C: A case report and review of the literature, *Arch. Pathol. Lab. Med.*, 1999, **123**, 642–643.
- 20 M. T. Elghetany and F. B. Davy, in *Clinical Diagnosis and Management by laboratory method*, ed. J. B. Henry, B Saunders Co USA, New York, 2001, pp. 561–563.
- 21 C. Baglioni and D. J. Weatherall, Abnormal human hemoglobins IX. Chemistry of hemoglobin J Baltimore, *Biochim. Biophys. Acta*, 1963, **78**, 637–643.
- 22 M. Angastiniotis and B. Modell, Global epidemiology of hemoglobin disorders, *Ann. N. Y. Acad. Sci.*, 1998, **850**, 251–269.
- 23 H. A. Itano, A Third Abnormal Hemoglobin Associated with Hereditary Hemolytic Anemia, *Proc. Natl. Acad. Sci. U. S. A.*, 1951, **37**, 775–784.





- 24 C. Baglioni, Abnormal human haemoglobins VIII. chemical studies on haemoglobin D, *Biochim. Biophys. Acta*, 1962, **59**, 437–449.
- 25 L. de Souza Torres, J. V. Okumura, D. G. Silva and C. R. Bonini-Domingos, Hemoglobin D-Punjab: origin, distribution and laboratory diagnosis, *Rev. Bras. Hematol. Hemoter.*, 2015, **37**, 120–126.
- 26 G. Fioretti, M. De Angioletti, L. Pagano, G. Lacerra, A. Viola and C. de Bonis, DNA polymorphisms associated with Hb D-Los Angeles [ $\beta$ 121(GH4)Glu→Gln] in southern Italy, *Hemoglobin*, 1993, **17**, 9–17.
- 27 H. Husquinet, M. T. Parent, S. Schoos-Barbette, J. Dodinval-Versie, C. Lambotte and F. Galacteros, Hemoglobin D-Los Angeles [ $\beta$ 121(Gh4)Glu → Gln] in the Province of Liège, Belgium, *Hemoglobin*, 1986, **10**, 587–592.
- 28 A. Lischka, A. Pollak, K. Bauer, H. Aschauer and G. Braunitzer, Hemoglobin D “Los Angeles” in an Austrian Family: Biochemical Identification, Clinical Aspects, and Kindred Study, *Hemoglobin*, 1984, **8**, 353–361.
- 29 E.Ö Atalay, H. Koyuncu, B. Turgut, A. Atalay, S. Yildiz and A. Bahadir, Rare Hemoglobin Variant Hb Yaizu Observed in Turkey, *Hemoglobin*, 2005, **29**, 307–310.
- 30 J. Dass, A. Gupta, S. Mittal, A. Saraf, S. Langer and M. Bhargava, Comparison of the characteristics of two hemoglobin variants, Hb D-Iran and Hb E, eluting in the Hb A2 window, *Blood Res.*, 2017, **52**, 130–134.
- 31 S. Rahbar, Haemoglobin D Iran:  $\beta$ 222 Glutamic Acid→Glutamine (B4), *Br. J. Haematol.*, 1973, **24**, 31–35.
- 32 Hemoglobinopathies: Current Practices for Screening, Confirmation and Follow-up. [https://www.cdc.gov/ncbddd/sicklecell/documents/nbs\\_hemoglobinopathy-testing\\_122015.pdf](https://www.cdc.gov/ncbddd/sicklecell/documents/nbs_hemoglobinopathy-testing_122015.pdf). 2015 (accessed March 11 2021).
- 33 NHS Sickle Cell and Thalassaemia Screening Programme. [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/file/585126/NHS\\_SCT\\_Handbook\\_for\\_Newborn\\_Laboratories.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/585126/NHS_SCT_Handbook_for_Newborn_Laboratories.pdf) (accessed March 11 2021).
- 34 M. Strohm, M. Hassman, B. Košata and M. Koldíček, mMass Data Miner: An Open Source Alternative for Mass Spectrometric Data Analysis, *Rapid Commun. Mass Spectrom.*, 2008, **22**, 905–908.
- 35 M. Strohm, D. Kavan, P. Novakand; and V. Havlicek, mMass 3: A Cross-Platform Software Environment for Precise Analysis of Mass Spectrometric Data, *Anal. Chem.*, 2010, **82**, 4648–4651.
- 36 H. L. Bandey, S. M. Bleay, V. J. Bowman, R. P. Downham and V. G. Sears, *Fingermark Visualisation Manual*, ed H. Bandey, Home Office, St Albans, 2014.
- 37 J. Yergey, A general approach to calculating isotopic distributions for mass spectrometry, *Int. J. Mass Spectrom. Ion Phys.*, 1983, **52**, 337–349.

