PAPER

Check for updates

Propyl acetate/butyronitrile mixture is ideally suited for investigating the effect of dielectric stabilization on (photo)chemical reactions†

Pragya Verma, [ID] Arnulf Rosspeintner [ID] and Tatu Kumpulainen [ID] *

Characterization of propyl acetate/butyronitrile (PA/BuCN) mixtures by various spectroscopic techniques is described. The neat solvents have identical viscosities and refractive indices but their dielectric constants differ significantly. Detailed solvatochromic and titration data show that the mixtures do not exhibit specific solute–solvent interactions or significant dielectric enrichment effects. Therefore, the mixtures are ideally suited for investigating the effect of dielectric stabilization on (photo)chemical reactions. Dynamic Stokes shift experiments performed on two push–pull probes demonstrate that the solvation dynamics are significantly decelerated in the mixtures as compared to the neat solvents. Therefore, the mixtures allow for varying both the extent and time scale of the dielectric stabilization in a predictable manner.

Introduction

Variation in chemical reactivity in solutions is often correlated with empirical solvent scales or macroscopic solvent parameters derived from a continuous description of the liquid.[1,2] In addition to "static" or equilibrium solvent effects, solvation dynamics play an important role in many ultrafast photochemical reactions in solutions.[3–9] Understanding the effect of the macroscopic solvent parameters on chemical reactions is of great fundamental importance but it is challenging to investigate one single parameter at a time. The main problem arises from the fact that most solvents do not have a single property that is different from the properties of other solvents. For example, while studying the effect of increasing dielectric constants, other properties like viscosity and refractive index might also change thus influencing the reaction under study. One solution to overcome this problem is to use solvent mixtures. This approach has been successfully utilized for investigating both chemical and photochemical reactions.[10–20]

Solvent mixtures of varying dielectric constant enable systematic studies on the influence of solvent stabilization on photochemical reactions.[18–21] Ideally, other solvent parameters such as viscosity and refractive index should remain constant. In addition, the mixtures should not exhibit any specific hydrogen-bonding (HB) interactions or dielectric enrichment effects that might influence the reactivity. Specific solvent interactions can disrupt hydrogen bonds between the reaction partners or alter the solvent stabilization in a non-systematic way.[22–26] On the other hand, solvents with a large difference in the dielectric constants (ε_r) are particularly prone to dielectric enrichment effects.[27–29]

A binary mixture between propyl acetate (**PA**) and butyronitrile (**BuCN**) is a promising candidate for investigating the effect of dielectric stabilization. The dielectric constant increases from 6.0 to 24.8 upon going from **PA** to **BuCN** and varies linearly with the weight fractions of the solvents according to:

$$\varepsilon_r = w_{\mathbf{PA}} \cdot \varepsilon_r(\mathbf{PA}) + (1 - w_{\mathbf{PA}}) \cdot \varepsilon_r(\mathbf{BuCN}), \tag{1}$$

where $w_{\mathbf{PA}}$ is the weight fraction of **PA**, whereas the viscosity (η) and refractive index (n) remain constant (Section S1, ESI†). The Kamlet–Taft parameter β, related to the HB accepting ability of the solvent, differs slightly whereas the α parameter (HB donating ability) is zero for both solvents (Table 1).[30] This specific mixture has been used to investigate the influence of dielectric constant on photoinduced electron transfer and

Department of Physical Chemistry, University of Geneva, 30 Quai Ernest Ansermet, Geneva, Switzerland. E-mail: tatu.kumpulainen@unige.ch; Fax: +41 22 379 6518; Tel: +41 22 379 6530

† Electronic supplementary information (ESI) available: Additional experimental details, characterization of the macroscopic solvent properties, supplementary experimental data, fits and parameters. See DOI: 10.1039/d0ra04525j

Table 1 The main solvent properties of interest for PA and BuCN[a]

Solvent	η (cP)	ε_r	n	α	β	π^*	μ (D)
PA	0.551	6.0	1.382	0.00	0.40	0.53	1.78
BuCN	0.549	24.8	1.382	0.00	0.45	0.63	4.07

[a] α, β, and π^* are the Kamlet–Taft parameters related to HB donating, HB accepting ability, and dipolarity/polarizability respectively and μ is the dipole moment of the solvent. All parameters were taken from ref. 45.

exciplex formation.[18–21] However, a detailed characterization of the properties of the mixtures has not been reported. We are interested in using the mixture for investigating ultrafast excited-state proton-transfer reactions and, therefore, we pay special attention to any possible specific interactions that might interfere with proton transfer dynamics. Secondly, the dynamics of the solvent relaxation in the **PA**/**BuCN** mixtures are characterized for the first time.

The article is structured as follow: first, we describe the steady-state solvatochromic behavior of betaine-30 (ref. 23) (**B30**), coumarin 153 (**C153**), and coumarin 152A (**C152A**) (Chart 1). The extremely large ground-state dipole moment of *ca.* 15 D (ref. 31) and the presence of the negatively charged phenolate oxygen render **B30** absorption band frequency very sensitive towards both HB donors and dielectric enrichment effects.[32,33] On the other hand, **B30** is nearly non-fluorescent.[34–36] Therefore, the two coumarins were used and their solvatochromism was investigated by using both steady-state absorption and emission. Additionally, we present a detailed investigation on the solvent relaxation parameters utilizing the time-dependent Stokes shift method[37] on broadband fluorescence spectra with a sub-100 fs time-resolution.[38,39] Time constants for the solvent relaxation were determined for both **C153** and **C152A** in the mixtures. In the third part, we present a detailed investigation on the microscopic viscosity by investigating both the rotational and translational diffusion of **C153** in the mixtures. Both of these coefficients depend on the viscosity and hence should remain constant in isoviscous mixtures in the absence of specific solute–solvent interactions.[40–42]

Since our main goal is to use the mixtures for proton transfer reactions, we demonstrate in the last part that the mixture does not disrupt the HB interactions between the reaction partners and that the dielectric constant has a significant impact on the driving force of the proton-transfer reaction. For this purpose, we use a 1,8-naphthalimide-based photoacid (Chart 1) complexed with either a weak base (*N*-methylimidazole, **NMI**) or deprotonated by a strong base (1,8-diazabicyclo[5.4.0]undec-7-ene, **DBU**).[43,44]

Experimental section

Materials

The dyes, BBOT and **C153** (Radiant Dyes), DCM, **C152A** and coumarin 6H (Exciton) and **B30** (Sigma Aldrich) were of highest

available grade and used without further purification. The solvents, *n*-butyronitrile (99%, Acros Organics, distilled at 154 °C) and propyl acetate (99%, Alfa Aesar) were stored over 3 Å molecular sieves under argon atmosphere to avoid moisture. Synthesis of the 1,8-naphthalimide-based photoacid, **C₄-dHONI**, has been described elsewhere.[43] The bases, **DBU** (99%, Alfa Aesar) and **NMI** (99%, Sigma Aldrich), were used as received.

Experimental methods

Steady-state absorption spectra were recorded using a Varian Cary 50 UV-vis spectrometer and emission spectra using a Horiba Scientific FluoroMax-4 (Jobin Yvon) fluorimeter corrected for the spectral sensitivity using a set of secondary emissive standards.[46] The fluorescence anisotropies were measured using a fs single wavelength fluorescence upconversion setup that has been described in previous publications.[47,48] The NMR spectra were recorded on a Bruker Avance III HD-NanoBay 300 MHz spectrometer at a temperature of 298 K. Time-resolved broadband fluorescence spectra were recorded using a setup similarly to those described in ref. 49 and 50. The full width at half-maximum (FWHM) of the cross correlation of the gate with solvent Raman signal was approximately 90 fs. The raw spectra were corrected for the spectral sensitivity with the use of a set of secondary emissive standards[46] and subsequently transferred into the transition dipole moment representation.[51] The chirp due to group velocity dispersion was determined by measuring the instantaneous response of BBOT in the same solvent. All measurements were performed at room temperature (21 ± 1 °C). A sample cell with a 1 cm optical path length was used for steady-state measurements and with a 1 mm path length for the time-resolved measurements.

Data analysis

The transition energies of all probes were determined by fitting the bands with a log-normal function according to:[52,53]

$$F(\tilde{\nu}) = I_0 \begin{cases} \exp\left[-\ln(2)\{\ln(1 + a(\tilde{\nu}))/b\}^2\right], & \alpha(\tilde{\nu}) > -1 \\ 0, & \alpha(\tilde{\nu}) \le -1 \end{cases} \quad (2)$$

$$\alpha(\tilde{\nu}) = \frac{2b(\tilde{\nu} - \tilde{\nu}_0)}{\Delta x}, \quad (3)$$

where I_0 is the band intensity, $\tilde{\nu}_0$ the peak frequency, b the asymmetry parameter and Δx the width parameter. Due to extensive literature data on $E_T(30)$ scales,[23,45,54] the transition energies of **B30** were determined from the absorption spectra in wavenumber and the corresponding $E_T(30)$ values of the mixtures were calculated according to:

$$E_T(30) \ [\text{kcal mol}^{-1}] = hc\tilde{\nu}_0 N_A = 2.8591 \cdot \tilde{\nu}_0 \ [\text{cm}^{-1}], \quad (4)$$

where $\tilde{\nu}_0$ is the peak frequency of the lowest-energy transition determined by fitting with the log-normal function.

The transition energies of the coumarins were determined from the peak frequencies of the log-normal fits in transition dipole moment (TDM) representation since this corresponds to the true lineshape of the underlying transitions.[51] This is
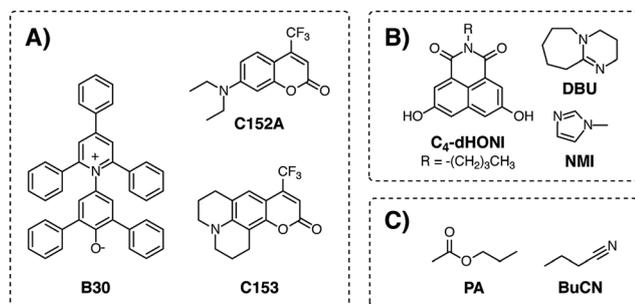

Chart 1 Chemical structures of (A) solvatochromic probes, (B) photoacid and organic base, and (C) solvents used in the study. Same abbreviations are used throughout the text.

supported by the bandwidths of the absorption and emission spectra, presented in Fig. S7 (ESI†). The absorption and emission bandwidths are nearly identical in the TDM representation whereas they differ by more than 500 cm^{-1} in the wavenumber representation.

All transition energies and Stokes shifts are represented as a function of the reaction field factor, Δf, defined as:

$$\Delta f = f(\varepsilon_r) - f(n^2) = \frac{2(\varepsilon_r - 1)}{2\varepsilon_r + 1} - \frac{2(n^2 - 1)}{2n^2 + 1}. \quad (5)$$

The factor, derived from Onsager's reaction-field theory, accounts for the dielectric stabilization of a non-polarizable point dipole in a spherical cavity due orientational dipole–dipole interactions.[55] Since the refractive index of the mixtures is constant, the transition energies are expected to show linear behavior with respect to Δf with slopes proportional to a combination of the ground- and excited-state permanent dipole moments.[56,57] Any deviations from linearity would indicate the presence of specific solute–solvent interactions or dielectric enrichment effects. For consistency, all solvent parameters are plotted against Δf and the weight fractions of **PA** were chosen to be equally spaced in Δf. Relation between Δf and the weight fraction (Fig. S4, ESI†) is nonlinear.

Rotational diffusion times of **C153** were determined from the fluorescence anisotropy decays. These decays were calculated from intensities measured at a parallel ($I_\parallel$) and perpendicular ($I_\perp$) pump polarization with respect to the detected signal according to:

$$r(t) = \frac{I_\parallel - I_\perp}{I_\parallel + 2I_\perp}. \quad (6)$$

The rotational times were obtained by fitting the anisotropy decays with a bi-exponential function. The goodness of fit was judged based on the weighted residual and the χ_r^2 value. Translational diffusion coefficients of **C153** were determined from the ^{19}F-DOSY NMR experiments (see ESI† for details).

Solvation dynamics were determined from the fs-broadband fluorescence spectra using the time-dependent Stokes shift method.[37] The spectra were analyzed globally using the approach discussed in detail in ref. 58. In brief, the fluorescence band is fitted with a time-dependent log-normal function where the dynamic Stokes shift is expected to follow a multi-exponential function.

Results and discussion

Steady-state solvatochromism

The absorption spectra of **B30** in the **PA/BuCN** mixtures are presented in Fig. 1A. The absorption band undergoes a blue shift of *ca.* 2000 cm^{-1} upon changing the solvent from neat **PA** ($\Delta f = 0.39$) to neat **BuCN** ($\Delta f = 0.56$), which can be attributed to the well-known strong negative solvatochromism of **B30**.[23] The peak frequencies were converted to $E_T(30)$ values according to eqn (4) and are plotted as a function of Δf in Fig. 1B.
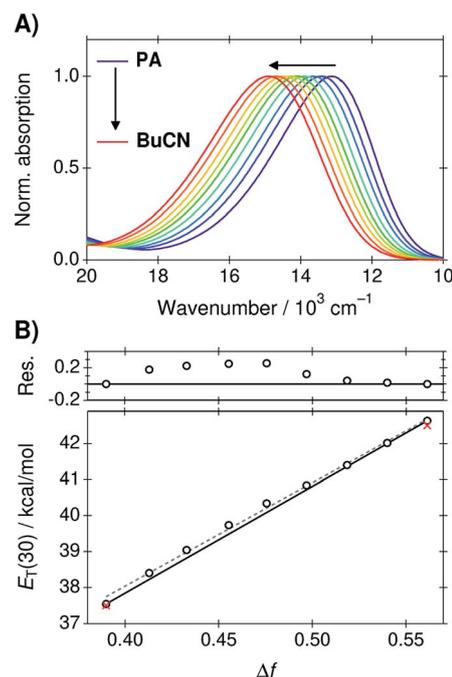

Fig. 1 (A) Absorption spectra of **B30** in the **PA/BuCN** solvent mixtures. (B) Corresponding $E_T(30)$ values as a function of Δf. The black line represents ideal behavior connecting the two extreme points corresponding to the neat solvents and deviation of the experimental values is shown in the top panel in kcal mol^{-1}. The dashed gray line represents a linear regression line to all points. Red crosses indicate the $E_T(30)$ values of the neat solvents taken from ref. 45.

The $E_T(30)$ values show a good linear correlation as a function Δf ($R^2 = 0.997$) represented by the dashed gray line in Fig. 1B. Minor deviations are, however, visible at large weight fractions of **PA** ($\Delta f < 0.5$). In the ideal case, the $E_T(30)$ values would vary linearly between the extreme points corresponding to the neat solvents represented by the black solid line in Fig. 1B. The solvation energy increases more rapidly in **PA** rich mixtures but returns to ideal behavior at around $\Delta f = 0.52$ corresponding to a nearly 50/50 mixture ($w_{PA} = 0.56$). Deviation of the experimental values from the ideal behavior is presented in the top panel of Fig. 1B. It should be noted, that **B30** is particularly susceptible to both dielectric enrichment as well HB donor interactions. We observed significant deviations from linearity before purification of the solvents. Therefore, the purity of the solvents was confirmed by comparing the measured $E_T(30)$ values of the neat solvent to the literature values (red crosses in Fig. 1B) prior any further measurements. After purification the agreement between the measured and literature values was around 0.1 kcal mol^{-1}. Therefore, the observed deviation from linearity is unlikely due to any impurities but can be attributed to the dielectric enrichment. However, the maximum deviation of 0.25 kcal mol^{-1} at $\Delta f = 0.48$ represents only 5% of the total variation and overall linear correlation between the solvation energy and Δf is excellent.

The peak frequencies of the absorption and emission spectra of **C153** and **C152A** were determined in the TDM representation and the Stokes shifts were calculated as a difference between the

absorption and emission frequencies. Experimental spectra and spectral parameters are given in ESI.† The peak frequencies and Stokes shifts as a difference to the values determined in neat **PA** are plotted as a function of Δf in Fig. 2. The negative slopes of the peak frequencies indicate red shifts of the bands whereas the positive slopes of the Stokes shifts indicate an increase.

For both coumarins, the linear correlation between the transition energies and Δf is reasonably good. **C153** shows slight scattering of the points, which are overemphasized in the Stokes shifts. Deviations from the linear regression lines are shown in the top panels. However, this largely originates from uncertainties in the fitted peak maxima due to the log-normal function and the deviations do not show any clear patterns. For **C153** in particular, the log-normal function is a sub-optimal band shape function in solvents of medium polarity. For **C152A**, the log-normal function reproduces the spectra better and the spectral shifts are larger resulting in an excellent linear correlation. Therefore, the observed solvatochromism of both compounds can be attributed to non-specific dielectric stabilization.

In summary, dielectric stabilization in the mixtures follows nearly ideal behavior for all compounds investigated. Although slight dielectric enrichment was observed for **B30**, the linear correlation between the solvation energies and Δf is remarkably good. Therefore, the mixture is ideally suited for controlling the extent of the dielectric stabilization.

Solvation dynamics

Solvation dynamics were determined from the time-resolved fluorescence spectra of **C153** and **C152A** in the neat solvents and three mixtures using the time-dependent Stokes shift method.[37] The spectra were analyzed globally in order to extract the time constants for the dynamic Stokes shifts.[37,58] A three-

exponential function was required to model the fluorescence peak frequencies in **PA** and in the mixtures whereas two-exponential function was adequate in **BuCN**. Experimental spectra, global fits, and weighted residuals are given in ESI.† Time-dependent fluorescence band positions and exponential fits of **C152A** are presented in Fig. 3A. The amplitudes and lifetimes as an average from the two coumarins are summarized in Table 2. All the individual best-fit parameters are given in ESI.†

Similarly to the steady-state spectra, magnitudes of the time-dependent Stokes shifts increase upon increasing Δf, **C152A** showing slightly larger shifts compared to **C153**. The relaxed fluorescence band maxima resulting from the global fit decrease linearly with Δf and are within 100 cm^{-1} from the peak maxima of the steady-state spectra. However, the band maxima of the initial time-zero spectra ($\tilde{\nu}_{t=0}$) as well as the total shifts ($\Delta \tilde{\nu}_{tot}$) are significantly overestimated in the global fits for both compounds. This is largely due to the non-exponential behavior of the spectral shifts at $t < 150$ fs that is sometimes modeled as a Gaussian decay.[26,59,60] In this region, the exponential fit significantly deviates from the experimental band maxima as can be seen in Fig. 3A. Therefore, the total shifts were additionally calculated using the time-zero spectra estimated from
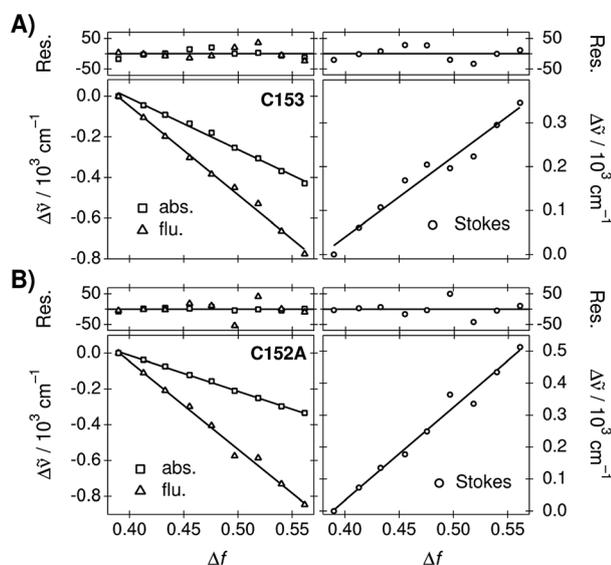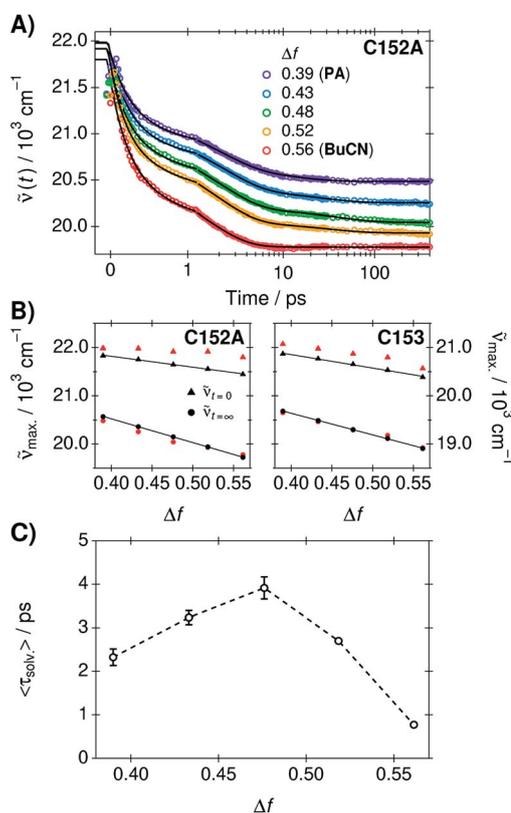

Fig. 3 (A) Time-dependent fluorescence band position of **C152A** in the PA/BuCN mixtures. The solid lines represent the band positions obtained from the global analysis corresponding to two- or three-exponential functions. (B) Fluorescence band maxima of the relaxed ($\tilde{\nu}_{t=\infty}$, circles) and time–zero ($\tilde{\nu}_{t=0}$, triangles) spectra estimated from steady–state spectra (black) or resulting from the global analysis (red). (C) The mean solvation times corresponding to the average value from the two coumarins.

Fig. 2 Absorption/emission frequencies and Stokes shifts of (A) **C153** and (B) **C152A** in PA/BuCN mixtures as a function of Δf. The frequencies are given as a difference to the value measured in neat **PA**. The lines represent linear fits and deviations of the experimental points from the fits are shown in the top panels in cm^{-1}.

Table 2 Lifetimes and amplitudes (in %) for the solvation relaxation in the **PA/BuCN** mixtures. All values correspond to the average values obtained for **C153** and **C152A**[a]

Δf	α_1	τ_1/ps	α_2	τ_2/ps	α_3	τ_3/ps	$\langle\tau_{\text{solv.}}\rangle$/ps
0.39	0.54	0.25	0.37	2.1	0.09	15.8	2.3 ± 0.2
0.43	0.54	0.24	0.37	2.3	0.09	25.2	3.2 ± 0.2
0.48	0.53	0.25	0.39	2.1	0.08	36.1	3.9 ± 0.3
0.52	0.55	0.24	0.38	2.0	0.07	27.4	2.7 ± 0.1
0.56	0.61	0.26	0.39	1.6	—	—	0.8 ± 0.1

[a] Average solvation time was calculated according to $\langle\tau_{\text{solv.}}\rangle = \Sigma\alpha_i\tau_i$.

the steady-state spectra in non-polar solvent according to a previously published procedure (see ESI† for details and representative spectra).[58,61]

The peak frequencies of the time-zero and relaxed fluorescence estimated from the steady-state spectra (black markers) and those resulting from the global fit (red markers) are presented in Fig. 3B. The total shifts, reflecting the solvation free energy,[62] increase linearly with Δf in the estimated spectra whereas the shifts resulting from the global fit are overestimated by *ca.* 200–300 cm^{-1}. However, the experimental band positions (markers in Fig. 3A) at $t = 0$ agree within 100 cm^{-1} with the estimated time-zero spectra. This further supports that the dielectric stabilization follows ideal behavior and is not complicated by specific solute–solvent interactions or dielectric enrichment effects.[62]

Despite the different magnitude of the total shifts, the time constants and relative amplitudes of the two coumarins are very similar. This demonstrates that the solvation dynamics are independent of the coumarin, in agreement with previous results.[58] Therefore, the characteristic solvent relaxation parameters, presented in Table 2, are given as an average of the individual parameters obtained for **C153** and **C152A**. The mean solvation times correspond to amplitude-weighted average lifetimes and are presented in Fig. 3C as a function of Δf.

Surprisingly, the solvation dynamics are significantly decelerated in the mixtures. Due to this unexpected behavior, the experiments were performed in duplicate reproducing the same behavior. The solvent relaxation becomes gradually slower upon increasing the amount of **BuCN** reaching a maximum at $\Delta f = 0.48$ corresponding to $w_{\text{PA}} = 0.79$. The first two lifetimes do not show significant differences, but the third time constant increases significantly in the mixtures. After this point, the solvation dynamics become faster upon increasing fraction of **BuCN**. In neat **BuCN**, the solvation dynamics are the fastest and slowest decay component (τ_3) disappears. The physical origin of this effect remains unclear and is beyond the scope of the present study. However, similar deceleration has been observed in other binary mixtures where it has been attributed to deceleration of the intrinsic solvent reorientation dynamics or translational components.[18,63,64] The rather long time scale of the deceleration observed in **PA/BuCN** mixtures points towards translational components that likely involve a significant reorientation of the first solvation shell.

Rotational and translational diffusion coefficients

The experimentally measured kinematic viscosity of the mixtures remains constant (Fig. S3, ESI†). However, we wanted to investigate whether molecular effects might influence the diffusion of solutes in the mixtures. Both the rotational and translational diffusion of solutes are related to the macroscopic viscosity of the solvent. Hence, the coefficients are expected to remain constant in solutions of constant viscosity for a specific solute. Deviations from hydrodynamic theories are often used as indicators for specific solute–solvent interactions that increase the friction.[42]

The rotational diffusion coefficient is proportional to the inverse of the rotational correlation time ($\tau_{\text{rot.}}$). For a spherical molecule, $\tau_{\text{rot.}}$ is directly proportional to the viscosity of solvent according to the Stokes–Einstein–Debye equation:[41,42]

$$\tau_{\text{rot.}} = \frac{V\eta}{k_{\text{B}}T} \tag{7}$$

where V is the hydrodynamic volume, η viscosity and k_{B} the Boltzmann constant, and T the temperature. On the other hand, the translational diffusion coefficient is inversely proportional to the viscosity according to the Stokes–Einstein equation:[40]

$$D = \frac{k_{\text{B}}T}{6\pi\eta R_{\text{rdW}}} \tag{8}$$

where R_{rdW} is the van der Waals radius of the molecule.

The rotational correlation times of **C153** were determined from the fluorescence anisotropy decays in the **PA/BuCN** mixtures.[42] Representative fluorescence decays measured in parallel and perpendicular polarizations together with the constructed anisotropy decay are shown in Fig. S12 (ESI†). The anisotropy decays were analyzed with a bi-exponential function and the best-fit parameters are summarized in Table S9 (ESI†). The bi-exponential decay originates from the non-spherical shape of **C153** (ref. 65) but both individual decay times are expected to show linear correlation with viscosity, as has been observed in other binary mixtures.[16] The mean rotational correlation times, $\langle\tau_{\text{rot.}}\rangle$, were calculated as an amplitude-weighted average lifetime and are presented as a function of Δf in Fig. 4A. The translational diffusion coefficients of **C153** were determined from the ^{19}F-DOSY experiments in the **PA/BuCN** mixtures. Experimental details and representative data are given in ESI.† The diffusion coefficients are presented as a function of Δf in Fig. 4B.

Both the rotational correlation times and translational diffusion coefficients are nearly constant in the mixtures. This unambiguously demonstrates that the viscosity of the mixtures remains constant. The invariant rotational correlation times are additionally a further indication for the absence of strong solute–solvent interactions. Both specific interactions and dielectric enrichment has been shown to influence the rotational correlation times.[42,66] Our results show that the mixture behaves ideally and does not show any deviations from hydrodynamic theories.

Ground-state association and deprotonation constants

As mentioned in the introduction, our main aim is to use the **PA/BuCN** mixtures in further studies on bimolecular excited-
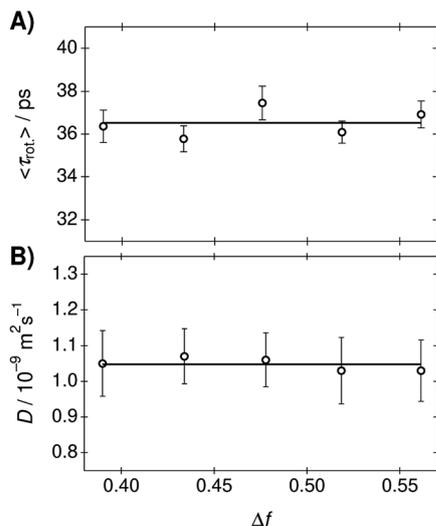
Fig. 4 (A) Rotational correlation times and (B) translational diffusion coefficients of **C153** in the **PA/BuCN** mixtures as a function of Δf. The lines indicate the mean values in the mixtures.

Fig. 5 Steady-state absorption spectra of **C$_4$-dHONI** ($c = 30$ μM) upon addition of (A) **NMI** and (C) **DBU** in a **PA/BuCN** solvent mixture ($\Delta f = 0.48$). Note the different wavelength scales in (A) and (C). Logarithmic association or deprotonation constants (B) with **NMI** and (D) **DBU** in all mixtures as a function of Δf. The lines in (B) represent the mean values of the association constants whereas in (D) they simply connect the data points.

state proton transfer from a photoacid to organic bases. Therefore, it is crucial that the mixture does not disrupt the HB equilibrium between the reaction partners. To demonstrate this, we determined the association constants between **C$_4$-dHONI** photoacid and a weak base (**NMI**) from a titration experiment. **NMI** forms hydrogen bonds to the hydroxyl groups but is not strong enough to deprotonate the photoacid in the ground state. On the other hand, a strong base (**DBU**) will deprotonate one of the hydroxyl groups already in the ground state.[44] Therefore, the impact of dielectric stabilization on the deprotonation constant can be demonstrated in the presence of a strong base.

Since **C$_4$-dHONI** has two binding sites, a model based on 1 : 2 host–guest stoichiometry was used. The association constants, K_1 and K_2, were obtained from the steady-state absorption spectra using a previously described global analysis procedure (see ESI† for details).[44] Representative absorption spectra of **C$_4$-dHONI** upon addition of **NMI** and **DBU** in a **PA/BuCN** mixture with $\Delta f = 0.48$ are presented in Fig. 5A and C and the association or deprotonation constants as a function of Δf in all the mixtures in Fig. 5B and D.

The absorption spectra of **C$_4$-dHONI** shows a broadening and a red shift upon addition of **NMI**. We have previously shown that **NMI** forms neutral hydrogen-bonded 1:1 and 1:2 complexes with **C$_4$-dHONI**.[44] The association constants, K_1 and K_2, determined from the global analysis remain constant in the mixtures. This clearly demonstrates that the solvent mixtures do not influence the HB equilibrium between the reaction partners. Therefore, the mixtures could be additionally used to investigate the effect of dielectric constant on different HB association reactions.

The absorption spectra upon addition of **DBU** show a significant decrease of the neutral band around 380 nm and an appearance of a long-wavelength band at 450 nm. This has been attributed to a deprotonation of a single hydroxyl group by
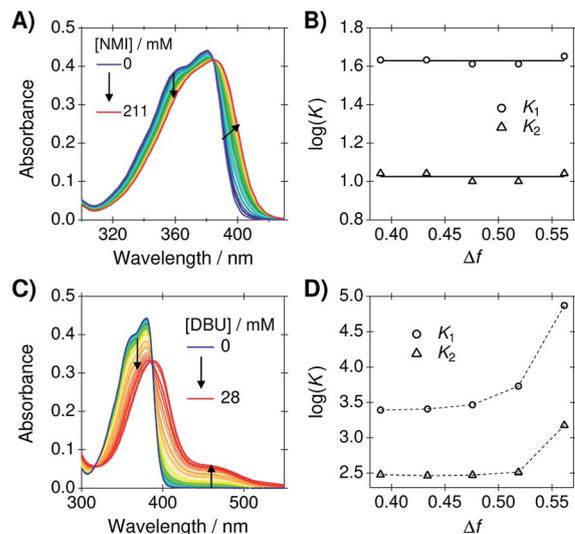
the strong base followed by HB association of another base molecule on the remaining hydroxyl group of **C$_4$-dHONI**. This is due to the fact that deprotonation of the first hydroxyl group significantly increases the pK_a of the second. The deprotonation constants, K_1, increase drastically upon increasing Δf, being 30 times higher in **BuCN** compared to **PA**. This shows that the reaction free energy for proton transfer reactions significantly decreases (more negative ΔG) upon increasing Δf. On the other hand, the association constant for the binding of a second **DBU**, K_2, remains nearly constant in neat **PA** and the mixtures but increases by a factor of four neat **BuCN**. This further demonstrates that the mixtures do not significantly influence HB equilibrium. The increase in **BuCN** might originate from incomplete description of the experimental spectra by the association model because different ion pair species (contact and non-contact) can further complicate the spectral behavior in polar solvents.[44] In any case, the impact of the dielectric stabilization on the ground-state deprotonation constant is very clear. The next step is to investigate the impact on photoinduced excited-state proton-transfer reactions and work in this direction is under way in our laboratory.

Conclusions

We have used a variety of spectroscopic techniques to characterize several properties of propyl acetate/butyronitrile binary mixtures. Refractive index and viscosity of the mixtures is constant whereas the dielectric constant varies linearly with the weight fractions of the solvents. Detailed solvatochromic studies on three push–pull probes demonstrate that the dielectric stabilization is mostly accounted by the bulk dielectric properties and is not complicated by specific solute–solvent

interactions or dielectric enrichment effects. Furthermore, the mixtures do not disrupt hydrogen-bonding equilibrium between the reaction partners. Therefore, the mixtures are ideally suited for investigating the effect of dielectric stabilization on chemical or photochemical reactions. Interestingly, the solvation dynamics are significantly slower in the mixtures as compared to the neat solvents. This additionally enables controlling the time scale of the dielectric stabilization in a predictable manner. Hence the mixtures can be used to investigate both static and dynamic solvent effects on chemical reactivity in a wide variety of systems.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Notes and references

1 *Solvent Effects and Chemical Reactivity*, ed. O. Tapia and J. Bertrán, Kluwer Academic Publishers, Dordrecht, 2002.

2 C. Reichardt, *Solvents and Solvent Effects in Organic Chemistry*, Wiley-VCH Verlag GmbH & Co., Weinheim, 3rd edn, 2003.

3 B. Bagchi, *Annu. Rev. Phys. Chem.*, 1989, **40**, 115–141.

4 M. J. Weaver and G. E. McManis, *Acc. Chem. Res.*, 1990, **23**, 294–300.

5 P. F. Barbara, G. C. Walker and T. P. Smith, *Science*, 1992, **256**, 975–981.

6 H. Heitele, *Angew. Chem., Int. Ed.*, 1993, **32**, 359–377.

7 P. M. Kiefer and J. T. Hynes, *J. Phys. Chem. A*, 2002, **106**, 1834–1849.

8 T. Kumpulainen, B. Lang, A. Rosspeintner and E. Vauthey, *Chem. Rev.*, 2017, **117**, 10826–10939.

9 T. Kumpulainen, A. Rosspeintner, B. Dereka and E. Vauthey, *J. Phys. Chem. Lett.*, 2017, **8**, 4516–4521.

10 E. Tommila, *Acta Chem. Scand.*, 1959, **13**, 1019–1030.

11 E. Niki, Y. Kamiya and N. Ohta, *Bull. Chem. Soc. Jpn.*, 1969, **42**, 3224–3229.

12 H. Kwart and P. A. Silver, *J. Org. Chem.*, 1975, **40**, 3019–3026.

13 R. A. Ransdell and C. C. Wamser, *J. Phys. Chem.*, 1992, **96**, 10572–10575.

14 P. M. Mancini, G. Fortunato, C. Adam, L. R. Vottero and A. J. Terenzani, *J. Phys. Org. Chem.*, 2002, **15**, 258–269.

15 C. A. Rumble and M. Maroncelli, *J. Chem. Phys.*, 2018, **148**, 193801.

16 G. Angulo, M. Brucka, M. Gerecke, G. Grampp, D. Jeannerat, J. Milkiewicz, Y. Mitrev, C. law Radzewicz, A. Rosspeintner, E. Vauthey and P. Wnuk, *Phys. Chem. Chem. Phys.*, 2016, **18**, 18460–18469.

17 J. Gardecki and M. Maroncelli, *Chem. Phys. Lett.*, 1999, **301**, 571–578.

18 G. Angulo, J. Jedrak, A. Ochab-Marcinek, P. Pasitsuparoad, C. Radzewicz, P. Wnuk and A. Rosspeintner, *J. Chem. Phys.*, 2017, **146**, 244505.

19 S. Richert, A. Rosspeintner, S. Landgraf, G. Grampp, E. Vauthey and D. R. Kattnig, *J. Am. Chem. Soc.*, 2013, **135**, 15144–15152.

20 H. M. Hoang, T. B. V. Pham, G. Grampp and D. R. Kattnig, *J. Phys. Chem. Lett.*, 2014, **5**, 3188–3194.

21 D. R. Kattnig, A. Rosspeintner and G. Grampp, *Angew. Chem., Int. Ed.*, 2008, **47**, 960–962.

22 D. McMorrow and M. Kasha, *J. Phys. Chem.*, 1984, **88**, 2235–2243.

23 C. Reichardt, *Chem. Rev.*, 1994, **94**, 2319–2358.

24 O. K. Abou-Zied, R. Jimenez, E. H. Z. Thompson, D. P. Millar and F. E. Romesberg, *J. Phys. Chem. A*, 2002, **106**, 3665–3672.

25 P. Zhou, M. R. Hoffmann, K. Han and G. He, *J. Phys. Chem. B*, 2015, **119**, 2125–2131.

26 J. L. Pérez-Lustres, F. Rodriguez-Prieto, M. Mosquera, T. A. Senyushkina, N. P. Ernsting and S. A. Kovalenko, *J. Am. Chem. Soc.*, 2007, **129**, 5408–5418.

27 P. Suppan, *J. Chem. Soc., Faraday Trans. 1*, 1987, **83**, 495–509.

28 J. G. Dawber, J. Ward and R. A. Williams, *J. Chem. Soc., Faraday Trans. 1*, 1988, **84**, 713–727.

29 N. K. Petrov, *High Energy Chem.*, 2006, **40**, 22–34.

30 M. J. Kamlet, J. L. M. Abboud, M. H. Abraham and R. W. Taft, *J. Org. Chem.*, 1983, **48**, 2877–2887.

31 M. C. Beard, G. M. Turner and C. A. Schmuttenmaer, *J. Phys. Chem. A*, 2002, **106**, 878–883.

32 C. Lerf and P. Suppan, *J. Chem. Soc., Faraday Trans.*, 1992, **88**, 963–969.

33 M. Rosés, C. Ràfols, J. Ortega and E. Bosch, *J. Chem. Soc., Perkin Trans. 2*, 1995, 1607–1615.

34 E. Åkesson, G. C. Walker and P. F. Barbara, *J. Chem. Phys.*, 1998, **95**, 4188–4194.

35 N. E. Levinger, A. E. Johnson, G. C. Walker and P. F. Barbara, *Chem. Phys. Lett.*, 1992, **196**, 159–165.

36 V. Kharlanov and W. Rettig, *J. Phys. Chem. A*, 2009, **113**, 10693–10703.

37 M. L. Horng, J. A. Gardecki, A. Papazyan and M. Maroncelli, *J. Phys. Chem.*, 1995, **99**, 17311–17337.

38 R. Schanz, S. Kovalenko, V. Kharlanov and N. Ernsting, *Appl. Phys. Lett.*, 2001, **79**, 566.

39 L. Zhao, J. L. P. Lustres, V. Farztdinov and N. P. Ernsting, *Phys. Chem. Chem. Phys.*, 2005, **7**, 1716–1725.

40 J. T. Edward, *J. Chem. Educ.*, 1970, **47**, 261–270.

41 G. T. Evans and D. Kivelson, *J. Chem. Phys.*, 1986, **84**, 385.

42 M. L. Horng, J. A. Gardecki and M. Maroncelli, *J. Phys. Chem. A*, 1997, **101**, 1030–1047.

43 T. Kumpulainen, B. H. Bakker, M. Hilbers and A. M. Brouwer, *J. Phys. Chem. B*, 2015, **119**, 2515–2524.

44 T. Kumpulainen, B. H. Bakker and A. M. Brouwer, *Phys. Chem. Chem. Phys.*, 2015, **17**, 20715–20724.

45 Y. Marcus, *The Properties of Solvents*, Wiley, Chichester, 1st edn, 1998.

46 J. A. Gardecki and M. Maroncelli, *Appl. Spectrosc.*, 1998, **52**, 1179–1189.

47 A. Morandeira, L. Engeli and E. Vauthey, *J. Phys. Chem. A*, 2002, **106**, 4833–4837.

48 G. Duvanel, J. Grilj, H. Chaumeil, P. Jacques and E. Vauthey, *Photochem. Photobiol. Sci.*, 2010, **9**, 908–915.

49 X.-X. Zhang, C. Würth, L. Zhao, U. Resch-Genger, N. P. Ernsting and M. Sajadi, *Rev. Sci. Instrum.*, 2011, **82**, 063108.

50 M. Gerecke, G. Bierhance, M. Gutmann, N. P. Ernsting and A. Rosspeintner, *Rev. Sci. Instrum.*, 2016, **87**, 053115.

51 G. Angulo, G. Grampp and A. Rosspeintner, *Spectrochim. Acta, Part A*, 2006, **65**, 727–731.

52 R. D. B. Fraser and E. Suzuki, *Anal. Chem.*, 1969, **41**, 37–39.

53 P. F. Rusch and J. P. Lelieur, *Anal. Chem.*, 1973, **45**, 1541–1543.

54 V. G. Machado, R. I. Stock and C. Reichardt, *Chem. Rev.*, 2014, **114**, 10429–10475.

55 L. Onsager, *J. Am. Chem. Soc.*, 1936, **58**, 1486–1493.

56 E. Lippert, *Z. Naturforsch., A: Phys. Sci.*, 1955, **10**, 541–545.

57 N. Mataga, Y. Kaifu and M. Kolzumi, *Bull. Chem. Soc. Jpn.*, 1956, **29**, 465–470.

58 T. Kumpulainen, A. Rosspeintner and E. Vauthey, *Phys. Chem. Chem. Phys.*, 2017, **19**, 8815–8825.

59 R. Jimenez, G. R. Fleming, P. V. Kumar and M. Maroncelli, *Nature*, 1994, **369**, 471–473.

60 X.-X. Zhang, M. Liang, N. P. Ernsting and M. Maroncelli, *J. Phys. Chem. B*, 2013, **117**, 4291–4304.

61 R. S. Fee and M. Maroncelli, *Chem. Phys.*, 1994, **183**, 235–247.

62 G. van der Zwan and J. T. Hynes, *J. Phys. Chem.*, 1985, **89**, 4181–4188.

63 F. Cichos, A. Willert, U. Rempel and C. von Borczyskowski, *J. Phys. Chem. A*, 1997, **101**, 8179–8185.

64 B. M. Luther, J. R. Kimmel and N. E. Levinger, *J. Chem. Phys.*, 2002, **116**, 3370.

65 R. L. Christensen, R. C. Drake and D. Phillips, *J. Phys. Chem.*, 1986, **90**, 5960–5967.

66 C. Qiu and G. J. Blanchard, *J. Phys. Chem. B*, 2015, **119**, 1986–1993.