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Thiolated poly(aspartic acid)-functionalized two-dimensional MoS₂, chitosan and bismuth film as a sensor platform for cadmium ion detection†

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To address this critical challenge, it is predicted that controllable surface modification of MoS_2 could significantly improve the sensitivity of MoS_2 based sensor.^{51,52} Thus, in this work, thiolated polymers are chemically adsorbed on electron-rich MoS_2 through covalent bonding. The sulfur on the MoS_2 surface plays as a soft Lewis base showing a high affinity for heavy metal ions (e.g., Bi^{3+} , Cd^{2+}). MoS_2 demonstrates both a high adsorption capacity due to abundant sulfur adsorption sites and fast kinetics due to easy access to these sites. Therefore, thiolated polymer modified MoS_2 plays as one of the most effective platforms for detecting Cd^{2+} . It shows that this sensor possesses superior stability, sensitivity and LOD. The proposed sensor can be used as an environmental-friendly “pre-alarm” tool for rapid determining Cd^{2+} in aquaculture water, Chinese drug, food and beverage.

2. Experimental section

2.1. Materials and solutions

Cadmium chloride, bismuth chloride, and other chemicals are from Tianjin Kermel (Tianjin, China). Thiolated poly(aspartic acid) is obtained from Covestro company (Shanghai, China). 100 mmol L^{-1} pH 6.5 acetate buffer is used as the electrolyte in electrochemical experiments.

2.2. Apparatus

A JEM-2100 instrument (Japan) is used for obtaining transmission electron microscopy (TEM) images. Scan electron microscopy (SEM) images are obtained by Zeiss Sigma 300 (Germany). A CHI 660B Electrochemical Workstation (Shanghai, China) is used for testing Cd^{2+} through DPASV. A functionalized MoS_2 and chitosan film modified glassy carbon (GC) electrode is the working electrode, the Ag/AgCl electrode is the reference electrode, and the platinum wire is the auxiliary electrode. The concentration of Cd^{2+} is also confirmed through atomic absorption spectrometry (AAS, Jena, Germany).

2.3. Preparation of MoS_2 and modification of GC electrode

MoS_2 nanosheets are prepared by an ultrasonic exfoliation method as literature reported.⁵³ 1.0 g MoS_2 powder is added into 10 mL *N*-methyl-pyrrolidone solution under ultrasonic wave for 2 h at 0 °C. After centrifugation at 7000 rpm for 30 min, the

prepared MoS_2 nanosheets is dispersed in the supernatant. The 1 mL of 1 mg mL^{-1} prepared MoS_2 is mixed with 1 mL 2 mg mL^{-1} thiolated poly(aspartic acid) (TPA), and vibrated by vortex for 12 h at room temperature.

The GC electrode is polished by alumina powder (0.05 μm) and washed by ethanol for three times, respectively. Then, 5 μL 0.6 mg mL^{-1} functionalized MoS_2 solution, 5 μL chitosan (8 wt%) and 10 μL Milli-Q water are mixed. Finally, 5 μL mixture is immobilized onto the GC electrode to obtain a TPA- MoS_2 /GC electrode.

2.4. Cd^{2+} detection by TPA- MoS_2 /GC electrode

Bismuth film is plated on the TPA- MoS_2 /GC electrode through the electrodeposition of 10 mg L^{-1} Bi^{3+} for 210 s with stirring. After coating bismuth film, Cd^{2+} is monitored by the TPA- MoS_2 /GC electrode in 8 mL 100 mmol L^{-1} acetate buffer (pH 6.5).

3. Results and discussion

3.1. TEM characterization of MoS_2

MoS_2 was obtained through ultrasonic exfoliation method. Fig. 1 showed representative TEM images of the MoS_2 nanosheets (A), the functionalized MoS_2 (B) and the functionalized MoS_2 deposited with bismuth and cadmium (C). It suggested that the prepared MoS_2 nanosheets were very thin and clear. After TPA non-covalent modified on the MoS_2 nanosheets, the original morphology of the MoS_2 nanosheets was retained in the composites. Fig. 1C is a TEM image of the TPA- MoS_2 -Bi-Cd nanocomposites and clearly showed the modification of the MoS_2 by Bi and Cd nanocluster. As shown in Fig. S1,[†] TPA- MoS_2 surface becomes rough and there are lots of nanoparticles on the surface of TPA- MoS_2 after detection. It is consistent with TEM images. FTIR was also used to confirm the formation of TPA on the MoS_2 nanosheets. As shown in Fig. 2, the peak of 2550 cm^{-1} was the SH group of TPA. After TPA modified on the MoS_2 nanosheets, the absorption bands at 1390 (symmetric stretching of $-\text{COOH}$), 1600 (asymmetric stretching of $-\text{COOH}$), and 3000–3500 cm^{-1} (NH_3^+ stretch) became weak. The change in their dipole moment was due to TPA modification on the MoS_2 nanosheets. These results showed the successful modification of TPA on the MoS_2 nanosheets. X-ray photoelectron spectroscopy (XPS) was used to determine the chemical composition and states of the TPA- MoS_2

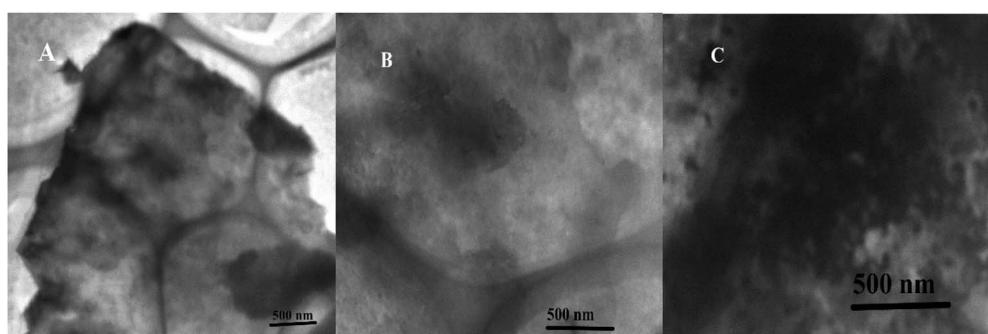
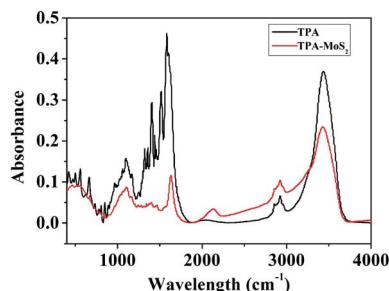


Fig. 1 TEM image of MoS_2 nanosheets (A), the TPA functionalized MoS_2 (B), and the TPA- MoS_2 with bismuth and cadmium (C).



Fig. 2 FTIR image of TPA and TPA–MoS₂.

with cadmium. As shown in Fig. S2A,† two peaks at 229 and 232 eV energy values ascribed to Mo core peaks, and the peak at 226 eV ascribed to S core peak. Fig. S2B† showed two peaks at 163 and 161.7 eV. These binding energy values are consistent with the expected charge states of Cd²⁺, Mo⁴⁺ and S²⁻. Furthermore, the specific surface area of the MoS₂ nanosheets was estimated by the Brunauer–Emmett–Teller (BET) method. Its specific surface area achieved to be 603 m² g⁻¹. The high effective surface area and high-quality of MoS₂ could provide good performances for GC electrode.

3.2. Electrochemical characterization of the TPA–MoS₂-modified GC electrode

Cyclic voltammetry (CV) was an electrochemical measurement method which could be used for monitoring the preparation procedure of GC electrode. As shown in Fig. 3, the immobilization process of the TPA–MoS₂/GC electrode was performed. Fig. 3 displayed the CVs of the bare GC, MoS₂/GC and TPA–MoS₂/GC electrodes in 2 mmol L⁻¹ K₃[Fe(CN)₆]/K₄[Fe(CN)₆] solution. The CV of the bare GC electrode was highest among these electrodes. It demonstrated that the electron transfer rate between [Fe(CN)₆]^{3-/-4-} and the bare electrode was very fast. The CV of the TPA–MoS₂/GC electrode was the lowest, which demonstrated that a chitosan film and TPA–MoS₂ had covered on the surface of the bare GC electrode. Without modification of

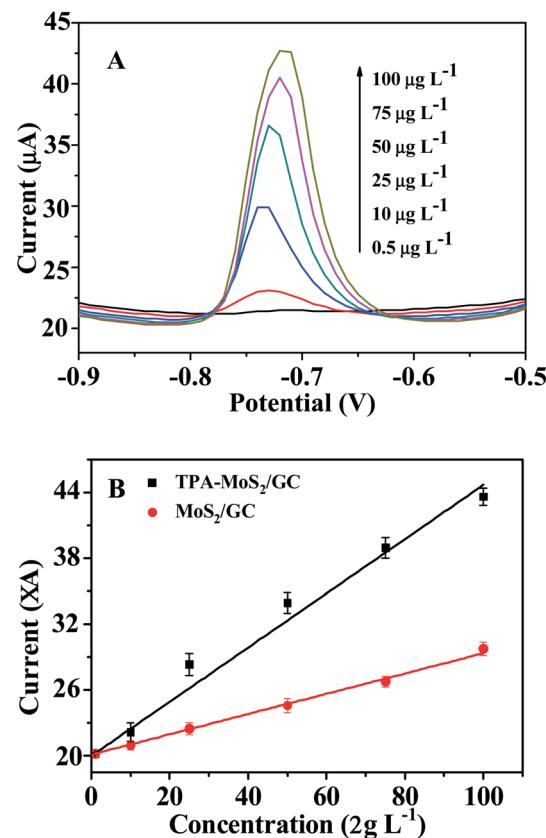
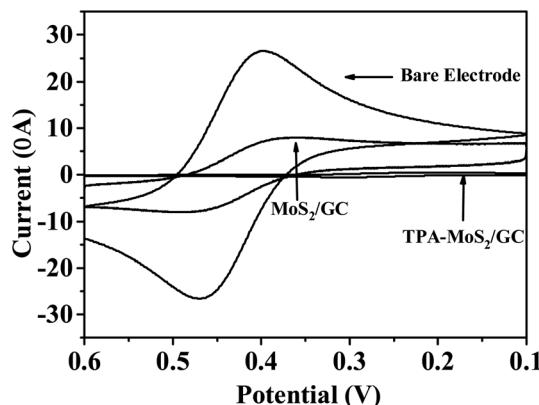


Fig. 4 (A) DPASVs obtained for different concentrations of Cd²⁺ using a bismuth-coated TPA–MoS₂/GC electrode; from bottom to top, 0.5, 10, 25, 50, 75, 100 $\mu\text{g L}^{-1}$ with 10 mg L^{-1} Bi³⁺ electrodeposited for 210 s under stirring in 100 mmol L^{-1} acetate buffer (pH 5.0). (B) Calibration curves of MoS₂/GC (red color) and TPA–MoS₂/GC (black color) electrodes for Cd²⁺.

Fig. 3 Cyclic voltammograms of bare GC, MoS₂/GC and TPA–MoS₂/GC electrodes in 2 mmol L⁻¹ [Fe(CN)₆]^{3-/-4-} at a scan rate of 100 mV s⁻¹.

TPA, the MoS₂ nanosheets possessed good conductivity, and the CV of the chitosan–MoS₂/GC electrode was higher than the TPA–MoS₂/GC electrode. The results might be due to the introduction of TPA, which played a key role in improving the binding sites of Cd²⁺. It also showed that the TPA–MoS₂ and chitosan film had covered on the GC electrode.

3.3. The TPA–MoS₂-modified GC electrode for Cd²⁺ analysis

In this study, DPASV was used for detecting Cd²⁺. Fig. 4A showed that the DPASVs of the MoS₂/GC and the TPA–MoS₂/GC electrodes increased with the concentrations of Cd²⁺ from 0.5 to 100 $\mu\text{g L}^{-1}$ in 100 mmol L^{-1} acetate buffer (pH 6.5) with 10 mg L^{-1} Bi³⁺ electrodepositing for 210 s. After the concentrations of Cd²⁺ achieved 150, 200 and 250 $\mu\text{g L}^{-1}$, the DPASVs of these electrodes increased unobvious. The DPASVs response signals increased linear from 0.5 to 100 $\mu\text{g L}^{-1}$ then become flat. As shown in Fig. 4B, the current peaks increased linearly with Cd²⁺ concentration, and the correlation coefficients was approximate 0.99. The sensitivity of the MoS₂/GC and the TPA–MoS₂/GC electrodes were 101.8 and 178.2 $\text{mA cm}^{-2} \text{m}^{-1}$, respectively. Furthermore, the detection limit (DL) was an



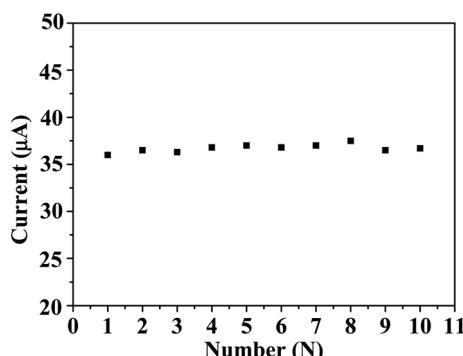


Fig. 5 The stability of 10 repetitive measurements of $50 \mu\text{g L}^{-1} \text{Cd}^{2+}$ in 0.1 mol L^{-1} acetate buffer (pH 5.0) containing $3 \text{ mg L}^{-1} \text{Bi}^{3+}$.

Table 1 Comparison of Cd^{2+} concentration in different aquaculture water samples by the sensor and AAS method

Sample	Concentration ($\mu\text{g L}^{-1}$) (sensor method)	Concentration ($\mu\text{g L}^{-1}$) (AAS method)	Difference ^a (%)
A	1.9	1.7	11.76
B	—	—	—
C	—	—	—
D	2.1	2.4	-12.5
E	—	—	—
F	—	—	—

^a Difference = $(\text{sensor}(\text{value}) - \text{AAS}(\text{value})) / \text{AAS}(\text{value}) \times 100\%$.

important parameter as sensors. The DL of the TPA–MoS₂/GC and the MoS₂/GC electrodes achieved $0.2 \mu\text{g L}^{-1}$ and $0.5 \mu\text{g L}^{-1}$ at a signal-to-noise ratio of 3, respectively. It demonstrated that the DL ($0.2 \mu\text{g L}^{-1}$) of the TPA–MoS₂/GC electrode was much lower than the standard for detecting tap water quality in China (GB 5749-2006, Cd 0.005 mg L^{-1}). The DL of the TPA–MoS₂/GC electrode was superior to that of the chitosan-multiwalled carbon nanotubes/GC electrode ($0.4 \mu\text{g L}^{-1}$, electrodepositing Bi³⁺ for 300 s). Reproducibility was another important parameter of sensors. As shown in Fig. 5, 10 repeated measurements for $50 \mu\text{g L}^{-1} \text{Cd}^{2+}$ had excellent reproducibility with relative standard deviation (RSD) of 0.41%. Therefore, this prepared sensor could be potentially applied to rapid detection of Cd²⁺ in the aquaculture water.

Table 2 Analysis of Cd²⁺ by different reported methods

Electrode	Detection limit ($\mu\text{g L}^{-1}$)	Deposition time (s)	Ref.
TPA–MoS ₂	0.17	210	Present work
Bi-coated GC	50	120	54
Bi-GP-CNT/GC	0.6	150	55
Bi-Nafion/GC	1.39	120	56

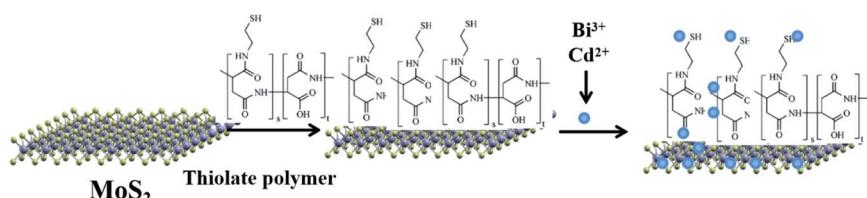
3.4. The TPA–MoS₂/GC electrode testing aquaculture water samples

This sensor was used for detecting Cd²⁺ in the aquaculture water samples. The test samples were directly added into the bottle without enrichment, and then detected by the TPA–MoS₂/GC electrode according to previous procedure. The Cd²⁺ standard solution ($50 \mu\text{g L}^{-1}$) was added into the bottle to calculate and correct the results of the test samples. A $0.22 \mu\text{m}$ filter membrane was used for extracting 6 aquaculture water samples. As shown in Table 1, the Cd²⁺ concentration in aquaculture water A and aquaculture water D were 1.9 and $2.1 \mu\text{g L}^{-1}$, respectively. For other samples (sample B, sample C, sample E and sample F), there was no Cd²⁺. The concentration of aquaculture water samples was validated by AAS. It indicated that the sensor was a reliable and sensitive tool for the rapid detection of Cd²⁺ in real samples.

The good sensing performance of the TPA–MoS₂/GC electrode might be attributed to the following factors: the large surface area of MoS₂ greatly increased the active surface area of the GC electrode for Cd²⁺ adsorption, and the abundant thiol group of TPA significantly increased the binding sites for Cd²⁺ (Scheme 1); the good electric conductivity of MoS₂ could improve the electron transfer rate between the surface of the GC electrode and Cd²⁺ in the bulk solution; and the 2D nanostructure provided a very broad space for the easy transfer of Cd²⁺. Based on these reasons, the TPA–MoS₂/GC electrode exhibited low DL and good reproducibility (Table 2). Thus, the TPA–MoS₂/GC electrode showed greater potential for use as a “pre-alarm” tool.

4. Conclusion

In summary, the TPA–MoS₂/GC electrode was prepared for Cd²⁺ detection in the aquaculture water samples. It showed that the TPA–MoS₂ nanocomposites offered significant advantages, such as low DL and good reproducibility. The good results were



Scheme 1 Schematic diagram of Cd²⁺ absorption on TPA–MoS₂ nanocomposites.



attributed to the large specific surface area, abundant thiol group from TPA and high electrical conductivity of the TPA-MoS₂ nanocomposites. Furthermore, the bismuth electrode provided more environmental-friendly electrode for researcher health. The TPA-MoS₂/GC electrode possessed greatly potential for its application in the on-site rapid analysis of Cd²⁺ in the water or food.

Conflicts of interest

There is no conflicts of interest.

Acknowledgements

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