


Cite this: *RSC Adv.*, 2020, **10**, 28786

Magnetic hyperthermia with ϵ -Fe₂O₃ nanoparticles†

Yuanyu Gu,^{ab} Marie Yoshikiyo,^c Asuka Namai,^c Debora Bonvin,^d Abelardo Martinez,^e Rafael Piñol,^b Pedro Téllez,^f Nuno J. O. Silva,^g Fredrik Ahrentorp,^h Christer Johansson,^h Joaquín Marco-Brualla,ⁱ Raquel Moreno-Loshuertos,ⁱ Patricio Fernández-Silva,ⁱ Yuwen Cui,^a Shin-ichi Ohkoshi^c and Angel Millán^{id}*^b

Biocompatibility restrictions have limited the use of magnetic nanoparticles for magnetic hyperthermia therapy to iron oxides, namely magnetite (Fe₃O₄) and maghemite (γ -Fe₂O₃). However, there is yet another magnetic iron oxide phase that has not been considered so far, in spite of its unique magnetic properties: ϵ -Fe₂O₃. Indeed, whereas Fe₃O₄ and γ -Fe₂O₃ have a relatively low magnetic coercivity, ϵ -Fe₂O₃ exhibits a giant coercivity. In this report, the heating power of ϵ -Fe₂O₃ nanoparticles in comparison with γ -Fe₂O₃ nanoparticles of similar size (~20 nm) was measured in a wide range of field frequencies and amplitudes, in uncoated and polymer-coated samples. It was found that ϵ -Fe₂O₃ nanoparticles primarily heat in the low-frequency regime (20–100 kHz) in media whose viscosity is similar to that of cell cytoplasm. In contrast, γ -Fe₂O₃ nanoparticles heat more effectively in the high frequency range (400–900 kHz). Cell culture experiments exhibited no toxicity in a wide range of nanoparticle concentrations and a high internalization rate. In conclusion, the performance of ϵ -Fe₂O₃ nanoparticles is slightly inferior to that of γ -Fe₂O₃ nanoparticles in human magnetic hyperthermia applications. However, these ϵ -Fe₂O₃ nanoparticles open the way for switchable magnetic heating owing to their distinct response to frequency.

Received 15th May 2020

Accepted 27th July 2020

DOI: 10.1039/d0ra04361c

rsc.li/rsc-advances

Introduction

Magnetic heating with magnetic nanoparticles (MNPs) is an elegant method for non-contact heating that has been implemented in industrial¹ and clinical applications.² The generation

of heat by MNPs in internal parts of the body (mainly tumors) by application of an alternating current (AC) magnetic field in the radio-frequency-range was proposed as a hyperthermia therapy decades ago,³ and it is now in clinical practice^{4,5} in a limited number of European hospitals. Unfortunately, the low heating power achievable is still a strongly limiting factor in these^{6–10} and other similar applications.¹¹

Few nanoparticle (NP) magnetic materials are trusted by health authorities for in-body use, where only iron oxides, and particularly maghemite (γ -Fe₂O₃), are generally accepted for hyperthermia cancer therapy. Although many other materials present advantages in terms of heating power, their safety is not yet guaranteed. Thus, the only viable opportunity to enhance clinical hyperthermia performance has been the optimization of γ -Fe₂O₃ NP structural features, such as crystallinity, size, shape and state of aggregation.^{14–16} Another magnetic iron(III) oxide, ϵ -Fe₂O₃, that could open new perspectives in the field was first prepared in the lab in 1934 by Forestier and Guiot-Guilain.¹⁷ But its exceptional magnetic properties remained unexplored until 2004 when it was prepared as a pure phase by Jin *et al.*,¹⁸ who showed that the magnetic behaviour of this phase differs drastically from the rest of iron oxides, in that it has a gigantic coercivity. Based on the development of reliable synthesis methods for the fabrication of pure ϵ -Fe₂O₃ NPs by

^aSchool of Materials Science and Engineering, Nanjing Tech University, 210009, Nanjing, PR China

^bInstituto de Ciencia de Materiales de Aragón, ICMA-CSIC University of Zaragoza, C/ Pedro Cerbuna 10, 50006, Zaragoza, Spain. E-mail: angel.millan@csic.es

^cDepartment of Chemistry, School of Science, The University of Tokyo, 7-3-1 Hongo, Bunkyo-ku, Tokyo, 113-0033, Japan

^dPowder Technology Laboratory, Institute of Materials, Ecole Polytechnique Fédérale de Lausanne, 1015 Lausanne, Switzerland

^eDepartamento de Electrónica de Potencia, I3A Universidad de Zaragoza, 50018 Zaragoza, Spain

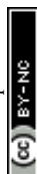
^fServicio de Apoyo a la Investigación, University of Zaragoza, C/ Pedro Cerbuna 10, 50006 Zaragoza, Spain

^gDepartamento de Física, CICECO-Aveiro Institute of Materials, Universidade de Aveiro, 3810-193 Aveiro, Portugal

^hRISE Research Institutes of Sweden, 411 33 Göteborg, Sweden

ⁱDepartamento de Bioquímica, Biología Molecular y Celular, Instituto de Biocomputación y Física de Sistemas Complejos, University of Zaragoza, C/ Pedro Cerbuna 10, 50006 Zaragoza, Spain

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra04361c



some of the authors of this report,^{18–23} we have investigated for the first time the utility of these NPs in hyperthermia therapy.

The thermal power density of MNPs under an applied AC magnetic field is the energy dissipated in a hysteresis cycle multiplied by the frequency. Depending on the intensity of the applied magnetic field, H , and the coercivity, H_c , of the NPs, the magnetization will revert by rotation of magnetic moment (*i.e.*, Néel relaxation²⁴) or by rotation of the entire NP (*i.e.*, Brownian relaxation²⁵). Each type of magnetization reversal has a different response to the frequency. Moreover, the choice of frequencies, f , and field intensities, H for human use are limited owing to safety considerations, and therefore hyperthermia studies should cover a wide range of frequencies. The acceptable field limits vary with the magnet coil diameter and configurations. Estimates resulting from human subjective impressions using a one-turn coil around the chest with a diameter of 30 cm fixed the limit for that configuration at $Hf = 5 \times 10^8 \text{ Am}^{-1} \text{ s}^{-1}$.¹² Later, a limit of $Hf = 5 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$ was established for smaller loops,¹³ and a limit of $Hf = 18 \times 10^8 \text{ Am}^{-1} \text{ s}^{-1}$ was proposed for gap magnets.⁴ The question then arises regarding the best strategy to improve the performance of clinical magnetic hyperthermia: using high frequencies and low fields, or using low frequencies and high fields.

In this report we measured and compared the heating power of $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NPs with a similar particle size ($\sim 20 \text{ nm}$). A wide range of frequencies (20–900 kHz) and field amplitudes ($4\text{--}95 \text{ kA m}^{-1}$) were used to determine the optimal field conditions for the application. The measurements were carried out at various NP mobility conditions: in pure water, under cell cytoplasm viscosity and at complete mobility restriction. We also studied the influence of a particle polymer coating on the heating power. Finally, some conclusions about the utility of $\epsilon\text{-Fe}_2\text{O}_3$ NPs, and the optimal frequency range for clinical hyperthermia therapy were drawn.

Experimental

Synthesis

Block copolymer P4VP-*b*-P(MPEGA-*co*-RhodPEGMA-*co*-carboxylicPEGMA) (poly(4-vinylpyridine)-*block*-poly(methoxypoly ethylenglycolacrylate-*co*-Rhodamine polyethylenglycolmethacrylate-*co*-carboxylic polyethylenglycolmethacrylate)), used for the coating of iron oxide nanoparticles, was prepared by atom transfer radical polymerization (ATRP) according to methods described elsewhere.²⁶ Details on materials and synthesis procedures are given in ESI† A scheme of the synthesis route is shown in ESI Fig S1.†

$\epsilon\text{-Fe}_2\text{O}_3$ nanoparticles were synthesized by partially arranging the ferrihydrite seed sol-gel method.²⁰ Basically, a precursor of iron oxide hydroxide nanoparticles embedded in silica matrix was prepared by the sol-gel technique; tetraethyl orthosilicate (TEOS) was added to aqueous dispersion of iron oxide hydroxide nanoparticles, to form silica by the hydrolysis process. Then, the precursor was sintered in air to form $\epsilon\text{-Fe}_2\text{O}_3$ nanoparticles embedded in silica matrix. Finally the silica matrix was etched by NaOH treatment and washed with water several times, and the obtained NPs were dispersed in

tetramethylammonium aqueous solution using supersonic waves to obtain a stable basic NP suspension.

$\gamma\text{-Fe}_2\text{O}_3$ nanoparticles were synthesized following a protocol modified from Bonvin *et al.* described previously.²⁷ Briefly, $\gamma\text{-Fe}_2\text{O}_3$ nanoparticles were synthesized by co-precipitation in combination with a hydrothermal treatment performed at 120°C for 15 h.

The NPs were coated with the P4VP-*b*-P(MPEGA-*co*-RhodPEGMA-*co*-carboxylicPEGMA) copolymer as described previously.²⁶ Briefly, the uncoated iron oxide nanoparticles are dispersed in slightly acidic medium ($\text{pH} = 2$) and mixed with a polymer solution at the same pH. At this pH, the P4VP block is hydrophilic, and, as the pH is increased to 7.4, it becomes hydrophobic encapsulating the nanoparticles. Finally, the suspension is filtered through a $0.22 \mu\text{m}$ membrane filter to obtain the final ferrofluid. The final iron oxide concentrations in the $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NP suspension samples were $4.0 \text{ g(Fe}_2\text{O}_3 \text{ per l)}$ and $3.86 \text{ g(Fe}_2\text{O}_3 \text{ per l)}$, respectively.

Media for hyperthermia experiments emulating cell cytoplasm viscosity consisted on polyethylene glycol (PEG 8000) aqueous solution. Most of reports on cell cytoplasm viscosity indicate values from 1.2 to 1.5 times that of water.^{28–32} However, some authors report values as high as 10 fold.³³ Considering that the viscosity of water is 0.89 mPa s , at 25°C , media with a viscosity 1.9 and 10 times that of water have been obtained from 3 wt% and 10 wt% PEG solutions in water^{34,35} with a viscosities of 1.68 mPa s and 8.9 mPa s , respectively. NPs dispersions in these media were prepared by dissolving under sonication 30 mg of PEG8000 in $970 \mu\text{l}$ of $\epsilon\text{-Fe}_2\text{O}_3$ NP suspension ($7.0 \text{ mg(Fe}_2\text{O}_3 \text{ per ml)}$), and by dissolving 100 mg of PEG8000 in $900 \mu\text{l}$ $\epsilon\text{-Fe}_2\text{O}_3$ ($c = 7.0 \text{ mg ml}^{-1}$).

SAR calculations

Specific absorption rate (SAR) values of NPs' suspensions were measured on self-designed and self-made equipment. 2 ml of sample and 2 ml of water were placed in the magnet gap, and their corresponding temperatures were measured by two GaAs fiber optic temperature sensors (OptoCon), which were inserted into the liquids and connected to a fiber optic temperature monitoring system. When the temperatures of the sample and the water were stable, they started to be recorded: (i) 300 s with the field off, (ii) 30 s with the field on, and (iii) 300 s with the field off. The SAR values were extracted from the $T(t)$ curves (Fig. 1), by using the equation:

$$\text{SAR} = \frac{C_p(\text{H}_2\text{O})}{c(\text{Fe}_2\text{O}_3)} \frac{d(\Delta T)}{dt} \quad (1)$$

where $C_p(\text{H}_2\text{O})$ is the heat capacity of water, $c(\text{Fe}_2\text{O}_3)$ is the concentration of iron oxide NPs, ΔT represents the temperature difference between suspension of iron oxide NPs and water reference, and $\frac{d(\Delta T)}{dt}$ stands for the initial slop of heating curves obtained by fitting the experimental measurements to a second order polynomial (see Fig. 1).

Transmission electron microscope (TEM) observations were carried in a JEOL 2000-FXII microscope on carbon coated



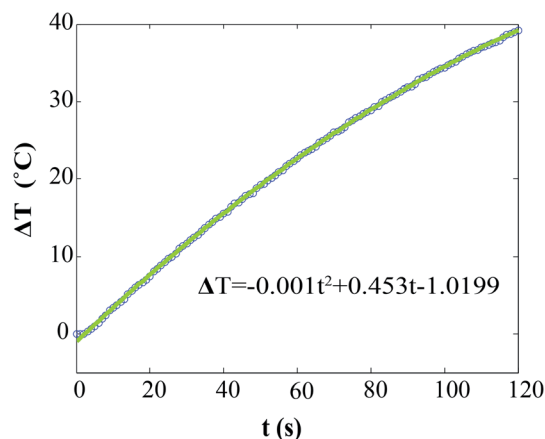


Fig. 1 Heating curves of ϵ -Fe₂O₃ in aqueous suspension exposed to an AC field (101 kHz, 51 kA m⁻¹). The line corresponds to a fitting to a second order polynomial. The y-axis corresponds to the temperature difference between the NPs suspension sample and the control pure water sample placed in the ferrite magnet gap.

copper grids after dip coating of the grids in the ferrofluid samples. Dynamic light scattering (DLS) measurements were performed in Zetasizer Nano ZS from Malvern Laser. Chemical analysis of iron content in the samples was carried out in by coupled plasma atomic emission spectrometry (ICP-AES).

The 2θ - θ scan X-ray powder diffraction (XRD) measurements were performed using Rigaku Ultima IV with Cu K α radiation ($\lambda = 1.5418$ Å). Rietveld analyses were performed using the PDXL program of RIGAKU.

Magnetic measurements of samples were carried out on a superconducting quantum interference device (SQUID)-based magnetometer MPMS-XL5 from Quantum Design. AC magnetic susceptibility measurements *versus* temperature were carried out in a 10 K to 300 K temperature range, and magnetization *vs.* field measurements at 5 K and 300 K were carried out in a -500 Oe to 50 000 Oe field range. AC magnetic susceptibility *vs.* frequency measurements at 300 K were carried out in a 1 Hz to 500 kHz frequency range, using the DynoMag AC susceptometer (RISE Research Institutes of Sweden).

SAR measurements at several amplitudes and frequencies of the magnetic field were carried out in a homemade magnetic heating source²⁶ consisting of a signal generator, a high power amplifier and a matching transformer connected a *RCL* circuit. The magnetic field was produced in between the gap of a ferrite nucleus with Litz wires windings. The field intensity and frequency during the measurements were varied in the ranges 8–92 kA m⁻¹ and 25 to 100 kHz respectively. The measurements were performed in aqueous suspensions of uncoated and coated ϵ -Fe₂O₃ NPs, uncoated and coated γ -Fe₂O₃ NPs and agar-agar gels of these suspensions.

Cell experiments

For these experiments, MDA-MB468 breast cancer cell line was purchased from Leibniz Institute and grown in DMEM medium (GIBCO) supplemented with 10% FBS (GIBCO) and with penicillin/streptomycin (GIBCO).

About 10 000 cells were seeded in 96-well plates 24 hours prior to the treatments with ferrofluids. Then, the cells were incubated with TNPs for 24 hours, with different concentrations of ϵ -Fe₂O₃ and γ -Fe₂O₃ (0.1, 0.2, 0.5 and 1.0 mg Fe₂O₃ per ml). As a control, MDA-MB-468 cells were cultured in the absence of NPs and prepared under the same conditions. Cells were collected by trypsinization and NPs incorporation and cytotoxicity were analyzed by flow cytometry in a FACSCalibur flow cytometer (BD Biosciences).

NPs cytotoxicity was evaluated by Annexin-V binding assay. Briefly, NPs treated cells were stained for 20 min at room temperature in the darkness with Annexin-Dy634, which binds to the phosphatidylserine exposed in the cell surface, in annexin-binding buffer (140 mM NaCl, 2.5 mM CaCl₂, 10 mM HEPES/NaOH, pH 7.4). Cell suspension was diluted to 200 μ l with the corresponding buffer and analyzed by flow cytometry.

In order to evaluate the internalization capacity of the NPS, rhodamine fluorescence was measured by cytometry in the same samples.

The effect of NPs on the viability and cell growth of cells was evaluated using the MTT reduction assay according to Mosmann *et al.*³⁶ Briefly, cells were seeded and treated with ferrofluids in the same way as explained above. After 24 hours of culture with NPs, the medium is removed and the cells are washed to eliminate NPs in suspension. Then each well was mixed with 10 μ l of a MTT dye solution (3-(4,5-dimethylthiazol-2-yl)-2,5 diphenyltetrazolium bromide, 5 mg ml⁻¹ in PBS). Intact cells contain mitochondrial dehydrogenases that can reduce the yellowish water-soluble MTT to insoluble purple formazan crystals, while dead cells do not produce this reduction. After 2–3 h of incubation, all formed crystals were centrifuged and solubilized in isopropanol. Finally, the absorbance of each well was measured in a microplate reader at 550 nm and compared to that of untreated cells. A reduction in absorbance reveals a reduced number of living cells.

Results and discussion

Structural characterization of the nanoparticles

The ϵ -Fe₂O₃ and γ -Fe₂O₃ NPs were characterized by DLS, TEM and XRD (Fig. 2). Rietveld analysis of the XRD pattern indicates a pure ϵ -Fe₂O₃ phase (orthorhombic, space group *Pna2*₁) (Fig. 2(a) and (b)). A comparison of the sample pattern with those of α -Fe₂O₃ and γ -Fe₂O₃ confirms the absence of these phases in the sample (ESI Fig. S2†). The crystallite sizes calculated by the Scherrer formula were 20.5 and 17.4 nm, for the ϵ -Fe₂O₃ NPs and γ -Fe₂O₃ NPs, respectively. The DLS results (Fig. 2(c) and (d)) indicated that the hydrodynamic diameters, D_H , of the uncoated NPs were 18 and 27 nm for ϵ -Fe₂O₃ and γ -Fe₂O₃ NPs, respectively, which increased after coating to 29 and 36 nm, respectively. The TEM images of the ϵ -Fe₂O₃ and γ -Fe₂O₃ iron oxide-copolymer NPs (Fig. 2(e) and (f), respectively) exhibited a mix of rectangular and hexagonal NPs with a mean size D_p (standard deviation: SD) of 19.1 (5.3) nm for ϵ -Fe₂O₃ NPs and D_p (SD) = 18.3 (7.3) nm γ -Fe₂O₃ NPs, respectively. Histograms of the particle size distributions derived from the TEM



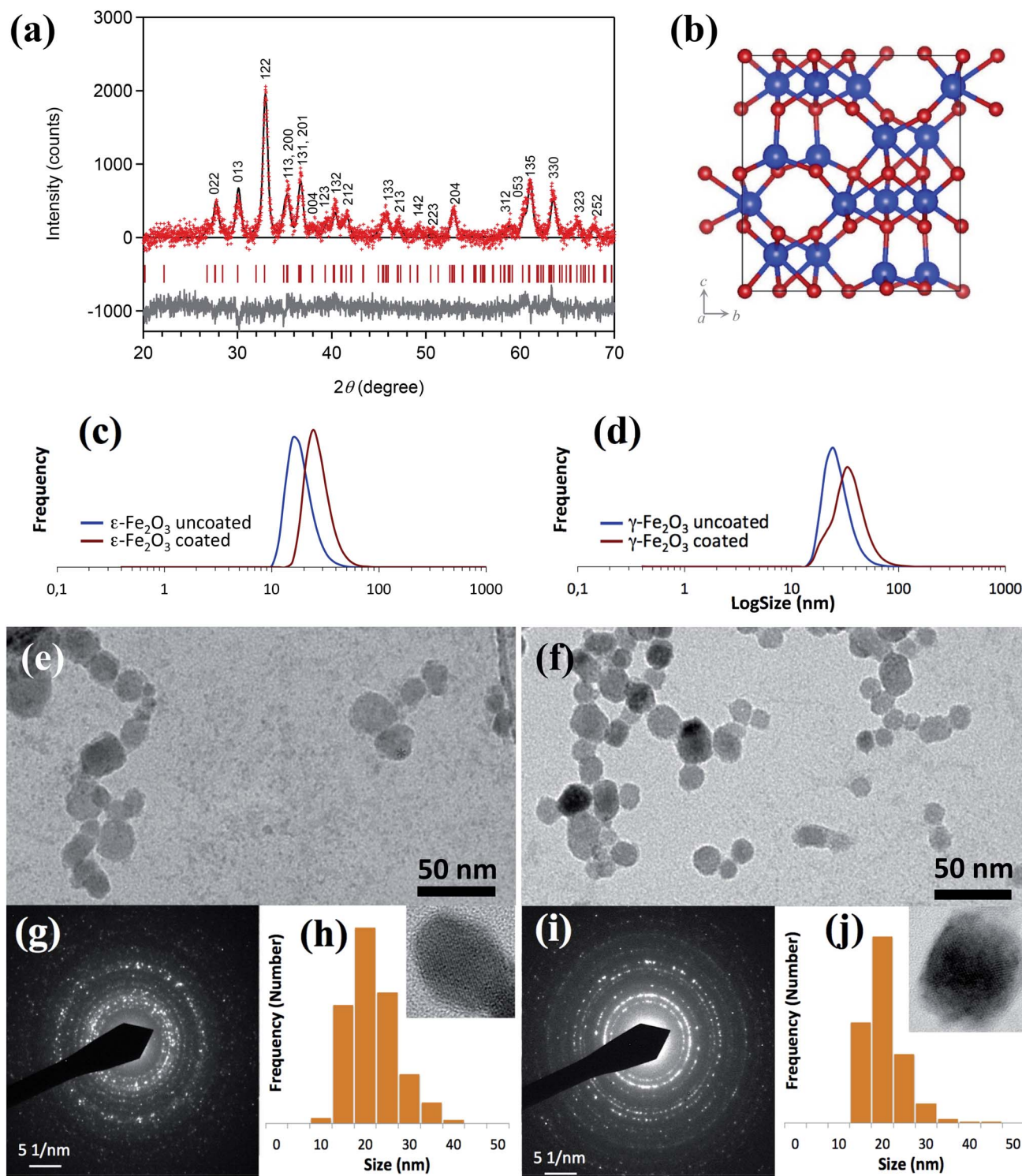


Fig. 2 (a) XRD pattern with Rietveld analysis of ϵ -Fe₂O₃ NPs. Red dots, black lines, and grey lines are the observed patterns, calculated patterns, and their differences, respectively. Red bars represent the calculated positions of the Bragg reflections of the ϵ -Fe₂O₃ phase (orthorhombic, *Pna*2₁). (b) Crystal structure of ϵ -Fe₂O₃. Blue and red balls indicate Fe and O atoms, respectively. (c) Distribution of hydrodynamic diameters, D_{H} , from dynamic light scattering (DLS) measurements of coated and uncoated ϵ -Fe₂O₃ NPs and (d) γ -Fe₂O₃ NPs. (e) TEM images of ϵ -Fe₂O₃ NPs and (f) γ -Fe₂O₃ NPs. (g) ED patterns of ϵ -Fe₂O₃ NPs. (h) Particle size histograms from TEM images of ϵ -Fe₂O₃ NPs, in the inset HRTEM image of a single NP. (i) ED patterns of γ -Fe₂O₃ NPs. (j) Particle size histograms from TEM images of γ -Fe₂O₃ NPs. In the inset, HRTEM images of a single NP.

images of the ϵ -Fe₂O₃ and γ -Fe₂O₃ iron oxide-copolymer NPs are shown in Fig. 2(h) and (j), respectively. The crystalline structure of the ϵ -Fe₂O₃ and γ -Fe₂O₃ NP samples was also established

from electron diffraction (ED) patterns (Fig. 2(g) and (i), respectively). Finally, a detailed structural characterization of these two NP types can be found in Ohkoshi *et al.*^{18–23} in the case

of $\epsilon\text{-Fe}_2\text{O}_3$ NPs, and in Bonvin *et al.*^{14,27} in the case of $\gamma\text{-Fe}_2\text{O}_3$ NPs.

Magnetic properties of the nanoparticles

The different magnetic characters of the $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NPs were clearly reflected in the $M(H)$ measurements at different temperatures. The hysteresis cycles of $\epsilon\text{-Fe}_2\text{O}_3$ NPs suspensions at a temperature well below the freezing point (*i.e.*, 200 K) exhibited a huge coercivity (Fig. 3(a), Table 1). The coercivity was considerable reduced when the temperature approached the water melting point, although still well above the higher field amplitude used in SAR measurements. On the contrary, the coercivity in $\gamma\text{-Fe}_2\text{O}_3$ NPs hysteresis cycles (Fig. 3(b), Table 1) was consistently below the SAR field amplitudes (inset in Fig. 3(b)). At 300 K, when the NPs are free to rotate, the coercivity of the suspensions decreased to 0 in both types of NPs.

The AC magnetic susceptibility *vs.* temperature measurements (ESI Fig. S4†) of uncoated $\epsilon\text{-Fe}_2\text{O}_3$ NPs suspension exhibit a peak in both $\chi'(T)$ and $\chi''(T)$ between 80 and 120 K. Both of the $\chi'(T)$ and $\chi''(T)$ peaks do not exhibit the typical frequency dependence of the Néel relaxation process. An anomaly in this temperature range was also observed previously in Mössbauer measurements, and was attributed to a structural transformation and possible spin reorientation effects.¹² Apart from this peak, $\chi''(T)$ remains zero in the entire 10–260 K temperature range, implying that no Néel relaxation processes are present in this range. At around 260 K the liquid begins to thaw, allowing free rotation of the NPs. Simultaneously, the χ'' peak increases abruptly in a frequency-dependent manner, which can be attributed to Brownian relaxation.

SAR measurements

The SAR values of the $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NPs were measured in a wide range of field frequencies using two different sets of equipment. Fig. 4(a) and (b) show the variation of SAR value with the safety parameter (Hf) for the $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NPs, respectively. Whereas for $\gamma\text{-Fe}_2\text{O}_3$ NPs all of the SAR values fit to a single straight line; those of $\epsilon\text{-Fe}_2\text{O}_3$ NPs were higher at frequencies from 25–61 kHz, being optimal at 45 kHz. It is evident that the heating power is very low at the safety limits proposed for wide coils (*i.e.*, $Hf = 5 \times 10^8 \text{ Am}^{-1} \text{ s}^{-1}$)¹² for both types of NPs herein. Extrapolating to the limit proposed for small coils (*i.e.*, $Hf = 5 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$)¹³ the SAR of $\epsilon\text{-Fe}_2\text{O}_3$ NPs was $80 \text{ W g}^{-1} \text{ Fe}_2\text{O}_3$ (at 75 kHz), and therefore inferior to the value obtained for $\gamma\text{-Fe}_2\text{O}_3$ NPs ($160 \text{ W g}^{-1} \text{ Fe}_2\text{O}_3$). In the high-frequency regime the different magnetic characters of the two types of NPs were evident (Fig. 4(c) and (d)). Within the safety limit, the performance of both types of NPs diminished with respect to that observed in the low-frequency regime. However, the diminished performance was especially acute for $\epsilon\text{-Fe}_2\text{O}_3$ NPs, whose SAR values dropped one order of magnitude. This performance disparity caused the difference in SAR value between the two phases to be about 15-fold. Conversely, although the highest absolute value of SAR in both cases was obtained at $f = 710 \text{ kHz}$, the best performance in terms of the

safety parameter was obtained at the lowest frequency (419 kHz). It should be noted that most of the experimental points in the high-frequency region lay outside the health safety limit adopted in this report (*i.e.*, $Hf = 5 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$). An interesting fact derived from these experiments is that, in the case of a suspension containing a mixture of $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NPs, a change from low to high frequency could switch off the heating *via* the $\epsilon\text{-Fe}_2\text{O}_3$ NPs while increasing that of the $\gamma\text{-Fe}_2\text{O}_3$ NPs. Such a switchable system could be useful, for instance, in catalytic cascade reactions.

Fig. 4(e) and (f) show the variation of SAR with the field amplitude, H , at a fixed f , at different mobility conditions for the $\epsilon\text{-Fe}_2\text{O}_3$ and $\gamma\text{-Fe}_2\text{O}_3$ NPs, respectively. During complete immobilization of the NPs, the SAR value of $\epsilon\text{-Fe}_2\text{O}_3$ NPs falls to 0 in the entire range of field amplitudes (Fig. 4(e)), indicating no heating whatsoever by Brownian relaxation. This should be expected from the high coercivity of these NPs (Fig. 3 and Table 1) that results in a very high degree of thermally-blocked MNPs. In fact, the characteristic Néel relaxation time for 20 nm-diameter $\epsilon\text{-Fe}_2\text{O}_3$ NPs with an anisotropy constant $K = 10^5\text{--}10^6 \text{ J m}^{-3}$ would be around 1000 years (using $\tau_0 = 10^{-10} \text{ s}$).^{18,20} Nevertheless, in media with a viscosity in the range of the reported cell cytoplasm values, the SAR values of the $\epsilon\text{-Fe}_2\text{O}_3$ NPs were similar to that in water, indicating that $\epsilon\text{-Fe}_2\text{O}_3$ NPs are fully useful for hyperthermia therapy. NP mobility in the interior of cells can also be restrained by membrane binding or cell-mediated aggregation. However, a dramatic fall of SAR has only been observed in NPs with a D_H of several hundreds of nm whereas in our case they are around 30 nm.^{37,38} On the other hand, it has been reported that pure magnetomechanical forces induced cell apoptosis and tumor reduction,³⁹ and in this case $\epsilon\text{-Fe}_2\text{O}_3$ NPs could kill cancer cells both by heating and mechanical stress. Experiments on life cells are currently in their way in our lab to sort out this matter.

In the case of $\gamma\text{-Fe}_2\text{O}_3$ NPs, the heating was still appreciable after gelification (Fig. 4(f)), although the SAR values dropped to half those obtained in the liquid state. This indicates, therefore, a contribution from both Néel and Brownian relaxation to the heating *via* $\gamma\text{-Fe}_2\text{O}_3$ NPs. In fact, considering $K = 10^4 \text{ J m}^{-3}$ for $\gamma\text{-Fe}_2\text{O}_3$ (one order of magnitude lower than the K of $\epsilon\text{-Fe}_2\text{O}_3$), NPs with a size of 16 nm would have a Néel relaxation time of 10^{-7} s , while NPs with a size of 35 nm would have a Néel relaxation time of 1000 s. This is indeed in the range of NP sizes of this sample as measured by TEM. The smaller-sized NPs are therefore expected to have no contribution to heating. At intermediate sizes, heating will arise from the Néel relaxation; while larger NPs may only dissipate heat by Brownian relaxation.

NP polymer coating

To be useful in biological applications, MNPs must be endowed with certain biological functionalities; *i.e.*, stability in biological media, hemocompatibility, long blood circulation times or imaging tags. This is realized herein by covering the NPs with adequate biopolymer coatings, which in turn can affect the hyperthermia performance of the NPs. Consequently, we studied the effect of polymer coating on the magnetic heating



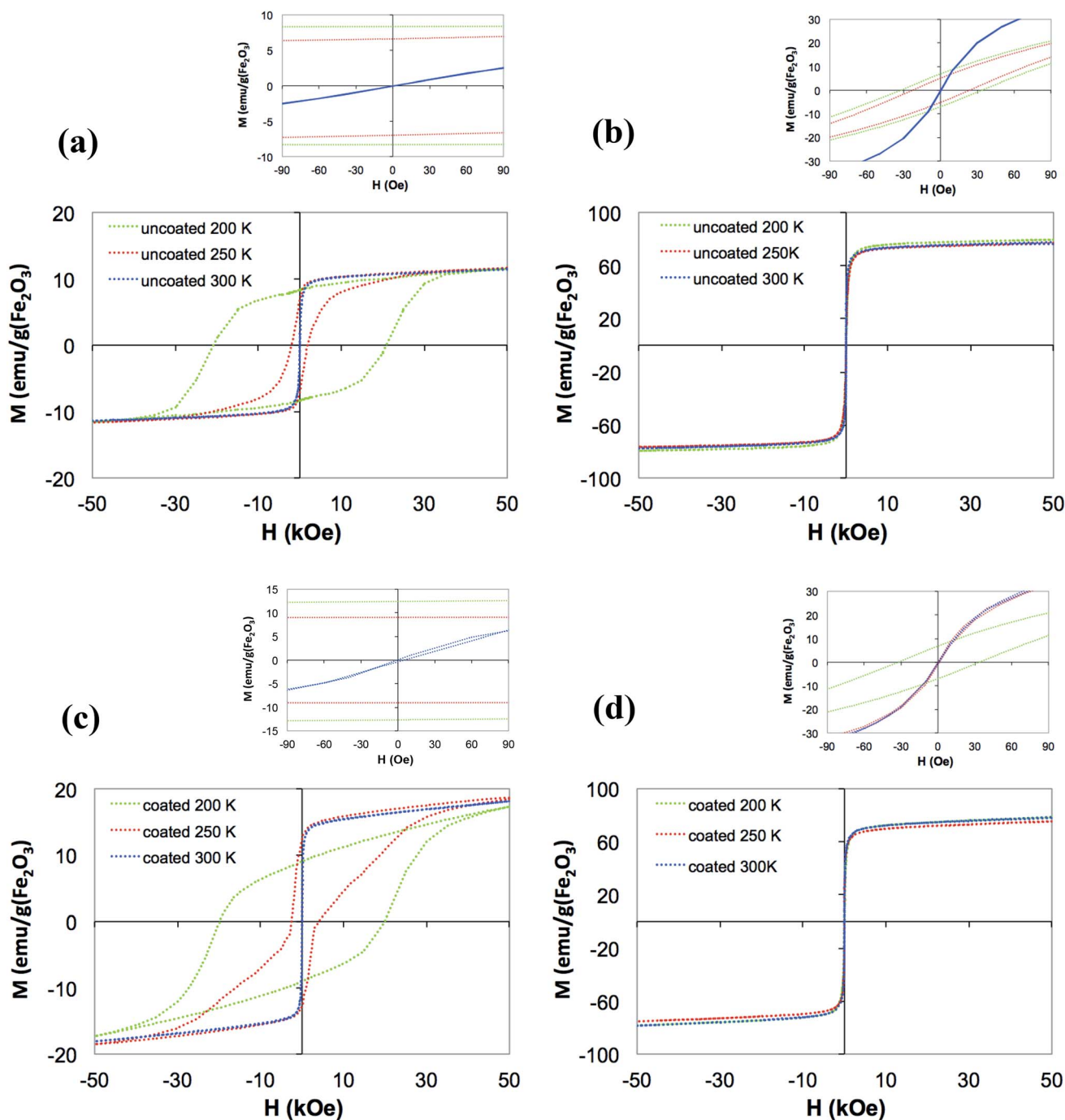


Fig. 3 Magnetization vs. field of water suspension of (a) uncoated and (c) coated ϵ -Fe₂O₃ NP, and (b) uncoated and (d) coated γ -Fe₂O₃ NPs at different temperatures. In the insets, details of the samples magnetization within the range of field amplitudes used in SAR experiments.

properties of both types of NPs. The coating polymer used herein was the P4VP-*b*-P(MPEGA-*co*-RhodPEGMA-*co*-carboxylicPEGMA) copolymer functionalized with a fluorescent tag (Rhodamine), in anticipation of cellular hyperthermia experiments. Moreover, this polymer possesses carboxylate residues at the ends of some of the polyethylene glycol (PEG) side chains that are suitable for conjugation to antibodies with specific binding properties to targeted body tissues (e.g., cancer tumors). Details on the polymer preparation and coating

procedures are given in the ESI.† After being coated with the copolymer, both the ϵ -Fe₂O₃ and γ -Fe₂O₃ NPs were very stable in water suspensions at the physiological pH (7.4). It is worth noting that this type of copolymer coating has been developed in our lab for the last two decades, and it provides excellent biodistribution capacity⁴⁰ and cell compatibility.^{41–45}

As respectively shown in Fig. 5(a) and (b), the polymer coatings reduced the SAR values of the ϵ -Fe₂O₃ and γ -Fe₂O₃ NPs at low frequencies, and especially in the case of γ -Fe₂O₃ NPs. At

Table 1 Coercivity Fields, H_c , and saturation magnetization, M_s , of ϵ - Fe_2O_3 and γ - Fe_2O_3 NPs suspensions, before and after polymer coating, at different temperatures

Phase	Coating	H_c (Oe)			M_s (emu per g Fe_2O_3)		
		200 K	250 K	300 K	200 K	250 K	300 K
ϵ - Fe_2O_3	No	20 500	1900	0	9	10	10
	Yes	20 000	3300	0	10	15	15
γ - Fe_2O_3	No	33	23	0	76	74	74
	Yes	36	19	0	72	69	72

high frequencies, however, the SAR values of the ϵ - Fe_2O_3 NPs dropped drastically after coating (Fig. 5(c)). Specifically, the SAR was practically 0 within the Hf safety range. Conversely, the SAR values of the coated γ - Fe_2O_3 NPs showed only a moderate decrease in SAR value (Fig. 5(d)).

Frequency dependent AC magnetic susceptibility

The power density dissipation assuming linear response theory (*i.e.*, low field amplitudes) is directly related to the out-of-phase susceptibility (*i.e.*, imaginary part), χ'' , by the relation

$$\text{SAR}(f) = \pi\mu_0 H_0^2 f \chi''(f) \quad (2)$$

where μ_0 is the permeability of free space and H_0 is the field amplitude. Thus, we measured the AC susceptibility (ACS; low field amplitude) of ϵ - Fe_2O_3 NPs and γ - Fe_2O_3 (uncoated and coated) MNPs as a function of the frequency in a range from 10 Hz to 500 kHz (Fig. 6). The χ'' vs. $\log(f)$ curves for the uncoated ϵ - Fe_2O_3 NPs exhibited a maximum in χ'' at 15.8 kHz (*i.e.*, Brownian relaxation frequency). In the case of uncoated γ - Fe_2O_3 NPs, the χ'' maximum appeared at about 8 kHz. Obviously, the power density will diminish when shifting the frequency up from this maximum, but an increase in the number of field loop cycles (frequency) may compensate for that loss. Thus, the optimal frequency for heating may deviate from this maximum and the heating power will be favored by the frequency. The absolute values of ACS (both the in-phase and out-of-phase components) were larger for the γ - Fe_2O_3 MNPs than the ϵ - Fe_2O_3 MNPs. Because the particle sizes were almost identical for both samples, the difference in ACS value is likely owing to differences in the intrinsic saturation magnetization between the two samples. The different values of the out-of-phase component will affect the heating properties. All samples exhibited Brownian relaxation as determined by low-field ACS analysis.

The shift of the χ'' peak in the polymer-coated sample with respect to the uncoated one was remarkable in ϵ - Fe_2O_3 NPs (from 15.8 to 1.0 kHz). The consequence is a decrease of the SAR value in the coated sample compared with the uncoated one, as observed in Fig. 5, which is particularly noticeable at high frequencies. However, at the lowest experimental frequencies the ACS of the coated sample is maximal while that of the

uncoated sample is very low, and then the situation may be reversed. Thus, we could expect an enhancement of hyperthermia performance for coated ϵ - Fe_2O_3 samples at frequencies well below the range used in SAR experiments. This large shift in frequency after coating was not observed in the γ - Fe_2O_3 NPs.

From eqn (2) it is possible to estimate SAR from the imaginary part of the AC susceptibility at a given frequency, if the imaginary part of the dynamic magnetization is linear to the applied AC field (*i.e.* low field amplitudes). The low field range also implies that SAR will almost vary with a quadratic Hf behaviour at a constant frequency when assuming small field dependence in the imaginary part of the AC susceptibility. From the SAR results shown in Fig. 4(c) and (d) we have low field amplitudes and the almost quadratic behaviour can be seen, and therefore we can apply eqn (2). If we take the uncoated ϵ - Fe_2O_3 and γ - Fe_2O_3 particles as an example we get for the cases $Hf = 4 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$ and $12 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$ at $f = 542 \text{ kHz}$ (that give fields of 7380 A m^{-1} and $22\,000 \text{ A m}^{-1}$), we obtain $\text{SAR} = 4 \text{ W g}^{-1}$ and 35 W g^{-1} for ϵ - Fe_2O_3 , and 35 W g^{-1} and 315 W g^{-1} for γ - Fe_2O_3 as estimated from the ACS response. The corresponding measured SAR values given in Fig. 4(c) and (d) (for $Hf = 4 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$ and $12 \times 10^9 \text{ Am}^{-1} \text{ s}^{-1}$ at $f = 542 \text{ kHz}$) is 4 W g^{-1} and 33 W g^{-1} for ϵ - Fe_2O_3 and 30 W g^{-1} and 300 W g^{-1} for γ - Fe_2O_3 , which is quite in accordance with the SAR estimations from the ACS results.

Cell experiments

Annexin-V cytotoxicity assays on ϵ - Fe_2O_3 and γ - Fe_2O_3 NPs coated with the rhodamine functionalized copolymer revealed that none of the NPs had an appreciable effect on the viability of MDA-MB468 cells in the entire range of concentrations used in the incubation stage (0.1 to 1 mg ml^{-1}) (Fig. 7(a)). The measurements were single-point, and therefore the differences between the control and NP seeded samples have no statistical significance. As shown in Fig. 7(b) and (c), MDA-MB-468 cells present high rhodamine fluorescence intensity, indicating a high internalization rate for the ϵ - Fe_2O_3 and γ - Fe_2O_3 NPs, respectively. As observed in Fig. 7(c), cells treated with the γ - Fe_2O_3 complex exhibited higher fluorescence intensity than those treated with the ϵ - Fe_2O_3 complex. Specifically, the Rhodamine median fluorescence intensity of γ - Fe_2O_3 was 1.43-, 1.29-, 1.55- and 1.65-fold higher than that of ϵ - Fe_2O_3 for 0.1, 0.2, 0.5 and 1 mg ml^{-1} concentrations, respectively), indicating greater incorporation of the γ - Fe_2O_3 complex.

Although ferrofluids do not seem to induce apoptosis in cells, MTT reduction assay analysis of cell growth inhibition *via* NPs indicated a similar reduction in the growth rate of cells treated with both complexes (Fig. 7(d)). Further, the cell growth rate reduction was concentration-dependent and reached values of around 40% at 0.5 and 1.0 mg ml^{-1} concentration.

Brownian versus Néel heating

It is clear herein that MNPs with a high coercivity such as ϵ - Fe_2O_3 NPs can only heat by the Brownian mechanism. It is also clear that the Brownian and Néel mechanisms operate in different frequency ranges. While the Brownian mechanism



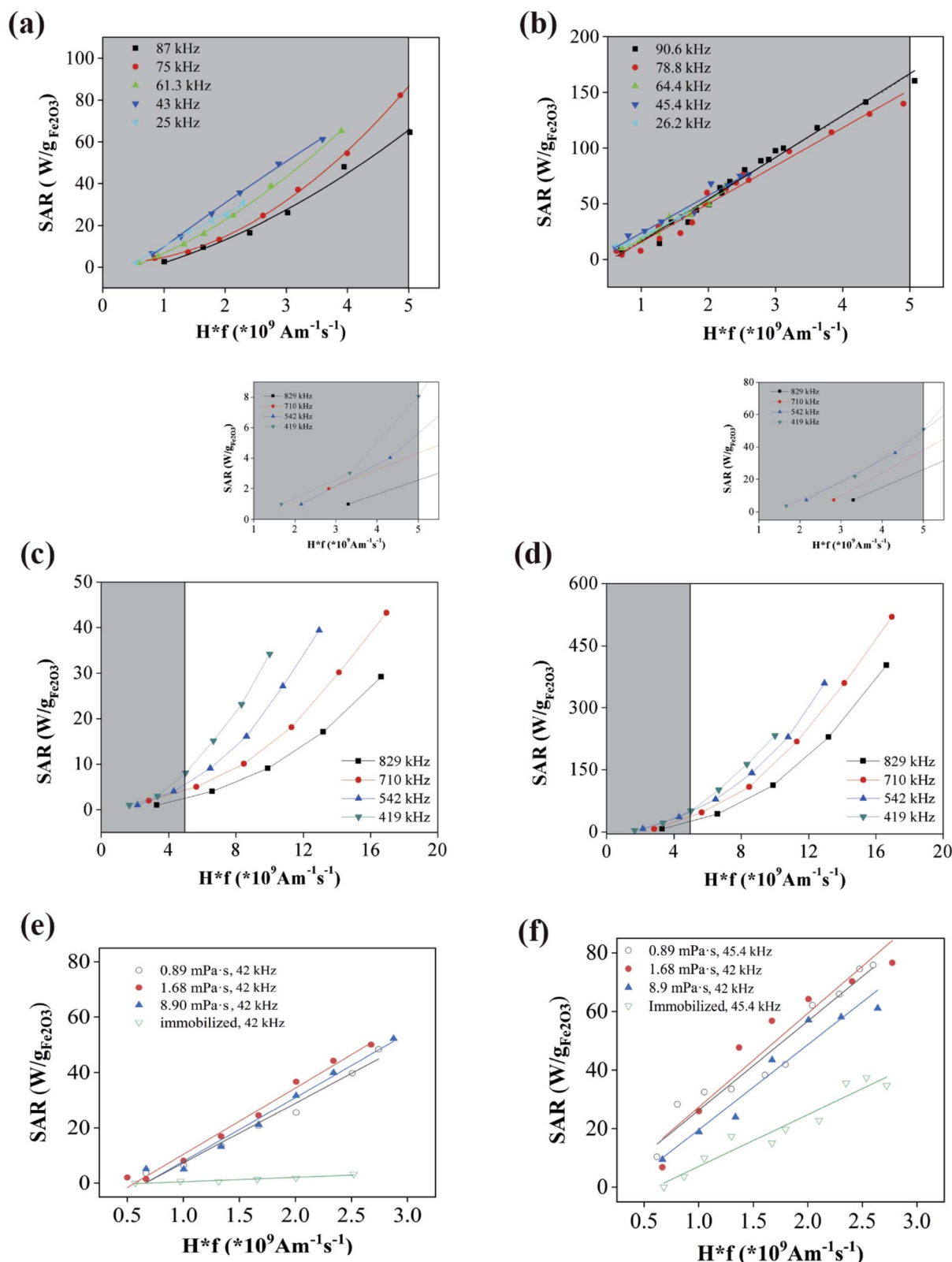


Fig. 4 SAR values in the low frequency range of aqueous suspension of (a) ϵ - Fe_2O_3 NPs and (b) γ - Fe_2O_3 NPs as a function of health parameter Hf . SAR values in the high frequency range of aqueous suspensions of (c) ϵ - Fe_2O_3 NPs and (d) γ - Fe_2O_3 NPs. In the insets, details of the SAR in the health safety range of Hf . Shaded areas mark the health safety region in a–d. SAR vs. Hf at different NP mobility conditions of (e) ϵ - Fe_2O_3 NPs and (f) γ - Fe_2O_3 NPs.

operates below 100 kHz, the Néel mechanism is mostly effective above 400 kHz. Consequently, ϵ - Fe_2O_3 NPs can be efficient in magnetic hyperthermia only at low frequencies. However,

Brownian NPs can be advantageous when the pursued effect is a mechanical one instead of heating. Indeed, it has been suggested in the literature that cell death might also be caused by



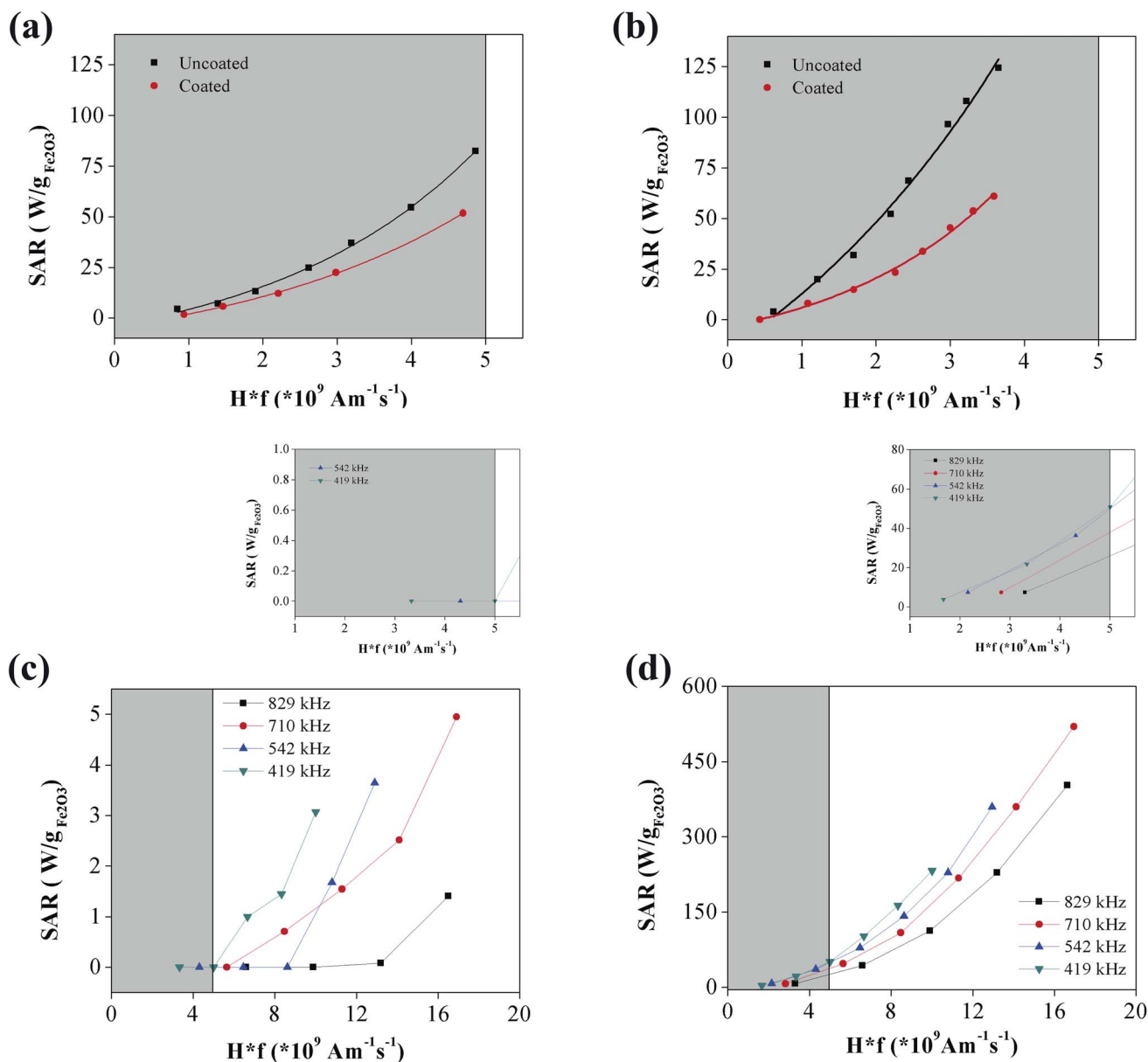


Fig. 5 SAR values in the low frequency range of aqueous suspension of (a) ϵ -Fe₂O₃ polymer coated NPs and (b) γ -Fe₂O₃ polymer coated NPs as a function of health parameter Hf . SAR values in the high frequency range of aqueous suspensions of (c) ϵ -Fe₂O₃ polymer coated NPs and (d) γ -Fe₂O₃ polymer coated NPs. In the insets, details of the SAR in the health safety range of Hf . Shaded areas mark the health safety region in (a–d).

mechanical effects.^{46–50} Moreover, there is increasing interest in using magnetic forces to activate transport through cell membranes.^{51,52} Moreover the strong dependence of magnetic heating on particle mobility can be used as an advantage to heat selectively tumors with a soft texture in a hard healthy tissue environment.

It was observed that the working frequency range is shifted downwards after coating the NPs owing to their increased size. However, the large frequencies shift of the χ'' peak of ϵ -Fe₂O₃ NPs after coating cannot be fully ascribed to the size increase because the size change is relatively small. Another determinant, at least in part, is the strong interactions of the PEG chains with water molecules and the flexibility of these chains.

Owing to the frequent use of a PEG coating of NPs for *in vivo* applications, this subject deserves a deeper insight.

The distinct behavior of Brownian and Néel NPs can be usefully applied toward switchable NP heating systems. For instance, consider a mixture of ϵ -Fe₂O₃ and γ -Fe₂O₃ NPs hosting different substances (*i.e.*, two different catalysts) activated at a distance by the heat generated in the NPs. One could switch-off the activity of the substance hosted by the ϵ -Fe₂O₃ NPs while simultaneously increasing the activity of the substance hosted by the γ -Fe₂O₃ NPs by turning the field from the low-frequency range to the high-frequency range. This is probably the most important outcome of this report, as it opens a new tool in



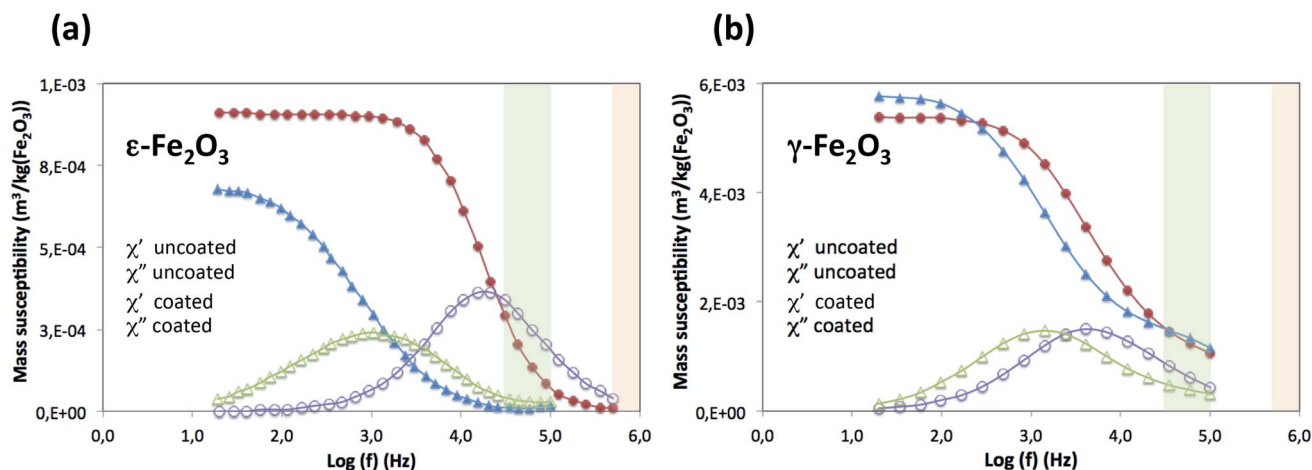


Fig. 6 Variation of the in-phase (χ') and out-of-phase (χ'') magnetic AC susceptibilities with the frequency of the alternating magnetic field of aqueous suspensions of a coated and uncoated ϵ -Fe₂O₃ NPs, and b coated and uncoated γ -Fe₂O₃ NPs. Low and high frequency ranges used in SAR experiments are shaded green and orange, respectively. The ac susceptibility is normalized to total mass of iron(III) oxide (to give the mass susceptibility).

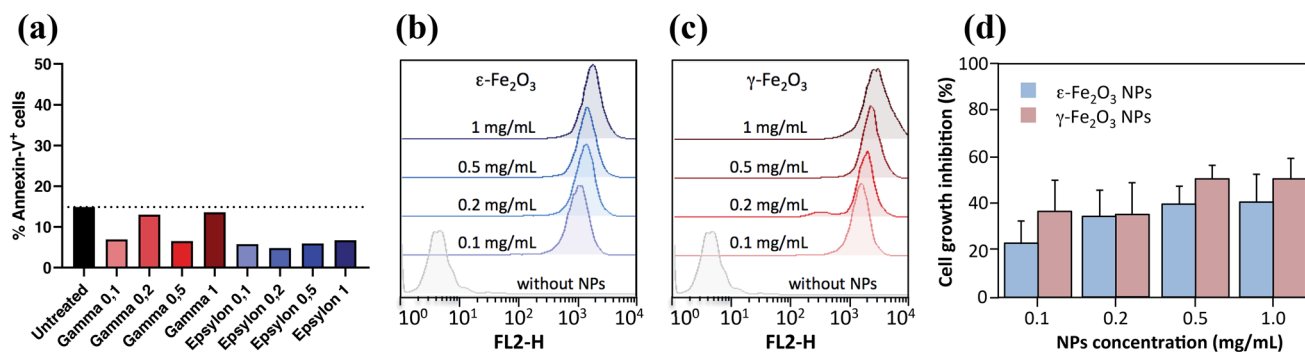


Fig. 7 (a) Annexin-V cell viability plot of MDA-MB468 cells by flow cytometry. Cells were treated with either γ -Fe₂O₃ or ϵ -Fe₂O₃ NPs. (b) Fluorescence intensity of ϵ -Fe₂O₃ and (c) γ -Fe₂O₃ NPs coated with a rhodamine copolymer at the indicated concentrations, determined by flow cytometry. (d) Growth inhibition of MDA-MB468 cells measured by MTT reduction assays after 24 hours of culture in the presence of the indicated concentrations of both types of NPs. Data are expressed as mean \pm S.D. of the mean ($n \geq 3$).

magnetic nano-heating that broadens the scope of applications for these systems.

Conclusions

The utility of ϵ -Fe₂O₃ NPs in magnetic hyperthermia has been evaluated in comparison with γ -Fe₂O₃ NPs of the same particle size. In accordance with their different magnetic natures, these two NP types exhibited very different responses to field parameters in magnetic hyperthermia applications. While γ -Fe₂O₃ NPs possessing a small coercivity achieved a higher heating power at high frequencies (*i.e.*, 400–800 kHz), ϵ -Fe₂O₃ NPs possessing a very large coercivity exhibited a poor heating performance in this frequency range and only operated at low frequencies (*i.e.*, 20–100 kHz). While in γ -Fe₂O₃ NPs possessing a small coercivity the magnetic moment follows the external ac field by rotating internally across magnetocrystalline energy barriers, in ϵ -Fe₂O₃ NPs possessing a very large coercivity the magnetic moment follows the external ac field by the

mechanical rotation of the NPs. As a consequence, γ -Fe₂O₃ NPs achieved a higher heating power at high frequencies (*i.e.*, 400–800 kHz), whereas ϵ -Fe₂O₃ NPs exhibited a poor heating performance in this frequency range and only operated at low frequencies (*i.e.*, 20–100 kHz), at which they relax with the Brownian characteristic time. This difference in behavior opens the possibility of switching the heating from one NP type to the other when both are used simultaneously as heating sources by a simple change of the field frequency.

Author contributions

A. Mi., S. O. and Y. C. conceived and planned the experiments, M. Y., A. N., D. B. and R. P. prepared the samples, M. Y., A. N., S. O., A. Mi., Y. G. and R. P. performed the structural characterization, C. J., F. A., N. J. O. S and Y. G. performed the magnetic measurements, Y. G., A. Ma. and P. T. carried out the SAR measurements, J. M., R. M. and P. F. carried out the cell experiments, A. Mi. and Y. G. wrote the manuscript



with input from all authors, A. Mi., Y. G., C. J., N. J. O. S. and D. B. contributed to the interpretation of the results. All authors provided critical feedback and helped shape the final manuscript.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was supported by European Union's Horizon 2020 FET Open program [Grants no: 801305 and 829162] Spanish Ministry of Science Innovation and Universities [Grant no: PGC2018_095795_B_I00] and Diputación General de Aragón [E11/17R]. Authors would like to acknowledge the use of Servicio General de Apoyo a la Investigación-SAI, Universidad de Zaragoza. This work was developed within the scope of the projects CoolPoint P2020-PTDC-CTMNAN-4511-2014 and CICECO-Aveiro Institute of Materials, UIDB/50011/2020 & UIDP/50011/2020, financed by national funds through the FCT/MEC and co-financed by FEDER under the PT2020 Partnership Agreement. We acknowledge support of the publication fee by the CSIC Open Access Publication Support Initiative through its Unit of Information Resources for Research (URICI). We thank Sara Maccagnano-Zacher, PhD, from Edanz Group (www.edanzediting.com/ac) for editing a draft of this manuscript.

References

- 1 A. Millán, F. Palacio, E. Snoeck, V. Serin and P. Lecante, Magnetic Polymer Nanocomposites, in *Polymer Nanocomposites*, ed. Y.-W. Mai and Z.-Z. Yu, Woodhead Publishing Ltd, Cambridge, 2006, (CRC, Boca Ratón, 2006).
- 2 Q. A. Pankhurst, J. Connolly, S. K. Jones and J. Dobson, Applications of magnetic nanoparticles in biomedicine, *J. Phys. D: Appl. Phys.*, 2003, **36**, 167–181.
- 3 R. K. Gilchrist, *et al.*, Selective inductive heating of lymph nodes, *Ann. Surg.*, 1957, **146**, 596–606.
- 4 B. Thiesen and A. Jordan, Clinical applications of magnetic nanoparticles for Hyperthermia, *Int. J. Hyperthermia*, 2008, **24**, 467–474.
- 5 A. Chiu-Lam and C. Rinaldi, Nanoscale Thermal Phenomena in the Vicinity of Magnetic Nanoparticles in Alternating Magnetic Fields, *Adv. Funct. Mater.*, 2016, **26**, 3933–3941.
- 6 D. Chang, *et al.*, Biologically Targeted Magnetic Hyperthermia: Potential and Limitations, *Front. Pharmacol.*, 2018, **9**, 831.
- 7 M. B. Cortie, D. L. Cortie and V. Timchenko, Heat transfer from nanoparticles for targeted destruction of infectious organisms, *Int. J. Hyperthermia*, 2018, **34**, 157–167.
- 8 S. Dutz and R. Hergt, Magnetic nanoparticle heating and heat transfer on a microscale: Basic principles, realities and physical limitations of hyperthermia for tumour therapy, *Int. J. Hyperthermia*, 2013, **29**, 790–800.
- 9 E. Kita, *et al.*, Ferromagnetic nanoparticles for magnetic hyperthermia and thermoablation therapy, *J. Phys. D: Appl. Phys.*, 2010, **43**, 474011.
- 10 V. Gavrilov-Isaac, *et al.*, Synthesis of Trimagnetic Multishell $\text{MnFe}_2\text{O}_4@\text{CoFe}_2\text{O}_4@\text{NiFe}_2\text{O}_4$ Nanoparticles, *Small*, 2015, **11**, 2614–2618.
- 11 M. B. Cortie, D. L. Cortie and V. Timchenko, Heat transfer from nanoparticles for targeted destruction of infectious organisms, *Int. J. Hyperthermia*, 2018, **34**, 157–167.
- 12 W. J. Atkinson, I. A. Brezovich and D. P. Chakraborty, Usable Frequencies in Hyperthermia with Thermal Seeds, *IEEE Trans. Biomed. Eng.*, 1984, **31**, 70–75.
- 13 R. Hergt and S. Dutz, Magnetic particle hyperthermia-biophysical limitations of a visionary tumour therapy, *J. Magn. Magn. Mater.*, 2007, **311**, 187–192.
- 14 D. Bonvin, *et al.*, Controlling structural and magnetic properties of IONPs by aqueous synthesis for improved Hyperthermia, *RSC Adv.*, 2017, **7**, 13159–13170.
- 15 S. Tong, C. A. Quinto, L. Zhang, P. Mohindra and G. Bao, Size-Dependent Heating of Magnetic Iron Oxide Nanoparticles, *ACS Nano*, 2017, **11**, 6808–6816.
- 16 S. Dutz, *et al.*, Ferrofluids of magnetic multicore nanoparticles for biomedical applications, *J. Magn. Magn. Mater.*, 2009, **321**, 1501–1504.
- 17 H. Forestier and G. C. Guioit-Guillain, Une nouvelle variété ferromagnétique de sesquioxyde de fer, *C. R. Acad. Sci.*, 1934, **199**, 720.
- 18 J. Jin, S. Ohkoshi and K. Hashimoto, Giant coercive field of nanometer-sized iron oxide, *Adv. Mater.*, 2004, **16**, 48–51.
- 19 S. Ohkoshi and H. Tokoro, Hard Magnetic Ferrite: $\epsilon\text{-Fe}_2\text{O}_3$, *Bull. Chem. Soc. Jpn.*, 2013, **86**, 897–907.
- 20 S. Ohkoshi, *et al.*, Nanometer-size hard magnetic ferrite exhibiting high optical-transparency and nonlinear optical-magnetoelectric effect, *Sci. Rep.*, 2015, **5**, 14414.
- 21 S. Ohkoshi, *et al.*, Large coercive field of 45 kOe on oriented magnetic film composed of metal-substituted ϵ -iron oxide, *J. Am. Chem. Soc.*, 2017, **139**, 13268.
- 22 S. Ohkoshi, *et al.*, Visible-light and THz-light induced Faraday effect on ϵ -iron oxide film, *J. Am. Chem. Soc.*, 2019, **141**, 1775.
- 23 J. Tucek, R. Zboril, A. Namai and S. Ohkoshi, $\epsilon\text{-Fe}_2\text{O}_3$: An Advanced Nanomaterial Exhibiting Giant Coercive Field, Millimeter-Wave Ferromagnetic Resonance, and Magnetoelectric Coupling, *Chem. Mater.*, 2010, **22**, 6483–6505.
- 24 L. Néel, Théorie du traînage magnétique des substances massives dans le domaine de Rayleigh, *J. Phys. Radium*, 1950, **11**, 49–61.
- 25 W. F. Brown, Thermal fluctuations of a single-domain particle, *Phys. Rev.*, 1963, **130**, 1677–1686.
- 26 R. Piñol, *et al.*, Joining Time-Resolved Thermometry and Magnetic-Induced Heating in a Single Nanoparticle Unveils Intriguing Thermal Properties, *ACS Nano*, 2015, **9**, 3134–3142.
- 27 D. Bonvin, H. Hofmann and M. M. Ebersold, Optimisation of aqueous synthesis of iron oxide nanoparticles for biomedical applications, *J. Nanopart. Res.*, 2016, **18**, 376.



- 28 K. Fushimi and A. S. Verkman, Low Viscosity in the Aqueous Domain of Cell Cytoplasm Measured by Picosecond Polarization Microfluorimetry, *J. Cell Biol.*, 1991, **112**, 719–725.
- 29 H. P. Kao, J. R. Abney and A. S. Verkman, Determinants of the Translational Mobility of a Small Solute in Cell Cytoplasm, *J. Cell Biol.*, 1993, **120**, 175–184.
- 30 H. C. Chang, Y. C. Lin and C. T. Kuo, A two-dimensional diffusion model quantifying intracellular transport with independent factors accounting for cytosol viscosity, binding, and steric hindrance, *Biochem. Eng. J.*, 2008, **41**, 217–227.
- 31 S. Bicknese, N. Periasamy, S. B. Shohet and A. S. Verkman, Cytoplasmic viscosity near the cell plasma membrane: measurement by evanescent field frequency-domain microfluorimetry, *Biophys. J.*, 1993, **165**, 1272–1282.
- 32 K. Luby-Phelps, S. Mujundar, R. Mujundar, L. Ernst, W. Galbraith and A. Waggoner, A novel fluorescence ratiometric method confirms the low solvent viscosity of the cytoplasm, *Biophys. J.*, 1993, **65**, 236–242.
- 33 R. Swaminathan, S. Bicknese, N. Periasamy and A. S. Verkman, Cytoplasmic Viscosity Near the Cell Plasma Membrane: Translational Diffusion of a Small Fluorescent Solute Measured by Total Internal Reflection-Fluorescence Photobleaching Recovery, *Biophys. J.*, 1996, **71**, 1140–1151.
- 34 P. Gonzalez-Tello, F. Camacho and G. Blazquez, Density and viscosity of concentrated aqueous solutions of polyethylene glycol, *J. Chem. Eng. Data*, 1994, **39**, 611–614.
- 35 V. K. Syal, A. Chauhan and S. Chauhan, Ultrasonic velocity, viscosity and density studies of poly (ethylene glycols)(PEG-8.000, PEG-20.000) in acetonitrile (AN) and water (H₂O) mixtures at 25 °C, *Indian J. Pure Appl. Phys.*, 2005, **27**, 61–69.
- 36 T. Mosmann, Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays, *J. Immunol. Methods*, 1983, **65**, 55–63.
- 37 M. L. Etheridge, K. R. Hurley, J. Zhang, S. Jeon, H. L. Ring, C. Hogan, C. L. Haynes, M. Garwood and J. C. Bischof, Accounting for biological aggregation in heating and imaging of magnetic nanoparticles, *Technology*, 2014, **2**, 214–228.
- 38 S. Dutz, M. Kettering, I. Hilger, R. Müller and M. Zeisberger, Magnetic multicore nanoparticles for hyperthermia-influence of particle immobilization in tumour tissue on magnetic properties, *Nanotechnology*, 2011, **22**, 265102.
- 39 M. Chen, *et al.*, Remote Control of Mechanical Forces via Mitochondrial-Targeted Magnetic Nanospinners for Efficient Cancer Treatment, *Small*, 2020, **16**, 1905424.
- 40 V. Gómez-Vallejo, *et al.*, PEG-Copolymer-coated Iron Oxide Nanoparticles that Avoid the Reticuloendothelial System and Act as Kidney MRI Contrast Agents, *Nanoscale*, 2018, **10**, 14153–14164.
- 41 L. M. A. Ali, *et al.*, F. Hemostasis disorders caused by polymer coated iron oxide nanoparticles, *J. Biomed. Nanotechnol.*, 2013, **9**, 1272.
- 42 L. M. A. Ali, *et al.*, Cell compatibility of a maghemite/polymer biomedical nanopatform, *Toxicol. In Vitro*, 2015, **29**, 962.
- 43 H. Amiri, *et al.*, Multifunctional Polymer-based Nanostructured Bio-ferrofluids as MRI Contrast Agents, *Magn. Reson. Med.*, 2011, **66**, 1715.
- 44 R. Bustamante, *et al.*, Influence of structural and magnetic properties in the heating performance of multicore bioferrofluids, *Phys. Rev. B*, 2013, **88**, 184406.
- 45 P. Díez, *et al.*, Functional Insights into the Cellular Response Triggered by Bile-Acid Platinum Compound Conjugated to Biocompatible Ferric Nanoparticles Using Quantitative Proteomic Approaches, *Nanoscale*, 2017, **9**, 9960–9972.
- 46 R. Mansell, *et al.*, Magnetic particles with perpendicular anisotropy for mechanical cancer cell destruction, *Sci. Rep.*, 2017, **7**, 4257.
- 47 E. M. Zhang, Dynamic Magnetic Fields Remote-Control Apoptosis via Nanoparticle Rotation, *ACS Nano*, 2014, **8**, 3192–3201.
- 48 M. H. Cho, *et al.*, A magnetic switch for the control of cell death signalling in in vitro and in vivo systems, *Nat. Mater.*, 2012, **11**, 1038–1043.
- 49 S. Leulmi, *et al.*, Triggering the apoptosis of targeted human renal cancer cells by the vibration of anisotropic magnetic particles attached to the cell membrane, *Nanoscale*, 2015, **7**, 15904.
- 50 D. H. Kim, *et al.*, Biofunctionalized magnetic-vortex microdiscs for targeted cancer-cell destruction, *Nat. Mater.*, 2010, **9**, 165–171.
- 51 D. Cheng, X. Li, G. Zhang and H. Shi, Morphological effect of oscillating magnetic nanoparticles in killing tumor cells, *Nanoscale Res. Lett.*, 2014, **9**, 195.
- 52 M. Domenech, I. Marrero-Berrios, M. Torres-Lugo and C. Rinaldi, Lysosomal Membrane Permeabilization by Targeted Magnetic Nanoparticles in Alternating Magnetic Fields, *ACS Nano*, 2013, **7**, 5091–5101.

