



Cite this: *RSC Adv.*, 2020, 10, 30723

The effect of saccharides on equilibrium swelling of thermo-responsive gels†

A. D. Drozdov * and J. deClaville Christiansen

Mechanical and optical properties of thermo-responsive (TR) gels change drastically at their volume phase transition temperature. As the critical temperature is strongly affected by the presence of small amounts of additives in aqueous solutions, TR gels can be employed as sensors for detection and recognition of multiple analytes (from specific ions to hazardous biochemicals to pathogenic proteins) and actuators for biomedical applications. A simplified mean-field model is developed for equilibrium swelling of TR gels in aqueous solutions of additives. Its advantage is that the model involves a relatively small (compared with the conventional approaches) number of material constants and accounts for changes in the thermo-mechanical response at transition from the swollen to collapsed state. The ability of the model to describe experimental swelling diagrams and to predict the influence of additives on the equilibrium degree of swelling and the volume phase transition temperature of TR gels is confirmed by comparison of observations on poly(*N*-isopropylacrylamide) gel in aqueous solutions of saccharides (glucose, sucrose and galactose) with results of numerical analysis.

Received 4th July 2020
Accepted 10th August 2020

DOI: 10.1039/d0ra05845a

rsc.li/rsc-advances

1 Introduction

Hydrogels are three-dimensional networks of polymer chains bridged by covalent and/or physical junctions. Thermo-responsive (TR) gels form a special group of hydrogels whose equilibrium degree of swelling, as well as mechanical and optical properties are strongly affected by temperature.¹ Equilibrium and transient swelling of TR gels in various solvents and their mixtures has recently attracted substantial attention due to a wide area of potential applications of these materials in technology² and medicine.³

Two types of TR hydrogels are conventionally distinguished. Gels of the LCST (low critical solution temperature)-type swell noticeably in water at temperatures T below their volume phase transition temperature T_c and shrink (collapse) at temperatures above T_c .⁴ Gels of the UCST (upper critical solution temperature)-type immersed into water shrink at temperatures below T_c and swell at $T > T_c$.⁵ Pronounced changes in degree of swelling Q at the critical temperature T_c are accompanied by morphological transformations of TR gels (a homogeneous micro-structure of a swollen gel becomes strongly inhomogeneous in the collapsed state due to phase separation into polymer-poor and polymer-rich domains).⁶

Temperature is not the only trigger causing abrupt changes in degree of swelling of TR hydrogels. Similar changes are

observed when relatively small amounts of additives (starch,⁷ saccharides,⁸ phenols,⁹ inorganic salts,¹⁰ surfactants,¹¹ oligomers¹²) are dissolved in water. The ability of TR gels to recognize host molecules make them suitable for applications as chemical and biochemical sensors.¹³ Development of hydrogel-based sensors for analytical, chemical and biomedical applications has recently become a focus of attention.^{14–17} Design of high-speed sensors with robust responsiveness to additives and optimization of their properties require adequate models for the thermo-mechanical behavior of TR gels in aqueous solutions.

Conventional models for swelling of TR gels in water are grounded on the Flory–Rehner concept¹⁸ that presumes the specific Helmholtz free energy of a gel Ψ to consist of three parts: (i) the specific free energy of water not interacting with the polymer network Ψ_1 , (ii) the specific strain energy of the network not interacting with water Ψ_2 , and (iii) the specific energy of interaction between water molecules and segments of chains Ψ_{int} . In the Flory–Rehner theory, the function Ψ_{int} is adopted in the form

$$\Psi_{\text{int}} = k_B T_0 (C_f \ln \phi_f + \chi C_f \phi_n), \quad (1)$$

where k_B is the Boltzmann constant, C_f stands for concentration of fluid phase (number of water molecules per unit volume in the initial state of a gel), ϕ_f and ϕ_n are volume fractions of fluid (water) and solid (polymer network) phases in the actual state, and χ is the Flory–Huggins (FH) parameter. The first term in eqn (1) characterizes the entropy and the other term describes the enthalpy of mixing of water molecules and segments of

Department of Materials and Production, Aalborg University, Fibigerstraede 16, Aalborg 9220, Denmark. E-mail: aleksey@m-tech.aau.dk

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra05845a



chains. As the analysis focuses on a narrow interval of temperatures near T_c , the actual temperature T is replaced with a fixed temperature T_0 in the thermodynamic factor $k_B T$.

When the FH parameter χ is treated as a function of temperature T only, this approach results in rather poor agreement with experimental swelling diagrams.¹⁹ To improve the quality of fitting, this coefficient is traditionally replaced with an effective FH parameter χ_{eff} , which is presumed to depend on two arguments, T and ϕ_n . Adopting an analytical expression for χ_{eff} (as a polynomial²⁰ or rational²¹ function of ϕ_n with temperature-dependent coefficients), an acceptable agreement can be reached between experimental data and results of simulation.

Two shortcomings of this approach are to be mentioned: (i) observations^{22,23} show that the volume phase transition temperature T_c is a characteristic parameter for a TR gel (this temperature is practically unaffected by preparation conditions and volume fractions of monomers and cross-linker in a pre-gel solution). However, the model (1) does not involve T_c as a parameter and does not permit it to be determined in simulation. Another approach to the description of swelling of TR gels was suggested in ref. 24 and 25. It is based on the Landau theory of phase transition and allow T_c to be found by fitting equilibrium swelling diagrams. (ii) Experimental data^{26,27} demonstrate that the elastic moduli of TR gels increase strongly (by an order of magnitude) at temperatures above T_c . This growth (driven by aggregation of hydrophobic segments into clusters that serve as physical junctions between chains in the network²³) is disregarded in the models based on eqn (1). An explicit account for the evolution of shear moduli of TR gels with temperature was performed in ref. 28 and 29.

To study equilibrium swelling of a TR gel in a mixture of two solvents (solvent-1 and solvent-2), the Flory–Rehner concept is generalized by including all terms describing interactions between segments of chains and solvent-1 and solvent-2 molecules into eqn (1). The following expression is adopted for the specific free energy $\Psi_{\text{int}}^{30,31}$

$$\Psi_{\text{int}} = k_B T_0 [(C_1 \ln \phi_1 + C_2 \ln \phi_2) + (\chi_{13} C_1 \phi_n + \chi_{23} C_2 \phi_n + \chi_{12} C_1 \phi_2)], \quad (2)$$

where C_1 , C_2 denote concentrations of cosolvents, ϕ_1 , ϕ_2 are their volume fractions in the gel, and χ_{13} , χ_{23} , χ_{12} stand for the FH parameters.

Observations in equilibrium swelling tests are reproduced poorly when the coefficients χ_{ij} are treated as functions of temperature T only.³² To ameliorate the accuracy of fitting, three approaches were suggested: (i) introduction of prescribed dependencies of the coefficients χ_{ij} on volume fractions of constituents,³³ (ii) extension of eqn (2) to account for size and shape of solvent molecules and segments of chains, as well as for attractive and repulsive interactions between them,³⁴ and (iii) inclusion of higher-order terms with respect to ϕ_1 , ϕ_2 , ϕ_n into eqn (2).³⁵ Although these refinements improve the quality of matching experimental data, they lead to a strong increase in the number of adjustable parameters, which makes

questionable the ability of the models to predict the thermo-mechanical behavior of TR gels in mixtures of solvents.

The objective of this study is threefold: (i) to develop a model with a reasonably small number of material constants for equilibrium swelling of TR gels in aqueous solutions of additives that accounts for changes in the micro-structure of the polymer network and its elastic moduli at transition from the swollen to collapsed state and allows the critical temperature T_c to be determined explicitly, (ii) to find adjustable parameters in the governing equations by matching equilibrium swelling diagrams on covalently cross-linked poly(*N*-isopropylacrylamide) (PNIPA) gel in water and aqueous solutions of glucose, galactose and sucrose at various temperatures and molar fractions of saccharides, (iii) to verify the ability of the model to predict observations by comparing results of simulation with experimental data in independent tests.

We focus on equilibrium swelling of LCST-type covalently cross-linked thermo-responsive gels in aqueous solutions with relatively low volume fractions of additives (below the concentration at which reentrant swelling starts³⁶). Analysis of the physical mechanisms for the co-solvency and co-nonsolvency phenomena in TR gels in mixed solvents³⁷ is beyond the scope of this work. Unlike previous studies, we do not confine ourselves to the response of TR gels at a fixed temperature. Temperature T and volume fraction of additive in the bath ϕ_2^{bath} are treated as independent parameters. This allows the critical temperature $T_c(\phi_2^{\text{bath}})$ and the critical concentration of additive $\phi_{2c}^{\text{bath}}(T)$ to be predicted at which the volume phase transition occurs.

A strong decay in the equilibrium degree of swelling at the critical temperature T_c for a TR gel in pure water is conventionally explained by pronounced changes in the effective hydrophilicity of chains involving hydrophilic and hydrophobic segments.^{38,39} At temperatures below T_c , each hydrophobic segment is surrounded by a cage-like structure formed by water molecules bridged by hydrogen bonds.⁴⁰ When temperature grows, clustered water molecules are destabilized by thermal fluctuations. Breakage of cages formed by water molecules induces agglomeration of hydrophobic segments and formation of hydrophobic aggregates from which water molecules are expelled. At temperatures above T_c , most cages are broken, and the structure of a TR gel becomes inhomogeneous: it consists of a number of deswollen hydrophobic aggregates bridged by hydrophilic segments and separated by nano-channels in which water molecules are located.⁴¹

The similarity between the equilibrium swelling diagrams on TR gels in pure water, $Q(T)$, and in aqueous solution of additives at a fixed temperature, $Q(\phi^{\text{bath}})$, see Fig. 1 below, can be explained by the same mechanism of destabilization of cage-like structures surrounding hydrophobic segments. In swelling tests with increasing temperature, breakage of cages is induced by thermal fluctuations, whereas in experiments with growing concentration of additives, it is caused by complexation of additives with water molecules (formation of molecular complexes linked by hydrogen bonds).⁴² When water molecules leave the cage-like structures around hydrophobic segments to form complexes with molecules of additives, these structures



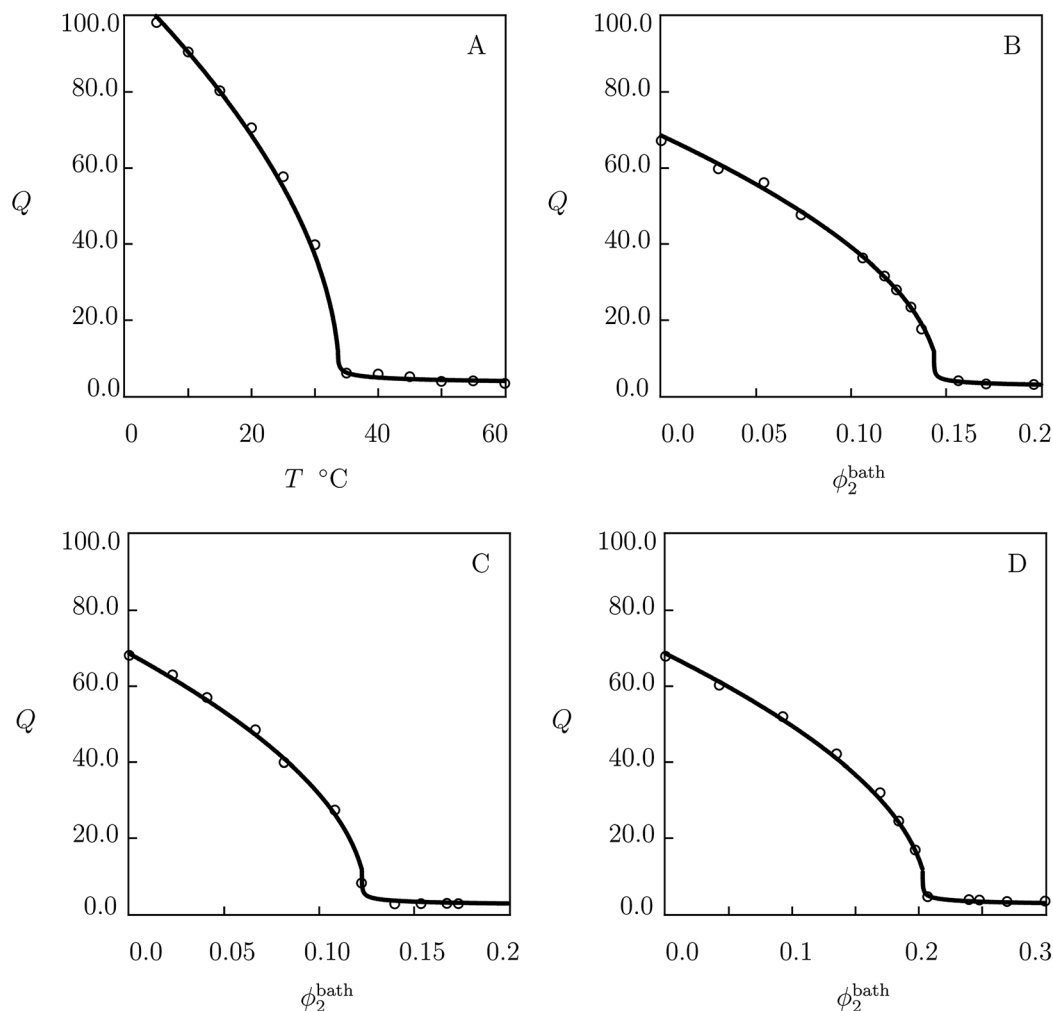


Fig. 1 (A) Degree of swelling Q versus temperature T . (B–D) Degree of swelling Q versus volume fraction of saccharides in the bath ϕ_2^{bath} . Circles: experimental data^a on PNIPA gel in pure water (A) and in aqueous solutions of glucose (B), galactose (C) and sucrose (D) at temperature $T = 20$ °C. Solid lines: results of simulation.

are destroyed. As a result, the “naked” segments aggregate into hydrophobic clusters, the polymer chains dehydrate, and the network collapses.^{43–45}

The novelty of our study consists in the following: (i) a model is developed for the mechanical response and equilibrium swelling of TR gels in aqueous solutions of additives. Unlike the conventional approach, we do not introduce phenomenological dependencies of the FH parameters χ_{ij} on volume fractions of polymer chains or cosolvent molecules. A pronounced decay in degree of swelling when temperatures and/or concentration of additives exceed their critical values is described by taking into account an increase in the elastic modulus of a gel caused by aggregation of hydrophobic segments. (ii) An advantage of the model is that it involves a small number of material constants with transparent physical meaning. In particular, to describe equilibrium swelling of a TR gel in an aqueous solution of an additive, only one extra coefficient is introduced in the governing equations. This allows the model to be applied not only for the description of experimental data, but also for prediction of equilibrium swelling diagrams on TR gels (see Fig. 3 and 4

below). (iii) Adjustable parameters are determined by fitting observations on PNIPA gel in aqueous solutions of saccharides. Application of TR gels with boronic acid derivatives for continuous glucose monitoring has recently attracted noticeable attention.^{46,47} The proposed approach can be helpful for the development of insulin-regulatory systems as interactions between glucose molecules and segments of PNIPA chains are conventionally disregarded in their analysis.

The exposition is organized as follows. Equilibrium swelling of TR gels in pure water is discussed in Section 2. The model is extended in Section 3 for the analysis of swelling in solutions of additives. The governing equations are verified in Section 4, where results of simulation are compared with experimental data. Concluding remarks are formulated in Section 5. A detailed derivation of the governing equations is provided in ESI.†

2 Swelling of TR gels in pure water

A TR gel is modeled as a two-phase medium composed of solid (an equivalent polymer network) and fluid (water) constituents.



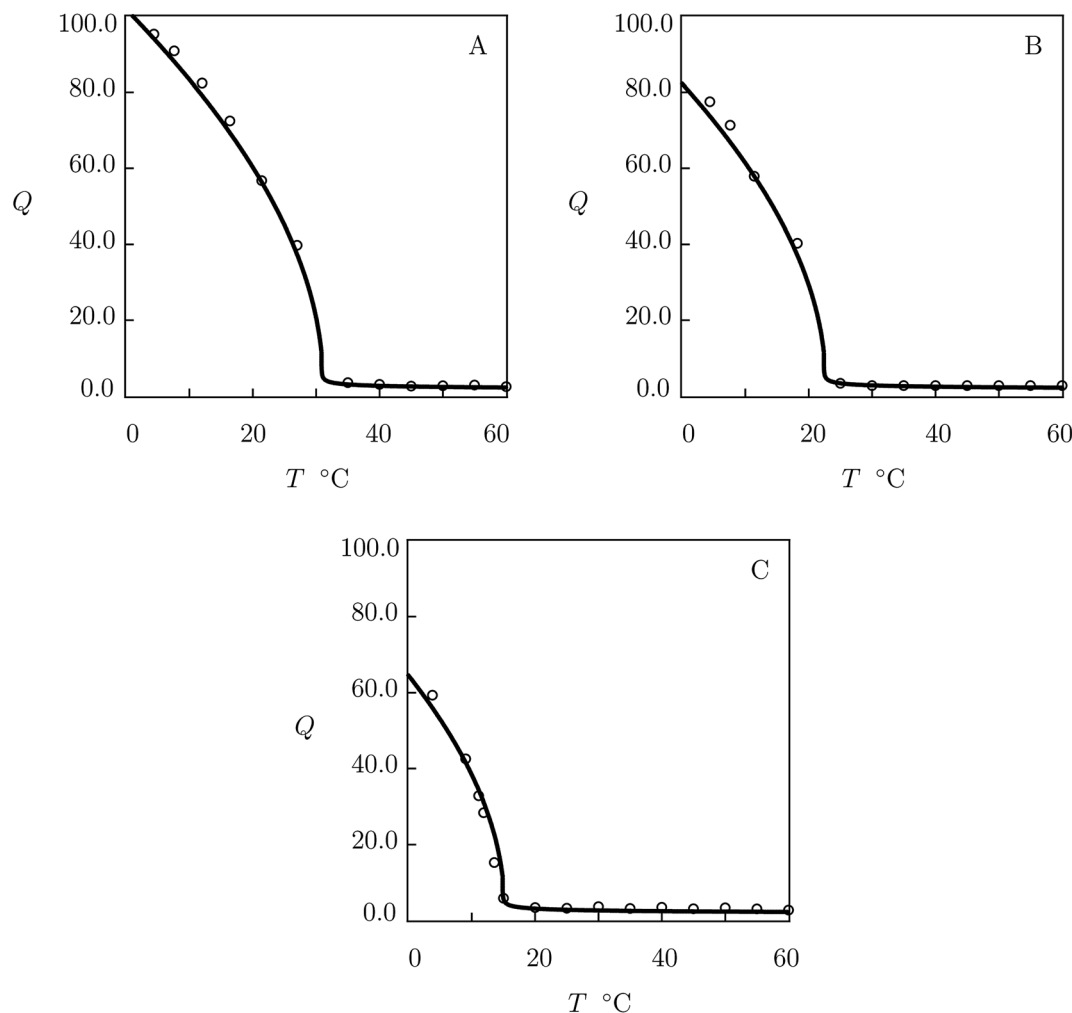


Fig. 2 Degree of swelling Q versus temperature T . Circles: experimental data⁸ on PNIPAA gel in aqueous solutions of glucose with volume fractions $\phi_2^{\text{bath}} = 0.031$ (A), 0.122 (B) and 0.188 (C). Solid lines: predictions of the model.

The solid and fluid phases are treated as immiscible (mass exchange between the phases is disregarded) interpenetrating (any elementary volume contains both phases) continua.

The initial configuration of a gel coincides with that of an undeformed dry specimen at some temperature $T_0 < T_c$. Adopting the affinity hypothesis, we suppose that macro-deformation of a gel coincides with deformation of its polymer network. Transformation of the initial configuration into the actual configuration at an arbitrary temperature T is described by the deformation gradient \mathbf{F} . Disregarding thermally induced volume expansion, we write the molecular incompressibility condition in the form

$$\det \mathbf{F} = 1 + C_f \nu, \quad (3)$$

where C_f denotes concentration of water molecules (number of molecules per unit volume in the initial state), and ν stands for the characteristic volume of a molecule. Eqn (3) means that volume deformation of a gel is driven by changes in water content only.

The polymer network involves two components. The first network with covalent bonds is built under cross-linking polymerization of a pre-gel solution. The other network with physical junctions is formed at temperatures $T > T_c$ due to aggregation of hydrophobic segments belonging to different chains. Disregarding the viscoelastic phenomena, we treat both networks as permanent.

For the covalently cross-linked network, transformation of the initial state into the reference (stress-free) state is described by the deformation gradient \mathbf{f}_1 . Presuming the network to be isotropic, we set

$$\mathbf{f}_1 = f_1^{\frac{1}{3}} \mathbf{I}, f_1 = 1 + Q_0, \quad (4)$$

where \mathbf{I} is the unit tensor, $Q_0 = C_{f0} \nu$, and C_{f0} stands for concentration of water in the reference state.

Keeping in mind that all water molecules are expelled from hydrophobic aggregates, we suppose that the reference state of the network with physical bonds coincides with the initial (dry) state of the gel.



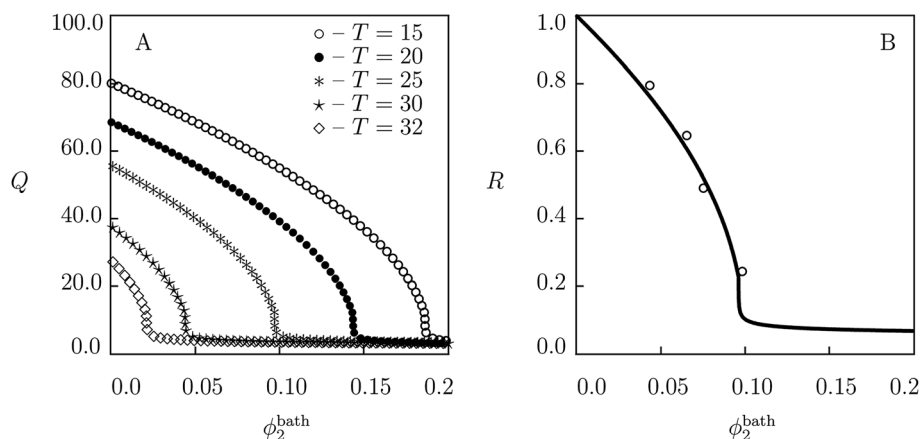


Fig. 3 (A) Degree of swelling Q versus volume fraction of glucose in the bath ϕ_2^{bath} . Symbols: predictions of the model for PNIPA gel in aqueous solution of glucose at various temperatures T °C. (B) The ratio of masses $R = M(\phi_2^{\text{bath}})/M(0)$ of a gel sample versus volume fraction of glucose in the bath ϕ_2^{bath} . Solid line: predictions of the model for PNIPA gel in aqueous solution of glucose at temperature $T = 25$ °C. Circles: experimental data.⁶⁴

Applying the multiplicative decomposition formula, we find the deformation gradients for elastic deformation of the networks

$$\mathbf{F}_e^{(1)} = f_1^{-\frac{1}{3}} \mathbf{F}, \mathbf{F}_e^{(2)} = \mathbf{F}$$

and the corresponding Cauchy–Green tensors for elastic deformation

$$\mathbf{B}_e^{(1)} = f_1^{-\frac{2}{3}} \mathbf{B}, \mathbf{B}_e^{(2)} = \mathbf{B}. \quad (5)$$

Here $\mathbf{B} = \mathbf{F} \cdot \mathbf{F}^T$ is the Cauchy–Green tensor for macro-deformation, the dot denotes inner product, and T stands for transpose.

The Helmholtz free energy (per unit volume in the initial state) Ψ equals the sum of the specific energies of fluid and solid constituents not interacting with each other and the energy of their interaction,

$$\Psi = \Psi_1 + \Psi_2 + \Psi_{\text{int}}, \quad (6)$$

where Ψ_1 is the specific energy of water molecules not interacting with segments of chains, Ψ_2 is the specific energy of polymer chains not interacting with water, and Ψ_{int} is the specific energy of mixing of water molecules with segments of chains.

The specific energy of water reads

$$\Psi_1 = \mu^0 C_f, \quad (7)$$

where μ^0 denotes chemical potential of water molecules not interacting with the polymer network.

The specific energy of the polymer network (consisting of two parts with chemical and physical bonds) is given by

$$\Psi_2 = \sum_{m=1}^2 W_m \left(I_{e1}^{(m)}, I_{e2}^{(m)}, I_{e3}^{(m)} \right), \quad (8)$$

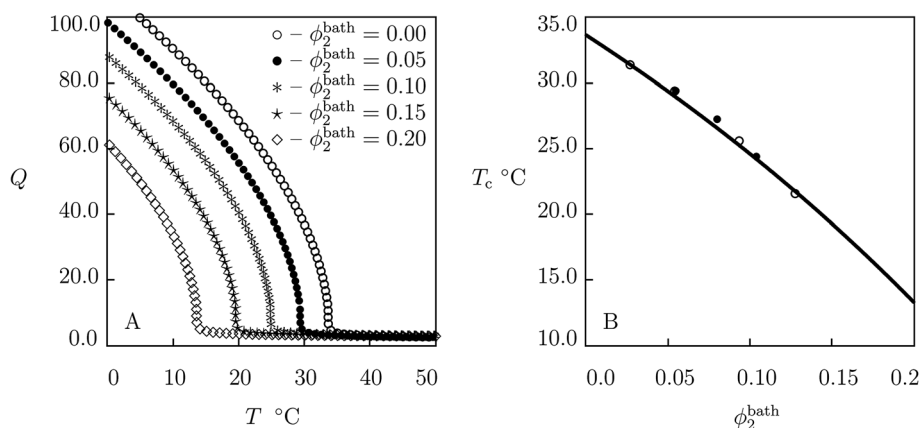


Fig. 4 (A) Degree of swelling Q versus temperature T . Symbols: predictions of the model for PNIPA gel in aqueous solutions of glucose with various volume fractions ϕ_2^{bath} . (B) Volume phase transition temperature T_c versus volume fraction of glucose in the bath ϕ_2^{bath} . Solid line: predictions of the model. Symbols: experimental data (○ – ref. 65, ● – ref. 66).



where the specific mechanical energy W_m stored in chains of the m th network depends on principal invariants $I_{e1}^{(m)}, I_{e2}^{(m)}, I_{e3}^{(m)}$ of the corresponding Cauchy–Green tensor for elastic deformation $\mathbf{B}_e^{(m)}$. For definiteness, the neo-Hookean formulas are accepted for the strain energy densities of polymer networks with covalent and physical bonds,

$$W_m = \frac{1}{2}G_m \left[\left(I_{e1}^{(m)} - 3 \right) - \ln I_{e3}^{(m)} \right], (m = 1, 2), \quad (9)$$

where G_1 and G_2 stand for the shear moduli. More sophisticated expressions for the functions W_m were developed and verified by comparison with observations in ref. 48 and 49.

Confining ourselves to the analysis of swelling in a rather narrow interval of temperatures near the volume phase transition temperature T_c , we treat the modulus G_1 of the covalently cross-linked network as a constant. The modulus G_2 of the network with physical bonds vanishes below T_c and grows with the difference $T - T_c$ above the volume phase transition temperature. The following relation is adopted for the function $G_2(T)$:

$$G_2(T) = 0, \\ (T < T_c), G_2(T) = \bar{G}_2 \left[1 - \exp \left(-\beta_2 \sqrt{T - T_c} \right) \right], (T \geq T_c), \quad (10)$$

where \bar{G}_2 and β_2 are adjustable parameters. The ability of eqn (10) to describe observations is confirmed in Fig. S-1,† where experimental data⁵⁰ are reported on the temperature-induced increase in the Young's modulus $E = \frac{3}{2}(G_1 + G_2)$ of PNIPA gel in water and NaCl solution together with their fit by eqn (10) with the material parameters listed in Table S-1.†

Introducing an order parameter

$$\eta = |T - T_c|^{\frac{1}{2}} \quad (11)$$

for the volume phase transition in a TR gel, we present eqn (10) in the form

$$\frac{dG_2}{d\eta} = \beta_2(\bar{G}_2 - G_2), G_2(0) = 0. \quad (12)$$

Bearing in mind that G_2 is proportional to concentration of physical bonds (aggregates from which molecules of water are expelled) that bridge polymer chains in the network above T_c , eqn (12) can be treated as a simple kinetic relation for aggregation of hydrophobic segments. More sophisticated relations for the kinetics of aggregation were discussed in ref. 51 and 52.

The specific energy of mixing of water molecules with segments of chains is adopted in the Flory–Huggins form (1), where ϕ_f and ϕ_n are given by

$$\phi_f = \frac{C_f v}{1 + C_f v}, \phi_n = \frac{1}{1 + C_f v}. \quad (13)$$

The following expression is accepted for the FH parameter χ :

$$\chi = \chi_0 + \chi_1 T (T < T_c), \quad \chi = \chi_{\max} (T > T_c), \quad (14)$$

where T and T_c are measured in °C, and the coefficients χ_0 , χ_1 , χ_{\max} obey the continuity condition

$$\chi_{\max} = \chi_0 + \chi_1 T_c. \quad (15)$$

Eqn (14) means that breakage of cage-like structures formed by water molecules surrounding hydrophobic segments leads to an increase in the effective hydrophobicity of chains (characterized by the parameter χ) at temperatures T below T_c . When T exceeds the volume phase transition temperature T_c , further growth of χ is prohibited by formation of aggregates of hydrophobic segments.

Applying the free energy imbalance inequality, we find that under equilibrium swelling of a TR gel, its degree of swelling $Q = C_f v$ obeys the nonlinear equation

$$\ln \frac{Q}{1+Q} + \frac{1}{1+Q} + \frac{\chi}{(1+Q)^2} + \frac{g_1}{1+Q} \left[\left(\frac{1+Q}{1+Q_0} \right)^{\frac{2}{3}} - 1 \right] \\ + \frac{g_2}{1+Q} \left[(1+Q)^{\frac{2}{3}} - 1 \right] \\ = 0, \quad (16)$$

where

$$g_m = \frac{G_m v}{k_B T_0} (m = 1, 2)$$

stand for the dimensionless shear moduli. Derivation of eqn (16) is given in Appendix A (ESI).†

Eqn (16) together with eqn (14) for the Flory–Huggins parameter χ and eqn (10) for the shear modulus G_2 provides governing relations for the equilibrium degree of swelling Q at an arbitrary temperature T . These equations involve seven adjustable parameters: (i) g_1 stands for the dimensionless shear modulus of the polymer network with covalent bonds, (ii) Q_0 denotes degree of swelling of a gel in its stress-free state, (iii) χ_0 , χ_1 characterize temperature-induced changes in the FH parameter χ below the critical temperature T_c , (iv) χ_{\max} is the FH parameter above the volume phase transition temperature, and (v) \bar{G}_2 , β_2 describe the kinetics of aggregation of hydrophobic segments of chains above the critical temperature. Given material constants, the critical temperature T_c is found from eqn (15).

To confirm the ability of the model to describe equilibrium swelling diagrams, we fit experimental data⁵³ on PNIPA gel prepared by irradiation cross-linking of an aqueous solution of NIPA chains with the irradiation dose $I = 20$ kGy. Observations in equilibrium water uptake tests are depicted in Fig. S-2A.† The data are matched by means of a two-step procedure. At the first step, observations below T_c (where $g_2 = 0$) are fitted. We set $Q_0 = 12.81$ (which means that the reference state coincides with the as-prepared state⁵⁴), determine χ from eqn (16), and approximate the experimental dependence $\chi(T)$ by means of eqn (14), where χ_0 and χ_1 are calculated by the least-squares technique. The dimensionless elastic modulus g_1 is found from the condition of the best-fit of the dependence $\chi(T)$. The coefficient χ_{\max} is determined from eqn (15). At the other step, the data



above T_c are matched by means of two parameters, β_2 and \bar{g}_2 , with $\bar{g}_2 = \bar{G}_2 v / (k_B T_0)$.

Fig. S-2A† demonstrates good agreement between the observations and results of simulation with the material constants listed in Table S-2.† Fig. S-2B† shows that eqn (14) describes correctly an increase in χ with temperature below the critical temperature T_c . An advantage of the governing eqn (10), (14) and (16) compared with previous models for swelling of TR gels (see, for example,^{55,56} where similar observations are approximated) is that it correctly describes equilibrium swelling diagrams by means of a rather limited set of adjustable parameters.

3 Swelling of TR gels in solutions of additives

Our aim now is to describe how degree of swelling of TR gels is affected by the presence of additives in aqueous solutions.

Experimental data show that volume phase transition in a TR gel immersed in water occurs at a critical temperature T_c which is practically independent of preparation conditions and concentrations of monomers and cross-linkers.²² This observation serves as a basis for treatment of T_c as a parameter characterizing the transition point, see eqn (12), and the function (9) (proportional to the difference $T - T_c$) as the order parameter η for volume phase transition.

Under swelling of a TR gel in a mixture of water with another solvent, the temperature T_c is strongly affected by the chemistry and concentration of the cosolvent. As T_c cannot be used as a criterion for the volume phase transition, three questions arise: (i) which parameter can replace T_c as a measure for coil-to-globule transition in polymer chains, (ii) how to express the order parameter η (that describes the kinetics of aggregation of hydrophobic segments) in terms of this quantity, and (iii) how to define χ in eqn (16) in order to account for the influence of cosolvents or additives. The main requirement for appropriate answers to these questions is that they should result in eqn (10), (14) and (16) when volume fraction of additives in the bath ϕ_2^{bath} vanishes.

We introduce the coefficient χ_{max} as a criterion for the volume phase transition in TR gels in binary mixtures. In other words, we suppose that some “equivalent” FH parameter χ_{eq} exists (treated as a function of temperature, volume fraction of additive in the gel, *etc.*) such that

$$\chi_{\text{eq}} < \chi_{\text{max}}$$

when the gel is in the swollen state, and

$$\chi_{\text{eq}} \geq \chi_{\text{max}}$$

when it is in the collapsed state.

It follows from eqn (14) that $\chi_{\text{eq}} = \chi_0 + \chi_1 T$ for a TR gel in pure water. This equality together with eqn (15) implies that when a TR gel collapses in pure water,

$$T - T_c = \frac{1}{\chi_1} (\chi_{\text{eq}} - \chi_{\text{max}}).$$

Combining this equality with eqn (11), we introduce the order parameter η by the formula

$$\eta = |\chi_{\text{eq}} - \chi_{\text{max}}|^{\frac{1}{2}}. \quad (17)$$

According to eqn (12), the shear modulus G_2 of the polymer network with physical bonds obeys the differential equation

$$\frac{dG_2}{d\eta} = \beta(\bar{G}_2 - G_2), G_2(0) = 0 \quad (18)$$

with $\beta = \beta_2 / \sqrt{\chi_1}$. The solution of eqn (18) reads

$$G_2 = 0, (\chi_{\text{eq}} < \chi_{\text{max}}), \\ G_2 = \bar{G}_2 [1 - \exp(-\beta \sqrt{\chi_{\text{eq}} - \chi_{\text{max}}})], (\chi_{\text{eq}} \geq \chi_{\text{max}}). \quad (19)$$

Eqn (19) means that G_2 vanishes when a gel is in the swollen state, and the modulus grows (due to aggregation of hydrophobic segments) with η when the gel is in the collapsed state.

To derive an expression for χ_{eq} of a TR gel in an aqueous solution, we transform eqn (2) for the specific energy of interaction between segments of chains and molecules of solvent-1 (water) and solvent-2 (additives). Bearing in mind that

$$\phi_n = \frac{1}{1 + (C_1 + C_2)v}, \phi_1 = \frac{C_1 v}{1 + (C_1 + C_2)v}, \phi_2 = \frac{C_2 v}{1 + (C_1 + C_2)v} \quad (20)$$

and setting

$$C_f = C_1 + C_2, \phi_1 = \frac{C_1}{C_f}, \phi_2 = \frac{C_2}{C_f}, \quad (21)$$

we find from eqn (2) that

$$\Psi_{\text{int}} = k_B T_0 C_f \left[\ln \frac{C_f v}{1 + C_f v} + \frac{\chi_{13} \phi_1 + \chi_{23} \phi_2 - \chi_{12} \phi_1 \phi_2}{1 + C_f v} \right. \\ \left. + (\phi_1 \ln \phi_1 + \phi_2 \ln \phi_2 + \chi_{12} \phi_1 \phi_2) \right]. \quad (22)$$

Measurements of concentration of additives in PNIPa gels immersed in aqueous solutions of saccharides^{7,8} reveal that the partitioning coefficient

$$P = \frac{\phi_2}{\phi_2^{\text{bath}}} \quad (23)$$

is very close to unity when these gels are in the swollen state. Setting $P = 1$ (the validity of this assumption will be examined in what follows), which means that

$$\phi_1 = \phi_1^{\text{bath}}, \phi_2 = \phi_2^{\text{bath}}, \quad (24)$$

we find from eqn (22) that

$$\Psi_{\text{int}} = k_B T_0 C_f \left[\ln \frac{C_f v}{1 + C_f v} + \frac{\chi^*}{1 + C_f v} + (\phi_1^{\text{bath}} \ln \phi_1^{\text{bath}} \right. \\ \left. + \phi_2^{\text{bath}} \ln \phi_2^{\text{bath}} + \chi_{12} \phi_1^{\text{bath}} \phi_2^{\text{bath}}) \right] \quad (25)$$

with



$$\chi^* = \chi_{13}\phi_1^{\text{bath}} + \chi_{23}\phi_2^{\text{bath}} - \chi_{12}\phi_1^{\text{bath}}\phi_2^{\text{bath}}. \quad (26)$$

When a TR gel is in the swollen state, and the specific energy of interaction between segments of chains and solvent-1 and solvent-2 molecules is given by eqn (25), its equilibrium degree of swelling Q obeys the equation

$$\ln \frac{Q}{1+Q} + \frac{1}{1+Q} + \frac{\chi^*}{(1+Q)^2} + \frac{g_1}{1+Q} \left[\left(\frac{1+Q}{1+Q_0} \right)^{\frac{2}{3}} - 1 \right] = 0. \quad (27)$$

Derivation of eqn (27) is provided in Appendix B (ESI).†

Comparison of eqn (16) and (27) implies that under condition (24), the equilibrium degree of swelling Q of a TR gel in an aqueous solution of additives obeys eqn (16), where χ is given by an extension of eqn (14),

$$\chi = \chi_{\text{eq}} (\chi_{\text{eq}} < \chi_{\text{max}}), \quad \chi = \chi_{\text{max}} (\chi_{\text{eq}} > \chi_{\text{max}}), \quad (28)$$

and χ_{eq} is determined by eqn (26). Eqn (26) was first proposed for the analysis of dilute solutions of copolymers in ref. 57. It was applied to the analysis of equilibrium swelling of covalently cross-linked poly(propylene glycol) gels in organic solvents in ref. 58.

It follows from eqn (26) that

$$\chi_{\text{eq}} = \chi_{13}(1 - \phi_2^{\text{bath}}) + \chi_{23}\phi_2^{\text{bath}} - \chi_{12}\phi_2^{\text{bath}}(1 - \phi_2^{\text{bath}}).$$

At small volume fractions of additives in an aqueous solution, $\phi_2^{\text{bath}} \ll 1$, this relation is simplified

$$\chi_{\text{eq}} = \chi_{13}(1 - \phi_2^{\text{bath}}) + (\chi_{23} - \chi_{12})\phi_2^{\text{bath}}. \quad (29)$$

The coefficient χ_{13} in eqn (29) describes interactions between water molecules and segments of chains. This quantity coincides with the FH parameter χ in eqn (1), and its dependence on temperature T is given by eqn (14).

The coefficient χ_{12} accounts for interactions between water molecules and molecules of an additive. Although this parameter depends slightly on temperature,⁵⁹ we disregard this dependence, as an appropriate activation energy is lower (by at least an order of magnitude) than the enthalpy characterizing the effect of T on χ_{13} (see Fig. S-3,† where experimental data⁶⁰ on aqueous solutions of glucose and sucrose are presented).

Observations on equilibrium swelling of PNIPA gels in water/ethanol and water/methanol mixtures^{20,61,62} show that at high concentrations of cosolvents, Q becomes independent of temperature, which implies that χ_{23} is temperature-independent for these additives. This conclusion is explained in ref. 63 by the fact that intensity of thermal fluctuations in the interval of temperatures under investigation (between 0 °C and T_c) is insufficient to activate oscillations of heavy alcohol molecules (their molecular mass exceeds that of water by several times) and to induce breakage of cage-like structures formed by these molecules around hydrophobic segments of chains. The same argument can be applied to the saccharides under consideration as their molecular mass exceeds mass of water molecules by an order of magnitude.

Based on this reasoning, we disregard the effect of temperature on χ_{12} and χ_{23} , and find from eqn (14) and (28) that

$$\chi_{\text{eq}} = (\chi_0 + \chi_1 T)(1 - \phi_2^{\text{bath}}) + K\phi_2^{\text{bath}}, \quad (30)$$

where

$$K = \chi_{23} - \chi_{12}. \quad (31)$$

It follows from eqn (30) that K is the only extra constant required to describe equilibrium swelling of a TR gel in an aqueous solution of additives. The other material constants are determined by fitting observations in water uptake tests at various temperatures.

Combination of eqn (28) and (30) results in the formula for the volume phase transition temperature T_c of a TR gel in an aqueous solution

$$T_c = \frac{1}{\chi_1} \left(\frac{\chi_{\text{max}} - K\phi_2^{\text{bath}}}{1 - \phi_2^{\text{bath}}} - \chi_0 \right). \quad (32)$$

It follows from eqn (32) that T_c increases with volume fraction of additives in the bath ϕ_2^{bath} at $\chi_{\text{max}} > K$ and decreases at $\chi_{\text{max}} < K$.

The governing eqn (16), (19), (28), (30) together with eqn (32) for the critical temperature T_c express the unknown parameters Q and T_c in terms of the volume fraction of additives in the bath ϕ_2^{bath} . These relations are based on the assumption (24) that volume fractions of additives in the bath and in the fluid phase of the gel coincide. Eqn (24) is confirmed by observations^{7,8} on PNIPA gels in aqueous solutions of saccharides with relatively low concentrations of additives at which the gels remain in the swollen state. Our aim now is to prove that condition (24) is fulfilled, and the partition coefficient P is close to unity (independently of the type of additive) when degree of swelling Q is sufficiently large.

For this purpose, we study equilibrium swelling of a TR gel with the energy of interaction between segments of chains and molecules of solvent-1 and solvent-2 described by eqn (2). A detailed derivation of the governing equations for this gel is provided in Appendix C (ESI).† Under the condition that the volume fraction of additives in the bath ϕ_2^{bath} is small compared with unity, our analysis implies that the partition coefficient P reads

$$P = \exp \left(\frac{\chi_0 + \chi_1 T - K}{1 + Q} \right). \quad (33)$$

It follows from eqn (33) that P is close to unity (which implies that the model (16), (19), (28) and (30) coincides with the conventional relations for swelling of a TR gel in a binary solvent) when the equilibrium degree of swelling Q is sufficiently large, that is below the volume phase transition temperature T_c .

4 Fitting of observations

To demonstrate the ability of the model to describe equilibrium swelling diagrams on TR gels in aqueous solutions of



saccharides and to predict changes in degree of swelling Q and critical temperature T_c induced by an increase in volume fraction of additives in the bath, we study observations in swelling tests on PNIPA gel in solutions of monosaccharides (D-glucose and D-galactose) and disaccharide (sucrose).

The gel is prepared by free radical cross-linking polymerization (24 h at 5 °C) of an aqueous solution of *N*-isopropylacrylamide (NIPA) monomers (0.7 M) by using *N,N'*-methylenebis(acrylamide) (BIS, 7.0 mM) as a cross-linker, ammonium persulfate (APS) as an initiator, and *N,N',N',N'*-tetramethylethylenediamine (TEMED) as an accelerator.⁸

We begin with fitting experimental data in equilibrium swelling tests on PNIPA gel in pure water (Fig. 1A). First, observations at temperatures $T < T_c$ are approximated by using the universal coefficients χ_0 , χ_1 reported in Table S-2.† Parameters Q_0 and g_1 are found from the best-fit condition for the experimental dependence $\chi(T)$ below T_c . The coefficient χ_{\max} is determined from measurements of T_c . The quantities β_2 and \bar{g}_2 are found by matching the experimental data above T_c . Adjustable parameters in the governing equations are determined by the method of nonlinear regression from the condition of minimum for the function

$$\sum(Q_{\text{exp}} - Q_{\text{sim}})^2,$$

where summation is conducted over all experimental points, Q_{exp} stands for the degree of swelling measured in tests, and Q_{sim} is calculated from solution of eqn (10), (14) and (16) by the Newton–Raphson algorithm. Fig. 1A demonstrates good agreement between the data and results of simulation with the material constants listed in Table S-3.†

We proceed with fitting equilibrium swelling diagrams in aqueous solutions of glucose, galactose and sucrose at $T = 20$ °C (Fig. 1B, C and D). Each set of data is approximated separately by means of the only parameter K . Given a volume phase transition temperature T_c for PNIPA gel in an aqueous solution of additive with volume fraction ϕ_2^{bath} , this coefficient is calculated from eqn (32). The best-fit values of K are collected in Table S-4.†

To examine the accuracy of predictions of swelling diagrams, we present experimental data in equilibrium swelling tests on PNIPA gel in solutions with three molar fractions of additives (0.3, 1.2 and 2.0 mol L⁻¹) together with results of simulation with the material parameters reported in Table S-3 and S-4.† Observations and results of numerical analysis for swelling tests on PNIPA gel in solutions of glucose are depicted in Fig. 2. Similar results for solutions of galactose and sucrose are reported in Fig. S-4 and S-5.† These figures confirm the ability of the model to predict experimental data.

To evaluate the effect of temperature T on equilibrium swelling curves, simulation is conducted of the governing equations for swelling of PNIPA gel in solutions of glucose with various volume fractions ϕ_2^{bath} at temperatures $T = 15, 20, 25, 30$ and 32 °C. Results of numerical analysis are reported in Fig. 3A. This figure shows that the growth of temperature induces a pronounced shift of the volume phase transition point to smaller volume fractions of the additive. To confirm

this conclusion, results of the numerical analysis at temperature $T = 25$ °C are re-plotted in the dimensionless form in Fig. 3B together with experimental data⁶⁴ on PNIPA gel with the same preparation conditions and a similar composition (the only difference is in the molar fraction of cross-linker: 18.4 mM (ref. 64) compared with 7 mM (ref. 8)).

To examine the influence of volume fraction of additives ϕ_2^{bath} on swelling diagrams, simulation of the governing equations is performed for PNIPA gel immersed in solutions of glucose, galactose and sucrose with ϕ_2^{bath} ranging from 0 to 0.3. Results of numerical analysis are reported in Fig. 4A (glucose), Fig. S-6A† (galactose) and Fig. S-7A† (sucrose), where equilibrium degree of swelling Q is plotted *versus* temperature T . These figures demonstrate that the presence of saccharides in aqueous solutions leads to a noticeable shift of the critical temperature T_c to smaller values. The decay in the volume phase transition temperature T_c with ϕ_2^{bath} is illustrated in Fig. 4B (glucose), Fig. S-6B† (galactose) and Fig. S-7B† (sucrose), where predictions of eqn (32) are compared with experimental data.^{65–67} Good agreement is demonstrated between predictions of the model and experimental data for the critical temperature of PNIPA gel in solutions of glucose and galactose. Slight deviations (less than 2 °C, that is the width of the volume phase transition interval for PNIPA gel) between the data and results of simulation in Fig. S-7C† may be explained by the non-standard technique used in ref. 67 for measurements of T_c .

The coefficient K in Table S-4† can be connected with hydration number n of additives in aqueous solutions. As the latter parameter is model-dependent (its value for saccharides is strongly affected by the method used to find n experimentally), two sets of data are reported in Fig. S-8A and S-8B.† In Fig. S-8A,† K is plotted *versus* n determined from measurements of density and sound velocity of aqueous solutions (parameter n is taken from ref. 68 for glucose and galactose and from ref. 69 for sucrose). In Fig. S-8B,† the same K is depicted *versus* n found by means of terahertz reflection spectroscopy.⁷⁰ In both cases, linear dependencies are revealed between the K values calculated by matching swelling diagrams on PNIPA gel in aqueous solutions of saccharides and their hydration numbers n . To demonstrate that these dependencies have some physical meaning, we calculate K for mannose from the graph in Fig. S-8A,† and apply it to predict (by means of eqn (32)) the effect of volume fraction of mannose ϕ_2^{bath} on the critical temperature T_c of PNIPA gel in mannose solution. Results of simulation are presented in Fig. S-8C† together with experimental data taken from ref. 65. An acceptable agreement is shown between the observations and predictions of the model.

Keeping in mind that the hydration number characterizes interaction between molecules of water and molecules of an additive, the proportionality between the coefficient K and hydration number n means that the parameter χ_{12} in eqn (31) (that accounts for this interaction in the Flory–Rehner model) plays the key role in the volume phase transition of TR gels. This conclusion is in accord with the concept of complexation of cosolvent molecules,^{43–45} according to which hydration of polymer chains requires formation of cage-like structures of water molecules surrounding hydrophobic segments. When water



molecules leave these structures to form complexes with molecules of an additive, the chains dehydrate and collapse.

5 Concluding remarks

A mean-field model is developed for equilibrium swelling of thermo-responsive gels in aqueous solutions of additives. Three issues are worth to be mentioning that distinguish our model from the conventional approach to swelling of TR gels in binary mixtures: (i) an increase in the shear modulus of a gel in the collapsed state (driven by aggregation of hydrophobic segments into clusters from which solvent molecules are expelled) is taken into account, see eqn (10) and (19), (ii) the governing equations allow the volume phase transition temperature T_c to be determined explicitly as a function of volume fraction of additive, see eqn (32), and (iii) to reduce the number of adjustable parameters, the entire set of FH coefficients χ_{ij} is not employed in the analysis (although connection (31) is established between these quantities and coefficient K in the model).

The model is grounded on the following assumptions: (i) hydrophobicity of polymer chains in a network is characterized by the only scalar coefficient χ_{eq} . In the swollen state of a gel, evolution of this quantity with temperature and volume fraction of additive is governed by semi-phenomenological eqn (30). Volume phase transition in a TR gel occurs when the equivalent FH parameter χ_{eq} reaches its critical value χ_{max} . (ii) In the collapsed state, the FH parameter adopts its ultimate value χ_{max} , and the kinetics of association of hydrophobic segments into clusters is governed by the order parameter (17). (iii) In the simplified model, volume fraction of additives in the fluid phase of a gel is presumed to coincide with that in the bath, see eqn (24). This hypothesis is confirmed by experimental data, and its accuracy is verified analytically, see eqn (33).

The model for the thermo-mechanical response and equilibrium swelling of a TR gel in a solution of additives is derived by means of the free energy imbalance inequality (Fig. S-8†). The governing eqn (16), (19), (28) and (30) involve eight material constants with transparent physical meaning: (i) g_1 stands for the dimensionless shear modulus of the polymer network with covalent bonds, (ii) Q_0 denotes degree of swelling of a gel in the stress-free state, (iii) χ_0 , χ_1 characterize the effect of temperature on the equivalent FH parameter χ_{eq} , (iv) K accounts for evolution of χ_{eq} with volume fraction of additives, (v) χ_{max} is the ultimate value of the equivalent FH parameter characterizing volume phase transition in the gel, and (vi) \bar{g}_2 and β describe the kinetics of aggregation of hydrophobic segments of chains into clusters in the collapsed state. The critical temperature T_c is found from eqn (32).

The coefficients χ_0 , χ_1 and χ_{max} are treated as “universal,” which means that their values are determined by chemical structure of monomers only. Parameters Q_0 , \bar{g}_2 and β are considered as “semi-universal”: these quantities are affected by the type and concentration of cross-linker, but are independent of temperature and volume fraction of additives. The coefficient K is the only parameter that accounts for the chemistry of additives.

The model is applied to the analysis of equilibrium swelling of PNIPAA gel in aqueous solutions on saccharides at various temperatures T . Fig. 1 demonstrates good agreement between the experimental swelling diagrams and results of simulation. The ability of the model to predict the effect of temperature on the equilibrium degree of swelling of PNIPAA gel in solutions of glucose, galactose and sucrose with various concentrations is confirmed by Fig. 2, 3, S-4 and S-5.† Its ability to predict changes in the volume phase transition temperature T_c with volume fraction of saccharides in the bath is illustrated in Fig. 4, S-6–S-8.†

Conflict of interest

There are no conflicts of interest to declare.

Acknowledgements

Financial support by Innovationsfonden (Innovation Fund Denmark, project 9091-00010B) is gratefully acknowledged.

References

- 1 A. Gandhi, A. Paul, S. O. Sen and K. K. Sen, *Asian J. Pharm. Sci.*, 2015, **10**, 99–107.
- 2 M. A. Cohen-Stuart, W. T. Huck, J. Genzer, M. Muller, C. Ober, M. Stamm, G. B. Sukhorukov, I. Szleifer, V. V. Tsukruk, M. Urban, F. Winnik, S. Zauscher, I. Luzinov and S. Minko, *Nat. Mater.*, 2010, **9**, 101–113.
- 3 P. Zarrintaj, M. Jouyandeh, M. R. Ganjali, B. S. Hadavand, M. Mozafari, S. S. Sheiko, M. Vatankehah-Varnoosfaderani, T. J. Gutierrez and M. R. Saeb, *Eur. Polym. J.*, 2019, **117**, 402–423.
- 4 Y. Hirokawa and T. Tanaka, *J. Chem. Phys.*, 1984, **81**, 6379–6380.
- 5 K. K. Bansal, P. K. Upadhyay, G. K. Saraogi, A. Rosling and J. M. Rosenholm, *EXPRESS Polym. Lett.*, 2019, **13**, 974–992.
- 6 M. Shibayama, *Polym. J.*, 2011, **43**, 18–34.
- 7 T. Ishidao, I.-S. Song, N. Ohtani, K. Sato, Y. Iwai and Y. Arai, *Fluid Phase Equilib.*, 1997, **136**, 163–171.
- 8 H. Kawasaki, S. Sasaki, H. Maeda, S. Mihara, M. Tokita and T. Komai, *J. Phys. Chem.*, 1996, **100**, 16282–16284.
- 9 S. Koga, S. Sasaki and H. Maeda, *J. Phys. Chem. B*, 2001, **105**, 4105–4110.
- 10 M. Annaka, K. Motokawa, S. Sasaki, T. Nakahira, H. Kawasaki, H. Maeda, Y. Amo and Y. Tominaga, *J. Chem. Phys.*, 2000, **113**, 5980–5985.
- 11 E. Kokufuta, S. Nakaizumi, S. Ito and T. Tanaka, *Macromolecules*, 1995, **28**, 1704–1708.
- 12 T. Ishidao, M. Akagi, H. Sugimoto, Y. Iwai and Y. Arai, *Macromolecules*, 1993, **26**, 7361–7362.
- 13 E. Kokufuta and T. Tanaka, *Macromolecules*, 1991, **24**, 1605–1607.
- 14 H. R. Culver, J. R. Clegg and N. A. Peppas, *Acc. Chem. Res.*, 2017, **50**, 170–178.
- 15 I. Y. Jung, J. S. Kim, B. R. Choi, K. Lee and H. Lee, *Adv. Healthcare Mater.*, 2017, **6**, 1601475.



- 16 L. Hu, Q. Zhang, X. Li and M. J. Serpe, *Mater. Horiz.*, 2019, **6**, 1774–1793.
- 17 J. R. Clegg and N. A. Peppas, *Soft Matter*, 2020, **16**, 856–869.
- 18 P. J. Flory and J. Rehner, *J. Chem. Phys.*, 1943, **11**, 521–526.
- 19 N. R. Richbourg and N. A. Peppas, *Prog. Polym. Sci.*, 2020, **105**, 101243.
- 20 S. Hirotsu, *J. Chem. Phys.*, 1988, **88**, 427–431.
- 21 Y. C. Bae, J. J. Shim, D. S. Soane and J. M. Prausnitz, *J. Appl. Polym. Sci.*, 1993, **47**, 1193–1206.
- 22 M. Shibayama and T. Tanaka, *Adv. Polym. Sci.*, 1993, **109**, 1–62.
- 23 H. Kojima, *Polym. J.*, 2018, **50**, 411–418.
- 24 A. D. Drozdov, *Eur. Phys. J. E*, 2014, **37**, 93.
- 25 A. D. Drozdov, *Acta Mech.*, 2015, **226**, 1283–1303.
- 26 L. Hanykova, J. Spevacek, M. Radecki, A. Zhigunov, J. Stastna, H. Valentova and Z. Sedlakova, *Colloid Polym. Sci.*, 2015, **293**, 709–720.
- 27 M. Lehmann, P. Krause, V. Miruchna and R. von Klitzing, *Colloid Polym. Sci.*, 2019, **297**, 633–640.
- 28 A. D. Drozdov and J. deClaville Christiansen, *Polymer*, 2017, **132**, 164–173.
- 29 A. D. Drozdov, *Int. J. Eng. Sci.*, 2018, **128**, 79–100.
- 30 A. R. Shultz and P. J. Flory, *J. Polym. Sci.*, 1955, **15**, 231–242.
- 31 O. Okeowo and J. R. Dorgan, *Macromolecules*, 2006, **39**, 8193–8202.
- 32 R. Xiao, J. Qian and S. Qu, *Int. J. Appl. Mech. Eng.*, 2019, **11**, 1950050.
- 33 R. Koningsveld and L. A. Kleintjens, *Macromolecules*, 1971, **4**, 637–641.
- 34 D. S. Abrams and J. M. Prausnitz, *AIChE J.*, 1975, **21**, 116–128.
- 35 Q. Xin, C. Peng, H. Liu and Y. Hu, *Ind. Eng. Chem. Res.*, 2008, **47**, 9678–9686.
- 36 S. Katayama, Y. Hirokawa and T. Tanaka, *Macromolecules*, 1984, **17**, 2641–2643.
- 37 D. Mukherji, C. M. Marques and K. Kremer, *J. Phys.: Condens. Matter*, 2018, **30**, 024002.
- 38 V. Aseyev, H. Tenhu and F. M. Winnik, *Adv. Polym. Sci.*, 2011, **242**, 29–89.
- 39 A. Halperin, M. Kroger and F. M. Winnik, *Angew. Chem., Int. Ed.*, 2015, **54**, 15342–15367.
- 40 Y. Ono and T. Shikata, *J. Am. Chem. Soc.*, 2006, **128**, 10030–10031.
- 41 D. Kurzbach, M. J. N. Junk and D. Hinderberger, *Macromol. Rapid Commun.*, 2013, **34**, 119–134.
- 42 L. Comez, M. Paolantoni, P. Sassi, S. Corezzi, A. Morresi and D. Fioretto, *Soft Matter*, 2016, **12**, 5501–5514.
- 43 G. Zhang and C. Wu, *J. Am. Chem. Soc.*, 2001, **123**, 1376–1380.
- 44 F. Tanaka, T. Koga and F. M. Winnik, *Phys. Rev. Lett.*, 2008, **101**, 028302.
- 45 T. Zuo, C. Ma, G. Jiao, Z. Han, S. Xiao, H. Liang, L. Hong, D. Bowron, A. Soper, C. C. Han and H. Cheng, *Macromolecules*, 2019, **52**, 457–464.
- 46 W. L. A. Brooks and B. S. Sumerlin, *Chem. Rev.*, 2016, **116**, 1375–1397.
- 47 B. J. van Enter and E. von Hauff, *Chem. Commun.*, 2018, **54**, 5032–5045.
- 48 A. D. Drozdov and J. de. C. Christiansen, *Int. J. Solids Struct.*, 2013, **50**, 3570–3585.
- 49 A. D. Drozdov and J. deClaville Christiansen, *Acta Mech.*, 2018, **229**, 5067–5092.
- 50 A. Ikehata and H. Ushiki, *Polymer*, 2002, **43**, 2089–2094.
- 51 A. M. Morris, M. A. Watzky and R. G. Finke, *Biochim. Biophys. Acta*, 2009, **1794**, 375–397.
- 52 A. S. Buchelnikov, V. P. Evstigneev and M. P. Evstigneev, *Phys. Chem. Chem. Phys.*, 2019, **21**, 7717–7731.
- 53 T. Norisuye, Y. Kida, N. Masui, Q. Tran-Cong-Miyata, Y. Maekawa, M. Yoshida and M. Shibayama, *Macromolecules*, 2003, **36**, 6202–6212.
- 54 M. Shibayama, Y. Shirotani, H. Hirose and S. Nomura, *Macromolecules*, 1997, **30**, 7307–7312.
- 55 K. Poschlad and S. Enders, *J. Chem. Thermodyn.*, 2011, **43**, 262–269.
- 56 H. E. Yang and Y. C. Bae, *J. Polym. Sci., Part B: Polym. Phys.*, 2017, **55**, 455–463.
- 57 W. H. Stockmayer, L. D. Moore, M. Fixman and B. N. Epstein, *J. Polym. Sci.*, 1955, **16**, 517–530.
- 58 Z. S. Petrovic, W. J. MacKnight, R. Koningsveld and K. Dusek, *Macromolecules*, 1987, **20**, 1088–1096.
- 59 R. G. M. van der Sman, *Food Funct.*, 2017, **8**, 360–371.
- 60 P. K. Banipal, N. Aggarwal and T. S. Banipal, *J. Chem. Eng. Data*, 2016, **61**, 1992–2001.
- 61 F. M. Winnik, H. Ringsdorf and J. Venzmer, *Macromolecules*, 1990, **23**, 2415–2416.
- 62 S. Hirotsu, T. Okajima and T. Yamamoto, *Macromolecules*, 1995, **28**, 775–777.
- 63 F. Rodriguez-Ropero, T. Hajari and N. F. A. van der Vegt, Mechanism of polymer collapse in miscible good solvents, *J. Phys. Chem. B*, 2015, **119**, 15780–15788.
- 64 N. Manukovsky, A. Shpigelman, R. Edelman and Y. D. Livney, *J. Polym. Sci., Part B: Polym. Phys.*, 2011, **49**, 523–530.
- 65 A. Shpigelman, I. Portnaya, O. Ramon and Y. D. Livney, *J. Polym. Sci., Part B: Polym. Phys.*, 2008, **46**, 2307–2318.
- 66 A. Shpigelman, Y. Paz, O. Ramon and Y. D. Livney, *Colloid Polym. Sci.*, 2011, **289**, 281–290.
- 67 P. Narang, S. B. Vepuri, P. Venkatesu and M. E. Soliman, *J. Colloid Interface Sci.*, 2017, **504**, 417–428.
- 68 R. Edelman, I. Kusner, R. Kisiliak, S. Srebnik and Y. D. Livney, *Food Hydrocolloids*, 2015, **48**, 27–37.
- 69 M. Starzak, S. D. Peacock and M. Mathlouthi, *Crit. Rev. Food Sci. Nutr.*, 2000, **40**, 327–367.
- 70 K. Shiraga, Y. Ogawa, N. Kondo, A. Irisawa and M. Imamura, *Food Chem.*, 2013, **140**, 315–320.

