


 Cite this: *RSC Adv.*, 2020, **10**, 29682

The statistical fusion identification of dairy products based on extracted Raman spectroscopy†

 Zheng-Yong Zhang *^{ab}

At present, practical and rapid identification techniques for dairy products are still scarce. Taking different brands of pasteurized milk as an example, they are all milky white in appearance, and their Raman spectra are very similar, so it is not feasible to identify them directly using the naked eye. In the current work, a clear feature extraction and fusion strategy based on a combination of Raman spectroscopy and a support vector machine (SVM) algorithm was demonstrated. The results showed a 58% average recognition accuracy rate for dairy products as based on the original Raman full spectral data and up to nearly 70% based on a single spectral interval. Data normalization processing effectively improved the recognition accuracy rate. The average recognition accuracy rate of dairy products reached 91% based on the normalized Raman full spectral data or nearly 85% based on a normalized single spectral interval. The fusion of multispectral feature regions yielded high accuracy and operation efficiency. After screening and optimizing based on SVM algorithm, the best spectral feature intervals were determined to be 335–354 cm^{-1} , 435–454 cm^{-1} , 485–540 cm^{-1} , 820–915 cm^{-1} , 1155–1185 cm^{-1} , 1300–1414 cm^{-1} , and 1415–1520 cm^{-1} under the experimental conditions, and the average identification accuracy rate here reached 93%. The developed scheme has the advantages of clear feature extraction and fusion, and short identification time, and it provides a technical reference for food quality control.

 Received 20th July 2020
 Accepted 28th July 2020

DOI: 10.1039/d0ra06318e

rsc.li/rsc-advances

1. Introduction

The quality and safety of dairy products have, due to their importance as food products, always been major concerns of ordinary consumers, regulators and scientists. The evaluation of the safety of dairy products mainly involves the analysis of any illegal additives (such as melamine and sodium thiocyanate),^{1,2} pollutants (such as lead, mercury, arsenic, and chromium, nitrate),³ mycotoxins (such as aflatoxin B1 and aflatoxin M1),^{4,5} microorganisms (such as *Staphylococcus aureus* and *Salmonella*),⁶ etc. The evaluation of the quality of dairy products mainly involves evaluating the fluctuation and conformity of the products; that is, the products should maintain high stability.⁷ In the previous published reports, the research has mainly focused on the safety factors of dairy products, and trace harmful substances can be determined qualitatively and quantitatively.⁸ However, the existing research methods also face some challenges, especially in the field of quality evaluation.⁹ For example, in recent years, law enforcement officers in China have found many cases of the

manufacturing and selling of fake dairy products. Recently these counterfeit products have been low-quality and low-price versions of brand products; and while in line with the national standards and harmless to the human body, the counterfeiters use them as fake mimics of the high-quality and high-prices products in order to deceive consumers and make illegal profits.¹⁰ There are many kinds of dairy products, and different dairy products show considerably different appearances and contents of their main components, so it is relatively easy to distinguish them. However, the dairy products of the different brands do have similar appearances and compositions, which has become one of the challenges in efficiently identifying them.^{11–13} In the work described in the current paper, different brands of pasteurized milk were taken as examples. Their all having milky white appearances make their direct identifications by the naked eye not feasible. Thus we set out to conduct research into the more intelligent identification of the quality of dairy products.

This research included two aspects: quality characterization and statistical identification. The technologies available for characterizing the quality of dairy products include chromatography, mass spectrometry and spectroscopy. Chromatography and mass spectrometry detection generally requires sample pretreatment, which is time consuming and laborious.¹⁴ The spectral detection methods include ultraviolet spectroscopy, fluorescence spectroscopy, infrared spectroscopy and Raman spectroscopy. The ultraviolet spectroscopy and fluorescence spectroscopy techniques provide relatively little information about dairy products.¹⁵

*State Key Laboratory of Dairy Biotechnology, Shanghai Engineering Research Center of Dairy Biotechnology, Dairy Research Institute, Bright Dairy & Food Co., Ltd., Shanghai 200436, The People's Republic of China

^bSchool of Management Science and Engineering, Nanjing University of Finance and Economics, Nanjing, Jiangsu 210023, The People's Republic of China. E-mail: zyzhang@nufe.edu.cn

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra06318e



Infrared spectroscopy is sensitive to water molecules. Raman spectroscopy shows many advantages: (1) it can be used to detect the sample directly; (2) the cross section of Raman scattering by water molecules is small, and hence does not affect the sample detection; (3) a portable Raman spectrometer can be used to realize on-site sample detection; and (4) Raman spectroscopy can be used to characterize the rich molecular vibration information of a sample, and can thus provide a “fingerprint” of the sample.^{16–18} Therefore, research on the quality identification of dairy products based on Raman spectroscopy has become a major pursuit. The Raman spectrum of each sample can be formulated as a numerical matrix, which can be used as a sample quality characteristic to input into the subsequent statistical model.¹⁹

Statistical identification is an important technology for the high-efficiency, in-depth mining of the internal relationships between characterization data and for realizing the scientific quality control of dairy products. The existing corresponding research methods mainly include principal component analysis (PCA), *etc.* Because PCA is a mathematical transformation feature extraction method, it can achieve a reduction in data dimensionality, but it ignores the chemical information of Raman spectra of samples, and the practical application is not particularly convenient.^{20–22} Therefore, on the basis of collecting and analyzing the Raman spectra of dairy products, in the work described in this paper, we comprehensively developed a support vector machine (SVM) algorithm for the feature extraction and fusion identification of Raman spectral characterization data, and developed a novel identification method approach.

2. Experimental

2.1 Samples and instruments

Momchilovtsi (MS)-pasteurized heat-treated flavored yoghurt products were obtained from Bright Dairy & Food Co., Ltd. Ambpoeial (AM)-pasteurized heat-treated flavored yoghurt products were obtained from Inner Mongolia Yili Industrial Group Co., Ltd. ChunZhen (CZ)-pasteurized heat-treated flavored yoghurt products were obtained from Mengniu Dairy Group Co. Ltd. The samples of the dairy products of each brand were original flavors; that is, they did not include any additional chemical. These samples all appeared milky white. Each dairy brand was tested 30 times. Information on the contents of the main components of these samples is shown in Table S1.†

The Raman spectra of the samples were obtained by using a Prott-ezRaman-D3 portable laser Raman spectrometer (Enwave Optronics, U.S.A.). The excitation wavelength of the laser was 785 nm, the laser power was 450 mW, the fiber coupled laser output was 100 μm, and the integration time was 150 s. The spectrometer was operated from 255 to 1974 cm⁻¹ with a resolution 1 cm⁻¹. Raman spectra of these samples were obtained without any physical and chemical pretreatments except that the samples were fully shaken.

2.2 Data analysis

The baseline calibration of the collected Raman spectra was carried out by using SLSR Reader V8.3.9 software (Enwave

Optronics, U.S.A.). The support vector machine (SVM) algorithm used for feature extraction and fusion identification of Raman spectral characterization data was run using MATLAB software (MathWorks, Natick, MA, U.S.A.).

A brief description of the basic idea of the SVM classification algorithm is provided here.^{23–25} First, a known training set (T) can be described using the formula

$$T = \{(x_1, y_1), \dots, (x_m, y_m)\} \in (X \times Y)^m,$$

where $x_i \in X = R^n$, $y_i \in Y = \{1, -1\}$ ($i = 1, 2, \dots, m$). Here x_i is the Raman spectrum data, and y_i is the category label. A two-classification model is described here. A multiple classification problem can be solved by constructing a multiple two-classification SVM. By selecting the appropriate kernel function $K(x, x')$ and parameter C , the optimization problem may be constructed and solved using the term

$$\min_{\alpha} \frac{1}{2} \sum_{i=1}^j \sum_{j=1}^m y_i y_j \alpha_i \alpha_j K(x_i, x_j) - \sum_{j=1}^m \alpha_j$$

and equation

$$\text{s.t.} \sum_{i=1}^m y_i \alpha_i = 0, \quad 0 \leq \alpha_i \leq C, \quad i = 1, \dots, m.$$

To obtain the optimal solution $\alpha^* = (\alpha_1^*, \dots, \alpha_m^*)^T$. A positive component of α^* is selected as $0 < \alpha_j^* < C$, and the threshold is calculated according to the equation $b^* = y_j - \sum_{i=1}^m y_i \alpha_i^* K(x_i - x_j)$. Also constructed is the decision function $f(x) = \text{sgn}(\sum_{i=1}^m \alpha_i^* y_i K(x, x_i) + b^*)$.

In the current work, the radial basis function (RBF) was applied as kernel functions using the equation

$$K(x, x_i) = \exp \left[-\frac{\|x - x_i\|^2}{2\sigma^2} \right],$$

where σ represents the kernel width of the RBF.

3. Results and discussion

3.1 Characterization of dairy products using Raman spectroscopy

As shown in Fig. 1, Raman spectra of three dairy products were collected, and their main Raman peak assignments were attributed to the corresponding molecular vibrations according to previous reports.^{26–28} The Raman band at 1759 cm⁻¹ was assigned to the ester C=O stretching of fatty acids. The strong Raman band at 1468 cm⁻¹ was attributed to CH₂ deformation of fats and sugars. And the strong band at 1016 cm⁻¹ was assigned to the ring-breathing mode, which was mainly derived from the phenylalanine of proteins. More information on Raman peak attribution is shown in Table 1. In Fig. 1, MS represents the momchilovtsi-pasteurized heat-treated flavored yoghurt products from Bright Dairy & Food Co., Ltd. Also, AM represents the ambpoeial-pasteurized heat-treated flavored yoghurt products



from Inner Mongolia Yili Industrial Group Co., Ltd., and CZ represents the ChunZhen-pasteurized heat-treated flavored yoghurt products from Mengniu Dairy Group Co. Ltd., respectively. Nearly 20 main narrow peaks were identified in the Raman spectrum of each experimental sample. The results showed that the samples were rich in fat, sugar and protein and yielded strong Raman fingerprints. Yet the Raman spectra of the experimental samples were very similar. Fig. S1–S3† show the Raman spectra of 30 samples of MS, AM and CZ dairy product brands, respectively. As shown in Fig. S4,† the dairy products of the various brands were milky white in appearance.

Furthermore, a single value moving range control chart was used to evaluate the quality fluctuation of the spectral data of each brand sample based on Raman peak intensities (at 1468 cm^{-1}), as shown in Fig. S5–S7.† The results showed that the spectra of each brand sample show statistically explainable quality fluctuations; the Raman spectral intensities of MS, AM and CZ were found to be located in the ranges 462.6–757.8, 397.4–682.2 and 482.5–739.8, respectively.^{7,29} The experimental results suggested that the spectral data conformed to certain statistical rules. At the same time, analysis also revealed spectral intensity overlap among brands, and therefore spectral intensity analysis alone cannot achieve brand identification. The nutritional components of the three dairy products were also similar (Table S1†), so the traditional identification methods would have been difficult to carry out and the compositional analysis would have been time-consuming and laborious. Therefore, taking their identification as an example, a new statistical identification method was provided.

3.2 Data preprocessing and feature extraction of the Raman spectra of the dairy products

First we investigated identification based on original Raman spectral data of the dairy products. The average recognition rate of the SVM model based on the original Raman full spectral

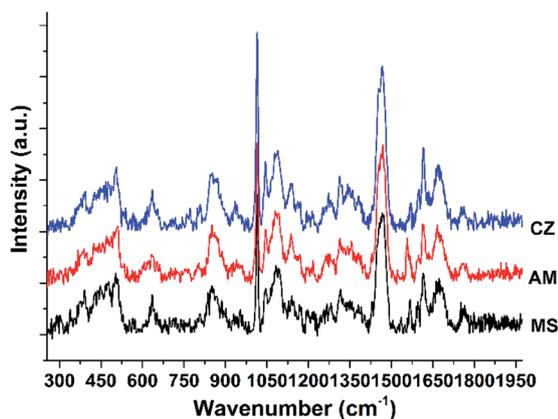


Fig. 1 Raman spectra of the different dairy products. (MS represents the momchilovtsi-pasteurized heat-treated flavored yoghurt products from Bright Dairy & Food Co., Ltd. AM represents the ambpoeial-pasteurized heat-treated flavored yoghurt products from Inner Mongolia Yili Industrial Group Co., Ltd. CZ represents the ChunZhen-pasteurized heat-treated flavored yoghurt products from Mengniu Dairy Group Co. Ltd.).

Table 1 The wavenumbers (cm^{-1}) of the main Raman peaks and their respective tentative assignments^{26–28a}

Wavenumber (cm^{-1})	Assignment
1759	$\nu(\text{C}=\text{O})_{\text{ester}}$
1670	$\nu(\text{C}=\text{O})$ amide I; $\nu(\text{C}=\text{C})$
1615	$\nu(\text{C}-\text{C})_{\text{ring}}$
1566	$\delta(\text{N}-\text{H})$; $\nu(\text{C}-\text{N})$ amide II
1468	$\delta(\text{CH}_2)$
1315	$\tau(\text{CH}_2)$
1279	$\gamma(\text{CH}_2)$
1139	$\nu(\text{C}-\text{O}) + \nu(\text{C}-\text{C}) + \delta(\text{C}-\text{O}-\text{H})$
1085	$\nu(\text{C}-\text{O}) + \nu(\text{C}-\text{C}) + \delta(\text{C}-\text{O}-\text{H})$
1046	$\nu(\text{C}-\text{O}) + \nu(\text{C}-\text{C}) + \delta(\text{C}-\text{O}-\text{H})$
1016	Ring-breathing (phenylalanine); $\nu(\text{C}-\text{C})_{\text{ring}}$
953	$\delta(\text{C}-\text{O}-\text{C}) + \delta(\text{C}-\text{O}-\text{H}) + \nu(\text{C}-\text{O})$
851	$\delta(\text{C}-\text{C}-\text{H}) + \delta(\text{C}-\text{O}-\text{C})$
807	$\delta(\text{C}-\text{C}-\text{O})$
633	$\delta(\text{C}-\text{C}-\text{O})$
502	Glucose
445	$\delta(\text{C}-\text{C}-\text{H}) + \tau(\text{C}-\text{O})$
389	Lactose

^a ν —stretching vibration, δ —deformation vibration, τ —twisting vibration, γ —out-of-plane bending vibration.

data was measured to be 58%. The test condition was that two-thirds of the samples were randomly selected as the training set, and the remaining samples were employed as the validation set. Ten runs of random tests were conducted to obtain the average accuracy rate. The purpose of feature extraction and analysis in general is to reduce the effects of interference of redundant information on the identification model, improve the detailed differences between samples, and then realize accurate identifications of the different samples and save running time. In the current work, the moving window method and spectral band selection method were employed to screen Raman spectral intervals. The width of the moving window was 20 wavenumbers. The experimental results are shown in Tables S2 and S3.† The highest average recognition rate reached 69%, and did so for 1755–1774 cm^{-1} . This recognition rate was higher than the recognition rate based on a full spectrum, indicating an expected recognition effect improvement to result from feature extraction.

Table 2 The identification results of dairy products based on different pretreatment methods and the use of the SVM recognition algorithm. The Raman spectroscopy range was 255–1974 cm^{-1}

Spectral pretreatment method	Average accuracy rate (%)	Running time (seconds)
Original	58	820
First derivative	53	833
Second derivative	51	841
Third derivative	40	831
Fourth derivative	53	838
Normalization	91	823



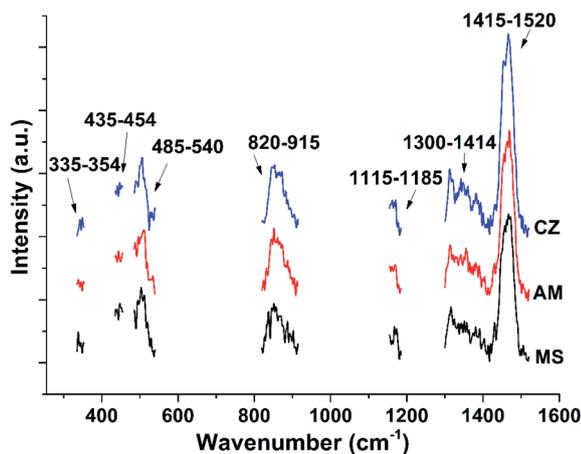


Fig. 2 Extracted Raman spectral feature intervals of dairy products. (MS represents the momchilovtsi-pasteurized heat-treated flavored yoghurt products from Bright Dairy & Food Co., Ltd. AM represents the ambpoelial-pasteurized heat-treated flavored yoghurt products from Inner Mongolia Yili Industrial Group Co., Ltd. CZ represents the ChunZhen-pasteurized heat-treated flavored yoghurt products from Mengniu Dairy Group Co. Ltd., respectively).

Then we investigated identification based on preprocessing Raman spectral data of dairy products. Spectral preprocessing was expected to help us highlight data differences and improve the recognition rate of models. Calculations of the first, second, third, and fourth derivatives and normalization processing were carried out on the Raman spectral data, as shown in Fig. S8–S12,[†] respectively. Mere visual inspection of these spectra did not show any very obvious difference between them. However, the average recognition rates calculated using the SVM model did show some differences, and the results are shown in Table 2. The recognition rates based on derivative processing decreased, while the recognition rates based on normalization processing increased significantly. These results suggested

a great impact of the normalization operation on the SVM model and suggested an effective removal of the dimensional influence of the original spectral data by the normalization.^{30–32} Furthermore, two thirds of the sample data were randomly selected as the training set, and the remaining data were used as the verification set. The average recognition effect of the moving window selection and spectral band feature selection was investigated in ten random experiments. The identification results are shown in Tables S4 and S5.[†] Each of the identification accuracy rates in the Raman spectral ranges 335–354 cm^{-1} , 435–454 cm^{-1} , and 835–854 cm^{-1} for moving window method was greater than 70%. The average of the recognition rates based on the Raman ranges 485–540 cm^{-1} , 1155–1185 cm^{-1} , 1300–1415 cm^{-1} and 1415–1520 cm^{-1} interval reached 83%. The results suggested that compared with the corresponding original data band mentioned above (Tables S2 and S3[†]), most of the recognition rates based on the feature band were improved after normalization, the contribution of different spectral bands to the accurate recognition of the SVM model were different, and the recognition rate was lower than that of the full spectral data (91%, Table 2), which may have been due to the lack of spectral data input from the SVM algorithm. Obvious differences between the samples were identified in the spectral feature intervals, as shown in Fig. 2.

3.3 Statistical fusion identification of dairy products based on Raman spectra

The data fusion methods reported in the literature mainly include data layer fusion, feature layer fusion and model level fusion. These methods are used with the expectation that they improve the efficiency of data utilization and improve the accuracy of model discrimination. Direct data fusion often involves redundant information. Model level fusion requires multiple classifiers, its operation is relatively complex, and the chemical information of the data tends to be fuzzy, which is not conducive to practical

Table 3 The identification results of dairy products based on the fusion of their Raman spectroscopy features and the use of the SVM recognition algorithm

Raman spectroscopy interval ranges (cm^{-1})	Average accuracy rate (%)	Running time (seconds)
335–354, 435–454	76	30
335–354, 435–454, 835–854	84	38
485–540, 820–915	84	78
485–540, 820–915, 1115–1185	85	90
485–540, 820–915, 1115–1185, 1300–1415	88	143
485–540, 820–915, 1115–1185, 1300–1414, 1415–1520	89	192
485–540, 820–915, 1115–1185, 1415–1520	87	138
335–354, 435–454, 485–540, 835–854	83	61
335–354, 435–454, 835–854, 1115–1185	83	51
335–354, 435–454, 835–854, 1115–1185, 1300–1415	90	102
335–354, 435–454, 835–854, 1115–1185, 1300–1414, 1415–1520	90	151
335–354, 435–454, 485–540, 835–854, 1115–1185, 1300–1414, 1415–1520	91	174
335–354, 435–454, 485–540, 820–915, 1115–1185, 1300–1414, 1415–1520	93	208



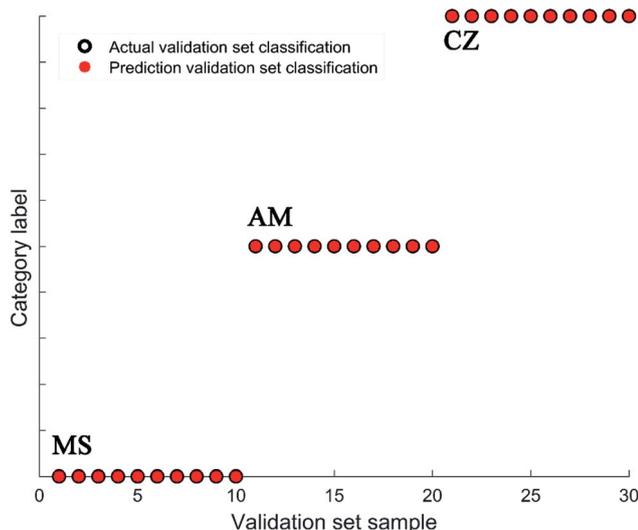


Fig. 3 Classification results for dairy products based on the use of the SVM. (MS represents momchilovtsi-pasteurized heat-treated flavored yoghurt products from Bright Dairy & Food Co., Ltd; AM represents ambpoelial-pasteurized heat-treated flavored yoghurt products from Inner Mongolia Yili Industrial Group Co., Ltd; and CZ represents ChunZhen-pasteurized heat-treated flavored yoghurt products from Mengniu Dairy Group Co. Ltd, respectively).

applications. However, the data fusion strategy based on clear chemical feature extraction is simple, intuitive and practical.^{33–35} The above-described research revealed different feature regions of Raman spectra of dairy products making different contributions to the identification methods of sample categories. After normalization, the recognition accuracy of the model improved, and at the same time more than one spectral interval with an accuracy rate of greater than 70% was found. The results suggested that we can further improve the accuracy rate of the identification model through the fusion of feature spectral intervals. To arrive at the test conditions, all spectral data were normalized, and then two-thirds of the spectral data were randomly selected as the training set, and the remaining one-third of the spectral data was employed as the validation set. Ten runs of random tests were conducted to obtain the average accuracy rate. As shown in Table 3, the average recognition rate of the model was higher than that based on the corresponding single feature spectral interval through different combinations of fusion. The results show that the best spectral feature fusion intervals included 335–354 cm^{-1} , 435–454 cm^{-1} , 485–540 cm^{-1} , 820–915 cm^{-1} , 1155–1185 cm^{-1} , 1300–1414 cm^{-1} and 1415–1520 cm^{-1} , and the average accuracy rate was increased from 58% (based on full spectral data without normalization) to 91% (based on full spectral data with normalization) and finally here to 93% with a savings of the running time (from about 820 s to 208 s). The best classification result based on the feature interval fusion reached 100%, as shown in Fig. 3.

4. Conclusions

In this work, a new and convenient research strategy for the feature extraction and fusion identification of dairy products

was proposed. Through the comprehensive application of Raman spectroscopy and SVM algorithm techniques, the Raman spectroscopy feature intervals could be extracted clearly and effectively, and the best spectral feature intervals in the experimental system were determined to be 335–354 cm^{-1} , 435–454 cm^{-1} , 485–540 cm^{-1} , 820–915 cm^{-1} , 1155–1185 cm^{-1} , 1300–1414 cm^{-1} , and 1415–1520 cm^{-1} . The identification accuracy rate could be further improved through normalization treatment and feature fusion, and increased to 93% from 58% (based on full spectral data without normalization) and 91% (based on full spectral data with normalization). The total running time of the model was greatly reduced from about 820 s to 208 s. The scheme has many advantages, such as the simplicity of sample signal acquisition, the high speed of analysis, the portability of the equipment, *etc.* This approach has potential application prospects in the fields of food anti-counterfeiting and quality control.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This research was financially supported by the Open Project Program of State Key Laboratory of Dairy Biotechnology (No. SKLDB2018-007) and the National Natural Science Foundation of China (No. 61602217).

References

- Z. Zhang, J. Liu and H. Wang, *Anal. Lett.*, 2015, **48**, 1930–1940.
- A. Kaleem, M. Azmat, A. Sharma, G. Shen and X. Ding, *Food Chem.*, 2019, **277**, 624–631.
- P. Singh, M. K. Singh, Y. R. Beg and G. R. Nishad, *Talanta*, 2019, **191**, 364–381.
- Z. Xue, Y. Zhang, W. Yu, J. Zhang, J. Wang, F. Wan, Y. Kim, Y. Liu and X. Kou, *Anal. Chim. Acta*, 2019, **1069**, 1–27.
- L. Qu, Q. Jia, C. Liu, W. Wang, L. Duan, G. Yang, C. Han and H. Li, *J. Chromatogr. A*, 2018, **1579**, 115–120.
- S. Stöckel, J. Kirchoff, U. Neugebauer, P. Rösch and J. Popp, *J. Raman Spectrosc.*, 2016, **47**, 89–109.
- D. C. Montgomery, *Introduction to Statistical Quality Control*, John Wiley & Sons, Inc., Sixth edn, 2009.
- X. Hu, J. Shi, Y. Shi, X. Zou, M. Arslan, W. Zhang, X. Huang, Z. Li and Y. Xu, *Food Chem.*, 2019, **272**, 58–65.
- X. Wang, C. Esquerre, G. Downey, L. Henihan, D. O. Callaghan and C. O. Donnell, *Talanta*, 2018, **183**, 320–328.
- Z. Zhang, M. Sha, J. Liu and H. Wang, *China Dairy Ind.*, 2017, **45**, 49–51.
- Y. Zhang, Z. Zhang, Y. Zhao, R. Dian, Y. Cheng, X. Qin and H. Wang, *Talanta*, 2020, **211**, 120681.
- N. N. Yazgan Karacaglar, T. Bulat, I. H. Boyaci and A. Topcu, *J. Food Drug Anal.*, 2019, **27**, 101–110.



- 13 C. M. McGoverin, A. S. S. Clark, S. E. Holroyd and K. C. Gordon, *Anal. Chim. Acta*, 2010, **673**, 26–32.
- 14 K. Slimani, Y. Pirottais, P. Maris, J. Abjean and D. Hurtaud-Pessel, *Food Chem.*, 2018, **262**, 168–177.
- 15 S. Xiong, B. Adhikari, X. D. Chen and L. Che, *J. Dairy Sci.*, 2016, **99**, 9652–9658.
- 16 A. Nedeljkovic, I. Tomasevic, J. Miocinovic and P. Pudja, *Food Chem.*, 2017, **232**, 487–492.
- 17 Z. Tan, T. Lou, Z. Huang, J. Zong, K. Xu, Q. Li and D. Chen, *J. Agric. Food Chem.*, 2017, **65**, 6274–6281.
- 18 Z. Zhang, M. Sha and H. Wang, *J. Raman Spectrosc.*, 2017, **48**, 1111–1115.
- 19 S. Weng, H. Yuan, X. Zhang, P. Li, L. Zheng, J. Zhao and L. Huang, *Analyst*, 2020, **145**, 4827–4835.
- 20 R. Ullah, S. Khan, H. Ali, M. Bilal, M. Saleem, A. Mahmood and M. Ahmed, *J. Raman Spectrosc.*, 2017, **48**, 692–696.
- 21 N. N. Yazgan Karacaglar, T. Bulat, I. H. Boyaci and A. Topcu, *J. Food Drug Anal.*, 2019, **27**, 101–110.
- 22 A. Amjad, R. Ullah, S. Khan, M. Bilal and A. Khan, *Vib. Spectrosc.*, 2018, **99**, 124–129.
- 23 S. Ren and L. Gao, *Analyst*, 2011, **136**, 1252–1261.
- 24 H. Sun, G. Lv, J. Mo, X. Lv, G. Du and Y. Liu, *Optik*, 2019, **184**, 214–219.
- 25 S. Bakhtiaridoost, H. Habibiyani, S. Muhammadnejad, M. Haddadi, H. Ghafoorifard, H. Arabalibeik and S. Amanpour, *RSC Adv.*, 2016, **6**, 50027–50033.
- 26 Z. Zhang, D. Gui, M. Sha, J. Liu and H. Wang, *J. Dairy Sci.*, 2019, **102**, 68–76.
- 27 P. H. R. Júnior, K. D. S. Oliveira, C. E. R. D. Almeida, L. F. C. D. Oliveira, R. Stephani, M. D. S. Pinto, A. F. D. Carvalho and Í. T. Perrone, *Food Chem.*, 2016, **196**, 584–588.
- 28 M. R. Almeida, K. D. S. Oliveira, R. Stephani and L. F. C. de Oliveira, *J. Raman Spectrosc.*, 2011, **42**, 1548–1552.
- 29 Z. Zhang, M. Sha, J. Liu and H. Wang, *J. Inst. Brew.*, 2017, **123**, 464–467.
- 30 F. Tian, F. Tan and H. Li, *Vib. Spectrosc.*, 2020, **107**, 103017.
- 31 H. Chen, L. Xu, W. Ai, B. Lin, Q. Feng and K. Cai, *Sci. Total Environ.*, 2020, **714**, 136765.
- 32 W. Li, X. Yan, J. Pan, S. Liu, D. Xue and H. Qu, *Spectrochim. Acta, Part A*, 2019, **218**, 271–280.
- 33 Z. Zhang and J. Liu, *J. Appl. Spectrosc.*, 2020, **87**, 99–104.
- 34 L. Zhou, C. Zhang, Z. Qiu and Y. He, *TrAC, Trends Anal. Chem.*, 2020, **127**, 115901.
- 35 M. Sha, Z. Zhang, D. Gui, Y. Wang, L. Fu and H. Wang, *Food Anal. Methods*, 2017, **10**, 3415–3423.

