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The synthesis and biological activity of marine alkaloid derivatives and analogues

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The ocean is the origin of life, with a unique ecological environment, which has given birth to a wealth of marine organisms. The ocean is an important source of biological resources and tens of thousands of monomeric compounds have been separated from marine organisms using modern separation technology. Most of these monomeric compounds have some kind of biological activity that has attracted extensive attention from researchers. Marine alkaloids are a kind of compound that can be separated from marine organisms. They have complex and special chemical structures, but at the same time, they can show diversity in biological activities. The biological activities of marine alkaloids mainly manifest in the form of anti-tumor, anti-fungus, anti-viral, anti-malaria, and anti-osteoporosis properties. Many marine alkaloids have good medicinal prospects and can possibly be used as anti-tumor, anti-viral, and anti-fungal clinical drugs or as lead compounds. The limited amounts of marine alkaloids that can be obtained by separation, coupled with the high cytotoxicity and low selectivity of these lead compounds, has restricted the clinical research and industrial development of marine alkaloids. Marine alkaloid derivatives and analogues have been obtained *via* rational drug design and chemical synthesis, to make up for the shortcomings of marine alkaloids; this has become an urgent subject for research and development. This work systematically reviews the recent developments relating to marine alkaloid derivatives and analogues in the field of medical chemistry over the last 10 years (2010–2019). We divide marine alkaloid derivatives and analogues into five types from the point-of-view of biological activity and elaborated on these activities. We also briefly discuss the optimization process, chemical synthesis, biological activity evaluation, and structure–activity relationship (SAR) of each of these compounds. The abundant SAR data provides reasonable approaches for the design and development of new biologically active marine alkaloid derivatives and analogues.

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1. Introduction

Alkaloids are natural compounds (natural products) that contain nitrogen and have significant biological activities.¹ They generally have chemical properties similar to bases. Some nitrogenous compounds derived from plants are not alkaline, but are still classified as alkaloids because of their obvious biological activity.^{2–4} It can be seen from the chemical structure of alkaloids that most of them have a complex nitrogen heterocyclic structure, with only a few organic amine alkaloid compounds not exhibiting such a structure.^{5–9} The definition of alkaloids by the International Union of Pure and Applied Chemistry (IUPAC) is not strict and exact. For example, some vitamins, amino acids, peptides and other nitrogenous compounds derived from natural products do not belong to the

category of alkaloids.¹⁰ Alkaloids are widely distributed in nature, especially in plants, and also in animals, microorganisms and marine organisms. There are many known types of alkaloids, with around 10 000 of them having structural formulas that have not yet been fully determined.^{11,12} As there are many types of alkaloids, each of which has a different structural formula, their properties are different from each other. However, the levels of alkaloids contained in living organisms are small, and this is similar in human beings. It has long been recognized that some plants or extracts containing alkaloids can cure diseases or be used as poisons.¹³ Since 1806, when the German pharmacist Serturmer isolated morphine from poppies, there have been more than 6000 alkaloids isolated from plants and animals. Through the joint efforts of medical chemists, pharmacologists and other scholars, people have found that a variety of alkaloids have anti-tumor, anti-bacterial, anti-viral and other biological activity.^{14–16} So far, nearly 100 alkaloid compounds have been previously used or are currently being used in clinical trials. In the treatment of many diseases, many alkaloids make good medicines, but some alkaloids can be used to make pesticides for agriculture.^{17–19}

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Because of their wide range of biological activity, alkaloids are of great interest.

The ocean is the birthplace of life and is rich in biological materials, making it a huge natural products screening resource.²⁰ So far, researchers have found more than 300 000 species of marine organisms in the ocean and more than one million new species are thought to be undiscovered.^{21–24} Marine organisms are secondary metabolites, which have formed and accumulated a large number of special chemical structures, significant biological activity in the process of long-term evolution, metabolism in special environments such as high salt, high pressure, low temperature, hypoxia, and lack of light.²⁵ As such, they show significant anti-viral, anti-inflammatory and anti-tumor properties, among others. Marine alkaloids are a secondary metabolite of marine organisms, mainly derived from marine organisms such as sponges, algae, coelenterates and tunica.^{26–29} Marine alkaloids are alkaline natural products that have important biological activity, the chemical structures of which include amine nitrogen functional groups and a complex carbon skeleton ring.³⁰ The main biological activities of marine alkaloids are their anti-tumor, anti-viral, anti-malaria, anti-fungal and anti-osteoporosis properties. Many marine alkaloids may be used as anti-tumor, anti-viral and anti-fungal clinical drugs or as the lead compounds for structural modification, as they have good medicinal prospects.³¹ In recent years, marine drugs research has been paid more and more attention by scholars, especially in the field of marine alkaloid drugs. Many types of marine alkaloids have been found and extracted from marine organisms, but most often the extraction quantity is small and the efficiency is low.^{32–35} Representative compounds of these marine alkaloids are shown in Fig. 1, including 3-alkyl-pyridinium and pyridine derivatives (e.g. viscosaline, compound 1, and theonelladin C, compound 2, respectively), N-fused heterocycles such as pyrroloquinoline (makaluvamine A, compound 3), carbazoles (ellipticine, compound 4), pyridoacridines (neoamphimedine, compound 5), N-phthalimides (e.g. granulatimide, compound 10, and isogranulatimide, compound 11), pyrrolopyrimidine (rigidin A–D, compounds 12–15), bis-indole derivatives (e.g. coscinamide A–C, compounds 16–18, asterriquinone, compound 19, dragmacidin, compound 20, dragmacidin A–C, compounds 21–23, dragmacidin D, compound 24, topsentin A, B₁ and B₂, compounds 25–27, nortopsentin A–C, compounds 28–30), and 2-aminoimidazolone derivatives (e.g. dispacamide, compound 6, polyandrocarpamine A and B, compounds 7–8, leucettamine B, compound 9). In addition, these marine alkaloids have the disadvantages of high cytotoxicity, low selectivity index and low activity.³⁶ These problems thus limit the clinical research and industrial development of marine alkaloid drugs.^{37–39} Therefore, it has become an urgent task in research and development to find marine alkaloid analogues and derivatives that are economical and environmentally friendly. A large number of target compounds can be obtained for activity screening by rational drugs design, structural modification of lead compounds and chemical synthesis.⁴⁰ In this way, the shortage of natural marine alkaloids may be solved, providing a good platform for the development of marine alkaloid

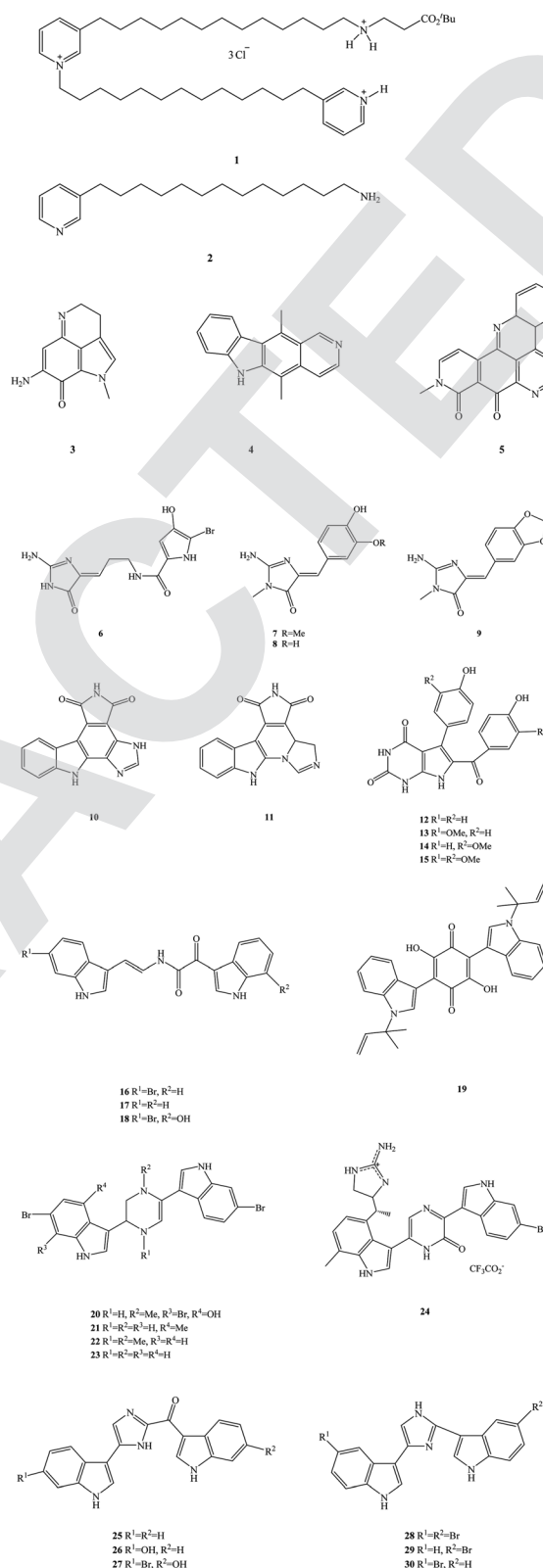


Fig. 1 The chemical structures of marine alkaloids.

derivatives and analogues.⁴¹ This work systematically reviews the recent development of marine alkaloid derivatives and analogues in the field of medical chemistry over the last 10 years



(2010–2019). We divide marine alkaloid derivatives and their analogues into five types from the point-of-view of their biological activity, including anti-tumor, anti-malarial, anti-bacterial, anti-viral (HCV), and other types of properties, elaborating on these activities. We also briefly discuss the optimization process, chemical synthesis, biological activity evaluation and structure–activity relationship (SAR) of each type of compound. The SAR is discussed, which provides a reasonable insight into the design and development of novel marine alkaloid derivatives and their analogues.

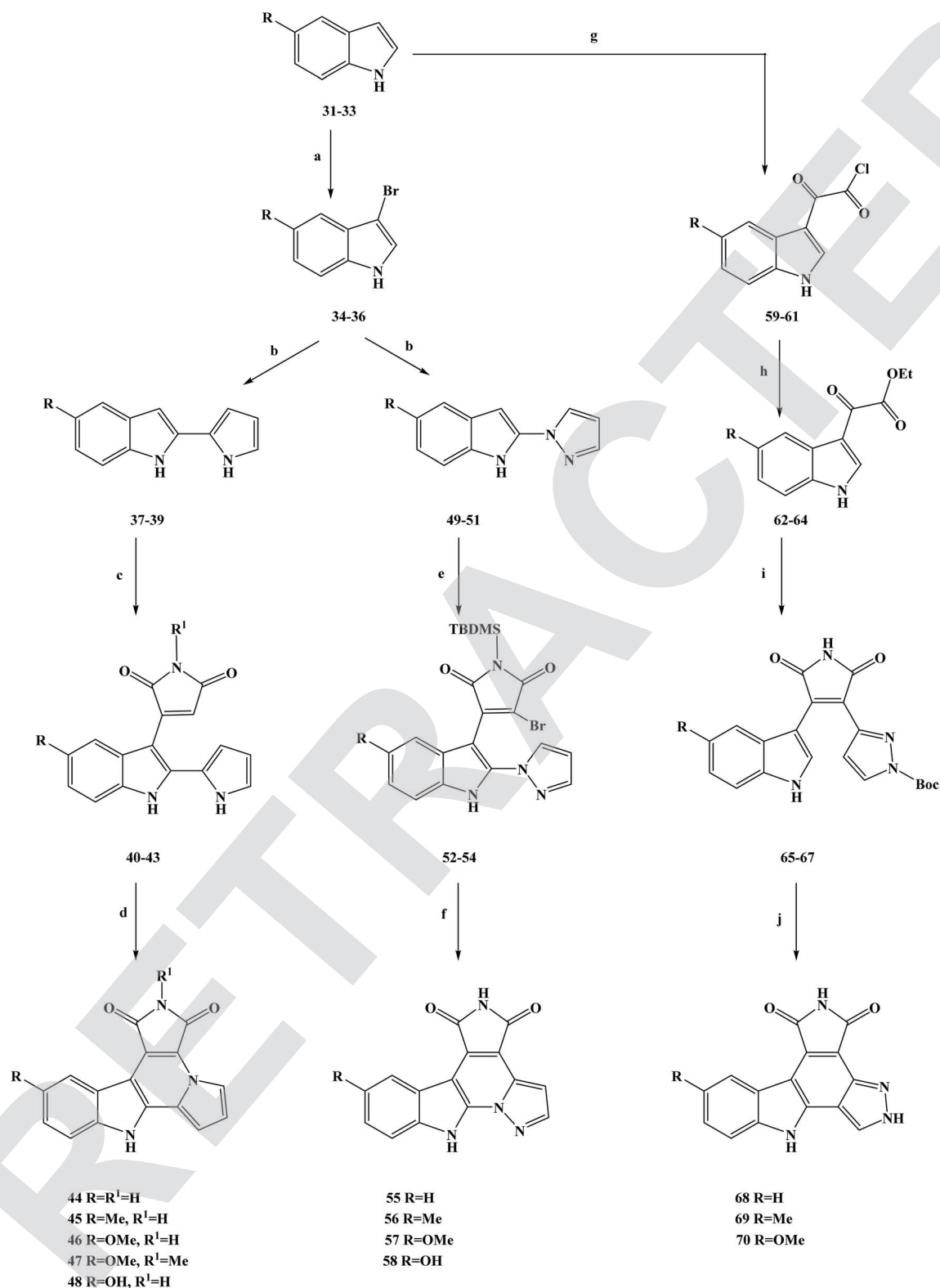
2. Synthesis and biological activities

2.1 Anti-tumor activity

In medicine, cancer refers to a malignant tumor that originates from epithelial tissue. It is the most common kind of disease related to malignant tumors.⁴² Cancer is characterized by abnormal cell differentiation and proliferation, uncontrolled growth, invasion and metastasis.^{43–45} Intercellular division is the preparation period for mitosis or meiosis in eukaryotic cells, during which the cell completes the replication of DNA molecules and the synthesis of related proteins. In cell division, the interphase is much longer than the interphase.⁴⁶ The interphase is usually divided into G₁, S and G₂ phases. The mitotic interphase is an important preparatory process for the whole process of mitosis. Studies have shown that if the G₁ and G₂ checkpoints are activated during the intermitotic period, the normal replication of DNA can be disturbed, thereby blocking subsequent cell division.⁴⁷ In this case, it can provide valuable time to repair normal DNA. A study has shown that in more than 50% of cancer cells, the G₁ checkpoint was damaged because of mutations in the p53 anti-oncogene. Relevant research has suggested that if damage to DNA and the G₂/M checkpoint inhibitor occurs simultaneously, this may selectively kill cancer cells *via* premature mitosis. This does not affect normal cell mitosis, with this combination being used in the treatment of cancer as a preliminary and promising method of chemotherapy.⁴⁸ With this biological background in mind, the marine alkaloids granulatimide (**10**) and isogranulatimide (**11**) were isolated and extracted from marine organisms showing G₂/M checkpoints inhibiting in the cell cycle.⁴⁹ The acquisition of these two marine alkaloids has attracted the attention of researchers, and they are a class of inhibitors with great research value. From the perspective of their mechanism of action, these marine alkaloids have been identified as inhibitors that can selectively inhibit checkpoint kinase 1 (Chk1), a key enzyme in the G₂/M checkpoint.^{50,51} Relevant studies have shown that the maleimide groups of these compounds play an important role in affecting their biological activity, and they can establish two hydrogen bonds with Glu⁸⁵ and Cys⁸⁷ in the ATP binding pocket of the enzyme.⁵² On this basis, Deslandes *et al.*⁵³ designed three series of granulatimide and isogranulatimide analogues for *in vitro* evaluation of anti-tumor activity, kinase inhibition and G₂/M checkpoint abrogation. It can be seen from the chemical structure of the designed target compounds that the newly designed compounds retain the skeleton structure of the lead compounds, and that the

main modification was to introduce different substituents into the benzene ring to obtain the active compounds. In the process of synthesizing the target compounds granulatimide and isogranulatimide analogues, they used the process shown in Scheme 1. In the first series of the target compounds **45–48**, the substituted indole compounds **31–33** were used as starting materials and brominated with bromine in *N,N*-dimethylformamide (DMF) to obtain the 3-bromide derivative compounds **34–36**. Compounds **34–36** were coupled with pyrrole in methylene dichloride to obtain the intermediate compounds **37–39**, which then underwent a condensation reaction with maleimide or methylmaleimide. In this step, the toluene was used as the reaction solvent and SnCl₂ as the reaction catalyst, and compounds **40–43** were obtained by Michael addition. Compounds **40–43** were cyclized and prepared by microwave irradiation with Pd as a catalyst to obtain the target compounds **44–47**. Meanwhile, the target compound **46** was demethylated in methylene dichloride with tribromo-based boron to obtain the target compound **48**. In the second series of target compounds, compounds **34–36** were used as starting materials for the reaction, and the intermediate compounds **49–51** were obtained *via* a coupling reaction with pyrazole in methylene dichloride. Compounds **49–51** underwent a condensation reaction with *N*-*t*-butyldimethylsilyl (TBDMS)-dibromo-maleimide in tetrahydrofuran (THF) to obtain the low yield intermediate compounds **52–54**. The intermediate formed in this step does not form a ring, and the TBDMS protective group does not detach during the reaction. Compounds **52–54** were cycloformed in acetonitrile under mild conditions to obtain the target compounds **55–57**. Meanwhile, the target compound **57** was demethylated in methylene dichloride with tribromo-based boron to obtain the target compound **58**. In the third series of target compounds, compounds **31–33** were first converted into compounds **62–64** in two steps by referring to related literature. Then, compounds **62–64** reacted with *N*-Boc-pyrazole-3-acetamide in THF to obtain the intermediate compounds **65–67**. Finally, compounds **65–67** in acetonitrile, under mild conditions formed a ring reaction, and then the protection with the Boc group (cracking reaction), to obtain the target compounds **68–70**. This synthesis route used cheap reagents, the procedures were relatively simple, the functional group protection was easy to achieve, and in the subsequent process deprotection was easy. In addition, microwave irradiation was used in the synthesis method, which reduced the reaction time and increased the yield. However, this synthesis route also has some shortcomings, the yield of some compounds was relatively low (3%), which brings great difficulties for its industrialization in the future. Further optimization of the process conditions is needed to improve the reaction yield and reduce the synthesis cost. In terms of their biological activity, the growth inhibition activity of the synthetic compounds was evaluated *in vitro*. The test results showed that in cancer cells, these compounds showed a degree of sensitivity and resistance to pro-apoptotic stimuli. Of the three series of target compounds, the second series had the highest average growth inhibition activity on tumor cells, with properties such as those of the target compounds **55** and **58**, where the average





Scheme 1 Synthesis of the marine alkaloid granulatimide and isogranulatimide analogue compounds **44–48**, **55–58**, and **68–70**. Reagents and conditions: (a) Br_2 , DMF, 2 h; (b) TFA, CH_2Cl_2 , r.t., 4 h; (c) $SnCl_2$, toluene, reflux, 24 h; (d) Pd, microwave 300 W, 190 °C, 15 min; (e) LiHMDS, THF, –15 °C then r.t., 12 h; (f) $h\nu$, CH_3CN , r.t., 5 h; (g) $(COCl)_2$, Et_2O , 25 °C, 3 h; (h) EtOH, Et_3N , 78 °C, 2 h; (i) t -BuOK, THF, r.t., 12 h; ii: HCl; (j) $h\nu$, CH_3CN , r.t., 5 h.



IC₅₀ growth inhibition concentrations were 11 and 16 nM, respectively. In addition, compounds **40–43** and **52–54** were found to have good anti-tumor activities during the activity screening of the intermediates, with an average IC₅₀ growth inhibition concentration of around 20 nM, among which compound **43** had the best activity (IC₅₀ = 13 nM). Studies and analyses on drug structure–activity relationships (SAR) have shown that isoprenodiamine has weak *in vitro* anti-tumor activity, while intermediates with open structures, such as the first and second series of target compounds, have good inhibitory activity. The substituents on the benzene ring also showed a certain influence on the activity, among which methoxy substituents had the best anti-tumor activity. At the same time, the anti-tumor activity of the final target compounds was decreased after the open structures of some intermediates were closed. On the basis of *in vitro* screening, the ability of the compounds to eliminate the activated G₂/M checkpoint in HCT116p53 cells was further investigated (Table 1). The test data showed that the target compound **55** had good activity (23.9), while the activity of the other compounds was low. To gain insight into the kinase selectivity of the target compounds **44–47**, **55–58** and **68–69**, serine/threonine kinases, namely CDK5/p25, CK1, CLK1, DYRK1A and GSK-3 were selected as targets for drug binding to proteins. The results of the selectivity tests showed that these target compounds had weak inhibitory activity and poor selectivity against kinases. Such results have no significant correlation with the anti-tumor activity *in vitro*, and the ability of the target compounds to inhibit other kinase proteins may be the cause of cytotoxicity. In addition, compounds **55**, **40** and **43** were quantitatively examined by electron microscopy. The results showed that compounds **55**, **40** and **43** induced cellular inhibition, which reduced cell proliferation by significantly expanding the size of cancer cells, thus reflecting the tissue changes in the actin cytoskeleton. In general, according to the existing data analysis, the target compound **55** and the intermediate compounds **40** and **43** have good anti-tumor activities. These compounds are worthy of further structural modification and other pharmacological studies, to fully understand these drugs and become ideal anti-cancer drugs as soon as possible.

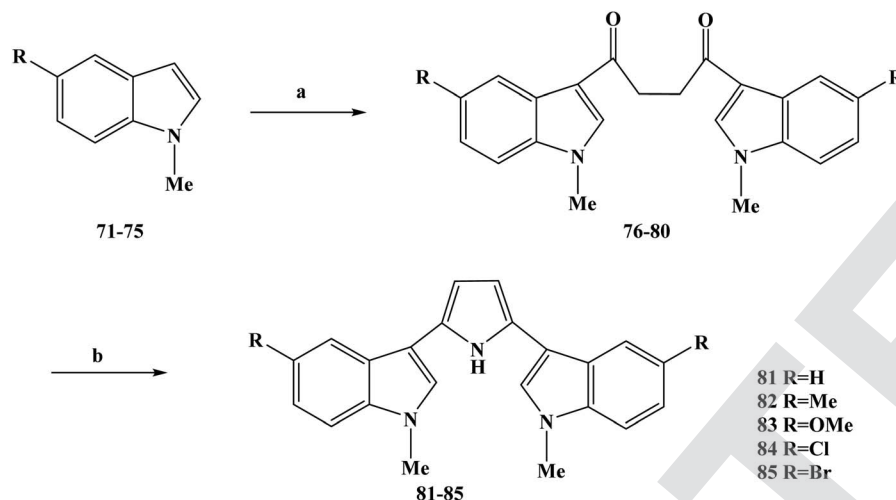
Natural products with biological activity can be isolated from marine organisms, including effective anti-tumor drugs found

in them.^{54–56} These anti-tumor drugs that are obtained from marine organisms are structurally diverse and complex. Most of these compounds with anti-tumor activity are alkaloids, which are commonly referred to as marine alkaloids.^{56–59} Among these marine alkaloids, a class of marine alkaloids with a bis-indole structure is well known.⁶⁰ This kind of bis-indole marine alkaloid is usually isolated from the metabolites of deep-sea sponges, and has been found to have good anti-inflammatory, anti-tumor, anti-viral and anti-bacterial biological activities in screening processes.⁶¹ The chemical structure of this type of bis-indole marine alkaloids is characterized by the fact that they are bound by two indoles through their 3-positions to intermediate ligands (including ring and chain structures). Related studies found that nortopsentins A–C (**28–30**) bis-indole marine alkaloids with imidazole as the five-membered ring as the connector showed IC₅₀ values of 4.5–20.7 μM to P388 cells *in vitro* during anti-tumor activity testing, while the IC₅₀ of P388 cells was 0.8–2.1 μM for *N*-methylated derivatives.^{62–64} The results of *in vitro* cytotoxicity studies indicated that the methylated derivatives significantly increased the activity of P388.⁶⁵ In order to obtain a large number of compounds, several complete synthesis methods of nortopsentins and analogues have been reported.^{66–68} Because of the abundant biological activity of marine alkaloids, they are often regarded as new drugs or lead compounds that have biological activity. Therefore, on this basis, nortopsentin was structurally modified, and dragmacidin (**20**) analogues were synthesized using six-membered cyclopyridine, pyrimidine, pyrazine and pyrazinone as intermediate ligands. These analogues have been found to have a broad spectrum of anti-tumor activity in human tumor cells. However, nortopsentin analogues have been synthesized using a five-membered-heterocyclic imidazole ring as a ligand.⁶⁹ In activity testing, it was found that these compounds had significant anti-proliferative activity, and the IC₅₀ value was at the sub-micromolar level. In the process of modifying the structure of the lead compound, one or two indoles were modified in addition to the indoles.^{70,71} In the activity testing results, it was found that the 3-indolyl-5-phenylpyridine derivative exhibited anti-proliferative activity in the range of 5–15 μM and effectively inhibited CDK1 at the 0.3–0.7 μM. On the basis of previous work, Carbone *et al.*⁷² took nortopsentine as the lead compound, modified its structure with pyrrole as the intermediate connector, and modified the substituents on the two indoles to design nortopsentine analogues with a 2,5-bis(3'-indolyl)pyrrole structure. In the process of the synthesis of the target compounds **81–85**, they selected the specific synthesis operation shown in Scheme 2. *N*-methylindoles (compounds **71–75**) were used as starting materials, and the compounds were synthesized *via* a two-step reaction. In the first step reaction, the Vilsmeier–Haack reaction was used to treat compounds **71–75** with phosphorus oxychloride and tetramethylsuccinamide, and the symmetrical intermediate compounds **76–80** were obtained. In the second step, compounds **76–80** were refluxed with ammonium acetate and acetic anhydride in acetic acid, and the corresponding 2,5-bis(3'-indolyl)pyrroles as the target compounds **81–85** were obtained after treatment. The synthesis route required only two

Table 1 Evaluation of the G₂/M checkpoint abrogation

Compounds	Mitotic index
44	9.6
55	23.9
56	13.9
58	3.9
69	0
38	5.0
39	8.3
53	5.5
Isogranulatimide	17.2
Granulatimide	19.4





Scheme 2 Synthesis of 2,5-bis(3'-indolyl)pyrrole analogue compounds **81–85**. Reagents and conditions: (a) POCl_3 , N,N,N',N' -tetramethylsuccinamide, then $55\text{--}60^\circ\text{C}$, 8 h or r.t., 20 h; (b) NH_4OAc , $(\text{CH}_3\text{CO})_2\text{O}$, CH_3COOH , reflux, 4 h.

steps to obtain the target compounds, is simple to carry out with few reaction steps, the total yields were medium to good, and the reagents used were cheap and easy to obtain, laying a solid foundation for its future industrialization. However, these synthesis methods are all classical chemical reactions, there was no innovation involved. In terms of subsequent process optimization, new synthesis methods can be considered. In the process of screening the biological activity of the synthesized target compounds **81–85**, human tumor cell lines were used for monolayer cell survival and proliferation tests were carried out to detect the anti-tumor activity of the target compounds *in vitro*. Using panels of human tumor cell lines of different source/tissue types, the anti-tumor activity, the selectivity of the tested compounds and evaluation of the eligible target compounds were analyzed for entry into preclinical studies. The *in vitro* anti-tumor activity of the target compounds **81–85** against 12 human tumor cell lines was screened using a monolayer cell survival and proliferation assay (Table 2). The results of the screening experiments showed that all of the target compounds showed cytotoxicity activity at the maximum test concentration of $100\text{ }\mu\text{M}$, with average IC_{50} values ranging from 4.4 to $0.37\text{ }\mu\text{M}$. The substituents on the indole ring in the target compounds **81–85** are electron-withdrawing and electron-donating groups. The analysis of the drug structure–activity relationship (SAR) shows that the anti-tumor activities of the electron-donating groups on the indole ring in the target

compounds were significantly higher than those of the electron-withdrawing groups. At the same time, there was also a SAR observed in which the stronger the ability of electron absorption or electron donation, the better the anti-tumor activity. In addition, the solid space of these substituents also affected the anti-tumor activity of the target compounds. These target compounds showed good *in vitro* anti-tumor activity, such as the characteristics of the target compounds **81** and **82**, so these compounds were selected for further anti-tumor activity study. The target compounds **81** and **82** were further analyzed in monolayer cultures of 42 human tumor cell lines, including 15 different types of solid tumors. The relevant test results showed that the target compounds **81** and **82** had a concentration-dependent inhibitory effect on tumor cell growth and showed obvious cytotoxicity, with average IC_{50} values of 1.54 and $0.67\text{ }\mu\text{M}$, respectively. Meanwhile, in *in vitro* cloning experiments, the target compounds **81** and **82** showed selective anti-tumor activity for human tumor transplantation, and the sensitive tumor models were dispersed among different tumor tissue types. Therefore, combined with these relevant data, the results indicate that the target compounds **81** and **82** are likely to become new anti-tumor drugs, which needs further studies, including evaluation of their *in vivo* anti-tumor activity and pharmacokinetic properties.

Cancer is one of the biggest threats to health in the world, and the number of deaths from it remains high, especially in developing countries.⁷³ In order to better treat cancer, new drugs with low toxicity and good anti-tumor activity, as well as new treatments, are urgently required.^{74–76} In the search for new anti-cancer drugs, people have set their sights on natural products. In recent years, many new compounds have been found in the secondary metabolites of marine organisms.⁷⁷ These marine organisms have been shown to be an important source of new compounds. Many new compounds obtained from marine organisms have been found to have a good inhibitory effect on tumor cell lines during the activity

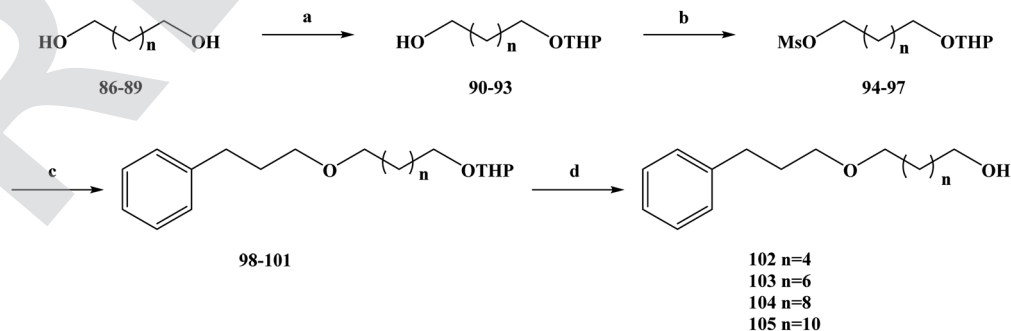
Table 2 *In vitro* activities of compounds **81–85**

Compounds	IC_{50} (μM)
81	0.37
82	0.37
83	3.4
84	3.4
85	4.4
Adriamycin	0.007



screening process. The secondary metabolites of sponges are abundant in marine organisms, the discovery of which has been widely reported by researchers.^{78–80} Some compounds were found to have anti-tumor activity during *in vitro* activity screening, and on this basis, analogues of these metabolites were synthesized.^{81–83} Many compounds were also found to have anti-tumor activity in activity evaluation. Rich in structural diversity and bioactive peptides were separated from marine sponges, compounds including the cyclopeptide perthamides G–K, glycopeptide theonellamides A–G and 3-alkylpyridine alkaloids (3-APAs) and theonelladines A–D.^{84–86} Different types of metabolites have been separated from marine sponges, and these compounds and synthetic analogues have been widely investigated in terms of their cytotoxicity activity towards human tumor cell lines. Theonelladine C (**2**) is an analogue of a 3-alkylpyridine alkaloid, which was found to have anti-protozoa activity and a pro-apoptotic effect on a human colon cancer (RKO-AS-45-1) cell line in activity screening. Therefore, on this basis, Gonçalves *et al.*⁸⁷ further modified the structure of theonelladine C to obtain target compounds with anti-tumor activity. During the structural design of the compounds, they introduced oxygen-containing carbon chains into the 3-alkyl chain of the lead compound and changed the length of the chain to alter the anti-tumor activity of the compounds.^{88,89} They designed marine alkaloid analogues of 3-alkylpyridine with alkyl chains ranging in length from 6 to 12 carbon atoms. At the same time, a hydroxyl group was attached to the end of the alkyl chain to improve the water solubility of the analogues and to provide the necessary preparation for further structural modification in the future. In the synthesis route selection of the new 3-APA analogues compounds **102–105**, they used the synthesis shown in Scheme 3. From this synthesis route, it can be seen that the key step in the synthesis of these analogues is the Williamson etherification reaction under mild conditions using phase transfer catalysis (PTC). The first step of this synthesis route was to selectively protect different diols (compounds **86–89**) to produce the corresponding tetrahydropyronic acetals (compounds **90–93**) in 74–89% yields. Compounds **90–93** were further treated in dichloromethane with methyl sulfonyl chloride to obtain the intermediate compounds **94–97**. Next, compounds **94–97** were etherized with 3-(pyrid-3-yl) propan-1-ol

using the phase transfer catalyst tetrabutylammonium bromide, and compounds **98–101** with a protective group were obtained. Finally, the protective groups of compounds **98–101** were removed with hydrochloric acid, and the target compounds **102–105** were obtained with in yields of 71–100%. The synthesis route has the advantages of being simple to carry out, low in cost and uses readily available reagents. At the same time, the yields of each step of the synthesis route are relatively high, and the reaction conditions are relatively mild, which is conducive for the use of this process in industrial production in the future. In the course of the anti-tumor biological activity study, the anti-tumor activities of all of the synthesized target compounds **102–105** and intermediates **98–101** were evaluated *in vitro* for colon cancer (RKO-AS-45-1) and uterine cancer (HeLa) (Table 3). Meanwhile, the target compounds **102–105** and intermediates **98–101** were tested in non-cancerous human lung fibroblasts (WI-26VA4) to evaluate the selectivity index (SI) of the compounds. *In vitro*, the MTT assay showed that the compounds **100**, **101**, and **104** had good anti-tumor activity. In RKO-AS-45-1 cells, the IC_{50} values were 5.1, 3.2 and 19.1 μ M, respectively. In anti HeLa cells, the IC_{50} values ranged from 4.0 to 9.4 μ M. The drug SAR analysis showed that there was a relationship between the IC_{50} value and the length of the alkyl chain of the compounds. Upon increasing the length of the carbon alkyl chain, the IC_{50} values of the two tumor cell lines decreased continuously, among which the compounds with 10 carbon atoms had the best anti-tumor activity (compounds **100** and **104**). However, as the carbon chain continued to lengthen, the anti-tumor activity decreased when the carbon chain exceeded 10 carbon atoms. In addition, the selectivity of compound **104** in the *in vitro* activity evaluation was found to be the highest among the compounds tested, with an SI of 5.18 for RKO-AS-45-1 and 11.65 for HeLa cells. On this basis, compounds **100** and **104** were selected for further related activities. Micronucleus and TUNEL tests showed that compounds **100** and **104** had mutational effects and induced apoptosis. These compounds induced DNA strand breaks and/or aneuploidy under the conditions used. The activation of p53 led to cell cycle arrest, preventing the proliferation of damaged cells and allowing DNA to replicate and undergo mitosis or induce apoptosis to eliminate irreparably damaged cells. At the same time, compound



Scheme 3 Synthesis of the 3-alkylpyridine marine alkaloid analogue compounds **102–105**. Reagents and conditions: (a) $NaHSO_4$, DHP, DMSO, hexane, 40 °C, 16 h; (b) $MsCl$, Et_3N , CH_2Cl_2 , r.t., 10 h; (c) 3-(pyrid-3-yl)propan-1-ol, $NaOH/H_2O$, $Bu_4N^+Br^-$, Et_2O , r.t., 72 h; (d) $MeOH$, HCl , r.t., 12 h.



Table 3 *In vitro* cytotoxicity activities of the compounds

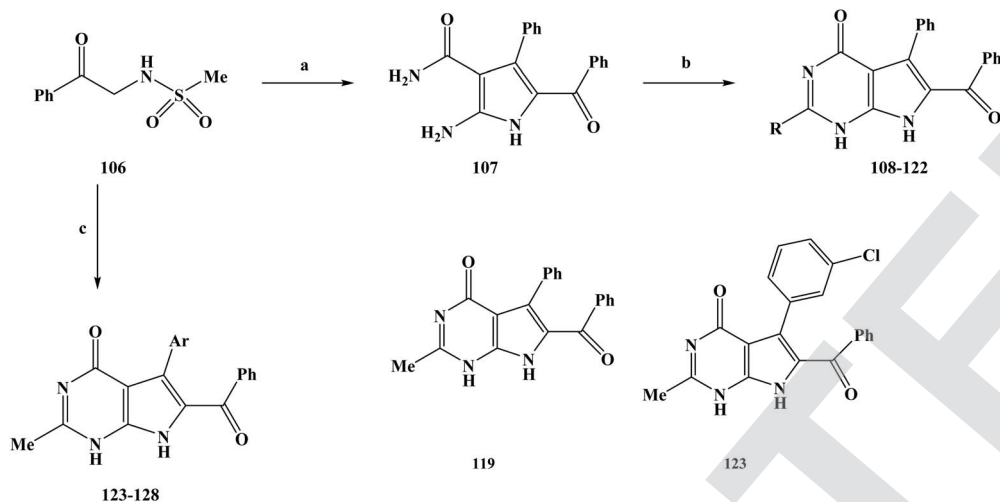
Compounds	IC ₅₀ (μM) ± SD			SI	
	PKO AS-45-1	HeLa	WL-26VA4	PKO AS-45-1	HeLa
98	>300	>300	>300	nd	nd
99	23.7 ± 1.7	8.1 ± 2.7	37.8 ± 5.0	1.59	4.66
100	5.1 ± 1.1	4.0 ± 0.8	6.4 ± 0.7	1.25	1.60
101	3.2 ± 1.7	9.4 ± 0.7	11.3 ± 1.4	3.53	1.20
102	>400	>400	>400	nd	nd
103	>300	191.8 ± 10.1	167.0 ± 10.5	nd	0.87
104	19.1 ± 4.4	8.5 ± 2.4	99.1 ± 11.2	5.18	11.65
105	131.9 ± 16.8	8.8 ± 1.9	34.1 ± 6.5	0.25	3.87
Etoposide	1.4 ± 0.6	2.7 ± 0.4	nd	—	—

104 also altered the myogenic RKO-AS-45-1 cells in the cytoskeleton. After treatment with compound **104**, the actin polymerization was changed. Drug molecules that can change the dynamics of actin could easily block the mitosis of cells, thus demonstrating the cytotoxicity of compound **104** to block mitosis. Overall, according to the analysis of the existing test results, compounds **100** and **104** could be used as potential anti-tumor drug candidates, or as lead compounds for structural modification, but this needs further research and development.

Anti-tumor drugs are a class of drugs that are used to treat cancer, including chemotherapy drugs and biological agents.⁹⁰ In recent years, the development of molecular oncology and molecular pharmacology has gradually clarified the nature of tumors.^{91–93} The invention and application of advanced technologies such as large-scale rapid screening, combinatorial chemistry and genetic engineering have accelerated the process of drug development. The research and development of anti-tumor drugs has entered a new era.⁹⁴ After years of development, many important advances have been made in the development of anti-tumor drugs. However, in the face of the most serious threats to human life and health, which account for more than 90% of malignant tumors, there is still a lack of effective and highly specific drugs.⁹⁵ On the one hand, the reflects the difficulty in developing anti-tumor drugs. On the other hand, it also means that the development of anti-tumor drugs still needs the application of new ideas, technologies and methods. At present, drug treatment has become one of the most important methods for the clinical treatment of cancer. The sales of anti-tumor drugs has been increasing year on year due to the high incidence and mortality of cancer.⁹⁶ Marine alkaloids with a pyrrole structure that are obtained from marine organisms have been found to have abundant biological activities during activity screening processes. Among these marine alkaloids, are rigidins A, B, C, and D (**12–15**). Related studies have shown that these compounds have some potential for biological activity, but few researchers have conducted in-depth studies to explore their biological activity in more detail. Frolova *et al.*⁹⁷ completed the total synthesis of rigidins A, B, C and D and carried out a comprehensive study on their biological activities. It was found that L1210 cells in a model of mouse leukemia showed good anti-proliferation activity, but the

activity of cultured human cancer cells was very low. On this basis, the structures of 7-deazahypoxanthine, 7-deazaadenine and 7-deazapurine were synthesized by modifying the 7-azathathine skeleton of the rigidins. In the study of the activities of the compounds, the anti-proliferation activity of the tumor cells was greatly improved. These compounds have also been found to damage the microtubule tissue in cancer cells by binding to the colchicine site of β -tubulin. On the basis of previous work, Scott *et al.*⁹⁸ further modified the structure of the lead compound and introduced different substituents on the C2 position to change the biological activity of the target compounds. In their structural design, two series of target compounds were designed. During the synthesis of these target compounds, **108–128**, they used the synthesis process outlined in Scheme 4. In the first series, the target compounds were obtained *via* a two-step reaction. *N*-Methylsulfonamidoacetophenone (compound **106**) was used as the starting material in the reaction. The reactant was stirred with benzaldehyde and cyanoacetamide in ethanol, and potassium carbonate was added as a base to obtain the intermediate compound **107**. In the second step, a ring formation reaction was carried out. Sodiated ethanol was prepared by reacting Na with ethanol, and compound **107** was reacted with the corresponding ester to obtain the target compounds **108–122**. In the second series, the target compounds **123–128** were synthesized in one step. Also, compound **106** was used as the starting material of the reaction, and the corresponding aldehydes, cyanoacetamide and triethyl orthoacetate (MeC(OEt)₃) were synthesized in ethanol using a one-pot method. In this step, potassium carbonate was used as a base. A reaction temperature of 90 °C was used for 24 h, followed by 150 °C for 3–6 h, with yields of 40–65% being obtained. The synthesis route has the advantages of being simple to use and low in cost with readily available reagents, and the desired target compounds could be obtained in either one- or two-step reactions. The one-pot method was applied to synthesize the target compounds or intermediates, which greatly improved the reaction efficiency and reduced the synthesis cost. However, this synthesis route also has some shortcomings. The yields of the reactions are generally low, in particular the yield of the target compound **111** was only 18%, which brings great difficulties for future industrialization. Therefore, it was necessary to further optimize the reaction conditions in the subsequent process





Scheme 4 Synthesis of the marine alkaloid rigidin analogue compounds **108–128**. Reagents and conditions: (a) benzaldehyde, cyanoacetamide, K_2CO_3 , EtOH, reflux, 14 h; (b) the corresponding ester, Na, EtOH, reflux, 10 h; (c) the corresponding aldehydes, cyanoacetamide, $MeC(OEt)_3$, K_2CO_3 , EtOH, 90 °C, 24 h, then 150 °C, 3–6 h.

optimization to increase the yield of the reaction. During the activity study, the HeLa cell line was used as a human cervical adenocarcinoma model and MCF-7 cells were used as a breast adenocarcinoma model to evaluate the anti-tumor proliferation activity of the first series of target compounds *in vitro*. *In vitro* screening showed that except for the target compound **108**, most of the other target compounds introduced at C2 exhibited anti-tumor cell proliferation activity. The activities of this series of compounds were studied based on drug SAR analysis: the C2-aryl (compounds **111** and **112**), C2-OEt (compound **114**), and branched C2-alkyl (compound **115**) activities were only in the two-digit to single-digit micromolar range; those of the linear C2-alkyl derivative compounds **116–118** showed an inhibitory effect at sub-micromolar concentrations; and the C2-methyl compound (compound **119**) could also show nanomolar tumor cell proliferation inhibition activity. When fluorine-containing groups were introduced on C2-Me, the activity gradually decreased with an increase in the space of the C2 position in the order of CH_3 (compound **119**) > CH_2F (compound **120**) > CHF_2 (compound **121**) > CF_3 (compound **123**). It can be seen from the activity results of these target compounds that the anti-proliferative activity of this series of compounds is dictated by the spatial properties of the C2 substituents rather than by the electronic properties. In the *in vitro* screening of the second series of compounds, the anti-proliferative activity of the target compounds was measured against U87 and A549. The results showed that the anti-proliferation activity of this series of compounds was lower than that of the target compound

119 (Table 4), and most of the target compounds exhibited an inhibitory effect at the sub-micromolar concentration, except for the target compound **126**. The second series of target compounds is based on target compound **119** with different substituents introduced at the C7. The result of this is that the polarity of the target compound was reduced, which damages the permeability of the cells and thus reduces the anti-tumor proliferation activity of the target compounds. On the basis of previous work, the 7-deazahypoxanthines were proposed to target to the colchicine site on β -tubulin, based on their effective inhibition of [3H] colchicine binding to tubulin. The results of docking studies on the target compounds **117** and **119** showed that linear C2-alkyl substituents on the 7-deazahypoxanthine scaffold could be accommodated in a small channel in the region of Lys352 and Asn258. The co-crystallized colchicine and target compound **119** adopt similar coordination modes in the colchicine pocket. On this basis, target compound **119** with the best activity was selected to study the effect on microtubules in cells. The results showed that **119** achieves the anti-proliferation of tumor cells by acting on microtubule-targeted proteins. According to the existing data, C2-aryl and C2-alkyl-denitroxanthine marine alkaloid analogues have good anti-tumor activities. In particular, **119** could be used as a candidate for anti-tumor drugs and has the potential to be developed into a new anti-tumor drug, which needs further research and development in the future.

There are many ways in which to discover drugs, but natural products are an important way to obtain new drugs or lead compounds.⁹⁹ Sources of natural products include compounds with certain biological activity obtained from plants, animals, and microbial metabolites using modern separation techniques. The isolation of drugs with biological activity from naturally occurring secondary metabolites has become an important field in the development of new drugs.^{100–102} These natural products have a unique chemical structure, and play an important role in drug discovery and innovation. They have irreplaceable significance in the treatment and prevention of

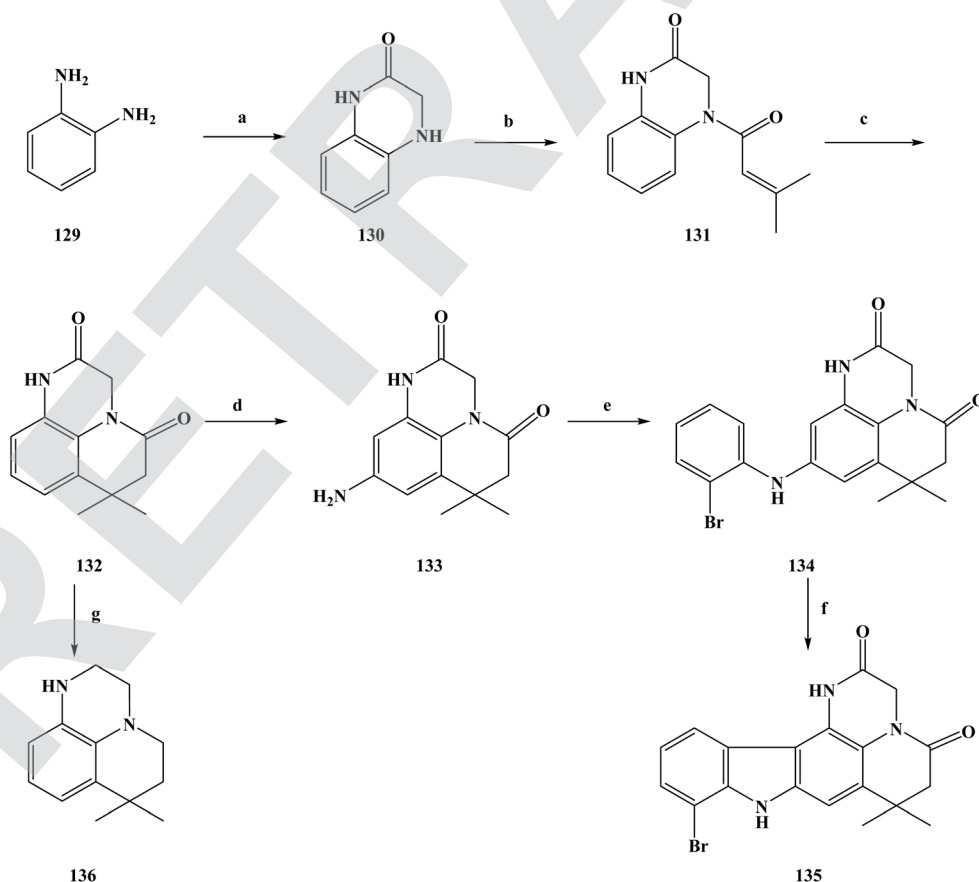
Table 4 Anti-proliferative activities of the synthesized compounds

Compounds	IC ₅₀ (μM)			
	HeLa	MCF-7	U-87	A549
119	0.029 ± 0.001	0.035 ± 0.003	0.077 ± 0.002	0.25 ± 0.01
123	0.27 ± 0.01	0.23 ± 0.00	0.90 ± 0.16	0.60 ± 0.23



human diseases. In natural product extraction and separation, alkaline compounds and alkaloids are widely investigated by researchers. Marine organisms are diverse species with diverse biological activity.¹⁰³ So far, researchers have found more than 300 000 species in the ocean, and it has been estimated that more than 1 million new ocean species have not yet been found. With this quantitative advantage, obtaining new compounds with biological activity from marine organisms has become an important pathway. At present, the research direction of people has turned towards marine organisms, and many marine alkaloids with biological activity have been separated from marine organisms.¹⁰⁴ So far, the anti-tumor alkaloids separated from marine sponges include makaluvamine A (3), ellipticine (4) and neoamphimedine (5). Between 2001 and 2010, most of the compounds separated from marine sponges have been alkaloids (marine alkaloids) and macrocyclic compounds. During this period, marine alkaloids with a pyrrolidone structure were separated from marine sponges.^{105,106} These substances were found to have significant cytotoxicity effects on a variety of tumor cells *in vitro* in the course of biological activity studies, as well as potential biological activity against topoisomerase II. Based on previous work, Boucle *et al.*¹⁰⁷ designed compounds with a praziquantel structure for anti-tumor activity. As could be seen from its chemical structure, the new

compounds were the makaluvamine A analogue compounds **135** and **136**. Meanwhile, target compounds with a pentacyclic structure have been investigated as potential anti-topoisomerase II inhibitors. In the synthesis process of the target compounds, the specific operations shown in Scheme 5 were used to synthesize the target compounds. To obtain the target compounds **135** and **136**, 1, 2-phenylenediamine (compound **129**) was used as the starting material to react with bromoacetyl bromide in THF to produce dihydroquinoxalinone compound **130**. The condensation reaction of compound **130** and dimethylacryloyl chloride in pyridine produced compound **131**, which was then cyclized in methylene dichloride using anhydrous aluminum chloride as a catalyst to obtain the tricyclic intermediate compound **132** (yield 83%). The intermediate compound **133** containing an amino group was obtained *via* the two-step nitration and catalytic hydrogenation of compound **132**, in which the nitration of was carried out with nitric acid in methylene dichloride. Then, the product was reduced *via* catalytic hydrogenation without separation to obtain compound **133**. Compound **133** was reacted with 2-bromoiodobenzene in anhydrous dioxane to obtain compound **134**, with a yield of 71%. Finally, compound **134** underwent cyclization, and *via* the Heck reaction generated pentacyclic compound **135**. In the reaction process, anhydrous dioxane was used as the solvent,



Scheme 5 Synthesis of compounds **135**–**136**. Reagents and conditions: (a) bromoacetyl bromide, THF, Et₃N, reflux, 2 days; (b) dimethylacryloyl chloride, pyridine, reflux, 2 h; (c) AlCl₃, DCM, 24 h; (d) i: HNO₃, DCM/DMF, reflux, 24 h; ii: H₂, Pd/C, DMF/DMSO, reflux, 18 h; (e) 2-bromoiodobenzene, Pd₂dba₃, Xantphos, K₂CO₃, dioxane, Schlenk, 18 h; (f) Pd(OAc)₂, Pcy₃HBF₄, K₂CO₃, DMA, reflux, 24 h; (g) BH₃/THF, THF, reflux, 2 h.



Table 5 Cytotoxicities of the synthesized compounds

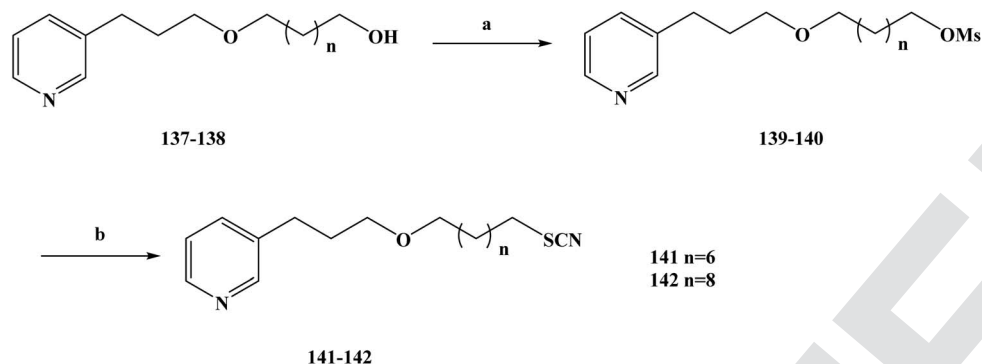
Compounds	IC ₅₀ (μM)						
	HUH-7	CaCo-2	MDA-MB-231	HCT-116	Pc-3	NCI	Fib.Hum
134	20	20	>25	25	25	>25	25
135	20	20	20	10	6	10	6
136	20	15	20	15	15	10	>25

palladium acetate as the catalyst and potassium carbonate as a base, with a yield of up to 40%. At the same time, compound **132** was reduced by boron/tetrahydrofuran and the target compound **136** was obtained in a yield of 81%. The synthesis route is simple to use, inexpensive and uses readily available raw materials, where moderate to good yields can be obtained at each step. However, this synthesis route also has some shortcomings, as it uses the precious metal Pd as a catalyst, which brings certain difficulties for the future industrialization of this method. At the same time, in the process of the compound **133** synthesis, the yield of this step was only 29%, and low yields increase the synthesis cost, which needs to be improved in the subsequent process optimization. In the process of studying the biological activity of the synthesized compounds, the anti-tumor activity of these compounds and the inhibition efficiency of topoisomerase II were evaluated. First, the compounds were tested against different cancer cell lines to assess their anti-tumor effects (Table 5). The inhibitory efficiency of the synthesized compounds was then evaluated against topoisomerase II in order to determine the mechanism of action of these newly synthesized marine alkaloid analogues. Different human cancer cell lines such as CACO-2, HCT-116, HUH-7, MDA-MB-231, PC-3, and NCI were selected for the *in vitro* cytotoxicity activity evaluation. The *in vitro* anti-tumor activity evaluation test results showed that compounds **134–136** have good anti-tumor activity, among which **136** exhibited cytotoxicity to CACO-2, HCT-116, PC-3 and NCI cell lines of 15, 15, 15 and 10 μM, respectively, but had no obvious toxicity to human fibroblasts. However, **134** and **135** also had certain inhibitory effects on human fibroblasts cell, so there was no obvious selectivity for cell inhibition. In the process of screening the anti-topoisomerase II activity of the synthesized compounds, the concentration of the compounds used for electrophoretic analysis was 100 μM. The results of the anti-topoisomerase II activity testing of the compounds showed that **135** better inhibited activity of human DNA topoisomerase II than the positive control doxorubicin at 100 μM. Other synthetic compounds showed no inhibition of topoisomerase II. From the *in vitro* anti-tumor activity of synthetic compound screening and testing of topoisomerase II activity, it was determined that compounds **135** or **136** could provide a basis for the further design of more analogues. The SAR and drug research can be used to identify compounds that have the best anti-tumor activity and greatest selectivity.

As aforementioned, natural products are compounds rich in biological activity that have the potential to be developed into

drugs.^{108–111} With the research and development of modern drugs, obtaining new drugs or lead compounds from marine organisms has become an important source. Marine sponges have been well studied and many biological activity compounds have been separated from these relatively simple organisms in recent years, including some new structural compounds.^{112–116} In the process of screening these compounds for biological activity *in vitro*, the results show that many of them have some specific biological activity and play an important role in the development of new drugs.¹¹⁷ Lithistida is a type of sponge that contains rich compounds that are structurally diverse with biological activity based on 3-alkylpyridine alkaloids (3-APAs), such as viscosaline (**1**) and theonelladin A–D.^{118–121} From the analysis of the structures of these obtained marine alkaloids, it was found that 3-APAs have a pyridine structure and an alkyl chain with variable length on the side chain, and the position of this alkyl chain is usually in the 1 or 3 position. The discovery that most marine alkaloids are cytotoxic (*i.e.* they have anti-tumor activity) has prompted wide study and interest in these.^{122–126} Previous studies have shown that in the evaluation of their biological activity, synthesized 3-APA analogues exhibit anti-malarial or anti-tumor properties¹²⁷ and inhibition potential, and some of these compounds may even become ideal new drugs.^{129–133} Tumors are a major health problem that has drawn worldwide concern.^{134–136} In previous work, 3-APA analogues were found to promote DNA damage, induce apoptosis, and alter two human tumor cell lines, RKO-AS-45-1 and HeLa actin cytoskeleton, during active screening.¹³⁷ Naturally occurring molecules containing thiocyanate structures are rare, and these functional groups are mainly present in the formation of cruciferae glucosides during the desaccharification of some natural products with anti-tumor activity. It has also been shown that compounds containing thiocyanate groups also have anti-parasite effects, showing high cytotoxicity in anti-parasite studies. In view of these data, Barbosa *et al.*¹³⁸ designed two new 3-APA analogues to study their anti-tumor activity using theonelladin C as a lead compound and introduced thiocyanate groups into its chemical structure. To synthesize the target compounds **141** and **142**, they chose the route shown in Scheme 6. They selected the previously reported synthetic compounds **1a–b** as the starting materials and synthesized the designed target compounds **141** and **142** in two steps. In the first step, the hydroxyl compounds **137** and **138** were reacted with methyl sulfonyl chloride in methylene dichloromethane to obtain the intermediates **139** and **140**, in yields of 52–83% after a simple treatment. The second step was





Scheme 6 Synthesis of compounds **141** and **142**. Reagents and conditions: (a) MsCl, Et₃N, CH₂Cl₂, r.t., 10 h; (b) TBAB, KSCN, THF, reflux, 3 h.

accomplished *via* S_N2 nucleophilic substitution, in which the intermediate compounds **139** and **140** were converted to the corresponding target compounds **141** and **142** containing thiocyanate groups. In this step, **139** and **140** were substituted with potassium thiocyanate in THF and tetrabutylammonium bromide was used as a catalyst. The desired target compounds **141** and **142** were obtained *via* nucleophilic substitution, in yields of 55–78%. This synthesis route has the advantages of short reaction steps, high yields in each step, and it is easy to control the total yield of the reaction. Also, the reagents used are cheap and easy to obtain, and the reaction conditions are mild, which lays a solid experimental foundation for future industrialization. In the activity screening process of the target compounds **141** and **142**, the anti-malarial activity of the target compounds was evaluated (Table 6). *In vitro* test results showed that target compounds **141** and **142** inhibited the growth of parasites, with IC₅₀ values of 5.5 and 2.3 μM, respectively. These results indicate that the compounds have anti-malarial activity, but their selectivity is not high, which reflects that they have high cytotoxicity to the control cell line and are not conducive to use in proprietary medicine. Because these compounds have good cytotoxicity, they have certain anti-tumor properties. So next, the anti-tumor activity of target compounds **141** and **142** was examined. In *in vitro* activity evaluation, they evaluated the cytotoxicity of the compounds towards human cancer lines of colon cancer (RKO-AS-45-1) and cervical cancer (HeLa) to evaluate the anti-tumor activity of **141** and **142**. Meanwhile, the non-tumor human cell line (lung fibroblast, WI-26VA4) was used as a reference to determine the selectivity index of the target compounds, as the selectivity index is one of the most important indexes by which to evaluate the activity of target compounds. The anti-tumor activity of the target compounds

141 and **142** was investigated *in vitro*. The IC₅₀ values of the target compound **141** against RKO-AS-45-1 and HeLa cells were 0.8 and 12.4 μM, respectively, and the IC₅₀ values of target compound **142** against RKO-AS-45-1 and HeLa cells were 6.36 and 3.8 μM, respectively. Anti-tumor drugs need to be able to be highly selective towards effective (active) compounds, as in the process of killing tumor cells it should be almost impossible to kill normal cells. In determining the selectivity index of target compounds **141** and **142**, it was found that **141** was selective for the tumor cell line RKO-AS-45-1 (SI = 3.59). In order to further investigate the cytotoxicity mechanism of **141** and **142**, a variety of gene toxicity tests were conducted *in vitro*, including micro-nucleus assay, comet assay, ames assay and annexin-V/propidium iodide staining. The results of the relevant experiments showed that the synthetic alkaloids induce the wrong separation of chromosomes and damage DNA during the cell division of human tumor cells, resulting in cell death. These results preliminarily indicate that **141** may be a promising candidate as an anti-tumor chemotherapy drug, which needs to be further studied and developed in the future for its rapid clinical use to treat more cancer patients.

Cancer is one of the major diseases that seriously endanger human health in today's society.^{139–143} In the treatment of cancer, there are mainly surgical treatment, chemotherapy, radiotherapy and other means, among which chemotherapy is the most important treatment method currently used.^{144–147} The research and development of novel compounds with a unique chemical structure has become an important research direction in the field of anti-cancer drugs. The discovery of new drugs or lead compounds from natural products is an important route.¹⁴⁸ On the basis of natural products, many derivatives or analogues with biological diversity have been synthesized, especially in

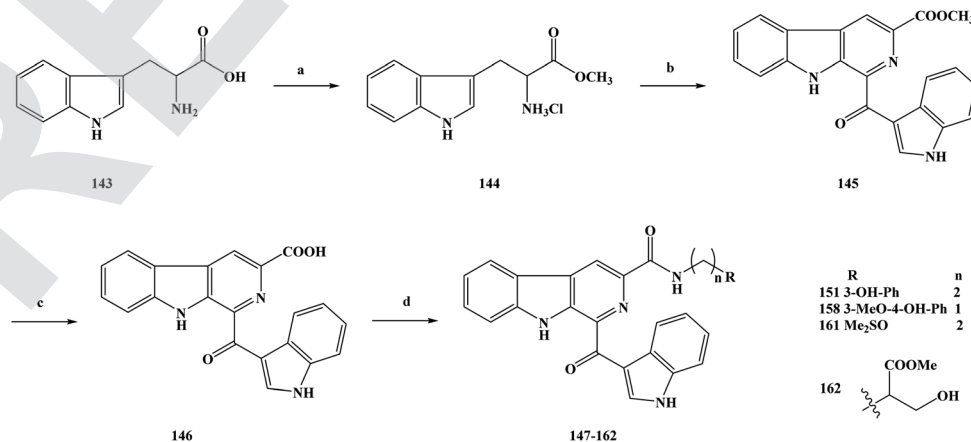
Table 6 *In vitro* inhibitory concentrations (IC₅₀) and selectivity indexes (SI) obtained for the human cancer cell lines RKO-AS45-1 and HeLa cells after exposure to different concentrations of the target compounds **141** and **142**

Compounds	IC ₅₀ (μM) ± SD			SI	
	PKO-AS45-1	HeLa	WI-26VA4	PKO-AS45-1	HeLa
141	0.80 ± 0.11	12.40 ± 2.25	2.87 ± 0.85	3.59	0.23
142	6.36 ± 1.30	3.80 ± 0.58	4.83 ± 1.14	0.76	1.27



terms of their anti-tumor activity. Alkaloids are natural products that are widely distributed in plants.^{149–153} They have a variety of chemical structures and play an important role in medicine. Among these natural alkaloids, β -carboline alkaloids are a type of widely distributed natural product with biological activities, which belong to the indole alkaloids and have a unique structure containing a tricyclic pyrido [3,4-*b*] indole ring.^{154–158} Pityriacitrin is a marine alkaloid separated from marine organisms. In terms of chemical structure analysis, it was found that the pityriacitrin tricyclic pyrido [3,4-*b*] ring structure is connected to the indole ring structure at the C-1 position.^{159–162} Pityriacitrin has been widely studied because of its unique chemical structure. In recent years, some derivatives and analogues of pityriacitrin have been separated or synthesized by researchers and their biological activities have been studied.¹⁶³ It has been found that these compounds have a wide range of biological activities, especially good anti-tumor activity and cytotoxicity. Natural products are always present in small amounts in nature, so in order to obtain large amounts of these compounds it is impossible to extract them from their sources.^{164–167} The use of synthetic methods changes this situation, and a large number of desired target compounds can be obtained through synthesis, and then applied to relevant research or used directly as drugs. Xu *et al.*¹⁶⁸ used pityriacitrin as the lead compound to modify its structure to obtain a large number of cytotoxic compounds. From the structure of the designed target compounds, it can be seen that they retain the chemical structure of pityriacitrin, and different amide groups were introduced at the C-6 position of the tricyclic pyrido [3,4-*b*] indole ring to obtain marine alkaloid β -carboline analogues. In this work, they combined the pharmacophores of β -carboline with biologically functional amide groups, thus synthesizing target compounds **147–162** with the structure of the β -carboline analogues. For the synthesis of the compounds, they selected the route shown in Scheme 7. Compounds **147–162** were prepared *via* a four-step reaction using tryptophan (compound **143**) as the starting material. In the first step, compound **143** was added to anhydrous methanol and then reacted with sulfide chloride to obtain the crude product methyl tryptophan

hydrochloride (compound **144**), without separation. In the second step, iodine and 1-(1*H*-indol-3-yl) ethanone were added to dimethylsulfoxide (DMSO) for the reaction, then compound **144** was added to continue the reaction, and 1-(1*H*-indole-3-carbonyl)-9*H*-pyrido [3,4-*b*] indole-3-carboxylate (compound **145**) was generated *via* a simple condensation ring reaction. In the direct nucleophilic substitution compound **145**, it was found that the reaction could not be carried out, so the reaction pathway was changed. The third step was to add **145** to a solution of sodium hydroxide. Hydrolysis readily occurred, and the intermediate **146** was obtained. In the final step, **146** was reacted with different substituted amines. DMF was used as the reaction solvent and HOBt/EDCI/Et₃N as the reaction catalyst to obtain the target compounds **147–162**. This synthesis uses inexpensive and readily available reagents, and the yields of the target compounds were moderate to good, obtained *via* a simple synthetic route, which lays foundation for its future industrialization. However, there are some problems in terms of the environmental friendliness of this route. Reagents such as chlorinated sulfoxide were used in the experiment, and some polluting gases were generated in the reaction process, which means that the route does not meet the requirements of green chemistry, and the subsequent process conditions therefore need to be optimized. In the course of the biological activity study, the cytotoxic effects of the marine alkaloid β -carboline analogue compounds **147–162** were tested *in vitro*. The SGC-790, A875, HepG2 and MARC145 cell lines were used as test subjects. *In vitro* cytotoxicity test results showed that the target compounds exhibited moderate to good inhibitory activity against all four cell lines. The activities of **159** and **160** with aromatic groups directly on their amides were lower than those of the benzylamine (**153–158**) and phenylethylamine (**147–152**) substituted compounds. The compound containing a 3-hydroxyl group (**151**) showed significantly better biological activity than the compound containing a 3-methoxy substituent (**148**). In order to further study the biological activity of these target compounds and to screen out the more active compounds, the IC₅₀ values of the target compounds were also tested *in vitro*. The results showed that the target compounds



Scheme 7 Synthesis of the marine alkaloid oriented β -carboline analogue compounds **147–162**. Reagents and conditions: (a) SOCl₂, MeOH; (b) 1-(1*H*-indol-3-yl) ethanone, I₂, DMSO; (c) NaOH, MeOH, r.t.; (d) NH₂(CH₂)_nR, HOBt, EDCI, Et₃N, DMF, r.t.

Table 7 Cytotoxic activities of the target compounds

Compounds	IC ₅₀ (μM)			
	SGC-7901	A875	HepG2	MARC145
151	17.65 ± 5.84	11.91 ± 0.80	8.63 ± 3.31	13.77 ± 3.75
158	16.18 ± 2.31	8.47 ± 2.96	11.08 ± 3.33	21.77 ± 6.94
161	6.82 ± 0.98	8.43 ± 1.93	7.69 ± 2.17	7.19 ± 1.43
162	14.30 ± 2.57	10.46 ± 1.34	9.99 ± 1.82	16.27 ± 0.07
5-FU	53.58 ± 1.99	62.12 ± 17.83	66.42 ± 12.99	115.54 ± 8.30

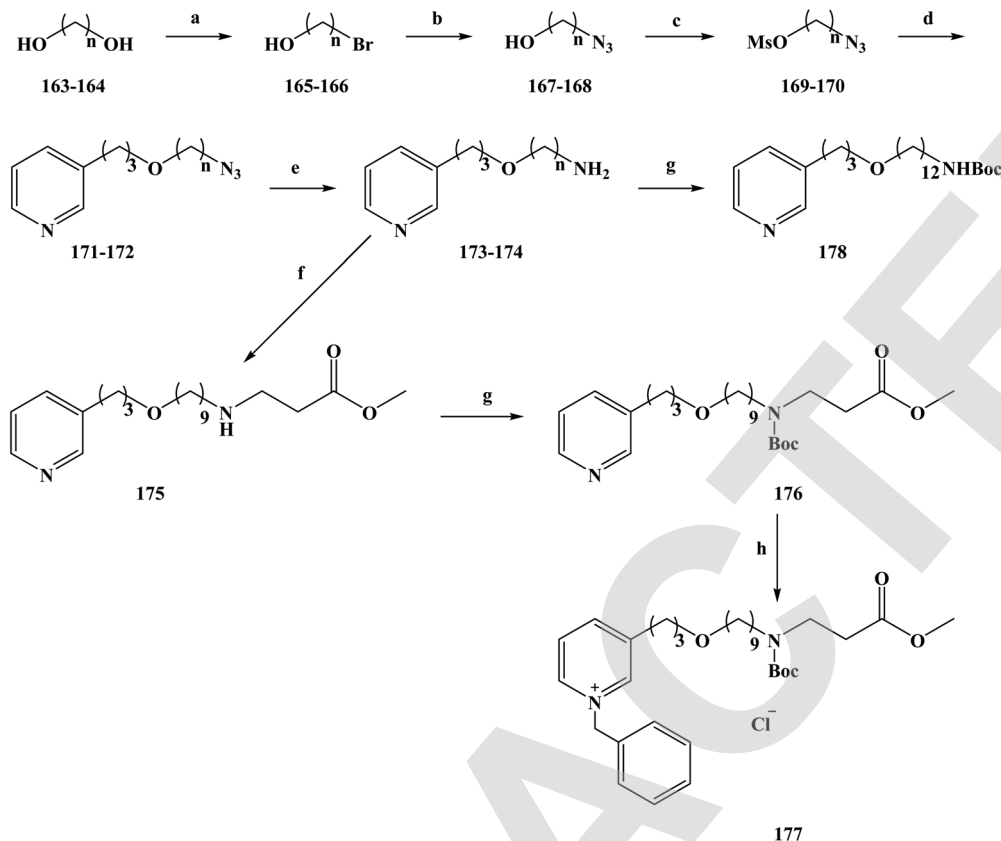
151, **158**, **161** and **162** showed good inhibitory activity (Table 7). The IC₅₀ values of the target compound **161** were 6.82 ± 0.98, 8.43 ± 1.93, 7.69 ± 2.17 and 7.19 ± 1.43 μM, respectively. The SAR results showed that benzylamine substituted compounds usually had better activity than phenylethylamine substituted compounds, and the activities of these two target compounds were obviously better than those of aromatic amine substituted compounds, which demonstrated the importance of the link chain between the aromatic rings and amides. In addition, the biological activities of most of the target compounds were better than those of β-carboline and β-carboline B, which indicated that the presence of amide groups in the compounds was beneficial to the anti-tumor activity of the target compounds. In addition, hydroxyl phenyl substituted compounds (**158** and **151**) have significantly higher activity than other corresponding compounds, possibly because the polarity of the hydroxyl phenyl group is favorable for binding to the target protein. The target compound **161** contains a special sulfonyl group and has the highest inhibitory activity. The special properties of the hydroxyphenyl and sulfur groups help to improve the biological activities of the target compounds. On the basis of the dose response analysis of **151**, **158**, **161** and **162** the results showed that these target compounds had a concentration-dependent inhibitory effect on cell lines. In general, these target compounds have good cytotoxic activity and can be used as potential cell agents.

2.2 Anti-malarial activity

In 2008, the World Health Organization released the world malaria report, which showed that malaria kills about one million people worldwide each year.^{169–173} Malaria is more serious in countries or regions that have poor living conditions and medical conditions, especially in Africa.¹⁷⁴ There is no effective vaccine to control malaria, and control programmes to reduce the spread of the disease still rely on chemotherapy (drug therapy) to prevent malaria and treat malaria-infected people. The existing drugs for the prevention and treatment of malaria mainly include mefloquine and artemisinin analogues, but new drug-resistant strains appear in the clinical, resulting in ineffective drug treatment.¹⁷⁵ Therefore, it is urgent to research and develop new and effective anti-malarial drugs for better prevention and treatment of malaria. In the research and development of anti-malarial drugs, medicinal chemists have focused on the research of marine alkaloids and their

analogues in recent years, hoping to find effective anti-malarial drugs based on these compounds. In the process of screening the anti-malarial activity of marine alkaloids, makaluvamine A (**2**) was the first molecule to be studied for anti-malarial activity.^{176,177} Makaluvamine A is a complex alkaloid with a heterocyclic structure separated from marine resources. However, this marine alkaloid was found to have a very low therapeutic index in an *in vitro* anti-malarial activity test, which means that it is not an ideal candidate as an anti-malarial drug for clinical treatment. Alkaloids containing piperidine and pyridine structures are widely distributed in marine sponges of different genera, and 3-alkyl pyridine alkaloids have been separated from sponges.¹⁷⁸ In 2004, Volk and Kceck reported a viscosaline (**1**) alkaloid separated from a marine sponge, and found that the compound showed high anti-microbial activity towards a variety of bacteria.^{179,180} In 2007, the Shorey team first reported the total synthesis of viscosaline, and described the key intermediate theonelladin C (**2**) in the synthesis method. The study found that theonelladin C is also a natural product compounds found in sponges.¹⁸¹ The chemical structures of viscosaline and theonelladin C are relatively simple, with viscosaline showing the highest anti-microbial activity. Hilario *et al.*¹⁸² used viscosaline as a lead compound to modify in order to prepare marine alkaloid viscosaline analogues. These 3-alkylpyridine analogues have a viscosaline pyridine ring in the alkyl chain, and oxygen-containing groups were introduced to alter the biological activity of the analogues. To synthesize compounds **177–178**, a relatively simple total synthesis method was chosen. The specific route for the synthesis of the new 3-alkylpyridine compounds is shown in Scheme 8. 1,9-Nonanodiol (compound **163**) and 1,12-dodecandiol (compound **164**) were used as starting materials in the reaction. In refluxing toluene, the reactants were selectively brominated with hydrobromic acid to obtain the corresponding brominated alcohol compounds **165** (yield 70%) and **166** (yield 80%). Compounds **165** and **166** were converted to their corresponding azides **167** and **168** in dimethyl sulfoxide via S_N2 substitution. Then, **167** and **168** were treated with methyl sulfonyl chloride in methylene dichloride to obtain the intermediate compounds **169** and **170**. Using tetrabutylammonium bromide as a phase transfer catalyst, compounds **171** and **172** were etherified with 3-pyridinepropanol, in good yields. The amine-containing compounds **173** (72% yield) and **174** (79% yield) were obtained by reducing terminal azides **171** and **172** in THF with lithium aluminum hydride. Compound **173** and methyl acrylate were refluxed in





Scheme 8 Synthesis of the oxygenated 3-alkylpyridine marine alkaloid analogue compounds **177** and **178**. Reagents and conditions: (a) HBr (aq), toluene, reflux; (b) NaN₃, DMSO, r.t., 12 h; (c) MsCl, Et₃N, methylene chloride, stirring, 3 h; (d) NaOH, Bu₄N⁺Br[−]; (e) LiAlH₄, THF; (f) methyl acrylate, toluene, reflux; (g) Boc₂O, methylene chloride; (h) BnCl, reflux.

toluene to form **175** (yield 70%). Compounds **174** and **175** were treated with di-tert-butyl carbonate in methylene dichloride to obtain **176** (yield 94%) and **178** (yield 75%). Compound **176** was alkylated with benzyl chloride to obtain another target compound, **177**. The synthetic enables the total synthesis of the target products, where the raw materials and reagents used are cheap and easy to obtain, and the yield of each step is relatively high, laying a solid foundation for its future industrialization. During the study of anti-malarial biological activity, the anti-malarial effect of the 3-alkylpyridine marine alkaloid analogues *in vitro* using a [³H]-hypoxanthine doping method

(Table 8) was established. On this basis, the cytotoxic effects of seven marine alkaloid analogues (compounds **172–178**) were tested using the MTT method to evaluate the safety of the target compounds. The seven tested marine alkaloid analogues were evaluated for their anti-malarial activity at half the maximum inhibitory concentration (IC₅₀) and half the lethal drug concentration (LC₅₀). *In vitro* anti-malarial results showed that the compounds tested were all active *versus* chloroquine against the plasmodium falciparum clone W2. Drug SAR studies have shown that the alkyl chain lengths of these tested compounds (compounds **173** and **174**) have an impact on their anti-malarial and cytotoxic properties. As the length of the alkyl chain increased from 9 to 12 carbon chains, the anti-malarial activity and selectivity index of the compounds were increasingly improved (compound **174** SI = 10.82). The length of the alkyl chain of compound **175** was also extended, significantly reducing its IC₅₀ value compared with that compound **173**, and significantly increasing its selectivity index from 5.48 to 12.36. However, after the amino group of compound **175** was protected with a Boc group, the IC₅₀ value of product **176** was significantly increased, and the selectivity was also significantly reduced (SI = 2.78). These results suggest that the position of the amine plays an important role in the anti-malarial activity of the compounds. At the same time, the target compound **177** of the quaternary ammonium reaction had a significantly reduced

Table 8 *In vitro* anti-plasmodial activities of the oxygenated 3-alkylpyridine marine alkaloid analogues

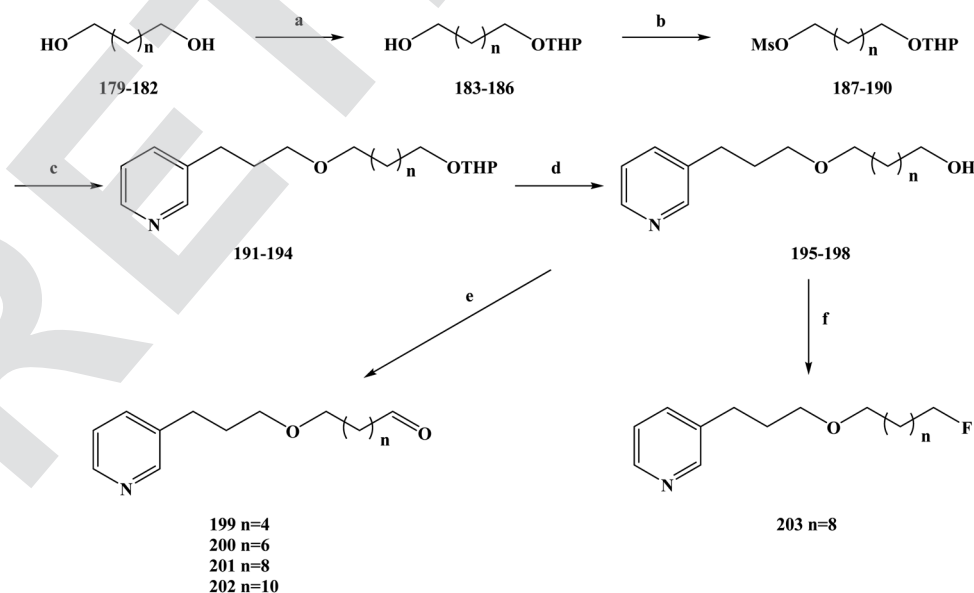
Compounds	IC ₅₀ (μM) ± SD	LC ₅₀ (μM) ± SD	SI
172	4.33 ± 0.87	45.52 ± 2.71	10.51
173	17.96 ± 3.59	98.43 ± 3.77	5.48
174	6.24 ± 0.31	67.54 ± 1.92	10.82
175	8.50 ± 0.14	105.12 ± 5.73	12.36
176	43.04 ± 4.30	120.00 ± 8.15	2.78
177	<3.38	52.67 ± 2.25	>15.58
178	<4.76	46.78 ± 4.65	>9.82
Chloroquine	0.625 ± 0.003	>100	>160



IC₅₀ value, and its selectivity was improved, making it the compound with the highest selectivity index (SI > 15.58) among the tested compounds. Preliminary *in vitro* anti-malarial activity studies showed that 3-alkylpyridine marine alkaloid analogues have ideal anti-malarial activity and cytotoxicity. Among the synthesized compounds, compound **177** had the best anti-malarial activity and the highest selectivity index, thus making it a promising candidate for the development of novel anti-malarial drugs and deserving of further study.

Malaria is a serious infection caused by malaria-carrying parasites and can be transmitted in the blood by female mosquitoes.¹⁸³ Malaria is a global public disease that has high morbidity and mortality.¹⁸⁴ According to the World Health Organization, there are more than 100 species of malaria parasites that cause malaria.^{185–188} However, only a few species of plasmodium have attracted research attention, including *Plasmodium falciparum*, *Plasmodium malariae*, *Plasmodium vivax*, *Plasmodium knowlesi* and *Plasmodium ovale*. Malaria, caused by malignant protozoa, is a severe disease and can cause other syndromes.¹⁸⁹ Drug therapy is one of the main strategies for the treatment and control of malaria.^{190–192} Because of the emergence of artemisinin-resistant parasites in the clinic. It is imperative to find new antimalarial drugs, especially those that are lethal to resistant parasites. In the process of developing new anti-malarial drugs, the target and mechanism of action of the drugs need to be further determined.^{193–195} Natural products are characterized by their complex structure and biological activity diversity.¹⁹⁶ Due to these reasons, research on anti-malarial drugs has shifted to natural products in recent years.^{197,198} In the process of screening the activity of natural products, 3-alkylpyridine marine alkaloids and their analogues have been found to have anti-malarial activity. On this basis, de Souza Barbosa *et al.*¹⁹⁹ further modified the structure of the lead

compound and designed new types of 3-alkylpyridine marine alkaloid analogues. To improve the anti-malarial activity, selectivity and safety of the compounds, they selected different lengths of alkyl chains. The types of functional groups attached to the ends of the chains were used to further investigate the SAR of the designed compounds. At the same time, in order to improve the anti-malarial activity, the selectivity, improve the safety and enhance the affinity of the compounds, the functional groups at the end of the side chain of the compound were fluorinated. During the synthesis route selection of 3-APA analogues, the specific synthesis operations were carried out as shown in Scheme 9. Diols (compounds **179–182**) with different length carbon chains were used as the starting materials of the reaction, and their single hydroxyl groups were selectively protected to obtain the corresponding monohydropyronic acetals (compounds **183–186**). Then, the hydroxyl groups of compounds **183–186** were further protected to obtain the compounds **187–190**. The compounds **187–190** were etherized with 3-(pyrid-3-yl)propan-1-ol using a phase transfer catalyst to obtain compounds **191–194**, which were then protected using hydrochloric acid to obtain compounds **195–198**. The Swern oxidation was used to convert the alcohols **195–198** to the corresponding aldehydes **199–202**, in yields of 60–95%. At the same time, compound **197** and diethylamine-based sulfur trifluoride underwent a fluorine-substituted halogenation reaction in methylene chloride to obtain compound **203** (yield 17%). This synthesis is simple to carry out under mild reaction conditions using cheap and readily available reagents. The yield per step was generally medium to good, but the fluorine substitution produced a low yield of only 16% (compound **203**), which would increase the costs of this process if it were to be industrialized. The halogenation reaction will be a key problem in the subsequent process optimization, which needs to be further



Scheme 9 Synthesis of the fluorinated 3-alkylpyridine marine alkaloid analogue compounds **199–203**. Reagents and conditions: (a) NaHSO₄, DHP, DMSO, hexane, 40 °C, 16 h; (b) MsCl, Et₃N, CH₂Cl₂, r.t., 10 h; (c) 3-pyridinepropanol, NaOH/H₂O, Bu₄N⁺Br[−], Et₂O, r.t., 72 h; (d) MeOH, HCl, r.t., 12 h; (e) (COCl)₂, Et₃N, DMSO, DCM, −60 °C, 25 min; (f) DAST, DCM, 0 °C, 18 h.



Table 9 *In vitro* anti-malarial activities, cytotoxicities and SIs of the 3-APA analogues

Compounds	IC ₅₀ (μM) ± SD		SI
	<i>P. falciparum</i> (W2)	WI-26VA4	
191	53.7 ± 0.7	>300	>5.6
192	37.7 ± 0.2	37.8 ± 5.0	1.0
193	14.0 ± 0.6	6.4 ± 0.7	0.4
194	8.9 ± 0.4	11.3 ± 1.4	1.2
195	210.6 ± 12.7	>400	1.9
196	>188	167.7 ± 10.5	nd
197	14.7 ± 0.2	99.1 ± 11.2	6.7
198	15.1 ± 0.8	34.1 ± 6.5	2.2
199	180.2 ± 8.3	nd	nd
200	52.4 ± 8.9	nd	nd
201	20.3 ± 0.2	nd	nd
202	30.6 ± 2.7	nd	nd
203	2.5 ± 0.1	28.2 ± 3.7	11.2
Chloroquine	0.4 ± 0.066	>100	>250

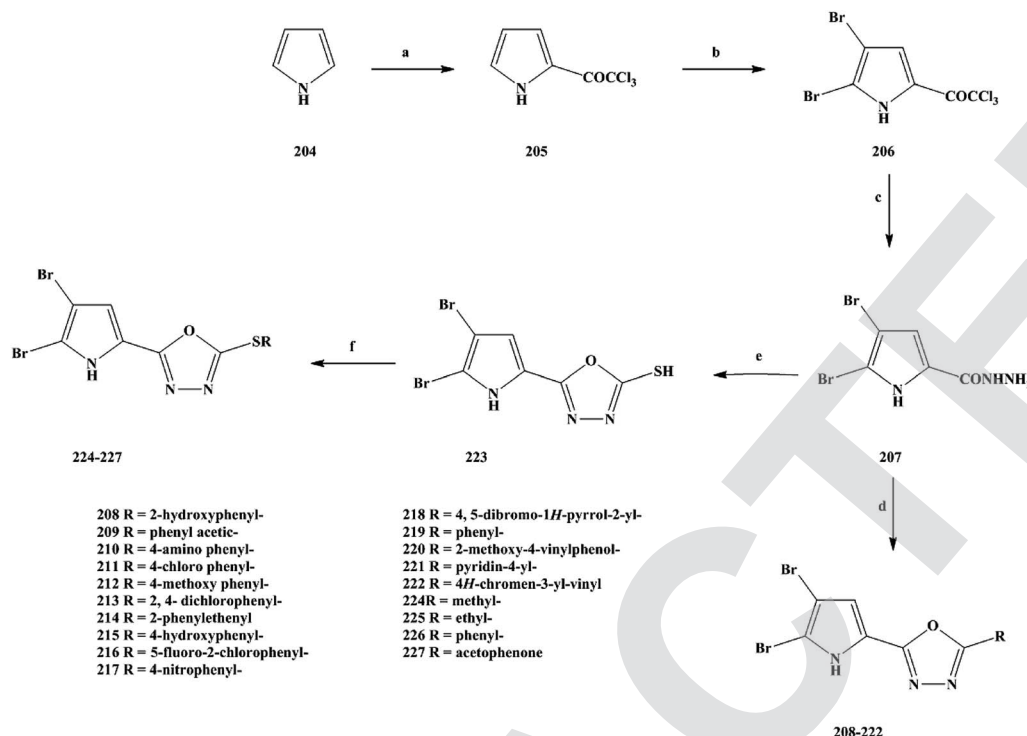
improved to improve the yield and reduce the reaction costs. Upon *in vitro* screening of the anti-malarial activity of the abovementioned compounds, they were screened against *Plasmodium falciparum* (strain W2) and were found to be cytotoxic to human fibroblasts (WI-26VA4). *In vitro* anti-malarial activity test results showed that the compounds tested had moderate activity against *Plasmodium falciparum*, with IC₅₀ values ranging from 8.9 to 210.6 μM (Table 9). The results of the SAR showed that there was a certain relationship between the anti-malarial activity and the selectivity index of these compounds. With the extension of the side chain from 6 to 12 carbon atoms, the anti-malarial activity of the side chain increased, and the activity decreased after the chain exceeded 12 carbon atoms. A similar SAR was also observed for the selectivity index, with the length of the carbon chain also affecting the selectivity index of the compounds. At the same time, the compounds containing hydroxyl groups had higher antimalarial activity than those containing other groups. The selectivity index is a key factor in determining whether or not an anti-malarial drug can be used in treatments. Compound **197** had one of the lowest IC₅₀ values (IC₅₀ = 14.7 μM) among the compounds tested, but its selectivity index was the highest (SI = 6.7). Therefore, compound **197** was selected for further structure optimization, and the its hydroxyl group was converted to a fluoride group to obtain compound **203**. This substitution was made to improve the anti-malarial activity of compound **197**. The *in vitro* anti-malarial activity of compound **203** was thus significantly increased (5.8 times) compared to that of compound **197**. The IC₅₀ value of the chloroquine resistant strain W2 of *Plasmodium falciparum* was 2.5 μM and the SI value was 11.2. The IC₅₀ value of the chloroquine sensitive strain 3D7 was 2.3 μM and the SI value was 12.2. On this basis, the binding ability of the heme group to compound **203** was further determined. Using an organo-silicone method, the results showed that compound **203** more favorably formed stable complexes (hematin/3-APA complex) with heme molecules in a molecular ratio of 2 : 1 after fluorine

was used to replace the hydroxyl groups. In summary, they synthesized a new halogenated 3-APA analogue, compound **203**, with good anti-malarial properties and increased activity against *Plasmodium falciparum* as well as an increased selectivity index. Compound **203** has great development potential as a new anti-malarial drug, and the mechanism of action and pharmacokinetic characteristics of the compound need to be further determined in follow-up work.

2.3 Anti-bacterial activity

Bacteria are one of the main groups of living organisms and as the most abundant of all living organisms, have a great influence on human activities.²⁰⁰ Bacteria are the cause of many diseases, including tuberculosis, gonorrhea, anthrax poisoning, syphilis, plague, trachoma and other diseases. In plants, bacteria cause leaf spots, fire blight and wilting.²⁰¹ Anti-microbial drugs generally have bactericidal or antibacterial activity, drugs such as antibiotics, sulfonamides, imidazoles, nitroimidazoles, quinolones and other chemical synthetic drugs. Some products obtained from the culture of microorganisms such as bacteria, actinomycetes and fungi, or the same or similar substances made by chemical semi-synthesis, may also be synthesized chemically.^{202–204} Anti-microbial agents have inhibitory and killing effects towards pathogens at certain concentrations. 1,3,4-Oxadiazoles are compounds that have special chemical structures²⁰⁵ and biological activities that include anti-inflammatory, hypoglycemic, anti-depressant, anti-tumor cell proliferation, anti-bacterial properties. In molecular docking and pharmacokinetic studies, it was found that the introduction of a 1,3,4-oxadiazole structure into the drug molecule could change its polarity, flexibility and metabolic stability. The chemical structure of 1,3,4-oxadiazole plays an irreplaceable role in drug efficacy.²⁰⁶ Previous work found that heterocycles containing a pyrrole ring system had diverse biological activity, with several pyrrolidone analogues showing obvious anti-microbial activity in the activity evaluation process. Bromopyrrole marine alkaloid, derived from the secondary metabolites of marine sponges, has been widely investigated. These marine alkaloids have a special chemical structure, most of which are composed of 4,5-dibromopyrrole rings.^{207,208} In the evaluation of the biological activity of these marine alkaloids containing a 4,5-dibromopyrrole ring, it was found that they have diverse biological activities, including blocking serotonin receptor antagonists of α-adrenoceptor receptors, acting as anti-histamines and serotonin receptor antagonists, against myosin aptase, inhibiting kinase activation, and exhibit anti-tumor, anti-microbial, anti-tuberculosis, and anti-fungal properties. On the basis of previous work, Rane *et al.*²⁰⁹ designed novel bromopyrrole marine alkaloid derivatives. The chemical structures of these designed target compounds showed that two groups could be joined together by splicing to form a hybrid molecule. Specifically, a 4,5-dibromopyrrole ring and 1,3,4-oxadiazole ring were combined *via* chemical bonds to form a new class of target compounds, **208–222** and **224–227**. Different substituents were introduced into the 1,3,4-oxadiazole ring to change its biological activity. To synthesize the target





Scheme 10 Synthesis of 2-(4,5-dibromopyrrol-2-yl)-5-substituted-1,3,4-oxadiazole compounds **208–222** and **224–227**. Reagents and conditions: (a) CCl_3COCl , anhydrous ether, K_2CO_3 ; (b) $\text{Br}_2/\text{chloroform}$; (c) $\text{NH}_2\text{NH}_2 \cdot \text{H}_2\text{O}$, r.t., 1 h; (d) $\text{R}-\text{COOH}$, POCl_3 , reflux; (e) CS_2 , KOH ethanol, reflux, 12 h; (f) alkyl/aryl halide, NaOH , ethanol, stirring, 10 h.

compounds, the synthesis route in Scheme 10 was used. In the first step of the reaction, pyrrole (compound **204**) was used as a starting material to conduct an esterification reaction with trichloroacetyl chloride. In the reaction, anhydrous ethyl ether was used as the reaction solvent and potassium carbonate as a base, to obtain a good yield of 2-trichloroacetylpyrrole (compound **205**). In the second step, compound **205** was brominated with bromine in chloroform to obtain 4,5-dibromo-2-trichloroacetyl-1H-pyrrole (compound **206**) in good yield. The third step was to stir compound **207** with hydrazine hydrate to react at room temperature. In the presence of phosphorus chloride, 2-(4,5-dibromo-1H-pyrrol-2-yl)-5-aryl-1,3,4-oxadiazoles (compounds **208–222**) could be prepared *via* the refluxing of compound **207** with aromatic acids containing different substituents. Meanwhile, 5-(4,5-dibromo-1H-pyrrol-2-yl)-1,3,4-oxadiazole-2-thiol (compound **223**) could be obtained by the reaction of compound **207** with carbon disulphide in ethanol under alkaline conditions in the presence of potassium hydroxide. When compound **223** was mixed with different halide substituents in ethanol, in the presence of sodium hydroxide to provide alkaline conditions, an alkylation reaction took place to obtain the target compound *S*-alkylation derivatives **224–227**. The synthetic route can be used to synthesize 20 derivatives, meaning that it has good universality. The reagents used in the synthesis process are cheap, easy to get, the synthesis is simple to carry out and produces compounds in good yield, laying a good foundation for its future industrialization. During the biological activity study, the anti-microbial

activity of the target compounds was evaluated *in vitro*, against *Escherichia coli*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae* bacterial strains. The minimum concentration (maximum dilution) required to inhibit bacterial growth was used as the minimum inhibitory concentration (MIC) of the target compound to evaluate the activity of the compound. Meanwhile, the anti-fungal activity of candida albicans in DMSO was screened using the agar dilution method. In addition, the synthesized compounds were screened *in vitro* against *Mycobacterium tuberculosis* H₃₇Rv to determine the minimum inhibitory concentration. The results of the *in vitro* anti-microbial activity screening showed that the synthesized compounds have good anti-bacterial, anti-fungal and anti-tuberculous activities. These spliced compounds showed anti-bacterial activity against both Gram-positive and Gram-negative bacteria, showing that they have extensive antibacterial activity. The results of drug SAR showed that the presence of electric-donating groups on the aromatic rings of the compounds, such as amino, chlorine, fluorine and nitro groups, was very important in inhibiting the activity of *Escherichia coli*. For example, the anti *Escherichia coli* MIC of compounds **210**, **211** and **216–218** was 1.56 μM , and the activity was similar to that of the positive control. For Gram-positive *Staphylococcus aureus*, compounds **208**, **211**, **216–218** and **220–222** had an equivalent activity of 1.56 μM at MIC compared with the positive control. However, the compounds **223–227** containing mercaptan groups or *S*-alkyl/aryl groups were less active against *Staphylococcus aureus*. All compounds tested had no inhibitory



Table 10 Anti-microbial test results of the target compounds

Compounds	MIC (μM)					
	<i>E. coli</i>	<i>S. aureus</i>	<i>P. aeruginosa</i>	<i>K. pneumoniae</i>	<i>C. albicans</i>	Antitubercular activity
211	1.56	1.56	6.25	100	12.5	9.50
213	12.5	6.25	6.25	100	12.5	1.60
216	1.56	1.56	6.25	100	12.5	6.50
217	1.56	1.56	6.25	100	1.56	9.00
218	1.56	1.56	6.25	100	1.56	3.50
227	3.125	12.5	12.5	100	1.56	1.50

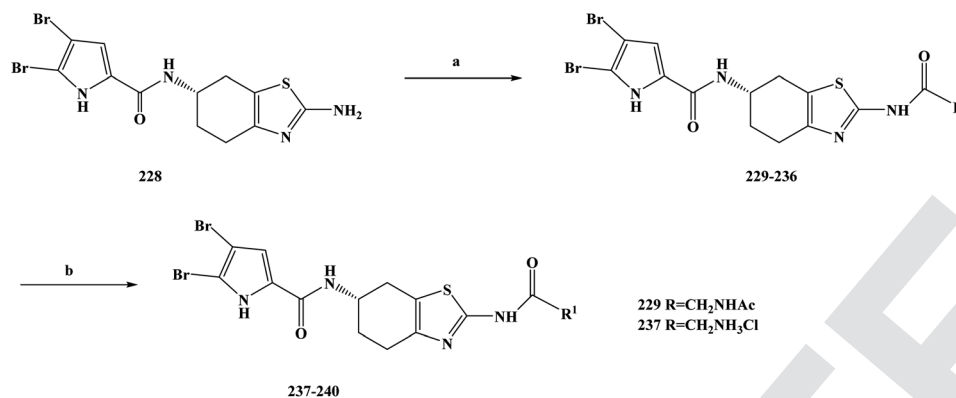
activity against pneumonia bacteria, while compounds **210–219** showed moderate activity against *pseudomonas aeruginosa* (MIC was 6.25 μM). In the *in vitro* anti-fungal activity screening, the three compounds with the best anti-fungal activity were **217**, **218** and **227**, and the MIC was 1.56 μM . In the *in vitro* evaluation of *Mycobacterium tuberculosis* activity, it was found that the compounds **213**, **218**, **221**, **222** and **227** showed obvious *Mycobacterium tuberculosis* activity when the MIC were 1.60, 3.50, 3.50, 2.00 and 1.50 μM , respectively, and the compound **227** showed the highest anti-tuberculosis activity. The SAR analysis showed that by introducing *S*-aryl substitution and free SH or *S*-alkyl or *S*-aryl on the C5 sites of the 1,3,4-oxadiazole ring, the anti-tuberculous activity of the compound increased. At the same time, when heterocyclic rings such as 4,5-dibromopyrrole, pyridine-4-yl and 4*H*-chromen-3-yl-vinyl were introduced, the anti-tuberculosis activities of the compounds were significantly enhanced. For example, the MICs of the compounds **218**, **221** and **222** were 3.50, 3.50 and 2.00 μM , respectively. In general, the compounds **221** and **216–218** have good anti-bacterial activity, compounds **217**, **218** and **227** have good anti-fungal activity, and compounds **213** and **227** inhibit *Mycobacterium tuberculosis* at low concentrations. These compounds have been developed into new anti-bacterial, anti-fungal or anti-tuberculosis drugs, which need to be further developed (Table 10).

2.4 Anti-viral (HCV) activity

Hepatitis C is a virus hepatitis caused by a hepatitis C virus (HCV) infection, mainly as a result of blood transfusion, acupuncture, drug abuse and other transmissions.²¹⁰ According to World Health Organization statistics, the global infection rate of HCV is around 3%, and there are an estimated 180 million people infected with HCV, with a 35 000 rise in new cases of hepatitis C each year. The mortality rate associated with HCV infection will continue to increase in the next 20 years, which is extremely harmful to the health and life of patients, making it one of the most serious social and public health problems.²¹¹ Anti-viral drugs that act directly on the replication cycle of the HCV virus have been developed to a certain extent, offering significant promise for treatment and improvement in patients with HCV infection. At present, the anti-viral drugs that can directly act include non-structural protease 3/4A inhibitors, non-structural 5A complex inhibitors and non-structural 5B polymerase inhibitors. These anti-HCV drugs can act on

different viral targets.^{212,213} At each stage of virus replication, the complementary strategy to directly target viral proteins is to target host cytokines required for the virus life cycle. Targeted host therapy includes inhibition of virus entry, translation, replication, assembly and release. In recent years, it has been found that analogues of marine alkaloid globulin can inhibit the replication of HCV by interacting with cellular chaperone heat shock protein 90 (Hsp90). This compound can bind to its ATP binding site through the N-terminal domain. HSP90 is one of the most common protein types in cytoplasm and plays an important role in eukaryotic cells.²¹⁴ Almost all viruses need the presence of Hsp90 to successfully spread. Hsp90 inhibitors have been studied in clinical trials for more than 20 years due to their beneficial role in the cell life cycle.^{215–217} However, the clinical development of Hsp90 inhibitors is also hindered to some extent, mainly because of the untargeted effect of Hsp90 inhibitors and the lack of sufficient specificity and mechanism of action. Previous studies have shown that HCV replication could be inhibited by targeting the host cell protein Hsp90. Preliminary work Lillsunde *et al.*²¹⁸ found that compounds with a pyrrolamide and 4,5,6,7-tetrahydrobenzo[1,2-*d*] thiazole structure could inhibit the replication of HCV type 1b. It was found that compounds containing the unsubstituted 2-amino group had the most effective biological activity in terms of drug SAR. Therefore, they further introduced different substituents on the lead 2-amino compound to change its biological activity. They designed a series of 4,5,6,7-tetrahydrobenzo[1,2-*d*] thiazole-2-amines with the 2-amino group replaced by a basic group or an *N*-heterocyclic ring to increase binding to Hsp90, thereby enhancing the inhibition of HCV replication by the target compounds **229–240**. In the process of synthesizing the target compounds, they used the route shown in Scheme 11, in which two steps were used to complete the preparation of the desired target compounds, and compound **228** was used as the starting material. The first step in the reaction, the acylation of compound **228** with the corresponding carboxylic acids, 1-ethyl-3-(3-dimethylaminopropyl) carbon diimide (EDC) and 1-hydroxy benzyl triazole (HOBt) as a catalyst, and *N,N*-dimethylformamide (DMF) as a reaction solvent, the preparation gave the required amide structures needed for the target compounds **229–237**. Since some of these compounds contained the Boc protection group in their molecular structure, the second step was performed in order to obtain the compounds with the protective group. In the second reaction,





Scheme 11 Synthesis of 4,5,6,7-tetrahydrobenzo [1,2-d] thiazole derivative compounds 229–240. Reagents and conditions: (a) corresponding carboxylic acids, EDC, HOBT, NMM, DMF, r.t., 18 h; (b) acetyl chloride, MeOH, 0 °C, 1 h, then r.t., 18 h.

the compounds 230–232 and 236 were placed in acetyl chloride to protect them, and finally the target compounds 237–240 were obtained. Twelve target compounds were prepared according to the synthesis route, showing that it has good universality. The synthetic route is relatively short and relatively easy to use. However, a major disadvantage of this route is that the yield of the reactions was generally low, except for the yields of target compounds 237 and 238, which were high. The yield of the second deprotection reaction was relatively high, while the yield of the first step amide reaction was relatively low, which would bring great difficulties for the future industrialization of this process. Therefore, in the subsequent process optimization, the process should be further improved to increase the yield of the reaction and reduce the synthesis cost. During the biological activity study, a HCV replication cell model was used to assess the anti-viral activity of the marine alkaloid analogue compounds 229–240. The target compounds 229–240 were first tested in a HCV genotype 1b replicator model to preliminarily evaluate the anti-viral activity of the target compounds *in vitro*. In the initial test results, the inhibition rates of the target compounds 229, 231 and 237–240 on HCV replicons exceeded 90%, and the inhibitory activities of the target compounds 230 and 233–236 were low, ranging from 30% to 87%, while the target compound 232 showed no inhibition. It was found from the SAR that the anti-viral activities of the target compounds containing chain substituents were better than those containing rings. The alkyl chain of the compounds containing the chain substituents was increased from one to two carbon atoms, and it was noted that the anti-viral activity was improved. At the same time, the anti-viral activity of the five-membered heterocyclic substituent was higher than that of the six-membered heterocyclic substituent, and the anti-viral activity of the six-membered heterocyclic azacyclic was higher than that of the benzene ring. Based on *in vitro* tests, the target compounds 229, 233, 234 and 237–240 were further studied to determine the dose-response (IC_{50}) and cytotoxicity (CC_{50}) of the target compounds to the HCV genotype 1b replication subsystem (Table 11). The results showed that all the selected target compounds inhibited the HCV replicons in a dose-dependent manner and had no toxicity at the replicon-

inhibiting concentration. At the same time, the SI results showed that the selectivities of the target compounds 237, 229 and 239 were relatively high, with SI values of 98, 62 and 40, respectively. The selectivities of these target compounds were higher than that of the positive control. To determine whether the target compounds were also effective in inhibiting full-length virus replication, the target compounds were evaluated for their ability to inhibit HCV genotype 2a replicons. The results showed that the target compounds 229, 237 and 239 had anti-viral activity against HCV, with IC_{50} values ranging from 0.49 to 1.52 μ M, while other target compounds had no inhibitory activity. The inhibitory activity of the target compounds 229 and 237 against genotype 2a was even slightly higher than that against genotype 1b. HCV transmission in hepatocytes could be interacted with cellular partners in a variety of ways. Hsp90 could assist in protein maturation, folding, and stabilization and it was used for the replication of HCV. Relevant studies have shown that the target Hsp90 could effectively inhibit the replication of HCV. In order to evaluate the inhibition of Hsp90 as a potential mechanism of action, the binding ability of the target compounds to Hsp90 was evaluated by microscale thermophoresis (MST). The results of the evaluation indicate that Hsp90 is indeed a potential target for the target compounds and may provide important guidance for the further development of anti-viral drugs targeting the progenitor of this cell. The

Table 11 Replicon inhibitory activities of the target compounds and control in HCV genotype 1b and 2a replicon models

Compounds	HCV replicon genotype			2a IC_{50} (μ M)
	1b IC_{50} (μ M)	CC_{50} (μ M)	IS	
229	2.5 \pm 0.25	154 \pm 2.9	62	1.52 \pm 0.15
233	5.5 \pm 1.8	204 \pm 4.9	37	nd
234	21 \pm 2.2	86 \pm 5.2	4.1	nd
237	1.3 \pm 0.23	128 \pm 6.6	98	0.49 \pm 0.06
238	22 \pm 2.8	67 \pm 2.0	3.0	nd
239	1.2 \pm 0.41	48 \pm 2.5	40	1.32 \pm 0.1
240	7.2 \pm 0.46	72 \pm 4.2	10	nd
17-DMAG	0.06 \pm 0.01	1.7 \pm 0.25	28	0.04 \pm 0.006
Ribavirin	64 \pm 7.0	>300	nd	nd

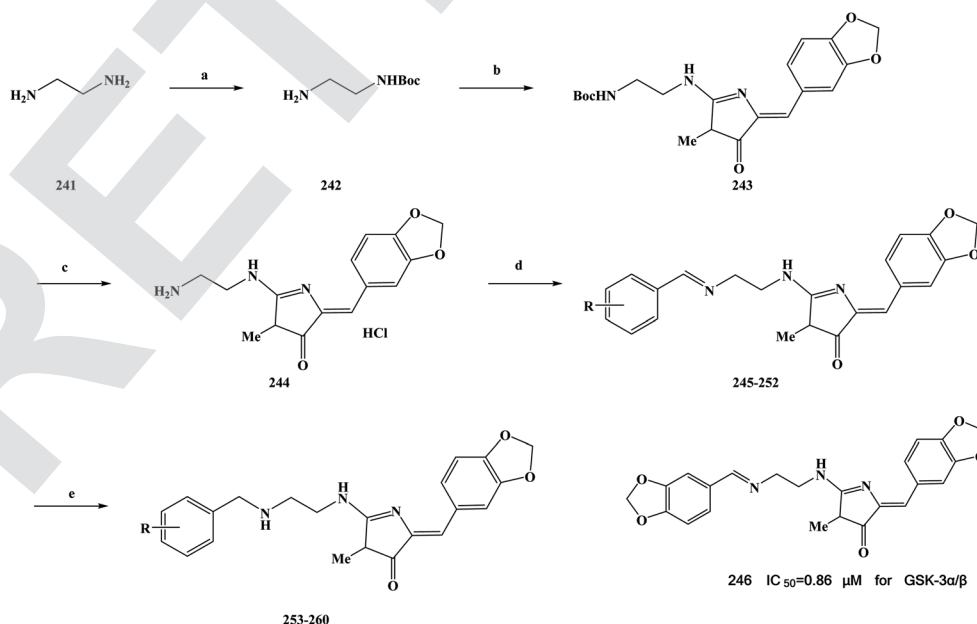


replication and infectivity of different viruses is the key to the successful development of anti-viral Hsp90 inhibitors. In general, compound **237** has good anti-viral activity and high selectivity, which needs to be further studied.

2.5 Other types of activity

Natural products from the sea have been paid much attention, especially the compounds with rich biological activity separated from marine sponges.²¹⁹ In the process of activity screening, the diversity biological activity of these compounds was observed, and these marine natural products were also found to have diverse chemical structures.²²⁰ The abundant metabolites produced by marine organisms offer a promising contribution to the discovery of new drugs for the treatment of diseases, or to the development of lead compounds for structural modification. Among these marine natural products, a class of marine alkaloids containing 2-aminoidazolidone structures have been widely studied.²²¹ This chemical structure is an attractive structure with the potential for further structural modification, which can be used as a starting point for the development and research of similar drugs with diverse chemical structures biological activities. These target compounds modified on the basis of the 2-aminoidazolidone structure constitute a huge candidate drug screening library, which can be fully researched and developed.^{222,223} Dispacamide is a marine alkaloid with anti-histamine activity. Polyandrocarpamine A (**7**) is selectively cytotoxicity to SF268 cell line (central nervous system) with a GI₅₀ value of 65 μ M. These marine alkaloids obtained from marine organisms were found to contain a 2-aminoidazolidone structure by chemical analysis. At the same time, leucettamine B (**9**) is also a marine alkaloid separated from marine sponges, which has been obtained by chemical synthesis²²⁴ However, no biological activity related to this substance has been reported so far. These marine

alkaloids containing 2-aminoidazolidone structures have been found to have abundant biological activity in activity studies. Based on previous work, Debdab *et al.*²²⁵ designed derivatives of the marine alkaloid leucettamine B. From the chemical structure of the designed target compounds **253–260**, it could be seen that these target compounds are derivatives of *N*-functionalized leucettamine B, in which aryl methyl groups are linked to the 2-amino structure *via* a short amino-alkyl chain. During the synthesis of the target compounds, they selected the method described in Scheme 12 to synthesis the marine alkaloid Lucettamine B derivatives. The target compounds were synthesized in five steps, the starting material with commercial ethylenediamine (compound **241**). The first step was to prepare a single *N*-Boc protected diamine (compound **242**) in methanol with ethylenediamine and di-*tert*-butyldicarbonate according to a method in the literature, in a yield of 65%. The second step of the reaction is the key step to compounds **242** and (5*Z*) 5-[(1,3-benzodioxo-5-yl)methylene]-3-methyl-2-ethylsulfanyl-3,5-dihydroimidazol-4-one with a microwave irradiation synthesis used to generate the intermediate compound **243** (yield 69%). The reaction was carried out for 2 h at 105 °C to achieve good conversion. In the third step, the protection group of compound **243** was removed, meaning that the *N*-Boc group was cracked. Compound **243** was treated with acetyl chloride and methanol in acetonitrile at room temperature, and a high yield of compound **244** (yield 98%) could be obtained by effective deprotection. In the fourth step, compound **244** was reacted with substituted benzaldehyde in methanol, and triethanolamine was used as the catalyst for the reaction. Compounds **245–252** were obtained after reaction at 50 °C for 24 h, reaching yields of 63–81%. In the final step, compounds **245–252** were reacted with NaBH₄ in methanol at 50 °C for 24 h to obtain the target compounds **253–260** (yield 72–97%). According to this method, the target



Scheme 12 Synthesis of the leucettamine B analogue compounds **253–260**. Reagents and conditions: (a) (Boc)₂O, MeOH, 0 °C, 10 min, then 25 °C, 1 h; (b) 105 °C, 2 h; (c) MeCOCl, MeOH, MeCN, 25 °C, 24 h; (d) Et₃N, MeOH, 50 °C, 24 h; (e) NaBH₄, MeOH, 50 °C, 24 h.

Table 12 The kinases inhibition values

Compounds	IC ₅₀ (μM)		
	CK1	CDK5/p25	GSK-3α/β
244	>100	>100	1.4
245	>100	>100	7.0
246	>100	>100	0.86
247	>100	>100	15.0
248	>100	>100	38.0
249	>100	>100	7.3
253	>100	>100	15.0
254	>100	>100	>10
255	>100	>100	>10
256	>100	>100	>10
257	>100	>100	>10
Leucettamine B	>100	>100	>10

compounds **253–260** were prepared in five steps with a total yield of 24% to 31%. This synthesis route is relatively simple to use, the reagents used are cheap and easy to obtain, and the yield is moderate to good, laying a foundation for its future industrialization. At the same time, the new method of microwave irradiation synthesis used in this synthesis route greatly reduced the reaction time and increased the reaction yield. During the biological activity study, the group tested the target compounds **253–260**, intermediate compound **244** and **245–252** *in vitro* for three protein kinases associated with Alzheimer's disease (Table 12). The protein kinases include casein kinase 1α/β (CK1α/β), cyclin-dependent kinase 5 (CDK5)/p25 and glycogen synthase kinase 3α/β (GSK-3α/β). *In vitro* biological activity test results showed that some marine alkaloid leucettamine B analogues have moderate to strong inhibitory activity of GSK-3α/β, with IC₅₀ values of between 0.86 and 38 μM. The intermediate compound **246** with an IC₅₀ of 0.86 μM, which inhibited GSK-3α/β, was the most active of the compounds tested. These compounds showed poor CK1α/β and CDK5/p25 inhibitory activity compared with GSK-3α/β inhibitory activity; obviously the good selective inhibition of GSK-3α/β exists. The target compounds **253–257** were product obtained from the addition reaction on the basis of the intermediate compounds **245–249**, but it was found in the activity test results that with the addition of the imine group, the inhibitory activity on GSK-3α/β was lost. After the comparison of the activity test results of compounds **253–257**, a drug SAR was established. The results of SAR showed that the types of substituents and the positions of the substituents on the phenyl groups had certain effects on the biological activity. At the same time, the intermediate compound **244** showed high inhibitory activity to GSK-3α/β, with an IC₅₀ of 1.4 μM, and a good selective inhibitory effect. The amino group in the aminoalkyl chain of compound **244** was replaced, and it retained good inhibitory activity, meaning that it can be further modified as a skeleton structure (the lead compound) for the discovery of novel selective GSK-3α/β inhibitors. In general, according to the available data, compound **246** could be used as a selective GSK-3α/β inhibitor for further *in vivo* activity evaluation and pharmacokinetic property studies.

3. Conclusions and prospects

Many types of compounds with biological activity have been extracted and separated from marine organisms, with marine alkaloids being some of the earliest compounds studied. *In vitro* or *in vivo* biological activity screening of marine alkaloids has shown that these compounds have diverse biological activities, including anti-tumor, anti-fungal, anti-viral, anti-malaria, anti-osteoporosis, and other biological activities, with the anti-tumor activity being studied in depth. With the increasing interest in the research of marine drugs, marine alkaloids have a promising future and may be further converted into anti-tumor, anti-viral, and anti-fungal clinical drugs or lead compounds. The amounts of these compounds that can be separated are limited, and it is easy to damage the marine organism environment. In addition, these lead compounds have the disadvantages of high cytotoxicity and low selectivity, which limit their clinical research potential and their industrial development. Therefore, marine alkaloid derivatives or analogues with new structures have been designed by applying modern drug design concepts to the structural modification of lead compounds, and these compounds were obtained *via* efficient classical or modern methods of chemical synthesis. In the activity screening, these marine alkaloid derivatives or analogues also show biological activity diversity and could make up for the shortcomings of natural marine alkaloids. This has become a focus of research. This work systematically reviewed the recent advances in marine alkaloid derivatives and analogues in the field of medical chemistry over the last 10 years (2010–2019). In the review process, we divided marine alkaloid derivatives and analogues into five types from the perspective of biological activity, including research into anti-tumor, anti-malarial, anti-bacterial, anti-viral (HCV), and other types, and specifically elaborated on these activities. We discussed the optimization process, chemical synthesis, biological activity evaluation and SAR of each type of compound. On this basis, we evaluated the synthesis route of each type of compounds, so as to provide a necessary reference for the future optimization of these processes. The SAR may provide a reasonable approach for the design and development of novel marine alkaloid derivatives or analogues. It is an important way to obtain lead compounds from marine alkaloids and it is very promising to obtain a large number of derivatives and analogues *via* structural modification. These derivatives and analogues have diverse biological activity, which, combined with the SAR, provides a good research platform for the discovery of new drugs in the future. Researchers should take advantage of the complex and special chemical structures and diverse biological activities of natural marine alkaloids. We should use modern drug design and synthesis methods to obtain more derivatives and analogues for activity screening and clinical research. As a result, the earlier these marine alkaloid derivatives and analogues enter the market, the better they will serve the public to prevent and treat diseases.



Conflicts of interest

There are no conflicts to declare.

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References

- 1 T. Rodrigues, D. Reker, P. Schneider and G. Schneider, *Nat. Chem.*, 2016, **8**, 531–541.
- 2 S. Z. Wang, G. Q. Dong and C. Q. Sheng, *Chem. Rev.*, 2019, **119**, 4180–4220.
- 3 J. Kobayashi, *Chem. Pharm. Bull.*, 2016, **64**, 1079–1083.
- 4 W. B. Wu, Y. Tang, J. L. Yang, E. Idehen and S. M. Sang, *J. Agric. Food Chem.*, 2018, **66**, 8005–8014.
- 5 Y. G. Han, Y. Luo, S. R. Qin, L. Xi, B. Wan and L. F. Du, *Pestic. Biochem. Physiol.*, 2014, **111**, 14–18.
- 6 M. G. Ciulla and K. Kumar, *Tetrahedron Lett.*, 2018, **59**, 3223–3233.
- 7 M. J. Martinez, L. M. Olmo and P. B. Benito, *Stud. Nat. Prod. Chem.*, 2015, **30**, 393–418.
- 8 M. M. Gendy, M. Shaaban, K. A. Shaaban, A. M. Bondkly and H. Laatsch, *J. Antibiot.*, 2008, **61**, 149–157.
- 9 U. Battaglia and C. J. Moody, *J. Nat. Prod.*, 2010, **73**, 1938–1939.
- 10 L. H. Stoodley, J. W. Costerton and P. Stoodley, *Nat. Rev. Microbiol.*, 2004, **2**, 95–108.
- 11 E. Tee, T. Karoli, S. Ramu, J. Huang, M. Butler and M. Cooper, *J. Nat. Prod.*, 2010, **73**, 1940–1942.
- 12 E. S. Zuniga, A. Korkegian, S. Mullen, E. J. Hembre, P. L. Ornstein, G. Cortez, K. Biswas, N. Kumar, J. Cramer, T. Masquelin, P. A. Hipskind, J. Odingo and T. Parish, *Bioorg. Med. Chem.*, 2017, **25**, 3922–3946.
- 13 J. J. Richards and C. Melander, *ChemBioChem*, 2010, **10**, 2287–2294.
- 14 W. B. Parker, *Virus Res.*, 2005, **107**, 165–171.
- 15 P. Lv, Y. L. Chen, T. Z. Shi, X. W. Wu, Q. X. Li and R. M. Hua, *Pestic. Biochem. Physiol.*, 2018, **147**, 3–10.
- 16 X. Y. Li, G. F. Hao, Q. M. Wang, Z. Chen, Y. Ding, L. Yu, D. Y. Hu and B. A. Song, *Oncotarget*, 2017, **8**, 82446–82458.
- 17 T. L. Simmons, E. Andrianasolo, K. M. Phail, P. Flatt and W. H. Gerwick, *Mol. Cancer Ther.*, 2005, **2005**(4), 333–342.
- 18 C. Cuevas and A. Francesch, *Nat. Prod. Rep.*, 2009, **26**, 322–337.
- 19 R. J. Worthington, J. J. Richards and C. Melander, *Org. Biomol. Chem.*, 2012, **10**, 7457–7474.
- 20 B. J. Petek, E. T. Loggers, S. M. Pollack and R. L. Jones, *Mar. Drugs*, 2015, **13**, 974–983.
- 21 F. Mascilini, G. Amadio, M. G. Stefano, M. Ludovisi, A. D. Legge, C. Conte, R. D. Vincenzo, C. Ricci, V. Masciullo, V. Salutari, G. Scambia and G. Ferrandina, *OncoTargets Ther.*, 2014, **7**, 1273–1284.
- 22 E. Adrian, *Curr. Pharm. Des.*, 2007, **13**, 3417–3426.
- 23 B. Haefner, *Drug Discovery Today*, 2003, **8**, 536–544.
- 24 K. B. Glaser and A. M. Mayer, *Biochem. Pharmacol.*, 2009, **78**, 440–448.
- 25 R. A. Davis, L. V. Christensen, A. D. Richardson, R. M. Rocha Da and C. M. Ireland, *Mar. Drugs*, 2003, **1**, 27–33.
- 26 T. F. Molinski, D. S. Dalisay, S. L. Lievens and J. P. Saludes, *Nat. Rev. Drug Discovery*, 2008, **8**, 69–85.
- 27 G. Schwartzmann, A. B. Rocha, J. Mattei and R. Lopes, *Expert Opin. Invest. Drugs*, 2003, **12**, 1367–1383.
- 28 R. Singh, M. Sharma, P. Joshi and D. S. Rawat, *Anti-Cancer Agents in Medicinal Chemistry*, 2008, **8**, 603–617.
- 29 J. W. Blunt, B. R. Copp, M. H. Munro, P. T. Northcote and M. R. Prinsep, *Nat. Prod. Rep.*, 2006, **23**, 26–78.
- 30 G. B. Sela, R. Epelbaum and M. Schaffer, *Curr. Med. Chem.*, 2010, **17**, 190–197.
- 31 G. Kuznetsov, K. T. Dyke, M. J. Towle, H. Cheng, J. Liu, J. P. Marsh, S. E. Schiller, M. R. Spyvee, H. Yang, B. M. Seletsky, C. J. Shaffer, V. Marceau, Y. Yao, E. M. Suh, S. Campagna, F. G. Fang, J. J. Kowalczyk and B. A. Littlefield, *Mol. Cancer Ther.*, 2009, **8**, 2852–2860.
- 32 E. A. Guzman, J. D. Johnson, M. K. Carrier, C. I. Meyer, T. P. Pitts, S. P. Gunasekera and A. E. Wright, *Anti-Cancer Drugs*, 2009, **20**, 149–155.
- 33 S. Deslandes, S. Chassaing and E. Delfourne, *Mar. Drugs*, 2009, **7**, 754–786.
- 34 R. Dasari and A. Kornienko, *Chem. Heterocycl. Compd.*, 2014, **50**, 139–144.
- 35 D. V. La-Barbera, K. Modzelewska, A. I. Glazar, P. D. Gray, M. Kaur, T. Liu, D. Grossman, M. K. Harper, S. K. Kuwada, N. Moghal and C. M. Ireland, *Anti-Cancer Drugs*, 2009, **20**, 425–436.
- 36 R. Campana, L. Casettari, L. Fagioli, M. Cespi, G. Bonacucina and W. Baffone, *Int. J. Food Microbiol.*, 2017, **241**, 132–140.
- 37 F. M. Ranaivoson, B. Gigant, S. Berritt, M. Joullie and M. Knossow, *Acta Crystallogr.*, 2012, **68**, 927–934.
- 38 E. Hamel, *Cell Biochem. Biophys.*, 2003, **38**, 1–22.
- 39 J. Kluza, M. A. Gallego, A. Loyens, J. C. Beauvillain, J. M. Sousa-Faro, C. Cuevas, P. Marchetti and C. Bailly, *Cancer Res.*, 2006, **66**, 3177–3187.
- 40 C. Ballot, J. Kluza, S. Lancel, A. Martoriati, S. M. Hassoun, L. Mortier, J. C. Vienne, G. Briand, P. Formstecher, C. Bailly, R. Nevriere and P. Marchetti, *Apoptosis*, 2010, **15**, 769–781.



- 41 J. Sun, J. Wu, B. An, N. J. Voogd, W. Cheng and W. Lin, *Mar. Drugs*, 2018, **16**, 1–15.
- 42 K. Vermeulen, D. R. Bockstaele and Z. N. Berneman, *Cell Proliferation*, 2003, **36**, 131–149.
- 43 A. R. Cuddihy and M. J. Connell, *Int. Rev. Cytol.*, 2003, **222**, 99–140.
- 44 C. H. Gowan, *Mutat. Res.*, 2003, **532**, 75–84.
- 45 R. G. Berlinck, R. Britton, E. Piers, L. Lim, M. Roberge, R. Moreira da Rocha and R. Andersen, *J. Org. Chem.*, 1998, **63**, 9850–9856.
- 46 X. Jiang, B. Zhao, R. Britton, L. Y. Lim, D. Leong, J. S. Sanghera, B. B. Zhou, E. Piers, R. J. Andersen and M. Roberge, *Mol. Cancer Ther.*, 2004, **3**, 1221–1227.
- 47 R. Boutros, V. Lobjois and B. Ducommun, *Nat. Rev. Cancer*, 2007, **7**, 495–507.
- 48 S. Deslandes, S. Chassaing and E. Delfourne, *Mar. Drugs*, 2009, **7**, 754–786.
- 49 B. Hugon, B. Pfeiffer, P. Renard and M. Prudhomme, *Tetrahedron Lett.*, 2003, **44**, 3927–3930.
- 50 B. Hugon, F. Anizon, C. Bailly, R. M. Golsteyn, A. Pierré, S. Léonce, J. Hickman, B. Pfeiffer and M. Prudhomme, *Bioorg. Med. Chem.*, 2007, **15**, 5965–5980.
- 51 E. Piers, R. Britton and R. J. Andersen, Improved synthesis of isogranulatimide, a G2 checkpoint inhibitor, *J. Org. Chem.*, 2000, **65**, 530–535.
- 52 M. Contour-Galcéra, A. Sidhu, P. Plas and P. Roubert, *ACS Med. Chem. Lett.*, 2005, **15**, 3555–3559.
- 53 S. Deslandes, D. Lamoral-Theys, C. Frongia, S. Chassaing, C. Bruyère, O. Lozach, L. Meijer, B. Ducommun, R. Kiss and E. Delfourne, *Eur. J. Med. Chem.*, 2012, **54**, 626–636.
- 54 D. J. Faulkner, *Nat. Prod. Rep.*, 2002, **19**, 1–48.
- 55 A. Casapullo, G. Bifulco, I. Bruno and R. Riccio, *J. Nat. Prod.*, 2000, **63**, 447–451.
- 56 B. Bao, Q. Sun, X. Yao, J. Hong, C. Lee, C. J. Sim and J. H. Jung, *J. Nat. Prod.*, 2005, **68**, 711–715.
- 57 W. Gul and M. T. Hamann, *Life Sci.*, 2005, **78**, 442–453.
- 58 H. R. Bokesch, L. K. Pannell, T. C. McKee and M. R. Boyd, *Tetrahedron Lett.*, 2000, **41**, 6305–6308.
- 59 C. J. Moody and J. R. Roffey, *Arkivoc*, 2002, **1**, 393–401.
- 60 F. Y. Miyake, K. Yakushijin and D. A. Horne, *Org. Lett.*, 2000, **2**, 2121–2123.
- 61 P. M. Fresneda, P. Molina and M. A. Sanz, *Synlett*, 2001, **2**, 218–221.
- 62 B. Jiang and X. H. Gu, *Bioorg. Med. Chem.*, 2008, **8**, 363–371.
- 63 B. Jiang, C. G. Yang, C. Xiong and J. Wang, *Bioorg. Med. Chem.*, 2001, **9**, 1149–1154.
- 64 B. Jiang, W. N. Xiong and C. G. Yang, *Bioorg. Med. Chem. Lett.*, 2011, **11**, 475–477.
- 65 W. N. Xiong, C. G. Yang and B. Jiang, *Bioorg. Med. Chem.*, 2001, **9**, 1773–1780.
- 66 P. Diana, A. Carbone, P. Barraja, A. Montalbano, A. Martorana, G. Dattolo, O. Gia, L. D. Via and G. Cirrincione, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 2342–2346.
- 67 P. Diana, A. Carbone, P. Barraja, A. Martorana, O. Gia, L. D. Via and G. Cirrincione, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 6134–6137.
- 68 P. Diana, A. Carbone, P. Barraja, H. H. Kelter, G. Fiebig and G. Cirrincione, *Bioorg. Med. Chem.*, 2010, **18**, 4524–4529.
- 69 D. Kumar, N. M. Kumar, K. H. Chang, R. Gupta and K. Shah, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 5897–5900.
- 70 A. Carbone, B. Parrino, G. D. Vita, A. Attanzio, V. Spanò, A. Montalbano, P. Barraja, L. Tesoriere, M. A. Livrea, P. Diana and G. Cirrincione, *Mar. Drugs*, 2015, **13**, 460–492.
- 71 B. Parrino, A. Carbone, G. D. Vita, C. Ciancimino, A. Attanzio, V. Spanò, A. Montalbano, P. Barraja, L. Tesoriere, M. A. Livrea, P. Diana and G. Cirrincione, *Mar. Drugs*, 2015, **13**, 1901–1924.
- 72 A. Carbone, B. Parrino, P. Barraja, V. Spanò, G. Cirrincione, P. Diana, A. Maier, G. Kelter and H. H. Fiebig, *Mar. Drugs*, 2013, **11**, 643–654.
- 73 P. Vineis and C. P. Wild, *Lancet*, 2014, **383**, 549–557.
- 74 W. R. Sawadogo, M. Schumacher, M. H. Teiten, C. Cerella, M. Dicato and M. Diederich, *Molecules*, 2013, **18**, 3641–3673.
- 75 I. Bhatnagar and S. K. Kim, *Mar. Drugs*, 2010, **8**, 2702–2720.
- 76 D. Sipkema, M. C. Franssen, R. Osinga, J. Tramper and R. H. Wijffels, *Mar. Biotechnol.*, 2005, **7**, 142–162.
- 77 A. Carbone, M. Pennati, B. Parrino, A. Lopergolo, P. Barraja, A. Montalbano, V. Spanò, S. Sbarra, V. Doldi and M. Cesare, *J. Med. Chem.*, 2013, **56**, 7060–7072.
- 78 A. Carbone, M. Pennati, P. Barraja, A. Montalbano, B. Parrino, V. Spanò, A. Lopergolo, S. Sbarra, V. Doldi and N. Zaffaroni, *Curr. Med. Chem.*, 2014, **21**, 1654–1666.
- 79 V. Stonik and S. Fedorov, *Mar. Drugs*, 2014, **12**, 636–671.
- 80 D. T. Youssef, L. AShaala, G. AMohamed, J. M. Badr, F. H. Bamanie and S. R. Ibrahim, *Mar. Drugs*, 2014, **12**, 1911–1923.
- 81 J. R. Pereira, F. F. Hilário, A. B. Lima, M. L. Silveira, L. M. Silva, R. B. Alves, R. P. Freitas and F. P. Varotti, *Prev. Nutr.*, 2012, **2**, 145–148.
- 82 P. AMachado, F. F. Hilário, L. O. Carvalho, M. L. Silveira, R. B. Alves, R. P. Freitas and E. S. Coimbra, *Chem. Biol. Drug Des.*, 2012, **80**, 745–751.
- 83 G. Iannolo, C. Conticello, L. Memeo and R. Maria, *Critical Reviews in Oncology/Hematology*, 2008, **66**, 42–51.
- 84 D. Rossi and G. Gaidano, *Haematologica*, 2003, **88**, 212–218.
- 85 I. Decordier, L. Dillen, E. Cundari and M. K. Volders, *Mutagenesis*, 2002, **17**, 337–344.
- 86 I. Decordier, E. Cundari and M. K. Volders, *Mutagenesis*, 2005, **20**, 173–179.
- 87 A. M. Gonçalves, A. B. Lima, M. C. Barbosa, L. F. Camargos, J. T. Oliveira, C. S. Barbosa, J. A. Villar, A. C. Costa, I. V. Silva, L. M. Silva, F. P. Varotti, F. V. Santos and G. H. Viana, *Mar. Drugs*, 2014, **12**, 4361–4378.
- 88 K. H. Khan, M. B. Codesido and L. R. Molife, *Critical Reviews in Oncology/Hematology*, 2013, **90**, 200–219.
- 89 M. K. Volders, T. Sofuni, M. Aardema, S. Albertini, D. Eastmond, M. Fenech, M. Ishidate, S. Kirchner, E. Lorge and T. Morita, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2003, **540**, 153–163.
- 90 R. A. Hill, *Annu. Rep. Prog. Chem., Sect. B: Org. Chem.*, 2005, **101**, 124–136.



- 91 Y. Nakao and N. Fusetani, *J. Nat. Prod.*, 2007, **2007**(70), 689–710.
- 92 M. Chittchang, P. Batsomboon, S. Ruchirawat and P. Ploypradith, *ChemMedChem*, 2009, **4**, 457–465.
- 93 S. Mabuchi, T. Hisamatsu, C. Kawase, M. Hayashi, K. Sawada, K. Mimura, K. Takahashi, T. Takahashi, H. Kurachi and T. Kimura, *Clin. Cancer Res.*, 2011, **17**, 4462–4473.
- 94 M. Sugumaran and W. E. Robinson, *Mar. Drugs*, 2010, **8**, 2906–2935.
- 95 H. Fan, J. Peng, M. T. Hamann and J. F. Hu, *Chem. Rev.*, 2008, **108**, 264–287.
- 96 M. Tsuda, K. Nozawa, K. Shimbo and J. Kobayashi, *J. Nat. Prod.*, 2003, **66**, 292–294.
- 97 L. V. Frolova, I. V. Magedov, A. E. Romero, M. Karki, I. Otero, K. Hayden, N. M. Evdokimov, L. M. Y. Banuls, S. K. Rastogi, W. R. Smith, S. L. Lu, R. Kiss, C. B. Shuster, E. Hamel, T. Betancourt, S. Rogelj and A. Kornienko, *J. Med. Chem.*, 2013, **56**, 6886–6990.
- 98 R. Scott, M. M. R. Karki, M. R. Reisenauer, R. Rodrigues, R. Dasari, W. R. Smith, S. C. Pelly, W. A. Otterlo, C. B. Shuster, S. Rogelj, I. V. Magedov, L. V. Frolova and A. Kornienko, *ChemMedChem*, 2014, **9**, 1428–1435.
- 99 D. J. Newman and G. M. Cragg, *J. Nat. Prod.*, 2007, **70**, 461–477.
- 100 K. H. Attmann and J. Gertsch, *Nat. Prod. Rep.*, 2007, **24**, 327–357.
- 101 M. F. Mehbub, J. Lei, C. Franco and W. Zhang, *Mar. Drugs*, 2014, **12**, 4539–4577.
- 102 S. Nag, D. H. Nadkarni, J. J. Qin, S. Voruganti, T. Nguyen, S. Xu, W. Wang, H. Wang, S. E. Velu and R. Zhang, *Mol. Cell. Pharmacol.*, 2012, **4**, 69–81.
- 103 F. M. Deane, E. C. Sullivan, A. R. Maguire, J. Gilbert, J. A. Sakoff, A. M. Cluskey and F. O. Carthy, *Org. Biomol. Chem.*, 2011, **11**, 1334–1344.
- 104 G. Lang, A. Pinkert, J. W. Blunt and M. H. Munro, *J. Nat. Prod.*, 2005, **68**, 1796–1798.
- 105 J. Ponder, B. H. Yoo, A. D. Abraham, Q. Li, A. K. Ashley, C. L. Amerin, Q. Zhou, B. G. Reid, P. Reigan and R. Hromas, *Mar. Drugs*, 2011, **9**, 2397–2408.
- 106 L. M. West, P. T. Northcote and C. N. Battershill, *J. Org. Chem.*, 2000, **65**, 445–449.
- 107 S. Boucle, C. Melin, M. Clastre and J. Guillard, *Mar. Drugs*, 2001, **513**, 655–665.
- 108 A. Martins, H. Vieira, H. Gaspar and S. Santos, *Mar. Drugs*, 2014, **12**, 1066–1101.
- 109 M. Köck, J. Muñoz, C. Cychon, C. Timm and G. Schmidt, *Phytochem. Rev.*, 2013, **12**, 391–406.
- 110 J. R. Pereira, F. F. Hilário, A. B. Lima, M. L. Silveira, L. M. Silva, R. B. Alves, R. P. De Freitas, F. P. Varotti and G. H. Viana, *Biomedicine & Preventive Nutrition*, 2012, **2**, 145–148.
- 111 R. R. Soares, J. M. Silva, B. C. Carlos, C. C. Fonseca, L. S. Souza, F. V. Lopes, R. M. Paula Dias, P. O. Moreira, C. Abramo, G. H. Viana, F. P. Varotti, A. D. Silva and K. K. Scopel, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 2308–2313.
- 112 I. Leto, M. Coronello, C. Righeschi, M. C. Bergonzi, E. Mini and A. R. Bilia, *ChemMedChem*, 2016, **11**, 1745–1751.
- 113 M. Burow, A. Bergner, J. Gershenzon and U. Wittstock, *Plant Mol. Biol.*, 2007, **63**, 49–61.
- 114 R. M. Viana, A. P. Butera, E. S. Santos, C. A. Tischer, R. B. Alves, R. P. Freitas, L. Guimaraes, F. Varotti, G. H. Viana and C. S. Nascimento, *J. Chem. Inf. Model.*, 2016, **56**, 571–579.
- 115 J. Beard, *Nature*, 2013, **92**, 60–61.
- 116 M. Lu, L. Sun, J. Zhou and J. Yang, *Tumor Biol.*, 2014, **35**, 5307–5314.
- 117 M. Fenech, *Nat. Protoc.*, 2007, **2**, 1084–1104.
- 118 J. T. Oliveira, M. C. Barbosa, L. F. Camargos, I. V. Silva, F. P. Varotti, L. M. Silva, L. M. Moreira, J. P. Lyon, V. J. Santos and F. V. Santos, *Cytotechnology*, 2017, **69**, 699–710.
- 119 E. Elhalem, B. N. Bailey, R. Docampo, S. H. Szajnman and J. B. Rodriguez, *J. Med. Chem.*, 2002, **86**, 3984–3999.
- 120 G. G. Linares, S. Gismondi, N. O. Codesido, S. N. Moreno, R. Docampo and J. B. Rodriguez, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 5068–5071.
- 121 H. Noedl, C. Wongsrichanalai, R. S. Miller, K. S. Myint, S. Looareesuwan, Y. Sukthana, V. Wongchotigul, H. Kollaritsch, G. Wiedermann and W. H. Wernsdorfer, *Exp. Parasitol.*, 2002, **102**, 157–163.
- 122 R. H. Lee, B. Kim, I. Choi, H. Kim, S. Choi, K. Suh, Y. C. Bae and J. S. Jung, *Cell. Physiol. Biochem.*, 2004, **14**, 311–324.
- 123 F. V. Santos, M. Andreo, A. L. Nasser, L. M. Moreira, W. Vilegas, I. M. Cólus and E. A. Varanda, *Arch. Biol. Sci.*, 2013, **65**, 191–195.
- 124 D. M. Cottrell, J. Capers, M. M. Salem, K. DeLuca-Fradley, S. L. Croft and K. A. Werbovetz, *Bioorg. Med. Chem.*, 2004, **12**, 2815–2824.
- 125 C. G. Havens, N. Bryant, L. Asher, L. Lamoreaux, S. Perfetto, J. J. Brendle and K. A. Werbovetz, *Mol. Biochem. Parasitol.*, 2000, **110**, 223–236.
- 126 C. C. Gomes, L. M. Moreira, V. J. Santos, A. S. Ramos, J. P. Lyon, C. P. Soares and F. V. Santos, *Genet. Mol. Biol.*, 2011, **34**, 116–121.
- 127 P. L. Olive and J. P. Banáth, *J. Nat. Prod.*, 2006, **1**, 23–29.
- 128 A. R. Collins, *Mol. Biotechnol.*, 2004, **26**, 249–261.
- 129 V. S. Gontijo, P. F. Espuri, R. B. Alves, L. F. Camargos, F. V. Dos Santos, W. A. Souza Judice, M. J. Marques and R. P. Freitas, *Eur. J. Med. Chem.*, 2015, **101**, 24–33.
- 130 L. Recio, C. Hobbs, W. Caspary and K. L. Witt, *J. Toxicol. Sci.*, 2010, **35**, 149–162.
- 131 C. Aiub, A. Giannerini, F. Ferreira, J. Mazzei, L. Stankevics, G. L. Hajdu, P. Guimarães, E. Hajdu and I. Felzenszwalb, *Mutat. Res.*, 2006, **611**, 34–41.
- 132 V. Alcolea, D. Plano, I. Encio, J. A. Palop, A. K. Sharma and C. Sanmartín, *Eur. J. Med. Chem.*, 2016, **123**, 407–418.
- 133 R. Abbotts, N. Thompson and S. Madhusudan, *Cancer Manage. Res.*, 2014, **6**, 77–92.
- 134 M. K. Volders, S. Bonassi, S. Knasmueller, N. Holland, C. Bolognesi and M. F. Fenech, *Mutat. Res.*, 2014, **759**, 49–58.



- 135 P. Villani, M. Spanò, F. Pacchierotti, M. Weimer and E. Cordelli, *Reprod. Toxicol.*, 2010, **30**, 44–49.
- 136 L. Stankevicius, C. Aiub, L. C. Santa Maria, G. L. Hajdu and I. Felzenszwalb, *Toxicol. In Vitro*, 2008, **22**, 1869–1877.
- 137 K. Utani, Y. Kohno, A. Okamoto and N. Shimizu, *PLoS One*, 2010, **5**, 10089.
- 138 M. C. Barbosa, C. S. Barbosa, J. T. Oliveira, N. C. Moreira, N. R. Martins, G. K. Gomes, C. A. Caldeira, M. L. Costa, D. S. Martins, L. Guimaraes, C. S. Nascimento, F. P. Varotti, G. H. Viana and F. V. Santos, *Mutat. Res., Genet. Toxicol. Environ. Mutagen.*, 2018, **825**, 31–39.
- 139 R. Tandon, I. Singh, V. Luxami, N. Tandon and K. Paul, *Chem. Rec.*, 2018, **18**, 1–33.
- 140 R. L. Siegel, K. D. Miller and A. Jemal, *Ca-Cancer J. Clin.*, 2018, **68**, 7–30.
- 141 Y. Sumii, N. Kotoku, A. Fukuda, T. Kawachi, M. Arai and M. Kobayashi, *Mar. Drugs*, 2015, **13**, 7419–7432.
- 142 D. J. Newman and G. M. Cragg, *J. Nat. Prod.*, 2007, **70**, 461–477.
- 143 D. J. Newman and G. M. Cragg, *Future Med. Chem.*, 2009, **1**, 1415–1427.
- 144 D. D. Luo, K. A. Carter, D. Miranda and J. F. Lovell, *Adv. Sci.*, 2017, **4**, 1600106.
- 145 K. V. Kudryavtsev, C. C. Yu, P. M. Ivantcova, V. I. Polshakov, A. V. Churakov, S. Brase, N. S. Zefirov and J. H. Guh, *Chem.-Asian J.*, 2015, **10**, 383–389.
- 146 B. C. Gerwick and T. C. Sparks, Natural products for pest control: an analysis of their role, value and future, *Pest Manage. Sci.*, 2014, **70**, 1169–1185.
- 147 O. F. Hüter, *Phytochem. Rev.*, 2011, **10**, 185–194.
- 148 T. Rodrigues, D. Reker, P. Schneider and G. Schneider, *Nat. Chem.*, 2016, **8**, 531–541.
- 149 K. Kaur, V. Kumar, A. K. Sharma and G. K. Gupta, *Eur. J. Med. Chem.*, 2014, **77**, 121–133.
- 150 I. Boulogne, P. Petit, H. Ozier-Lafontaine, L. Desfontaines and G. Loranger-Merciris, *Environ. Chem. Lett.*, 2012, **10**, 325–347.
- 151 T. C. Sparks, D. R. Hahn and N. V. Garizi, *Pest Manage. Sci.*, 2017, **73**, 700–715.
- 152 S. Miresmailli and M. B. Isman, *Trends Plant Sci.*, 2014, **19**, 29–35.
- 153 J. H. Schrittwieser and V. Resch, *RSC Adv.*, 2013, **3**, 17602–17632.
- 154 B. Parrino, A. Attanzio, V. Spano, S. Cascioferro, A. Montalbano, P. Barraja, L. Tesoriere, P. Diana, G. Cirrincione and A. Carbone, *Eur. J. Med. Chem.*, 2017, **138**, 371–383.
- 155 M. Ishikura, T. Abe, T. Choshi and S. Hibino, *Nat. Prod. Rep.*, 2015, **32**, 1389–1471.
- 156 M. Salmoun, C. Devijver, D. Daloze, J. C. Braekman and R. W. Soest, *J. Nat. Prod.*, 2002, **65**, 1173–1176.
- 157 V. Spano, A. Attanzio, S. Cascioferro, A. Carbone, A. Montalbano, P. Barraja, L. Tesoriere, G. Cirrincione, P. Diana and B. Parrino, *Mar. Drugs*, 2016, **14**, 226.
- 158 S. Cascioferro, A. Attanzio, V. D. Sarno, S. Musella, L. Tesoriere, G. Cirrincione, P. Diana and B. Parrino, *Mar. Drugs*, 2019, **17**, 35.
- 159 N. Mexia, G. Gaitanis, A. Velegraki, A. Soshilov, M. S. Denison and P. Magiatis, *Arch. Biochem. Biophys.*, 2015, **571**, 16–20.
- 160 C. Tan, Y. Zhang, N. Kong, Y. Di and X. Hao, *Helv. Chim. Acta*, 2015, **98**, 72–77.
- 161 X. Shang, X. Guo, B. Li, H. Pan, J. Zhang, Y. Zhang and X. Miao, *J. Ethnopharmacol.*, 2016, **192**, 350–361.
- 162 H. M. Viart, A. Bachmann, W. Kayitare and R. Sarpong, *J. Am. Chem. Soc.*, 2017, **139**, 1325–1329.
- 163 P. Zhang, X. Sun, B. Xu, K. Bijian, S. Wan, G. Li, M. A. Jamali and T. Jiang, *Eur. J. Med. Chem.*, 2011, **46**, 6089–6097.
- 164 S. Manda, S. Sharma, A. Wani, P. Joshi, V. Kumar, S. K. Guru, S. S. Bharate, S. Bhushan, R. A. Vishwakarma, A. Kumar and S. B. Bharate, *Eur. J. Med. Chem.*, 2016, **107**, 1–11.
- 165 T. Suzuki, M. N. Khan, H. Sawada, E. Imai, Y. Itoh, K. Yamatsuta, N. Tokuda, J. Takeuchi, T. Seko, H. Nakagawa and N. Miyata, *J. Med. Chem.*, 2012, **55**, 5760–5773.
- 166 N. Pradidphol, N. Kongkathip, P. Sittikul, N. Boonyalai and B. Kongkathip, *Eur. J. Med. Chem.*, 2012, **49**, 253–270.
- 167 S. Ke, L. Shi, Z. Zhang and Z. Yang, *Sci. Rep.*, 2017, **7**, 44439.
- 168 T. Xu, L. Shi, Y. Zhang, K. Wang, Z. Yang and S. Ke, *Eur. J. Med. Chem.*, 2019, **168**, 293–300.
- 169 C. P. Sanchez, A. Dave, W. D. Stein and M. Lanzer, *Int. J. Parasitol.*, 2010, **40**, 1109–1118.
- 170 M. Dhanawat, N. Das, R. C. Nagarwal and S. K. Shrivastava, *Mini-Rev. Med. Chem.*, 2009, **9**, 1447–1469.
- 171 N. V. Dharia, A. Chatterjee and E. A. Winzeler, *Mini-Rev. Med. Chem.*, 2010, **11**, 131–138.
- 172 K. Sepcic, *J. Toxicol., Toxin Rev.*, 2000, **19**, 139–160.
- 173 J. Rodriguez, *Stud. Nat. Prod. Chem.*, 2000, **24**, 573–681.
- 174 C. Timm and M. Kock, *Synthesis*, 2006, **113**, 2580–2584.
- 175 D. L. Gardiner, T. S. Skinner-Adams, C. L. Brown, K. T. Andrews, C. M. Stack, J. S. Carthy, J. P. Dalton and K. R. Trenhome, *Expert Rev. Anti-Infect. Ther.*, 2009, **7**, 1087–1098.
- 176 E. Fattorusso and O. Tagliatela-Scafati, *Mar. Drugs*, 2009, **7**, 130–152.
- 177 R. C. Larock and Y. Wang, *Tetrahedron Lett.*, 2002, **43**, 21–23.
- 178 Y. Wang, X. Y. Dong and R. C. Larock, *J. Org. Chem.*, 2003, **68**, 3090–3098.
- 179 C. A. Volk and M. Kock, *Org. Biomol. Chem.*, 2004, **2**, 1827–1830.
- 180 B. J. Shorey, V. Lee and J. E. Baldwin, *Tetrahedron*, 2007, **63**, 5587–5592.
- 181 J. B. Lekana-Douki, J. B. Bongui, S. L. Liabagui, S. E. Edou, R. Zatra and U. Bisvigou, *J. Ethnopharmacol.*, 2011, **133**, 1103–1108.
- 182 F. F. Hilario, R. C. Paula, M. L. Silveira, G. H. Viana, R. B. Alves, J. R. Pereira, L. M. Silva, R. P. Freitas and F. P. Varotti, *Chem. Biol. Drug Des.*, 2011, **78**, 477–482.
- 183 I. NNkumama, W. P. Meara and F. H. Osier, *Trends Parasitol.*, 2017, **33**, 128–140.
- 184 J. N. Burrows, S. Duparc, W. E. Gutteridge, R. H. Huijsduijn, W. Kaszubska, F. Macintyre,



- S. Mazzuri, J. J. Möhrle and T. N. CWells, *Malar. J.*, 2017, **16**, 26.
- 185 T. S. Skinner-Adams, S. D. Sumanadasa, G. M. Fisher, R. A. Davis, D. L. Doolan and K. T. Andrews, *Drug Discovery Today*, 2016, **21**, 725–739.
- 186 K. C. Nicolaou, *Angew. Chem., Int. Ed.*, 2014, **53**, 9128–9140.
- 187 A. Rossati, O. Bargiacchi, V. Kroumova, M. Zaramella, A. Caputo and P. L. Garavelli, *Infez. Med.*, 2016, **24**, 93–104.
- 188 N. Tiberti, S. L. Latham, S. Bush, A. Cohen, R. O. Opoka, C. C. John, A. Juillard, G. E. Grau and V. Combes, *Sci. Rep.*, 2016, **6**, 37871.
- 189 V. C. Corey, A. K. Lukens, E. S. Istvan, M. C. Lee, V. Franco, P. Magistrado, O. Coburn-Flynn, T. Sakata-Kato, O. Fuchs and E. A. Winzeler, *Nature Communications*, 2016, **7**, 11901.
- 190 R. M. Ribeiro-Viana, A. P. Butera, E. S. Santos, C. A. Tischer, R. B. Alves, R. P. Freitas, L. Guimaraes, F. P. Varotti, G. H. Viana and C. S. Nascimento, *J. Chem. Inf. Model.*, 2016, **56**, 571–579.
- 191 T. Liang, C. N. Neumann and T. Ritter, *Angew. Chem., Int. Ed.*, 2013, **52**, 8214–8264.
- 192 A. R. Collins, *Mol. Biotechnol.*, 2004, **26**, 249–261.
- 193 M. Fenech, *Nat. Protoc.*, 2007, **2**, 1084–1104.
- 194 C. C. Gomes, L. M. Moreira, V. J. Santos, A. S. Ramos, J. P. Lyon, C. P. Soares and F. V. Santos, *Genet. Mol. Biol.*, 2011, **34**, 116–121.
- 195 F. V. Santos, M. Andreo, A. L. Nasser, L. M. Moreira, W. Vilegas, I. M. Cylus and E. A. Varanda, *Arch. Biol. Sci.*, 2013, **65**, 191–195.
- 196 K. Katsuno, J. N. Burrows, K. Duncan, R. H. Huijsduijnen, T. Kaneko, K. Kita, C. E. Mowbray, D. Schmatz, P. Warner and B. T. Slingsby, *Nat. Rev. Drug Discovery*, 2015, **14**, 751–758.
- 197 A. Carbone, B. Parrino, M. G. Cusimano, V. Spanò, A. Montalbano, P. Barraja, D. Schillaci, G. Cirrincione, P. Diana and S. Cascioferro, *Mar. Drugs*, 2018, **16**, 274.
- 198 R. P. Araldi, T. C. Melo, T. B. Mendes, P. L. SáJunior, B. H. Nozima, E. Tito, R. F. Carvalho, E. B. Souza and R. C. Stocco, *Biomed. Pharmacother.*, 2015, **72**, 74–82.
- 199 C. S. Barbosa, D. S. Guimaraes, A. M. Goncalves, M. C. Barbosa, M. L. Costa, C. S. Nascimento, L. Guimaraes, R. M. Ribeiro-Viana, F. V. Santos, C. F. Brito, F. P. Varotti and G. H. Viana, *ACS Omega*, 2017, **2**, 8264–8272.
- 200 P. J. Durai, V. Padma, P. Vijaya, P. N. Sylaja and J. M. Murthy, *Stroke and thrombolysis in developing countries*, *Int. J. Stroke*, 2007, **2**, 17–26.
- 201 J. Wu, J. Ling, X. Wang, T. Li, J. Liu, Y. Lai, H. Ji, S. Peng, J. Tian and Y. Zhang, *J. Med. Chem.*, 2012, **55**, 7173.
- 202 G. Bartosz, *Biochem. Pharmacol.*, 2009, **77**, 1303–1315.
- 203 P. Talalay, A. T. Dinkova-Kostova and W. D. Holtzclaw, *Adv. Enzyme Regul.*, 2003, **43**, 121–134.
- 204 R. Sane, S. Agarwal and W. F. Elmquist, *Drug Metab. Dispos.*, 2012, **40**, 1612.
- 205 P. Paumard, J. Vaillier, B. Coulary, J. Schaeffer, V. Soubannier, D. M. Mueller, D. Brethes, R. J. Di and J. Velours, *EMBO J.*, 2002, **21**, 221–230.
- 206 E. R. Stadtman, Role of oxidant species in aging, *Curr. Med. Chem.*, 2004, **11**, 1105–1112.
- 207 H. Chen, H. Yoshioka, G. S. Kim, J. E. Jung, N. Okami, H. Sakata, C. M. Maier, P. Narasimhan, C. E. Goeders and P. H. Chan, *Antioxid. Redox Signaling*, 2011, **14**, 1505–1517.
- 208 H. Liu, C. J. Li, J. Z. Yang, N. Na, Y. K. Si, L. Li, N. H. Chen and D. M. Zhang, *J. Nat. Prod.*, 2012, **75**, 677–682.
- 209 R. A. Rane, S. D. Gutte and N. U. Sahu, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 6429–6432.
- 210 J. M. Pawlotsky, *Gastroenterology*, 2014, **146**, 1176–1192.
- 211 M. B. Zeisel, E. Crouchet, T. F. Baumert and C. Schuster, *Viruses*, 2015, **7**, 5659–5685.
- 212 K. E. Lillsunde, T. Tomasic, D. Kikelj and P. Tammela, *Chem. Biol. Drug Des.*, 2017, **90**, 1147–1154.
- 213 J. I. MacPherson, B. Sidders, S. Wieland, J. Zhong, P. Targett-Adams, V. Lohmann, P. Backes, O. Delpuech-Adams, F. Chisari, M. Lewis, T. Parkinson and D. L. Robertson, *PLoS One*, 2011, **6**, 25584.
- 214 P. T. Dolan, C. Zhang, S. Khadka, V. Arumugaswami, A. D. Vangeloff, N. S. Heaton, S. Sahasrabudhe, G. Randall, R. Sun and D. J. LaCount, *Mol. Biosyst.*, 2019, **3**, 3199–3209.
- 215 M. Taipale, D. F. Jarosz and S. Lindquist, *Nat. Rev. Mol. Cell Biol.*, 2010, **11**, 515–528.
- 216 R. Geller, S. Taguwa and J. Frydman, *Biochim. Biophys. Acta*, 2012, **1823**, 698–706.
- 217 M. A. Germain, L. Chatel-Chaix, B. Gagn, P. Bonneil, P. Thibault, F. Pradezynski, B. Chassey, L. Meyniel-Schicklin, V. Lotteau, M. Baril and D. Lamarre, *Mol. Cell. Proteomics*, 2014, **13**, 184–203.
- 218 K. E. Lillsunde, T. Tomašič, P. Schult, V. Lohmann, D. Kikelj and P. Tammela, *ChemMedChem*, 2019, **14**, 334–342.
- 219 J. R. Chérouvrier, F. Carreaux and J. P. Bazureau, *Tet. Lett.*, 2002, **43**, 3581–3584.
- 220 M. Rahaus, N. Desloges and M. H. Wolff, *Cellular Signalling*, 2007, **19**, 312–320.
- 221 B. W. Doble, S. Patel, G. A. Wood, L. K. Kockeritz and J. R. Woodgett, *Deve. Cell.*, 2007, **12**, 957–971.
- 222 Q. Wang, Y. Zhou and B. M. Evers, *Neoplasia*, 2006, **8**, 781–787.
- 223 S. Leclerc, M. Garnier, R. Hoessel, D. Marko, J. A. Bibb, G. L. Snyder, P. Greengard, J. Biernat, E.-M. Mandelkow, G. Eisenbrand and L. Meijer, *J. Biol. Chem.*, 2001, **276**, 251–260.
- 224 A. Primot, B. Baratte, M. Gompel, A. Borgne, S. Liabeuf, J. L. Romette, F. Costantini and L. Meijer, *Protein Expression Purif.*, 2000, **20**, 394–404.
- 225 M. Debda, S. Renault, O. Lozach, L. Meijer, L. Paquin, F. Carreaux and J. P. Bazureau, *Eur. J. Med. Chem.*, 2010, **45**, 805–810.

