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## Introduction

$\alpha$ -Amanitin, the deadliest of the amatoxins produced by the death-cap mushroom *Amanita phalloides*, is a potent, orally available inhibitor of RNA polymerase II (pol II) ( $K_i \sim 10$  nM),<sup>1</sup> that has been validated as a payload for targeted cancer therapy.<sup>2</sup> First described in 1907<sup>3</sup> and isolated in 1941,<sup>4</sup>  $\alpha$ -amanitin is a compact bicyclic octapeptide that has been indispensable for probing RNA pol II-catalysed transcription in eukaryotes. While the toxin makes numerous contacts with RNA pol II, the lack of synthetic access has limited the ability to generate discrete analogs to probe these contacts, most notably the role of the 6'-hydroxyl group on the indole. Of the few naturally occurring amatoxins wherein the 6'-OH is replaced by H, amaninamide and amanin are only 25–50% as toxic as the natural product (Fig. 1) indicating moderate importance of the OH group.<sup>5</sup>

Its putative contribution to affinity is understood from two apparent interactions seen in a recent co-crystal structure of  $\alpha$ -amanitin-RNA pol II: (i) the 6'-hydroxyl group forms an H-bond with Ser-782 accounting for the difference in  $K_i$ 's between mammalian and yeast RNA pol II; (ii) the hydroxyl group increases the electron density of the indole which forms a cation– $\pi$  interaction with Arg-749.<sup>6</sup> Besides contributing to toxicity, the 6'-OH provides a preferred handle for chemical functionalization for grafting to antibodies.

Recently,  $\alpha$ -amanitin has been showcased as a highly effective payload in antibody drug conjugates (ADCs).<sup>2a</sup> Unlike other ADC payloads,  $\alpha$ -amanitin targets both dividing and quiescent cells and shows potential for preventing cancer relapse from

## Meeting key synthetic challenges in amanitin synthesis with a new cytotoxic analog: 5'-hydroxy-6'-deoxy-amanitin†

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Appreciating the need to access synthetic analogs of amanitin, here we report the synthesis of 5'-hydroxy-6'-deoxy-amanitin, a novel, rationally-designed bioactive analog and constitutional isomer of  $\alpha$ -amanitin, that is anticipated to be used as a payload for antibody drug conjugates. In completing this synthesis, we meet the challenge of diastereoselective sulfoxidation by presenting two high-yielding and diastereoselective sulfoxidation approaches to afford the more toxic (*R*)-sulfoxide.

drug-tolerant cell subpopulations.<sup>7</sup> Examples include ADCs for targeting human epidermal growth factor receptor 2 and prostate specific membrane antigen.<sup>8</sup> With but a few exceptions,<sup>9</sup> nearly all bioconjugates to a cytotoxic amanitin have emerged from naturally-sourced  $\alpha$ -amanitin. To date, conjugation handles used for  $\alpha$ -amanitin-based bioconjugates include the  $\delta$ -hydroxyl of (2*S*,3*R*,4*R*)-4,5-dihydroxyisoleucine (DHile),<sup>2a</sup> the asparagine side chain,<sup>10</sup> and the 6'-hydroxyl of the tryptathionine staple.<sup>11</sup> Of these, increasingly, the latter has been favored as a site for bioconjugation to the intact toxin.

As biosynthetic fermentation is currently low-yielding,<sup>12</sup> synthesis will likely serve as the main means for accessing large amounts of toxin needed to address structure–activity relationships and provide new handles for bioconjugation. This heightened interest in  $\alpha$ -amanitin has inspired three total syntheses<sup>13</sup> that address synthetic challenges embedded in three key components: (i) an enantioselective synthesis of DHile, (ii) creation of an oxidatively challenged 6'-OH-tryptathionine-sulfoxide that demands formal oxidation at

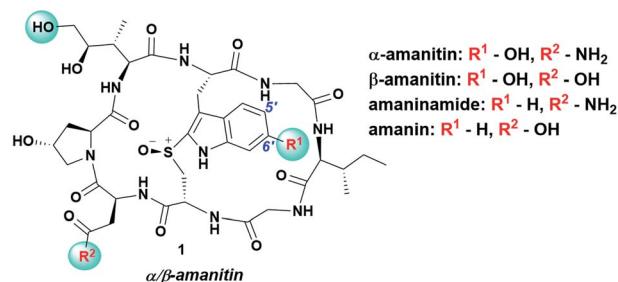


Fig. 1 Chemical structure of  $\alpha/\beta$ -amanitin and structure–activity relationships of natural and synthetic amatoxins with indole modifications; a 2–4 fold-increase in  $K_i$  as a result of indole modification is relative to the 6'-OH containing counterparts; commonly used conjugation sites are highlighted.

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positions 2 and 6 of the indole, and (iii) stereoselective oxidation of the thioether to give (*R*)-sulfoxide, which is approximately 8-fold more toxic than the (*S*)-sulfoxide.<sup>13a</sup>

In 2018, Perrin *et al.* opted for a 6-boronated-L-tryptophan as a masked phenol equivalent that underwent oxidative fluorocyclization followed by Savige–Fontana reaction to form the tryptathionine staple<sup>14</sup> with the pendent boronate.<sup>13a</sup> Oxidative deborylation, peptide elongation, and macrocyclization gave the *S*-deoxy-amanitin that was readily sulfoxidized with moderate diastereoselectivity (*ca.* 2 : 1) favoring the (*R*)-sulfoxide.

In 2020, Süssmuth *et al.* described a convergent “5 + 1 + 2” strategy whereby the tryptathionine staple was synthesized first by reacting protected cysteine-sulfenyl chloride with orthogonally protected 6-OBn-L-tryptophan. The tryptathionine was then elaborated to a pentapeptide followed by addition of DHile and Pro-Asn and macrolactamization.<sup>13b</sup> A third report advanced a scalable synthesis of 6-OAc-Trp, and utilized Savige–Fontana reaction<sup>14</sup> to introduce the 6'-OH-tryptathionine staple.<sup>13c</sup> While all three reports provide access to 6'-OH-tryptathionine, they noted the severe challenges of selective oxidative thiolation of the electron-rich indole.

In contrast to the immediate interest in accessing the 6'-hydroxyl group, interest in generating the (*R*)-sulfoxide remains academic. Nevertheless, because Nature exclusively produces the (*R*)-sulfoxide, logic suggests that the (*R*)-sulfoxide may provide privileged pharmacological/toxicological properties. As such, diastereoselective access of the (*R*)-sulfoxide continues to pose a significant synthetic challenge that was met with but moderate selectivity in the first total synthesis and remained unprogressed in the two subsequent ones.

Despite these synthetic efforts, there remains a demand for biologically active *and* more synthetically accessible  $\alpha$ -amanitin analogs that: (i) capture the putative  $\pi$ -cation interaction for enhanced affinity and (ii) provide an indole-situated conjugation handle for ADC development. Towards these ends, herein we report the synthesis of a constitutional isomer of near-native toxicity that introduces a 5'-OH-tryptathionine, which is readily achieved from cheap, commercially available 5-OH-L-tryptophan (5-OH-Trp).

In addition, we identify new methodologies to create the (*R*)-sulfoxide with high diastereoselectivity ( $>15 : 1$ ) along with the (*S*)-sulfoxide diastereomer and sulfone for comparison. Anticipating success with chemoselective conjugation akin to aforementioned conjugates to the 6'-OH-indole, this work now expands the toolbox of available synthetic amanitin analogs for consideration as payloads for use in translational applications.

## Results

In approaching the synthetic challenge of creating a hydroxy-tryptathionine staple, we appreciated the high natural abundance of 5-OH-tryptophan in a number of biosynthetic pathways along with its commercial availability as a single enantiomer. Hence, we hypothesized that 5-OH-L-tryptophan could be introduced *in lieu* of the synthetically-demanding 6-OH-L-tryptophan. We also reasoned that the 5'-hydroxyl could

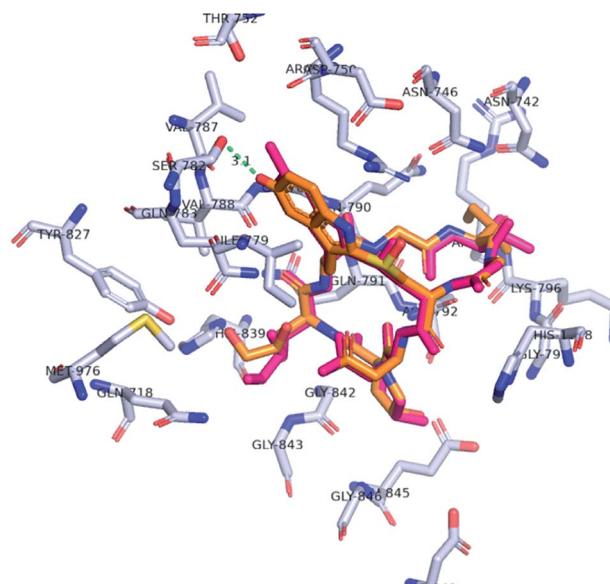


Fig. 2 Docking pose of 5'-OH-6'-deoxy-amanitin 2 showing favorable interactions with RNA-pol II; 5'-OH-6'-deoxy-amanitin 2 – orange,  $\alpha$ -amanitin 1 – magenta, green dashed line – distance between Ser-782 and indole alcohol.

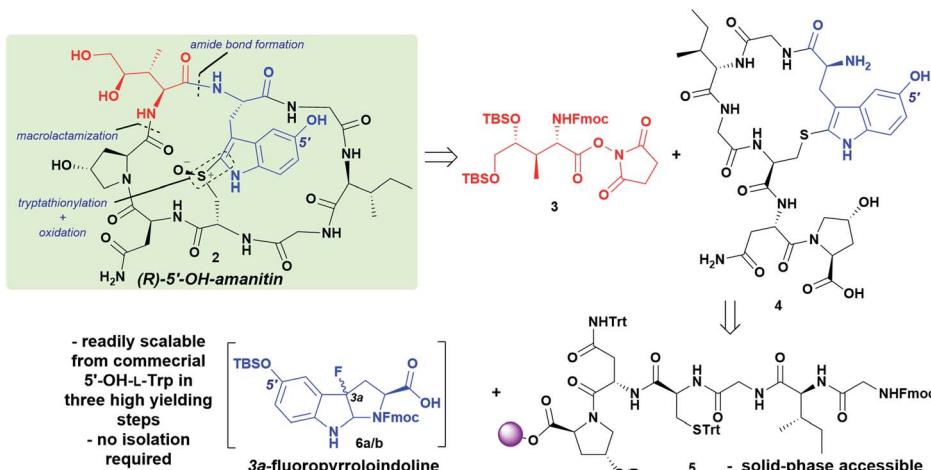
also engage in an H-bonding interaction with Ser-782. A docking study validated this hypothesis: the 5'-OH-6'-deoxy-amanitin (2) was docked into the RNA pol II using Autodock Vina molecular docking program.<sup>15</sup>

The most energetically favorable pose of 5'-OH-6'-deoxy-amanitin was compared to the crystal structure pose of  $\alpha$ -amanitin (Fig. 2). Further assessment using PyMOL Molecular Graphics System confirmed that the aforementioned key interactions between the enzyme and the indole ring are still achievable and the modified indole does not alter the overall confirmation of the fit.<sup>16</sup> Importantly, the docking pose suggested that the 5'-hydroxyl group should still form the anticipated H-bond with Ser-782 (Fig. 2, green line). Based on these observations, we reasoned that the incorporation of 5-OH-tryptophan should maintain the toxicity of the new amanitin analog.

Encouraged by this result, we undertook a synthesis of 5'-OH-6'-deoxy-amanitin (2) that would rely on two key components: (i) introduction of the fully protected DHile-NHS ester as previously reported, and (ii) synthesis and characterization of the 3a-fluoro-pyrroloindoline produced from 5-OH-tryptophan that could be subjected to late-stage Savige–Fontana reaction to give the 5'-OH-tryptathionine (Scheme 1). In addition to this synthetic feat, we addressed mid- and late-stage sulfoxidation with high diastereoselectivity to afford the desired (*R*)-sulfoxide.

Following the approach used in the first total synthesis of  $\alpha$ -amanitin<sup>13a</sup> and supported by more recent work on the oxidative fluorocyclization of tryptophan to give fluoropyrroloindoline (FPI) for Savige–Fontana tryptathionylation,<sup>17</sup> we started with commercially available 5-OH-tryptophan. Following Fmoc-protection, oxidative fluorocyclization with *N*-fluoro-2,4,6-trimethylpyridinium triflate (FP-T300) gave a diastereomeric





**Scheme 1** Retrosynthetic of 5'-OH-6'-deoxy-amanitin 2, showing major disconnection sites and key intermediates. DHLe-NHS ester 3 is coupled to the monocyclic 5'-OH-heptapeptide 4 obtained from a 5-OH-tryptophan-derived FPI precursor 6a/b and solid-phase accessible hexapeptide 5.

pair of fluoropyrroloindolines (*syn-cis/anti-cis*). Unfortunately, these decomposed upon isolation, presumably due to the electron-rich indole ring (data not shown). To mitigate this problem, we employed a facile protection of the hydroxyl group as a *tert*-butyldimethylsilyl ether that greatly improved the stability of the diastereomeric pair of FPI's 6a/b. Thus, TBS-protected 8 was treated with FP-T300 in DCM to yield 6a/b as a diastereomeric mixture in 87% yield (Scheme 2, 6a/b *syn-cis/anti-cis*, 1.8 : 1).

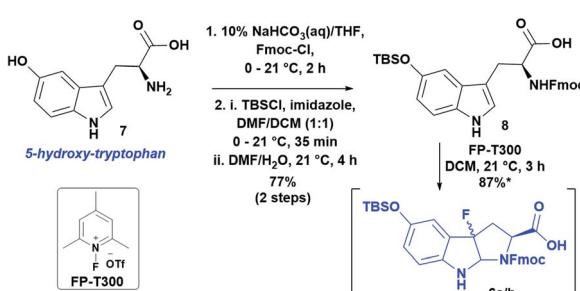
With fluorocyclization proceeding in high yield, we directly coupled the diastereomeric mixture 6a/b to hexapeptide 5 that had been prepared on the solid-phase using standard Fmoc solid-phase peptide synthesis conditions (Scheme 3). It is easily appreciated that the 1.8 : 1 diastereomeric distribution of the FPI precursor is immaterial since the stereochemistry is lost during the Savage-Fontana reaction. Fmoc-deprotection, followed by TFA/DCM (5 : 1) treatment resulted in global deprotection, resin cleavage, and thioetherification (Savage-Fontana reaction) to afford the desired 5'-OH-tryptathionine-containing monocyte 4. The monocyclic heptapeptide 4 was coupled with the fully protected NHS-ester of DHLe (3).<sup>13a</sup> Subsequent Fmoc-deprotection revealed a free amine, while TBS

deprotection decreased steric encumbrance to facilitate macrocyclization that successfully effected 11, the penultimate and first appreciably toxic species that would await sulfoxidation to give the target (R)-5'-OH-6'-deoxy-amanitin (Scheme 3).

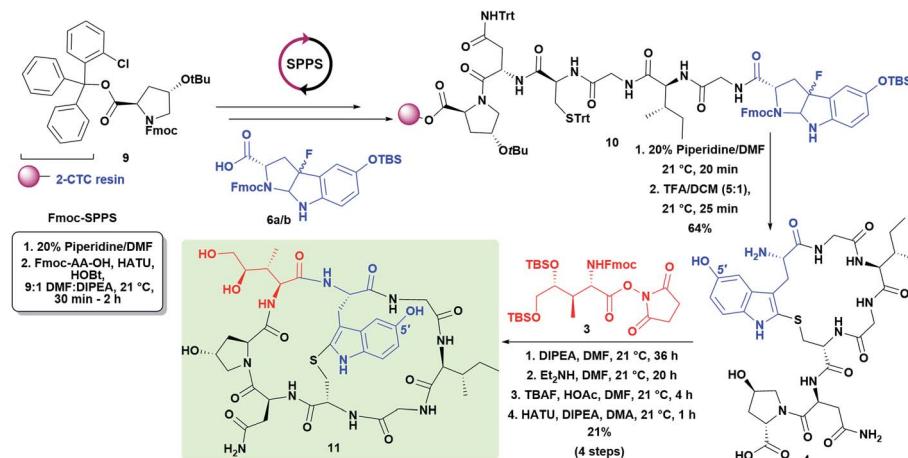
With a robust synthetic approach to a new thioether analog with equal toxicity compared to *S*-deoxy-amanitin (*vide infra*), the final obstacle towards 5'-OH-6'-deoxy-amanitin was the diastereoselective sulfoxidation of the tryptathionine bridge to the (R)-configured sulfoxide. Of note, in the *α*-amanitin series, the (R)-sulfoxide is the most potent of the toxins, followed by the equipotent sulfone and thioether and then the unnatural (*S*)-sulfoxide that shows 8-fold reduced potency.<sup>18</sup> The relationship between the stereochemical configuration of the sulfoxide and the biological activity has fascinated scientists for decades as numerous efforts fail to provide a structural rationalization for this intriguing difference in toxicity (*vide infra*, discussion).<sup>5b,18,19</sup>

Achieving diastereoselective sulfoxidation still poses an unmet synthetic challenge in this context. Previously, it was shown that oxidation of the thioether with H<sub>2</sub>O<sub>2</sub> in acetic acid produces sulfoxides in a 1 : 2 ratio favoring the (*S*)-diastereomer.<sup>5b,20</sup> In the 2018 report, *m*-chloroperbenzoic acid (*m*CPBA)-mediated sulfoxidation successfully yielded the (R)-sulfoxide in modest ~2 : 1 excess over (*S*)-sulfoxide. Inexplicably, selectivity could be tuned by varying the reaction solvent; iPrOH/EtOH and *t*BuOH/EtOH co-mixtures were effective at providing moderate selectivity for the desired (R)-sulfoxide while polar solvents (e.g. DMF, HOAc, CF<sub>3</sub>CH<sub>2</sub>OH) greatly favored the (*S*)-sulfoxide.<sup>13a,c</sup>

In order to verify that 11 would undergo similar sulfoxidation as *S*-deoxy-amanitin, we commenced our investigation by subjecting it to *m*CPBA treatment in iPrOH/EtOH (2 : 1) (Fig. 3). The HPLC and mass spectrometry analysis of the reaction revealed full consumption of 11 after one hour and formation of three distinct products in a ratio of 4.4 : 4.6 : 1, which were identified by mass spectrometry as two sulfoxides



**Scheme 2** Synthesis of *N'*-Fmoc-FPI(5-OTBS)-OH (6a/b) as a pair of diastereomers; \*NMR yield.



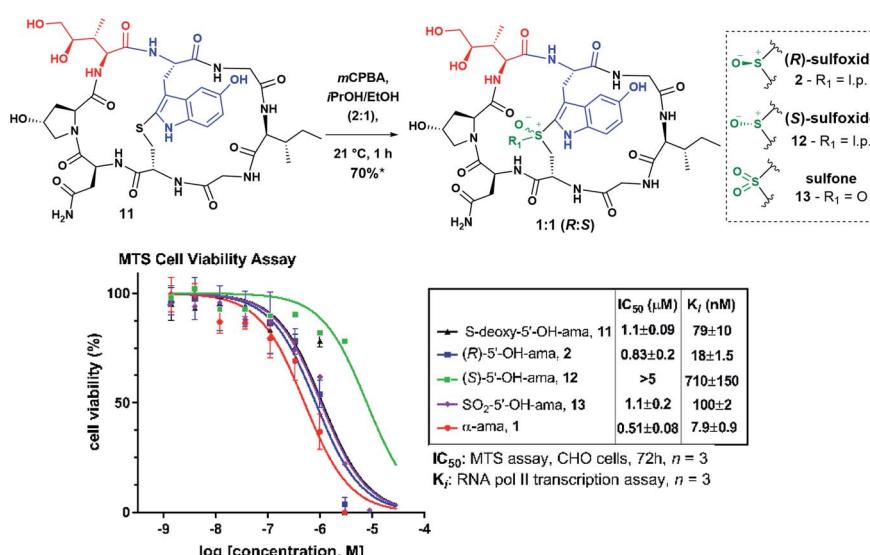
Scheme 3 Synthesis of 5'-OH-tryptathionine and incorporation of DHlle to give bicyclic precursor 11.

and a sulfone (see Fig. S2 in ESI†). This transformation was accompanied by a predictable change in the UV-Vis absorbance signifying a modification of the chromophore. The first of the two sulfoxides eluting from the HPLC was tentatively assigned as the desired (*R*)-sulfoxide based on the shorter HPLC retention time observed for  $\alpha$ -amanitin with respect to its (*S*)-sulfoxide diastereomer. It is well-known that the (*S*)-sulfoxides in a series of amanitin analogs are significantly less potent inhibitors of RNA pol II *in vitro*, compared to the corresponding (*R*)-sulfoxides<sup>5c</sup> (although previous reports did not provide unambiguous structural elucidation and relied only on correlations with chromatographic retention times and toxicity to assign stereochemistry).

Applying the correlation between chromatographic retention times of these compounds and authentic  $\alpha$ -amanitin to make a tentative stereochemical assignment, we then assessed the biological activity of the new amanitin analogs 2 and 11–13.

using MTS colorimetric cell viability and RNA pol II transcription assays (Fig. 3). To our delight, MTS assay on Chinese Hamster Ovary (CHO) cells revealed that 2 and 11–13 showed toxicities in keeping with the chromatographic elution profile observed for the (*R*)-, (*S*)-sulfoxides and the sulfone of  $\alpha$ -amanitin (Fig. 3) and leading us to assign a tentative (*R*)-designation to the sulfoxide born on 2.

Whereas 2 was nearly as cytotoxic as  $\alpha$ -amanitin itself, strikingly, the putative (*S*)-sulfoxide 12 showed significantly reduced cytotoxicity even compared to (*S*)-sulfoxide of  $\alpha$ -amanitin.<sup>13a,18,21</sup> While many synthetic amanitin analogs reported in the recent studies have relied solely on IC<sub>50</sub> values from cytotoxicity assays as a measure of activity,<sup>9,22</sup> such analysis assumes equal cellular uptake. To obtain a direct measure of activity against the intracellular target RNA pol II, we performed an *in vitro* transcription assay using HeLa-Scribe nuclear extract transcription system and [<sup>32</sup>P]- $\alpha$ -rGTP. Interestingly, the K<sub>i</sub>

Fig. 3 MTS and RNA pol II assay results of oxidized products; \*yield in sulfoxide; IC<sub>50</sub>: CHO cells, MTS, 72 h, n = 3; K<sub>i</sub>: RNA pol II transcription assay, n = 3.

values for compounds **2** and **11–13** were spread over a far greater range compared to the  $IC_{50}$  values (Fig. 3). Thioether **11** and sulfone **13** showed similar affinity ( $79 \pm 10$  and  $100 \pm 2.2$  nM, respectively), while (*R*)-sulfoxide **2** was approximately five-fold more active ( $18 \pm 1.5$  nM) and the (*S*)-sulfoxide **12** showed greatly reduced inhibitory activity ( $710 \pm 150$  nM) compared to (*R*)-sulfoxide **2**.

The near-native toxicity of **2** would suggest significant potential for use as an ADC payload and prompted us to develop diastereoselective sulfoxidation methods in anticipation of future translational needs. Hence, we turned our attention to catalytic sulfoxidation as it represents one of the dominant methods for enantioselective sulfoxidation. One of the earliest and most convenient methods for asymmetric sulfoxidation employs titanium tetra-isopropoxide [ $Ti(OiPr)_4$ ] and (+)-diethyl-L-tartrate (L-DET) as a ligand.<sup>23</sup> The initial sulfoxidation screening in the presence of  $Ti(OiPr)_4$  and L-DET was carried out with two different oxidizing agents, cumene hydroperoxide (CHP) and urea hydroperoxide (UHP), in MeOH : DCM (1 : 1) at 4 °C (Table 1, entries 1–3). Use of CHP as an oxidant yielded (*S*)-sulfoxide in excess (Table 1, entry 1) while UHP gave the (*R*)-sulfoxide in 2.6-fold excess when used in the presence of L-DET :  $Ti(OiPr)_4$  (Table 1, entry 2). In the absence of L-DET :  $Ti(OiPr)_4$ , UHP gave the (*S*)-sulfoxide in 2.8-fold excess (Table 1, entry 3). When L-DET :  $Ti(OiPr)_4$  were used in 4 : 1 ratio, the diastereoselectivity was improved to 95 : 5 (*R* : *S*) when applied to 100-nmol (Table 1, entry 4) (lower selectivity was observed on a 5-nmol scale). Use of excess L-DET ligand with respect to  $Ti(OiPr)_4$  (4 : 1 ratio) in 1 : 1 MeOH/DCM at 4 °C over 20 hours (Table 1, entry 4), yielded the desired (*R*)-sulfoxide in 71% yield and without over-oxidation to the sulfone. Nevertheless, approximately 25% of thioether **11** remained unconsumed after 20 hours. Increasing the amount of UHP in the presence of L-DET :  $Ti(OiPr)_4$  (8 : 1) and using DCM as the major solvent improved both the yield and the diastereoselectivity (5–

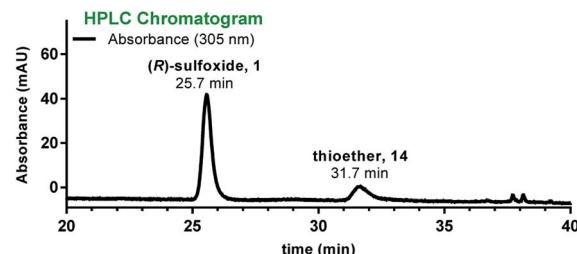
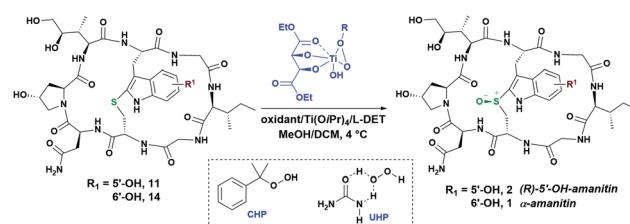


Fig. 4 HPLC chromatogram of the sulfoxidation reaction run in the presence of  $Ti(OiPr)_4$  and L-DET showing formation of  $\alpha$ -amanitin **1** as the major diastereomer along with unreacted thioether **14**.

nmol scale, Table 1, entry 5). To illustrate the utility of this protocol in production of  $\alpha$ -amanitin, we subjected *S*-deoxy-amanitin **14** to these optimized conditions. Gratifyingly, the (*R*)-sulfoxide was obtained in 84% yield and with high diastereoselectivity (Table 1, entry 6 and Fig. 4). The product identity was confirmed by HPLC co-injection with an authentic sample of  $\alpha$ -amanitin, illustrating the first case of a highly diastereoselective access to  $\alpha$ -amanitin (see Fig. S7, ESI†).

While these results represent the first-ever successful approach to a highly diastereoselective installation of the (*R*)-sulfoxide in any amanitin analog, we wondered if there could be yet other means of accessing the desired (*R*)-sulfoxide without resorting to chiral catalysis. Appreciating that the monocyclic heptapeptide intermediate **4** likely adopts a different geometry compared to the bicyclic **11**, we hypothesized that the sulfoxidation of this intermediate may offer a unique opportunity for inducing the desired (*R*)-configuration at an earlier stage in the synthesis. We started our initial screening of the sulfoxidation with *m*CPBA in *iPrOH/EtOH* (2 : 1); two products eluted from the HPLC one minute apart. The first product, designated as compound **15**, appearing to be a single diastereomer, was found to be a sulfoxide by MS but of unknown stereochemistry, while

Table 1 Oxidation of thioether **11** and **14** to (*R*)- and (*S*)-sulfoxides in the presence of  $Ti(OiPr)_4$  and L-DET<sup>a</sup>



Entry	Conditions	Substrate	( <i>R</i> : <i>S</i> )	<b>11</b> or <b>12</b> (%)	Yield (%)
1	CHP/ $Ti(OiPr)_4$ /L-DET (1 : 1 : 2), DCM/MeOH (1 : 1), 17 h	<b>11</b>	1 : 1.1	39	61 <sup>b</sup>
2	UHP/ $Ti(OiPr)_4$ /L-DET (1 : 1 : 2), DCM/MeOH (1 : 1), 17 h	<b>11</b>	2.6 : 1	44	66 <sup>b</sup>
3	UHP, DCM/MeOH (1 : 1), 17 h	<b>11</b>	1 : 2.8	20	80 <sup>b</sup>
4	UHP/ $Ti(OiPr)_4$ /L-DET (1.2 : 0.8 : 3.2), DCM/MeOH (1 : 1), 20 h	<b>11</b>	<b>19 : 1</b>	25	<b>71<sup>c</sup></b>
5	UHP/ $Ti(OiPr)_4$ /L-DET (2.5 : 1 : 8), DCM/MeOH (3 : 1), 17 h	<b>11</b>	6.4 : 1	4	96 <sup>b</sup>
6	UHP/ $Ti(OiPr)_4$ /L-DET (2.5 : 1 : 8), DCM/MeOH (3 : 1), 17 h	<b>14</b>	>50 : 1	<b>16</b>	<b>84<sup>b</sup></b>

<sup>a</sup> (*R* : *S*) distribution was determined by HPLC. <sup>b</sup> HPLC sulfoxide yield. <sup>c</sup> Isolated yield.

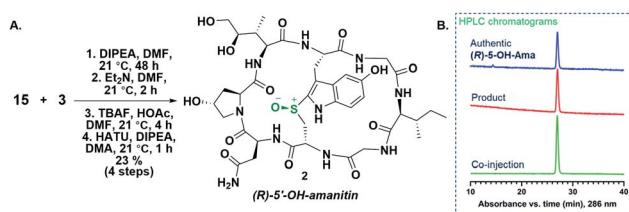


the second proved to be an inseparable mixture of the second sulfoxide and sulfone **16** (Fig. 5). To reduce the amount of sulfoxide/sulfone by-products, solvent and temperature screening was performed (Table S4,† entries 1–16). The lowest amount of the by-products was obtained with dimethylformamide (DMF) as the solvent (Table S4,† entry 11). The sulfoxidation was further optimized by varying the reaction temperature (Table S4,† entries 14–16). Temperatures below 0 °C were found to be optimal for suppressing the production of sulfoxide/sulfone. Running the reaction in DMF at –20 °C on a 1-μmol scale yielded the first sulfoxide in 5-fold excess over the second sulfoxide and inseparable sulfone (Fig. 5). This protocol resulted in complete consumption of the thioether **4** in less than 5 minutes.

To confirm the stereochemical configuration of the monocyclic sulfoxide, **15** was converted to the final bicyclic 5'-OH-6'-deoxy-amanitin **2** by the following approach (Scheme 4): the heptapeptide sulfoxide **15** was coupled to a suitably protected dihydroxyisoleucine to give the octapeptide, which was then Fmoc- and TBS-deprotected and finally macrolactamized to produce the bicyclic amanitin analog **2** (Scheme 4). The final product was analysed by HPLC, including co-injection with an “authentic sample” of (R)-5'-OH-6'-deoxy-amanitin (**2**) (Fig. 2), that was obtained through an *m*CPBA oxidation of the bicyclic precursor **11** following a previously reported procedure for  $\alpha$ -amanitin synthesis (Scheme 4A).<sup>13a</sup>

Based on the HPLC co-injection data, we therefore confirmed that the sulfoxide **15** was indeed of the (R)-configuration (Scheme 4B). This method thus represents a highly diastereoselective, mild, facile, and cost-effective approach for introduction of (R)-sulfoxide at an earlier stage in the synthesis and, importantly, before the incorporation of the costly DHle or other analogs that may be of interest.

To examine whether this approach is generally applicable to other monocyclic precursors, we attempted this *m*CPBA sulfoxidation on the monocyclic 5'-OH-pentapeptide **S4**, analogous



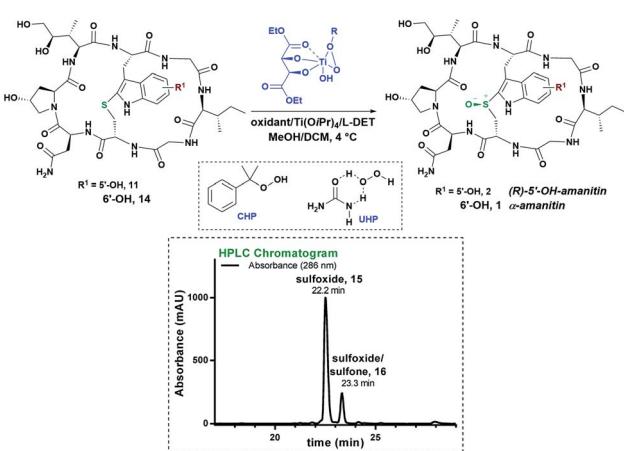
**Scheme 4** Synthesis of the (R)-5'-OH-6'-deoxy-amanitin **2** to confirm the stereochemistry of the heptapeptide sulfoxidation, incorporation of dihydroxyisoleucine and macrolactamization (A). HPLC chromatogram of the sulfoxide containing 5'-OH-6'-deoxy-amanitin obtained through sulfoxide **15** and co-injection with (R)-5'-OH-6'-deoxy-amanitin **2** produced via a reported *m*CPBA oxidation route confirming (R)-sulfoxide formation during heptapeptide sulfoxidation (B).

to the building block employed in the “5 + 1 + 2” strategy developed by Süssmuth and coworkers. Somewhat surprisingly, this led to formation of two sulfoxides in 1 : 2 ratio. Similarly, when the monocyclic octapeptide (*seco*-5'-OH-6'-deoxy-octapeptide **S3**) was subjected to *m*CPBA in DMF, two sulfoxides were formed with little diastereoselectivity (1 : 1.7) suggesting that neither the pentapeptide nor the octapeptide adopts a conformation that is preordered to confer highly diastereoselective attack by the oxidant.

In an attempt to rationalize the outstanding selectivity in oxidizing the heptapeptide to obtain the (R)-sulfoxide, we examined the differences in geometries between the bicyclic octapeptide **11** and monocyclic heptapeptide **4**. Based on the geometry-optimized structure of the heptapeptide monocycle **4**, we hypothesize that the extensive H-bonding interactions observed between Asn-1, Hyp-2, 5'-OH-Trp-4 (Fig. 6A) and the backbone may cause an increase in steric bulk on the *si*-face of the indole, disfavoring oxidant attack on the pro-*S* lone pair on the sulfur atom while leaving the pro-*R* lone pair more solvent-accessible.

This can be observed by measuring distances between sulfur and the neighboring atoms in the heptapeptide monocycle **4** and comparing these to bicyclic **11** (Fig. 6B). Based on this analysis, five out of six neighboring atoms are in closer proximity to sulfur in the heptapeptide. The Asn-1 side chain is noteworthy since it shows the largest degree of displacement. In contrast, the conformation of amanitin **11** shows that both lone pairs on the sulfur atom share a similarly congested environment thus explaining the generally poor diastereoselectivity.

Finally, to complete this study, we evaluated certain key products by circular dichroism (CD) to prove for key structural elements such as  $\alpha$ -helices,  $\beta$ -sheets, and  $\beta$ -turns that are consistently found in non-random portions of proteins. Amanitins are characterized by a type-II  $\beta$ -turn in ring II (formed by Trp4-Gly5-Ile6-Gly7-Cys8), which is recognized to be crucial for binding, and numerous natural toxins in this class as well as synthetic derivatives have been characterized by CD. Notably, (R)-sulfoxide **2** strongly matched the conformation of  $\alpha$ -amanitin; however, the nearly-equivalent thioether **11** gave a spectrum that differed significantly from both **2** and  $\alpha$ -



**Fig. 5** Oxidation of heptapeptide **4** to (R)-sulfoxide and (S)-sulfoxide in 5 : 1 ratio with *m*CPBA as an oxidant in DMF at –20 °C; HPLC chromatogram of *m*CPBA oxidation; in reporting this, we show **15** to have undefined stereochemistry at sulphur, which was determined subsequently.



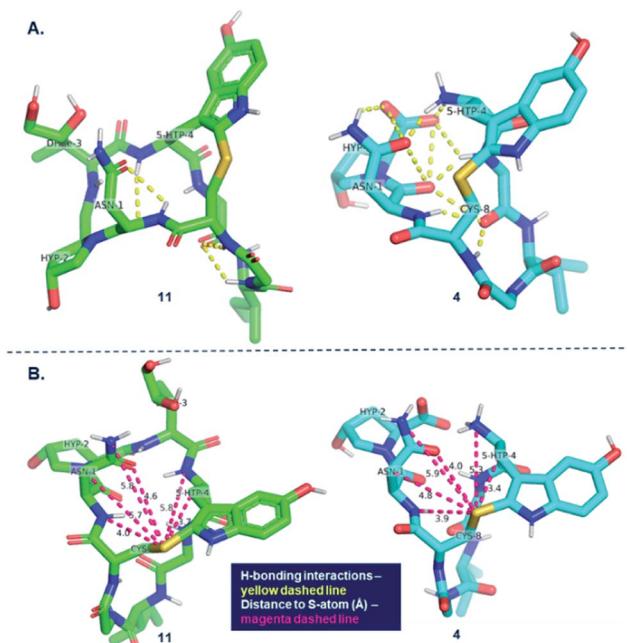


Fig. 6 Modelling of bicyclic precursor **11** (in green, left) and mono-cyclic heptapeptide precursor **4** (in blue, right) shows greater exposure of the pro-*R* lone pair compared to the pro-*S* lone pair; H-bonding interactions highlighted in yellow (A); distances from neighboring atoms to S-atom (B).

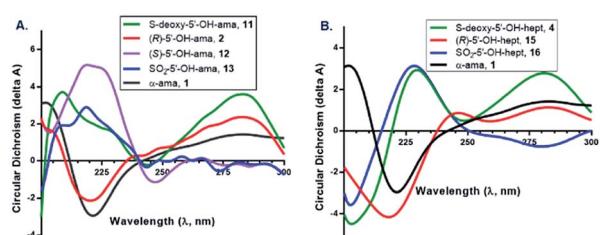


Fig. 7 CD spectra of amanitins **2**, **11–13** (A) and heptapeptides **4**, **15**, **16** (B).

amanitin in the 210–240 nm range (Fig. 7). The (*S*)-sulfoxide **12** and sulfone **13** both differed significantly from **2** and  $\alpha$ -amanitin at nearly all wavelengths, and while both showed more similar CD-spectra, these showed vastly different cytotoxicities and  $K_i$  values *in vitro*. A similar trend is observed for the heptapeptides. The implications of these finding are discussed below.

## Discussion

Herein we describe the synthesis of a new amanitin analog that is a constitutional isomer of the natural product. In light of the importance of the 6'-OH for both enhancing toxicity and providing a conjugation handle, and given the synthetic challenges in accessing the 6-OH-tryptophan or its equivalents, we aimed to design an analog that would: (i) impart greater electron density into the indole system, (ii) provide an H-bond for

added recognition, and (iii) preserve a conjugation handle on the indole while bypassing the need to install the synthetically challenging 6'-OH-tryptathionine staple. To direct our efforts, we first examined the interactions between the indole core and the enzyme in the crystal structure along with the biological activity scope of the known indole-modified analogs that have been prepared from the natural product.

A review of the vast literature on derivatives of naturally sourced amatoxins suggests that the indole core is relatively tolerant to modifications. Small modifications generally result in negligible reductions in RNA pol II affinity (Fig. 1). For example, 6'-ether derivatives have been synthesized successfully in the past by treating  $\alpha$ -amanitin with alkyl halides.<sup>24</sup> Similarly, a two-fold reduction in affinity was observed when  $\alpha$ -amanitin was methylated or ethylated at position-6 to give 6'-OR- $\alpha$ -amanitin ( $R = Me, Et$ ); methyl and ethyl ethers were two-fold less active in an RNA pol II inhibition assay. However, longer chain lipophilic derivatives exhibited a much-reduced affinity towards RNA pol II even though their cell toxicity was increased likely due to improved cell penetration capacity.<sup>24</sup> Other 7'-modified derivatives of  $\alpha$ -amanitin have been synthesized, and this position was generally shown to be quite tolerant to modifications. Amine and arylazo substitutions were introduced at the 7'-position of tryptophan on  $\alpha$ -amanitin.<sup>25</sup> In 1982, Morris *et al.* evaluated a series of six secondary-amine modifications at the 7'-position and seven 7'-arylazo-amatoxins for affinity towards RNA pol II and III. All of the tested analogs were active towards RNA pol II, with  $K_i$  values up to 8 times that of  $\alpha$ -amanitin.<sup>26</sup> These reports further underscored the plasticity that the indole core offers and encouraged us to develop a new tryptophan-modified analog while addressing the remaining challenge of diastereoselective sulfoxidation.

As part of our characterization, we provide CD spectra to demonstrate what appears to be conformational differences between the thioether, (*R*)- and (*S*)-sulfoxides along with the sulfone. Since the sulfoxides show dramatic differences in the CD spectra, it suggests that they adopt significantly different conformations. Previously, however, full structural elucidation by NMR spectroscopy unexpectedly showed that the (*R*)- and (*S*)-sulfoxides had nearly overlapping structures,<sup>18</sup> and while in the crystalline state, structures of the (*S*)-sulfoxide and sulfone were nearly identical.<sup>19a</sup> To date, molecular dynamics calculations have provided little additional insight into an enduring mystery as to why the (*S*)-sulfoxide is uniquely less active than the sulfone, the (*R*)-sulfoxide<sup>27</sup> and the thioether. Hence, in terms of this work, a structural rationalization for the differences in CD spectra or cytotoxicity/ $K_i$  values, which correlate to those seen with other amanitins, is not readily intuited.

While the advantages of using an (*R*)-sulfoxide over either the thioether or sulfone as an ADC payload remain to be assessed in preclinical animal models and human clinical studies, it is reasonable to hypothesize that Nature may have chosen to install the (*R*)-sulfoxide deliberately to enhance systemic toxicity. Here, we have identified a means of achieving highly diastereoselective sulfoxidation on the precursor heptapeptide, which now enables production of a configurationally stable precursor that awaits conjugation with DHLe, the most

synthetically challenging monomer used in the synthesis. In addition, the use of a chiral catalyst to introduce the (*R*)-sulfoxide late-stage now provides access to late-stage diastereoselective sulfoxidation on bicyclic octapeptides.

The use of chiral sulfoxidation catalysts on *S*-deoxy-amanitin recapitulated similar diastereoselectivity that we report here for the 5'-hydrox-6'-deoxy analog demonstrating utility for resolving this question in the context of providing the natural product as well. Although we did not evaluate this on the heptapeptide precursor of amanitin, we believe that this method should provide similar results and thus likely augurs for success in accessing the natural product in high diastereomeric excess without resorting to chiral catalysts. Finally, we posit that one might use this approach to accomplish sulfoxidation on amanitin analogs that are produced from circularly-permuted macrocycles resembling phakellistatins.<sup>28</sup>

## Conclusions

5'-OH-6'-deoxy-amanitin, a novel and bioactive analog of  $\alpha$ -amanitin that takes advantage of the naturally abundant, commercially available 5-OH-tryptophan to incorporate a conjugation handle, was successfully synthesized. The 5-OTBS-fluoropyrrololoindoline (5-OTBS-FPI) was accessed in three simple high yielding steps from commercially available 5-OH-L-tryptophan, in contrast to addressing key oxidations on the synthetically challenging 6-OH-tryptophan. MTS cell viability and RNA pol II inhibition assays demonstrated near-native potency. It is readily appreciated that the 5'-OH can serve as a handle for further conjugation to various antibodies through a multitude of currently available strategies.<sup>11a,e</sup>

The (*R*)-sulfoxide 2 is significantly more toxic than the (*S*)-sulfoxide 12 in the context of this new analog. To access this diastereomer, we developed a late-stage sulfoxidation approach utilizing  $Ti^{IV}/L$ -DET-based catalytic system that is conveniently applied to the bicyclic toxin as the ultimate synthetic step. We suggest that this methodology has a high potential of being translatable to a broad range of bicyclic amanitin analogs due to structural similarities observed among active amatoxins. In addition, a mild, facile, and cost-effective early stage sulfoxidation approach at the monocycle stage was successfully employed. This new methodology capitalizes on what appear to be unique structural components of the heptapeptide macrocycle that must determine the diastereoselectivity of sulfoxidation that uses readily available reagents. Taken together, these sulfoxidation methods represent a great improvement in diastereoselectivity over the previously reported *mCPBA*-assisted sulfoxidation. Finally, we believe this highly selective method for (*R*)-sulfoxide formation may be broadly applicable to use on other monocyclic amanitin precursors and presents a significant advancement for future large scale amanitin syntheses.

## Conflicts of interest

There are no conflicts to declare.

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## Notes and references

- 1 T. J. Lindell, F. Weinberg, P. W. Morris, R. G. Roeder and W. J. Rutter, *Science*, 1970, **170**, 447–449.
- 2 (a) G. Moldenhauer, A. V. Salnikov, S. Luttgau, I. Herr, J. Anderl and H. Faulstich, *J. Natl. Cancer Inst.*, 2012, **104**, 622–634; (b) K. C. Nicolaou and S. Rigol, *Angew. Chem., Int. Ed.*, 2019, **58**, 11206–11241.
- 3 H. Schlesinger and W. W. Ford, *J. Biol. Chem.*, 1907, **3**, 279–283.
- 4 H. Wieland and R. Hallermayer, *Justus Liebigs Ann. Chem.*, 1941, **548**, 1–18.
- 5 (a) K. Baumann, K. Munter and H. Faulstich, *Biochemistry*, 1993, **32**, 4043–4050; (b) A. Buku, R. Altmann and T. Wieland, *Justus Liebigs Ann. Chem.*, 1974, 1580–1586; (c) G. Zanotti, C. Mohringer and T. Wieland, *Int. J. Pept. Protein Res.*, 1987, **30**, 450–459.
- 6 X. Y. Liu, L. Farnung, C. Wigge and P. Cramer, *J. Biol. Chem.*, 2018, **293**, 7189–7194.
- 7 K. Kume, M. Ikeda, S. Miura, K. Ito, K. A. Sato, Y. Ohmori, F. Endo, H. Katagiri, K. Ishida, C. Ito, T. Iwaya and S. S. Nishizuka, *Sci. Rep.*, 2016, **6**, 15.
- 8 (a) J. Anderl, C. Muller, B. Heckl-Ostreicher and R. Wehr, *Cancer Res.*, 2011, **71**, 1; (b) T. Hechler, M. Kulke, C. Mueller, A. Pahl and J. Anderl, *Cancer Res.*, 2014, **74**, 1.
- 9 L. Zhao, J. P. May, A. Blanc, D. J. Dietrich, A. Loonchanta, K. Matinkhoo, A. Pryyma and D. M. Perrin, *ChemBioChem*, 2015, **16**, 1420–1425.
- 10 G. Barbantibrodano and L. Fiume, *Nature (London), New Biol.*, 1973, **243**, 281–283.
- 11 (a) K. W. Swiderska, A. Szlachcic, L. Opalinski, M. Zakrzewska and J. Otlewski, *Int. J. Mol. Sci.*, 2018, **19**, 16; (b) L. Bodero, P. L. Rivas, B. Korsak, T. Hechler, A. Pahl, C. Muller, D. Arosio, L. Pignataro, C. Gennari and U. Piarulli, *Beilstein J. Org. Chem.*, 2018, **14**, 407–415; (c) M. A. Krzysik, L. Opalinski and J. Otlewski, *Mol. Pharm.*, 2019, **16**, 3588–3599; (d) S. Park, S. Y. Kim, J. Cho, D. Jung, D. Seo, J. Lee, S. Lee, S. Yun, H. Lee, O. Park, B. Seo, S. Kim, M. Seol, S. H. Woo and T. K. Park, *Bioconjugate Chem.*, 2019, **30**, 1969–1978; (e) S. Park, S. Y. Kim, J. Cho, D. Jung, D. Seo, J. Lee, S. Lee, S. Yun, H. Lee, O. Park, B. Seo, S. H. Woo and T. K. Park, *Bioconjugate Chem.*, 2019, **30**, 1957–1968.
- 12 (a) H. Luo, S. Y. Hong, R. M. Sgambelluri, E. Angelos, X. Li and J. D. Walton, *Chem. Biol.*, 2014, **21**, 1610–1617; (b) H. Luo, B. DuBois, R. M. Sgambelluri, E. R. Angelos, X. Li, D. Holmes and J. D. Walton, *Toxicol.*, 2015, **103**, 60–64.
- 13 (a) K. Matinkhoo, A. Pryyma, M. Todorovic, B. O. Patrick and D. M. Perrin, *J. Am. Chem. Soc.*, 2018, **140**, 6513–6517; (b)



M. A. J. Siegert, C. H. Knittel and R. D. Süssmuth, *Angew. Chem., Int. Ed.*, 2020, **59**, 5500–5504; (c) C. Lutz, W. Simon, S. Werner-Simon, A. Pahl and C. Muller, *Angew. Chem., Int. Ed.*, 2020, **59**, 11390–11393.

14 W. E. Savage and A. Fontana, *Chem. Commun.*, 1976, **15**, 600–601.

15 (a) O. Trott and A. J. Olson, *J. Comput. Chem.*, 2010, **31**, 455–461; (b) D. Seeliger and B. L. de Groot, *J. Comput.-Aided Mol. Des.*, 2010, **24**, 417–422.

16 W. L. DeLano, *The PyMOL Molecular Graphics System*, DeLano Scientific, San Carlos CA, USA, 2002.

17 A. Pryyma, Y. J. Bu, Y. Wai, B. O. Patrick and D. M. Perrin, *Org. Lett.*, 2019, **21**, 8234–8238.

18 T. Wieland, C. Gotzendorfer, J. Dabrowski, W. N. Lipscomb and G. Shoham, *Biochemistry*, 1983, **22**, 1264–1271.

19 (a) G. Shoham, W. N. Lipscomb and T. Wieland, *J. Am. Chem. Soc.*, 1989, **111**, 4791–4809; (b) G. Shoham, D. C. Rees, W. N. Lipscomb, G. Zanotti and T. Wieland, *J. Am. Chem. Soc.*, 1984, **106**, 4606–4615.

20 G. Zanotti, C. Birr and T. Wieland, *Int. J. Pept. Protein Res.*, 1981, **18**, 162–168.

21 (a) A. Buku, T. Wieland, H. Bodenmuller and H. Faulstich, *Experientia*, 1980, **36**, 33–34; (b) T. Wieland, C. Gotzendorfer, G. Zanotti and A. C. Vaisius, *Eur. J. Biochem.*, 1981, **117**, 161–164.

22 J. P. May, P. Fournier, B. O. Patrick and D. M. Perrin, *Chem.–Eur. J.*, 2008, **14**, 3410–3417.

23 (a) F. Difuria, G. Modena and R. Seraglia, *Synthesis*, 1984, 325–326; (b) J. L. Han, V. A. Soloshonok, K. D. Klika, J. Drabowicz and A. Wzorek, *Chem. Soc. Rev.*, 2018, **47**, 1307–1350; (c) P. Pitchen and H. B. Kagan, *Tetrahedron Lett.*, 1984, **25**, 1049–1052.

24 H. Faulstich, H. Trischmann, T. Wieland and E. Wulf, *Biochemistry*, 1981, **20**, 6498–6504.

25 (a) E. Falckpedersen, P. W. Morris and D. L. Venton, *Int. J. Pept. Protein Res.*, 1983, **21**, 431–439; (b) P. W. Morris, D. L. Venton and K. M. Kelley, *Biochemistry*, 1978, **17**, 690–698.

26 E. Falckpedersen, W. Neuman and P. W. Morris, *Biochemistry*, 1982, **21**, 5164–5170.

27 A. B. Pomilio, M. E. Battista and A. A. Vitale, *J. Mol. Struct.: THEOCHEM*, 2001, **536**, 243–262.

28 J. P. May and D. M. Perrin, *Chem.–Eur. J.*, 2008, **14**, 3404–3409.

