


 Cite this: *RSC Adv.*, 2020, 10, 37989

Thiolated poly(aspartic acid)-functionalized two-dimensional MoS₂, chitosan and bismuth film as a sensor platform for cadmium ion detection†

 Qiang Cao,^{‡,ab} Yushi Xiao,^{‡,ab} Rong Huang,^{‡,ac} Na Liu,^{ab} Hai Chi,^{id d} Cheng-Te Lin,^{id e} Chi-Hsien Huang,^f Gang Han^a and Lidong Wu^{id *a}

In this work, a sensitive electrochemical platform for determination of cadmium ions (Cd²⁺) is obtained using thiolated poly(aspartic acid) (TPA)-functionalized MoS₂ as a sensor platform by differential pulse anodic stripping voltammetry (DPASV). The performance of the TPA–MoS₂-modified sensor is systemically studied. It demonstrates that the TPA–MoS₂ nanocomposite modified sensor exhibits superior analytical performance for Cd²⁺ over a linear range from 0.5 μg L⁻¹ to 50 μg L⁻¹, with a detection limit of 0.17 μg L⁻¹. Chitosan is able to form a continuous coating film on the surface of the GC electrode. The good sensing performance of the TPA–MoS₂-modified sensor may be attributed to the following factors: the large surface area of MoS₂ (603 m² g⁻¹), and the abundant thiol groups of TPA. Thus, the TPA–MoS₂-modified sensor proves to be a reliable and environmentally friendly tool for the effective monitoring of Cd²⁺ existing in aquacultural environments.

 Received 16th July 2020
 Accepted 6th October 2020

DOI: 10.1039/d0ra06197b

rsc.li/rsc-advances

1. Introduction

Cadmium (Cd) is one of the priority pollutants due to its high toxicity.^{1–3} Cd²⁺ not only causes renal tubular dysfunction and bone degeneration (itai-itai disease),^{4–6} but is also classified as a carcinogenic chemical by the International Agency for Research on Cancer (IARC).^{7–9} Traditionally, inductively coupled plasma mass spectrometry (ICP-MS),^{10,11} atomic absorption spectrometry (AAS)^{12–14} and ion chromatography^{15,16} are used for detecting Cd²⁺. Although these instruments have good precision and high resolution, they are too expensive and labor-intensive.^{17,18} Therefore, it is urgent to develop a sensitive method to protect our food, medicine and drinking water from Cd²⁺ pollution.

The electrochemical sensor is one of the most promising tools for detecting Cd²⁺ due to its low cost, rapid speed, portability and reliability.^{19–21} Mercury modified electrode has been intensively used for detecting trace Cd²⁺ by differential pulse anodic stripping voltammetry (DPASV).^{22,23} However, the toxicity of mercury severely limits its applications in constructing sensors.^{24,25} Therefore, bismuth, the less toxicity heavy metal,^{26,27} is used for constructing sensors in this work. Except electrode materials, the sensing material also plays an important role in constructing sensors.²⁸

As is well known, 2D materials have been established for the sensitive detection methods of environmental pollutants.^{29–32} Molybdenum disulfide (MoS₂) is one of transition-metal dichalcogenides (TMDs), which has a typical layered structure formed by a stack of planes.^{33–35} Each layer of MoS₂ consists of three atom layers (S–Mo–S), a layer of molybdenum atoms sandwiched between two layers of sulfur atoms.^{36–38} These triple layers are weakly held together by van der Waals forces.^{39–41} MoS₂ has unique properties including good flexibility, a tunable bandgap energy controlled by the number of layers and planar electric transportation property.^{42–45} Due to its fascinating characteristics, MoS₂ has received much attention for construction of promising electronics devices, for example, field-effect transistors, lithium-ion batteries, *etc.*^{46–49} However, exploring more binding sites on MoS₂ for Cd²⁺ is still challenging, and this is the key to achieve excellent low detection limit and sensitivity. The poor surface modification of MoS₂ limits its application as electrode material for constructing electrochemical sensors.⁵⁰

^aKey Laboratory of Control of Quality and Safety for Aquatic Products, Chinese Academy of Fishery Sciences, Beijing, 100141, China. E-mail: wulidong19849510@hotmail.com; Fax: +86-10-68690712; Tel: +86-10-68690712

^bShanghai Ocean University, Shanghai 201306, China

^cBeijing Engineering and Technology Research Center of Food Additives, Beijing Technology and Business University (BTBU), Beijing, 100048, China

^dEast China Sea Fisheries Research Institute, Chinese Academy of Fishery Sciences, Shanghai, 201306, China

^eNingbo Institute of Materials Technology and Engineering, Chinese Academy of Sciences, Ningbo, 315201, China

^fDepartment of Materials Engineering, Mingchi University of Technology, 243303, Taiwan

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra06197b

‡ Authors equally contribution to this work.



To address this critical challenge, it is predicted that controllable surface modification of MoS₂ could significantly improve the sensitivity of MoS₂ based sensor.^{51,52} Thus, in this work, thiolated polymers are chemically adsorbed on electron-rich MoS₂ through covalent bonding. The sulfur on the MoS₂ surface plays as a soft Lewis base showing a high affinity for heavy metal ions (*e.g.*, Bi³⁺, Cd²⁺). MoS₂ demonstrates both a high adsorption capacity due to abundant sulfur adsorption sites and fast kinetics due to easy access to these sites. Therefore, thiolated polymer modified MoS₂ plays as one of the most effective platforms for detecting Cd²⁺. It shows that this sensor possesses superior stability, sensitivity and LOD. The proposed sensor can be used as an environmental-friendly “pre-alarm” tool for rapid determining Cd²⁺ in aquaculture water, Chinese drug, food and beverage.

2. Experimental section

2.1. Materials and solutions

Cadmium chloride, bismuth chloride, and other chemicals are from Tianjin Kermel (Tianjin, China). Thiolated poly(aspartic acid) is obtained from Covestro company (Shanghai, China). 100 mmol L⁻¹ pH 6.5 acetate buffer is used as the electrolyte in electrochemical experiments.

2.2. Apparatus

A JEM-2100 instrument (Japan) is used for obtaining transmission electron microscopy (TEM) images. Scan electron microscopy (SEM) images are obtained by Zeiss Sigma 300 (Germany). A CHI 660B Electrochemical Workstation (Shanghai, China) is used for testing Cd²⁺ through DPASV. A functionalized MoS₂ and chitosan film modified glassy carbon (GC) electrode is the working electrode, the Ag/AgCl electrode is the reference electrode, and the platinum wire is the auxiliary electrode. The concentration of Cd²⁺ is also confirmed through atomic absorption spectrometry (AAS, Jena, Germany).

2.3. Preparation of MoS₂ and modification of GC electrode

MoS₂ nanosheets are prepared by an ultrasonic exfoliation method as literature reported.⁵³ 1.0 g MoS₂ powder is added into 10 mL *N*-methyl-pyrrolidone solution under ultrasonic wave for 2 h at 0 °C. After centrifugation at 7000 rpm for 30 min, the

prepared MoS₂ nanosheets is dispersed in the supernatant. The 1 mL of 1 mg mL⁻¹ prepared MoS₂ is mixed with 1 mL 2 mg mL⁻¹ thiolated poly(aspartic acid) (TPA), and vibrated by vortex for 12 h at room temperature.

The GC electrode is polished by alumina powder (0.05 μm) and washed by ethanol for three times, respectively. Then, 5 μL 0.6 mg mL⁻¹ functionalized MoS₂ solution, 5 μL chitosan (8 wt%) and 10 μL Milli-Q water are mixed. Finally, 5 μL mixture is immobilized onto the GC electrode to obtain a TPA-MoS₂/GC electrode.

2.4. Cd²⁺ detection by TPA-MoS₂/GC electrode

Bismuth film is plated on the TPA-MoS₂/GC electrode through the electrodeposition of 10 mg L⁻¹ Bi³⁺ for 210 s with stirring. After coating bismuth film, Cd²⁺ is monitored by the TPA-MoS₂/GC electrode in 8 mL 100 mmol L⁻¹ acetate buffer (pH 6.5).

3. Results and discussion

3.1. TEM characterization of MoS₂

MoS₂ was obtained through ultrasonic exfoliation method. Fig. 1 showed representative TEM images of the MoS₂ nanosheets (A), the functionalized MoS₂ (B) and the functionalized MoS₂ deposited with bismuth and cadmium (C). It suggested that the prepared MoS₂ nanosheets were very thin and clear. After TPA non-covalent modified on the MoS₂ nanosheets, the original morphology of the MoS₂ nanosheets was retained in the composites. Fig. 1C is a TEM image of the TPA-MoS₂-Bi-Cd nanocomposites and clearly showed the modification of the MoS₂ by Bi and Cd nanocluster. As shown in Fig. S1,† TPA-MoS₂ surface becomes rough and there are lots of nanoparticles on the surface of TPA-MoS₂ after detection. It is consistent with TEM images. FTIR was also used to confirm the formation of TPA on the MoS₂ nanosheets. As shown in Fig. 2, the peak of 2550 cm⁻¹ was the SH group of TPA. After TPA modified on the MoS₂ nanosheets, the absorption bands at 1390 (symmetric stretching of -COOH), 1600 (asymmetric stretching of -COOH), and 3000–3500 cm⁻¹ (NH₃⁺ stretch) became weak. The change in their dipole moment was due to TPA modification on the MoS₂ nanosheets. These results showed the successful modification of TPA on the MoS₂ nanosheets. X-ray photoelectron spectroscopy (XPS) was used to determine the chemical composition and states of the TPA-MoS₂

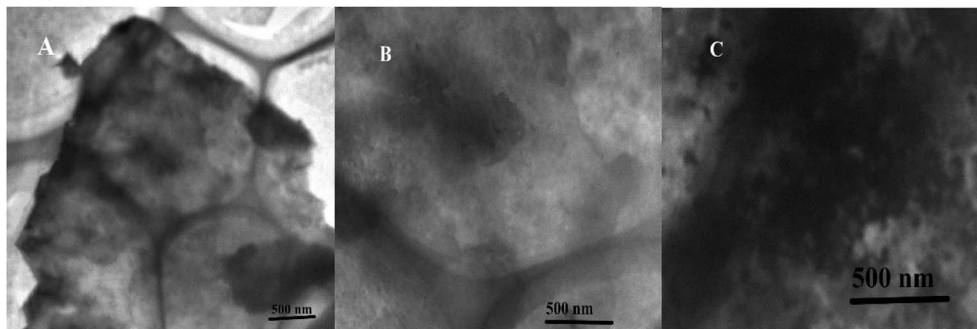


Fig. 1 TEM image of MoS₂ nanosheets (A), the TPA functionalized MoS₂ (B), and the TPA-MoS₂ with bismuth and cadmium (C).



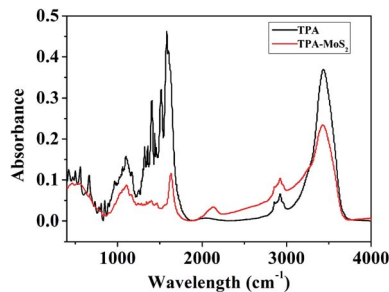


Fig. 2 FTIR image of TPA and TPA-MoS₂.

with cadmium. As shown in Fig. S2A,† two peaks at 229 and 232 eV energy values ascribed to Mo core peaks, and the peak at 226 eV ascribed to S core peak. Fig. S2B† showed two peaks at 163 and 161.7 eV. These binding energy values are consistent with the expected charge states of Cd²⁺, Mo⁴⁺ and S²⁻. Furthermore, the specific surface area of the MoS₂ nanosheets was estimated by the Brunauer-Emmett-Teller (BET) method. Its specific surface area achieved to be 603 m² g⁻¹. The high effective surface area and high-quality of MoS₂ could provide good performances for GC electrode.

3.2. Electrochemical characterization of the TPA-MoS₂-modified GC electrode

Cyclic voltammetry (CV) was an electrochemical measurement method which could be used for monitoring the preparation procedure of GC electrode. As shown in Fig. 3, the immobilization process of the TPA-MoS₂/GC electrode was performed. Fig. 3 displayed the CVs of the bare GC, MoS₂/GC and TPA-MoS₂/GC electrodes in 2 mmol L⁻¹ K₃[Fe(CN)₆]/K₄[Fe(CN)₆] solution. The CV of the bare GC electrode was highest among these electrodes. It demonstrated that the electron transfer rate between [Fe(CN)₆]^{3-/4-} and the bare electrode was very fast. The CV of the TPA-MoS₂/GC electrode was the lowest, which demonstrated that a chitosan film and TPA-MoS₂ had covered on the surface of the bare GC electrode. Without modification of

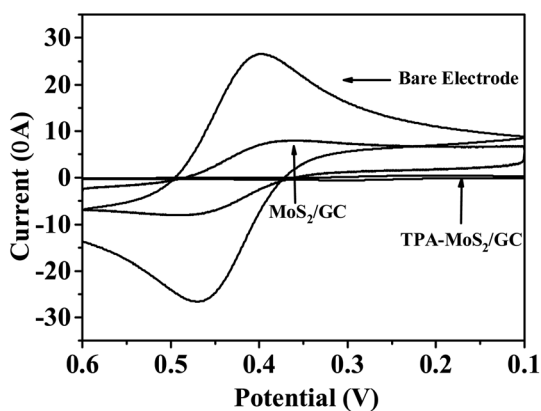


Fig. 3 Cyclic voltammograms of bare GC, MoS₂/GC and TPA-MoS₂/GC electrodes in 2 mmol L⁻¹ [Fe(CN)₆]^{3-/4-} at a scan rate of 100 mV s⁻¹.

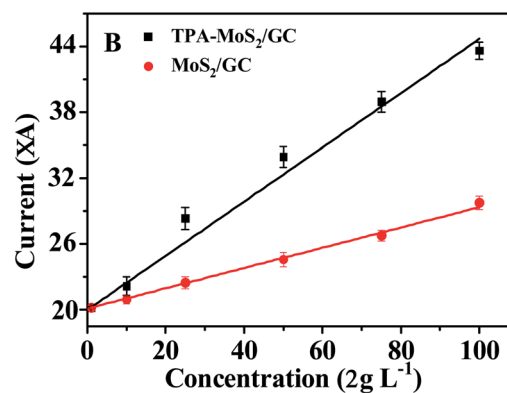
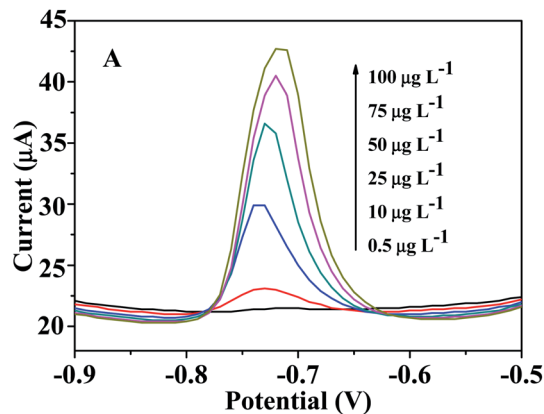


Fig. 4 (A) DPASVs obtained for different concentrations of Cd²⁺ using a bismuth-coated TPA-MoS₂/GC electrode; from bottom to top, 0.5, 10, 25, 50, 75, 100 µg L⁻¹ with 10 mg L⁻¹ Bi³⁺ electrodeposited for 210 s under stirring in 100 mmol L⁻¹ acetate buffer (pH 5.0). (B) Calibration curves of MoS₂/GC (red color) and TPA-MoS₂/GC (black color) electrodes for Cd²⁺.

TPA, the MoS₂ nanosheets possessed good conductivity, and the CV of the chitosan-MoS₂/GC electrode was higher than the TPA-MoS₂/GC electrode. The results might be due to the introduction of TPA, which played a key role in improving the binding sites of Cd²⁺. It also showed that the TPA-MoS₂ and chitosan film had covered on the GC electrode.

3.3. The TPA-MoS₂-modified GC electrode for Cd²⁺ analysis

In this study, DPASV was used for detecting Cd²⁺. Fig. 4A showed that the DPASVs of the MoS₂/GC and the TPA-MoS₂/GC electrodes increased with the concentrations of Cd²⁺ from 0.5 to 100 µg L⁻¹ in 100 mmol L⁻¹ acetate buffer (pH 6.5) with 10 mg L⁻¹ Bi³⁺ electrodepositing for 210 s. After the concentrations of Cd²⁺ achieved 150, 200 and 250 µg L⁻¹, the DPASVs of these electrodes increased unobvious. The DPASVs response signals increased linearly from 0.5 to 100 µg L⁻¹ then become flat. As shown in Fig. 4B, the current peaks increased linearly with Cd²⁺ concentration, and the correlation coefficients was approximate 0.99. The sensitivity of the MoS₂/GC and the TPA-MoS₂/GC electrodes were 101.8 and 178.2 mA cm⁻² m⁻¹, respectively. Furthermore, the detection limit (DL) was an



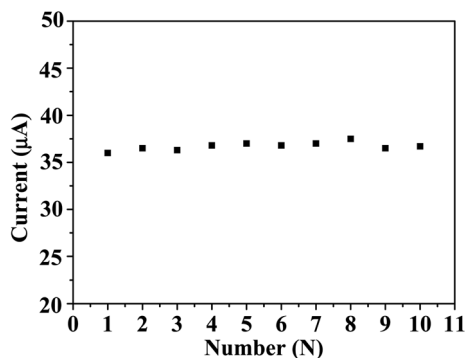


Fig. 5 The stability of 10 repetitive measurements of $50 \mu\text{g L}^{-1} \text{Cd}^{2+}$ in 0.1 mol L^{-1} acetate buffer (pH 5.0) containing $3 \text{ mg L}^{-1} \text{Bi}^{3+}$.

Table 1 Comparison of Cd^{2+} concentration in different aquaculture water samples by the sensor and AAS method

Sample	Concentration ($\mu\text{g L}^{-1}$) (sensor method)	Concentration ($\mu\text{g L}^{-1}$) (AAS method)	Difference ^a (%)
A	1.9	1.7	11.76
B	—	—	—
C	—	—	—
D	2.1	2.4	-12.5
E	—	—	—
F	—	—	—

^a Difference = (sensor(value) – AAS(value))/AAS(value) × 100%.

important parameter as sensors. The DL of the TPA-MoS₂/GC and the MoS₂/GC electrodes achieved $0.2 \mu\text{g L}^{-1}$ and $0.5 \mu\text{g L}^{-1}$ at a signal-to-noise ratio of 3, respectively. It demonstrated that the DL ($0.2 \mu\text{g L}^{-1}$) of the TPA-MoS₂/GC electrode was much lower than the standard for detecting tap water quality in China (GB 5749-2006, Cd 0.005 mg L^{-1}). The DL of the TPA-MoS₂/GC electrode was superior to that of the chitosan-multiwalled carbon nanotubes/GC electrode ($0.4 \mu\text{g L}^{-1}$, electrodeposition Bi³⁺ for 300 s). Reproducibility was another important parameter of sensors. As shown in Fig. 5, 10 repeated measurements for $50 \mu\text{g L}^{-1} \text{Cd}^{2+}$ had excellent reproducibility with relative standard deviation (RSD) of 0.41%. Therefore, this prepared sensor could be potentially applied to rapid detection of Cd²⁺ in the aquaculture water.

Table 2 Analysis of Cd²⁺ by different reported methods

Electrode	Detection limit ($\mu\text{g L}^{-1}$)	Deposition time (s)	Ref.
TPA-MoS ₂	0.17	210	Present work
Bi-coated GC	50	120	54
Bi-GP-CNT/GC	0.6	150	55
Bi-Nafion/GC	1.39	120	56

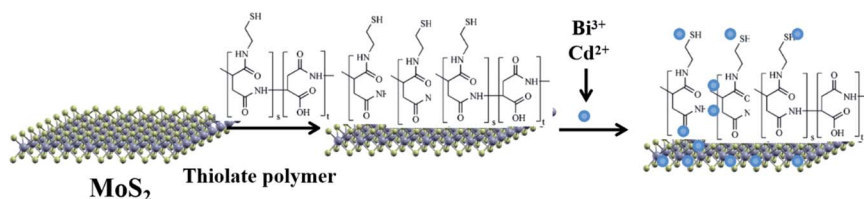
3.4. The TPA-MoS₂/GC electrode testing aquaculture water samples

This sensor was used for detecting Cd²⁺ in the aquaculture water samples. The test samples were directly added into the bottle without enrichment, and then detected by the TPA-MoS₂/GC electrode according to previous procedure. The Cd²⁺ standard solution ($50 \mu\text{g L}^{-1}$) was added into the bottle to calculate and correct the results of the test samples. A $0.22 \mu\text{m}$ filter membrane was used for extracting 6 aquaculture water samples. As shown in Table 1, the Cd²⁺ concentration in aquaculture water A and aquaculture water D were 1.9 and $2.1 \mu\text{g L}^{-1}$, respectively. For other samples (sample B, sample C, sample E and sample F), there was no Cd²⁺. The concentration of aquaculture water samples was validated by AAS. It indicated that the sensor was a reliable and sensitive tool for the rapid detection of Cd²⁺ in real samples.

The good sensing performance of the TPA-MoS₂/GC electrode might be attributed to the following factors: the large surface area of MoS₂ greatly increased the active surface area of the GC electrode for Cd²⁺ adsorption, and the abundant thiol group of TPA significantly increased the binding sites for Cd²⁺ (Scheme 1); the good electric conductivity of MoS₂ could improve the electron transfer rate between the surface of the GC electrode and Cd²⁺ in the bulk solution; and the 2D nanostructure provided a very broad space for the easy transfer of Cd²⁺. Based on these reasons, the TPA-MoS₂/GC electrode exhibited low DL and good reproducibility (Table 2). Thus, the TPA-MoS₂/GC electrode showed greater potential for use as a “pre-alarm” tool.

4. Conclusion

In summary, the TPA-MoS₂/GC electrode was prepared for Cd²⁺ detection in the aquaculture water samples. It showed that the TPA-MoS₂ nanocomposites offered significant advantages, such as low DL and good reproducibility. The good results were



Scheme 1 Schematic diagram of Cd²⁺ absorption on TPA-MoS₂ nanocomposites.



attributed to the large specific surface area, abundant thiol group from TPA and high electrical conductivity of the TPA-MoS₂ nanocomposites. Furthermore, the bismuth electrode provided more environmental-friendly electrode for researcher health. The TPA-MoS₂/GC electrode possessed greatly potential for its application in the on-site rapid analysis of Cd²⁺ in the water or food.

Conflicts of interest

There is no conflicts of interest.

Acknowledgements

This work was supported by the Central Public-interest Scientific Institution Basal Research Fund, CAFS (No. 2020GH09 and 2020TD75).

References

- 1 Q. Zhao, H. Wang, Y. Du, H. J. Rogers, Z. Wu, S. Jia, X. Yao, F. Xie and W. Liu, *J. Agric. Food Chem.*, 2020, **68**, 1974–1985.
- 2 C. O. Ogunkunle, D. A. Odulaja, F. O. Akande, M. Varun, V. Vishwakarma and P. O. Fatoba, *J. Biotechnol.*, 2020, **310**, 54–61.
- 3 Y. S. Kim, M. Y. Song, J. S. Kim, D. S. Rha, Y. J. Jeon, J. E. Kim, H. Y. Ryu, I. J. Yu and K. S. Song, *Toxicol. Res.*, 2009, **25**, 140–146.
- 4 A. Al-Ghafari, E. Elmorsy, E. Fikry, M. Alrowaili and W. G. Carter, *PLoS One*, 2019, **14**, e0225341.
- 5 L. Knani, M. Venditti, S. Kechiche, M. Banni, I. Messaoudi and S. Minucci, *Toxicol. Mech. Methods*, 2019, 1–9.
- 6 X. Li, R. Li, J. Yan, Y. Song, J. Huo, Z. Lan, J. Chen and L. Zhang, *J. Appl. Toxicol.*, 2020, **40**, 352–362.
- 7 P. Chen, X. Duan, M. Li, C. Huang, J. Li, R. Chu, H. Ying, H. Song, X. Jia, Q. Ba and H. Wang, *Toxicol. Appl. Pharmacol.*, 2016, **310**, 150–158.
- 8 IARC Monogr. Eval. Carcinog. Risk Chem. Man, 1976, **11**, 1–293.
- 9 N. Degraeve, *Mutat. Res.*, 1981, **86**, 115–135.
- 10 B. Yan, M. P. Isaure, S. Mounicou, H. Castillo-Michel, W. De Nolf, C. Nguyen and J. Y. Cornu, *Environ. Pollut.*, 2020, **260**, 113987.
- 11 N. Zhang, K. Shen, X. Yang, Z. Li, T. Zhou, Y. Zhang, Q. Sheng and J. Zheng, *Food Chem.*, 2018, **264**, 462–470.
- 12 N. A. Kasa, E. Akkaya, B. T. Zaman, G. Cetin and S. Bakirdere, *Environ. Monit. Assess.*, 2018, **190**, 589.
- 13 M. Shirani, S. Habibollahi and A. Akbari, *Food Chem.*, 2019, **281**, 304–311.
- 14 O. Zverina, J. Kuta, P. Coufalik, P. Koseckova and J. Komarek, *Food Chem.*, 2019, **298**, 125084.
- 15 B. Paull, E. Twohill and W. Bashir, *J. Chromatogr. A*, 2000, **877**, 123–132.
- 16 S. Tanikkul, J. Jakmune, S. Lapanantnoppakhun, M. Rayanakorn, P. Sooksamiti, R. E. Synovec, G. D. Christian and K. Grudpan, *Talanta*, 2004, **64**, 1241–1246.
- 17 M. Xu, R. Ma, C. Huang, G. Shi, T. Zhou and J. Deng, *Anal. Chim. Acta*, 2020, **1096**, 174–183.
- 18 M. Zhou, Y. Wu, J. Zhang, Y. Zhang, X. Chen, J. Ye and S. Wang, *Anal. Sci.*, 2019, **35**, 283–287.
- 19 K. M. Hassan, G. M. Elhaddad and M. AbdelAzzem, *Mikrochim. Acta*, 2019, **186**, 440.
- 20 Y. Chen, D. Zhang, D. Wang, L. Lu, X. Wang and G. Guo, *Talanta*, 2019, **202**, 27–33.
- 21 L. Wu, X. Fu, H. Liu, J. Li and Y. Song, *Anal. Chim. Acta*, 2014, **851**, 43–48.
- 22 M. F. De Oliveira, A. A. Saczk, L. L. Okumura, A. P. Fernandes, M. De Moraes and N. R. Stradiotto, *Anal. Bioanal. Chem.*, 2004, **380**, 135–140.
- 23 E. Nagles, V. Arancibia, C. Rojas and R. Segura, *Talanta*, 2012, **99**, 119–124.
- 24 Z. J. Suturovic, S. Z. Kravic, Z. S. Stojanovic, A. D. Durovic and T. Z. Brezo-Borjan, *J. Anal. Methods Chem.*, 2019, **2019**, 3579176.
- 25 M. Mladenov, V. Mirceski, I. Gjorgoski and B. Jordanoski, *Bioelectrochemistry*, 2004, **65**, 69–76.
- 26 N. B. Li, W. W. Zhu, J. H. Luo and H. Q. Luo, *Analyst*, 2012, **137**, 614–617.
- 27 H. Li, J. Li, Z. Yang, Q. Xu, C. Hou, J. Peng and X. Hu, *J. Hazard. Mater.*, 2011, **191**, 26–31.
- 28 S. Cerovac, V. Guzsvany, Z. Konya, A. M. Ashrafi, I. Svancara, S. Roncevic, A. Kukovecz, B. Dalmacija and K. Vytras, *Talanta*, 2015, **134**, 640–649.
- 29 J. Tu, Y. Gan, T. Liang, Q. Hu, Q. Wang, T. Ren, Q. Sun, H. Wan and P. Wang, *Front. Chem.*, 2018, **6**, 333.
- 30 Q. Wang, Z. Zhou, Y. Zhai, L. Zhang, W. Hong, Z. Zhang and S. Dong, *Talanta*, 2015, **141**, 247–252.
- 31 J. Liu, L. Han, T. Wang, W. Hong, Y. Liu and E. Wang, *Chem.–Asian J.*, 2012, **7**, 2824–2829.
- 32 J. M. Lim, D. Kim, Y. G. Lim, M. S. Park, Y. J. Kim, M. Cho and K. Cho, *J. Mater. Chem. A*, 2015, **3**, 7066–7076.
- 33 L. O. Jones, M. A. Mosquera, M. A. Ratner and G. C. Schatz, *ACS Appl. Mater. Interfaces*, 2020, **12**, 4607–4615.
- 34 Z. Han, Z. Tang, K. Jiang, Q. Huang, J. Meng, D. Nie and Z. Zhao, *Biosens. Bioelectron.*, 2020, **150**, 111894.
- 35 Z. Chen, C. Liu, J. Liu, J. Li, S. Xi, X. Chi, H. Xu, I. H. Park, X. Peng, X. Li, W. Yu, X. Liu, L. Zhong, K. Leng, W. Huang, M. J. Koh and K. P. Loh, *Adv. Mater.*, 2020, **32**, e1906437.
- 36 F. I. Alzakia, W. Sun, S. J. Pennycook and S. C. Tan, *ACS Appl. Mater. Interfaces*, 2020, **12**, 3096–3103.
- 37 W. Lu, P. Yu, M. Jian, H. Wang, H. Wang, X. Liang and Y. Zhang, *ACS Appl. Mater. Interfaces*, 2020, **12**, 16822.
- 38 C. L. C. Rodriguez, P. A. R. Munoz, K. Z. Donato, L. Seixas, R. K. Donato and G. J. M. Fechine, *Phys. Chem. Chem. Phys.*, 2020, **22**, 1457–1465.
- 39 E. M. Alexeev, D. A. Ruiz-Tijerina, M. Danovich, M. J. Hamer, D. J. Terry, P. K. Nayak, S. Ahn, S. Pak, J. Lee, J. I. Sohn, M. R. Molas, M. Koperski, K. Watanabe, T. Taniguchi, K. S. Novoselov, R. V. Gorbachev, H. S. Shin, V. I. Fal'ko and A. I. Tartakovskii, *Nature*, 2019, **567**, 81–86.
- 40 M. M. Furchi, A. Pospischil, F. Libisch, J. Burgdorfer and T. Mueller, *Nano Lett.*, 2014, **14**, 4785–4791.



- 41 S. Tsoi, P. Dev, A. L. Friedman, R. Stine, J. T. Robinson, T. L. Reinecke and P. E. Sheehan, *ACS Nano*, 2014, **8**, 12410–12417.
- 42 R. Zhang, Y. Qin, P. Liu, C. Jia, Y. Tang and H. Wang, *ChemSusChem*, 2020, **13**, 1354–1365.
- 43 Z. Wang, Q. Tu, S. Zheng, J. J. Urban, S. Li and B. Mi, *Nano Lett.*, 2017, **17**, 7289–7298.
- 44 Y. Yuan, H. Lv, Q. Xu, H. Liu and Y. Wang, *Nanoscale*, 2019, **11**, 4318–4327.
- 45 Y. Gao, K. Huang, X. Wu, Z. Hou and Y. Liu, *J. Alloys Compd.*, 2018, **741**, 174–181.
- 46 A. Dankert, L. Langouche, M. V. Kamalakar and S. P. Dash, *ACS Nano*, 2014, **8**, 476–482.
- 47 H. Liu, M. Si, S. Najmaei, A. T. Neal, Y. Du, P. M. Ajayan, J. Lou and P. D. Ye, *Nano Lett.*, 2013, **13**, 2640–2646.
- 48 W. Zhang, Y. Song, S. J. He, L. Shang, R. N. Ma, L. P. Jia and H. S. Wang, *Nanoscale*, 2019, **11**, 20910–20916.
- 49 P. Bi and W. Hong, *Mater. Chem. Phys.*, 2020, **255**, 123577–123583.
- 50 D. Ma, J. Yu, W. Yin, X. Zhang, L. Mei, Y. Zu, L. An and Z. Gu, *Chemistry*, 2018, **24**, 15868–15878.
- 51 Y. Chen, X. Wu and K. Huang, *Sens. Actuators, B*, 2018, **270**, 179–186.
- 52 H. L. Shuai, X. Wu and K. J. Huang, *J. Mater. Chem. B*, 2017, **5**, 5362–5372.
- 53 A. Gupta, V. Arunachalam and S. Vasudevan, *J. Phys. Chem. Lett.*, 2016, **7**, 4884–4890.
- 54 J. Wang, J. Lu, S. B. Hocevar, P. A. Farias and B. Ogorevc, *Anal. Chem.*, 2000, **72**, 3218–3222.
- 55 X. Xuan and J. Y. Park, *Sens. Actuators, B*, 2018, **255**, 1220–1227.
- 56 J. Li, J. Zhang, H. Wei and E. Wang, *Analyst*, 2009, **134**, 273–277.

