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Interaction of synthetic and lignin-based sulfonated polymers with hydrophilic, hydrophobic, and charged self-assembled monolayers†

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There is a need to understand the role of polymer structure on its interaction with surfaces to produce effective functional surfaces. In this work, we produced two anionic polymers of lignin-3-sulfopropyl methacrylate (L-S) and poly(vinyl alcohol-co-vinyl acetate)-3-sulfopropyl methacrylate (PVA-S) with similar charge densities and molecular weights. On the gold-coated surface, we deposited self-assembled monolayers (SAM) bearing different terminal moieties namely, hydroxyl, carboxyl, methyl, and amine groups of alkanethiols. This study highlighted the difference between the interaction of L-S and PVA-S and functionalized self-assembled surfaces. The information was generated using advanced tools, such as an X-ray photoelectron spectroscopy (XPS), and a quartz crystal microbalance with dissipation (QCM-D), which facilitated the correlation development between polymer properties and deposition performance on the functionalized surfaces. The higher deposition of PVA-S than L-S onto OH and COOH surfaces was observed due to its greater hydrogen bonding development and higher solubility. The solubility and structure of PVA-S were also beneficial for its higher adsorption than L-S onto CH₃ and NH₂ surfaces. However, the variation in pH, temperature, and salt significantly affected the adsorption of the macromolecules.

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1. Introduction

Polymeric films are ubiquitous in applications ranging from automobiles to construction. The majority of polymer films are multilayer polymeric materials with varied functionalities. To generate multilayer films, the interaction of polymers with different surfaces is critical. Polymer adsorption on different surfaces and at the solid/water interface can happen as a result of hydrophobic, hydrogen-bonding, and electrostatic interactions between polymer segments and surfaces. Polymer adsorption is influenced by the properties of the polymer and

the interaction between the polymer, surface, and solvent. In this context, the difference in the structure of the polymer, *e.g.*, linear or three-dimensional, has shown to have a crucial impact on the adsorption behavior of the polymer on surfaces.^{1,2}

Poly(vinyl alcohol-co-vinyl acetate) (PVA) is known as an odorless, whitish or creamy, nontoxic, biocompatible, thermo-stable linear synthetic polymer used widely in different applications. PVA polymer is used vastly in textile, papermaking, coating industry, 3D printing, optical gas and humidity sensors, emission sensors for vehicles and oral drug delivery, and solar cells.³⁻⁷ PVA, by having a linear structure,² has been functionalized with anionic and cationic monomers to improve its adsorption on fibrous pulp and broaden its application as adhesives and emulsifiers.^{6,8}

Lignin, an abundant phenolic polymer, is one of the alternatives to petroleum feedstocks,⁹ which recently attracted tremendous attention and applications.¹⁰⁻¹⁵ Lignin, by having a complex three-dimensional structure, has been reported to show a distinct interaction behavior compared to linear polymeric materials.^{16,17} However, as lignin has a complicated structure, the interaction mechanisms of lignin derivatives with different functionalized surfaces are still unclear. Revealing these mechanisms is believed to highly affect its end-used applications in wastewater treatment, surface coating, and biological applications, for instance.

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† Electronic supplementary information (ESI) available: ¹H NMR spectroscopy (Fig. S1 and S2), FTIR (Table S1 and Fig. S3), GPC (Table S2), zeta potential (Fig. S4), XPS (Table S3 and Fig. S5), water adsorption at a different temperature on different SAM surfaces (Fig. S6–S9), effect of temperature on the adsorption of L-S, and PVA-S on SAMs (Fig. S10–S17), effect of pH on the hydrodynamic radius (*R_h*) of L-S, and PVA-S (Fig. S18), water adsorption at different pH on different SAM surfaces (Fig. S19–S22), effect of pH on the adsorption of L-S and PVA-S on SAMs (Fig. S23–S30), contact angle of water–air (*θ_{W/A}*) interfaces of PVA-S and L-S polymers at different pH (Table S4), effect of salt concentration on the hydrodynamic radius (*R_h*) of L-S and PVA-S (Fig. S31), effect of salt concentration on the adsorption of L-S and PVA-S on SAMs (Fig. S32–S35). See DOI: 10.1039/d0ra07554j



Polymers could adsorb following altered mechanisms.^{18–20} Thus, interaction mechanisms between the polymer and adsorbing surface include, but are not limited to, charge interaction, hydrogen bonding, van der Waals forces, and hydrophobic interactions. Although the mechanisms behind the adsorption of various polymers on different surfaces in contrastingly charged systems have been studied, information on the adsorption in the absence of electrostatic interaction for lignin and PVA based polymers is limited. To address this, two types of branched anionic polymers of poly(vinyl alcohol-*co*-vinyl acetate)-3-sulfopropyl methacrylate (PVA-S) and lignin-3-sulfopropyl methacrylate (L-S) were produced. To eliminate the effect of molecular weight and charged group in comparing lignin and PVA, the reaction condition was controlled to produce polymers with a similar molecular weight and charge density. This will elucidate the role of the structure and surface chemistry of polymers in adsorption.

Self-assembled monolayers (SAMs) with different surface chemistries were used to study the adsorption of polymers and the adsorption kinetics.^{21–24} These surfaces are extensively used in electrochemical sensors²⁵ and also as model surfaces to study the adsorption of polymers, such as proteins.^{26,27} Various SAMs with various combinations of moieties can be used to elucidate the different contributions of the driving force for polymer adsorption.²³ In studying polymer adsorption on SAM surfaces, it was found that polyelectrolytes and the surface with a like-charges were shown adsorption.²³ In other words, although these systems had an interaction barrier of electrostatic origins, the adsorption of the polymer was fast.²³ In another study, the adhesion forces between a hydrophobic surface (CH₃-SAM) and alkali lignin was analyzed and the results revealed the importance of hydrophobic interaction between the CH₃ surface and in non-modified lignin.²⁴ As another objective of this work, the comparison of lignin and poly(vinyl alcohol-*co*-vinyl acetate) adsorption on different SAM surfaces could provide insights into the impact of polymer properties on the adsorption performance of polymers on altered surfaces. To the best of the authors' knowledge, the interaction of sulfonated PVA and lignin-based compounds on SAMs have not been studied.

In the present study, the adsorbed amount of PVA-S and L-S on OH, COOH, CH₃, and NH₂ functionalized surfaces were studied for the first time using QCM-D to provide information on their altered adsorption behavior. The selected SAM surfaces carried distinct terminal functional groups relevant to the surfaces used in different industries, such as surface coatings, mining, pharmaceutical, and cosmetics.^{4,7,9} Also, conducting adsorption studies under different saline and pH conditions would reveal the performance of these sulfonate-based polymers on altered surfaces in different environments and the interaction mechanisms of the adsorption processes on the surfaces.

The main goal of this paper was to identify how the interaction of lignin derivatives, *i.e.*, highly branched materials, is different from their synthetic linear equivalents when their molecular weights and charge densities are similar. This paper provides fundamental insights into quantitative adsorption fundamentals of lignin and synthetic macromolecules.

Demonstrating this difference would help establish methods to improve the characteristics of lignin for creating valorized lignin derivatives with desired functionality.

2. Experimental section

2.1 Materials

In this work, 11-mercapto-1-undecanol (–OH, 97%), 12-mercaptododecanoic acid (–COOH, 96%), 1-dodecanethiol (–CH₃, ≥98%), ammonium hydroxide, 6-amino-1-hexanethiol hydrochloride (–NH₂), vinyl acetate (99%), 3-trimethylsilyl-(2,2,3,3-*D*₄)-propionic acid sodium salt (TMSP) (99.8%), hydrochloric acid (37%), sodium hydroxide (99.0%), poly diallyl dimethylammonium chloride (PDADMAC) with the molecular weight of 100–200 kg mol^{–1}, potassium chloride (KCl), D₂O (with the isotopic purity of 99.8%), 3-sulfopropyl methacrylate potassium salt (98%) (S), sodium persulfate (Na₂S₂O₈), sodium chloride (99%), ethanol (99.8%), methanol (99.8%) and dimethyl sulfoxide-*D*₆ ([*D*₆]DMSO) (99.9%), and hydrogen peroxide (30 wt%) were all purchased from Sigma-Aldrich company. Cellulose acetate membrane with the molecular weight cut-off of 1000 g mol^{–1} was purchased from Spectrum Labs. Inc., USA. Also, softwood kraft lignin (L) was produced *via* the LignoForce technology of FPInnovations in Thunder Bay, ON, and received as a raw material. High-performance liquid chromatography grade water was produced using a Milli-Q water purifier with the resistivity of less than 18 MΩ cm^{–1} and used throughout this work. AT-cut gold-coated piezoelectric quartz crystal sensors (5 MHz resonant frequency) were purchased from Biolin Scientific Inc. Nylon filters with a pore size of 0.22 μm were purchased from the Celltreat Scientific company. Additionally, all of the chemicals utilized in this work were of analytical grades.

2.2 Synthesis of L-S and PVA-S polymers

Softwood kraft lignin (L) was polymerized with 3-sulfopropyl methacrylate potassium salt (S) based on the methodology described in a previous study.²⁵ The reaction was conducted at a molar ratio of 1.2 S/L, 10 wt% HCl, 90 wt% of the water in the presence of 1.5 wt% of potassium persulfate (as initiator) at 80 °C for 90 min. The produced polymer was purified with membrane dialysis for three days to remove any unreacted monomers, and the purified lignin-3-sulfopropyl methacrylate sample was denoted as L-S.

In another set of reactions, vinyl acetate was used, instead of lignin, in polymerizing with 3-sulfopropyl methacrylate potassium salt (S). The reaction was conducted by using vinyl acetate (VA) and S in a molar ratio of 0.5 S/VA, 10% NaOH, 9/1 v/v of methanol/water in the presence of 1.5 wt% of potassium persulfate (as initiator) at 80 °C for 90 min.^{6,28} The produced polymer was precipitated by methanol precipitation and centrifugation at 3500 rpm for 10 min and then purified using membrane dialysis for three days. The poly(vinyl alcohol-*co*-vinyl acetate)-3-sulfopropyl methacrylate sample was denoted as PVA-S. Polymeric solutions were prepared in the concentration of 1 g L^{–1} for different analyses in this study.



2.3 Static and dynamic light scattering

The light scattering analysis of produced L-S and PVA-S polymers were performed by a static light scattering instrument, Brookhaven BI-200SM, equipped with a goniometer. The laser polarized light was set at 633 nm.²⁹ The cell was set at different temperatures ranging from 15 to 65 °C. The samples were passed through a 0.22 μm diameter porous filter. The second virial coefficient (A_2), and the average radius of gyration (R_g) were obtained from the concentration dependence and slope of the angle based on Zimm plot eqn (1), respectively.^{30–32}

$$\frac{Kc}{\Delta R_\theta} = \frac{1}{M_w} \left[1 + \frac{16\pi n^2}{3\lambda^2} R_g^2 \sin^2\left(\frac{\theta}{2}\right) \right] + 2A_2c \quad (1)$$

where $K = 4\pi^2 n^2 (dn/dc)^2 / N_A \lambda^4$ with A_2 is the second virial coefficient, θ is the measurement angle, n is the refractive index of the liquid medium, R_g is the radius of gyration, N_A is Avogadro's number, λ is the laser wavelength (633 nm), and ΔR_θ is the excess Rayleigh ratio [$\Delta R_\theta = R_\theta(\text{solution}) - R_\theta(\text{solvent})$], respectively.

The hydrodynamic radius (R_h) of the polymers was determined using a dynamic light scattering (DLS) instrument, NanoBrook, ZetaPALS, Brookhaven Instruments Corp., USA, which was equipped with a 35 mW power laser ($\lambda = 637$ nm, wavelength). Measurements were made at 90° at different temperatures (15–65 °C), and sample solutions were filtered using 0.2 μm pore size filters for light scattering measurements. The hydrodynamic radius (R_h) was measured based on the diffusion coefficient (D) by using the Stoke–Einstein eqn (2):^{25,33,34}

$$R_h = \frac{K_B T}{6\pi\eta_s D} \quad (2)$$

where η_s , K_B , T are the viscosity of the solvent, Boltzmann constant, and the absolute temperature, respectively.

Differential refractometry (DR) technique was used to determine the macromolecular solutions' specific refractive index increments (dn/dc). It is essential to measure the dn/dc precisely to obtain an accurate M_w value. The refractive index increment of each polymer solution (dn/dc) was measured using a Brookhaven BI-DNDC instrument. Although many studies have reported dn/dc values for homopolymers, limited reports are found on the dn/dc values for heteropolymers.^{29,35}

The second virial coefficient (A_2), the average radius of gyration (R_g), hydrodynamic radius (R_h), and refractive index increments (dn/dc) were conducted three times and the average values were reported.

2.4 Characterization

The charge density of samples was determined with a Particle Charge Detector (PCD 04, BTG Mütek GmbH) using a 0.005 mol L⁻¹ PDADMAC or 0.005 mol L⁻¹ PVSK solution as the titrant, as explained elsewhere.^{14,36} The elemental analysis of the polymers was performed using an elemental analyzer (Vario EL Cube, Elemental Analyzer, Germany) as explained in detail elsewhere.³⁷ The molecular weight of PVA-S and L-S polymers was measured by a gel permeation chromatography (GPC,

Malvern GPCmax VE2001 with multi-detectors) after treating the samples at different pH values (4.0, 6.7, and 11.0) for 12 h and purifying the samples using membrane dialysis for 3 days.³⁸ The details of this analysis can be found in the ESI† file.

The phenolic hydroxyl and carboxylate group contents of L and L-S samples were determined using an automatic potentiometric titrator (785 DMP Titrino, Metrohm, Switzerland) with the HCl standard solution as a titrant (more information can be found in the ESI†).

The contact angle of water–air ($\theta_{w/A}$) of PVA-S and L-S polymers at different pH were conducted using Theta Lite contact angle analyzer (Biolin Scientific, Finland) associated with a camera-based on our previous experiment.²⁵ PVA-S and L-S were coated on glass slides using a spin coater (WS-650, Laurel Technologies Corp) and dried overnight. Then, the contact angle of 1.5 μL of a droplet on coated surfaces was determined.

The structure of L, PVA-S, and L-S was analyzed by a ¹H NMR spectroscopy with 32 scans. Samples were dissolved in D₂O or [D₆] DMSO and stirred until fully dissolved (*i.e.*, for 12 h).^{9,39} Trimethylsilyl propionic acid (TMSP) was used as the internal standard.⁴⁰ More information about this experiment is available in the ESI† file.

The zeta potential of lignin (L), L-S, and PVA-S polymers were analyzed using a NanoBrook Zeta PALS (Brookhaven Instruments Corp, USA) at pH ranging from 3.0 to 11.0. The analysis was performed three times, and the average values were reported in this study.

2.5 Self-assembled monolayers (SAM)s preparation

In the QCM-D measurements, AT-cut piezoelectric quartz crystals (Biolin Scientific Inc), which were covered with gold and had a fundamental frequency of 5 MHz, were used. The crystals were modified according to the procedure described by Hedin *et al.*⁴¹ The surfaces were initially cleaned for 10 min in a UV/ozone chamber, which was followed by immersing in a 1 : 1 : 6 mixture of hydrogen peroxide (25%), ammonium hydroxide (25%), and Milli-Q water for 8 min at 78 °C. To obtain chemically well-defined and electrically inert SAM surfaces, the crystals were immersed overnight (>15 h) in 20 mL of 2 mM of alkanethiol solution in degassed ethanol (99.8%) and an amber bottle, and stirred at 50 rpm and 25 °C in a water bath. Then, 11-mercapto-1-undecanol (–OH), 1-dodecanethiol (–CH), 12-mercaptododecanoic acid (–COOH), and 6-amino-1-hexanethiol hydrochloride (–NH₂) chemicals were used to form hydroxyl (OH), methyl (CH₃), (COOH) and (NH₂) terminated SAMs on the surfaces, respectively. The thiol group of *n*-alkanethiols bonds to Au surfaces by metal–sulfur bonds (*i.e.*, chemisorption) and forms close-packed SAMs (*i.e.*, self-assembled monolayers), which leaves –OH, –COOH, CH₃, and NH₂ (*i.e.*, the other end of the functional groups) available on the surface.^{23,41} To remove excess thiols from the treated surfaces, the surfaces were washed and ultrasonicated in ethanol five times, each time for 5 min. The SAM surfaces were dried under N₂ gas.

2.6 SAM characterization

Theta Lite Contact Angle analyzer (Biolin Scientific, Finland) associated with a camera was employed to quantitatively



analyze the wettability of the Au surfaces before and after SAM deposition. The contact angle analysis of water droplets (1.5 μL) on the bare and modified QCM sensors was conducted using the sessile drop method based on Young's equation with three independent measurements. The values of the water contact angle were evaluated for the four types of SAM are reported in Table 2 and are consistent with reports in the literature.^{23,42,43}

The X-ray photoelectron spectroscopy (XPS) measurements were performed on the bare and SAM-coated sensors by using a Kratos Axis Supra with a monochromatic Al K α radiation (1486.6 eV) with 1 mm spot diameter at a base pressure of about 3×10^{-10} mbar and 20 eV pass energy. Based on the surface plane, the take-off angle for the detected photoelectrons was adjusted to 60°. For energy referencing, spectra were calibrated to the C1 line peak at 284.6 eV.⁴⁴ Data analysis and peak fitting were conducted using ESCAPE software (V1.2.0.1325). The thickness of SAM-terminated surfaces on Au substrates was estimated by means of the attenuation of the Au 4f signal using the following eqn (3):

$$I = I_0 \exp\left[-\frac{d}{\lambda \sin \theta}\right] \quad (3)$$

where I and I_0 are the average intensities of the Au 4f_{5/2} and Au 4f_{7/2} peaks, attenuated by SAM and the bare gold surface, respectively. θ is the photoelectron take-off angle, λ is the effective attenuation length of the photoelectron, and d is the film thickness.^{44,45} The theoretical length values of SAM molecules used in this work have been attained from computational modeling Avogadro software (V1.2.0) (written in C++ (Qt) with General Public License) and force field model of MMFF94 (designed by Merck).

2.7 Quartz crystal microbalance with dissipation (QCM-D)

The detailed description of the QCM-D technique has been explained by Li *et al.*⁴⁶ and Pensini *et al.*⁴⁷ The adsorption studies of L-S and PVA-S were conducted on the above mentioned SAM-coated gold sensors by using a QCM-D, E1, 401, instrument (Q-Sense Inc., Gothenburg, Sweden). A peristaltic pump was used to pump solutions at the flow rate of 0.15 mL min⁻¹ through the chamber of the QCM instrument. The temperature was set to room temperature (25 °C) for all experiments. The adsorption experiment was initially conducted with a buffer solution of the desired aqueous pH and electrolytes, until achieving a baseline signal in the analysis. Then, the adsorption experiment was initiated by replacing the buffer solution with L-S or PVA-S polymer solutions with the same pH and electrolyte concentration. After reaching saturation in adsorption, the solutions were changed to the buffer solutions for removing unadsorbed components from the SAM coated surfaces. The solutions were degassed prior to the experiment. This analysis was conducted at different salt concentrations (1–1000 mM) and pH (3.0–11.0) with PVA-S and L-S solutions. It is worth noting that the PVA-S, L-S, and buffer solutions had the same pH and salinity in all experiments.

In this analysis, Sauerbrey and Voigt equations were used to evaluate the properties of the adsorbed layers and analyse

the changes in the adsorbed mass on the sensor (more information is available in the ESI†). The Sauerbrey equation was used when $\Delta D \leq 1 \times 10^{-6}$, which would be an indication of an elastic and rigid layer. However, data points were fitted into the Voigt model for layers with the higher dissipation using three harmonic overtones of 5, 7, and 9 of the Q-Tools software. The 9th overtone was used for the better representation of data (details available in the ESI†). Also, the fluid density and viscosity were considered to be 0.99 g cm⁻³ and 1.05 mPa, respectively. The error bars related to the adsorbed mass present the standard deviation of the repeated experiments.

3. Results and discussion

3.1 Characterization of produced polymers

The ¹H-NMR spectra of the produced samples are depicted in Fig. S1 and S2.† The appearance of a new peak at 0.96 and 1.10 ppm is assigned to the methyl group of S monomer in the ¹H-NMR spectra.^{25,48,49} The three peaks at 4.7, 4.5, and 4.2 ppm are assigned to the proton in the (OH) group of PVA (Fig. S2†).^{39,50} The peak at 1.97 ppm is also assigned to the methyl (CH₃) group of PVA.^{39,51} More information could be found in the ESI.† The results in Fig. S1 and S2† confirm the successful production of L-S and PVA-S. Also, FTIR analysis was performed on the obtained polymers and the spectroscopy, as well as the peak data, are shown in Fig. S3 and Table S1 in the ESI† file, respectively. The results of this analysis also confirmed the successful polymerization and the production of L-S and PVA-S. In addition, the reaction yield for the production of L-S (65.32%) was measured to be lower than the reaction yield for the production of PVA-S (74.21%). The properties of produced polymers are shown in Table 1. Based on the elemental analysis (Table 1), L (unmodified lignin), L-S, and PVA-S had 0.76, 8.03, and 8.17 wt% sulfur element, respectively, and no traceable nitrogen. The charge density (CD) of L, L-S, and PVA-S were -0.85, -3.17, and -3.20 meq. g⁻¹, respectively, which would indicate that the amount of introduced sulfur content to polyvinyl acetate and lignin was equal. The GPC analysis demonstrated the molecular weight of 1.8×10^4 , 11.4×10^4 , and 11.3×10^4 g mol⁻¹ for L, L-S, and PVA-S, respectively. Since the adsorption of the produced polymers is going to be analyzed on different surfaces at different pH, it is crucial to analyze the stability of the polymers at different pH. To do so, the molecular weight analysis was performed using GPC for the samples pretreated in both acidic and alkaline pH for 12 h. As seen in Table S2,† compared to neutral pH, the PVA-S, and L-S samples showed 7.6–8.9, and 4.0–0.2% reduction in their molecular weights, respectively. These results reveal that both polymers are relatively stable over the examined pH range. The charge of produced polymers at various pH was determined through zeta potential analysis and presented in Fig. S4 in the ESI† file. By changing the pH, the zeta potential of L was changed from about -7.5 to -17.6 mV. The zeta potential increment was observed more drastically for both PVA-S and L-S polymers than L, which could be due to their sulfonate groups.



Table 1 Chemical properties of produced polymers

Samples name	L	L-S	PVA-S
Nitrogen content, ^a wt%	<0.09 ^b	<0.09 ^b	<0.09 ^b
Sulfur content, ^a wt%	0.76	8.03	8.17
Charge density, ^a $\mu\text{eq. g}^{-1}$	-0.85	-3.17	-3.20
Carboxylate content, ^a mmol g^{-1}	0.17	0.16	—
Phenolic hydroxyl group content, ^a mmol g^{-1}	1.86	0.18	—
M_w (GPC), g mol^{-1}	1.8×10^4	11.4×10^4	11.3×10^4
M_n (GPC), g mol^{-1}	0.48×10^4	6.38×10^4	6.05×10^4
dn/dc , ^a mL g^{-1}	0.1320	0.1601	0.0660
Reaction yield, %	100	65.32	74.21

^a Error was <0.05%. ^b Method sensitivity <0.09.

3.2 Characterization of self-assembled monolayers (SAM)

A series of SAM-modified surfaces were produced by reacting thiols with different end groups on the QCM sensor. Fig. 1 demonstrates the X-ray photoelectron spectroscopy (XPS) of SAM with -OH, -NH₂, -CH₃, and -COOH end groups on the sensors. The presence of a peak at 162 eV is attributed to the Au-S energy binding, which indicates the successful modification of Au with SAMs.

Table S3 (in the ESI† file) includes the relative atomic concentrations of SAM-modified surfaces (C, O, S, and N), which confirms the successful formation of SAM. The excess amount of O in -CH₃ could be attributed to the atmospheric contamination. In the case of -NH₂ modified surface, the excess amount of O could be attributed to the atmospheric contamination (3.8%) and partial oxidation of the S-Au bond to sulfonate (7.3%). The S_{2p} level of -OH, -COOH, -CH₃, and -NH₂ SAM-terminated surfaces were studied, and it has been found that the partial oxidation of -SH to sulfonate happened only in NH₂ terminated SAM, which could be attributed to the HCl used to stabilize amino compound.⁵²

Table 2 demonstrates the properties of the four types of SAMs. The sessile drop method was used to measure the deionized water contact angle on SAM coated sensors.^{42,43} The

water contact angle of the gold sensor was $75 \pm 2^\circ$. The contact angle of -OH, -COOH, -CH₃, and -NH₂ surfaces were measured to be $<10^\circ$, 30° , 110° , and 41° , respectively. At neutral pH (pH 6.7), -OH and -CH₃ carry no charges while -NH₂ is positively charged and -COOH is negatively charged.⁵³ The contact angle measurements of SAM films suggested that the surface coated with the -CH₃ group was highly hydrophobic, while the surfaces coated with -OH, -COOH, and -NH₂ groups were wettable. SAMs functionalized with -OH could be considered as super-hydrophilic as the water droplet spread on the SAM surface (a contact angle of $<10^\circ$). According to the literature, the contact angle for -OH functionalized SAM was reported to be as low as 9.5° ^{23,52} and as high as $40\text{--}50^\circ$.^{54,55} The monolayer surface containing OH has been reported to be unstable.⁵⁶ The rate of contact angle variation was also revealed to be dependent on temperature. Also, freshly prepared surfaces were suggested to be used for more credibility of this analysis, which was performed accordingly in this work. These SAMs (-CH₃, -OH, -COOH, and -NH₂) were well characterized and used extensively as model surfaces.^{57,58}

The theoretical lengths of SAMs were obtained from computational modeling, Avogadro software (V1.2.0), are reported in Table 2. By considering the estimated values achieved

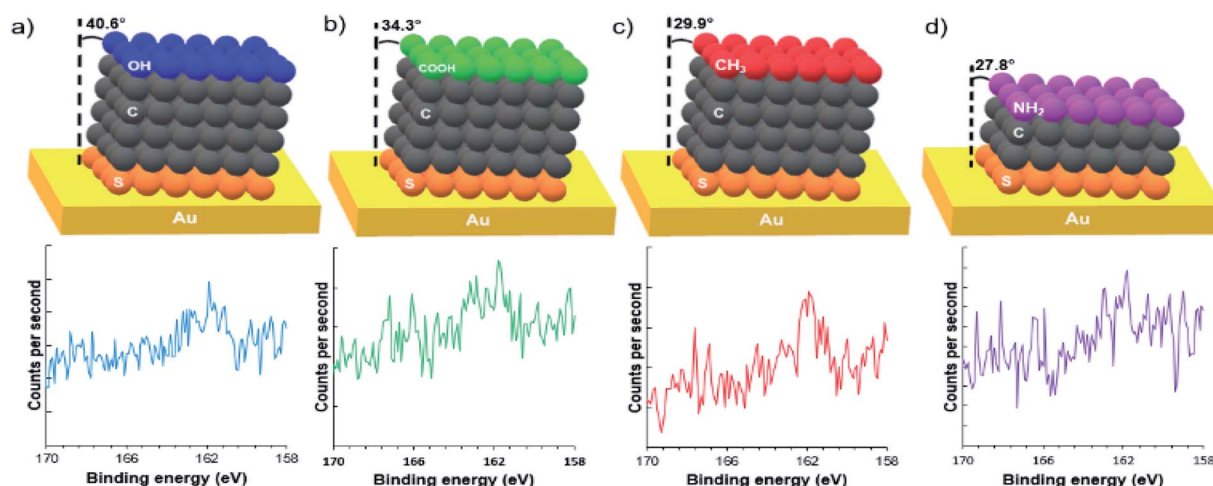


Fig. 1 SAMs with different terminated groups on gold: XPS S 2p spectra of (a) -OH, (b) -COOH, (c) -CH₃, and (d) -NH₂.



Table 2 Used SAMs and their properties

Surface	Chemical formula	SAMs name	Water contact angle, °	Features	Charge at pH 6.7	Theoretical length, Å	Thickness, Å	Tilt angle, °
OH	HS(CH ₂) ₁₁ OH	11-Mercapto-1-undecanol	<10	Hydrophilic	Neutral	17.1	12.97	40.6
COOH	HS(CH ₂) ₁₁ COOH	12-Mercaptododecanoic acid	30 ± 3	Hydrophilic	–	18.3	15.11	34.3
CH ₃	HS(CH ₂) ₁₁ CH ₃	1-Dodecanethiol	110 ± 2	Hydrophobic	Neutral	17.3	15.00	29.9
NH ₂	HS(CH ₂) ₆ NH ₂ ·HCl	6-Amino-1-hexanethiol hydrochloride	41 ± 1	Hydrophilic	+	10.8	9.55	27.8

from the Au 4f (Fig. S5†) attenuation analysis, the SAMs show an atilt angle in the range of 27.8–40.6° for the surface (Table 2 and Fig. S5†). If SAMs were to be defective, isolated domains would have been formed on the surfaces, which would have further resulted in single molecules to generate a lying-down configuration.⁵² Thus, our results reveal a defect-free and tightly packed SAMs. In other words, all SAMs had significant high-packing densities while exhibiting differential end groups.^{52,58}

Also, to analyze the interaction of SAM surfaces with water molecules, water adsorption on different SAM surfaces was conducted at different temperatures (Fig. S6–S9†) and pH (Fig. S10–S13†). Based on the obtained results, no significant difference (*i.e.*, within 10%) was observed between SAM surfaces. This indicates that the chemically modified SAM surfaces are stable under the examined conditions. Also, these conditions (different temperatures and pH) showed no specific effect on the swelling of SAMs (Fig. S6–S13†).

3.3 Measurements of the hydrodynamic radius (R_h), the radius of gyration (R_g), and second virial coefficient (A_2)

Fig. 2a depicts the hydrodynamic radius, R_h , of PVA-S, and L-S in pure water. As can be seen, the R_h of L-S increased by increasing

the temperature from 15 to 65 °C. The increase in the overall size of L-S with temperature enhancement could be due to more hydration of lignin structure at a higher temperature. In the case of PVA-S, the insignificant change in the R_h might be attributed to the dehydration of PVA-S caused by polyvinyl acetate chains at a high temperature. Similar behavior was reported for poly(ethylene oxide) in water.^{59,60} Fig. 2b depicts the radius of gyration, R_g , defined as the root mean square distance of a particle's components from its center of mass. As seen, enhancing the temperature drastically increased the R_g of L-S, while it did not significantly affect the R_g of PVA-S. It is worth mentioning that R_g is sensitive to the refractive index distribution (mass distribution), while R_h is sensitive to hydrodynamics.³³ It is well established that the ratio of R_g/R_h is a characteristic parameter related to the conformation of polymer chains in solutions. The values of R_g/R_h for uniform hard-sphere, random coil, and rod-like structures have been reported to be 0.778, 1.78, and ≥ 2 , respectively.^{61–64} In the case of L-S, both R_g/R_h and R_g have increased with increasing the temperature. At 15 °C, the R_g/R_h ratio for L-S was about 0.77, while the R_g/R_h ratio increased to about 1.27 at 65 °C, revealing that the L-S present a swollen structure.³² An increment in the

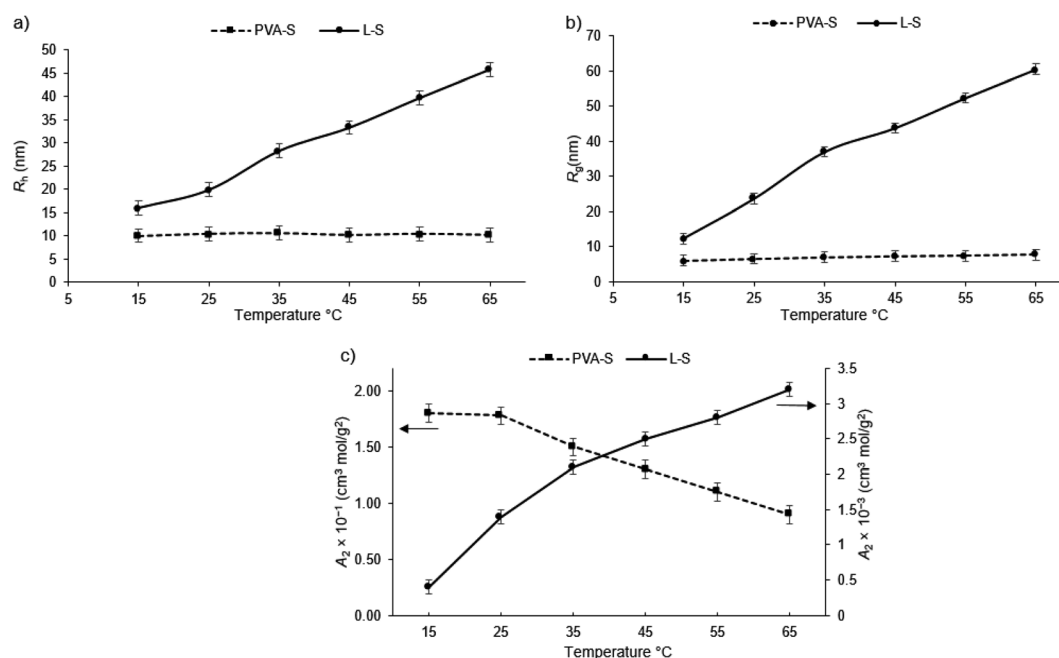


Fig. 2 Temperature dependence of (a) hydrodynamic radius (R_h), (b) radius of gyration (R_g), and (c) second virial coefficient (A_2) of PVA-S and L-S polymers.



R_g/R_h ratio for L-S in the range of 0.77 to 1.27 (from 15 to 65 °C) is in agreement with the intermediate structure from a hard-sphere to a looser structure due to the water swelling effect.⁶⁵ The respective R_g/R_h ratio of PVA-S changed from ~ 0.60 at 15 °C to ~ 0.76 at 65 °C. Thus, the spherical shape of the PVA-S polymer was insignificantly altered *via* temperature alteration. The higher R_g/R_h ratio at a high temperature also ascribed to the dehydration of PVA-S.⁶⁶

Fig. 2c indicates the second virial coefficient (A_2) of the polymers, which signifies the polymer–solvent interactions. A low value of A_2 means strong interactions among solute particles, indicating that the solute is in a poor solvent, which means that the polymers are partially or completely insoluble in the solvent. The magnitude of A_2 values denotes the strength of such interactions. The variation in A_2 with temperature depends on the hydrophilic/hydrophobic structure of polymers.^{67–69} Based on Fig. 2c, the A_2 value for L-S polymer increases as the temperature rises from 15 to 65 °C, implying that high temperature improved the interaction of L-S polymer and water. The intensive interactions between polymer segments could lead to a more compact structure with the hard-sphere configuration at low temperature, while the more efficient interaction between L-S and water loosened the structure of L-S polymer at a higher temperature, which is supported by results depicted in Fig. 2a and b. In the case of PVA-S (Fig. 2c), A_2 values decreased by increasing the temperature implying that water has become a poor solvent for this polymer at a higher temperature. Similar behavior has been reported for poly(vinyl alcohol).^{70,71} As reported in the literature, the PVA macromolecule chain becomes dehydrated above critical solution temperature (25 °C in our case).^{72,73}

The lower solubility of L-S than PVA-S, based on the second virial coefficient (A_2), could be due to two reasons: its aromatic

structure,^{74,75} and, its pK_a value (phenolic groups), which drops at higher temperatures and leads to an increase in its solubility.^{76,77}

3.4 Effect of temperature on the adsorption of L-S and PVA-S on SAMs

3.4.1 Adsorption on –OH functionalized surface. Fig. 3 illustrates the adsorption analysis of L-S and PVA-S on the OH-functionalized surface at different temperatures of 25 °C, 35 °C, and 45 °C (raw data is available in the ESI in Fig. S14 and S15†). Based on these results (Fig. 3a), limited adsorption was observed for L-S on the OH functionalized surface. Also, increasing the temperature did not improve the adsorption. Although the R_g and R_h of L-S increased with respect to temperature increment (Fig. 2a and b), they could not improve the development of hydrogen bonding between L-S polymer and OH-functionalized surface. Also, enhancing the temperature decreased the adsorption of PVA-S on the OH-functionalized surface, which might be due to a decrease in its solubility at higher temperatures (Fig. 2c).

3.4.2 Adsorption on –COOH functionalized surface. Fig. 3b shows the adsorption of L-S and PVA-S on COOH-functionalized surfaces at different temperatures (in the range of 25–45 °C) (raw data is shown in the ESI in Fig. S16 and S17†). The adsorption of L-S on the COOH-functionalized surface was improved marginally by increasing the temperature. At room temperature, the carboxylate group exists in its deprotonated ($-\text{COO}^-$) form. By increasing the temperature, the carboxylate group becomes protonated ($-\text{COOH}$),⁷⁸ resulting in a decrease in the negative charge density of the surface. This phenomenon paves the way for hydrophilic interactions of L-S with the surface to become more dominant than electrostatic attraction.

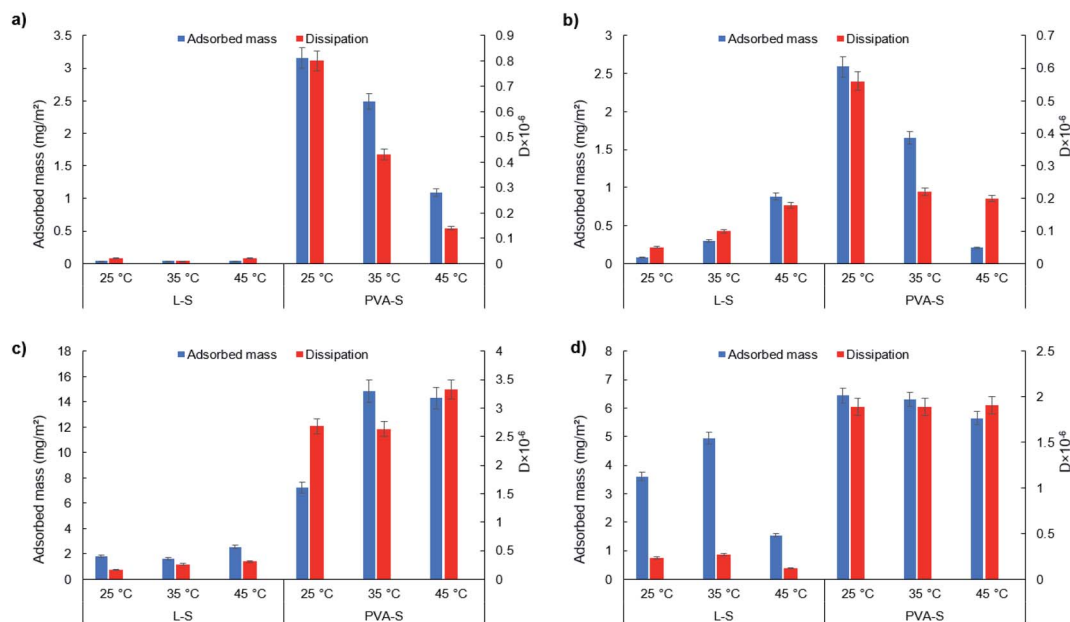


Fig. 3 Adsorbed mass and dissipation of L-S and PVA-S on (a) OH, (b) COOH, (c) CH₃, and (d) NH₂ functionalized surfaces at different temperatures.



However, it should be noted that even at a higher temperature (45 °C), the adsorption amount of L-S does not increase significantly, which is a clear indication of the limited adsorption of L-S on the -COOH-functionalized surface (Fig. 3b).

The adsorbed amount of PVA-S is observed to decrease sharply from 2.59 mg m⁻² at 25 °C to 0.21 mg m⁻² at 45 °C. As explained earlier, the solubility of PVA-S was reduced at higher temperatures (Fig. 2c), and thus its driving force for adsorption dropped. By increasing the temperature, the interaction between hydrophilic groups of PVA-S would decrease. The reason for this could be the denser structure of PVA at higher temperatures, which would result in dehydration in the structure of PVA⁷² and a lower adsorption amount on the -OH functionalized surface. On the other hand, by lowering the hydrogen bonding capability of PVA-S, more hydrophobic parts of PVA-S (*i.e.*, CH) would be exposed and interact with -CH₃ functionalized surface.

3.4.3 Adsorption on -CH₃-functionalized surface. Fig. 3c indicates the adsorption of polymers on the -CH₃-functionalized surface at different temperatures (in the range of 25–45 °C) (the raw data is presented in the ESI in Fig. S18 and S19†). According to Fig. 3c, the L-S adsorption was increased by enhancing the temperature. As the R_g and R_h values for L-S were increased by the temperature increment, its hydrophobic features were more exposed (Fig. 2), promoting its interaction and thus adsorption on the hydrophobic surface (*i.e.*, -CH₃-functionalized surface). Also, the adsorption of PVA-S on this surface (Fig. 3c) increased in general, which might be attributed to its solubility reduction at a higher temperature (Fig. 2c).

3.4.4 Adsorption on -NH₂-functionalized surface. Fig. 3d indicates the adsorption of L-S and PVA-S on the NH₂-functionalized surface (the raw data is demonstrated in the ESI in Fig. S20 and S21†). As seen, the highest adsorption of L-S was achieved at 35 °C. Increasing the adsorption from 25 to 35 °C could be due to the improvement in lignin solubility at higher temperatures (Fig. 2c). In the case of PVA-S (Fig. 3d) and L-S at higher than 35 °C, by increasing the temperature, the adsorption of PVA-S on the NH₂-functionalized surface decreased gradually. This reduction in the adsorption for both L-S and PVA-S polymers could be due to the weakening of electrostatic interaction between negatively charged polymers and the positively charged surface at higher temperatures.⁷⁸

Overall, comparing the adsorption of L-S on the NH₂ (Fig. 3d) and CH₃ (Fig. 3c) surfaces, the higher adsorption of L-S was found on the NH₂-functionalized surfaces. This implies that the electrostatic interaction could be more dominant than hydrophobic interactions for L-S. In other words, temperature increment could improve the electrostatic interaction development of L-S with surfaces more than its hydrophobic interaction. The similar trend observed for PVA-S adsorption on -NH₂ and -CH₃-functionalized surfaces could indicate that the electrostatic attraction and hydrophobic interaction were both playing important roles in PVA-S adsorption onto the functionalized surfaces. In addition, comparing L-S and PVA-S, the highest adsorption capacity of L-S (4.95 mg m⁻²) was achieved at 35 °C on -NH₂ functionalized surface (Fig. 3d). Meanwhile, the highest adsorption amount of PVA-S (14.83 mg m⁻²) was

obtained at 35 °C on the -CH₃ functionalized surface (Fig. 3c). The unmodified lignin (L) polymer contains different functional groups of phenolic hydroxyl (1.86 mmol g⁻¹ (Table 1), carboxylate (0.17 mmol g⁻¹ (Table 1), carbonyl, and methoxy groups, which could provide the macromolecule with hydrogen bonding capability. However, the rigid and three-dimensional structure of lignin¹¹ would limit the chance of available functional groups on L-S to interact with the solid surfaces containing -OH and COOH groups and lead the sites on lignin to prefer developing hydrogen bonds with the solution rather than the surfaces.

3.5 Effect of pH on the adsorption of L-S and PVA-S on SAMs

3.5.1 Adsorption on -OH-functionalized surface. Representative experimental traces for the QCM adsorption analysis of L-S and PVA-S on OH-functionalized surface are shown in Fig. 4 under three different pH of 3.0, 6.8, and 11.0, while the raw data is presented in the ESI file in Fig. S22 and S23.† As seen in Fig. 4a, L-S limitedly adsorbed on this surface at all pH ranges, which might reveal that L-S was incapable of developing noticeable hydrogen bonding with this surface in all pH.^{79,80} On the other hand, it is seen that PVA-S could adsorb on the OH-functionalized surface more than L-S at all pH, while having the most adsorption at pH 3.0, and the least at pH 11.0. This indicates a stronger electrostatic interaction of PVA-S with the surfaces (results shown in Fig. S22†), since variations in the pH affect the charge of polymers, begetting an attraction or a repulsion in the electrostatic interaction.⁸¹ Moreover, not only the hydrophilicity of PVA-S is higher than L-S (Table S4†), but PVA-S is also more soluble (Fig. 2c), which can adsorb more water. It is worth noting that the steric hindrance of lignin could also adversely affect its adsorption to the surfaces.⁸² Also, since the OH groups of lignin are substituted with S groups after the polymerization reaction, less free hydroxyl groups are left on lignin to develop hydrogen bonding interaction. While in the polymerization of PVA, the S groups do not replace the OH groups, leaving more hydroxyl groups available on PVA-S to interact through hydrogen bonding with OH-functionalized surface. The higher adsorption of PVA-S than L-S (Fig. S22 and S23†) increased the dissipation on the sensor.

3.5.2 Adsorption on -COOH-functionalized surface. Fig. 4b includes the adsorption of L-S and PVA-S on the -COOH-functionalized surface at different pH, while the raw data is depicted in the ESI in Fig. S24 and S25.† As seen, L-S adsorption to the surface was higher at pH 3.0, and it decreased by the pH elevation. At pH 3.0, L-S could develop a hydrophilic interaction with -COOH through its functional groups. However, this interaction diminishes due to the repulsion force resulted from the higher negativity of the -COOH surface at higher pH. The incapability of L-S in hydrogen bonding development is once more observable due to its minimal adsorption to the surface, which was discussed earlier.^{79,80}

A similar trend of reduction in the PVA-S adsorption to -COOH surface was observed by increasing the pH, which is due to the enhancement in the repulsion force between the polymer and the surface. While electrostatic interactions play a critical



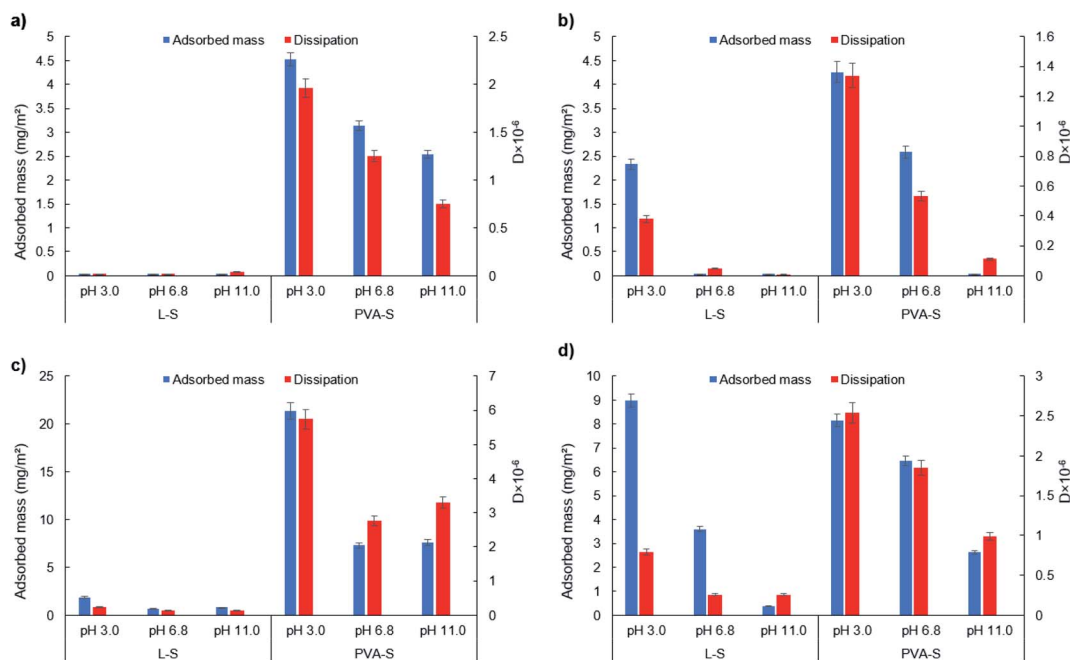


Fig. 4 Adsorbed mass and dissipation of L-S and PVA-S on (a) OH, (b) COOH, (c) CH₃, and (d) NH₂ functionalized surfaces at different pH.

role in the adsorption of like-charged polymers (*i.e.*, electrostatic repulsion), the interesting PVA-S adsorption on the negatively charged –COOH surface indicates the sensible development of nonelectrostatic interactions between PVA-S and COOH surface (*e.g.*, van der Waals and hydrogen bonding).

3.5.3 Adsorption on –CH₃-functionalized surface. Fig. 4c also reveals the adsorption of L-S and PVA-S polymers on the –CH₃ functionalized surface, while the raw data is presented in the ESI file in Fig. S26 and S27.† As seen, the adsorbed mass of L-S onto the surface was reduced at all pH. This interaction reveals that L-S has adsorbed on the –CH₃ functionalized surface *via* a nonelectrostatic interaction (hydrophobic interaction), which is hypothesized to develop through its aromatic units. It has also been reported that by increasing the pH, the diffuse layer potential for CH₃ becomes negative, which leads to a reduction in the adsorption amount of negatively charged polyacrylate (PAA) and polystyrene sulfonate (PSS) on this surface.²³ Similar results were obtained for the adsorption of PVA-S on –CH₃ as a function of pH.

It is seen that L-S showed less hydrophobic interaction than PVA-S with the CH₃ functionalized surface at all pH studied. This might be because of the linear structure of PVA-S, which facilitates the hydrophobic interaction of PVA-S with the surface, while the three-dimensional structure of L-S prohibits the hydrophobic interaction of the aromatic core of lignin with CH₃ (*i.e.*, steric hindrance).

3.5.4 Adsorption on –NH₂-functionalized surface. The adsorption of the polymers on the positively charged –NH₂ functionalized surface is also included in Fig. 4d, while the raw data is shown in the ESI file in Fig. S28 and S29.† As seen, a sharp and fast decrease in the adsorbed mass of L-S onto –NH₂ surface is observed, which reveals a strong electrostatic

attraction of this polymer with the surface at pH 3.0, since this surface bears ionized groups at this pH and appears in the form of NH₃⁺. It has also been reported that in the oppositely charged systems, adsorption kinetics are very fast.^{83,84} The polymer adsorption was reduced by increasing the pH, which might be due to the accumulation of a larger amount of hydroxyl ions at the NH₂/water interface. This leads to a reduction in the positivity of the surface, less electrostatic interaction with the polymer, and thus less adsorption.²³ Also, a similar trend of adsorption was observed for PVA-S as a function of pH. Comparing the adsorption of L-S on NH₂ and CH₃ (Fig. 4) surfaces, the higher adsorption of L-S was observed on the NH₂ surface.

3.5.5 Overall performance at different pH. The results in Fig. 4 suggest that L-S illustrated significantly different adsorption mechanisms than did PVA-S at different pH values. At pH 3.0, the maximum adsorption of L-S was observed on the NH₂ surface, while the highest adsorption of PVA-S was observed on the CH₃ surface. The monotonical decrease in the adsorption of L-S and PVA-S was observed with increasing the pH for almost all surfaces. The decrease in the adsorption was mainly reported to be attributed to the decrease in the magnitude of the diffuse-layer potential when this potential is contrary to that of the adsorbed polymer.^{23,83,85} This effect is due to the reduction in the attraction force and subsequently enhanced repulsion force at the solid/water interface between the deposited polymers.^{83,86} Also, the smaller changes in dissipation values of L-S than PVA-S were observed for almost all surfaces at different pH, which might be due to its lower adsorption to SAMs (Fig. S22–S29†).

Furthermore, increasing the pH (from 3.0 to 11.0) led to the enhancement in the hydrodynamic radius (R_h) of L-S and PVA-S



polymers, and this change was more sensible for L-S (Fig. S30[†]), which can be due to its better solubility at higher pH. According to the contact angle analysis (Table S4[†]), PVA-S was more hydrophilic than L-S at all pH. This means that PVA-S could trap more water in its structure, which might lead to more changes in the adsorbed mass upon its adsorption to SAM surfaces. Although these two polymers have the same amount of sulfonate groups, their structural difference¹ led to their altered interactions with varied surfaces. Also, its linear structure² favored the adsorption of PVA-S on the functionalized surfaces.

In general, the maximum adsorption amount of L-S (8.99 mg m⁻²) was attained at pH 3.0 on the -NH₂ functionalized surface. Meanwhile, the highest adsorption amount of PVA-S (21.33 mg m⁻²) was obtained at pH 3.0 on -CH₃ functionalized surface.

3.6 Effect of salt concentration on the adsorption of L-S and PVA-S on SAMs

The adsorption of L-S and PVA-S on different functionalized surfaces at equilibrium is shown in Fig. 5 and 6 (the original data is provided as Fig. S31–S34 in the ESI[†] file). In saline systems, the long-range electrostatic double-layer interactions between the polymers and surfaces become remarkably small while the nonelectrostatic forces come into action.²³ By increasing the salt concentration, the adsorption of L-S polymer increased on all surfaces (Fig. 5a). These results suggest that nonelectrostatic forces played a role in the adsorption of L-S on carboxyl and hydroxyl-coated surfaces. The more significant

change for OH and COOH might be due to nonelectrostatic forces becoming relatively important compared to electrostatic interaction in salty systems.

Fig. 5b illustrates the dissipation changes on different alkanethiol surfaces. It is seen that the increase in the salt concentration (from 1 mM to 1000 mM) elevated the dissipation of L-S polymer on the -OH surface while depicting the maximum dissipation (16.48 × 10⁶) at 1000 mM of salt concentration (Fig. 5b). Other surfaces also showed an increase in their dissipation when salt concentration increased from 100 mM to 1000 mM. This increase in dissipation might be raised by two phenomena; an increase in the adsorption of L-S on the surfaces and adaptation of an extended configuration in salty systems (Fig. S32[†]), which might be due to screening of the intramolecular interactions in the L-S.⁸⁷ Increasing the salt concentration affected the hydrodynamic radius (*R_h*) of the polymer due to electrostatic shielding and attraction between sulfonate groups of L-S and cations available from salt. In this case, the *R_h* increment was observed to be more in L-S than in PVA-S (Fig. S35[†]). The addition of salt would increase the attraction between adjacent polymer cores by eliminating the electrostatic interaction and exposing the hydrophobic sites of the polymer. This would lead to more L-S particles to aggregate and thus enhance L-S adsorption onto surfaces. Similar behaviour was observed for the self-aggregation of CNC (cellulose nanocrystals) at 160 mM KCl.⁸⁸

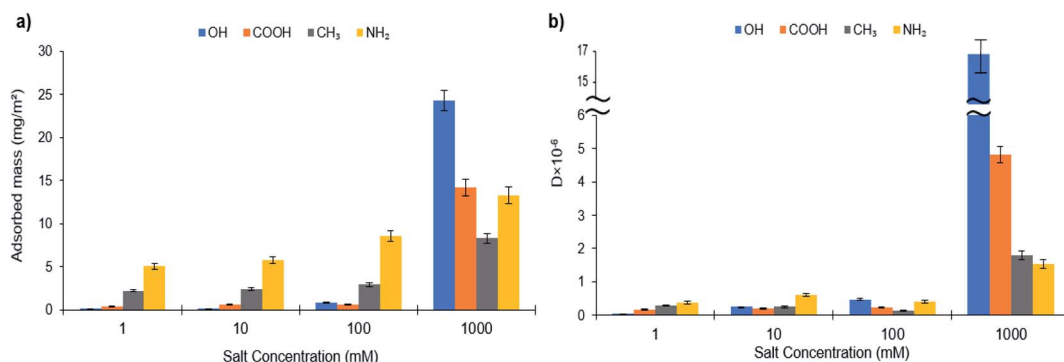


Fig. 5 (a) Adsorbed mass and (b) dissipation of L-S on SAMs of different surfaces at different salt concentrations.

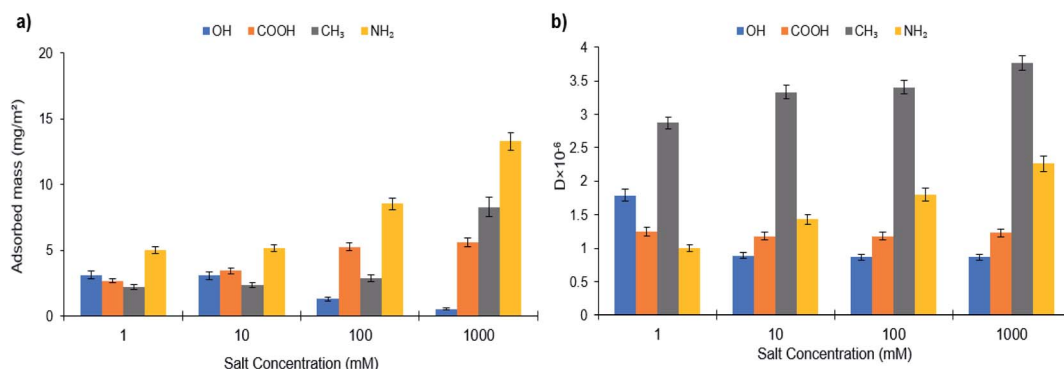


Fig. 6 (a) Adsorbed mass and (b) dissipation of PVA-S on SAMs of different chemistry at different salt concentrations.



Fig. 6 depicts the adsorbed mass and dissipation of PVA-S adsorption on different functionalized surfaces as a function of salt concentration. If electrostatic interactions between the polymer and oppositely charged substrates were dominant, there would be a cutoff of adsorption with increased ionic strength, above which the adsorption would be decreased. Normally, the cutoff is abrupt when there are only electrostatic interactions deriving the adsorption.^{86,89–91} The adsorption of PVA-S polymer decreased on the –OH surface, while it was increased on –COOH, –CH₃, and –NH₂ modified surfaces (Fig. 6a). Compared to the results obtained from different surfaces, having constant adsorption indicates that non-electrostatic forces dominated (rather than electrostatic interactions) when the polymer was not ionized.²³ Also, a decrease in the adsorption of PVA-S on the –OH surface in the saline system has been reported in other studies.^{23,92} The fast adsorption kinetics (Fig. S33†) seen on positively charged NH₂ terminated thiolates were observed to be unaffected by increasing the ionic strength (Fig. 6). This could be due to the electrostatic attraction force, which is always favorable and fast.^{88,92,93}

In contrast, the adsorption kinetics for liked-charged systems (PVA-S and L-S on –COOH SAMs) was slow while a late saturation was attained (Fig. S32 and S33†). Salt reduced the kinetic barrier between like-charged polymers and surfaces, further favoring the adsorption.^{21,23,94} The noticeable change in the adsorbed mass of the –CH₃ surface indicates that the hydrophobic interaction developed between the acetate group of the PVA-S and the surface, while all charges on the polymer were screened. The adsorbed mass was increased marginally for PVA-S on amine-rich monolayers by enhancing the ionic strength, which could be related to the higher adsorption of counterions from the solution, suppressing the polymer adsorption to this surface (Fig. 6).

Also, as mentioned earlier, the R_h of L-S and PVA-S increased as a result of an augment in salinity (Fig. S35†). This might be because increasing ionic strength causes the inter and intra-chain repulsion to develop a looser manner and expand polymer chains. These changes in the conformation of polymers lead to more sights become available for the interaction, which might have contributed to the increased adsorption of these polymers on most surfaces (Fig. 5 and 6). Polymerized kraft lignin-acrylic acid (KL-AA) with a molecular weight of $7.4 \times 10^5 \text{ g mol}^{-1}$ was reported to have the R_h value of 25.2 nm in salt-free solution.⁹⁵ Moreover, the same behavior has been reported in the literature for poly(potassium-2-sulfopropylmethacrylate) when salt was added.⁹⁶

In salinity, the highest adsorption amount for L-S polymer (24.32 mg m^{-2}) was attained at 1000 mM of salt concentration on the –OH functionalized surface. Meanwhile, the maximum adsorption of PVA-S (13.3 mg m^{-2}) was achieved at 1000 mM on the –NH₂ surface.

Fig. 6b shows the dissipation changes on different alkanethiol surfaces with PVA-S adsorption concerning the ionic strength. It can be seen that the reduction in dissipation could be due to the decrease in the adsorption amount. In the case of –COOH, although the adsorbed mass was increased by the salt addition, the reduction in the dissipation could be due to PVA-S

adopting a more compact structure, which was generated from nonelectrostatic forces in saline systems. In the case of –OH, –CH₃ and NH₂ surfaces, the dissipation value was changed proportionally to the adsorption of PVA-S on these surfaces.

3.7 Application

OH-coated surfaces interact with other materials, such as organic dyes polyesters,⁹⁷ fatty acids, and alkyl resins.⁹⁸ These surfaces are generally used in wastewater treatments, polymer blends, attachment of antibodies, and crystal engineering.^{9,99} The adsorption of L-S was less significant than PVA-S. This may make PVA-S a more effective coating material. Considering the temperature, salt, and pH variations, the results suggest that increasing the temperature, salt, and pH have an adverse impact on the adsorption of PVA-S onto the OH-functionalized surface.

For COOH coated surfaces that interact with other materials, such as proteins, surfactants, antigen detection,^{100,101} PVA-S showed to be a more effective adsorbent than L-S, but its effectiveness was significantly affected by pH and temperature (Fig. 3 and 4).

For CH₃ coated surfaces that interact with other materials, such as surfactants or surface-active agents,¹⁰² and have applications such as self-cleaning, anti-fogging, and anti-corrosion,¹⁰³ PVA-S showed to be a better candidate. The temperature analysis showed the limited impact of PVA-S adsorption onto this surface. Also, the pH analysis confirmed that at acidic pH, the hydrophobic interaction of both PVA-S and L-S polymers with the surface is more significant. The salinity analysis confirmed that a higher salt concentration (1000 mM) could increase the adsorption for both PVA-S and L-S polymers.

In NH₂ coated surfaces that interact with other materials, such as heavy metal, and have applications as flocculants, adsorbents, and dispersants, PVA-S observed to be a better candidate. The temperature analysis showed the limited impact of temperature variation on PVA-S adsorption and adverse effect on L-S adsorption. Also, the pH analysis confirmed that alkaline pH has a negative impact on the adsorption for both PVA-S and L-S polymers. The salt analysis confirmed that increasing the salt increases the adsorption for both PVA-S and L-S polymers.

Also, the contact angle analysis confirmed that PVA-S is more hydrophilic than the L-S polymer. Based on the second virial coefficient (A_2) obtained from static light scattering, increasing the temperature adversely impacted the water solubility of PVA-S, while increasing that of L-S. Based on the hydrodynamic radius obtained from dynamic light scattering, pH enhancement has a more intense impact on the L-S polymer than PVA-S. On the other hand, PVA-S showed sensitivity to temperature, pH, and salt. Although the L-S adsorption was more limited, it was insensitive to temperature. Therefore, L-S could be used in applications that are sensitive to temperature but do not require a high level of adsorption. Also, the adsorption of L-S showed improvement by increasing the salt concentration. This demonstrates the affinity of lignin in developing different interaction mechanisms with different functionalized surfaces over PVA-S in saline systems. Thus, the contribution of



nonelectrostatic forces seems to be more significant in the L-S polymer, illustrating practically an irreversible adsorption onto all surfaces, and its adsorption to the surfaces even when the electrostatic barrier exists. Therefore, it could be revealed that in applications that contain salts, *e.g.* wastewater treatment,¹⁵ additives for composites,¹⁰⁴ lignin could be a better candidate to be used than PVA-S.

3.8 Future trend

Understanding the interaction of sulfonated lignin and PVA would help advance scenarios to improve the properties of lignin for generating super functional lignin derivatives. The analysis in this paper reveals that the inherent steric hindrance of lignin may need to be reduced, *via* depolymerization, oxidation, for instance, for elevating its adsorption on surfaces. Furthermore, raised from its folded and compact molecular structure, most of the functional groups on lignin were not exposed but need to be activated. Alternatively, lignin can be decorated with more functional groups to boost its interaction with surfaces. For example, a combination of sulfonation, carboxylation, and polymerization may induce lignin with desired properties.

4. Conclusions

There has not been any systematic study in the literature on evaluating the performance of synthetic and lignin-based polymers having a similar charge density and molecular weight on well-defined self-assembled monolayers. In this study, we synthesized two types of anionic polymers of lignin-3-sulfopropyl methacrylate (L-S), and poly(vinyl alcohol-co-vinyl acetate)-3-sulfopropyl methacrylate (PVA-S) with similar charge densities and molecular weights. The R_h and R_g of PVA-S were smaller than those of L-S, while PVA-S had a larger A_2 . The A_2 of PVA-S decreased with augment in the temperature whereas it enhanced for L-S, which was in agreement with the more compact structure of PVA-S. On the OH surface, PVA-S was adsorbed more than L-S, which could be due to its limited hydrogen bonding. Also, temperature, salt, and pH variations adversely affected the adsorption of PVA-S onto this surface. Although PVA-S was a more effective adsorbent on the COOH coated surface due to its higher solubility, pH and temperature were observed to remarkably impact its adsorption performance. The adsorption of both polymers was also increased with salinity enhancement on the COOH-surface. For the CH₃ coated surface, although the PVA-S was observed to be a better candidate than L-S, increasing the temperature enhanced the L-S adsorption onto this surface due to more exposure of its hydrophobic parts. Interestingly, under acidic conditions, the hydrophobic interaction of both PVA-S and L-S polymers with the surface was increased. PVA-S also adsorbed more onto the NH₂ coated surface than L-S due to its solubility and polymer structure. The temperature was more significantly affected the L-S adsorption onto this surface than did PVA-S, while the similar negative impact was observed for the adsorption of both polymers onto this surface at higher pH. Also, in saline systems,

L-S adsorption was improved more significantly compared to PVA-S. Overall, the maximum adsorption of L-S (24.32 mg m⁻²) and PVA-S (21.33 mg m⁻²) polymers were observed on -OH functionalized surface at 1000 mM salt and on -CH₃ functionalized surface at pH 3.0, respectively.

Conflicts of interest

No competing financial interests are declared by authors.

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References

- 1 E. R. Van der Hage, M. M. Mulder and J. J. Boon, Structural characterization of lignin polymers by temperature-resolved in-source pyrolysis-mass spectrometry and Curie-point pyrolysis-gas chromatography/mass spectrometry, *J. Anal. Appl. Pyrolysis*, 1993, **25**, 149–183.
- 2 F. Wang, P. Chandler, R. Oszust, E. Sowell, Z. Graham, W. Ardito and X. Hu, Thermal and structural analysis of silk-polyvinyl acetate blends, *J. Therm. Anal. Calorim.*, 2017, **127**, 923–929.
- 3 S. Jie, Y. Li, B. Jie, Z. Chu-Shu, L. Shao-Fang, Z. Feng and Y. Qing-Li, Peanut protein-polyvinyl alcohol composite fibers extruded from an ionic liquid, *RSC Adv.*, 2013, **3**, 10619–10622.
- 4 A. Veerabhadraiah, S. Ramakrishna, G. Angadi, M. Venkatram, V. K. Ananthapadmanabha, K. N. M. H. NarayanaRao and K. Munishamaiah, Development of polyvinyl acetate thin films by electrospinning for sensor applications, *Appl. Nanosci.*, 2017, **7**, 355–363.
- 5 J. A. Gidigbi, S. A. Osemeahon, A. M. Ngoshe and A. Babanyaya, Modification of Polyvinyl Acetate with Hydroxylated Avocado Seed Oil as a Copolymer Binder for Possible Application in Coating Industry, *International Journal of Recent Innovations in Academic Research*, 2019, **3**, 231–244.
- 6 T. Moritani and J. Yamauchi, Functional modification of poly(vinyl alcohol) by copolymerization III. Modification with cationic monomers, *Polymer*, 1998, **39**, 559–572.
- 7 S. Ur-Rehman, M. Noman, A. D. Khan, A. Saboor, M. S. Ahmad and H. U. Khan, Synthesis of polyvinyl acetate/graphene nanocomposite and its application as an electrolyte in dye sensitized solar cells, *Optik*, 2020, **202**, 163591.
- 8 T. Moritani and J. Yamauchi, Functional modification of poly(vinyl alcohol) by copolymerization: II. Modification with a sulfonate monomer, *Polymer*, 1998, **39**, 553–557.



- 9 A. E. Kazzaz, Z. H. Feizi and P. Fatehi, Grafting strategies for hydroxy groups of lignin for producing materials, *Green Chem.*, 2019, **21**, 5714–5752.
- 10 M. Moradipour, E. K. Chase, M. A. Khan, S. O. Asare, B. C. Lynn, S. E. Rankin and B. L. Knutson, Interaction of lignin-derived dimer and eugenol-functionalized silica nanoparticles with supported lipid bilayers, *Colloids Surf., B*, 2020, 111028.
- 11 X. Luo, A. Mohanty and M. Misra, Lignin as a reactive reinforcing filler for water-blown rigid biofoam composites from soy oil-based polyurethane, *Ind. Crops Prod.*, 2013, **47**, 13–19.
- 12 Z. H. Feizi, A. E. Kazzaz, F. Kong and P. Fatehi, Evolving a flocculation process for isolating lignosulfonate from solution, *Sep. Purif. Technol.*, 2019, **222**, 254–263.
- 13 A. E. Kazzaz and P. Fatehi, Technical lignin and its potential modification routes: a mini-review, *Ind. Crops Prod.*, 2020, **154**, 112732.
- 14 A. E. Kazzaz, Z. H. Feizi and P. Fatehi, Interaction of sulfomethylated lignin and aluminum oxide, *Colloid Polym. Sci.*, 2018, **296**, 1867–1878.
- 15 J. Chen, A. E. Kazzaz, N. AlipoorMazandarani, Z. H. Feizi and P. Fatehi, Production of flocculants, adsorbents, and dispersants from lignin, *Molecules*, 2018, **23**, 868.
- 16 R. Vanholme, B. Demedts, K. Morreel, J. Ralph and W. Boerjan, Lignin biosynthesis and structure, *Plant Physiol.*, 2010, **153**, 895–905.
- 17 T. M. Garver and P. T. Callaghan, Hydrodynamics of kraft lignins, *Macromolecules*, 1991, **24**, 420–430.
- 18 S. L. Clark and P. T. Hammond, The role of secondary interactions in selective electrostatic multilayer deposition, *Langmuir*, 2000, **16**, 10206–10214.
- 19 X. Jiang, C. Ortiz and P. T. Hammond, Exploring the rules for selective deposition: interactions of model polyamines on acid and oligoethylene oxide surfaces, *Langmuir*, 2002, **18**, 1131–1143.
- 20 C. Friedsam, A. D. C. Bécares, U. Jonas, M. Seitz and H. E. Gaub, Adsorption of polyacrylic acid on self-assembled monolayers investigated by single-molecule force spectroscopy, *New J. Phys.*, 2004, **6**, 9.
- 21 P. Maroni, F. J. M. Ruiz-Cabello and A. Tiraferri, Studying the role of surface chemistry on polyelectrolyte adsorption using gold–thiol self-assembled monolayer with optical reflectivity, *Soft Matter*, 2014, **10**, 9220–9225.
- 22 H. Eto, N. Soga, H. G. Franquelim, P. Glock, A. Khmelinskaia, L. Kai and P. Schuille, Design of sealable custom-shaped cell mimics based on self-assembled monolayers on CYTOP polymer, *ACS Appl. Mater. Interfaces*, 2019, **11**, 21372–21380.
- 23 P. Maroni, F. J. Montes Ruiz-Cabello, C. Cardoso and A. Tiraferri, Adsorbed mass of polymers on self-assembled monolayers: effect of surface chemistry and polymer charge, *Langmuir*, 2015, **31**, 6045–6054.
- 24 Y. Song, J. Park, C. Lim and D. W. Lee, In-Depth Study of the Interaction Mechanism between the Lignin Nanofilms: Toward a Renewable and Organic Solvent-Free Binder, *ACS Sustainable Chem. Eng.*, 2020, **8**, 362–371.
- 25 A. E. Kazzaz and P. Fatehi, Fabrication of Amphoteric Lignin and its Hydrophilicity/Oleophilicity at Oil/Water Interface, *J. Colloid Interface Sci.*, 2020, **561**, 231–243.
- 26 J. J. Gooding, F. Mearns, W. Yang and J. Liu, Self-assembled monolayers into the 21st century: recent advances and applications, *Electroanalysis*, 2003, **15**, 81–96.
- 27 S. Huang, Q. Hou, D. Guo, H. Yang, T. Chen, F. Liu and J. Wang, Adsorption mechanism of mussel-derived adhesive proteins onto various self-assembled monolayers, *RSC Adv.*, 2017, **7**, 39530–39538.
- 28 C. K. Haweel and S. H. Ammar, Preparation of polyvinyl alcohol from local raw material, *J. Chem. Pet. Eng.*, 2008, **9**, 15–21.
- 29 S. Contreras, A. R. Gaspar, A. Guerra, L. A. Lucia and D. S. Argyropoulos, Propensity of lignin to associate: light scattering photometry study with native lignins, *Biomacromolecules*, 2008, **9**, 3362–3369.
- 30 S. M. Munzert, G. Schwarz and D. G. Kurth, Tailoring length and viscosity of dynamic metallo-supramolecular polymers in solution, *RSC Adv.*, 2016, **6**, 15441–15450.
- 31 F. C. Giacomelli, I. C. Riegel, C. L. Petzhold, N. P. da Silveira and P. Štěpánek, Aggregation behavior of a new series of ABA triblock copolymers bearing short outer A blocks in B-selective solvent: from free chains to bridged micelles, *Langmuir*, 2008, **25**, 731–738.
- 32 P. Dimitrov, M. Jamróz-Piegza, B. Trzebicka and A. Dworak, The influence of hydrophobic substitution on self-association of poly(ethylene oxide)-b-poly(n-alkyl glycidyl carbamate) sb-poly(ethylene oxide) triblock copolymers in aqueous media, *Polymer*, 2007, **48**, 1866–1874.
- 33 H. Hussain, B. H. Tan, G. L. Seah, Y. Liu, C. B. He and T. P. Davis, Micelle formation and gelation of (PEG-P (MA-POSS)) amphiphilic block copolymers via associative hydrophobic effects, *Langmuir*, 2010, **26**, 11763–11773.
- 34 C. L. Berhaut, D. Lemordant, P. Porion, L. Timperman, G. Schmidt and M. Anouti, Ionic association analysis of LiTDI, LiFSI and LiPF₆ in EC/DMC for better Li-ion battery performances, *RSC Adv.*, 2019, **9**, 4599–4608.
- 35 A. Ishikubo, J. Mays and M. Tirrell, Behavior of cationic surfactants in poly(styrene sulfonate) brushes, *Ind. Eng. Chem. Res.*, 2008, **47**, 6426–6433.
- 36 Z. H. Feizi and P. Fatehi, Carboxymethylated cellulose nanocrystals as clay suspension dispersants: effect of size and surface functional groups, *Cellulose*, 2020, **27**, 3759–3772.
- 37 M. K. Konduri, F. Kong and P. Fatehi, Production of carboxymethylated lignin and its application as a dispersant, *Eur. Polym. J.*, 2015, **70**, 371–383.
- 38 A. E. Kazzaz, Z. H. Feizi, F. Kong and P. Fatehi, Interaction of poly(acrylic acid) and aluminum oxide particles in suspension: particle size effect, *Colloids Surf., A*, 2018, **556**, 218–226.
- 39 C. J. Huang, F. S. Shieu, W. P. Hsieh and T. C. Chang, Acidic hydrolysis of a poly(vinyl acetate) matrix by the catalytic effect of Ag nanoparticles and the micellization of Ag-metal-containing polymer, *J. Appl. Polym. Sci.*, 2006, **100**, 1457–1464.



- 40 P. Maes, Y. B. Monakhova, T. Kuballa, H. Reusch and D. W. Lachenmeier, Qualitative and quantitative control of carbonated cola beverages using ¹H NMR spectroscopy, *J. Agric. Food Chem.*, 2012, **60**, 2778–2784.
- 41 J. Hedin, J. E. Löfroth and M. Nydén, Adsorption behavior and cross-linking of EHEC and HM-EHEC at hydrophilic and hydrophobic modified surfaces monitored by SPR and QCM-D, *Langmuir*, 2007, **23**, 6148–6155.
- 42 H. Wang, S. Chen, L. Li and S. Jiang, Improved method for the preparation of carboxylic acid and amine terminated self-assembled monolayers of alkanethiolates, *Langmuir*, 2005, **21**, 2633–2636.
- 43 M. L. Wallwork, D. A. Smith, J. Zhang, J. Kirkham and C. Robinson, Complex chemical force titration behavior of amine-terminated self-assembled monolayers, *Langmuir*, 2001, **17**, 1126–1131.
- 44 L. Herrer, V. Sebastian, S. Martín, A. González-Orive, F. Pérez-Murano, P. J. Low and P. Cea, High surface coverage of a self-assembled monolayer by in situ synthesis of palladium nanodeposits, *Nanoscale*, 2017, **9**, 13281–13290.
- 45 C. D. Bain and G. M. Whitesides, Attenuation lengths of photoelectrons in hydrocarbon films, *J. Phys. Chem.*, 1989, **93**, 1670–1673.
- 46 Q. Li, R. Yuan and Y. Li, Study on the molecular behavior of hydrophobically modified poly(acrylic acid) in aqueous solution and its emulsion-stabilizing capacity, *J. Appl. Polym. Sci.*, 2013, **128**, 206–215.
- 47 E. Pensini, C. M. Yip, D. O'Carroll and B. E. Sleep, Carboxymethyl cellulose binding to mineral substrates: characterization by atomic force microscopy-based Force spectroscopy and quartz-crystal microbalance with dissipation monitoring, *J. Colloid Interface Sci.*, 2013, **402**, 58–67.
- 48 G. Masci, D. Bontempo, N. Tiso, M. Diociaiuti, L. Mannina, D. Capitani and V. Crescenzi, Atom transfer radical polymerization of potassium 3-sulfopropyl methacrylate: direct synthesis of amphiphilic block copolymers with methyl methacrylate, *Macromolecules*, 2004, **37**, 4464–4473.
- 49 Y. Xu, A. Walther and A. H. Müller, Direct synthesis of poly(potassium 3-sulfopropyl methacrylate) cylindrical polymer brushes via ATRP using a supramolecular complex with crown ether, *Macromol. Rapid Commun.*, 2010, **31**, 1462–1466.
- 50 I. Korbag and S. Mohamed Saleh, Studies on the formation of intermolecular interactions and structural characterization of polyvinyl alcohol/lignin film, *Int. J. Environ. Stud.*, 2016, **73**, 226–235.
- 51 H. Lee, M. F. Rubner and R. E. Cohen, Coating, *US Pat.*, 13215869, 2012.
- 52 R. Hajiraissi, M. Hanke, Y. Yang, B. Duderija, A. Gonzalez Orive, G. Grundmeier and A. Keller, Adsorption and fibrillization of islet amyloid polypeptide at self-assembled monolayers studied by QCM-D, AFM, and PM-IRRAS, *Langmuir*, 2018, **34**, 3517–3524.
- 53 H. T. Phan, S. Bartelt-Hunt, K. B. Rodenhausen, M. Schubert and J. C. Bartz, Investigation of bovine serum albumin (BSA) attachment onto self-assembled monolayers (SAMs) using combinatorial quartz crystal microbalance with dissipation (QCM-D) and spectroscopic ellipsometry (SE), *PLoS One*, 2015, **10**, e0141282.
- 54 J. A. Jones, L. A. Qin, H. Meyerson, I. K. Kwon, T. Matsuda and J. M. Anderson, Instability of self-assembled monolayers as a model material system for macrophage/FBGC cellular behavior, *J. Biomed. Mater. Res., Part A*, 2008, **86**, 261–268.
- 55 J. Li, T. Guan, C. Hao, L. Li and Y. Zhang, Effects of self-assembled monolayers with different chemical groups on ovarian cancer cell line behavior in vitro, *J. Chem.*, 2015, **10**, 784626.
- 56 S. D. Evans, R. Sharma and A. Ulman, Contact angle stability: reorganization of monolayer surfaces?, *Langmuir*, 1991, **7**, 156–161.
- 57 K. S. Chong, S. Sun and G. J. Leggett, Measurement of the kinetics of photo-oxidation of self-assembled monolayers using friction force microscopy, *Langmuir*, 2005, **21**, 3903–3909.
- 58 J. C. Love, L. A. Estroff, J. K. Kriebel, R. G. Nuzzo and G. M. Whitesides, Self-assembled monolayers of thiolates on metals as a form of nanotechnology, *Chem. Rev.*, 2005, **105**, 1103–1170.
- 59 H. Hussain, K. Busse and J. Kressler, Poly(ethylene oxide)- and Poly(perfluorohexylethyl methacrylate)-Containing Amphiphilic Block Copolymers: Association Properties in Aqueous Solution, *Macromol. Chem. Phys.*, 2003, **204**, 936–946.
- 60 C. Booth and D. Attwood, Effects of block architecture and composition on the association properties of poly(oxyalkylene) copolymers in aqueous solution, *Macromol. Rapid Commun.*, 2000, **21**, 501–527.
- 61 Y. Kawata, S. Kozuka and S. I. Yusa, Thermo-responsive behavior of amphoteric diblock copolymers bearing sulfonate and quaternary amino pendant groups, *Langmuir*, 2018, **35**, 1458–1464.
- 62 K. Huber, S. Bantle, P. Lutz and W. Burchard, Hydrodynamic and thermodynamic behavior of short-chain polystyrene in toluene and cyclohexane at 34.5 degree. C, *Macromolecules*, 1985, **18**, 1461–1467.
- 63 T. Konishi, T. Yoshizaki and H. Yamakawa, On the “Universal Constants” ρ and Φ of flexible polymers, *Macromolecules*, 1991, **24**, 5614–5622.
- 64 H. Hussain, K. Y. Mya and C. He, Self-assembly of brush-like poly [poly(ethylene glycol) methyl ether methacrylate] synthesized via aqueous atom transfer radical polymerization, *Langmuir*, 2008, **24**, 13279–13286.
- 65 J. H. Yao, K. Y. Mya, X. Li, M. Parameswaran, Q. H. Xu, K. P. Loh and Z. K. Chen, Light scattering and luminescence studies on self-aggregation behavior of amphiphilic copolymer micelles, *J. Phys. Chem. B*, 2008, **112**, 749–755.
- 66 H. Hussain, B. H. Tan, C. S. Gudipati, Y. Liu, C. B. He and T. P. Davis, Synthesis and self-assembly of poly(styrene)-b-poly(N-vinylpyrrolidone) amphiphilic diblock copolymers



- made via a combined ATRP and MADIX approach, *J. Polym. Sci., Part A: Polym. Chem.*, 2008, **46**, 5604–5615.
- 67 T. Liu, Z. Zhou, C. Wu, B. Chu, D. K. Schneider and V. M. Nace, Self-Assembly of Poly(oxybutylene)-Poly(oxyethylene)-Poly(oxybutylene)(B₆E₄₆B₆) Triblock Copolymer in Aqueous Solution, *J. Phys. Chem. B*, 1997, **101**, 8808–8815.
- 68 B. Jeong, Y. H. Bae and S. W. Kim, Biodegradable thermosensitive micelles of PEG-PLGA-PEG triblock copolymers, *Colloids Surf., B*, 1999, **16**, 185–193.
- 69 E. Castro, S. Barbosa, J. Juarez, P. Taboada, I. A. Katime and V. Mosquera, Influence of external factors on the micellization process and aggregate structure of poly(oxy)styrene-poly(oxy)ethylene block copolymers, *J. Phys. Chem. B*, 2008, **112**, 5296–5304.
- 70 A. A. Tager, A. A. Anikeyeva, L. V. Adamova, V. M. Andreyeva, T. A. Kuz'mina and M. V. Tsilipotkina, The effect of temperature on the water solubility of polyvinyl alcohol, *Polym. Sci.*, 1971, **13**, 751–758.
- 71 V. T. Matsuo and H. Inagaki, Über den lösungszustand des polyvinylalkohols in wasser I. Mitt.: metastabiler zustand der lösung, *Macromol. Chem. Phys.*, 1962, **53**, 130–144.
- 72 J. Liu, C. Detrembleur, M. Hurtgen, A. Debuigne, M. C. De Pauw-Gillet, S. Mornet and C. Jérôme, Thermo-responsive gold/poly(vinyl alcohol)-b-poly(N-vinylcaprolactam) core-corona nanoparticles as a drug delivery system, *Polym. Chem.*, 2014, **5**, 5289–5299.
- 73 N. A. Cortez-Lemus and A. Licea-Claverie, Preparation of a mini-library of thermo-responsive star (NVCL/NVP-VAC) polymers with tailored properties using a hexafunctional xanthate RAFT agent, *Polymers*, 2018, **10**, 20.
- 74 J. P. Inwood, L. Pakzad and P. Fatehi, Production of sulfur containing kraft lignin products, *BioResources*, 2018, **13**, 53–70.
- 75 A. N. Evdokimov, A. V. Kurzin, O. V. Fedorova, P. V. Lukanin, V. G. Kazakov and A. D. Trifonova, Desulfurization of kraft lignin, *Wood Sci. Technol.*, 2018, **52**, 1165–1174.
- 76 M. Norgren and B. Lindström, Dissociation of phenolic groups in kraft lignin at elevated temperatures, *Holzforchung*, 2000, **54**, 519–527.
- 77 W. Zhu and H. Theliander, Precipitation of lignin from softwood black liquor: an investigation of the equilibrium and molecular properties of lignin, *BioResources*, 2015, **10**, 1696–1714.
- 78 R. Ou, H. Zhang, J. Wei, S. Kim, L. Wan, N. S. Nguyen and H. Wang, Thermoresponsive amphoteric metal-organic frameworks for efficient and reversible adsorption of multiple salts from water, *Adv. Mater.*, 2018, **30**, 1802767.
- 79 R. K. Sharma, J. B. Wooten, V. L. Baliga, X. Lin, W. G. Chan and M. R. Hajaligol, Characterization of chars from pyrolysis of lignin, *Fuel*, 2004, **83**, 1469–1482.
- 80 A. A. M. Nada, M. A. Yousef, K. A. Shaffei and A. M. Salah, Infrared spectroscopy of some treated lignins, *Polym. Degrad. Stab.*, 1998, **62**, 157–163.
- 81 A. D. Phan and T. X. Hoang, The pH-dependent electrostatic interaction of a metal nanoparticle with the MS2 virus-like particles, *Chem. Phys. Lett.*, 2019, **730**, 84–88.
- 82 X. Zhang, Z. Zhao, G. Ran, Y. Liu, S. Liu, B. Zhou and Z. Wang, Synthesis of lignin-modified silica nanoparticles from black liquor of rice straw pulping, *Powder Technol.*, 2013, **246**, 664–668.
- 83 I. Popa, B. P. Cahill, P. Maroni, G. Papastavrou and M. Borkovec, Thin adsorbed films of a strong cationic polyelectrolyte on silica substrates, *J. Colloid Interface Sci.*, 2007, **309**, 28–35.
- 84 M. Porus, P. Maroni and M. Borkovec, Structure of adsorbed polyelectrolyte monolayers investigated by combining optical reflectometry and piezoelectric techniques, *Langmuir*, 2012, **28**, 5642–5651.
- 85 D. Bauer, H. Buchhammer, A. Fuchs, W. Jaeger, E. Killmann, K. Lunkwitz, R. Rehmet and S. Schwarz, Stability of colloidal silica, sikron and polystyrene latex influenced by the adsorption of polycations of different charge density, *Colloids Surf., A*, 1999, **156**, 291–305.
- 86 I. Szilagyi, G. Trefalt, A. Tiraferri, P. Maroni and M. Borkovec, Polyelectrolyte adsorption, interparticle forces, and colloidal aggregation, *Soft Matter*, 2014, **10**, 2479–2502.
- 87 A. Sweity, W. Ying, M. S. Ali-Shtayeh, F. Yang, A. Bick, G. Oron and M. Herzberg, Relation between EPS adherence, viscoelastic properties, and MBR operation: Biofouling study with QCM-D, *Water Res.*, 2011, **45**, 6430–6440.
- 88 T. Phan-Xuan, A. Thuresson, M. Skepö, A. Labrador, R. Bordes and A. Matic, Aggregation behavior of aqueous cellulose nanocrystals: the effect of inorganic salts, *Cellulose*, 2016, **23**, 3653–3663.
- 89 M. Jiang, I. Popa, P. Maroni and M. Borkovec, Adsorption of poly(L-lysine) on silica probed by optical reflectometry, *Colloids Surf., A*, 2010, **360**, 20–25.
- 90 J. Forsman, Polyelectrolyte adsorption: electrostatic mechanisms and nonmonotonic responses to salt addition, *Langmuir*, 2012, **28**, 5138–5150.
- 91 F. Xie, T. Nylander, L. Piculell, S. Utsel, L. Wågberg, T. Åkesson and J. Forsman, Polyelectrolyte adsorption on solid surfaces: theoretical predictions and experimental measurements, *Langmuir*, 2013, **29**, 12421–12431.
- 92 S. J. De Carvalho, R. Metzler and A. G. Cherstvy, Critical adsorption of polyelectrolytes onto charged Janus nanospheres, *Phys. Chem. Chem. Phys.*, 2014, **16**, 15539–15550.
- 93 N. Hansupalak and M. M. Santore, Sharp polyelectrolyte adsorption cutoff induced by a monovalent salt, *Langmuir*, 2003, **19**, 7423–7426.
- 94 N. G. Hoogeveen, M. A. C. Stuart and G. J. Fleer, Polyelectrolyte adsorption on oxides: I. Kinetics and adsorbed amounts, *J. Colloid Interface Sci.*, 1996, **182**, 133–145.
- 95 F. Kong, S. Wang, W. Gao and P. Fatehi, Novel pathway to produce high molecular weight kraft lignin-acrylic acid



- polymers in acidic suspension systems, *RSC Adv.*, 2018, **8**, 12322–12336.
- 96 D. J. Liaw and C. C. Huang, Dilute solution properties of anionic poly(potassium-2-sulfopropylmethacrylate), *J. Appl. Polym. Sci.*, 1997, **63**, 175–185.
- 97 K. J. D. MacKenzie, Innovative applications of inorganic polymers (geopolymers), in *Handbook of Alkali-Activated Cements, Mortars and Concretes*, Woodhead Publ., 2015, pp. 777–805.
- 98 D. H. Solomon and J. J. Hopwood, Reactivity of functional groups in surface coating polymers. Part I. Hydroxyl groups in alkyd resins, *J. Appl. Polym. Sci.*, 1966, **10**, 981–991.
- 99 D. Andrews, G. Scholes and G. Wiederrecht, *Comprehensive nanoscience and technology*, Academic Press, 2010.
- 100 P. K. Ajikumar, J. K. Ng, Y. C. Tang, J. Y. Lee, G. Stephanopoulos and H. P. Too, Carboxyl-terminated dendrimer-coated bioactive interface for protein microarray: high-sensitivity detection of antigen in complex biological samples, *Langmuir*, 2007, **23**, 5670–5677.
- 101 M. Gasser, B. Rothen-Rutishauser, H. F. Krug, P. Gehr, M. Nelle, B. Yan and P. Wick, The adsorption of biomolecules to multi-walled carbon nanotubes is influenced by both pulmonary surfactant lipids and surface chemistry, *J. Nanobiotechnol.*, 2010, **8**, 31.
- 102 M. Jawaid, S. Boufi and A. Khalil H. P. S., *Cellulose-Reinforced nanofibre composites: Production, properties and applications*, Woodhead Publ., 2017.
- 103 M. K. Meena, B. K. Tudu, A. Kumar and B. Bhushan, Development of polyurethane-based superhydrophobic coatings on steel surfaces, *Philos. Trans. R. Soc., A*, 2020, **378**, 20190446.
- 104 C. Salas, O. J. Rojas, L. A. Lucia, M. A. Hubbe and J. Genzer, On the surface interactions of proteins with lignin, *ACS Appl. Mater. Interfaces*, 2013, **5**, 199–206.

