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Synthesis and evaluation of eco-friendly carboxymethyl cellulose/polyvinyl alcohol/CuO bionanocomposites and their use in coating processed cheese

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In the present study, we formulated and characterized CMC/PVA/CuO bionanocomposites to evaluate their use in coating processed cheese. Copper oxide nanoparticles (CuO-NPs) were prepared and added to a mixed solution of carboxymethyl cellulose (CMC)/polyvinyl alcohol (PVA) using compositions of 0.3, 0.6 and 0.9% (w/v). The CMC/PVA/CuO bionanocomposites were prepared by a solution casting method and used for coating processed cheese. The fabricated bionanocomposite films and CuO-NPs were characterized by TEM, SEM, EDEX, XRD, DLS, and FT-IR analysis. Inclusion of CuO-NPs decreased the gas transmission rate (GTR) and water vapor transmission rate (WVTR) of the prepared film. Also, the bionanocomposite suspensions exhibited high but variable inhibitory effects against several pathogenic bacteria and fungi. The impact of coating of processed cheese surfaces with the prepared bionanocomposite films on microbiological, physicochemical, textural and sensory properties of the processed cheese were assessed during 6 months of cold storage. Coating cheese with film containing CuO-NPs eliminated mould growth on the cheese surface and decreased significantly ($P < 0.05$) the total bacterial count of the cheese. Furthermore, coating of cheese decreased the moisture losses and retarded the increase in the cheese hardness during storage. The highest acceptability at the end of the storage period was given for processed cheese coated with the bionanocomposite containing 0.9% CuO-NPs. Thus, the obtained CMC/PVA/CuO bionanocomposite films could be a promising candidate for cheese packaging applications.

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1. Introduction

In the past years, nanotechnology has been increasingly used to meet the consumer's demand for quality foods and in developing antimicrobial agents to prolong the food lifetime during storage and distribution. Food producers continuously evaluate the benefits and disadvantages of packaging materials in order to choose suitable packaging materials for their food products.^{1,2} The application of nanomaterials has a wide range in various fields, such as physical, chemical, electronic, biomedical and materials sciences. Carboxymethyl cellulose (CMC) is an anionic decomposable and biocompatible polymer derivative from natural cellulose. CMC is usually blended with other stabilizers and gums to retain moisture, and to improve the structural consistency of bakery products due to its high water-absorbing capacity.³ Polyvinyl alcohol (PVA) is a biocompatible

polymer rich in hydroxyl groups, non-toxic, semi-crystalline and soluble in water.⁴ It is broadly used in food packaging, construction sectors and household, due to its properties, such as biocompatibility, mechanical performance and solvent resistance.⁵ The major disadvantages of using PVA in food packaging are its low biodegradation rate and its high moisture absorption. So, it is commonly mixed with biopolymers and/or bio-based materials to improve its performance and environmental properties. Also the limitations in both mechanical properties and water permeability of PVA films can be overcome *via* the addition of different nanomaterials or blending with other biopolymers.⁶

Copper oxide (CuO) is widely used for various applications in a number of areas such as catalyst, ceramic, thermoelectric sensors, glass, superconducting materials and antimicrobial agents.⁷ Fabrication of CuO-NPs is less expensive when compared to gold and silver nanoparticles. In addition, CuO-NPs possess excellent potent antimicrobial action because of its characteristic crystal structure. The physical and chemical properties of NPs, such as size, agglomeration state in liquids and surface charge, play a significant role in the final

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interactions of NPs with target cells. CuO-NPs were reported to have potent antimicrobial activity against *Escherichia coli* and *Staphylococcus* depending on the concentration and particle size.⁸ Harikumar & Aravind⁹ found that copper nanocomposites can inhibit *E. coli* and that its effect increased with the increase of concentration and contact time. The antimicrobial property of film containing heat treated CuO-NPs against the growth of *E. coli* were higher than the film containing NaBH₄-treated CuO-NPs.¹⁰ Almasi, Jafarzadeh & Mehryar¹¹ found that CuO-NPs exhibit antibacterial act against Gram negative and Gram positive bacteria and that its inclusion in bacterial cellulose nanofiber reduced its antibacterial activity.

Nanomaterials have been used in food packaging in various forms including nanocomposites polymers with high barrier properties, smart packaging, nano-coatings, surface biocides effect, active packaging and as antimicrobial agents.^{12–14} CuO-NPs have been incorporated in different composites used in food packaging. It is included in two-layer bags made of heat-resistant casein protein layer coated with sodium alginate-pectin.¹⁰ These pouches were used to package coconut edible oil which was found to retard oil oxidation. Low density polyethylene/CuO-NPs composite film was used to package UF cheese¹⁵ whereas it exhibited high antimicrobial activity and UV-light barrier. In addition, the metallic nanoparticles could be utilize in different applications^{16–19} the release of metallic nanoparticles into a food matrix was in the safe acceptable level. Also, CuO-NPs composite film was used successfully in the packing of an Indian fermented milk product (Peda) whereas it exhibited high antimicrobial activity against several pathogenic and spoilage microorganisms.²⁰

Processed cheeses are gaining worldwide continuous popularity due to its diverse composition and wide applications.²¹ The diversity of ingredients used for its formulation permits the production of a wide variety of processed cheeses with different flavours, textures and functions. Recently,²² more attention has been paid to processed cheese as a promising food vehicle able to deliver specific nutrients into human body. Packaging is a crucial step in processed cheese packaging due to its long shelf life at ambient temperatures.²¹ Improper packaging and storage conditions results on surface growth of yeasts and moulds conventionally overcome by adding mould inhibitor to the formulation. The presence of spoilage microorganisms is more critical to the safety of processed cheese. The use of packaging material containing antimicrobial agents may be a good alternative ensuring the safety of the product.

In the current work, novel CMC/PVA/CuO bionanocomposite materials containing different levels of CuO-NPs have been prepared, characterized and evaluated as surface coating film for processed cheese. Changes in the composition, textural and microbiological properties and acceptability of processed cheese from different treatments were followed during storage period.

2. Materials and methods

2.1. Materials

Carboxymethyl cellulose (CMC) polymer was gotten from Kelong Chemical Factory, China. Polyvinyl alcohol (PVA),

copper nitrate and sodium hydroxide were obtained from (Sigma Aldrich). Ras cheese (27.50% fat, 61.00% total solids) and Cheddar cheese (25.50% fat, and 60.00% total solids) were bought from Cairo Market. Soft cheese (13.00% fat, 40.00% total solids) and butter oil were gotten from the Dairy Department, Cairo University. Joha S9 emulsifying salts (BK Giulini Chemie GmbH, Ladenburg, Germany) and citric acid were obtained from the local market. Pathogenic strains: All test strains *Yersinia enterocolitica*, *Aspergillus niger*, *Bacillus cereus* B-3711, *Aspergillus flavus* 3357, *Listeria monocytogenes* 598, *Escherichia coli* and *Salmonella typhimurium* were collected from Dairy Microbiology Lab., National Research Centre, Egypt.

2.2. Methods

2.2.1. Synthesis of CuO nanoparticles by precipitation method.

CuO nanoparticles were manufactured by precipitation procedure *via* copper nitrate (Cu (NO₃)₂). Copper nitrate solution (0.1 M) was prepared in deionized water and (0.1 M) of sodium hydroxide (NaOH) solution was added gradually under strong stirring up to pH 14 where a black precipitate was formed. The precipitate was washed repeatedly with absolute ethanol and deionized water till pH 7 was gotten. The washed precipitate was dried at 80 °C for 16 h and calcined by heating at 500 °C for 4 h.

2.2.2. Preparation of CMC/PVA/CuO bionanocomposites.

Different CMC/PVA/CuO bionanocomposites were synthesized *via* solution casting technique. CMC and PVA solutions (5% w/v) were prepared separately and then mixed at the ratio of 1 : 1 (v/v). The prepared CuO-NPs were added at the ratios of 0.0, 0.3, 0.6 and 0.9% (w/v) to different aliquots of the prepared mixed CMC/PVA solution. The bionanocomposites were prepared by gradual addition of CuO-NPs suspension (1 mg mL⁻¹ in water) to the mixed polymer solution and sonicated for 3 h at 25 °C, 500 W, frequency, 20 kHz and amplitude 50% using ultrasonic homogenizer (Q500 Sonicator, Qsonica, USA). The CMC/PVA/CuO bionanocomposites solutions were transferred into transparent glass Petri dish and left at room temperature for 72 h in order to vaporize the solvent then create the CMC/PVA/CuO bionanocomposites film.

2.2.3. Characterization of CMC/PVA/CuO bionanocomposites

2.2.3.1. X-ray diffraction pattern (XRD). The prepared CuO-NPs powder was evaluated *via* Philips X-ray diffractometer (PW 1930 generator, PW 1820 goniometer) using Cu K α radiation (45 kV, 40 mA, with $\lambda = 0.15418$ nm) with scans were run in the range of a 2θ from 5 to 80° with a step size of 0.02 and step time of 1 s. The d -spacing was calculated in the diffraction patterns using Bragg's equation ($n\lambda = 2d \sin \theta$).

2.2.3.2. Morphological characterization. The morphology of the fabricated bionanocomposites samples was done on surfaces of the prepared bionanocomposite films and examined using scanning electron microscopy (SEM) model JEM-1230, Japan, operated at 120 kV, with a maximum magnification of 600×10^3 and a resolution until 0.2 nm. Also, the morphology of prepared CuO-NPs was evaluated through drying a drop of the solution on a carbon-coated copper grid of the Joel-100S



transmission electron microscope, of resolution of 0.3 nm. The SEM microscope was attached to a dispersive energy spectrometer (EDEX).

2.2.3.3. Evaluation of particle size of the prepared CuO-NPs. The particle size of prepared CuO nanoparticles was evaluated via dynamic light scattering (NICOMP 380 ZLS, Dynamic light scattering (DLS) instrument, USA).

2.2.3.4. Barrier properties. Water vapor transmission rate (WVTR) was carried out using GBI W303 (B) Water Vapor Permeability Analyzer (China) using the cup method. The water vapor transmission rate was calculated by way of the amounts of water vapor transferred through a unit area of the film in a unit time below exact conditions of temperature (38 °C) and humidity (4%) as stated by the subsequent standards (ASTM E96). Furthermore, the oxygen transmission rate (O₂) was evaluated by N530 Gas Permeability Analyzer (China), using (ISO2556-2001) as international standard.

2.2.3.5. FT-IR spectra. The interactions established in fabricated bionanocomposites were assessed by FT-IR spectra and recorded in the range from 400 to 4000 cm⁻¹ using (Shimadzu 8400S) FT-IR Spectrophotometer.

2.2.3.6. Biodegradation characterization. The biodegradation test of bionanocomposite films carried out in soil conditions by burying bionanocomposites samples in soil. Firstly, the bionanocomposites samples were dried till constant weight at 50 °C then the bionanocomposites samples (3 cm × 3 cm) with 2 mm thickness were buried in the soil (pH = 7.5) and the relative humidity was retained at 50–60% by spraying water and the temperature was 20–25 °C. Finally, the bionanocomposites samples were weighed every week for three times, and washed with distilled water, afterward; the bionanocomposites samples were dried at 50 °C for 24 h and then weighed. Results were repeated three times.

2.2.4. Preparation of processed cheese spread. Processed cheese spread was prepared by using Ras cheese (125 g), Cheddar cheese (125 g), soft cheese (350 g), emulsifying salt (30 g), citric acid (0.5 g), butter oil (100 g) and water (270 g). The constituents were mixed, sited in a processing pot of 2.5 kg volume, heated by direct steam up to 90 °C with irregularly mixing at 1400 rpm for 5 min and then heating was discontinued. The resulted hot and melted cheese was packaged in plastic containers (100 g capacity), allowed to cool to 40 °C under controlled conditions before cheese coating. The cheese containers were divided into 4 groups, the 1st was surface coated with CMC/PVA solution and the 2nd, 3rd and 4th groups were coated with CMC/PVA/CuO suspensions containing 0.3, 0.6 and 0.9% of CuO-NPs, respectively. The different coating suspensions were brushing on the cheese surfaces and left for 2 h at room temperature to film formation and containers were capped with their lids. Cheese containers were kept in refrigerator at ~7 °C for 6 months. Representative samples were taken every two months and analyzed.

2.2.4.1. Physicochemical analysis of cheese. Cheese samples were analyzed for moisture, fat, ash and total nitrogen (TN) contents total solid (TS) and titratable acidity (TA), as described in ref. 23. The pH values of cheeses were measured using pH meter via glass electrode (Hanna Co., Italy).

2.2.4.2. Textural profile analysis of cheese. The texture profile measurements (TPA) of processed cheese samples were evaluated according to the International Dairy Federation²⁴ using the double compression test (TMS-Pro texture analyzer, Food Technology Co., USA). The values for texture features, i.e., hardness (N), adhesiveness (N s), gumminess (N), cohesiveness, chewiness (N), and springiness were planned from the TPA graphic.

2.2.5. Microbiological analysis

2.2.5.1. Antimicrobial activity of fabricated CMC/PVA/CuO bionanocomposites as coating suspensions. Firstly, activated test bacterial strains in trypton soya broth, incubated at 37 °C for 24 h and activated tested fungi strains in potato dextrose broth, incubated at 25 °C for 72 h. The antimicrobial assay of bionanocomposites was carried out using the diffusion plate method according to Dodiya & Amin.²⁵ 0.1 mL of the different tested strains (~10⁵ cells per mL) was spread on the surface of the plate containing nutrient agar media and the plates were left at 37 °C for 2 h. Using cork borer (0.5 cm) to prepare wells in the agar layer and after that full each well with 50 µL of different bionanocomposites suspension. After incubation at 37 °C/24 h, the diameter of the clear inhibition zone surrounding the wells was measured.

2.2.5.2. Microbiological properties of processed cheese during storage. Ten grams of each processed cheese samples were homogenized individually with 90 mL of sterilized tri-sodium citrate solution (2% w/v). Decimal dilutions were prepared in 9 mL sterile saline solution. Potato dextrose agar medium was used to count yeasts and moulds after incubated at 25 °C for 4 days in aerobic condition.²⁶ Violet red bile agar medium was used to detect coliforms after incubated at 37 °C for 18 h. Also, plate count agar medium was used to counted total bacteria after incubated at 37 °C for 48 h.²⁷ Plate count agar medium was used to determined psychotropic bacteria after incubated at 7 °C for 7 days.²⁶ Also, spore forming counts were enumerated by plate count agar medium after heated the tubes contained samples for 80 °C for 10 min and cold rapidly²⁸ and plates were incubated at 35 °C for 48 h.

2.2.6. Sensory analysis. The sensory attributes (color, odor, taste, consistency, and oiling off) were judged by a panel taste of 10 experienced members of the dairy department, National Research Centre, Cairo. Panelists were asked to judge each sensory attribute out of 10 point scale.

2.2.7. Statistical analysis. Data were statistically analyzed by two way ANOVA and correlation between moisture and hardness using Vassar States computational site according to Lowry.²⁹

3. Results and discussion

3.1. Evaluation of the prepared CuO-NPs and the bionanocomposites films

3.1.1. X-Ray diffraction (XRD) examinations. The crystal phase records of the tested samples were carried out by XRD as a non-destructive technique widely used for the estimation of crystalline materials. The XRD patterns of the synthesized CuO-NPs as well as CMC/PVA/CuO bionanocomposites are presented



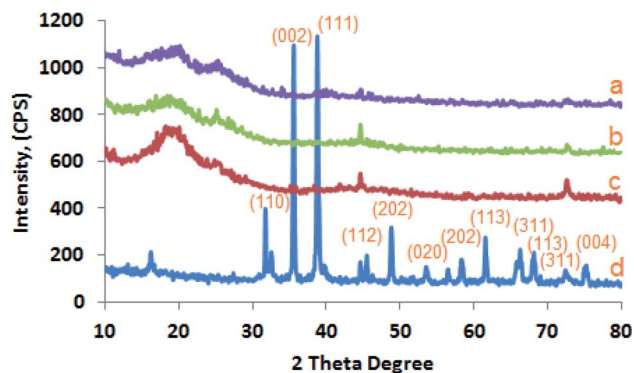


Fig. 1 X-ray diffraction patterns of CuO-NPs (d), as well as the prepared CMC/PVA/CuO bionanocomposites containing different ratios of CuO-NPs, (a) 0.3%, (b) 0.6 and (c) 0.9%.

in (Fig. 1). XRD peaks confirm that the formed CuO-NPs were in the monoclinic phase. Also, X-ray pattern of CMC/PVA/CuO bionanocomposites revealed a fairly well-matched monoclinic structure for pure CuO-NPs.^{30,31} The characteristic peaks located at $2\theta = 31.62^\circ, 35.34^\circ, 38.86^\circ, 48.54^\circ, 48.75^\circ, 53.41^\circ, 56.38^\circ, 58.19^\circ, 61.58^\circ, 66.29^\circ, 67.99^\circ$ and 71.54° are assigned to (110), (002), (111), (112), (202), (020), (113), (311) and (004) plane orientation of CuO nanoparticles (JCPDS 80-1268).³² Furthermore, Fig. 1a–c displayed the CMC/PVA/CuO bionanocomposites containing different 0.3, 0.6 and 0.9% CuO-NPs respectively. The XRD shows that the increase of CuO-NPs in the CMC/PVA matrix led to an increase in the intensity CuO-NPs peaks into the prepared CMC/PVA/CuO bionanocomposite films.

3.1.2. Morphology and particle size analyzer of the fabricated CuO-NPs. The size distribution of the fabricated CuO nanoparticles in this current study is assessed using dynamic light scattering (DLS) instrument (NICOMP 380 ZLS, USA). The particle size distribution of CuO nanoparticles is estimated from (Fig. 2a), is about 12 nm. This, enhanced surface area for interaction, greater thermodynamic stability, and developed bioavailability. The transmission electron microscope (TEM) micrographs of the fabricated and CuO-NPs are shown in (Fig. 2b). It is clear that the prepared CuO-NPs exhibits spherical

and uniform shape with a major particle size of about 10–15 nm. However, few aggregates of CuO-NPs were detected (Fig. 2b) suggesting that some interactive forces between precursor particles produced by OH⁻ bonding were reduced during drying process of the prepared CuO-NPs.

3.1.3. SEM examination of the prepared bionanocomposites. The prepared CMC/PVA/CuO bionanocomposites were evaluated using scanning electron microscope (SEM). Fig. 3 demonstrates the morphology of CuO nanoparticles, CMC/PVA blend and the prepared CMC/PVA/CuO bionanocomposites. The prepared CuO-NPs had sphere-shaped structure as shown in Fig. 3a, while Fig. 3b displays the fabricated CMC/PVA blend which revealed smooth surface indicating high compatibility between CMC and PVA. Fig. 3c and d show that the CuO-NPs were evenly dispersed and embedded into the bionanocomposites matrix in case the addition of 0.3% and 0.9% CuO-NPs, respectively.

The EDEX analysis of the CMC/PVA blend as well as the prepared CMC/PVA/CuO bionanocomposites film fabricated using different loadings % of CuO-NPs, the existences of CuO nanoparticles into the bionanocomposites matrix was proven using EDEX analysis (Fig. 4) which displays that the presence of CuO-NPs in the EDEX and the percent of CuO-NPs in the matrix about 0.3 and 0.9% that demonstrated by SEM analysis (Fig. 3).

3.1.4. FT-IR spectroscopy of the fabricated CMC/PVA/CuO bionanocomposites. FT-IR spectroscopy is a valuable instrument to recognize functional groups in a molecule, to establish possible intermolecular interactions among various constituents in the fabricated bionanocomposites system, Fourier transform infrared spectra of CMC/PVA blend as well as CMC/PVA/CuO bionanocomposites without and with added 0.3% and 0.9% CuO-NPs respectively were recorded in the range 500–4000 cm^{-1} (Fig. 5). Fig. 5a displays the FT-IR spectrum of the CuO-NPs fabricated through hydrothermal method. The symmetric and asymmetric stretching vibrations of the O–H bond appear around 2970 cm^{-1} and 3436 cm^{-1} . The occurrence of bands at 610 cm^{-1} and 1100 cm^{-1} designates various modes of bending vibration of the Cu–O bond. Also, the peak appears at 1578 cm^{-1} shows stretching vibration of the Cu–O bond of CuO-NPs. Furthermore, from Fig. 5 all bands of the CuO

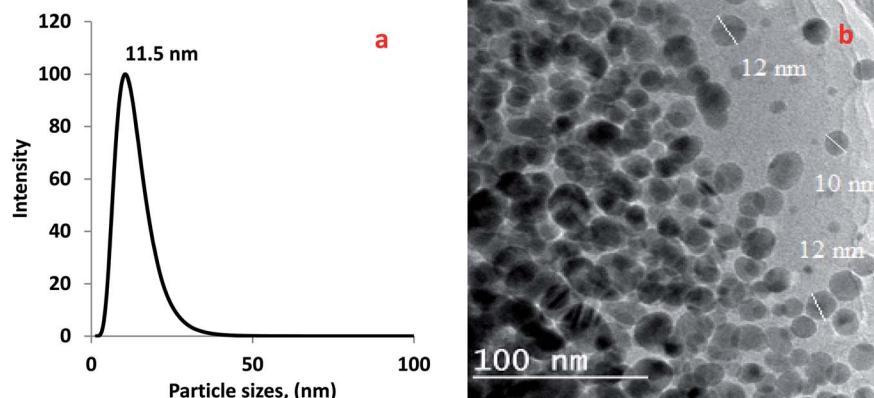


Fig. 2 (a) DLS of the prepared CuO-NPs, (b) TEM image of the prepared CuO-NPs.



nanoparticles, carboxymethyl cellulose (CMC), polyvinyl alcohol (PVA) as well as the prepared CMC/PVA blend were detected in the spectrum of CMC/PVA/CuO bionanocomposites films indicating no changes in their structures after compounding. CMC/PVA blend displays FT-IR absorption peaks appear at 3283 and 2925 cm^{-1} , correlated to the (OH) stretching in addition (CH_2) asymmetric stretching group associated to carboxymethyl cellulose. Additionally, the FT-IR spectrum of polyvinyl alcohol is displayed in Fig. 5, all the main bands accompanying acetate and hydroxyl groups were detected. Furthermore, the comprehensive peak noticed among 3580 and 3150 cm^{-1} is accompanying by the stretching O-H from the intermolecular and intermolecular hydrogen bonds.

The vibrational peak detected between 2860 and 3986 cm^{-1} are attributed to the stretching C-H from alkyl groups also the stretching of C=O and C-O from acetate group from PVA were detected at peaks around 1740 and 1720 cm^{-1} .

Correspondingly, the peaks corresponding to hydroxyl groups shifted to 3312 cm^{-1} and altered into weaker signs for CMC/PVA/CuO bionanocomposites films likened to CMC/PVA blend. Also, it has clear bands at 528 cm^{-1} and 580 cm^{-1} that are correlated to CuO stretching modes. Samples have the absorption bands in range from 1385 to 1642 cm^{-1} which might be associated to OH bending vibrations joined with copper atoms. Thus, FT-IR data proposes that the presence of CuO-NPs in the CMC/PVA blend.

3.1.5. GTR and WVTR of the fabricated CMC/PVA/CuO bionanocomposites films. The prepared bionanocomposites were evaluated using GTR and WVTR instruments; the WVTR is an essential criterion for the relevance of any materials to be used in applications such as food packaging. Therefore, the transportation of water vapor from the adjacent atmosphere to food products or humidity losses from the foodstuff to nearby atmosphere use noteworthy influence on the packaged food

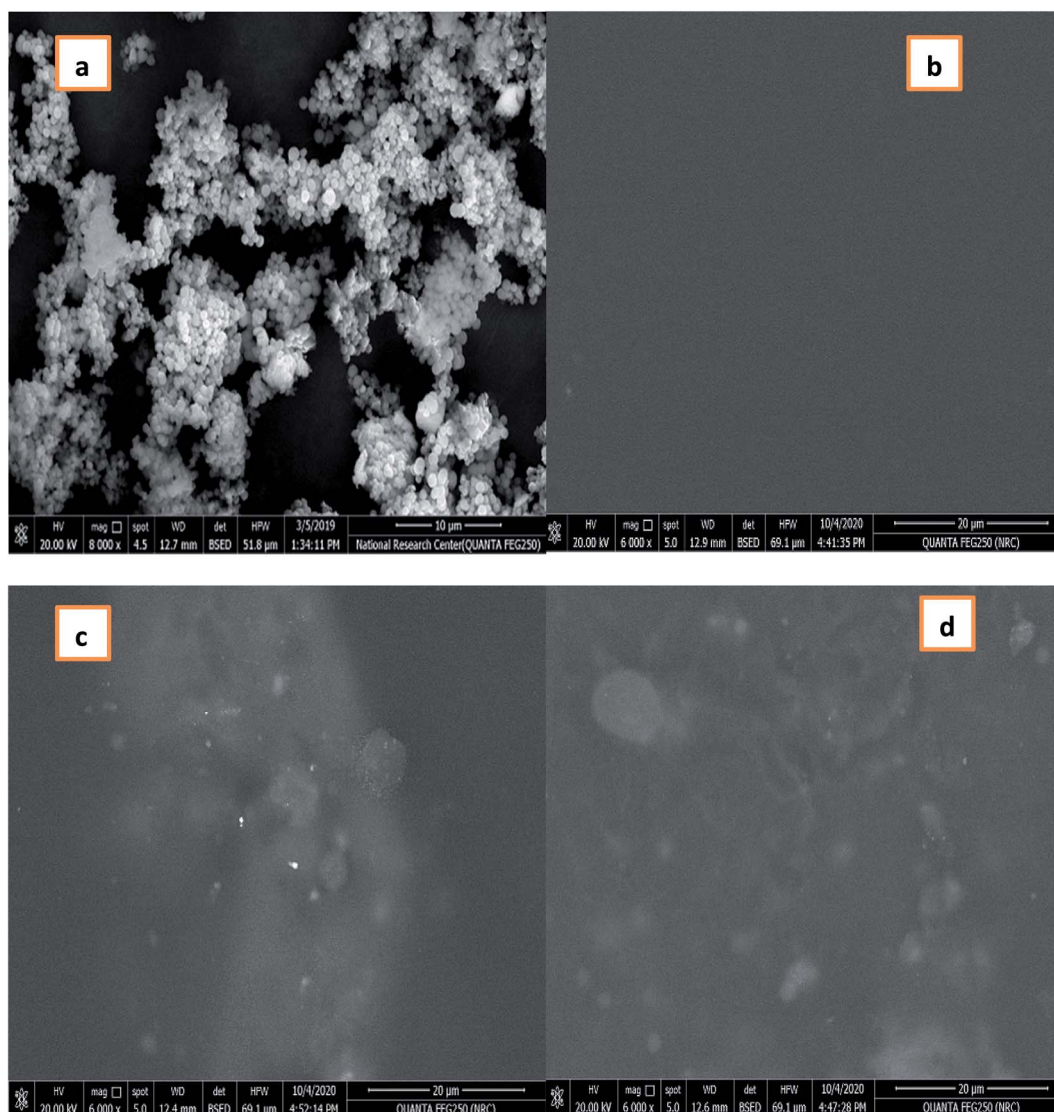


Fig. 3 SEM image of (a) CuO-NPs, (b) CMC/PVA blend, (c) and (d) CMC/PVA/CuO bionanocomposites containing different amounts of CuO-NPs (0.3% and 0.9%), respectively.



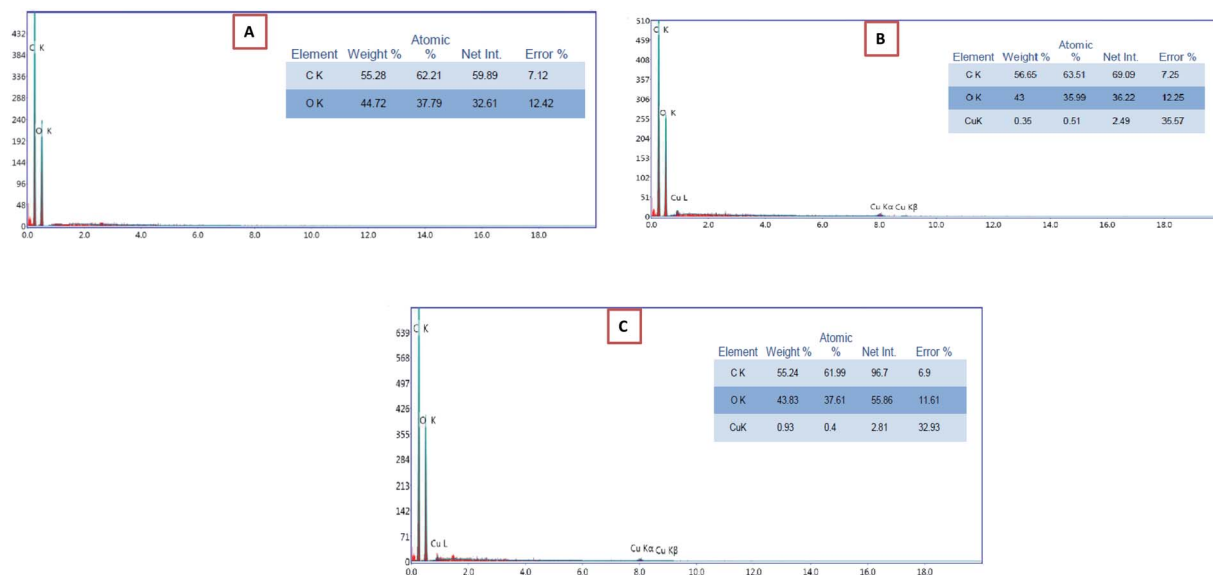


Fig. 4 EDEX analysis of (A) CMC/PVA blend, and (B, C) CMC/PVA/CuO bionanocomposites containing different amounts of CuO-NPs (0.3% and 0.9%), respectively.

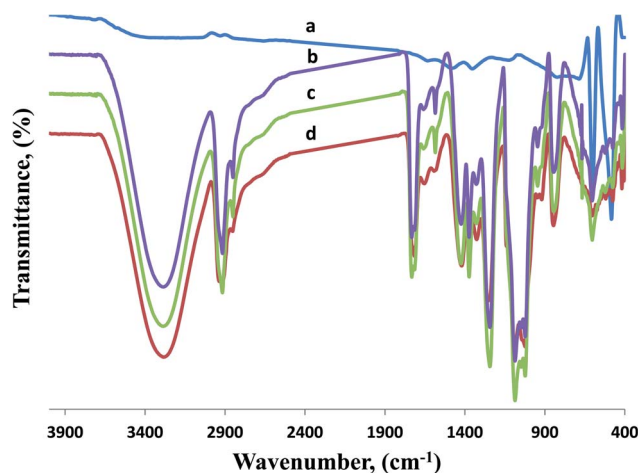


Fig. 5 FT-IR spectra of (a) CuO-NPs, (b) CMC/PVA blend as well as the prepared CMC/PVA/CuO bionanocomposites containing different amounts of CuO-NPs, (c) 0.3% and (d) 0.9%.

such as stability point of view during the shelf-life, the quality, and time. Table 1 displays the WVTR for the fabricated CMC/PVA/CuO bionanocomposites films. Data indicate that CMC/PVA film displayed higher WVTR as related with CMC/PVA/CuO bionanocomposites films. By rising the added percentage

of CuO-NPs in the CMC/PVA/CuO bionanocomposites reduced the WVTR value of the film.

Thus, the adding of 0.3% and 0.9% CuO-NPs decreased dramatically the WVTR value of the prepared bionanocomposites film to 669.35 and 169.13 g per m² per day respectively in comparison to 730.23 g per m² per day for CMC/PVA film. Also, Table 1 demonstrates that resistance for gas permeation of the fabricated bionanocomposites films develops by increasing the percentage of added CuO-NPs in the CMC/PVA blend. Hence the permeability of (O₂) decreased from 33.07 to 25.55 cc per m² per day compared by the CMC/PVA films which display higher GTR compared to the prepared bionanocomposites films. The present results can be explained by the tortuosity effect.³³ Incorporation of impermeable nanoparticles within a polymer matrix represents an obstacle to the diffusion of gases and water vapors. For the permeation of diffusing molecules it has to follow a more tortuous pathway. The volume fraction and shape of the added nanoparticles and their dispersion and distribution within the polymer matrix influences the tortuosity effect.

3.2. Soil burial degradation test

The biodegradation investigation was used to achieve the eco-friendly compatibility of prepared CMC/PVA/CuO bionanocomposites. The biodegradation of the CMC/PVA blend as well

Table 1 The WVTR and OTR of the prepared CMC/PVA/CuO bionanocomposites films^a

Samples	CuO-NPs, ratio	OTR, cc per m ² per day	WVTR, g per m ² per day
CMC/PVA	0.0	71.63 ± 0.67	730.23 ± 0.57
CMC/PVA/CuO	0.3% ± 0.88	33.07 ± 0.58	669.35 ± 0.88
CMC/PVA/CuO	0.9% ± 0.58	25.55 ± 0.33	169.13 ± 0.88

^a All samples are represented as mean of replicates ± standard deviation.



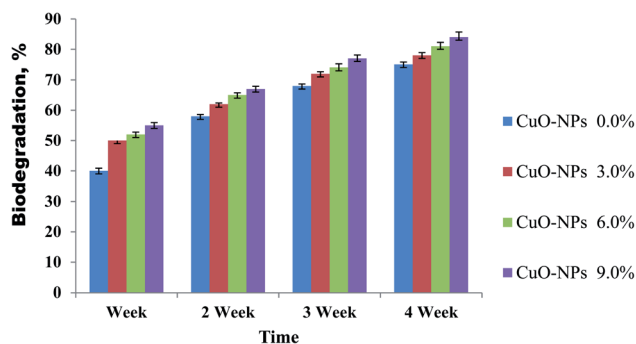


Fig. 6 The biodegradation activities of CMC/PVA/CuO bionanocomposites containing different ratios of CuO-NPs during 4 weeks of soil burial.

as the fabricated bionanocomposites was achieved through a soil burial degradation technique for four weeks. The CMC/PVA blend demonstrated degradation rate in soil around 75% weight loss after 4 weeks of soil burial degradation as shown in (Fig. 6). Furthermore, the biodegradation examination of CMC/PVA/CuO bionanocomposites containing different ratios of CuO-NPs during 4 weeks, displays the biofilms biodegradation (between 78 to 84%) after four week. It can be realized that the highest weight loss was observed for the bionanocomposite film containing high content of CuO-NPs. Furthermore, the obtained data revealed that with raising the CuO-NPs contents

from 0.3 to 0.9 wt%, the biodegradation rate was slightly improved as revealed in Fig. 6. It appears the following cause's for instance large cavity creation by presences of CuO-NPs in the bionanocomposites film surface and rising of agglomeration could improve the biodegradability of the prepared bionanocomposites.³⁴ Moreover, the percentage of weight loss are effected by the composition of bionanocomposites film, and many factors including the type of present microorganisms and its quantity, as well as the degradation conditions (for example; pH, temperature, moisture, and oxygen), impurities amount; and the soil features (specifically the particle size distribution in the soil).³⁵

3.3. Physicochemical properties of processed cheese

Table 2 shows that the moisture content of processed cheese was affected by packaging and storage period. The significant decrease in the moisture content was more pronounced in the surface coated cheese using a film without added CuO-NPs (control) compared to those coated with bionanocomposites containing different percentages of CuO-NPs. At the end of the storage period processed cheese coated with 0.9% CuO-NPs film had the highest moisture content whereas the lowest was in the control. The increased in moisture content in coated cheese during storage can be explained by the improved barrier properties of the used films as evident from the reduced WVTR as revealed in Table 1. Also, there were significant differences in

Table 2 Changes in the moisture, fat, total (TN) and ash contents and acidity and pH of processed cheese as affected by coating and storage period^a

		CMC/PVA/CuO bionanocomposites			
Storage period (month)		0.0 CuO-NPs%	0.3 CuO-NPs%	0.6 CuO-NPs%	0.9 CuO-NPs%
Moisture %	Fresh	59.76 ^{aA}	59.76 ^{aA}	59.76 ^{aA}	59.76 ^{aA}
	2	57.11 ^{dB}	57.53 ^{CB}	58.32 ^{BB}	59.00 ^{AB}
	4	56.16 ^{CC}	57.12 ^{BC}	57.78 ^{AC}	57.96 ^{AC}
	6	55.17 ^{BD}	56.89 ^{AC}	57.10 ^{AD}	57.29 ^{AD}
TN %	Fresh	1.92 ^{aD}	1.92 ^{aD}	1.92 ^{aD}	1.92 ^{aD}
	2	2.10 ^{aC}	2.05 ^{bC}	2.02 ^{bC}	1.98 ^{cC}
	4	2.18 ^{aB}	2.14 ^{bB}	2.08 ^{cB}	2.06 ^{cB}
	6	2.24 ^{aA}	2.21 ^{aA}	2.16 ^{bA}	2.12 ^{cA}
Fat %	Fresh	19.67 ^{aC}	19.67 ^{aC}	19.67 ^{aC}	19.67 ^{aC}
	2	20.67 ^{aB}	20.67 ^{aB}	20.50 ^{aB}	20.17 ^{bB}
	4	21.17 ^{aA}	21.00 ^{aA}	20.67 ^{bB}	20.67 ^{bA}
	6	21.17 ^{aA}	21.17 ^{aA}	21.00 ^{aA}	20.83 ^{aA}
Ash %	Fresh	4.37 ^{aB}	4.37 ^{aB}	4.37 ^{aC}	4.37 ^{aD}
	2	4.66 ^{aA}	4.65 ^{aA}	4.55 ^{bB}	4.47 ^{bC}
	4	4.72 ^{aA}	4.70 ^{aA}	4.62 ^{aB}	4.60 ^{aB}
	6	4.76 ^{aA}	4.74 ^{aA}	4.74 ^{aA}	4.73 ^{aA}
Acidity %	Fresh	1.55 ^{aD}	1.55 ^{aD}	1.55 ^{aD}	1.55 ^{aD}
	2	1.64 ^{bC}	1.65 ^{bC}	1.68 ^{aC}	1.70 ^{aC}
	4	1.75 ^{cB}	1.77 ^{cB}	1.82 ^{bB}	1.85 ^{aB}
	6	1.79 ^{bA}	1.84 ^{aA}	1.86 ^{aA}	1.88 ^{aA}
pH	Fresh	6.03 ^{aA}	6.03 ^{aA}	6.03 ^{aA}	6.03 ^{aA}
	2	5.93 ^{aB}	5.92 ^{aB}	5.89 ^{bB}	5.83 ^{cB}
	4	5.86 ^{aC}	5.82 ^{bC}	5.78 ^{cC}	5.73 ^{dC}
	6	5.79 ^{aD}	5.74 ^{bD}	5.71 ^{cD}	5.68 ^{dD}

^a Means with same superscript, lower case (effect of coating) and upper case (effect of storage) are not significantly different ($P < 0.05$).



Table 3 Changes in the texture parameters of processed cheese as affected by coating and storage period^a

Storage period (month)	CMC/PVA/CuO bionanocomposites			
	0.0 CuO-NPs%	0.3 CuO-NPs%	0.6 CuO-NPs%	0.9 CuO-NPs%
Hardness (N)				
Fresh	2.30 ^{aD}	2.30 ^{aD}	2.30 ^{aD}	2.30 ^{aD}
2	5.54 ^{aC}	5.47 ^{aC}	5.30 ^{bC}	5.20 ^{cC}
4	7.22 ^{aB}	7.00 ^{bB}	6.88 ^{cB}	6.53 ^{dB}
6	9.41 ^{aA}	9.16 ^{bA}	8.76 ^{cA}	8.48 ^{dA}
Cohesiveness (area B/A)				
Fresh	0.642 ^{aA}	0.642 ^{aA}	0.642 ^{aA}	0.642 ^{aB}
2	0.589 ^{aA}	0.571 ^{bB}	0.631 ^{bA}	0.808 ^{aA}
4	0.530 ^{cA}	0.541 ^{cB}	0.661 ^{bA}	0.847 ^{aA}
6	0.637 ^{bA}	0.652 ^{bA}	0.685 ^{bA}	0.881 ^{aA}
Springiness				
Fresh	0.119 ^{aC}	0.119 ^{aC}	0.119 ^{aC}	0.119 ^{aC}
2	0.705 ^{cA}	0.689 ^{cA}	0.758 ^{bA}	0.907 ^{aA}
4	0.633 ^{cB}	0.577 ^{dB}	0.738 ^{bA}	0.883 ^{aA}
6	0.691 ^{cA}	0.672 ^{cA}	0.780 ^{bA}	0.825 ^{aB}
Gumminess				
Fresh	1.477 ^{aD}	1.477 ^{aD}	1.477 ^{aD}	1.477 ^{aD}
2	3.263 ^{cC}	3.123 ^{dC}	3.344 ^{bC}	4.202 ^{aC}
4	3.826 ^{cB}	3.787 ^{dB}	4.547 ^{bB}	5.531 ^{aB}
6	5.994 ^{bA}	5.972 ^{bA}	6.000 ^{bA}	7.471 ^{aA}
Chewiness				
Fresh	0.914 ^{aD}	0.914 ^{aC}	0.914 ^{aD}	0.914 ^{aD}
2	2.300 ^{cC}	2.152 ^{dB}	2.530 ^{bC}	3.811 ^{aC}
4	2.422 ^{cB}	2.185 ^{dB}	3.356 ^{bB}	4.884 ^{aB}
6	4.142 ^{cA}	4.013 ^{dA}	4.680 ^{bA}	6.164 ^{aA}

^a Means with same superscript, lower case (effect of coating) and upper case (effect of storage) are not significantly different ($P < 0.05$).

the moisture content of processed cheese from all treatments as affected by the coating materials which can be attributed to differences in their WVTR.

The fat content of processed cheese increased through storage but without significant differences among treatments (Table 2). The increase in the fat percentage caused mainly from the increase in the total solid (TS) during storage.

Also, Table 2 shows that the total nitrogen (TN) content of coated processed cheese increased slightly at the end of the storage period basically due to loss of moisture. The control cheese had the highest TN content compared to the other coated processed cheese treatments (Table 2), which can be attributed to its low moisture content. There were no significant differences in the content of ash observed between all treatments of the cheese while, with advanced of the storage, ash contents increased significantly.

In all cheese treatments the acidity was significantly raised and pH decreased through storage period (Table 2). Also, slight but significant differences were found in percentage of the acidity of processed cheese from different treatments. These results point to that cheese coating didn't interfere with regular changes in processed cheese during storage.

3.4. Texture profile analysis (TPA) of processed cheese

The TPA parameters of processed cheese from different treatments are presented in Table 3. The hardness of processed cheese was affected by coating and storage period. With the progress of the storage period, hardness increased in all treatments compared to hardness of the fresh cheeses. This may be attributed mainly to the moisture losses and increases in the total solids contents of cheese. A close but negative relation was found between hardness and moisture content of cheese with r^2 value of 0.82. Also, significant ($P > 0.05$) differences were found in the hardness of cheese coated with bionanocomposites films containing different levels of CuO-NPs whereas cheese coated with bionanocomposites containing 0.9% CuO-NPs had the lowest hardness contrary the control treatment.

Correspondingly, the cheese hardness reduced with the rise in the percentage of the CuO-NPs in the coat film. This may be owed to the low water vapor permeability of the bionanocomposites used.³³ Both the storage period and coating had significant effects ($P < 0.05$) on the gumminess and chewiness of processed cheese. Gumminess and chewiness were increased with storage progress and these increases were more obvious in cheese coated with films containing 0.9% CuO-NPs. From the above results it can be established that the most pronounced



Table 4 Changes in the sensory attributes scores (out of 10 points) of processed cheese as affected by coating and storage period^d

Storage period (month)	CMC/PVA/CuO bionanocomposites			
	0.0 CuO-NPs%	0.3 CuO-NPs%	0.6 CuO-NPs%	0.9 CuO-NPs%
Colour				
Fresh	8.0 ^{aA}	8.0 ^{aA}	8.0 ^{aA}	8.0 ^{aA}
2	7.3 ^{aA}	7.6 ^{aA}	8.0 ^{aA}	8.0 ^{aA}
4	5.0 ^{cB}	7.0 ^{bA}	8.0 ^{aA}	8.0 ^{aA}
6	4.0 ^{dC}	6.0 ^{cB}	7.0 ^{bB}	8.0 ^{aA}
Odour				
Fresh	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}
2	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}
4	8.0 ^{aB}	8.6 ^{aB}	8.6 ^{aB}	9.0 ^{aB}
6	7.0 ^{bC}	8.0 ^{aB}	9.0 ^{aB}	9.0 ^{aB}
Taste				
Fresh	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}
2	7.7 ^{bB}	7.7 ^{bB}	9.7 ^{aA}	9.7 ^{aA}
4	7.3 ^{bB}	7.7 ^{bB}	8.7 ^{aB}	9.0 ^{aB}
6	6.3 ^{bC}	6.7 ^{bC}	8.7 ^{aB}	9.0 ^{aB}
Oiling off				
Fresh	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}	10.0 ^{aA}
2	8.3 ^{bB}	9.7 ^{aA}	10 ^{aA}	9.7 ^{aA}
4	8.3 ^{bB}	8.7 ^{bB}	9.7 ^{aA}	9.7 ^{aA}
6	7.3 ^{bC}	7.7 ^{bC}	8.7 ^{aB}	9.0 ^{aB}
Consistency				
Fresh	10 ^{aA}	10 ^{aA}	10 ^{aA}	10 ^{aA}
2	9.3 ^{aB}	9.0 ^{aB}	9.7 ^{aA}	9.7 ^{aA}
4	7.7 ^{cC}	8.0 ^{cC}	9.0 ^{bB}	9.7 ^{aA}
6	6.0 ^{dD}	8.0 ^{bC}	8.0 ^{bC}	8.7 ^{aB}
Total scores				
Fresh	48 ^{aA}	48 ^{aA}	48 ^{aA}	48 ^{aA}
2	42.6 ^{cB}	44 ^{bB}	47.4 ^{aA}	47.1 ^{aB}
4	36.3 ^{cC}	40.1 ^{bC}	44 ^{bB}	45.4 ^{aC}
6	30.6 ^{dD}	36.4 ^{cD}	41.4 ^{bC}	43.7 ^{aD}

^a Means with same superscript, lower case (effect of coating) and upper case (effect of storage) are not significantly different ($P < 0.05$).

changes in the textural properties of processed cheese as affected by storage period and coating happened in hardness, gumminess and chewiness in cheese coated with films

containing 0.9% CuO-NPs. Nevertheless, coating and storage time had no important ($P > 0.05$) effect on springiness of processed cheese.

Table 5 Antimicrobial activity(mm inhibition zone) of fabricated CMC/PVA/CuO bionanocomposites suspensions against some pathogenic microorganisms^a

Strains	CMC/PVA/CuO bionanocomposites			
	0.0, CuO-NPs%	0.3, CuO-NPs%	0.6, CuO-NPs%	0.9, CuO-NPs%
<i>Bacillus cereus</i>	9 ^c	17 ^d	20 ^b	23 ^a
<i>Staphylococcus aureus</i>	11 ^d	19 ^c	23 ^b	30 ^a
<i>Listeria monocytogenes</i>	12 ^d	21 ^c	26 ^b	32 ^a
<i>Escherichia coli</i>	13 ^d	16 ^c	20 ^b	25 ^a
<i>Yersinia enterocolitica</i>	14 ^c	23 ^b	25 ^b	31 ^a
<i>Salmonella typhimurium</i>	10 ^c	14 ^b	20 ^a	22 ^a
<i>Aspergillus niger</i>	10 ^d	14 ^c	23 ^b	30 ^a
<i>Aspergillus flavus</i>	14 ^d	20 ^c	26 ^b	35 ^a

^a Means with same superscript, lower case (effect of coating) in the same row are not significantly different ($P < 0.05$).



Table 6 Changes in the microbiological quality (cfu per g) of processed cheese as affected by coating and storage period^a

Storage period (month)	CMC/PVA/CuO bionanocomposites			
	0.0, CuO-NPs%	0.3, CuO-NPs%	0.6, CuO-NPs%	0.9, CuO-NPs%
Total bacterial count				
Fresh	30.3×10^{aD}	30.3×10^{aD}	30.3×10^{aD}	30.3×10^{aA}
2	17×10^{3aC}	7.6×10^{3bC}	7.0×10^{3bC}	23×10^{cA}
4	79.1×10^{3aB}	42.6×10^{3bB}	27×10^{3cC}	87.6×10^{dA}
6	36×10^{4aA}	81×10^{3bA}	58.3×10^{3cA}	92×10^{dA}
Coliforms				
Fresh	ND	ND	ND	Nil
2	1.1×10	ND	ND	ND
4	1.6×10	ND	ND	ND
6	1.6×10	ND	ND	ND
Moulds & yeasts				
Fresh	ND	ND	ND	ND
2	2×10	ND	ND	ND
4	5×10	ND	ND	ND
6	11×10	3.3×10	1.3×10	0.6×10
Psychrotropics				
Fresh	ND	ND	ND	ND
2	ND	ND	ND	ND
4	8×10	2.3×10	1.6×10	ND
6	15×10	7.6×10	4.3×10	2.0×10
Spore formers				
Fresh	2.3×10	2.3×10	2.3×10	2.3×10
2	4.6×10	3.6×10	3.3×10	ND
4	18.3×10	7.6×10	8.1×10	1.1×10
6	7.6×10	6.6×10	8.1×10	4.1×10

^a Means with same superscript, lower case (effect of coating) and upper case (effect of storage) are not significantly different ($P < 0.05$).

3.5. Sensory properties

The changes in the sensory scores of processed cheese during storage as affected by coating were presented in Table 4. Fresh coated processed cheeses ranked high scores for colour, odour, taste, oiling off and consistency. During storage the colour scores were reduced for all treatments up to the end of storage period. Colour scores in cheese coated with 0.9% CuO-NPs were constant. Control and coated cheeses gained the similar scores for the consistency when fresh and after then significantly decreased until the end of storage period. Oiling off scores in 0.0% loading of CuO-NPs coated processed cheese were lower than other all treatments.

The maximum scores of consistency, odour and taste were given to cheese coated with film containing 0.9% CuO-NPs. Cheese coated with film contain 0.3% and 0.6% CuO-NPs ranked lower scores than cheese coated with film contain 0.9% CuO-NPs for all parameters, while 0.0% CuO-NPs coated cheese take the minimum scores when fresh and during storage period. The total scores of the cheese showed that the cheese coated with film contain 0.9% CuO-NPs was better acceptable than control and other coated processed cheese treatments.

3.6. Antimicrobial activity of CMC/PVA/CuO nanocomposites

Table 5 shows that copper nanoparticles exhibited significant antimicrobial action against different Gram-positive, Gram-negative bacteria and fungi. It also shows that the antimicrobial effect increased significantly ($P > 0.05$) with increasing the mass of fabricated in the tested bionanocomposites suspensions but these increases were nonlinear. It is showed from the inhibition zone that fabricated CuO-NPs possess effective bactericidal action. An insufficient mechanisms contributing to the antimicrobial belongings of the CuO-NPs have been suggested. CuO-NPs might adhere to the bacterial cell walls, because of their positive charge. CuO-NPs interrelate with amine and carboxyl groups existing on the surfaces of microbial cells. Thus, bacteria with higher density of these ionic groups on their cell surfaces have greater affinity for and are highly subject to CuO-NPs. In living microorganisms, copper is a structural constituent for numerous enzymes. Therefore, a comparatively high concentration of Cu^{2+} is necessary to produce toxic effects contrary to microbial pathogens. When managed in high dose, Cu^{2+} ions aid in the creation of ROS, which thereby interact with DNA and intercalate nucleic acid



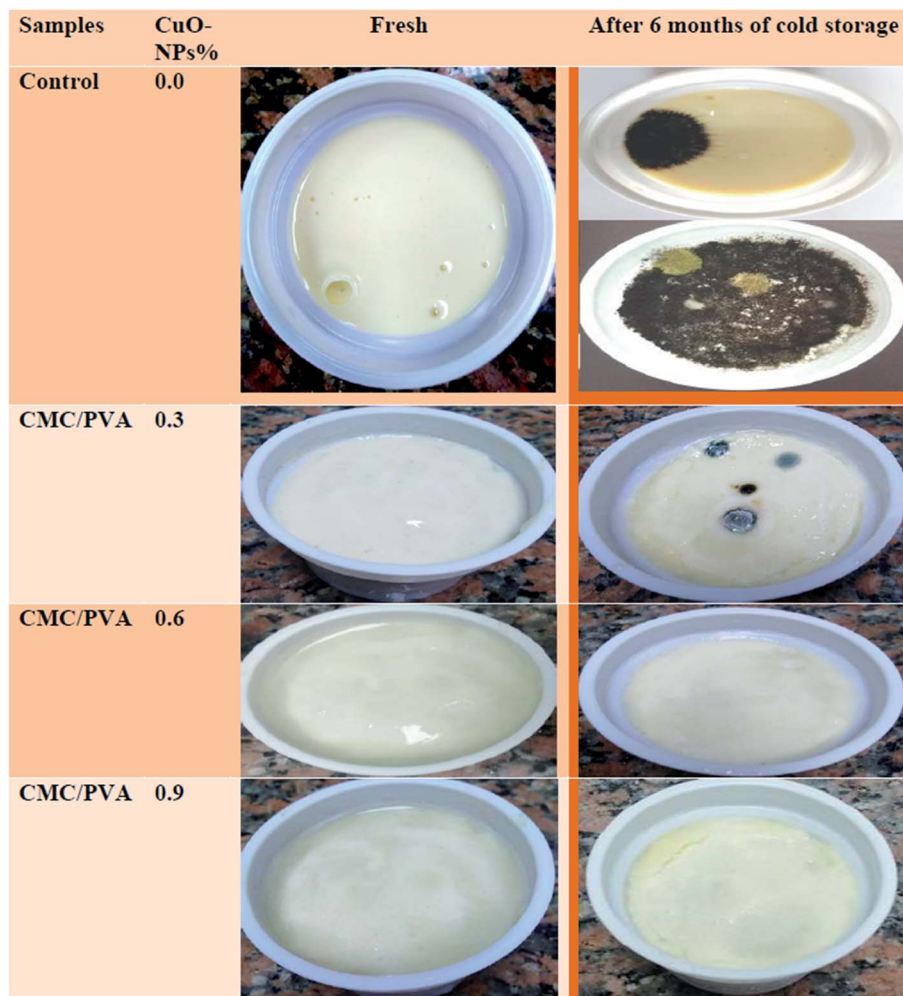


Fig. 7 Influence of the prepared CMC/PVA/CuO bionanocomposites coating film on the quality of processed cheese after 6 months of cold storage.

strands. The release of Cu^{2+} could also disturb the amino acid synthesis in many microbes.³⁶

It is clear that the main factor verbalizing antibacterial mechanism is credited to the presence of ROS creation *via* the CuO-NPs instead of soluble copper ions. Conversely, we cannot exclude certain degree of bacterial ability to solubilize the devoted CuO and so to excerpt harmful Cu^{2+} ions. Therefore, we suggest that collective actions of the strong adherence of the CuO nanoparticles to the cell membrane of bacterial, along with ROS generation on the surface of CuO nanoparticles, cause rise in cell permeability, leading to an unrestrained transport of CuO nanoparticles over the cytoplasmic membrane and ultimately to cells death. This approach of operation mainly rises in the case of small nanometric level of CuO-NPs as a result of their higher surface-to-volume ratio, resultant in the establishment of more ROS per unit weight, and to their higher probability to pass the cell membrane.

The fabricated CuO-NPs exhibited an average diameter of inhibition zone against pathogenic strains around 23–35 mm indicating biocidal action. The higher effect was observed against *A. flavus*, *L. monocytogenes* and *S. aureus* compared with

the CuO-NPs free solution. The soluble ions released from the nanoparticles caused cytotoxicity by interacting either directly with the cellular membrane or intracellularly.^{37,38} Also, polymer/copper metal nanocomposites were developed using polyvinyl methyl ketone, poly-(vinyl chloride), and polyvinylidene fluoride as the polymer matrices. These composites were able to display bacteriostatic and antifungal activities depending on the polymeric matrix properties.³⁹ Also, Harikumar & Aravind⁹ found that copper nanoparticles display antibacterial action against *E. coli* and gained zone was 12 mm.

3.7. Microbiological properties of processed cheese coated with fabricated copper bionanocomposites

Changes in microbiological population in different treatment of processed cheese coated with fabricated copper bionanocomposites are shown in Table 6 and Fig. 7. The total bacterial count of processed cheese from different treatments increased significantly ($P < 0.05$) during storage. However, the rate of the total bacterial growth was retarded with the inclusion of CuO-NPs in the coating film and the most pronounced effect was



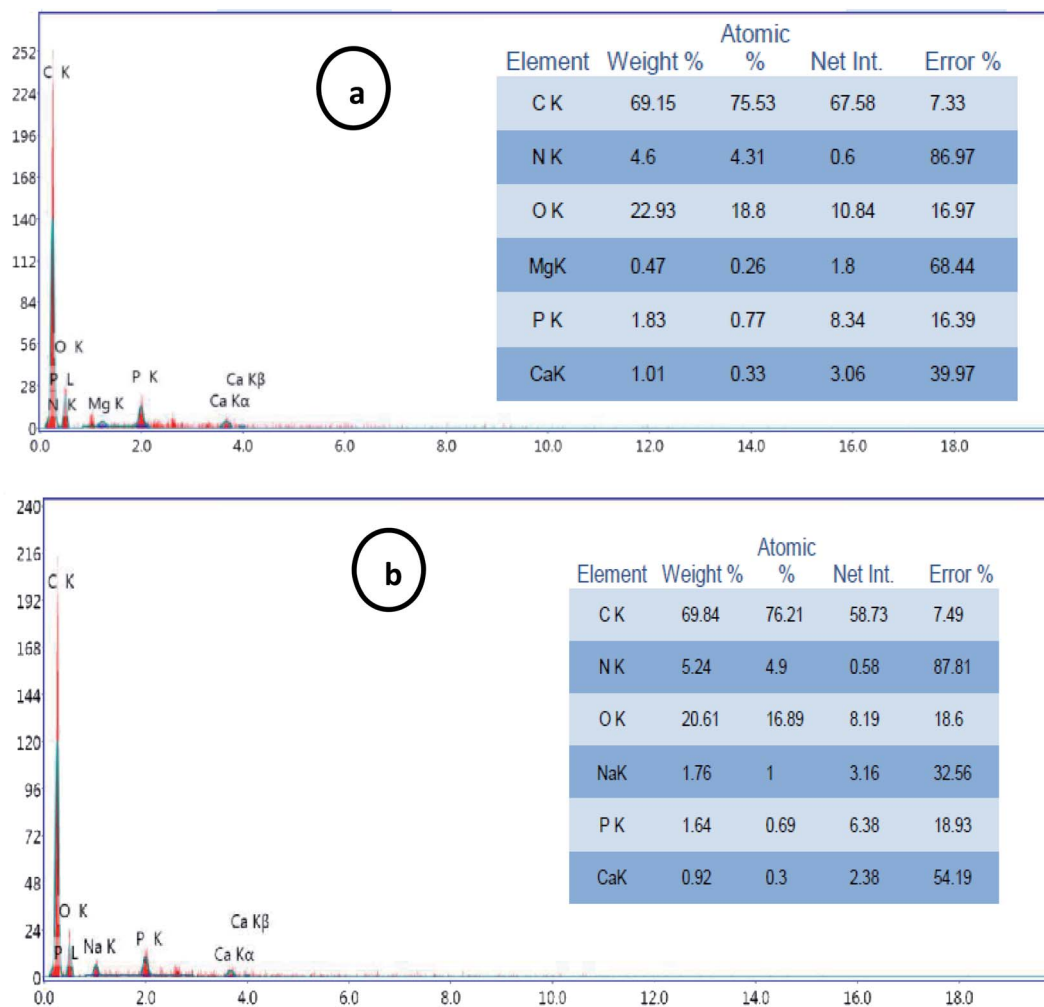


Fig. 8 EDEX data of processed cheese coated using CMC/PVA/CuO bionanocomposites containing CuO-NPs with different concentrations, (a) 0.3%, and (b) 0.9% after 6 months of cold storage.

found with the use of 0.9% CuO-NPs containing film. The retarded bacterial growth cannot be attributed solely to the changes in the moisture content, but the decrease in oxygen permeation may play a role in this respect. No coliforms were detected throughout storage in processed cheese coated with bionanocomposites containing the different levels of CuO-NPs while few numbers were found in control after 2 month of storage.

These results confirmed by Delgado, Quijada, Palma & Palza⁴⁰ who found that the release rate of Cu^{2+} from polypropylene composites based on copper nanoparticles can kill *E. coli* bacteria. No moulds and yeasts were detected in processed cheese coated with bionanocomposites containing the different levels of CuO-NPs up to the 4th month of storage and very few numbers were detected after 6 month of storage. Psychotropics were not detected in control and cheese from different treatments after 2 month of storage and very few were detected thereafter. Small numbers of spore formers were found in control cheese and cheese from the different treatments throughout storage period indicating that the bionanocomposites had no probable effect on the spore formers in processed

cheese. The forgoing results indicate coating processed cheese with the bionanocomposites containing CuO-NPs improved the microbiological quality of processed cheese and prevented surface growth of moulds and yeast and the most pronounced effect was obtained with the use of 0.9% CuO-NPs containing bionanocomposites.

3.8. EDEX investigation of cheese after storage period

Simultaneously, EDEX analysis of the processed cheese after storage period (6 months) coated with CMC/PVA/CuO bionanocomposite loaded with different concentrations of CuO-NPs (0.3 and 9%). The cheese was ashing and the resultant powder subject to EDEX investigation to evaluate the migration of CuO-NPs to cheese throughout the storage period. The EDEX data (Fig. 8) pointed out that no migration of CuO-NPs happened from the bionanocomposite film to coat processed cheese. These results demonstrate the safety of utilize CuO-NPs in direct contact with foodstuffs for small storage time (28 days) without paying care to the migration of nanomaterials from the bionanocomposite coating film to the packaged food.



4. Conclusions

In the present work CMC/PVA/CuO bionanocomposite comprising various concentrations of CuO-NPs have been established for processed cheese coating. Inclusion of CuO-NPs enhanced the water vapor permeability, mechanical properties, as well as the antimicrobial properties of the coating material. The coating material containing 0.9% CuO-NPs decreased moisture losses, improved the microbiological quality of cheese and prevented mould growth on the cheese surface during storage. Also, it gained high acceptability for sensory properties. These prepared bionanocomposites reduce growth of microbial contaminants and noticeably extend and enhance the shelf-life of processed cheese.

Conflicts of interest

There are no conflicts to declare.

References

- 1 N. A. Al-Tayyar, A. M. Youssef and R. R. Al-Hindi, Edible coatings and antimicrobial nanoemulsions for enhancing shelf life and reducing foodborne pathogens of fruits and vegetables: a review, *Sustainable Mater. Technol.*, 2020, **26**, e00215.
- 2 S. M. El-Sayed, H. S. El-Sayed, O. A. Ibrahim and A. M. Youssef, Rational design of chitosan/guar gum/zinc oxide bionanocomposites based on Roselle calyx extract for Ras cheese coating, *Carbohydr. Polym.*, 2020, 116234, DOI: 10.1016/j.carbpol.2020.116234.
- 3 A. M. Youssef, S. M. El-Sayed, H. S. El-Sayed, H. H. Salama and A. Dufresne, Enhancement of Egyptian soft white cheese shelf life using a novel chitosan/carboxymethyl cellulose/zinc oxide bionanocomposite film, *Carbohydr. Polym.*, 2016, **151**, 9–19, DOI: 10.1016/j.carbpol.2016.05.023.
- 4 A. M. El Sayed, S. El-Gamal, W. M. Morsi and G. Mohammed, Effect of PVA and copper oxide nanoparticles on the structural, optical, and electrical properties of carboxymethyl cellulose films, *J. Mater. Sci.*, 2015, **50**, 4717–4728, DOI: 10.1007/s10853-015-9023-z.
- 5 A. M. Youssef and S. M. El-Sayed, Bionanocomposites materials for food packaging applications: Concepts and future outlook, *Carbohydr. Polym.*, 2018, **193**, 19–27, DOI: 10.1016/j.carbpol.2018.03.088.
- 6 A. M. Youssef, F. M. Assem, M. E. Abdel-Aziz, M. Elaaser, O. A. Ibrahim, M. Mahmoud and M. H. Abd El-Salam, Development of bionanocomposite materials and its use in coating of Ras cheese, *Food Chem.*, 2019, **270**, 467–475, DOI: 10.1016/j.foodchem.2018.07.114.
- 7 F. Ijaz, S. Shahid, S. A. Khan, W. Ahmad and S. Zaman, Green synthesis of copper oxide nanoparticles using *Abutilon indicum* leaf extract: Antimicrobial, antioxidant and photocatalytic dye degradation activities, *Trop. J. Pharm. Res.*, 2017, **16**(4), 743–753, DOI: 10.4314/tjpr.v16i4.2.
- 8 S. Shahzadi, N. Zafar and R. Sharif, Antibacterial activity of metallic nanoparticles, *Bact. Pathog. Antibact. Control*, 2018, **51**, DOI: 10.5772/intechopen.72526.
- 9 P. S. Harikumar and A. Aravind, Antibacterial activity of copper nanoparticles and copper nanocomposites against *Escherichia coli* bacteria, *Int. J. Sci.*, 2016, **5**(2), 83–90, DOI: 10.1016/j.tox.2013.07.012.
- 10 G. Gautam and P. Mishra, Development and characterization of copper nanocomposite containing bilayer film for coconut oil packaging, *J. Food Process. Preserv.*, 2017, **41**(6), e13243, DOI: 10.1111/jfpp.13243.
- 11 H. Almasi, P. Jafarzadeh and L. Mehryar, Fabrication of novel nanohybrids by impregnation of CuO nanoparticles into bacterial cellulose and chitosan nanofibers: Characterization, antimicrobial and release properties, *Carbohydr. Polym.*, 2018, **186**, 273–281, DOI: 10.1016/j.carbpol.2018.01.067.
- 12 Y. Huang, L. Mei, X. Chen and Q. Wang, Recent developments in food packaging based on nanomaterials, *Nanomaterials*, 2018, **8**(10), 830, DOI: 10.3390/nano8100830.
- 13 N. A. Al-Tayyar, A. M. Youssef and R. Al-hindi, Antimicrobial food packaging based on sustainable bio-based materials for reducing foodborne pathogens: A review, *Food Chem.*, 2020, **310**, 125915.
- 14 H. F. Youssef, M. E. El-Naggar, F. K. Fouda and A. M. Youssef, Antimicrobial packaging film based on biodegradable CMC/PVA-zeolite doped with noble metal cations, *Food Packag. Shelf Life*, 2019, **22**, 100378.
- 15 F. Beigmohammadi, S. H. Peighambaroust, J. Hesari, S. Azadmard-Damirchi, S. J. Peighambaroust and N. K. Khosrowshahi, Antibacterial properties of LDPE nanocomposite films in packaging of UF cheese, *LWT-Food Sci. Technol.*, 2016, **65**, 106–111, DOI: 10.1016/j.lwt.2015.07.059.
- 16 A. M. Youssef and F. M. Malhat, Selective removal of heavy metals from drinking water using titanium dioxide nanowire, *Macromol. Symp.*, 2014, **337**(1), 96–101.
- 17 A. M. Youssef, F. M. Malhat and A. F. A. Abd El-Hakim, Preparation and Utilization of Polystyrene Nanocomposites Based on TiO₂ Nanowires, *Polym.-Plast. Technol. Eng.*, 2013, **52**(3), 228–235.
- 18 A. Z. Noah, M. A. El Semary, A. M. Youssef and M. A. El-Safty, Enhancement of yield point at high pressure high temperature wells by using polymer nanocomposites based on ZnO & CaCO₃ nanoparticles, *Egypt. J. Petrol.*, 2017, **26**(1), 33–40.
- 19 A. M. Youssef, F. M. Malhat, A. A. Abdel Hakim and I. Dekany, Synthesis and utilization of poly(methylmethacrylate) nanocomposites based on modified montmorillonite, *Arabian J. Chem.*, 2017, **10**(5), 631–642.
- 20 G. B. Lomate, B. Dandi and S. Mishra, Development of antimicrobial LDPE/Cu nanocomposite food packaging film for extended shelf life of peda, *Food Packag. Shelf Life*, 2018, **16**, 211–219, DOI: 10.1016/j.fpsl.2018.04.001.
- 21 R. Kapoor and L. E. Metzger, Process cheese: Scientific and technological aspects—A review, *Compr. Rev. Food Sci. Food*



- Saf.*, 2008, 7(2), 194–214, DOI: 10.1111/j.1541-4337.2008.00040.x.
- 22 E. El Dakhakhny and N. Dabour, Processed Cheese: Basics and Possibility for the Development of Healthier Products, *Alexandria J. Food Sci. Technol.*, 2016, 13(2), 45–62.
- 23 AOAC, *Official Methods of Analysis*, Association of the Official Analytical Chemists, Arlington, VA, USA, 25th edn, 2002.
- 24 IDF, *Rheological and fracture properties of cheeses*, Bulletin No. 268, International Dairy Federation, Brussels, Belgium, 1991.
- 25 B. Dodiya, B. Amin, S. Kamlaben and P. Patel, Antibacterial activity and phytochemical screening of different parts of *Moringa oleifera* against selected gram positive and gram negative bacteria, *J. Pharm. Chem. Biol. Sci.*, 2015, 3, 421–425.
- 26 APHA, *Standard Methods for Examination of Dairy Products*, American Public Health Association, Washington, USA, 16th edn, 1994.
- 27 APHA, *Compendium of Methods for the Microbiological Examination of Food*, American Public Health Association, Washington D.C., USA, 3rd edn, 1992.
- 28 FDA, *Bacteriological Analytical Manual*, AOAC International, Arlington, VA, USA, 9th edn, 2002.
- 29 R. Lowry, *VassarStats: web site for statistical computation* <http://faculty.vassar.edu/lowry.VassarStats.html>, 2009.
- 30 V. S. de Souza, H. O. da Frota and E. A. Sanches, Polyaniline-CuO hybrid nanocomposite with enhanced electrical conductivity, *J. Mol. Struct.*, 2018, 1153, 20–27, DOI: 10.1016/j.molstruc.2017.09.084.
- 31 C. Wolf, H. Angellier-Coussy, N. Gontard, F. Doghieri and V. Guillard, How the shape of fillers affects the barrier properties of polymer/non-porous particles nanocomposites: A review, *J. Membr. Sci.*, 2018, 556, 393–418, DOI: 10.1016/j.memsci.2018.03.085.
- 32 G. Ren, D. Hu, E. W. Cheng, M. A. Vargas-Reus, P. Reip and R. P. Allaker, Characterization of copper oxide nanoparticles for antimicrobial applications, *Int. J. Antimicrob. Agents*, 2009, 33(6), 587–590.
- 33 A. M. Youssef, S. M. El-Sayed, H. S. El-Sayed, H. H. Salama, F. M. Assem and M. H. Abd El-Salam, Novel bionanocomposite materials used for packaging skimmed milk acid coagulated cheese (Karish), *Int. J. Biol. Macromol.*, 2018, 115, 1002–1011, DOI: 10.1016/j.ijbiomac.2018.04.165.
- 34 Z. W. Abdullah and Y. Dong, Biodegradable and Water Resistant Poly (vinyl) Alcohol (PVA)/Starch (ST)/Glycerol (GL)/Halloysite Nanotube (HNT) Nanocomposite Films for Sustainable Food Packaging, *Front. Mater.*, 2019, 6, 1–17.
- 35 I. Korbog and S. Mohamed Saleh, Studies on mechanical and biodegradability properties of PVA/lignin blend films, *Int. J. Environ. Stud.*, 2016, 73(1), 18–24.
- 36 A. J. Huh and Y. J. Kwon, “Nanoantibiotics”: a new paradigm for treating infectious diseases using nanomaterials in the antibiotics resistant era, *J. Controlled Release*, 2011, 156, 128–145.
- 37 C. Gunawan, W. Yang Teoh, C. P. Marquis and R. Amal, Cytotoxic origin of copper (II) oxide nanoparticles: comparative studies with micron-sized particles, leachate, and metal salts, *ACS Nano*, 2011, 5(9), 7214–7225, DOI: 10.1021/nn2020248.
- 38 H. L. Karlsson, Pontus Cronholm, Yolanda Hedberg, Malin Tornberg, Laura De Battice, Sofia Svedhem, and Inger Odnevall Wallinder. “Cell membrane damage and protein interaction induced by copper containing nanoparticles—Importance of the metal release process”, *Toxicology*, 2013, 313(1), 59–69.
- 39 N. Cioffi, L. Torsi, N. Ditaranto, G. Tantillo, L. Ghibelli, L. Sabbatini, T. Bleve-Zacheo, M. D'Alessio, P. Giorgio Zambonin and E. Traversa, Copper nanoparticle/polymer composites with antifungal and bacteriostatic properties, *Chem. Mater.*, 2005, 17(21), 5255–5262, DOI: 10.1021/cm0505244.
- 40 K. Delgado, R. Quijada, R. Palma and H. Palza, Polypropylene with embedded copper metal or copper oxide nanoparticles as a novel plastic antimicrobial agent, *Lett. Appl. Microbiol.*, 2011, 53(1), 50–54, DOI: 10.1111/j.1472-765X.2011.03069.x.

