


 Cite this: *RSC Adv.*, 2020, 10, 18355

Combining amino acids and carbohydrates into readily biodegradable, task specific ionic liquids†

 Alina Brzęczek-Szafran,^a Przemysław Więcek,^a Maciej Guzik^b and Anna Chrobok^a

The growing interest in the application of ionic liquids (ILs) with simultaneous sustainability draws attention to their environmental impact in general and their biodegradability in particular. Considering this, we designed a series of novel bio-ionic liquids based on natural, abundant compounds: a carbohydrate [Carb], as the cation, and amino acids [AA], as the anions; these ILs can serve as viable alternatives to the well-known and utile cholinium AAILs. Several [Carb][AA] ILs were characterized by ¹H and ¹³C NMR, mass spectrometry, thermogravimetry (TGA) and differential scanning calorimetry (DSC). The biodegradability properties of the [Carb][AA] ILs were elucidated as well and showed biodegradation readily occurred, decomposing within 5–6 days. These novel materials were successfully utilized as catalysts for the Knoevenagel condensation reaction, where conversion values of 67–94% were achieved under exceptionally mild conditions using water as the solvent and reaction times as short as 15 minutes. These sugar based ILs were easily separated and recycled.

Received 29th March 2020

Accepted 28th April 2020

DOI: 10.1039/d0ra03664a

rsc.li/rsc-advances

Introduction

Increasing consumption of fossil fuels and its related impact on the environment have driven development of sustainable and environmentally friendly processes; ionic liquids (ILs) have contributed significantly to this development due to their intrinsic properties, which led to them being initially considered as “green”.¹ However, poor biodegradability of conventional ILs and their notable toxicity toward the environment² opened a route for development of biocompatible ILs (Bio-ILs) with the focus on using at least one ionic counterpart derived from biocompatible materials.^{3,4} Discovery of amino acid ILs (AAILs) by Fukumoto *et al.*⁵ led to intense replacement of conventional fossil cations and anions with amino acids that can be transformed into both ion types.^{6,7} An advance in this approach was recently observed making AAILs the most studied bio-derived ILs, with the most significant applications of these ILs coming in catalysis,⁸ CO₂ capture,⁹ electrochemistry¹⁰ or energy storage.¹¹ Additionally, amino acids reveal chirality so their IL derivatives constitute a diverse and sustainable pool of chiral solvents for application in catalysis and materials synthesis.¹²

On the other hand, despite origins of bio-ILs from biocompatible materials, they may not be fully biodegradable. Biodegradation aspects of different ILs classes were summarized in an

extensive review by Jordan and Gathergood, who defined several principles necessary to obtain readily biodegradable ILs, *i.e.* combination of hydroxyl functionalities and carboxylate counterions.¹³ The cholinium family of ILs has emerged here as the most readily biodegradable ILs with nearly 50% of all readily biodegradable ILs comprising a cholinium cation and an organic acid anion.¹³ Choline-based ILs have also been combined with amino acids, whose low viscosities and ease of preparation have found a wide range of applications.^{14–16}

Among other bio-renewable compounds facile for bio-ILs synthesis are carbohydrates, which due to abundance, renewability and environmental friendliness, have very recently been demonstrated as great candidates toward novel applications, which make use of their task-specific properties originating from rich hydrogen-bond network structure. In specific, carbohydrate-based ILs were used for metal complexation in thermochromic systems.¹⁷ Moreover, they were applied to catalytically enhance Diels–Alder transformations, where ILs possessing a carbohydrate cation were used as both the catalyst and the reaction solvent.¹⁸ Recently, the ability of sugar to form hydrogen bonds was used to favor gel formation between [NTf₂⁻]-based ILs and gluconate-based ILs; in addition, it was further used for desulfurization of fuels.¹⁹ The coordinating abilities of carbohydrate-based ILs were also used for cellulose dissolution, while a combination of sugars with nitrogen rich ionic moieties resulted in precursors for N-doped carbons.²⁰ Moreover, the presence of several OH groups on carbohydrate units contributed to a decrease in the toxicity of their corresponding ILs, making sugar moieties highly desirable functionalization agents from a “green chemistry” point of view.²¹

^aDepartment of Chemical Organic Technology and Petrochemistry, Silesian University of Technology, Krzywoustego 4, 44-100, Gliwice, Poland

^bJerzy Haber Institute of Catalysis and Surface Chemistry, Polish Academy of Sciences, Niezapominajek 8, 30-239 Kraków, Poland

† Electronic supplementary information (ESI) available: Synthetic procedures, NMR spectra, biodegradation patterns. See DOI: 10.1039/d0ra03664a



Considering the strategy of using both anionic and cationic components derived from biocompatible materials, principles for synthesis of biodegradable ionic systems outlined by Jordan and Gathergood and promising properties of carbohydrate-based ILs, we designed and developed readily biodegradable ILs by the first combination of carbohydrates (Carb) and amino acids (AA). Here, we report a comprehensive study revealing their thermal analysis and biodegradation patterns. Moreover, we demonstrate the catalytic performance of [Carb][AA] ILs towards the Knoevenagel condensation, featuring materials with hydrogen-bond-rich structures and amino acid based ionic moieties as task-specific ILs.

Results and discussion

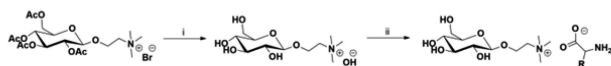
Synthesis of ILs

The synthesis of ILs was investigated using natural, abundant carbohydrate compound (glucose) and seven amino acids (L-glycine, L-leucine, L-serine, L-arginine, L-tyrosine, L-histidine, L-tryptophan). ILs were prepared using acylated *N*-[2-(D-glucopyranosyl)ethyl]-*N,N,N*-trimethylammonium bromide and a previously published procedure from our group.²² The bromide salt was transformed into the corresponding hydroxide using an ion exchange resin followed by neutralization with an amino acid as depicted in Scheme 1. That step was accompanied by acyl group hydrolysis to yield carbohydrate/amino acid-based ILs with yields of 67–87% and those structures are depicted in Fig. 1.

These ionic liquids (of the type [Carb][AAILs]) were very viscous; their viscosities were two orders of magnitude higher than corresponding cholinium ILs (Table 1).^{23,24} The profound influence on the increased viscosity had got the cation with multiple hydroxyl groups, responsible for hydrogen interactions. On the other hand, the differences in the viscosity among the particular IL types can be ascribed to larger anions and hence stronger intermolecular interactions (van der Waals, hydrogen bonding or π -stacking for derivatives with aromatic moieties). The IL with the lowest viscosity, similar to corresponding cholinium derivative was [Carb][Gly], while the highest was [Carb][Tyr]. The new ionic liquids were quite soluble in water and methanol, but not in other polar solvents like acetonitrile, ethyl acetate or DMF.

Thermal properties

Differential scanning calorimetric measurements showed no melting points for these compounds, but glass transition temperatures (T_g) were observed and ranged from -19 to 6 °C (Table 1). The systems were stable up to 198 °C with the exception of [Carb][Arg], which decomposed at 107 °C (T_d).



Scheme 1 General route for the synthesis of carbohydrate/amino acid-based ILs. Reaction conditions: (i) anion exchange resin; (ii) amino acid (1.2 eq.), ethanol, RT, 12–48 h.

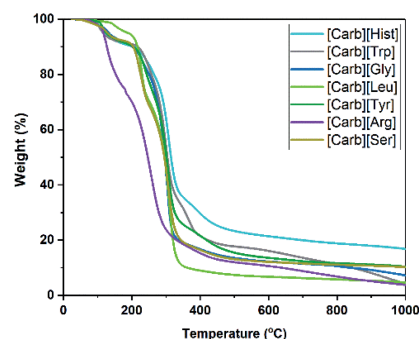


Fig. 1 Thermogravimetric analysis curves of carbohydrate/amino acid-based ILs.

Table 1 [Carb][AA] ILs properties

ILs	T_g (°C)	T_d (°C)	η /(mPa s)	
			50 °C	70 °C
[Carb][Gly]	-19	198	32 211	10 196
[Carb][Ser]	-18	198	147 953	28 645
[Carb][Leu]	4	208	—	359 841
[Carb][Arg]	-15	107	641 665	247 626
[Carb][His]	-9	207	159 815	106 930
[Carb][Trp]	0	211	—	408 284
[Carb][Tyr]	6	209	17 700 353	1 476 023

Thermogravimetric analysis curves of carbohydrate/amino acid based ILs are depicted in Fig. 2 with data presented in Table 1. As reported previously,^{5,23} the AAILs T_g and T_d values depended on the side-chain structure of the amino acid. The lowest T_g was observed for [Carb][Gly] (-19 °C), while elongating the chain resulted in higher T_g values. Similarly, the T_d values corresponded well to this trend with the exception of [Carb][Arg], which showed reduced stability. Compared to imidazolium based AAILs, these novel ILs showed slightly lower thermal stabilities (up to 211 °C),⁵ though higher than the corresponding cholinium AAILs.^{23,24}

Biodegradability

Biodegradability of novel materials is an important parameter that influences their environmental soundness. We determined whether these novel amino acid – sugar-based ILs readily decompose in controlled artificial waste waters. In general, it is worth noticing that biomass derived components, that are used to build ionic liquids, highly contribute to biodegradability increase, even when they are accompanied by a non-biodegradable component (*i.e.* tetrabutylammonium accompanied with a sugar²⁵ or a fatty acid²⁶). In our case both the hydroxy bond donors (HBD) and acceptors (HBA) were naturally sourced. Studies performed with activated sludge (Fig. 3 and ESI†) revealed that all ILs decomposed within 5–6 days. Therefore, all of these new derivatives can be classified as readily biodegradable.¹³ There were no obvious differences in degradation rates of the sugar component nor the amino acids for any IL (except for histidine). Also we did not notice any negative



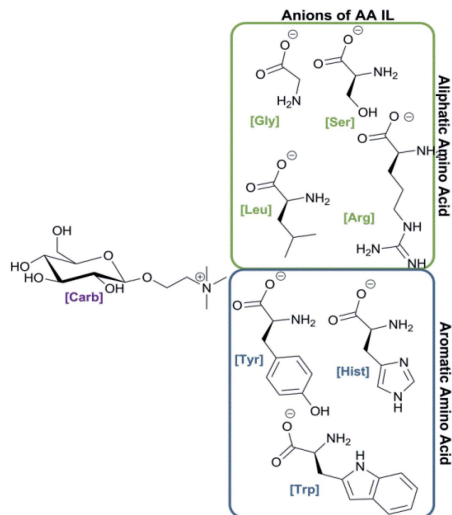


Fig. 2 Chemical structures of new carbohydrate/amino acid based ILs.

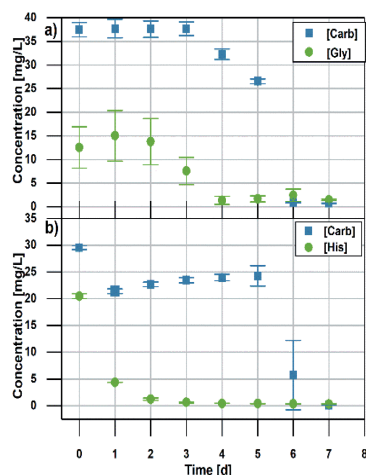


Fig. 3 Biodegradation of (a) aliphatic (b) aromatic amino acid/carbohydrate-based ILs.

impact of HBD to HBA and *vice versa* on biodegradation rates, which correlated to other studies.¹³ Similarly, ionic liquids constructed with choline and amino acids were shown to be readily biodegradable, where authors used a closed bottle and CO₂ headspace test accompanied with monitoring of the depletion of dissolved O₂.²⁷

Application example

These novel ILs were investigated as organocatalysts in the Knoevenagel condensation, a facile method for the synthesis of valuable α,β -unsaturated carbonyl compounds widely utilized in the pharmaceutical industry.²⁸ Amino acids catalyze the Knoevenagel condensation²⁹ with various activities leading to conversions from 2 to 92% after 1 h using DMSO as the solvent.³⁰ Recent studies have shown that transformation of an amino acid into an IL may lead to enhanced catalytic performance due to the role of the amino acid anion in the reaction mechanism.³¹ Thus, using amino acids which had shown poor catalytic activity towards the

condensation reaction (such as Tyr 3%, His 6%, Leu 9%), resulted in reaction time reductions to 30 minutes and conversions increased to 80–89% when these amino acids were incorporated as ILs.³¹ In the Knoevenagel condensation, AAILs possessing an anionic amino acid moiety and bound with tetrabutylammonium³¹ and cholinium cations,³² or possessing a cation with an amino acid amide anion³³ were investigated and served as easily separable and recyclable catalysts and solvents/co-solvents.

The catalytic performance of the carbohydrate AAILs was tested in the model reaction of benzaldehyde with malononitrile in water as the solvent/co-solvent. Application of water-soluble carbohydrate-based AAILs facilitated an easy separation of water-insoluble condensation product and readily recycling of the [Carb][AA] ILs for the next reaction cycle. The influence of the catalytic amounts of the IL on the example of [Carb][Leu], on the conversion of benzaldehyde is presented in Table 2. Furthermore, the influence of all investigated ILs on reaction rates are given in Fig. 4a. In the presence of all carbohydrate AAILs, high conversions of benzaldehyde (67–92%) were obtained immediately after 15 minutes with a 10% catalyst loading. The type of AAILs influenced the initial reaction rate with their activity corresponding to the activity of amino acids in the Knoevenagel condensation. In contrast to pristine amino acids, the use of AAILs resulted in shortened reaction times to 15 minutes. Longer reaction times did not improve product yields except for [Carb][Gly], which yielded a conversion of 94% after 30 min (only 67% after 15 min). The performance of investigated systems compared to other amino acid catalysts used in the Knoevenagel condensation is provided in the Table 1, ESI.† The products were exclusively *E* isomers, obtained with 100% selectivity. After separation of the water insoluble product, the aqueous phase was extracted with CH₂Cl₂ and reused in the next reaction cycle. Over four consecutive runs, no conversion drop was observed (Fig. 4b). Application of highly water soluble, hydrogen-bond-rich [Carb][AA] ILs in the Knoevenagel condensation afforded a clean, mild synthetic approach with the added benefit of product separation ease and short reaction time. Presented approach adhere to the Principles of Green Chemistry to a good extent, with Atom Economy (AE) of 90%, Reaction Mass Efficiency (RME) of 82%, Optimum Efficiency (OE) of 92% and Process Mass Intensity (PMI) of 15.3.³⁴

Table 2 The influence of catalytic amounts of the IL on the conversion of benzaldehyde^a

Catalyst loading (mol%)	Conversion (%)
0.5	24
1	37
2.5	73
5	84
10	92

^a Reaction conditions: 25 °C; 1000 rpm; benzaldehyde (1 mmol, 0.106 g); malononitrile (1 mmol, 0.661 g); H₂O 2 mL, catalyst: [Carb][Leu] (mol%); 15 min.



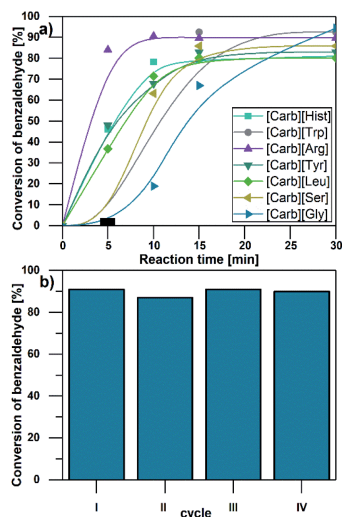


Fig. 4 (a) Influence of the amino acid anion on the conversion of benzaldehyde in the reaction with malononitrile. (b) Yield of 2-benzylidenomalononitrile from condensation of benzaldehyde with malononitrile (25 °C, 10% catalyst loading, 30 min) in consecutive recycling cycles.

Experimental

Synthesis of ILs together with the characterization data is provided in ESI (S2).†

Characterization

^1H NMR spectra were recorded at 400 MHz and ^{13}C spectra were recorded at 100 MHz (Agilent spectrometer). Chemical shifts (ppm) are reported relative to tetramethylsilane (TMS) as an internal standard. High-resolution mass spectrometry (HRMS) analyses were performed on a Waters Xevo G2 Q-TOF mass spectrometer equipped with an ESI source operating in the positive negative ion mode. Accurate mass and molecular ion compositions were calculated using the MassLynx software incorporated with the instrument. Thermogravimetric analyses (TGA) were performed using a Mettler-Toledo TGA/DSC 3+ thermobalance. Samples (approximately 5–10 mg) were heated from 25–900 °C at a rate of 10 °C min^{-1} in standard 70 μL Al_2O_3 crucibles under a dynamic nitrogen flow of 80 mL min^{-1} . Differential scanning calorimetry (DSC) were performed using a Mettler-Toledo DSC 822 analyzer. Samples were heated from –65–150 °C at a rate of 10 °C min^{-1} in standard 40 μL Al crucibles under a dynamic nitrogen flow of 80 mL min^{-1} . Viscosity measurements were carried out using a Brookfield RST-CPS rheometer with a cone and plate measuring system. Due to high viscosities, samples were analyzed at 70 °C, and where it was possible at 50 °C.

Biodegradation test

Biodegradation studies were based on principles outlined in the OECD 301 test. Briefly, an adapted minimal salt medium (MSM) was used [0.085 g L^{-1} KH_2PO_4 , 0.217 g L^{-1} K_2HPO_4 , 0.34 g L^{-1} $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$, 0.005 g L^{-1} NH_4Cl , 0.0364 g L^{-1} $\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$, 0.0225 g L^{-1} $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$, 0.00025 g L^{-1} $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$]. Activated

sludge (AS) was sourced from Waterworks Kraków, Poland. The inoculum (300 mL of AS) was preconditioned in the MSM (700 mL) for five days in the absence of a carbon source, with vigorous shaking (200 rpm) at a constant temperature of 20 °C in the absence of light. For the degradation studies, IL samples (50 mg L^{-1} in MSM) were inoculated with preconditioned activated sludge (30 mg L^{-1}). Experiments were conducted with vigorous shaking (200 rpm) at a constant temperature of 20 °C in the dark for 21 days. During that period, samples were collected and analyzed by HPLC-MS as described below.

High pressure liquid chromatography with mass detection (HPLC-MS)

The analyses were performed by HPLC measurements on an Agilent 1290 Infinity System with an autosampler and a MS Agilent 6460 Triple Quad Detector equipped with an Agilent Zorbax Eclipse Plus C18 column (2.1 \times 50 mm, 1.8 μm). To separate the IL components, the column was eluted at 30 °C, a flow rate of 0.8 mL min^{-1} and developed with an isocratic elution (95% solvent A, 0.1% formic acid in water; 5% solvent B, 0.1% formic acid in acetonitrile). The injection (2.5 μL) interval was 2.0 min. An MS Agilent 6460 Triple Quad tandem mass spectrometer with an Agilent Jet Stream ESI interface was used in positive ion mode for both HBDs and HBAs. Nitrogen (flow rate 10 L min^{-1}) was used as the drying gas and for collision-activated dissociation. Drying gas and sheath gas temperatures were set to 350 °C. The capillary voltage was 3500 V; the nozzle voltage was set to 500 V. Compounds were monitored in multiple reaction monitoring mode (MRM) with the following transitions, polarity, fragmentor (F) and collision energies (CE): ESI+: carbohydrate ($M + \text{ACN} + \text{H}$) 266.1 \rightarrow 104.1 m/z , $F = 142$ V, $\text{CE} = 30$ V; tyrosine ($M + \text{H}$) 182.1 \rightarrow 91.1 m/z , $F = 73$ V, $\text{CE} = 30$ V; arginine ($M + \text{H}$) 175.1 \rightarrow 70.1 m/z , $F = 93$ V, $\text{CE} = 26$ V; histidine ($M + \text{H}$) 156.1 \rightarrow 110.1 m/z , $F = 78$ V, $\text{CE} = 10$ V; leucine ($M + \text{H}$) 132.1 \rightarrow 86.1 m/z , $F = 78$ V, $\text{CE} = 6$ V; serine ($M + \text{H}$) 106.1 \rightarrow 42.1 m/z , $F = 64$ V, $\text{CE} = 22$ V; tryptophan ($M + \text{H}$) 205.1 \rightarrow 118.1 m/z , $F = 73$ V, $\text{CE} = 26$ V and glycine ($M + \text{H}$) 76.1 \rightarrow 30.8 m/z , $F = 44$ V, $\text{CE} = 30$ V. Standard curves were prepared in a premix of solvents A and B (95 : 5% v/v) and were used for quantitative analysis. MassHunter software (Agilent) was used for HPLC-MS system control, data acquisition and data processing.

Knoevenagel condensation

All reactions were carried out in a 10 mL round bottom flask, where benzaldehyde (1 mmol), malononitrile (1 mmol), AAIL (0.1 mmol) and 2 mL of water were placed. The mixture was vigorously stirred (1000 rpm) at 25 °C for 5–60 min. Afterwards, the mixture was homogenized with 5 mL of ethanol and the benzaldehyde conversion was monitored by GC-FID analysis with n -decane as an internal standard (50 μL of sample diluted with 1.5 mL of ethanol).

IL recycling

In typical IL recycling experiments, reactions were scaled up by a factor of ten. A mixture of benzaldehyde (10 mmol), malononitrile (10 mmol) and IL (10 mol%) were stirred in water for 20 min at 25 °C. The solid product was separated and the



aqueous phase was extracted with CH₂Cl₂ (4 × 25 mL) and reused in the next reaction.

Conclusions

Novel bio-ILs were synthesized using nontoxic, biodegradable and abundant materials (amino acids and glucose). A series of [Carb][AA] ILs was synthesized by neutralization of an *N*-[2-(D-glucopyranosyl)ethyl]-*N,N,N*-trimethylammonium hydroxide solution with seven different amino acids. Replacement of conventional, petroleum-derived imidazolium/quaternary ammonium cations with carbohydrates improved biodegradability of the ILs; most of the novel ILs decomposed within 5 to 6 days in an activated sludge environment which categorizes them as readily biodegradable. These ILs were successfully applied as catalysts for the Knoevenagel condensation reaction; conversion yields of 67–94% were achieved under exceptionally mild conditions. The use of these water soluble AAILs afforded easy separation of the products insoluble in water. Sugar based AAILs can serve as viable alternatives to cholinium AAILs for different task-specific applications and utilize their hydrogen-bond-rich structures.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This work was financed by the National Science Centre Poland (Grant no. UMO-2018/29/B/ST8/01784). We greatly acknowledge the joint consortium “Interdisciplinary Centre of Physical, Chemical and Biological Sciences” of ICSC PAS and INP PAS for providing access to the Agilent 1290 Infinity System with the autosampler and MS Agilent 6460 Triple Quad Detector.

Notes and references

- M. Kar, K. Matuszek, and D. R. MacFarlane, *Kirk-Othmer Encyclopedia of Chemical Technology*, 2019, p. 1.
- J. Pernak, J. Rogoża and I. Mirska, *Eur. J. Med. Chem.*, 2001, **36**, 313–320.
- J. Hulsbosch, D. E. De Vos, K. Binnemans and R. Ameloot, *ACS Sustainable Chem. Eng.*, 2016, **4**, 2917–2931.
- J. M. Gomes, S. S. Silva and R. L. Reis, *Chem. Soc. Rev.*, 2019, **48**, 4317–4335.
- K. Fukumoto, M. Yoshizawa and H. Ohno, *J. Am. Chem. Soc.*, 2005, **127**, 2398–2399.
- L. He, G.-H. Tao, D. A. Parrish and J. M. Shreeve, *J. Phys. Chem. B*, 2009, **113**, 15162–15169.
- Y. Qian, S. Xiao, L. Liu and Y. Wang, *Tetrahedron: Asymmetry*, 2008, **19**, 1515–1518.
- M. P. Pereira, R. de Souza Martins, M. A. L. de Oliveira and F. I. Bombonato, *RSC Adv.*, 2018, **8**, 23903–23913.
- F.-F. Chen, K. Huang, Y. Zhou, Z.-Q. Tian, X. Zhu, D.-J. Tao, D. Jiang and S. Dai, *Angew. Chem., Int. Ed.*, 2016, **55**, 7166–7170.
- X. Lu, X. Wang, J. Jin, Q. Zhang and J. Chen, *Biosens. Bioelectron.*, 2014, **62**, 134–139.
- H. Zhou, Y. Zhou, L. Li, Y. Li, X. Liu, P. Zhao and B. Gao, *ACS Sustainable Chem. Eng.*, 2019, **7**, 9281–9290.
- F. Tang, Q. Zhang, D. Ren, Z. Nie, Q. Liu and S. Yao, *J. Chromatogr. A*, 2010, **1217**, 4669–4674.
- A. Jordan and N. Gathergood, *Chem. Soc. Rev.*, 2015, **44**, 8200–8237.
- R. Wang, Y. Chang, Z. Tan and F. Li, *Sep. Sci. Technol.*, 2016, **51**, 1093–1102.
- Z. Zhang, N. Kang, J. Zhou, X. Li, L. He and H. Sui, *Nanomaterials*, 2019, **9**, 504.
- S. Bhattacharyya and F. U. Shah, *ACS Sustainable Chem. Eng.*, 2016, **4**, 5441–5449.
- F. Billeci, H. N. Gunaratne, F. D'Anna, G. G. Morgan, K. R. Seddon and N. V. Plechkova, *Green Chem.*, 2019, **21**, 1412–1416.
- K. Erfurt, I. Wandzik, K. Walczak, K. Matuszek and A. Chrobok, *Green Chem.*, 2014, **16**, 3508–3514.
- F. Billeci, F. D'Anna, H. N. Gunaratne, N. V. Plechkova and K. R. Seddon, *Green Chem.*, 2018, **20**, 4260–4276.
- A. Brzeczek-Szafran, K. Erfurt, A. Blacha-Grzechnik, M. Krzywiecki, S. Boncel and A. Chrobok, *ACS Sustainable Chem. Eng.*, 2019, **7**, 19880–19888.
- F. Billeci, F. D'Anna, M. Feroci, P. Cancemi, S. Feo, A. Forlino, F. Tonnelli, K. R. Seddon, H. N. Gunaratne and N. V. Plechkova, *ACS Sustainable Chem. Eng.*, 2020, **8**, 926–938.
- J. Pernak, K. Czerniak, A. Biedziak, K. Marcinkowska, T. Praczyk, K. Erfurt and A. Chrobok, *RSC Adv.*, 2016, **6**, 52781–52789.
- Q.-P. Liu, X.-D. Hou, N. Li and M.-H. Zong, *Green Chem.*, 2012, **14**, 304–307.
- D.-J. Tao, Z. Cheng, F.-F. Chen, Z.-M. Li, N. Hu and X.-S. Chen, *J. Chem. Eng. Data*, 2013, **58**, 1542–1548.
- N. Ferlin, M. Courty, S. Gatard, M. Spulak, B. Quilty, I. Beadham, M. Ghavre, A. Haifß, K. Kümmerer, N. Gathergood and others, *Tetrahedron*, 2013, **69**, 6150–6161.
- K. Harażna, K. Walas, P. Urbańska, T. Witko, W. Snoch, A. Siemek, B. Jachimska, M. Krzan, B. D. Napruszewska, M. Witko and others, *Green Chem.*, 2019, **21**, 3116–3126.
- X.-D. Hou, Q.-P. Liu, T. J. Smith, N. Li and M.-H. Zong, *PLoS One*, 2013, **8**, e59145.
- Y. Ono, *J. Catal.*, 2003, **216**, 406–415.
- F. S. Prout, *J. Org. Chem.*, 1953, **18**, 928–933.
- Y.-H. He, Y. Hu and Z. Guan, *Synth. Commun.*, 2011, **41**, 1617–1628.
- P. Ossowicz, Z. Rozwadowski, M. Gano and E. Janus, *Pol. J. Chem. Technol.*, 2016, **18**, 90–95.
- P. Moriel, E. J. Garcia-Suarez, M. Martinez, A. B. Garcia, M. Montes-Morán, V. Calvino-Casilda and M. A. Bañares, *Tetrahedron Lett.*, 2010, **51**, 4877–4881.
- P. A. Burate, B. R. Javle, P. H. Desale and A. K. Kinage, *Catal. Lett.*, 2019, **149**, 2368–2375.
- C. R. McElroy, A. Constantinou, L. C. Jones, L. Summerton and J. H. Clark, *Green Chem.*, 2015, **17**, 3111–3121.

