


Cite this: *RSC Adv.*, 2020, 10, 37449

Amino group dependent sensing properties of metal–organic frameworks: selective turn-on fluorescence detection of lysine and arginine†

Jing Dong,^{ab} Xiu-Du Zhang,^a Xia-Fei Xie,^a Fan Guo^a and Wei-Yin Sun^{id}*^a

Recently, metal–organic frameworks (MOFs) have been extensively investigated as fluorescence chemosensors due to their tunable porosity, framework structure and photoluminescence properties. In this paper, a well-known Zr(IV)-based MOF, UiO-66-NH₂ was demonstrated to have capability for detection of L-lysine (Lys) and L-arginine (Arg) selectively from common essential amino acids in aqueous media via a fluorescence turn-on mechanism. Further investigation reveals its high sensitivity and strong anti-interference properties. Moreover, the possible mechanism for sensing Lys and Arg was explored by FT-IR and ¹H-NMR, and the results indicate that the enhancement of the fluorescence could be ascribed to the adsorption of Lys/Arg and the hydrogen bonding interactions between Lys/Arg and the amino group of UiO-66-NH₂. The difference of the sensing capacity and sensitivity between UiO-66 and UiO-66-NH₂ revealed that the amino group plays an essential role in the sensing performance. This work presents a unique example of the functional group dependent sensing properties of MOFs.

Received 10th August 2020

Accepted 4th October 2020

DOI: 10.1039/d0ra06879a

rsc.li/rsc-advances

Introduction

As important biological molecules, amino acids (AAs) are an indispensable constituent of living systems, which are the basic substances that make up proteins as well as enzymes and exist widely in food, blood and tissues.¹ AAs are considered to have an inseparable relationship with the metabolism of proteins and peptides in a series of physiological processes.² Detection of definite AAs in food and organisms is particularly important in nutritional analysis and disease diagnosis.³ Among the amino acid family, lysine (Lys) is one of the essential AAs for human beings and mammals.⁴ Lys has a great influence on the Krebs–Henseleit cycle and polyamine synthesis, which is related to the metabolic function of human beings and animals.⁵ What is more, Lys cannot be synthesized by itself in the human body and must be supplemented from food. As a result, the content of Lys is usually proposed as an evaluation criterion to judge the nutritional value of food, and hence the detection of Lys has a positive nutritional significance.⁶ On the other hand, as another important natural amino acid, arginine (Arg) has been recognized as an essential amino acid for young/adult

mammals and birds. The increase of Arg metabolism can lead to the abnormality of human related system.⁷ Meanwhile, Arg has been demonstrated to be a biomarker of specific metabolic diseases, such as sulfite oxidase deficiency⁸ and has also been considered to be closely related to the pathogenesis of a number of diseases involving cardiovascular and renal.⁹ As a result, the detection of Arg is very helpful for drug analysis and medical diagnosis, which has great significance for further clinical diagnosis and treatment.^{8,10} Therefore, it is necessary to develop simple, sensitive and selective probe to distinguish Lys/Arg from other AAs. However, one of the biggest problem and challenge of biosensors for the detection of AAs is the stability and biocompatibility of the sensing materials in aqueous media.¹¹

Metal–organic frameworks (MOFs) are porous organic–inorganic hybrid materials formed by metal ions or metal clusters and organic bridging ligands through coordination interactions.¹² In the past several decades, they have been intensively investigated and successfully applied in varied areas including energy storage,¹³ heterogeneous catalysis,¹⁴ gas adsorption and separation,^{15,16} drug delivery¹⁷ and so on due to their structural diversity, controllability, high porosity and large surface area.^{18–20} In particular, the investigation employing MOFs as fluorescent materials to detect a variety of substances, for example, the hazardous gas molecules,²¹ volatile organic chemicals (VOCs),²² explosives²³ etc., has attracted great attention. On the one hand, the fluorescent properties of MOFs depend on the nature of the metal ions/clusters and organic ligands in the construction of MOFs and the regulation of the fluorescence behavior of MOFs could be realized through the

^aCoordination Chemistry Institute, State Key Laboratory of Coordination Chemistry, School of Chemistry and Chemical Engineering, Nanjing National Laboratory of Microstructures, Collaborative Innovation Center of Advanced Microstructures, Nanjing University, Nanjing 210023, China. E-mail: sunwy@nju.edu.cn

^bNanjing Tech University, Nanjing 211816, China

† Electronic supplementary information (ESI) available: PXRD, UV-Vis and additional emission spectra, comparison for lysine and arginine sensing. See DOI: 10.1039/d0ra06879a



rational selection and design.^{24,25} On the other hand, the pores and/or channels of MOFs could adsorb appropriate guest molecules and the interactions between the guest molecules and the metal nodes or organic ligands of MOFs may influence the fluorescence behavior to some extent.²⁶ In addition, quite a lot of MOFs are easy to be modified with different functional groups and thus high selectivity can be achieved by modification of specific functional groups.²⁷ Therefore, MOFs have been admired by researchers and gained popularity in the development of fluorescence sensing materials.

As a kind of Zr(IV)-based MOFs, the series of MOFs developed by Lillerud and coworkers^{28–31} including UiO-66, UiO-67 and UiO-68 (UiO = University of Oslo) have become one of the most famous and studied MOFs due to the high porosity, surface area, excellent stability and easy modification. In this paper, we studied the fluorescence properties of amino group-functionalized UiO-66, namely UiO-66-NH₂, and found that Lys and Arg could enhance its fluorescence emission intensity significantly in aqueous suspension, which realized the purpose of detecting Lys and Arg in aqueous media. Further experiments not only demonstrated its high sensing selectivity in various AAs, but also revealed that the emission intensity of UiO-66-NH₂ in aqueous suspension is in proportional to the concentration of Lys and Arg, which made it possible for the quantitatively detection of Lys and Arg. In addition, UiO-66-NH₂ also exhibits strong anti-interference in the detection of Lys and Arg. According to the FT-IR and ¹H-NMR spectral measurements, the fluorescence turn-on mechanism may be ascribed to the adsorption of Lys/Arg into the pores and the hydrogen bonding interactions between Lys/Arg and the amino groups in the framework of UiO-66-NH₂. Furthermore, the results of the sensing capacity and sensitivity of UiO-66 compared with that of UiO-66-NH₂ illustrate that the amino groups of UiO-66-NH₂ play an essential role in the sensing performance.

Experimental

Materials and methods

UiO-66-NH₂ and UiO-66 were synthesized following previously reported procedure with slight modification.^{28,32} All the chemicals and solvents were purchased commercially and used without further purification. FT-IR spectra were recorded in the range of 400–4000 cm^{−1} using a Bruker Vecotr22 FT-IR spectrophotometer with KBr pellets. Powder X-ray diffraction (PXRD) measurements were conducted using a Bruker D8 Advance X-ray diffractometer equipped with a Cu-K α radiation ($\lambda = 1.5418 \text{ \AA}$) source, in which the X-ray tube was operated at 40 kV and 40 mA. Fluorescence spectra were measured on a PerkinElmer LS-55 fluorescence spectrometer. ¹H NMR spectra were recorded on a Bruker-DRX 500 MHz instrument. UV-Vis spectral measurements were conducted at room temperature on a Shimadzu UV3600 spectrophotometer.

Fluorescence and sensing experiments

The solid-state fluorescence property of UiO-66-NH₂ was examined at room temperature. In order to explore the sensing

capacity for AAs, the as-synthesized samples of UiO-66-NH₂ were fully ground and then dispersed in deionized water or the aqueous solutions of different AAs (0.1 M) with ultrasonic treatment to prepare the steady suspension (0.5 mg mL^{−1}). The suspension was stirred at a constant rate during fluorescence measurements for homogeneity and the fluorescence emission spectra in the range of 370–680 nm were recorded at room temperature. Each experiment was repeated three times to obtain reliable data.

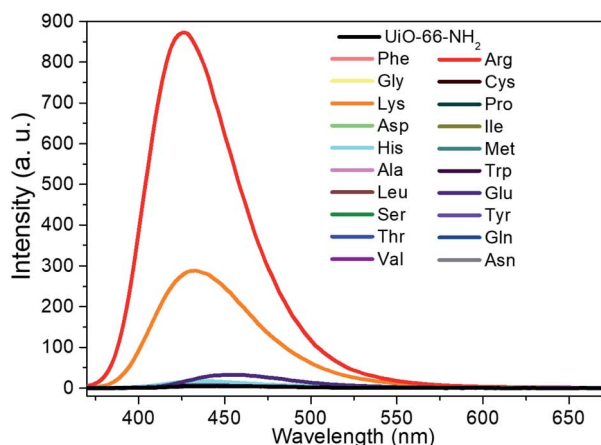
Quantitative titration and anti-interference experiments

In order to investigate the relationship between the emission enhancement ratio and the concentration of Lys/Arg, quantitative titration experiments were conducted by gradual addition of the Lys (0.1 M) or Arg (0.02 M) solution into the suspension of UiO-66-NH₂ (0.5 mg mL^{−1}) in deionized water and the fluorescent emission spectra were recorded every five minutes after the addition of Lys/Arg. Moreover, the anti-interference experiments were carried out by the addition in sequence of the solutions of other AAs and Lys/Arg (0.1 M) into the suspension of UiO-66-NH₂ (0.5 mg mL^{−1}) in water. All the experiments were repeated three times to achieve reliable data.

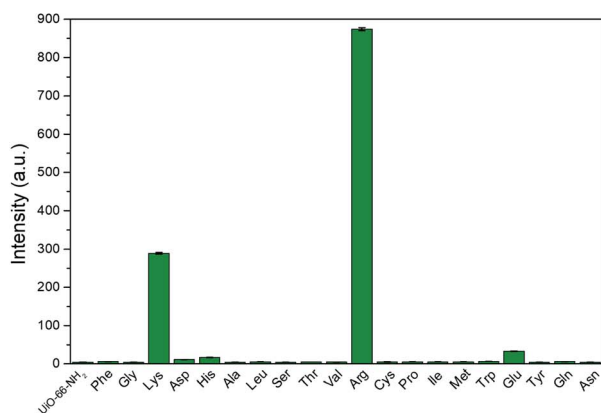
Results and discussion

The bulk-phase purity of the as-synthesized UiO-66-NH₂ was confirmed by using powder X-ray diffraction (PXRD) measurements. As depicted in Fig. S1 (ESI[†]), the peak positions of the obtained crystalline sample UiO-66-NH₂ matched well with the simulated ones from the single crystal X-ray diffraction data,³³ manifesting the phase purity of the synthesized samples. Then the solid-state fluorescence properties of the synthesized UiO-66-NH₂ were examined at room temperature. It can be observed in Fig. S2 (ESI[†]) that UiO-66-NH₂ exhibits characteristic emission band with maxima at 456 nm under the excitation wavelength of 375 nm. Furthermore, in order to investigate the sensing ability of UiO-66-NH₂ towards AAs in aqueous media, the as-synthesized samples of UiO-66-NH₂ were fully ground and dispersed in deionized water containing different AAs including L-phenylalanine (Phe), glycine (Gly), L-lysine (Lys), L-aspartic acid (Asp), L-histidine (His), L-alanine (Ala), L-leucine (Leu), L-serine (Ser), L-threonine (Thr), L-valine (Val), L-arginine (Arg), L-cysteine (Cys), L-proline (Pro), L-isoleucine (Ile), L-methionine (Met), L-tryptophan (Trp), L-glutamic acid (Glu), L-tyrosine (Tyr), L-glutamine (Gln) and L-asparagine (Asn) with concentration of 0.1 M to prepare the stable suspension (0.5 mg mL^{−1}) by ultrasonication, respectively. Then, the fluorescence emission spectra of these suspension were recorded in the range of 370–680 nm excited at 350 nm. Compared to the emission of the aqueous suspension of UiO-66-NH₂ without any AAs, increase in the emission intensities were observed with some shift in the maximum wavelength for all the suspensions with addition of AAs (Fig. 1a and S3, ESI[†]). As clearly shown in Fig. 1b, it is worth noting that only a slight emission increment could be monitored for the suspensions of most of the AAs but Lys and Arg could greatly enhance the emission intensity of





(a)

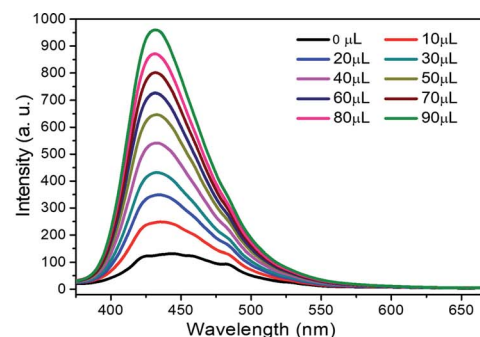


(b)

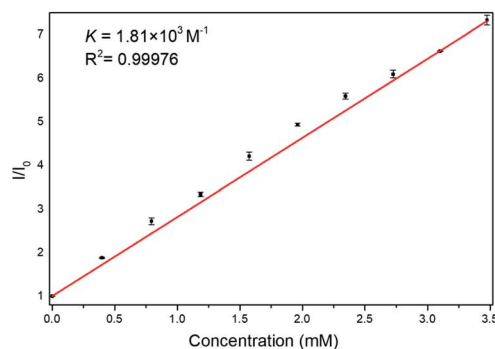
Fig. 1 (a) Fluorescence emission curves of the aqueous suspension of UiO-66-NH₂ (0.5 mg mL⁻¹) in the absence and presence of different AAs (0.1 M). (b) Maximum fluorescence intensities of the aqueous suspension of UiO-66-NH₂ in the absence and presence of different AAs (in consideration of the measuring range of emission intensity, the fluorescence spectra were obtained with an optical attenuator).

UiO-66-NH₂. Particularly, the presence of Arg could result in a greater than 181-fold increase in the emission intensity, which is much more significant than that of Lys (about 60 times). UV-Vis spectra of the suspension of UiO-66-NH₂ with various amino acids were measured and they show absorption between 250–400 nm (Fig. S4, ESI[†]).

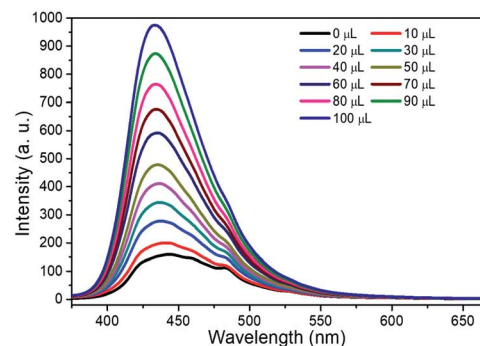
The great fluorescence increment behavior indicates that UiO-66-NH₂ is possible to be employed as a fluorescent sensor for detection of Lys and Arg. In order to explore the detection sensitivity of UiO-66-NH₂ towards Lys and Arg, quantitative fluorescence titration experiments upon the gradual addition of the aqueous solution of Lys (0.1 M) and Arg (0.02 M) to the aqueous suspension of UiO-66-NH₂ were carried out, respectively. As depicted in Fig. 2a and c, the emission intensities of the suspension of UiO-66-NH₂ increase continuously with the addition of Lys and Arg. The plots of the emission enhancement ratio I/I_0 vs. the concentration of Lys and Arg revealed a linear relationship between them in the concentration range of 0–



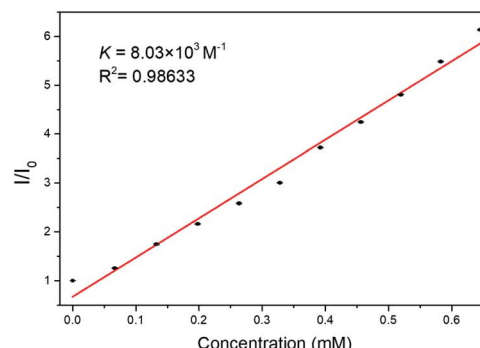
(a)



(b)



(c)



(d)

Fig. 2 The changes in the fluorescence emission of the aqueous suspension of UiO-66-NH₂ (0.5 mg mL⁻¹) with the gradual addition certain amount of Lys (0.1 M) (a) and Arg (0.02 M) (c). The plots of I/I_0 of the maximum intensities vs. the concentration of Lys (b) and Arg (d).

3.475 and 0–0.645 mM with the correlation coefficients (R^2) of 0.99976 (Fig. 2b) and 0.98633 (Fig. 2d) for Lys and Arg, respectively, where I_0 and I are the emission intensities of the aqueous suspension before and after the addition of the aqueous solution of Lys/Arg. Thus, the formula of $I/I_0 = K[A] + 1$ is used to fit the linear curves, where K is the slope, $[A]$ is the concentration of analyte. The slope (K) of the linear curves were estimated to be $1.81 \times 10^3 \text{ M}^{-1}$ for Lys and $8.03 \times 10^3 \text{ M}^{-1}$ for Arg (Fig. 2b and d), which is higher or comparable to some reported materials (Tables S1 and S2, ESI†). According to the standard equation $3\delta/K$ for the calculation of the limit of detection (LOD), where δ is the standard deviation for replicating detections of blank suspensions, the LOD is calculated to be 60.22 and 21.50 μM for Lys and Arg, respectively. The results manifested that UiO-66-NH₂ has high sensing sensitivity towards Arg than Lys.

In addition to the AAs, the sensing ability of UiO-66-NH₂ towards some biogenic amines was also checked. As shown in Fig. S5 (ESI†), the addition of putrescine, cadaverine and spermidine had little influence on the fluorescence of UiO-66-NH₂, while the presence of spermine could strengthen the fluorescence of UiO-66-NH₂. Such phenomenon indicated that UiO-66-NH₂ may be able to detect spermine as well.

In practical application, there are usually different AAs existing simultaneously in the real sample. Therefore, it is of great significance to be capable to identify the target analyte in the presence of other competing species. In order to verify its anti-interference, competing experiments were conducted by gradual addition of the aqueous solution of other AAs followed by Lys/Arg. As shown in Fig. 3a, it was found that the addition of the aqueous solutions of other AAs (1 mL, 0.1 M) only cause slight fluorescence emission increment. However, significant increase in the emission intensities could be observed upon the addition of aqueous solution of Lys (1 mL, 0.1 M). Similar phenomenon was also observed for Arg, furthermore, the emission intensity of the aqueous suspension of UiO-66-NH₂ could still be enhanced obviously in the presence of Lys (Fig. 3b). Such phenomenon implied that UiO-66-NH₂ is capable to detect Lys and Arg in the presence of interfering AAs.

The effects of anions on the sensing performance of UiO-66-NH₂ towards Lys and Arg were investigated. As exhibited in Fig. S6 (ESI†), the presence of sodium salt of acetate, benzoate, salicylate, Cl[−], Br[−] had almost no influence on the sensing capability of UiO-66-NH₂ towards Lys and Arg, while the addition of ATPs could induce the loss of sensing capacity. As for phosphate and pyrophosphate, the addition of them could also cause the sharp increase in the emission intensities of the suspension of UiO-66-NH₂ as reported previously³⁴ and the following addition of Lys/Arg could not induce further enhancement of the emission intensities. Such phenomenon indicated that the presence of ATPs, phosphate and pyrophosphate could affect the sensing performance of UiO-66-NH₂ towards Lys and Arg.

The reusability of UiO-66-NH₂ for sensing Lys and Arg has been checked. After centrifuging the suspension and washing with deionized water several times, the sample of the treated UiO-66-NH₂ was prepared into the suspension again and the fluorescence return to the weak emission. The sensing

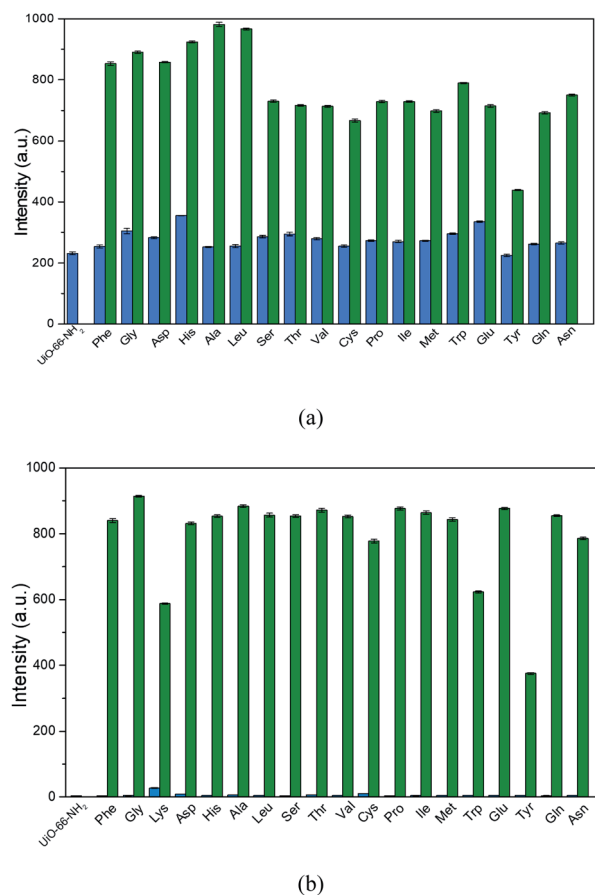


Fig. 3 The fluorescence intensities of the suspension of UiO-66-NH₂ upon the addition of competing AAs and Lys (a) and Arg (b) in sequence (in consideration of the measuring range of emission intensity, the fluorescence spectra of Arg were obtained with an optical attenuator).

experiments with the recovered samples revealed that the sensing capacity towards Lys without obvious changes, however, the sensing performance for Arg weakened (Fig. S7, ESI†), implying that UiO-66-NH₂ could be reused to detect Lys but not Arg.

In addition, we also conducted experiments to further understand the insight for the detection of Lys and Arg by UiO-66-NH₂. Firstly, the stability of UiO-66-NH₂ in aqueous solution of Lys and Arg were examined. As shown in Fig. 4, the correspondence between the PXRD patterns of the UiO-66-NH₂ before and after soaking in aqueous solution of Lys and Arg demonstrates the stability of the framework in the presence of Lys and Arg, suggesting that UiO-66-NH₂ could retain the framework structure, and accordingly the turn-on fluorescence caused by the collapse of the framework can be excluded. Afterwards, we speculated that the fluorescence enhancement may be originated by the adsorption of Lys or Arg into the pores of UiO-66-NH₂. Therefore, ¹H-NMR and FT-IR spectral measurements were carried out to examine this assumption. Prior to the ¹H NMR experiments, the samples of the untreated and Lys/Arg-treated UiO-66-NH₂ were digested in HF/DMSO-*d*₆. The ¹H-NMR spectra of Lys-treated UiO-66-NH₂ shows the



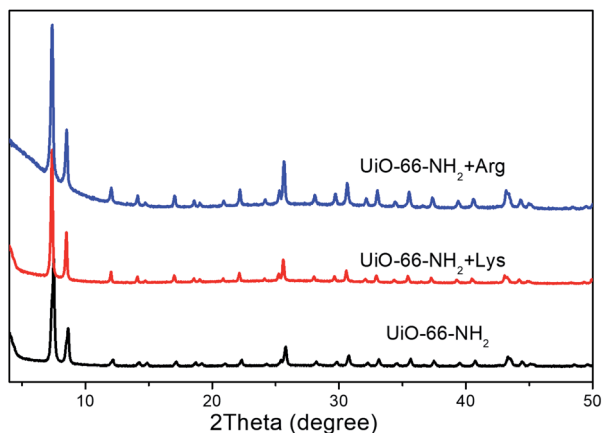
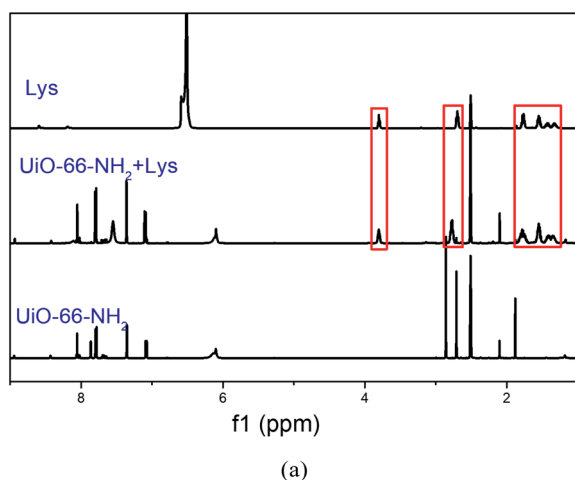
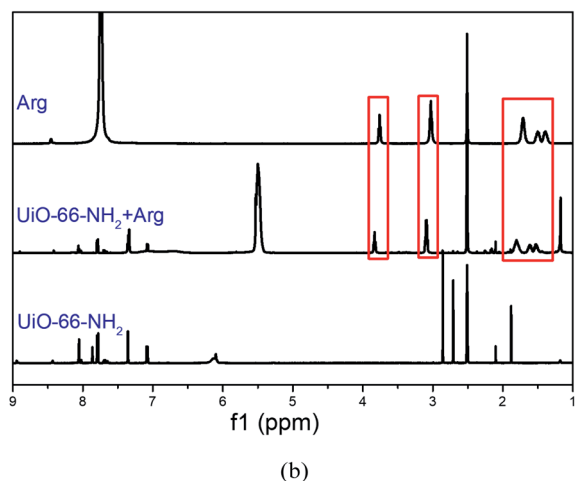


Fig. 4 PXRD patterns of UiO-66-NH₂ before and after soaking in aqueous solution of Lys/Arg.

appearance of new peaks at 1.28–1.48, 1.55, 1.70–1.85, 2.78, 3.80 ppm belonging to Lys (Fig. 5a and Table S3, ESI[†]), and also the new peaks at 1.38–1.69, 1.71–1.88, 3.09, 3.83 ppm belonging



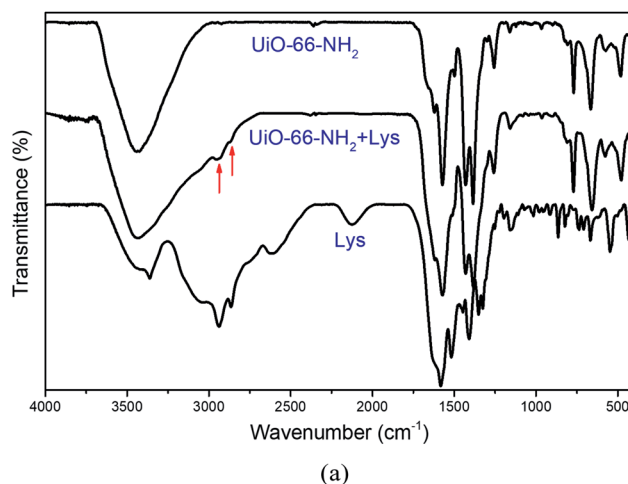
(a)



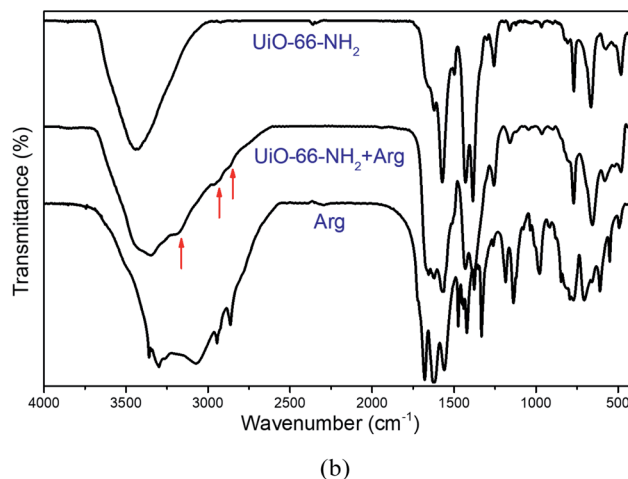
(b)

Fig. 5 ¹H-NMR spectra of Lys (a) and Arg (b) in DCI/DMSO-*d*₆, UiO-66-NH₂ digested in HF/DMSO-*d*₆ before and after the immersion in the solution of Lys (a) and Arg (b).

to Arg were observed in the ¹H-NMR spectra of Arg-treated UiO-66-NH₂ (Fig. 5b and Table S3, ESI[†]), which confirmed the adsorption of Lys and Arg into the pores of UiO-66-NH₂. The calculated molecular sizes of Lys and Arg are smaller than the pore size of UiO-66-NH₂ (Fig. S8, ESI[†]), which allow the adsorption. In addition, after incubation with the aqueous solution of Lys/Arg, the FT-IR spectra revealed the appearance of C–H stretching vibrations around 2952 and 2962 cm^{−1} for Lys and Arg respectively, and N–H stretching vibrations around 3178 cm^{−1} for Arg (Fig. 6a and b). Meanwhile, compared to the original FT-IR spectrum of UiO-66-NH₂, the shift of the peaks at 1621, 1499, 1256 cm^{−1} (Fig. 6a and b) could be observed, which not only demonstrated the adsorption of Lys/Arg into the pores of UiO-66-NH₂, but also implied the existence of hydrogen bonding interactions between Lys/Arg and UiO-66-NH₂.^{6,35} It has been reported that the weakened emission of UiO-66-NH₂, compared with the organic ligand due to the ligand-to-metal charge transfer (LMCT), could be recovered by addition of phosphate through the interactions between the analyte and



(a)



(b)

Fig. 6 (a) FT-IR spectra of Lys, UiO-66-NH₂ before and after the immersion in the aqueous solution of Lys. (b) FT-IR spectra of Arg, UiO-66-NH₂ before and after the immersion in the aqueous solution of Arg.

UiO-66-NH₂.³⁴ Furthermore, its sensing capacity towards Lys and Arg under different pH was examined as well and the results revealed that the sensing performance was weakened significantly at pH = 2 (Fig. S9, ESI†), which also suggested that the hydrogen bonding interactions between Lys/Arg and UiO-66-NH₂ play an important role in the detection of Lys/Arg. Compared to the other AAs, there are amino and guanidyl groups in the side chains of Lys and Arg, respectively, which may be responsible for the interactions of Lys/Arg with UiO-66-NH₂.

To further ensure the interactions between Lys/Arg and amino group in the UiO-66-NH₂ playing important role in detection of the analyte, we use the corresponding MOF without amino group functionalization, namely UiO-66, to recognize the AAs under the same conditions. As illustrated in Fig. 7a and b, the addition of the selected AAs except Arg could not induce fluorescence enhancements of the aqueous suspension of UiO-66, in contrast, Lys, Asp, His, Met, Trp and Tyr lead to the obvious fluorescence quenching. In addition, although the addition of Arg was still capable to cause the increment of the emission intensity of UiO-66, the emission enhancement

amplitude in the case of UiO-66 was far lower than that of UiO-66-NH₂, which distinctly confirmed the essential role of amino group in UiO-66-NH₂ for the fluorescent turn-on sensing of Lys/Arg. These phenomena indicate that the introduction of amino functional group may be the critical factor to realize the sensing capacity and high sensitivity of UiO-66-NH₂ towards Lys and Arg.

Conclusions

In summary, we demonstrate the sensing capability of a well-used MOF, UiO-66-NH₂ towards Lys and Arg in aqueous media *via* the fluorescence turn-on effect. Quantitative titration experiments revealed that it can detect Lys and Arg selectively in various amino acids and the emission enhancement had linear correlation with the concentration of Lys/Arg, which indicated the realization of quantitative determination of Lys/Arg. Moreover, the selective and sensitive sensing Lys/Arg could be ascribed to the adsorption of Lys/Arg and the hydrogen bonding interactions between Lys/Arg and the amino group in UiO-66-NH₂. Comparing the sensing capacity and sensitivity of UiO-66 with that of UiO-66-NH₂, it could be concluded that the amino functional group play an essential role in the sensing performance.

Conflicts of interest

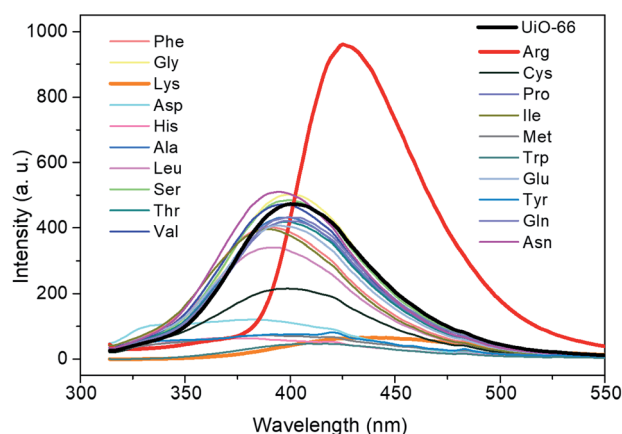
There are no conflicts to declare.

Acknowledgements

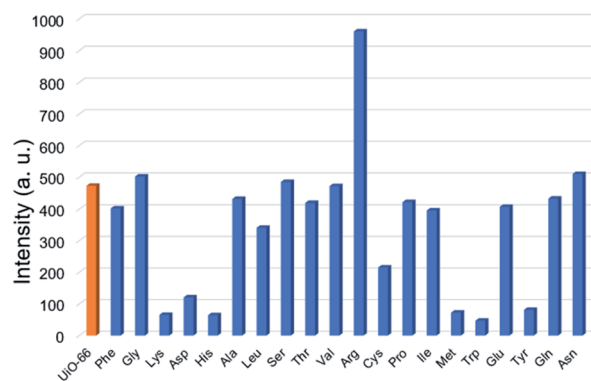
We gratefully acknowledge the National Basic Research Program of China (grant no. 2017YFA0303504) and the Fundamental Research Funds for the Central Universities (grant no. 14380232) for financial support of this work. This work was also supported by a Project Funded by the Priority Academic Program Development of Jiangsu Higher Education Institutions.

Notes and references

- 1 C. P. Han and H. B. Li, *Small*, 2008, **4**, 1344–1350.
- 2 J. Wang, H. B. Liu, Z. F. Tong and C. S. Ha, *Coord. Chem. Rev.*, 2015, **303**, 139–184.
- 3 Y. W. Zhao, Y. Wang and X. M. Zhang, *ACS Appl. Mater. Interfaces*, 2017, **9**, 20991–20999.
- 4 X. R. Wang, Z. Huang, J. Du, X. Z. Wang, N. Gu, X. Tian, Y. Li, Y. Y. Liu, J. Z. Huo and B. Ding, *Inorg. Chem.*, 2018, **57**, 12885–12899.
- 5 Y. Zhou and J. Yoon, *Chem. Soc. Rev.*, 2012, **41**, 52–67.
- 6 Y. H. Liu, M. J. Huangfu, P. Y. Wu, M. Jiang, X. L. Zhao, L. L. Liang, L. H. Xie, J. G. Bai and J. Wang, *Dalton Trans.*, 2019, **48**, 13834–13840.
- 7 X. Z. Wang, X. R. Wang, Y. Y. Liu, J. Z. Huo, Y. Li, Q. Wang, K. Liu and B. Ding, *Ultrason. Sonochem.*, 2019, **59**, 104734.
- 8 R. Cui, Y. Wan, G. Ji and Z. Liu, *Analyst*, 2019, **144**, 5875–5881.



(a)



(b)

Fig. 7 (a) The fluorescence emission curves of the aqueous suspension of UiO-66 in the absence and presence of different AAs. (b) Maximum fluorescence intensities of the aqueous suspension of UiO-66 in the absence and presence of different AAs.



- 9 W. D. Pu, H. W. Zhao, C. Z. Huang, L. P. Wu and D. Xu, *Anal. Chim. Acta*, 2013, **764**, 78–83.
- 10 R. S. Bhosale, G. V. Shitre, R. Kumar, D. O. Biradar, S. V. Bhosale, R. Narayan and S. V. Bhosale, *Sens. Actuators, B*, 2017, **241**, 1270–1275.
- 11 J. Dong, D. Zhao, Y. Lu and W. Y. Sun, *J. Mater. Chem. A*, 2019, **7**, 22744–22767.
- 12 S. R. Batten, N. R. Champness, X. M. Chen, J. Garcia-Martinez, S. Kitagawa, L. Ohrstrom, M. O'Keeffe, M. P. Suh and J. Reedijk, *Pure Appl. Chem.*, 2013, **85**, 1715–1724.
- 13 J. S. Chavez, K. L. Harrison and D. F. Sava Gallis, *RSC Adv.*, 2017, **7**, 24312–24320.
- 14 F. Vermoortele, B. Bueken, G. Le Bars, B. Van de Voorde, M. Vandichel, K. Houthoofd, A. Vimont, M. Daturi, M. Waroquier, V. Van Speybroeck, C. Kirschhock and D. E. De Vos, *J. Am. Chem. Soc.*, 2013, **135**, 11465–11468.
- 15 N. L. Rosi, J. Eckert, M. Eddaoudi, D. T. Vodak, J. Kim, M. O'Keeffe and O. M. Yaghi, *Science*, 2003, **300**, 1127–1129.
- 16 J. R. Li, J. Sculley and H. C. Zhou, *Chem. Rev.*, 2012, **112**, 869–932.
- 17 K. M. L. Taylor-Pashow, J. Della Rocca, Z. Xie, S. Tran and W. Lin, *J. Am. Chem. Soc.*, 2009, **131**, 14261–14263.
- 18 H. Li, M. Eddaoudi, M. O'Keeffe and O. M. Yaghi, *Nature*, 1999, **402**, 276–279.
- 19 M. Eddaoudi, H. L. Li and O. M. Yaghi, *J. Am. Chem. Soc.*, 2000, **122**, 1391–1397.
- 20 B. L. Chen, M. Eddaoudi, S. T. Hyde, M. O'Keeffe and O. M. Yaghi, *Science*, 2001, **291**, 1021–1023.
- 21 E. Haghighi and S. Zeinali, *Microporous Mesoporous Mater.*, 2020, **300**, 110065.
- 22 M. Zhang, G. Feng, Z. Song, Y. P. Zhou, H. Y. Chao, D. Yuan, T. T. Y. Tan, Z. Guo, Z. Hu, B. Z. Tang, B. Liu and D. Zhao, *J. Am. Chem. Soc.*, 2014, **136**, 7241–7244.
- 23 H. Xu, F. Liu, Y. Cui, B. Chen and G. Qian, *Chem. Commun.*, 2011, **47**, 3153–3155.
- 24 W. P. Lustig, S. Mukherjee, N. D. Rudd, A. V. Desai, J. Li and S. K. Ghosh, *Chem. Soc. Rev.*, 2017, **46**, 3242–3285.
- 25 Y. Zhang, S. Yuan, G. Day, X. Wang, X. Yang and H. C. Zhou, *Coord. Chem. Rev.*, 2018, **354**, 28–45.
- 26 L. E. Kreno, K. Leong, O. K. Farha, M. Allendorf, R. P. Van Duyne and J. T. Hupp, *Chem. Rev.*, 2012, **112**, 1105–1125.
- 27 A. Buragohain and S. Biswas, *CrystEngComm*, 2016, **18**, 4374–4381.
- 28 M. Kandiah, S. Usseglio, S. Svelle, U. Olsbye, K. P. Lillerud and M. Tilset, *J. Mater. Chem.*, 2010, **20**, 9848–9851.
- 29 S. Oien-Odegaard, B. Bouchevreau, K. Hylland, L. P. Wu, R. Blom, C. Grande, U. Olsbye, M. Tilset and K. P. Lillerud, *Inorg. Chem.*, 2016, **55**, 1986–1991.
- 30 M. Kandiah, M. H. Nilsen, S. Usseglio, S. Jakobsen, U. Olsbye, M. Tilset, C. Larabi, E. A. Quadrelli, F. Bonino and K. P. Lillerud, *Chem. Mater.*, 2010, **22**, 6632–6640.
- 31 S. Chavan, J. G. Vitillo, D. Gianolio, O. Zavorotynska, B. Civalieri, S. Jakobsen, M. H. Nilsen, L. Valenzano, C. Lamberti, K. P. Lillerud and S. Bordiga, *Phys. Chem. Chem. Phys.*, 2012, **14**, 1614–1626.
- 32 H. Chevreau, W. Liang, G. J. Kearley, S. G. Duyker, D. M. D'Alessandro and V. K. Peterson, *J. Phys. Chem. C*, 2015, **119**, 6980–6987.
- 33 C. A. Trickett, K. J. Gagnon, S. Lee, F. Gandara, H. B. Burgi and O. M. Yaghi, *Angew. Chem., Int. Ed.*, 2015, **54**, 11162–11167.
- 34 J. Yang, Y. Dai, X. Zhu, Z. Wang, Y. Li, Q. Zhuang, J. Shi and J. Gu, *J. Mater. Chem. A*, 2015, **3**, 7445–7452.
- 35 H. Zhu, J. Huang, Q. Zhou, Z. Lv, C. Li and G. Hu, *J. Lumin.*, 2019, **208**, 67–74.

