

Cite this: *Chem. Sci.*, 2020, **11**, 13079

All publication charges for this article have been paid for by the Royal Society of Chemistry

Received 28th July 2020
Accepted 19th October 2020

DOI: 10.1039/d0sc04136j

rsc.li/chemical-science

Direct, stereoselective thioglycosylation enabled by an organophotoredox radical strategy†

Peng Ji, Yueteng Zhang, Feng Gao, Fangchao Bi and Wei Wang *

While strategies involving a $2e^-$ transfer pathway have dictated glycosylation development, the direct glycosylation of readily accessible glycosyl donors as radical precursors is particularly appealing because of high radical anomeric selectivity and atom- and step-economy. However, the development of the radical process has been challenging owing to notorious competing reduction, elimination and/or S_N side reactions of commonly used, labile glycosyl donors. Here we introduce an organophotocatalytic strategy through which glycosyl bromides can be efficiently converted into corresponding anomeric radicals by photoredox mediated HAT catalysis without a transition metal or a directing group and achieve highly anomeric selectivity. The power of this platform has been demonstrated by the mild reaction conditions enabling the synthesis of challenging α -1,2-*cis*-thioglycosides, the tolerance of various functional groups and the broad substrate scope for both common pentoses and hexoses. Furthermore, this general approach is compatible with both sp^2 and sp^3 sulfur electrophiles and late-stage glycodiversification for a total of 50 substrates probed.

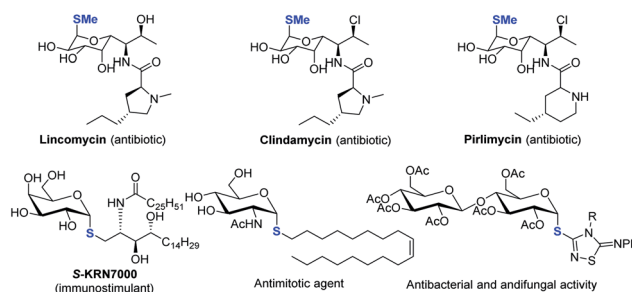
Introduction

Despite the fact that *O*-linked glycosides are a dominant form in biologically important glycoconjugates,¹ the replacement of “*O*” by *C*-, *N*- and *S*-linked glycosides offers the merits of improved hydrolytic stability and/or bioactivity while maintaining similar conformational preferences.² In particular, thioglycosides have emerged as a privileged class of structures owing to their broad spectrum of biological activities (see representative examples in Scheme 1).^{2–5} Moreover, they are widely used as glycosyl donors in glycosylation reactions.⁶ The broad biological and synthetic utility has triggered significant interest in the development of efficient methods to construct a C–S bond with a defined anomeric configuration, which plays key roles in biological activities.

Strategies involving an ionic $2e^-$ transfer pathway have dictated the C–S bond formation development.^{7–13} Direct replacement by a thiol with a glycosyl donor is an attractive approach in that both starting materials are readily accessible, but gives a mixture of α/β anomers in most cases (Scheme 2a).⁸ To overcome these limitations, the methods of reversing the polarity at the anomeric carbon have been developed (Scheme 2b).⁹ These elegant methods enable the stereoselective control formation of both α and β anomers but with limited scope of saccharides.^{9a} Indirect methods using preformed anomeric

thiols offer versatile approaches to thioglycosides (Scheme 2c).^{10–13} Nonetheless, the anomeric stereoselectivity of these processes depends on the nature of the anomeric thiols. In particular, few methods are capable of selectively constructing the challenging α -1,2-*cis*-thioglycosides,^{8b} featured in a number of natural products and bioactive molecules (Scheme 1).

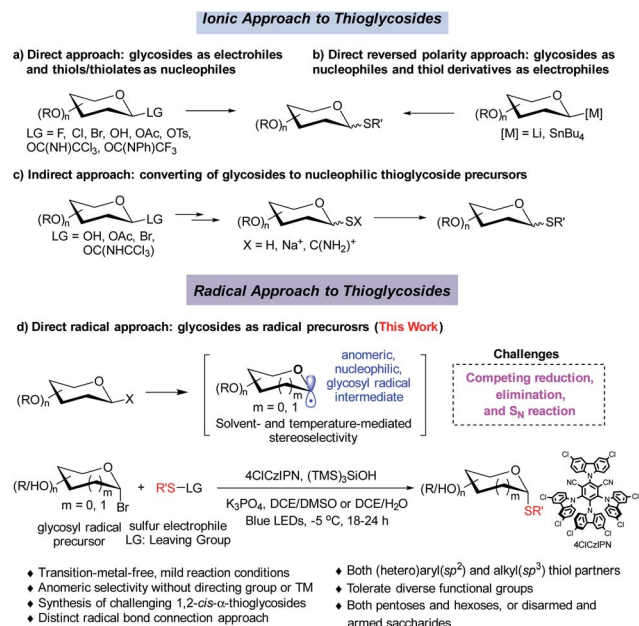
Radical cross coupling offers a distinct paradigm for stereoselective construction of glycosidic bonds.¹⁴ Anomeric radicals have been elegantly explored for highly stereoselective *C*-glycosidic bond formation with a transition metal (TM).^{15–17} However, stereoselective C–S bond formation through the glycosyl radical has remained elusive (Scheme 2d).¹⁷ This is attributed to: (1) reduction of glycosyl radicals by HAT (hydrogen atom transfer) donors;¹⁸ (2) elimination reaction of labile glycosyl donors with a TM catalyst;¹⁹ (3) competing S_N2 reaction with thiols, which could compromise the anomeric selectivity.^{2b,7} Therefore, stable radical precursors such as



Scheme 1 Selected examples of thioglycosides with α -1,2-*cis*-configuration.

Departments of Pharmacology and Toxicology and Chemistry and Biochemistry, BIO5 Institute, and University of Arizona Cancer Centre, University of Arizona, Tucson, AZ 85721, USA. E-mail: wwang@pharmacy.arizona.edu

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0sc04136j



Scheme 2 Ionic and radical thioglycosylation.

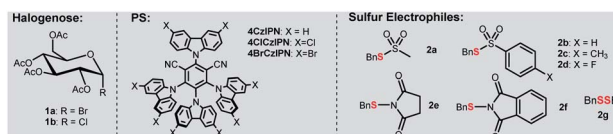
glycosyl stannanes are designed to minimize these issues.¹⁷ Given the fact that the glycosyl radical can favour the formation of anomeric C1 conformation, we deliberately push the limit by developing an organophotocatalytic approach without a directing group or a TM for stereoselective *S*-glycosylation. Herein, we wish to disclose the results of the investigation, which has led to a general organophotocatalyzed thiolation of glycosyl bromides with highly stereoselective control (Scheme 2d).

Results and discussion

In our own efforts, recently we have developed visible-light-mediated glycosyl radical reactions for the synthesis of *C*-glycosides.¹⁵ In addition, we reported an organophotocatalytic thiolation of acyl radical method with thiosulfonates.²⁰ These chemistries guided us to explore a new thioglycosylation reaction. The reaction of α-glucopyranosyl bromide **1a** with thiosulfonate **2a** and 4CzIPN²¹ as a photocatalyst (PS) was probed (Table 1 and Tables S1–S6†). First, we examined several commonly used reductants including *i*Pr₂NEt, Hantzsch ester, and ascorbic acid (Table S1,† entries 2, 6 and 7) for the

Table 1 Reaction optimization

Entry	Variation from the “standard conditions” ^a	Yield ^b (3a , %)	α : β ^c
1	None	76 (72) ^d	>20 : 1
2	4CzIPN (5 mol%), 2c , Na ₂ CO ₃ (4.0 equiv.), DMSO, rt	37	<10 : 1
3	4BrCzIPN (5 mol%), 2c , Na ₂ CO ₃ (4.0 equiv.), DMSO, rt	33	<10 : 1
4	4ClCzIPN (5 mol%), 2c , Na ₂ CO ₃ (4.0 equiv.), DMSO, rt	65	<10 : 1
5	CS ₂ CO ₃ instead of K ₃ PO ₄ , DCE : DMSO (1 : 1, v/v), rt	Trace	—
6	DCE : DMSO (1 : 1, v/v), rt	80	<10 : 1
7	2b instead of 2a , DCE : DMSO (1 : 1, v/v), rt	72	<10 : 1
8	2d instead of 2a , DCE : DMSO (1 : 1, v/v), rt	66	<10 : 1
9	2d instead of 2a , DCE : DMSO (1 : 1, v/v), rt	68	<10 : 1
10	2e instead of 2a , DCE : DMSO (1 : 1, v/v), rt	Trace	—
11	2f instead of 2a , DCE : DMSO (1 : 1, v/v), rt	66	<10 : 1
12	2g instead of 2a , DCE : DMSO (1 : 1, v/v), rt	Trace	—
13	1b instead of 1a	Trace	—
14	DCE instead of DCE : H ₂ O (2 : 1, v/v), rt	60	17 : 1
15	DCE instead of DCE : H ₂ O (2 : 1, v/v), −5 °C	67	>20 : 1
16	Without 4ClCzIPN, (TMS) ₃ SiOH or K ₃ PO ₄	Trace	—
17	Under dark conditions	Trace	—



^a Standard conditions: unless specified, a mixture of glycosyl bromide (0.2 mmol), sulfur electrophile (0.1 mmol), 4ClCzIPN (0.005 mmol), K₃PO₄ (0.4 mmol), and (TMS)₃SiOH (0.15 mmol) in DCE/DMSO (1 mL, 1 : 1, v/v) or DCE/H₂O (1.5 mL, 2 : 1, v/v) was irradiated with 40 W Kessil blue LEDs in a N₂ atmosphere at −5 °C for 24 h. ^b Yield determined by ¹H NMR using 1,1,2,2-tetrachloroethane as an internal reference. ^c Ratio determined by crude ¹H NMR. ^d Isolated yield.

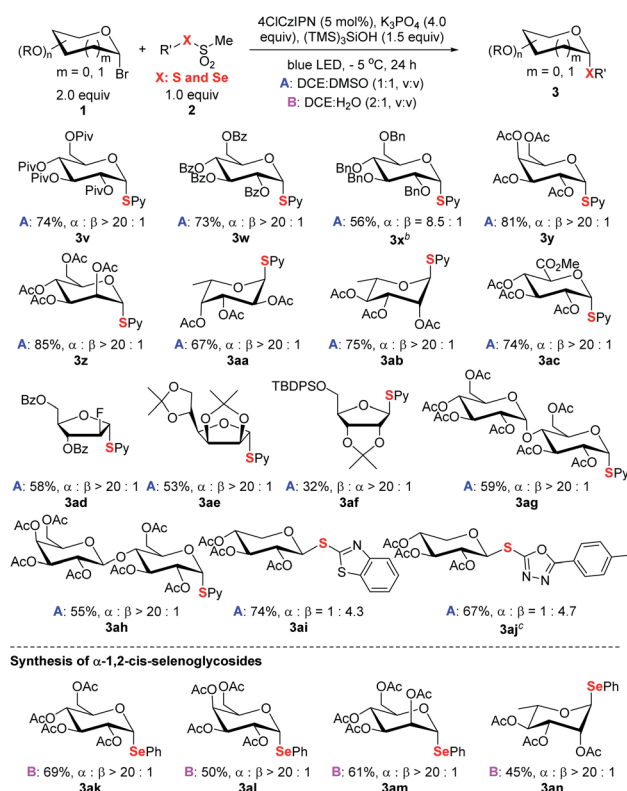
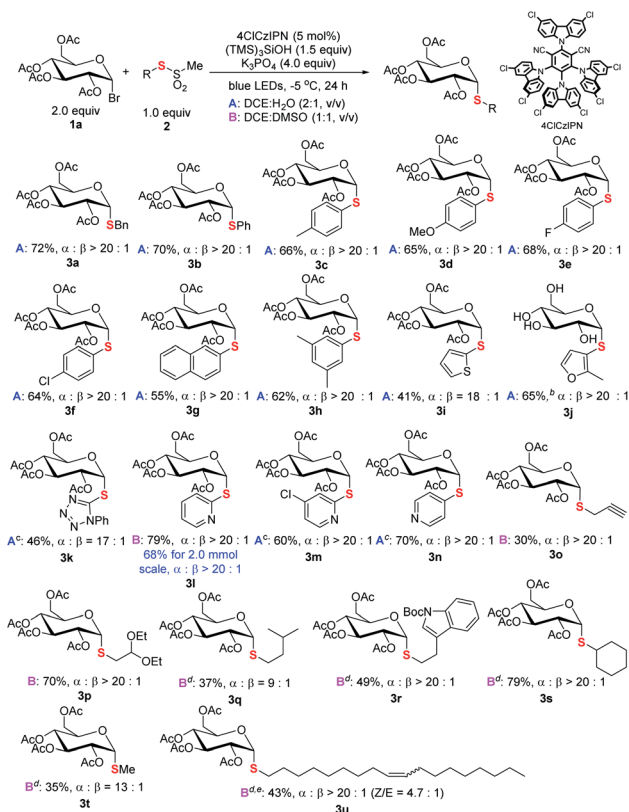


generation of the glycosyl radical. Disappointingly, only the reduced product **4** was obtained. It should be pointed out that this is a general problem in using glycosyl halides as radical progenitors in glycosylation.¹⁸ Minimizing the issue requires a radical capable of effective dehalogenation whereas the hydrogenated product should be a weak H-donor. A silyl or a silyloxy radical can induce dehalogenation while the strong Si-H and Si-O-H make them more difficult to abstract.²² Therefore, various silanes were screened and (TMS)₃SiOH was the best, giving **3a** in 37% yield (Table S1,† entries 3–5 and 8–9). A survey of PSs revealed 4ClCzIPN^{21b,c} as the optimal promoter (65% yield, Table S2† and 1, entries 2–4). The process was also sensitive to bases (entries 4–6 and Table S4†) and K₃PO₄ gave **3a** in high yield. Among the thiosulfonates probed (entries 6–12), methanethiosulfonate (**2a**) was the best, possibly attributed to the lower hindrance and relatively high redox stability ($E_{\text{red}} = -1.65$ V vs. SCE, Fig. S3†). Glycosyl chloride (**1b**) did not undergo the dechlorination presumably due to its strong C-Cl bond (entry 13). To further improve the stereoselectivity (entry 6), we conducted reaction optimization including the solvent and reaction temperature (Table 1, entries 14–15 and Tables S3 and S6†). It was found that the biphasic solvent (DCE : H₂O = 2 : 1) could not only retain the high anomeric selectivity but also increase the yield (76%, entry 1), and a low temperature (−5 °C)

is also required to maintain good yield and anomeric selectivity (entry 14, 15). The control experiments confirmed that base, light, (TMS)₃SiOH, and PS were essential for this transformation (entries 16–17).

The generality of the new *S*-glycosylation was examined. We first evaluated the performance using glucosyl bromide (**1a**) as a radical donor for coupling with various thiosulfonates **2** (Scheme 3). The process serves as a general approach to both aryl and alkyl thioglycosides. Uniformly high axial selectivities are observed regardless of the nature of the sulfur electrophiles. With respect to aryls, electron-neutral (**3b**), -donating (**3c–3d**, **3h**), and -withdrawing (**3e–3f**) groups on the phenyl ring and fused aromatic (**3g**) can be tolerated. Moreover, heteroaromatic thiosulfonates such as thiophenyl (**3i**) and furanyl (**3j**) enabled access to medicinally valued thioglycosides. The tetrazole derived disulfide instead of labile thiosulfonate could serve as an alternative and delivered the desired **3k**. The reaction performed in DCE : H₂O failed for pyridinyl thiosulfonate. Decent results (**3l**, 79%, $\alpha : \beta > 20 : 1$) were obtained with DCE : DMSO (condition B). The protocol can also be applied in gram scale synthesis. Notably, less reactive sp³ alkyl glycosides **3o–3s** could be synthesized with the protocol.¹⁷

For even less electrophilic substrates, *p*-tolylthiosulfonates (**3q–3u**) displayed better performance than methylthiosulfonates. Particularly, a long alkyl chain with a *Z*-double bond product (**3u**), which exhibits intriguing antitumor activity



Scheme 3 Scope of thiosulfonates. ^aReaction conditions: unless specified, see footnote a of Table 1 and the ESI;† isolated yield; the ratio of α and β anomers determined by crude ¹H NMR. ^bYield after hydrolysis of the acyl group. ^cDisulfide used. ^dToluenethiosulfonate used. ^e*Z/E* ratio determined by ¹H NMR.

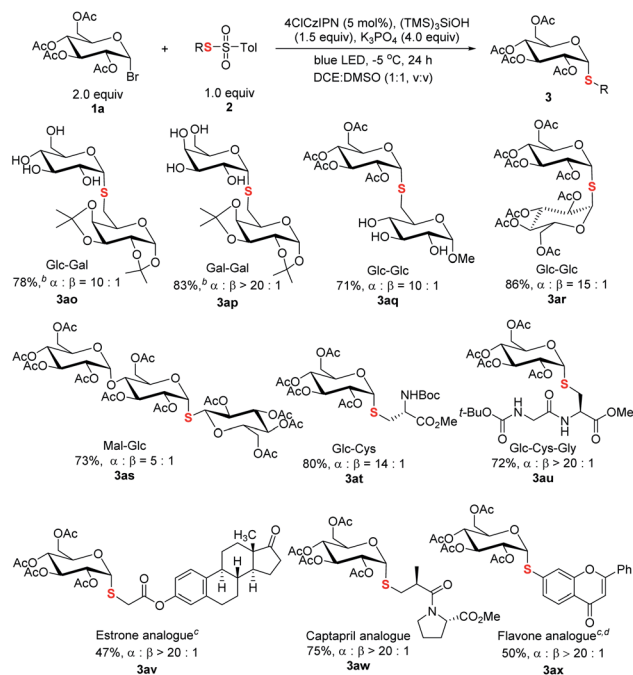
Scheme 4 Scope of saccharides and selenoglycosylation. ^aReaction conditions: unless specified see footnote a of Table 1 and the ESI;† isolated yield; the ratio of α and β anomers determined by crude ¹H NMR. ^b3.0 equiv. of glycosyl bromide used. ^cDisulfide used.



(Scheme 1), is efficiently prepared with high diastereoselectivity. The limitation of the method is also realized. C₂-N-Ac-saccharides such as D-glucosamine failed to react due to the lability of these reactants (see Fig. S5 in the ESI†).

The alternation of sugars was probed next (Scheme 4). Both common hexoses (glucose **3v–3x**, galactose **3y**, mannose **3z**, fucose **3aa**, rhamnopyranose **3ab**, and glucuronic acid **3ac**) and pentoses (**3ad–3af**) gave good yields and high stereoselectivity. Among the tested monosaccharides, except ribose (**3af**) adopting expected β selectivity owing to the steric effect, the others gave expected α -selectivity. Furthermore, disaccharides (**3ag** and **3ah**) could participate in the process smoothly. For xyloses (**3ai–3aj**), the obtained products adopted β orientation since the anomeric xylosyl radical is β selective.²³ Besides pyridyl (Py), other pharmaceutically relevant heteroaromatics such as benzothiazole and oxadiazole (**3ai**, **3aj**) could be efficiently incorporated. This offers a viable strategy for the synthesis of xylose-derived bioactive analogs.^{4c} Finally, the strategy can also be extended for the synthesis of synthetically challenging α -1,2-*cis*-selenoglycosides (Scheme 4 and Table S7†).²⁴ For example, under the reaction conditions (see footnote a of Table 1, DCE : H₂O, v/v, 2 : 1), four glycosyl bromides could couple with methyl phenylselenenyl sulfonate to deliver the corresponding α -seleno-glycosides **3ak–3an** with uniformly high stereoselectivity ($\alpha : \beta > 20 : 1$).

The capacity of selective functionalization of biologically relevant structures and therapeutics is the testament to the synthetic power of a methodology. As demonstrated (Scheme 5),



thiosulfonate **2**. In the reactions, we still observed a notable amount of the reduction product **4**. It is believed that it is produced from the reaction of **8** with (TMS)₃SiOH, which was confirmed by deuteration experiments with observed deuterated product **4-d** (Scheme 6b). This also rationalizes that 2 equiv. of glycosyl bromide **1** is used to ensure high efficiency of the thio-glycosylation process. Finally, a radical trapping study with TEMPO and methyl acrylate^{16d} further confirms the radical engaged process (Scheme 6c).

Conclusions

In conclusion, we have developed a metal-free, glycosyl radical strategy for the stereoselective synthesis of thioglycosides by employing commonly used glycosyl bromides as radical precursors. The uncovered organophotoredox mediated HAT radical pathway can highly stereoselectively induce the formation of an anomeric C–S bond while minimizing the side reactions. The power of the platform has been underscored by the mild reaction conditions enabling the synthesis of challenging α -1,2-*cis*-thioglycosides, the tolerance of various functional groups and the broad substrate scope for both common pentoses and hexoses. Furthermore, this general approach is compatible with both sp² and sp³ sulfur electrophiles and late-stage glycodiversification. It is expected that the strategy enabling the efficient generation of glycosyl radicals from labile glycosyl bromides can offer a reliable alternative for the synthesis of C- and other hetero-glycosides.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Financial support was provided by the NIH (5R01GM125920-04) and the NSF MRI for the acquisition of a 500 MHz NMR spectrometer (1920234).

Notes and references

- (a) P. M. Rudd, T. Elliot, P. Cresswell, I. A. Wilson and R. A. Dwek, *Science*, 2001, **291**, 2370; (b) B. G. Davis, *Chem. Rev.*, 2002, **102**, 579; (c) K. J. Doores, D. P. Gamblin and B. G. Davis, *Chem.–Eur. J.*, 2006, **12**, 656; (d) J. B. Lowe, *Cell*, 2001, **104**, 809.
- (a) D. A. Thayer, H. N. Yu, M. C. Galan and C.-H. Wong, *Angew. Chem., Int. Ed.*, 2005, **44**, 4596; (b) K. Pachamuthu and R. R. Schmidt, *Chem. Rev.*, 2006, **106**, 160; (c) C. S. Rye and S. G. Withers, *Carbohydr. Res.*, 2004, **339**, 699; (d) Z. Amso, S. W. Bisset, S.-H. yang, P. W. R. Harris, T. H. Wright, C. D. Navo, M. L. Patchett, G. E. Norris and M. A. Brimble, *Chem. Sci.*, 2018, **9**, 1686.
- (a) S. Schwarz, J. Shen, K. Kadlec, Y. Wang, G. B. Michael, A. T. Feßler and B. Vester, *Cold Spring Harb. Perspect. Med.*, 2016, **6**, a027037; (b) T. J. Oman, J. M. Boettcher, H. Wang, X. N. Okalibe and W. A. van der Donk, *Nat. Chem. Biol.*, 2011, **7**, 78; (c) Y. S. Y. Hsieh, B. L. Wilkinson, M. R. O'Connell, J. P. Mackay, J. M. Matthews and R. J. Payne, *Org. Lett.*, 2012, **14**, 1910; (d) S. Biswas, C. V. Garcia De Gonzalo, L. M. Repka and W. A. van der Donk, *ACS Chem. Biol.*, 2017, **12**, 2965.
- (a) Z. J. Witczak, *Curr. Med. Chem.*, 1999, **6**, 165; (b) Z. J. Witczak, P. Kaplon and P. Markus Dey, *Carbohydr. Res.*, 2003, **338**, 11; (c) W. A. El-Sayed, N. M. Fathi, W. A. Gad and E. S. H. El-Ashry, *J. Carbohydr. Chem.*, 2008, **27**, 357.
- (a) R. N. Comber, J. D. Friedrich, D. A. Dunshee, S. L. Petty and J. A. Secrist, *Carbohydr. Res.*, 1994, **262**, 245; (b) D. V. Mangte and S. P. Deshmukh, *Heteroat. Chem.*, 2007, **18**, 390; (c) W. A. El-Sayed, N. M. Fathi, W. A. Gad and E. S. H. El-Ashry, *J. Carbohydr. Chem.*, 2008, **27**, 357.
- (a) D. Ikuta, Y. Hirata, S. Wakamori, H. Shimada, Y. Tomabechei, Y. Kawasaki, K. Ikeuchi, T. Hagimori, S. Matsumoto and H. Yamada, *Science*, 2019, **364**, 674; (b) M. Lahmann and S. Oscarson, *Org. Lett.*, 2000, **2**, 3881; (c) S. S. Nigudkar and A. V. Demchenko, *Chem. Sci.*, 2015, **6**, 2687; (d) M. L. Spell, K. Deveau, C. G. Bresnahan, B. L. Bernard, W. Sheffield, R. Kumar and J. R. Ragains, *Angew. Chem., Int. Ed.*, 2016, **55**, 6515; (e) G. Lian, X. Zhang and B. Yu, *Carbohydr. Res.*, 2015, **403**, 13; (f) B. Dhakal and D. Crich, *J. Am. Chem. Soc.*, 2018, **140**, 15008; (g) H.-Y. Wang, S. A. Blaszczyk, G. Xiao and W. Tang, *Chem. Soc. Rev.*, 2018, **47**, 681.
- (a) M. Gerz, H. Matter and H. Kessler, *Angew. Chem., Int. Ed. Engl.*, 1993, **32**, 269; (b) J. Ramos-Soriano, U. Niss, J. Angulo, M. Angulo, A. J. Moreno-Vargas, A. T. Carmona, S. Ohlson and I. Robina, *Chem.–Eur. J.*, 2013, **19**, 17989; (c) P. J. Pfäffli, S. H. Hixson and L. Anderson, *Carbohydr. Res.*, 1972, **23**, 195; (d) T. Fujihira, T. Takido and M. Seno, *J. Mol. Catal. A: Chem.*, 1999, **137**, 65.
- (a) S. Ecoply, Y. Singh and A. V. Demchenko, *Org. Biomol. Chem.*, 2019, **17**, 8379; (b) S. Zhu, G. Samala, E. T. Sletten, J. L. Stockdill and H. M. Nguyen, *Chem. Sci.*, 2019, **10**, 10475; (c) D. Hirofumi and N. Yoshihiro, *Trends Glycosci. Glycotechnol.*, 2014, **26**, 119.
- (a) K. N. Baryal, D. Zhu, X. Li and J. Zhu, *Angew. Chem., Int. Ed.*, 2013, **52**, 8012; (b) F. Zhu, E. Miller, S. Zhang, D. Yi, S. O'Neill, X. Hong and M. A. Walczak, *J. Am. Chem. Soc.*, 2018, **140**, 18140.
- Alkylation/arylation methods: (a) H. Driguez, *ChemBiochem*, 2001, **2**, 311; (b) R. Komor, A. Kasprzycka, G. Pastuch-Gawolek and W. Szeja, *Carbohydr. Res.*, 2014, **396**, 37; (c) R. T. Dere, A. Kumar, V. Kumar, X. Zhu and R. R. Schmidt, *J. Org. Chem.*, 2011, **76**, 7539; (d) F. M. Ibatullin, K. A. Shabalin, J. V. Jänis and A. G. Shavva, *Tetrahedron Lett.*, 2003, **44**, 7961.
- Cross coupling: (a) D. Montoir, M. Amoura, Z. E. A. Ababsa, T. M. Vishwanatha, E. Yen-Pon, E. V. Robert, M. Beltramo, V. Piller, M. Alami, V. Aucagne and S. Messaoudi, *Chem. Sci.*, 2018, **9**, 8753; (b) M. Zhu, G. Dagousset, M. Alami, E. Magnier and S. Messaoudi, *Org. Lett.*, 2019, **21**, 5132.
- Thiol-ene reaction and Michael addition: (a) G. Zhao, S. Kaur and T. Wang, *Org. Lett.*, 2017, **19**, 3291; (b) M. Fiore,



- M. L. Conte, S. Pafifico, A. Marra and A. Dondoni, *Tetrahedron Lett.*, 2011, **52**, 444; (c) N. Merbouh, F. K. Wallner, O. M. Cociorva and P. H. Seeberger, *Org. Lett.*, 2007, **9**, 651; (d) Z. J. Witzak, H. Chen and P. Kaplon, *Tetrahedron: Asymmetry*, 2000, **11**, 519; (e) Y. Zhu and W. A. van der Donk, *Org. Lett.*, 2001, **3**, 1189.
- 13 Nucleophilic substitution: (a) D. P. Gamblin, P. Garnier, S. Kasteren, N. J. Oldham, A. J. Fairbanks and B. G. Davis, *Angew. Chem., Int. Ed.*, 2004, **43**, 828; (b) G. J. L. Bernardes, D. P. Gamblin and B. G. Davis, *Angew. Chem., Int. Ed.*, 2006, **45**, 4007; (c) B. G. Davis, M. A. T. Maughan, M. P. Green, A. Ullman and J. B. Jones, *Tetrahedron: Asymmetry*, 2000, **11**, 245; (d) D. P. Galonić, W. A. van der Donk and D. Gin, *J. Am. Chem. Soc.*, 2004, **126**, 12712; (e) D. P. Galonić, N. D. Ide, W. A. van der Donk and D. Y. Gin, *J. Am. Chem. Soc.*, 2005, **127**, 7359.
- 14 J. M. Smith, S. J. Harwood and P. S. Baran, *Acc. Chem. Res.*, 2018, **51**, 1807.
- 15 (a) P. Ji, Y. Zhang, Y. Wei, H. Huang, W. Hu, P. A. Mariano and W. Wang, *Org. Lett.*, 2019, **21**, 3086; (b) P. Ji, Y. Zhang, Y. Dong, H. Huang, Y. Wei and W. Wang, *Org. Lett.*, 2020, **22**, 1557; (c) Y. Ma, S. Liu, Y. Xi, H. Li, K. Yang, Z. Cheng, W. Wang and Y. Zhang, *Chem. Commun.*, 2019, **55**, 14657; (d) N. Kiya, Y. Hidaka, K. Usui and G. Hirai, *Org. Lett.*, 2019, **21**, 1588–1592; (e) Y. Hidaka, N. Kiya, M. Yoritake, K. Usui and G. Hirai, *Chem. Commun.*, 2020, **56**, 4712–4715.
- 16 (a) A review, see: Y. Yang and B. Yu, *Chem. Rev.*, 2017, **117**, 12281; (b) N. Miquel, G. Doisneau and J.-M. Beau, *Angew. Chem., Int. Ed.*, 2000, **39**, 4111; (c) H. Abe, S. Shuto and A. Matsuda, *J. Am. Chem. Soc.*, 2001, **123**, 11870; (d) R. S. Andrews, J. J. Becker and M. R. Gagné, *Angew. Chem., Int. Ed.*, 2010, **49**, 7274; (e) F. Toriyama, J. Cornella, L. Wimmer, T.-G. Chen, D. D. Dixon, G. Creech and P. S. Baran, *J. Am. Chem. Soc.*, 2016, **138**, 11132; (f) K. Masuda, M. Nagatomo and M. Inoue, *Nat. Chem.*, 2017, **9**, 207; (g) A. Dumoulin, J. K. Matsui, A. Gutierrez-Bonet and G. A. Molander, *Angew. Chem., Int. Ed.*, 2018, **57**, 6614; (h) S. O. Badir, A. Dumoulin, J. K. Matsui and G. A. Molander, *Angew. Chem., Int. Ed.*, 2018, **57**, 6610.
- 17 (a) During the preparation of this manuscript, Hong, Walczak and coworkers reported a Cu(i) and blue LED co-catalyzed stereoselective thioglycosylation of glycosyl stannanes with disulfides: F. Zhu, S.-Q. Zhang, Z. Chen, J. Rui, X. Hong and M. A. Walczak, *J. Am. Chem. Soc.*, 2020, **142**, 11102. (b) A similar strategy reported by Luo and Nguyen used for impressive stereoselective formation of glycosidic C–O bonds: F. Yu, J. L. Dickson, R. S. Loka, H. Xu, R. N. Schaugaaard, H. B. Schlegel, L. Luo and H. M. Nguyen, *ACS Catal.*, 2020, **10**, 5990.
- 18 (a) R. S. Andrews, J. J. Becker and M. R. Gagné, *Org. Lett.*, 2011, **13**, 2406; (b) S. Yamago and A. Matsumoto, *J. Org. Chem.*, 2008, **73**, 7300; (c) J. Guiard, Y. Rhali and J.-P. Praly, *Eur. J. Org. Chem.*, 2014, 4461.
- 19 (a) N. Miquel, G. Doisneau and J.-M. Beau, *Angew. Chem., Int. Ed.*, 2000, **39**, 4111; (b) H. Gong and M. R. Gagné, *J. Am. Chem. Soc.*, 2008, **130**, 12177; (c) R. P. Spencer, C. L. Cavallaro and J. Schwartz, *J. Org. Chem.*, 1999, **64**, 3987.
- 20 Y. Zhang, P. Ji, W. Hu, Y. Wei, H. Huang and W. Wang, *Chem.–Eur. J.*, 2019, **25**, 8225.
- 21 (a) H. Uoyama, K. Goushi, K. Shizu, H. Nomura and C. Adachi, *Nature*, 2012, **492**, 234; (b) F. Le Vaillant, M. Garreau, S. Nicolai, G. Gryn'ova, C. Corminboeuf and J. Waser, *Chem. Sci.*, 2018, **9**, 5883; (c) A. Kretzschmar, C. Patze, S. T. Schwaebel and U. H. F. Bunz, *J. Org. Chem.*, 2015, **80**, 9126; (d) a review, see: T.-Y. Shang, L.-H. Lu, Z. Cao, Y. Liu, W.-M. He and B. Yu, *Chem. Commun.*, 2019, **55**, 5408.
- 22 (a) C. Chatgililoglu, *Chem. Rev.*, 1995, **95**, 1229; (b) C. Le, T. Q. Chen, C. Liang, P. Zhang and D. W. C. MacMillan, *Science*, 2018, **360**, 1010; (c) R. T. Smith, X. Zhang, J. A. Rincon, J. Agejas, C. Mateo, M. Barberis, S. Garcia-Cerrada, O. de Frutos and D. W. C. MacMillan, *J. Am. Chem. Soc.*, 2018, **140**, 17433; (d) M. S. Lowry, J. I. Goldsmith, J. D. Slinker, R. Rohl, R. A. Pascal, G. G. Malliaras and S. Bernhard, *Chem. Mater.*, 2005, **17**, 5712.
- 23 (a) B. Giese, J. Dupuis, M. Leising, M. Nix and H. J. Lindner, *Carbohydr. Res.*, 1987, **171**, 329–341; (b) H. Abe, S. Shuto and A. Matsuda, *J. Am. Chem. Soc.*, 2001, **123**, 11870.
- 24 (a) A. W. McDonagh, M. F. Mahon and P. V. Murphy, *Org. Lett.*, 2016, **18**, 552; (b) S. Czernecki and D. Randriamandimby, *J. Carbohydr. Chem.*, 1996, **15**, 183; (c) F. Santoyo-Gonzalez, F. G. Calvo-Flores, P. Garcia-Mendoza, F. Hernandez-Mateo, J. Isac-Garcia and R. Robles-Dia, *J. Org. Chem.*, 1993, **58**, 6122; (d) V. Di Bussolo, A. Fiasella, F. Balzano, G. Uccello Barretta and P. Crotti, *J. Org. Chem.*, 2010, **75**, 4284.
- 25 (a) S. Izumi, Y. Kobayashi and Y. Takemoto, *Angew. Chem., Int. Ed.*, 2020, **59**, 14054; (b) J. Defaye and A. Gadelle, *Carbohydr. Res.*, 1991, **217**, 51.
- 26 F. Denes, M. Pichowicz, G. Povie and P. Renaud, *Chem. Rev.*, 2014, **114**, 2587.
- 27 <http://butane.chem.uiuc.edu/cyerkes/Chem104ACSpring2009/Genchemref/bondenergies.html>.
- 28 M. D. Paredes and R. Alonso, *J. Org. Chem.*, 2000, **65**, 2292.

