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Kinetically controlled Ag⁺-coordinated chiral supramolecular polymerization accompanying a helical inversion†

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We report kinetically controlled chiral supramolecular polymerization based on ligand–metal complex with a 3 : 2 (L : Ag⁺) stoichiometry accompanying a helical inversion in water. A new family of bipyridine-based ligands (D-L¹, L-L¹, D-L², and D-L³) possessing hydrazine and D- or L-alanine moieties at the alkyl chain groups has been designed and synthesized. Interestingly, upon addition of AgNO₃ (0.5–1.3 equiv.) to the D-L¹ solution, it generated the aggregate I composed of the D-L¹AgNO₃ complex (D-L¹ : Ag⁺ = 1 : 1) as the kinetic product with a spherical structure. Then, aggregate I (nanoparticle) was transformed into the aggregate II (supramolecular polymer) based on the (D-L¹)₃Ag₂(NO₃)₂ complex as the thermodynamic product with a fiber structure, which led to the helical inversion from the left-handed (M-type) to the right-handed (P-type) helicity accompanying CD amplification. In contrast, the spherical aggregate I (nanoparticle) composed of the D-L¹AgNO₃ complex with the left-handed (M-type) helicity formed in the presence of 2.0 equiv. of AgNO₃ and was not additionally changed, which indicated that it was the thermodynamic product. The chiral supramolecular polymer based on (D-L¹)₃Ag₂(NO₃)₂ was produced *via* a nucleation–elongation mechanism with a cooperative pathway. In thermodynamic study, the standard ΔG° and ΔH_e values for the aggregates I and II were calculated using the van't Hoff plot. The enhanced ΔG° value of the aggregate II compared to that of the formation of aggregate I confirms that aggregate II was thermodynamically more stable. In the kinetic study, the influence of concentration of AgNO₃ confirmed the initial formation of the aggregate I (nanoparticle), which then evolved to the aggregate II (supramolecular polymer). Thus, the concentration of the (D-L¹)₃Ag₂(NO₃)₂ complex in the initial state plays a critical role in generating aggregate II (supramolecular polymer). In particular, NO₃[−] acts as a critical linker and accelerator in the transformation from the aggregate I to the aggregate II. This is the first example of a system for a kinetically controlled chiral supramolecular polymer that is formed *via* multiple steps with coordination structural change.

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Introduction

Control of helicity in dynamic supramolecular assemblies based on metal–ligand coordinated architectures such as capsules, cages, and helices has rapidly developed into one of the most successful and versatile strategies in supramolecular chemistry,^{1–5} largely due to its essential role in biomolecules

such as DNA and proteins. In addition, numerous artificial helical systems, ranging from oligonucleotides and peptide-mimetics to oligomers and polymers, have been developed.

Metallosupramolecular polymers based on metal–ligand coordination have attracted much attention as a significant class of organic/inorganic hybrid supramolecular materials.^{6–30} Because of the incorporation of metal complexes in metallosupramolecular polymers, these metal-containing systems exhibit redox, optical, electrochromic, catalytic, and magnetic properties as well as the usual properties of organic polymers.^{31–34} Chiral metallosupramolecular polymers are particularly attractive because they are adaptable to chiral recognition and sensing, and asymmetric catalysis.^{35,36}

In particular, recent advances in kinetic control over the dynamic structural and functional state of the metastable state (*i.e.*, the out-of-equilibrium state) in supramolecular systems has allowed the development of supramolecular polymers with

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unprecedented structural complexity and sophisticated functionality.^{15–17,37–43} Such metastable states, in which spontaneous self-assembly is kinetically suppressed owing to competing aggregation and intramolecular hydrogen bonding, have been utilized to control the self-assembly behavior of supramolecular polymers.^{16,40,41} Furthermore, understanding artificial supramolecular systems from kinetic perspectives has improved in terms of pathway complexity.^{39,41,43–45} Moreover, the growth mechanisms of supramolecular polymerization can now be interpreted by the thermodynamic (cooperative or isodesmic) models proposed by Meijer *et al.*^{42,43,46} Such studies have demonstrated that self-assembly pathways play a significant role in determining thermodynamic or kinetic products. Thus, these previous works demonstrate that achieving kinetically programmable out-of-equilibrium states in supramolecular systems is a significant challenge related to achieving desired characteristics for dynamic self-assembly processes in supramolecular polymerization. For example, Besenius and co-workers demonstrated the kinetically controlled supramolecular polymerization of an Au(I)-metallopeptide in water.³⁷ This metal complex kinetically assembles into metastable nanorods at room temperature but is converted into a thermodynamic equilibrium fibril state upon heating. Similarly, Fernández and co-workers recently demonstrated packing polymorphism in a supramolecular polymer achieved from an amphiphilic Pt(II) complex through the kinetically controlled pathway.¹⁵ However, kinetically controlled chiral supramolecular polymerization accompanying helical inversion, based on metal-coordinated building blocks by controlling the concentration of the unique complex structure has not been successful in water. Thus, kinetically controlled metal-coordinated chiral supramolecular polymerization that exhibits pathway complexity with unique metal-ligand building blocks remains highly challenging.

Based on these considerations, we designed a new family of chiral bipyridine-based ligands (*D*-L¹, *L*-L¹, *D*-L², and *D*-L³) that have hydrazine and/or *D*- or *L*-alanine moieties at the alkyl chain groups as a chiral segment and a binding site for intermolecular hydrogen-bonding interactions in supramolecular polymerization (Fig. 1). The Ag⁺ ion was selected as a guest ion because it forms a variety of coordination geometries with bipyridine-based

ligands, such as linear, triangular, square planar, and tetrahedral structures with different coordination numbers.^{47–49} In particular, there is a rarely observed Ag–Ag interaction in complexes obtained from Ag⁺ ions when three ligand molecules are coordinated with two Ag⁺ ions. Furthermore, the Ag–Ag interaction also acts as an additional driving force and a metal-coordinated chiral seed in chiral supramolecular polymerization.

Surprisingly, we found that the left-handed (*M*-type) chiral bipyridine complexes (*D*-L¹)₂AgNO₃ and *D*-L¹AgNO₃ are formed at less than 0.5 equiv. and at over 1.4 equiv. of AgNO₃, respectively. More interestingly, the addition of 0.5–1.3 equiv. of AgNO₃ to a *D*-L¹ solution generates the *D*-L¹AgNO₃ complex, which forms aggregate I with a spherical structure. However, this eventually forms aggregate II with a fiber structure based on the (*D*-L¹)₃Ag₂NO₃ complex, involving left-handed (*M*-type) to right-handed (*P*-type) helicity conversion. The supramolecular polymerization that forms the fiber structure occurs *via* a cooperative pathway by a nucleation–elongation mechanism that is kinetically controlled by the concentrations of Ag⁺ and NO₃[−]. Thus, we herein report kinetically controlled chiral supramolecular polymerization in water based on a unique complex building block.

Results and discussion

Stoichiometric ratios of self-assembled complexes with Ag⁺ by ESI-MS

Because bipyridine-based ligands form a variety of complexes with Ag⁺, we first investigated the complexation behavior of ligand *D*-L¹, using high-resolution electrospray ionization mass spectrometry (HR ESI-MS) (Fig. S1–S4†). We confirmed that *D*-L¹ formed mainly (*D*-L¹)₂AgNO₃, (*D*-L¹)₃Ag₂(NO₃)₂, and *D*-L¹AgNO₃ complexes with 0.4, 1.0, and 2.0 equiv. of Ag⁺, respectively (see ESI: explained in detail†).

Helicity inversion in the formation of supramolecular polymers in the presence of Ag⁺ ion

To investigate the helicity of self-assembled complexes comprising *D*-L¹ (with *D*-alanine moieties) and *L*-L¹ (with *L*-alanine moieties) with Ag⁺, time-dependent circular dichroism (CD) and UV-Vis analysis of *D*-L¹ (8 mM) was performed for three different concentrations (0.4, 1.0, and 2.0 equiv.) of AgNO₃ at 25 °C (Fig. 2 and S5†). The negative CD signals immediately increase upon addition of 0.4 and 2.0 equiv. of AgNO₃ but were not inverted to a positive CD signal (Fig. 2C: (a) and (c)). Conversely, the negative CD signal at 334 nm was inverted to a positive CD signal with 1.0 equiv. of AgNO₃ after 70 min and was enhanced drastically (Fig. 2A and C: (b)). The transparent solution prepared with *D*-L¹ (8 mM) and AgNO₃ (1.0 equiv.) also changed to a colloidal suspension during the CD measurement (as observed by ESI/TOF-MS measurement). The CD inversion was due to the formation of the (*D*-L¹)₃Ag₂(NO₃)₂ complex, which has right-handed (*P*-type) helicity. Furthermore, the weak CD signal intensity of *D*-L¹ with 1.0 equiv. of AgNO₃ is due to the presence of three complexes with different helicities in the initial state (Fig. 2C). The plot of the CD intensity and UV-Vis

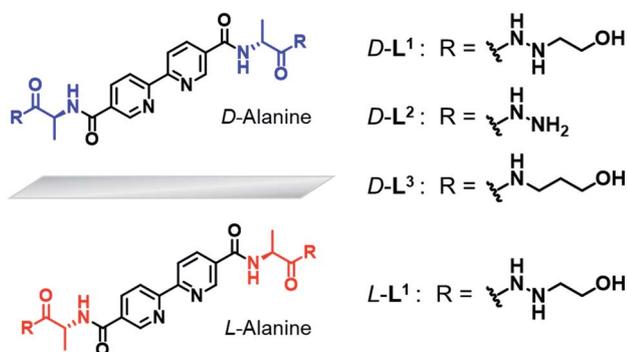


Fig. 1 Chemical structures of chiral bipyridine derivatives with hydrazinoethanol (*D*-L¹ and *L*-L¹), hydrazine (*D*-L²) and 3-aminoethanol (*D*-L³).



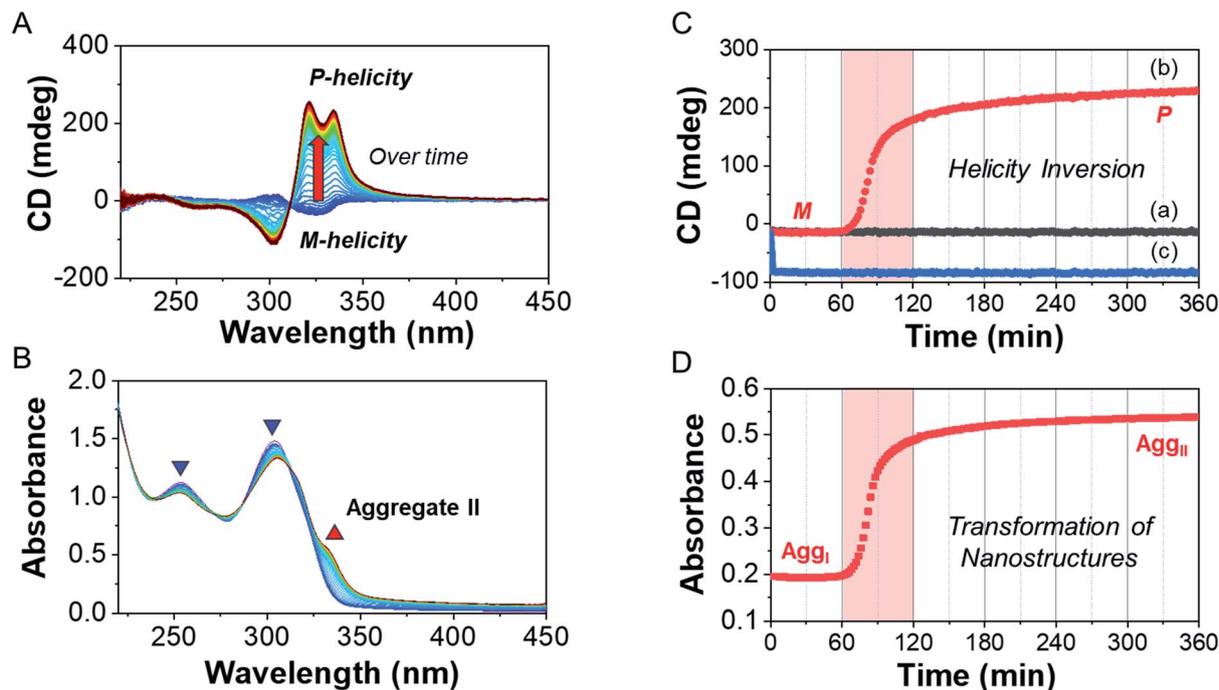


Fig. 2 Time-dependent (A) CD and (B) UV spectra changes of $D-L^1$ (8 mM) with $AgNO_3$ (1.0 equiv.) in water. (C) Plot for CD spectra changes of $D-L^1$ vs. time in the presence of $AgNO_3$ (a: 0.4, b: 1.0, and c: 2.0 equiv.) at 334 nm in water. (D) Plot for absorbance changes of $D-L^1$ vs. time in the presence of $AgNO_3$ (1.0 equiv.) at 334 nm in water.

spectral changes showed a non-linear sigmoidal transition characterized by a nucleation–elongation mechanism (Fig. 2C and D). The long lag time in the CD and UV-Vis observations implies the existence of the kinetic aggregate.

We also observed the CD spectra of the supramolecular polymeric $D-L^1$ complexes at various concentrations (0–2.0 equiv.) of Ag^+ after aging for 3 h to eliminate the time effect (Fig. 3). The CD signal of complex $D-L^1$ (8 mM) was observed at a mole ratio below 0.5 as a negative signal at 315 nm, which we ascribe to the bipyridine derivative $D-L^1$ chromophores. These negative signals in the CD analysis indicate that complex $(D-L^1)_2AgNO_3$ with four coordination was orientated to the left-handed direction (Schemes 1 and S1A[†]). Interestingly, the positive CD signal at 334 nm for supramolecular polymer $D-L^1$ in cases where the Ag^+ -to- $D-L^1$ mole ratio was greater than 0.4 was gradually enhanced (Fig. 3A and C), reaching a maximum at a Ag^+ -to- $D-L^1$ mole ratio of 1.0 (Fig. 3C). This CD inversion was due to the transformation of $(D-L^1)_2AgNO_3$ (anisotropic factor, $|g| = 1.0 \times 10^{-3}$ at 334 nm) and $D-L^1AgNO_3$ complexes into the $(D-L^1)_3Ag_2(NO_3)_2$ complex, as shown in Scheme S1B.[†] Thus, the addition of 0.5–1.3 equiv. of $AgNO_3$ generates aggregate I composed of the $D-L^1AgNO_3$ complex ($|g| = 8.0 \times 10^{-3}$ at 334 nm) with left-handed helicity as the kinetic product, and then finally aggregate II is formed based on the $(D-L^1)_3Ag_2(NO_3)_2$ complex ($|g| = 1.45 \times 10^{-2}$ at 334 nm), which has right-handed (P-type) helicity (Schemes 1 and S1B[†]) as the thermodynamic product.

Furthermore, the transparent solution became a colloidal suspension at 0.5–1.3 equiv. of Ag^+ . After separation, we observed the CD spectra of both the colloidal particles and the filtered

solution. The colloidal particles dispersed in water exhibited an intense positive CD signal (Fig. S6a[†]), which was almost the same as that for the solution before filtration. However, the filtrate exhibited an extremely weak negative CD signal (Fig. S6b[†]). These findings indicate that complexes $(D-L^1)_2AgNO_3$ and $D-L^1AgNO_3$ present before CD inversion change to $(D-L^1)_3Ag_2(NO_3)_2$ as the colloidal aggregate II upon achieving chemical equilibrium, as shown in Schemes 1 and S1B.[†] These results provide clear evidence that the $(D-L^1)_3Ag_2(NO_3)_2$ complex plays an essential role as a nucleus (seed) in the formation of aggregate II. In addition, it is noteworthy that $AgNO_3$ acted as a key component in forming aggregates I and II. More importantly, the positive CD signal for the supramolecular polymer obtained with 1.0 equiv. of Ag^+ did not change upon further addition of $AgNO_3$ (Fig. S7[†]), indicating that aggregate II is the thermodynamic product. When the Ag^+ -to- $D-L^1$ mole ratio increases beyond 1.0, the intensity of the CD signal for the supramolecular polymer gradually decreases again (Fig. 3A and C). This decrease in the positive CD signal was due to the coexistence of aggregate II and the monomeric $D-L^1AgNO_3$ complex species. At 1.3 equiv. of $AgNO_3$, time-dependent CD spectra of $D-L^1$ (8 mM) did not change after 180 min, indicating that the kinetic product was not existed (Fig. S8[†]). However, a small amount of monomeric 1 : 1 complex species, which does not form the spherical structure, exists with the thermodynamic product. Finally, the negative CD signal for $D-L^1$ at over 1.4 equiv. of $AgNO_3$ is attributed to the formation of the $D-L^1AgNO_3$ complex as the main product (Scheme S1C[†]), which indicates that the $D-L^1AgNO_3$ complex has M-helicity (left-handed helicity). In addition, the negative CD signal obtained at 1.5 equiv. of Ag^+ was



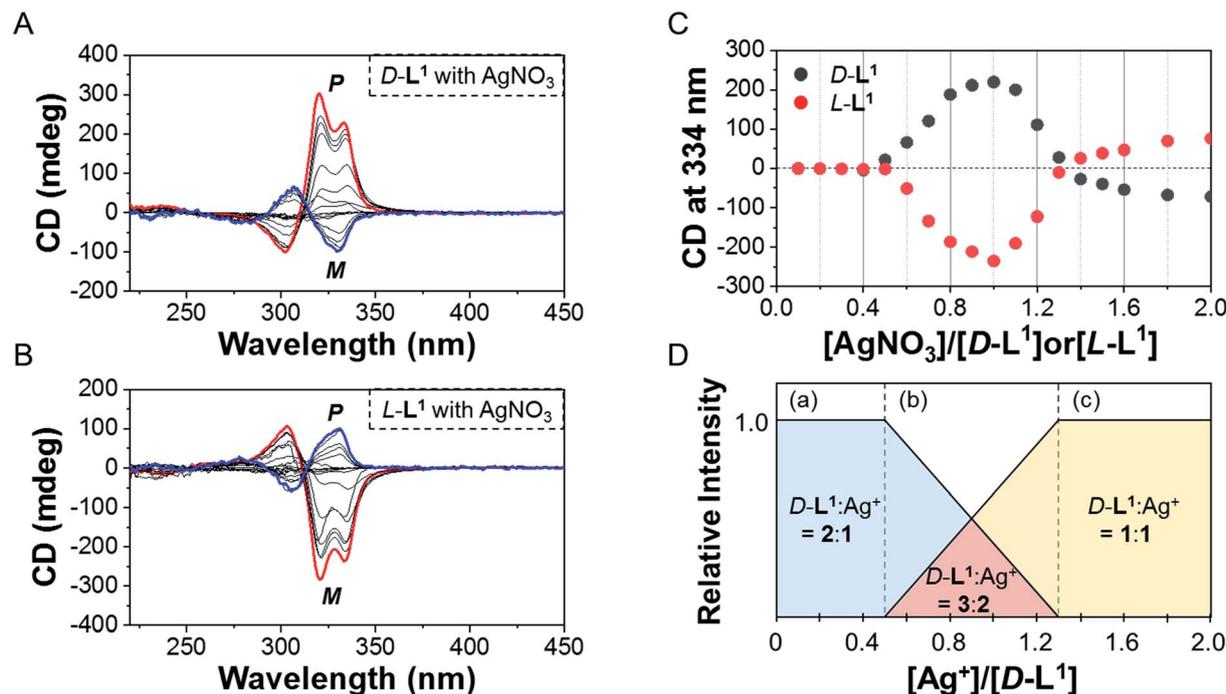
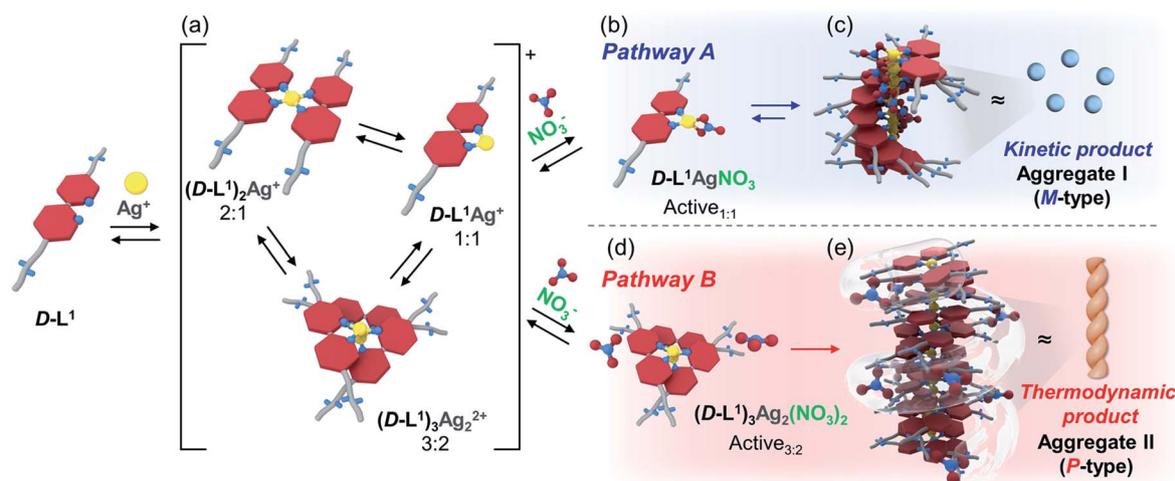


Fig. 3 CD spectra of (A) $D-L^1$ or (B) $L-L^1$ upon addition of $AgNO_3$ (0–2.0 equiv.) after aging for 3 h in water. (C) Plot for CD spectral changes of $D-L^1$ and $L-L^1$ vs. equivalent of $AgNO_3$ at 334 nm. (D) Complex structures existed in various concentration of $AgNO_3$; (a) $(D-L^1)_2AgNO_3$, (b) $(D-L^1)_3Ag_2(NO_3)_2$, and $D-L^1AgNO_3$, and (c) $D-L^1AgNO_3$.

inverted to the positive signal when 0.5 equiv. of free $D-L^1$ was added to a mixture of $D-L^1$ and Ag^+ (1.5 equiv.) (Fig. S9[†]). This CD inversion was due to the $(D-L^1)_3Ag_2(NO_3)_2$ complex, which was generated from the $D-L^1AgNO_3$ complex. We conclude that the aggregate II as the thermodynamic product was only produced for 0.5–1.3 equiv. of Ag^+ (Fig. 3D and Scheme 1).

In contrast, the CD spectra of complex $L-L^1$ (8 mM) upon addition of $AgNO_3$ (0–0.4 equiv.) exhibited a positive signal at

315 nm (Fig. 3B and C), which is exactly opposite to the behavior of complex $D-L^1$ with Ag^+ . This positive CD signal indicates that the bipyridine moiety of $L-L^1$ is orientated in the right-handed (P-type) direction. The positive CD signal of $L-L^1$ was inverted into an intensely negative CD signal when 0.5–1.3 equiv. of Ag^+ was added to $L-L^1$. This chiral inversion is also due to the transformation from $(L-L^1)_2AgNO_3$ and $L-L^1AgNO_3$ complexes into complex $(L-L^1)_3Ag_2(NO_3)_2$, which led to the formation of the



Scheme 1 Proposed chiral supramolecular polymerization for $D-L^1$ with $AgNO_3$ (0.5–1.3 equiv.) showing the assembly by nucleation–elongation mechanism; (a) complex formation ($D-L^1 : Ag^+ = 2 : 1, 1 : 1$ and $3 : 2$), (b) $1 : 1$ complex formation, (c) the formation of aggregate I based on $D-L^1AgNO_3$ complex with M-helicity, (d) $3 : 2$ complex formation, and (e) formation of the aggregate II based on $(D-L^1)_3Ag_2(NO_3)_2$ complex with P-helicity.



left-handed aggregate II. However, the positive CD signal of $D-L^1$ was also observed when 1.4–2.0 equiv. of $AgNO_3$ was added to $D-L^1$, which was attributed to the formation of aggregate I (nanoparticles) based on the $D-L^1AgNO_3$ complex by path A in Scheme 1.

In order to investigate their linear dichroism (LD) effects, the LD spectra of $D-L^1$ (8 mM) and $L-L^1$ (8 mM) with $AgNO_3$ (1.0 and 1.4 equiv.) were observed under the same conditions as those used for CD observations (Fig. S10†). Extremely weak LD signals were obtained for $D-L^1$ and $L-L^1$ in the presence of $AgNO_3$ (1.0 and 1.4 equiv.). These results imply that the LD effect is only marginally affected by the CD response of supramolecular polymers.

To determine how anions effect the production of supramolecular polymers, we further observed the handedness of the supramolecular polymer upon addition of $AgBF_4$, $AgClO_4$, $AgPF_6$, or $AgOTf$ instead of NO_3^- . As shown in Fig. S11,† the negative CD signals for complex $D-L^1$ formed with 0–2.0 equiv. of $AgBF_4$, $AgClO_4$, $AgPF_6$, or $AgOTf$ showed a slightly enhanced intensity, but they did not invert to the positive signal even though 3 : 2 complex ($D-L^1 : Ag^+$) formation was confirmed by HR ESI-MS (Fig. S12–S15†). These findings imply that the NO_3^- anion plays a critical role in the formation of aggregate II, as shown in Schemes 1 and S1B.†

Role of NO_3^- anion in the supramolecular polymerization

To demonstrate the essential role of NO_3^- in the formation of the supramolecular polymer, we also observed the CD spectral changes for $D-L^1$ in the presence of 1.0 equiv. of $AgOTf$, $AgClO_4$, $AgBF_4$, or $AgPF_6$ upon the addition of 2.0 equiv. of $NaNO_3$ (Fig. S16†). The weak negative CD signal became a positive CD signal when $NaNO_3$ (0.4 equiv.) was added to a mixed $D-L^1$ and $AgOTf$ (1 : 1 equiv.) solution. The continuous addition of $NaNO_3$ gradually enhanced the positive CD signals, and the signal reached a maximum at 2.0 equiv. of $NaNO_3$ (Fig. 4), which was due to the generation of the aggregate II based on $(D-L^1)_3Ag_2(NO_3)_2$. The peak for the $[(D-L^1)_3Ag_2(NO_3)_2]^+$ fragmented from the $(D-L^1)_3Ag_2(NO_3)_2$ complex was clearly observed in HR ESI/TOF-MS measurement (Fig. S17†). This led not only to the helical inversion but also to the formation of supramolecular polymers. These results indicate that NO_3^- plays a key role in not only the formation of the supramolecular polymer *via* the bridge effect but also helical inversion by path B, as shown in Scheme 1.

As a reference experiment, HR ESI-MS and CD spectra of $D-L^2$, and $D-L^3$ without ethyl alcohol or the hydrazine moiety at both ends were observed upon addition of $AgNO_3$. The formation of the $(D-L^2)_2AgNO_3$, $(D-L^2)_3Ag_2(NO_3)_2$, or $D-L^2AgNO_3$ complexes obtained at different concentrations of Ag^+ (0.4, 1.0, and 2.0 equiv. of Ag^+) was confirmed *via* HR ESI-MS (Fig. S18–S20†). As expected, the $D-L^2$ complex structures with different concentrations of Ag^+ showed a similar tendency as that $D-L^1$. The CD inversion of $D-L^2$ (8 mM) with 0.8–1.1 equiv. of $AgNO_3$ occurred due to the formation of aggregate II based on the $(D-L^2)_3Ag_2(NO_3)_2$ complex (Fig. S21†). However, the CD intensity for the supramolecular polymer $D-L^2$ was *ca.* 4.3-fold lower than

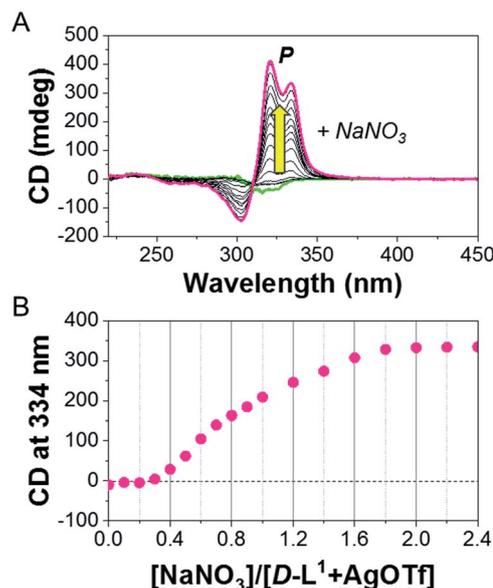


Fig. 4 (A) CD spectra of $D-L^1$ upon addition of $NaNO_3$ in the presence of $AgOTf$ (1.0 equiv.) in water. (B) Plot for CD intensity of $D-L^1 + AgOTf$ against added $NaNO_3$ concentration.

that of supramolecular polymer $D-L^1$ under a same condition, which was attributed to the weak complex formation between $D-L^2$ and Ag^+ . However, the negative CD signal did not invert to a positive CD signal when 1.0 equiv. of $AgNO_3$ was added to $D-L^3$ (8 mM), which does not have the hydrazine moiety (Fig. S22†), because of no formation of 3 : 2 complex ($D-L^3 : Ag^+$) as shown in Fig. S23.† These findings indicate that the amine in the hydrazine group plays a critical role in 3 : 2 complex formation ($D-L^3 : Ag^+$).

Interestingly, upon addition of $NaCl$ (0.9 equiv.) to aggregate I, a positive CD signal resulted, and the maximum positive CD signal was reached with 1.3 equiv. of $NaCl$ (Fig. S24†). The $(D-L^1)_2AgNO_3$ complex and free $D-L^1$ were generated by decomplexation of $D-L^1AgNO_3$ in the presence of $NaCl$, and this eventually produced $(D-L^1)_3Ag_2(NO_3)_2$, which enables the formation of aggregate II, as shown in Scheme S1C.† The largest positive CD signal induced *via* 1.3 equiv. of $NaCl$ was ascribed to the formation of the largest amount of the $(D-L^1)_3Ag_2(NO_3)_2$ complex. Thus, this CD inversion was due to transformation from the $D-L^1Ag$ to the $(D-L^1)_3Ag_2(NO_3)_2$ complex, which led to further formation of aggregate II.

NMR and IR studies of aggregates I and II

To determine the exact binding site of $D-L^1$ for NO_3^- in the formation of the supramolecular polymer, we obtained the 1H NMR spectrum of $D-L^1$ in the presence and the absence of $AgNO_3$ in D_2O and H_2O (1 : 1 v/v) (Fig. S25–S27†). These results suggest that the supramolecular polymer constructed from π – π stacking interactions, and the intermolecular hydrogen-bonding interactions between $D-L^1$ and $D-L^1$ were weakened due to the linkage with NO_3^- between the hydrazine moiety of $D-L^1$ and $D-L^1$ (see ESI: explained in detail†). The FT-IR results



provide evidence for the intermolecular hydrogen bonds between the hydrazine NH and the NO_3^- in the chiral supramolecular polymers (Fig. S28 and S29, see ESI: explained in detail†). In addition, a sharp powder X-ray diffraction (XRD) peak for aggregate II was observed at $2\theta = 32$ (Fig. S30†), which corresponds to an Ag–Ag interaction with a distance of 2.8 Å.⁴⁸ This suggests that NO_3^- did not coordinate with Ag^+ . The Ag–Ag interaction observed likely plays an important role in the chiral supramolecular polymerization. Thus, based on the FT-IR, powder XRD, and ^1H NMR results, we determined that NO_3^- acted as a bridge to produce the helical supramolecular polymeric structure. This is in agreement with results previously reported by the Miyake group.⁵⁰

Morphology observation of aggregates I and II by AFM

To confirm the morphology changes of the supramolecular polymers formed at different concentrations of AgNO_3 , we performed time-dependent atomic force microscopy (AFM) experiments (Fig. 5). For this purpose, freshly prepared solutions of D-L^1 with and without AgNO_3 (0.4, 1.0, 1.3, and 2.0 equiv.) were quickly dropped onto silicon wafers at 298 K after a constant aging time. This fixed the sample on the silicon wafer surface because it acts like a mode and prevents structural changes in the material during drying.

In the absence of AgNO_3 , the AFM image of pristine D-L^1 showed fiber structures with widths of *ca.* 30 nm and lengths of several-hundred micrometers (Fig. S31†), indicating nanostructural formation of D-L^1 by π - π stacking. This fiber structure did not change with time, which indicative that the fiber structure was the thermodynamic product. In contrast, no distinct morphology for the D-L^1 solution with 0.4 equiv. of AgNO_3 was observed regardless of aging time, indicating that the $(\text{D-L}^1)_2\text{AgNO}_3$ complex did not elongate to form aggregates I

or II. However, time-dependent AFM images of the D-L^1 solution containing AgNO_3 (2.0 equiv.), which formed complex $\text{D-L}^1\text{AgNO}_3$, were observed at different aging times. After aging for 10 min, the AFM image showed spherical structures with a diameter of *ca.* 50 nm (Fig. S32†). The spherical structures with dimensions close to 5 nm, as previously reported by Meijer's group,⁵¹ were similar to that of the micelle type containing a chiral molecular arrangement. This sphere would consist of the bilayer structure, because the full extended molecule length of D-L^1 is *ca.* 3 nm.

No significant morphological changes were observed after aging for two weeks, indicating that the spherical structure was the thermodynamic product. The morphology changes were further supported by dynamic light scattering (DLS) study (Fig. S33b†). Note that a growth pattern in the scattering intensity was not observed, which means that the morphology did not change. This finding suggests that the nanoparticles formed in the presence of 2.0 equiv. of AgNO_3 were a thermodynamic aggregate, as shown in Scheme S1C.†

Interestingly, the D-L^1 solution containing AgNO_3 (1.0 equiv.) had only small spherical structures with *ca.* 50 nm diameters at an aging time of 10 min (Fig. 5A), which originated from aggregate I (Scheme 1). After 90 min, the AFM image showed one-dimensional nanorods with 200 to 300 nm length and nanoparticles with *ca.* 50 nm diameter (Fig. 5B), which were due to aggregate II and the aggregate I, respectively. Upon allowing the solution to age for 120 min at 298 K, the nanorods grew into fibrous structures (Fig. 5C and D). This was clear evidence that the $(\text{D-L}^1)_3\text{Ag}_2(\text{NO}_3)_2$ complex grew into one-dimensional fibers with intermolecular interactions as shown in Scheme 1. These fibers had an average thickness of ~ 3 nm by AFM observation (Fig. S34†), which is consistent with the fully extended molecular length of ligand D-L^1 . Based on density functional theory (DFT) calculations, one fiber should consist of *ca.* 20 units of $(\text{D-L}^1)_3\text{Ag}_2(\text{NO}_3)_2$ complex in the width direction (Fig. S34†). As shown by the DLS study (Fig. S33a†), the growth pattern in scattering intensity was similar to the time-dependent change in the CD signal at 334 nm. This clearly indicates that the spherical structure as the kinetic product was transformed into the fiber structure as the thermodynamic product. Furthermore, only fibrous structures were observed in the presence of 1.3 equiv. of AgNO_3 after aging for 180 min (Fig. S35†), indicating that the thermodynamic product but not the kinetic product existed.

To confirm the anion effect in the formation of the supramolecular fibers, we obtained AFM images of the D-L^1 solution containing AgOTf (1.0 equiv.). No significant morphological changes were observed, regardless of aging time. However, a narrow size distribution for the nanoparticles was observed during the lag time when NaNO_3 (1.0 equiv.) was added to a D-L^1 solution containing AgOTf (1.0 equiv.) (Fig. S36A†). These nanoparticles were transformed into fibrous structures several micrometers in length upon 90 min of aging (Fig. S36B and C†). This is indirect evidence that NO_3^- acted as a bridging ligand between the hydrazine groups of complex D-L^1 to generate the chiral supramolecular polymer.

For D-L^2 with AgNO_3 (1.0 equiv.), the AFM image showed nanoparticles with *ca.* 50 nm diameter during the initial stage

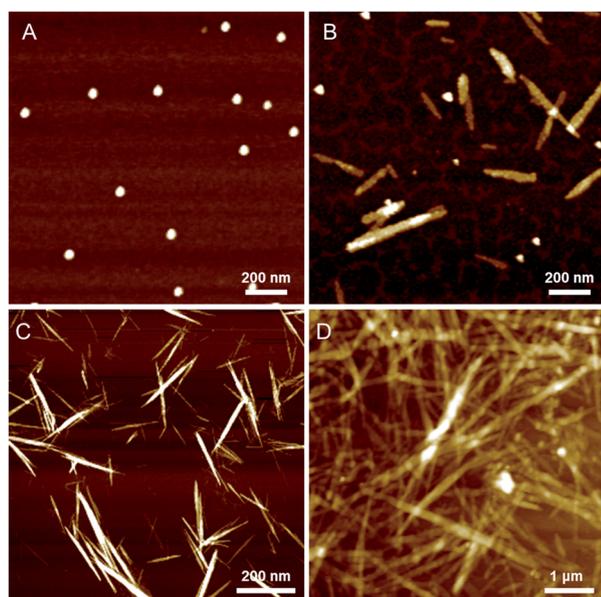


Fig. 5 AFM images of complex D-L^1 with 1.0 equiv. of AgNO_3 in water after aging of (A) 10 min, (B) 90 min, (C) 120 min and (D) 240 min.



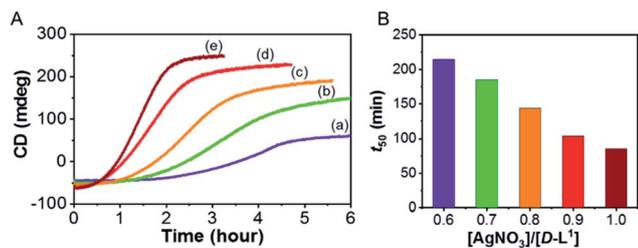


Fig. 8 (A) Time course evolution of the aggregate I of $D-L^1$ into the kinetically controlled aggregate II with different concentrations of $AgNO_3$; (a) 0.6, (b) 0.7, (c) 0.8, (d) 0.9 and (e) 1.0 equiv. at 334 nm in water. (B) Representation for t_{50} values vs. $AgNO_3$ concentration.

(Fig. 8). The kinetic profiles of supramolecular polymer $D-L^1$ (8 mM) that formed with different concentrations of $AgNO_3$ (0.6–1.0 equiv.) in aqueous solution at 25 °C showed non-sigmoidal curves with different lag times (Fig. 8). The t_{50} value, at which the transformation is 50% complete, became short as the $AgNO_3$ concentration increased (Fig. 8B). The influence of $AgNO_3$ concentration on the kinetics confirmed the initial formation of aggregate I consisting of $D-L^1AgNO_3$, which developed into aggregate II consisting of $(D-L^1)_3Ag_2(NO_3)_2$. The largest amount of the $(D-L^1)_3Ag_2(NO_3)_2$ complex produced during the initial stage in the presence of 1.0 equiv. of $AgNO_3$, kinetically accelerated the transformation of aggregate I into the aggregate II. Thus, the concentration of the $(D-L^1)_3Ag_2(NO_3)_2$ complex during the initial state played a critical role in producing aggregate II, which exhibited a helical fiber structure.

Furthermore, the kinetic profiles of the supramolecular polymer were also observed upon addition of $NaNO_3$ (1.0, 1.4, 1.7, and 2.0 equiv.) in the presence of $AgOTf$ (1.0 equiv.) (Fig. S45†). The highest concentration of $NaNO_3$ showed the largest positive CD intensity (Fig. S45d†) owing to formation of the largest amount of the $(D-L^1)_3Ag_2(NO_3)_2$ complex under these conditions. These results imply that the kinetics of aggregate II strongly influenced by NO_3^- anions. Furthermore, the lag time decreased with increasing $NaNO_3$ concentration, indicating that the NO_3^- anion plays an essential role in the helical inversion from aggregate I to aggregate II. To our knowledge, this is the first example of a system in which the NO_3^- anion acts as an accelerator and linker in the transformation of aggregates. In addition, the kinetic profile of the supramolecular polymer $D-L^2$ that does not have an ethyl alcohol moiety showed the same tendency as that of compound $D-L^1$ (Fig. S46†). However, the supramolecular polymerization of $D-L^2$ was observed in a narrower Ag^+ concentration range compared to that for $D-L^1$ because $D-L^2$ forms the $(D-L^2)_2AgNO_3$ complex as a main product, as shown by HR ESI-MS measurements.

Conclusions

We have demonstrated that supramolecular polymerization involving a distinct metal–ligand complex was controlled kinetically *via* the concentration of Ag^+ and anions in water.

This also enabled the controlled, reversible transformation between M- and P-type helices, with a change in complex structure as a function of Ag^+ concentration. A change in the complex structure also accompanied the helical inversion, where the M-type helices of $(D-L^1)_2AgNO_3$ and $D-L^1AgNO_3$ were inverted to the P-type helix of $(D-L^1)_3Ag_2(NO_3)_2$. In the kinetic study, the $(D-L^1)_3Ag_2(NO_3)_2$ complex was found to play an essential role in chiral inversion. The distinct $(D-L^1)_3Ag_2(NO_3)_2$ complex produced the chiral supramolecular polymer (aggregate II) *via* a nucleation–elongation mechanism with a cooperative pathway in the range of 0.5–1.3 equiv. of $AgNO_3$, and this process showed an unusually long lag time. In addition, $D-L^1$, and $L-L^1$ in the presence of 0.5–1.3 equiv. of $AgNO_3$ formed intermediate aggregate I (nanoparticles) as the kinetic product that evolved to afford a helical supramolecular polymer (aggregate II). This transformation can be accelerated according to the Ag^+ and NO_3^- concentration. Also, $AgNO_3$ acted as a key component in forming aggregates I and II. In particular, NO_3^- bridged between hydrazine–NH groups of the complex *via* intermolecular hydrogen bonds in the supramolecular polymer. In contrast, aggregate I formed at over 1.4 equiv. of $AgNO_3$ was the thermodynamic product.

This system is an extremely rare example of chiral supramolecular polymerization based on metal–organic building blocks *via* control of coordination number. We believe that this concept will greatly contribute to the development of metal-coordinated supramolecular systems with optimized functional properties and may provide new insights into the understanding of the chiral assembly process in complex biological systems.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 N. Ousaka, Y. Takeyama, H. Iida and E. Yashima, *Nat. Chem.*, 2011, **3**, 856–861.
- 2 J. Rebek, Jr, *Angew. Chem., Int. Ed.*, 2005, **44**, 2068–2078.
- 3 P. J. Stang, B. Olenyuk, D. C. Muddiman, D. S. Wunschel and R. D. Smith, *Organometallics*, 1997, **16**, 3094–3096.
- 4 E. Yashima, N. Ousaka, D. Taura, K. Shimomura, T. Ikai and K. Maeda, *Chem. Rev.*, 2016, **116**, 13752–13990.



- 5 A. R. A. Palmans and E. W. Meijer, *Angew. Chem., Int. Ed.*, 2007, **46**, 8948–8968.
- 6 M. Albrecht, *Chem. Rev.*, 2001, **101**, 3457–3498.
- 7 Q. Wan, X.-S. Xiao, W.-P. To, W. Lu, Y. Chen, K.-H. Low and C.-M. Che, *Angew. Chem., Int. Ed.*, 2018, **57**, 17189–17193.
- 8 Q. Wan, W.-P. To, C. Yang and C.-M. Che, *Angew. Chem., Int. Ed.*, 2018, **57**, 3089–3093.
- 9 J. L.-L. Tsai, T. Zou, J. Liu, T. Chen, A. O.-Y. Chan, C. Yang, C.-N. Lok and C.-M. Che, *Chem. Sci.*, 2015, **6**, 3823–3830.
- 10 A. Mishra, D. B. Korlepara, M. Kumar, A. Jain, N. Jonnalagadda, K. K. Bejagam, S. Balasubramanian and S. J. George, *Nat. Commun.*, 2018, **9**, 1295.
- 11 K. K. Kartha, N. K. Allampally, A. T. Politi, D. D. Prabhu, H. Ouchi, R. Q. Albuquerque, S. Yagai and G. Fernández, *Chem. Sci.*, 2019, **10**, 752–760.
- 12 F. Mamiya, N. Ousaka and E. Yashima, *Angew. Chem., Int. Ed.*, 2015, **54**, 14442–14446.
- 13 N. Ousaka, K. Shimizu, Y. Suzuki, T. Iwata, M. Itakura, D. Taura, H. Iida, Y. Furusho, T. Mori and E. Yashima, *J. Am. Chem. Soc.*, 2018, **140**, 17027–17039.
- 14 M. E. Robinson, A. Nazemi, D. J. Lunn, D. W. Hayward, C. E. Boott, M.-S. Hsiao, R. L. Harniman, S. A. Davis, G. R. Whittell, R. M. Richardson, L. De Cola and I. Manners, *ACS Nano*, 2017, **11**, 9162–9175.
- 15 A. Langenstroer, K. K. Kartha, Y. Dorca, J. Droste, V. Stepanenko, R. Q. Albuquerque, M. R. Hansen, L. Sánchez and G. Fernández, *J. Am. Chem. Soc.*, 2019, **141**, 5192–5200.
- 16 S. Ogi, C. Grzeszkiewicz and F. Würthner, *Chem. Sci.*, 2018, **9**, 2768–2773.
- 17 L. Herkert, J. Droste, K. K. Kartha, P. A. Korevaar, T. F. A. de Greef, M. R. Hansen and G. Fernández, *Angew. Chem., Int. Ed.*, 2019, **58**, 11344–11349.
- 18 A. Langenstroer, Y. Dorca, K. K. Kartha, M. J. Mayoral, V. Stepanenko, G. Fernández and L. Sánchez, *Macromol. Rapid Commun.*, 2018, **39**, 1800191.
- 19 N. A. M. S. Caturello, Z. Csók, G. Fernández and R. Q. Albuquerque, *Chem.–Eur. J.*, 2016, **22**, 17681–17689.
- 20 K. Y. Kim, J. Kim, C. J. Moon, J. Liu, S. S. Lee, M. Y. Choi, C. Feng and J. H. Jung, *Angew. Chem., Int. Ed.*, 2019, **58**, 11709–11714.
- 21 H. Park, K. Y. Kim, S. H. Jung, Y. Choi, H. Sato and J. H. Jung, *Chem. Mater.*, 2018, **30**, 2074–2083.
- 22 K. Y. Kim, J. Kim, H. Park, Y. Choi, K.-Y. Kwon and J. H. Jung, *Bull. Korean Chem. Soc.*, 2018, **39**, 988–994.
- 23 V. C.-H. Wong, C. Po, S. Y.-L. Leung, A. K.-W. Chan, S. Yang, B. Zhu, X. Cui and V. W.-W. Yam, *J. Am. Chem. Soc.*, 2018, **140**, 657–666.
- 24 S. Y.-L. Leung, K. M.-C. Wong and V. W.-W. Yam, *Proc. Natl. Acad. Sci. U. S. A.*, 2016, **113**, 2845–2850.
- 25 M. H.-Y. Chan, S. Y.-L. Leung and V. W.-W. Yam, *J. Am. Chem. Soc.*, 2018, **140**, 7637–7646.
- 26 Y. Sakata, S. Chiba, M. Miyashita, T. Nabeshima and S. Akine, *Chem.–Eur. J.*, 2019, **25**, 2962–2966.
- 27 T. Nakamura, H. Kimura, T. Okuhara, M. Yamamura and T. Nabeshima, *J. Am. Chem. Soc.*, 2016, **138**, 794–797.
- 28 R. Matsuoka and T. Nabeshima, *Front. Chem.*, 2018, **6**, 349.
- 29 B. Kemper, M. von Gröning, V. Lewe, D. Spitzer, T. Otremba, N. Stergiou, D. Schollmeyer, E. Schmitt, B. J. Ravoo and P. Besenius, *Chem.–Eur. J.*, 2017, **23**, 6048–6055.
- 30 B. Kemper, Y. R. Hristova, S. Tacke, L. Stegemann, L. S. van Bezouwen, M. C. A. Stuart, J. Klingauf, C. A. Strassert and P. Besenius, *Chem. Commun.*, 2015, **51**, 5253–5256.
- 31 A. Winter and U. S. Schubert, *Chem. Soc. Rev.*, 2016, **45**, 5311–5357.
- 32 L. Yu, Z. Wang, J. Wu, S. Tu and K. Ding, *Angew. Chem., Int. Ed.*, 2010, **49**, 3627–3630.
- 33 G. R. Whittell, M. D. Hager, U. S. Schubert and I. Manners, *Nat. Mater.*, 2011, **10**, 176–188.
- 34 M. Higuchi, *J. Mater. Chem. C*, 2014, **2**, 9331–9341.
- 35 D. Taura, S. Hioki, J. Tanabe, N. Ousaka and E. Yashima, *ACS Catal.*, 2016, **6**, 4685–4689.
- 36 L.-J. Chen, H.-B. Yang and M. Shionoya, *Chem. Soc. Rev.*, 2017, **46**, 2555–2576.
- 37 B. Kemper, L. Zengerling, D. Spitzer, R. Otter, T. Bauer and P. Besenius, *J. Am. Chem. Soc.*, 2018, **140**, 534–537.
- 38 A. Sorrenti, J. Leira-Iglesias, A. J. Markvoort, T. F. A. de Greef and T. M. Hermans, *Chem. Soc. Rev.*, 2017, **46**, 5476–5490.
- 39 T. Fukui, S. Kawai, S. Fujinuma, Y. Matsushita, T. Yasuda, T. Sakurai, S. Seki, M. Takeuchi and K. Sugiyasu, *Nat. Chem.*, 2017, **9**, 493–499.
- 40 W. Wagner, M. Wehner, V. Stepanenko, S. Ogi and F. Würthner, *Angew. Chem., Int. Ed.*, 2017, **56**, 16008–16012.
- 41 S. Ogi, V. Stepanenko, J. Thein and F. Würthner, *J. Am. Chem. Soc.*, 2016, **138**, 670–678.
- 42 M. F. J. Mabesoone, A. J. Markvoort, M. Banno, T. Yamaguchi, F. Helmich, Y. Naito, E. Yashima, A. R. A. Palmans and E. W. Meijer, *J. Am. Chem. Soc.*, 2018, **140**, 7810–7819.
- 43 P. A. Korevaar, S. J. George, A. J. Markvoort, M. M. J. Smulders, P. A. J. Hilbers, A. P. H. J. Schenning, T. F. A. De Greef and E. W. Meijer, *Nature*, 2012, **481**, 492–496.
- 44 S. Ogi, K. Sugiyasu, S. Manna, S. Samitsu and M. Takeuchi, *Nat. Chem.*, 2014, **6**, 188–195.
- 45 J. S. Valera, R. Gomez and L. Sánchez, *Small*, 2018, **14**, 1702437.
- 46 M. A. J. Gillissen, M. M. E. Koenigs, J. J. H. Spiering, J. A. J. M. Vekemans, A. R. A. Palmans, I. K. Voets and E. W. Meijer, *J. Am. Chem. Soc.*, 2014, **136**, 336–343.
- 47 T. Nakamura, H. Ube and M. Shionoya, *Angew. Chem., Int. Ed.*, 2013, **52**, 12096–12100.
- 48 A. Bellusci, M. Ghedini, L. Giorgini, F. Gozzo, E. I. Szerb, A. Crispini and D. Pucci, *Dalton Trans.*, 2009, 7381–7389.
- 49 A. Bellusci, A. Crispini, D. Pucci, E. I. Szerb and M. Ghedini, *Cryst. Growth Des.*, 2008, **8**, 3114–3122.
- 50 H. Miyake, K. Yoshida, H. Sugimoto and H. Tsukube, *J. Am. Chem. Soc.*, 2004, **126**, 6524–6525.
- 51 P. Besenius, G. Portale, P. H. H. Bomans, H. M. Janssen, A. R. A. Palmans and E. W. Meijer, *Proc. Natl. Acad. Sci. U. S. A.*, 2010, **107**, 17888–17893.
- 52 J. Tomasi and M. Persico, *Chem. Rev.*, 1994, **94**, 2027–2094.
- 53 M. Shen, Y. Xie, H. F. Schaefer, III and C. A. Deakyne, *J. Chem. Phys.*, 1990, **93**, 3379–3388.



- 54 T. Vijayakumar, I. H. Joe, C. P. R. Nair and V. S. Jayakumar, *J. Mol. Struct.*, 2008, **877**, 20–35.
- 55 M. H.-Y. Chan, M. Ng, S. Y.-L. Leung, W. H. Lam and V. W.-W. Yam, *J. Am. Chem. Soc.*, 2017, **139**, 8639–8645.
- 56 T. F. A. De Greef, M. M. J. Smulders, M. Wolffs, A. P. H. J. Schenning, R. P. Sijbesma and E. W. Meijer, *Chem. Rev.*, 2009, **109**, 5687–5754.
- 57 M. Wehner, M. I. S. Röhr, M. Bühler, V. Stepanenko, W. Wagner and F. Würthner, *J. Am. Chem. Soc.*, 2019, **141**, 6092–6107.
- 58 S. Dhiman, A. Sarkar and S. J. George, *RSC Adv.*, 2018, **8**, 18913–18925.
- 59 M. M. J. Smulders, M. M. L. Nieuwenhuizen, T. F. A. de Greef, P. van der Schoot, A. P. H. J. Schenning and E. W. Meijer, *Chem.–Eur. J.*, 2010, **16**, 362–367.

