RSC Advances



REVIEW

View Article Online
View Journal | View Issue



Cite this: RSC Adv., 2020, 10, 24951

the brown algae genus *Sargassum*Mohammed I. Rushdi, Iman A. M. Abdel-Rahman, Iman Saber, Iman Saber

Pharmacological and natural products diversity of

Eman Zekry Attia,^c Wedad M. Abdelraheem,^d Hashem A. Madkour,^e Hossam M. Hassan, of Abeer H. Elmaidomy^f and Usama Ramadan Abdelmohsen **p**

Sargassum (F. Sargassaceae) is an important seaweed excessively distributed in tropical and subtropical regions. Different species of Sargassum have folk applications in human nutrition and are considered a rich source of vitamins, carotenoids, proteins, and minerals. Many bioactive compounds chemically classified as terpenoids, sterols, sulfated polysaccharides, polyphenols, sargaquinoic acids, sargachromenol, and pheophytin were isolated from different Sargassum species. These isolated compounds and/or extracts exhibit diverse biological activities, including analgesic, anti-inflammatory, antioxidant, neuroprotective, anti-microbial, anti-tumor, fibrinolytic, immune-modulatory, anti-coagulant, hepatoprotective, and anti-viral activities. This review covers the literature from 1974 to 2020 on the genus Sargassum, and reveal the active components together with their biological activities according to their structure to create a base for additional studies on the clinical applications of Sargassum.

Received 21st April 2020 Accepted 13th June 2020

DOI: 10.1039/d0ra03576a

rsc.li/rsc-advances

Introduction

Marine natural products have been characterized by their chemical structural diversity, along with different biological activities. The Red Sea is considered one of the most important marine hotspots comprising high biodiversity. Many marine algae are native to the Red Sea, which could be classified into brown algae (Phaeophyta), green algae (Chlorophyta), and red algae (Rhodophyta). Brown seaweeds are predominantly brown in color because of their contents of carotenoid fucoxanthin and polysaccharides, namely alginates, laminarins, fucans, and cellulose. Green seaweeds are characterized by their chlorophyll a and b content with ulvan being the major polysaccharide. While, in red seaweeds, the principal pigments are phycoerythrin, phycocyanin, and the primary polysaccharides are agars

Sargassum (F. Sargassaceae), of the order Fucales, subclass Cyclosporeae, and class Phaeophyceae, is a genus of brown algae, communally known as gulfweed or sea holly, and is considered one of the most complex Phaeophyceae genera. Sargassum was discovered by Agardh in 1820 and is reported to contain 537 species names in the algae database, of which 358 species have been accepted taxonomically. It comprises many different species that are distributed worldwide, although primarily found in tropical and subtropical marine waters, and also generally growing on rocky reefs.

Sargassum thallus (10–200 cm) may be linear or bushy, with stipes 1–20 cm long arising from a discoid–conical holdfast, and do not penetrate the substratum. Its main axes are perennial, short, and cylindrical or flattened in sections, and bear the scars of deciduous branches. The shape of the leaves can be simple, bifid, or divided several times, round, spatulate, turbinate, lanceolate, ovoid, linear or of any intermediate form with air bladders (vesicles) normally present, subspherical to the ovoid, petiolate, mutic or apiculate, replacing the ramuli or axillary to the laterals. The basis of the leaves is rounded or attenuate, symmetrical or not. The pedicel may be variable in length, cylindrical, or flattened in sections and smooth. The leaves margin may be simple or doubled at the apex, and may be smooth, undulated, finely serrated, deeply dentated, or any intermediate aspect. The midrib may be short and thick or

and carrageenans. Also, marine algae are rich sources of structurally diverse bioactive compounds with various biological activities, although only a few species have been chemically examined in the last decades.

^aDepartment of Pharmacognosy, Faculty of Pharmacy, South Valley University, 83523, Oena, Egypt

^bDepartment of Botany and Microbiology, Faculty of Science, South Valley University, 83523, Qena, Egypt

Department of Pharmacognosy, Faculty of Pharmacy, Minia University, 61519 Minia, Egypt. E-mail: usama.ramadan@mu.edu.eg; Fax: +20-86-2369075; Tel: +20-86-2347759

^aDepartment of Medical Microbiology and Immunology, Faculty of Medicine, Minia University, 61519 Minia, Egypt

^eDepartment of Marine and Environmental Geology, National Institute of Oceanography and Fisheries, Red Sea Branch, 84511 Hurghada, Egypt

Department of Pharmacognosy, Faculty of Pharmacy, Beni-Suef University, Beni-Suef, 62514, Egypt

^{*}Department of Pharmacognosy, Faculty of Pharmacy, Deraya University, Universities Zone, 61111 New Minia City, Egypt



Fig. 1 Sargassum sanyaense from Hurghada City coast line, Red Sea (Egypt).

thinner and reaching the apex or any intermediate length. The apex may be acute, rounded, or truncated, simple, or showing a cup-like shaped depression (Fig. 1).²

A survey of folk uses of *Sargassum* spp. in the last 20 years period (1977–1996), showed that different regions in the tropical country use the brown seaweed differently. Commonly, *Sargassum* spp., is used as a cover or wrapper of fish to maintain their freshness. In the Ilocos Region in the Northern Philippines, *Sargassum* spp. is used as a vegetable; whereas in the Visayas and Northern Mindanao, natives utilize *Sargassum* spp. as a fertilizer, flower inducer, and insect repellant. The Boholano people also use the brown seaweed as animal feed. In certain parts of the island of Bohol, a *Sargassum* drink is made and is reported to have health benefits.¹

For nearly 2000 years, *Sargassum* spp. has been also used in Traditional Chinese Medicine (TCM) to treat a variety of diseases, including thyroid.¹ Although *Sargassum* has not attracted the attention of most researchers, the therapeutic potentials of pure compounds isolated from *Sargassum* are promising for their antiviral, antimicrobial, cytotoxic, antipyretic, anti-inflammatory, cardioprotective, hepatoprotective, and hypolipidemic properties.³ Some metabolites, such as embelin, tanacetol A, and oxygenated fucosterols, have attracted great attention because of their uncommon structural complexity and interesting pharmacological properties.¹¹³ Some species are economically important, especially in the food, textile, cosmetics, and pharmaceutical industries. This review covers the literature on the genus *Sargassum* along with its chemical and medicinal potential.

2. Sargassum angustifolium

S. angustifolium, commonly known as narrow leaf *Sargassum* weed, is widely distributed around the Western Indian Ocean.⁴ The *n*-hexane, dichloromethane, and *n*-butanol fractions isolated from *S. angustifolium*, collected from the Persian Gulf, were reported to have a cytotoxic effect against HeLa (cervical cancer) and MCF-7 (breast cancer) cells, where the cell survival was inversely proportional to the increase in the concentration of the different extracts, respectively, from 150 µg mL⁻¹ to 900

μg mL⁻¹. The, *n*-hexane fraction was shown to have a median growth inhibitory concentration value (MIC) of 71 and 77 μg mL⁻¹, against HeLa and MCF-7, while the MIC of the dichloromethane fraction was 36 and 88 μg mL⁻¹, respectively. The *n*-butanol fraction showed an MIC of 25 μg mL⁻¹ against MCF-7.⁴ Sulfated polysaccharides isolated from *S. angustifolium* were reported to have potent immuno-stimulant activity, through inducing RAW264.7 macrophage cells to release nitric oxide, IL-1β, TNF- α , IL-6, IL-10, and IL-12 through activation of the NF-κB and MAPKs signaling pathways.⁵ Also, the phosphate buffer extract of *S. angustifolium* had a low α -amylase inhibition activity with an IC₅₀ of 1.85 mg mL⁻¹.⁶

3. Sargassum aquifolium

GC-MS of a crude petroleum ether extract of *S. aquifolium* (Turner), collected from the Red Sea of Jazan (KSA), was reported containing phenol, 2,4-bis(1,1-dimethylethyl) 1, phenol, 2,2'-methylenebis[6-(1,1-dimethylethyl)-4-methyl 2, stigmasta-5,24(28)-dien-3-ol, (3\u00e1,24Z) 3, and stigmasterol 4 (Fig. 2).⁷ The crystals of a petroleum ether extract of *S. aquifolium* dissolved in Millipore water were tested for antibacterial activity against *S. aureus*, *S. pyogenes*, *B. subtilis*, *E. coli*, *P. aeruginosa*, and *K. pneumonia*. The maximum antibacterial activity was exerted against *E. coli*, with an MIC of 100–150 mg mL⁻¹.⁷ L-Fucose, D-galactose, D-mannose, D-glucuronic acid, D-xylose, and sulfate were found to be the main constituents of fucoidan detected in *S. aquifolium*, collected from Vietnam, with anticoagulant,

Fig. 2 Chemical structures of compounds 1–24.

cytotoxic, and antitumor activities, which increase with sulfation.⁸

4. Sargassum asperifolium

The polysaccharide extracts of *S. asperifolium*, collected from the Red Sea (Egypt), possessed anti-inflammatory activity, through the significant inhibition of nitric oxide generation, and LPS-induced TNF- α . Analysis of the volatile fraction of *S. asperifolium* led to the identification of fucosterol 5, cholesterol 6, and palmitic acid 7. Dictyone 8, dictyone acetate 9, saringosterone 10, and saringosterol 11 were also detected in *S. asperifolium* collected from Hurghada (Egypt) (Fig. 2). 11

5. Sargassum autumnale

Isonahocol D1 **12**, D2 **13**, nahocol-A **14**, nahocol-A1 **15**, nahocol-B, C, D1, and D2 **16–19**, were detected in a methanol extract of *S. autumnale*, collected from Omosu Bay (Japan). Nahocols were shown to have an aryl prenyl ether structure, which is considered a precursor of the prenyl hydroquinones or prenyl benzoquinones in the plant and animal kingdoms.¹²

6. Sargassum binderi

Fucoxanthin **20** was detected in Malaysian *S. binderi* and *S. duplicatum*. The amount of fucoxanthin **20** and the total lipid contents of *S. duplicatum* (1.01 \pm 0.10 and 21.3 \pm 0.10 mg g⁻¹ dry weight, respectively) were significantly higher than those of *S. binderi* (0.73 \pm 0.39 and 16.6 \pm 4.10, respectively). Docosahexanoic acid, eicosapentanoic acid, arachidonic acid, linoleic acid and α -linolenic unsaturated fatty contents in *S. duplicatum* were found to be higher (0.76%, 2.55%, 13.64%, 5.81%, and 5.35%, respectively) than in *S. binderi* (0.70%, 1.82%, 9.13%, 6.37% and 4.39%, respectively), while palmitic acid (7) the major fatty acid in both samples.¹³

7. Sargassum boveanum

The different fractions of *n*-hexane, trichloroethane, chloroform, and *n*-butanol of the methanol-ethyl acetate extract of *S. boveanum*, collected from the Persian Gulf, showed a decrease in the cell survival of HeLa cells with increasing the concentration of the extracts, with IC₅₀ values of 150.3 \pm 23.10, 437.0 \pm 147.3, 110.4 \pm 33.67, and 1025.0 \pm 15.20 μg mL $^{-1}$, respectively. 14

8. Sargassum carpophyllum

28ξ-Dihydroxy-24-ethylcholesta-5,23-*Z*-dien 21 and 2*a*-oxa-2-oxo-5*f*-hydroxy-3,4-dinor-24-ethylcholesta-24(28)-ene 22 together with five steroids, namely fucosterol 5, 24-ethylcholesta-4,24(28)-dien-3,6-dione 23, 24ξ-hydroperoxy-24-vinylcholesterol 24, 24-ketocholesterol 25, and 24*R*,28*R*- and 24*S*,28*S*-epoxy-24-ethylcholesterol 26 (Fig. 2 and 3) were detected in an ethanol extract of *S. carpophyllum*, collected from the Beihai (China) Sea. Compounds 21 and 24 reportedly had

Fig. 3 Chemical structures of compounds 21-44.

cytotoxic activities against HL-60, with IC_{50} values of 7.8 and 8.5 $\mu g \; mL^{-1}$, respectively, while compounds 5 and 23 showed potent cytotoxic activities against P-388, with IC_{50} values of 0.7 and 0.8 $\mu g \; mL^{-1}$, respectively. Compound 26 showed cytotoxic activity against MCF-7, HCT-8, 1A9, HOS, and PC-3, with IC_{50} values of 4.0, 8.8, 10.0, 10.0, and 7.2 $\mu g \; mL^{-1}$, respectively. Another compound, 24-ethyl-3-carbethoxy-3-hydroxy-A-nor-cholesta-5,24(28)-dien-2-one 27, was also detected in *S. carpo-phyllum*, collected from the South China Sea, which was the first reported non-steroidal ester from natural sources. ¹⁶

9. Sargassum cinereum

Fucoidan was detected in *S. cinereum*, collected from Tuticorin Lat. (India). Fucoidan was reported to contain 65.753% of L-fucose and 3.7% \pm 1.54% of sulfate, respectively, with a monosaccharide composition, such as L-fucose, D-galactose, D-mannose, and D-xylose. The maximum 2,2′-diphenyl-1-picrylhydrazyl (DPPH) scavenger activity for the fucoidan extract was found at the concentration of 100 μg , whereas the crude extract of *S. cinereum* showed 63.58% \pm 0.56% scavenging activity. Fucoidan extract showed 50% cell death after 24 h of incubation (75 \pm 0.9037 μg mL $^{-1}$) against the colon cancer cell line HCT-15. 17 The mechanism of fucoidan was a dosedependent inhibited growth of the colon cancer cell line Caco-2, (IC $_{50}=250~\mu g$ mL $^{-1}$), the increased production of reactive oxygen species (ROS), and augmented mitochondrial membrane permeability. 18

RSC Advances Review

10. Sargassum crassifolium

An *S. crassifolium* ethanol extract, collected from Diora-Zinungan, showed antimicrobial activity against *A. hydrophila*, with an MIC value of 24.33 ± 0.58 mm.¹⁹

11. Sargassum cristaefolium

Sodium alginate was detected in *S. cristaefolium*, collected from Poteran Island (Indonesia). This alginate had a molecular weight of 217.94 \pm 7.14 kDa with a mannuronic acid to guluronic acid ratio (M/G) of 0.28, and the L-guluronic acid block was 0.78, which was higher than the D-mannuronic acid block.²⁰

12. Sargassum dentifolium

Polysaccharide extracts of *S. dentifolium* collected from the Red Sea (Egypt) had a protective effect against cyclophosphamide (CP)-induced genotoxicity in mice bone marrow cells (BMCs).²¹ Also, the ethanol extract of *S. dentifolium*, reported as hepatoprotective in carbon tetrachloride (CCl₄)-induced hepatitis in rats, had antioxidant activity.²²

13. Sargassum fallax

Fallachromenoic acid **28**, an unusual halogenated meroditerpenoid, retroflexanone **29**, fallahydroquinone **30**, and fallaquinone **31** (Fig. 3), were isolated from the Southern Australian *S. fallax*, and were shown to have antitumor activities against a P388 Murine Leukaemia cell line, with an $IC_{50} > 27-29$ $\mu M.^{23}$

14. Sargassum filipendula

Heterofucan extracted from *S. filipendula* was shown to have antiproliferative and apoptosis activities against human cervical cancer (HeLa) cells, at a concentration of 0.1 to 2.0 mg mL⁻¹.²⁴

15. Sargassum fluitans

The *in vivo* anti-fibrotic effect of a fucoidan extract of *S. fluitans* (chemically composed of carbohydrates, sulfates, uronic acids, protein, and phenols), collected from Puerto Morelos (Mexico), was detected in a $\mathrm{CCl_4}$ -induced liver damage model in rats. The daily oral administration of fucoidan extract (50 mg kg $^{-1}$) showed a significant reduction in liver enzymatic activity, liver infiltration of inflammatory cells, collagen fiber deposition, and gene expression cytokines. 25

16. Sargassum fulvellum

Grasshopper ketone 32 was detected in an ethanol extract of *S. fulvellum* (SFEE) collected from Jindo (Korea). Grasshopper ketone 32 and SFEE were investigated on 2,4-dinitro-chlorobenzene (DNCB)-induced atopic dermatitis (AD)-like skin lesions in BALB/c mice. SFEE and grasshopper ketone 32 were found to have an inhibitory effect on AD by regulating immune

mediators and cells and thus may be a potentially effective alternative therapy for AD.26 Boiling water, ethanol, and dichloromethane extracts of S. fulvellum, collected from Wando aquaculture farm (Korea), were tested for anti-inflammatory, antipyretic, and analgesic activities in mice. The dichloromethane extract (0.4 mg per ear) significantly inhibited the inflammatory symptoms of mouse ear edema by 79.1%.27 Pheophytin A 33 was detected in S. fulvellum collected from the Japanese coastline. When PC12 cells were treated with a low concentration of pheophytin A 33 (3.9 μ g mL⁻¹) in the presence of a low level of nerve growth factor (10 ng mL⁻¹), the compound produced neuritis outgrowth similar to that produced by a high level of nerve growth factor (50 ng mL⁻¹). Pheophytin A 33 also enhanced signal transduction in the mitogen-activated protein kinase signaling pathway, which is also induced by nerve growth factor.28 1-O-Palmitoyl-2-O-oleoyl-3-O-(α-D-glucopyranosyl)glycerol (POGG) 34 and 1-O-myristoyl-2-O-oleoyl-3-O-(α-D-glucopyranosyl)-glycerol (MOGG) 35 were obtained from S. fulvellum collected from the East Sea in China. POGG 34 and MOGG 35 showed fibrinolytic activity in the reaction system of pro-u-PA and plasminogen.29 Fulvellic acid esters 36 and 37 are glycerides bearing a methacrylic acid moiety and were detected in a methanol extract of S. fulvellum collected from Awakominato (Chiba) (Fig. 3).30

17. Sargassum fusiforme

The polysaccharide fraction detected in *S. fusiforme* collected from Qingdao (China) consisted of L-fucose, p-mannose, L-rhamnose, p-glucose, p-galactose, and p-glucuronic acid with different molar ratios. Regarding the hypoglycemic and hypolipidemic effects, the oral administration of these polysaccharide fractions prominently restrained the loss of body weight and increase of water intake, and also significantly controlled the increase in the levels of fasting blood glucose of diabetic rats, as well as showed better effects in controlling fasting blood glucose, alanine aminotransferase (ALT), uric acid (UA) and urea nitrogen (BUN) levels.³¹ The polysaccharide also showed a significant protective effect against ultraviolet B (UVB) radiation in hairless Kun Ming (KM) mice.³²

18. Sargassum glaucescens

Stigmasterol 4, fucosterol 5, cholesterol 6, 24(S)-hydroxy-24-vinylcholesterol 38, 24(R)-hydroxy-24-vinylcholesterol 39, and β -sitosterol 40 (Fig. 3) were detected in *S. glaucescens* collected from Southern Iran. *In vitro* α -amylase inhibitory activities were detected with a methanol extract, whereby a potent inhibition (IC $_{50}=8.9\pm2.4~{\rm mg~mL}^{-1}$) of the enzyme compared to acarbose as a positive control was detected. Fucoidan was detected in *S. glaucescens* collected from Kenting (Taiwan), where the monosaccharide composition was L-fucose, sulfate, and uronic acid, and it was also shown to exhibit antioxidant activities, in a dose-dependent manner, against DPPH.

19. Sargassum hemiphyllum

A hot-water extract of S. hemiphyllum collected from the coast of Penghu County (Taiwan) was found to have antioxidant activity and immune-stimulating activities when using HB4C5, and J774.1 cells. The antioxidant activity increased with a concentration <3.5 mg mL⁻¹. The HB4C5 cells showed the maximum relative activities of cell proliferation (174%) and IgM secretion (132%) with 120 $\mu g \text{ mL}^{-1}$ of the hot-water extract at 80 $\mu g \text{ mL}^{-1}$, while J774.1 cells showed the maximum relative activities of cell proliferation (141%) and phagocytosis (147%).35 The major polyphenols (17.35 \pm 0.93–36.66 \pm 2.01 mg g⁻¹) were extracted using water, ethanol, and acetone (WES, EES, and AES, respectively) for S. hemiphyllum. The inhibition of α -amylase, α glucosidase, sucrose, and maltase activities and stimulation of insulin secretion were greater with AES than with WES or EES. Moreover, 250 μg mL⁻¹ EES and AES significantly increased insulin secretion in the presence of 25 mg mL⁻¹ glibenclamide compared to with 50 mg mL⁻¹ glibenclamide.³⁶ Sulfated polysaccharide extracted from S. hemiphyllum (SHSP) was detected on the mouse macrophage cell line (RAW 264.7) activated by

lipopolysaccharide (LPS), which can used as a model system. The secretion profiles of pro-inflammatory cytokines, including IL-1β, IL-6, TNF-α, and NO, were found to be significantly reduced in 1-5 mg mL⁻¹ dose ranges of SHSP treatments, in which the anti-inflammatory properties of SHSP may be attributed to the downregulation of NF-κB in the nucleus.37

Sargassum henslowianum 20.

Saringosterol 11, aurantiamide acetate 41, aurantiamide 42, vestitone 43, 7-hydroxy-2',4'-dimethoxyisoflavanone 44, liquiritigenin 45, 5,6-dihydroxy-7-methoxyflavone 46, and 5,7dihydroxy-8-methoxyflavone 47 (Fig. 3 and 4) were detected in an ethanol extract of S. henslowianum collected from Guangdong and Fujian Provinces (China).38 Fucoidan was also detected in S. henslowianum collected from Hai Van-Son Cha peninsula (Vietnam). The oral dose (100 mg kg⁻¹) of fucoidan was shown to decrease cholesterol, triglyceride, and LDLcholesterol levels in obese mice.39 Fucose-containing sulfated polysaccharides (FCSPs) extracted from S. henslowianum

Chemical structures of compounds 45-66.

notably had dose-dependent growth-inhibitory effects on the proliferation of melanoma B16 cells.⁴⁰

21. Sargassum horneri

The anti-inflammatory effects of an ethanol extract of *S. horneri* (ESH) on the RAW 264.7 murine macrophage cell line were tested. ESH was not cytotoxic to RAW 264.7, while a dose of ESH 200 μg mL⁻¹ was shown to reduce the mRNA level of cytokines, including IL-1β, and pro-inflammatory genes as iNOS and COX-2 in LPS-stimulated macrophage activation. ESH was also found to elicit anti-inflammatory effects by inhibiting ERK, p-p38, and NF-κB phosphorylation.⁴¹ Also, 6-hydroxy-4,4,7*a*-trimethyl-5,6,7,7*a*-tetrahydrobenzofuran-2(4*H*)-one **48** (Fig. 4) has been detected in ESH.⁴²

22. Sargassum kjellmanianum

(+)-Kjellmanianone 49 (Fig. 4), is a highly oxygenated cyclopentenone isolated from S. kjellmanianum collected from the Bay of Hiroshima (Japan), and was shown to have moderate antibacterial activity against E. coli and B. subtilis var. niger, Gram-positive microorganisms. 43 Sargassumlactam 50 (Fig. 4) was also detected in S. kjellmanianum and was shown to have a moderate antibacterial effect against S. aureus and E. coli. 44 An aqueous extract of S. kjellmanianum collected from Hahakimoku (Japan) was effective in the in vivo growth inhibition of implanted Sarcoma-180 cells, but showed no effect against L-1210-bearing mice. Polysaccharide fractions containing Lfucose and ester sulfate in the amounts of 12.6%, and 15.4%, 23.5%, and 17.2%, respectively, were extracted from S. kjellmanianum. The sulfated polysaccharide was effective against L-1210 leukemia with an ILS value of 26%. 45 The in vitro immuneregulatory and anti-tumor activities of sulfated polysaccharides from a hot-water crude extract of S. kjellmanianum (SKP) were detected; the polysaccharides were composed of L-fucose, Dgalactose, D-xylose, and D-mannose. All three types of polysaccharides were sulfated, with the sulfate group accounting for 3.4%, 25.6%, and 29.9% of SKP1, SKP2, and SKP3, respectively. SKP2 and SKP3 significantly enhanced the immune function of immunocytes, and SKP2 increased the proliferation rate of spleen lymphocytes and peritoneal macrophages by up to 53.56% and 51.41%. Cell culture showed that SKP1, SKP2, and SKP3 all inhibited the proliferation of HT-29 and HeLa cells. 46

23. Sargassum ilicifolium

The anticonvulsant activity of *S. ilicifolium* collected from Bhatkarwada (India) in maximal electroshock (MES)- and pentylenetetrazole (PTZ)-induced convulsion in mice was detected. Chloroform (600 mg kg $^{-1}$) and ethanol extracts (400 mg kg $^{-1}$ and 600 mg kg $^{-1}$) of *S. ilicifolium* significantly downregulated the duration of tonic hind limb extension in a maximal electroshock (MES) model and upregulated the latency to the onset of convulsions in a pentylenetetrazol (PTZ) model. These results were comparatively similar to the effects of phenytoin (25 mg kg $^{-1}$) and phenobarbitone (20 mg kg $^{-1}$).

24. Sargassum integerrimum

Heteropolysaccharides were detected in *S. integerrimum*, and were shown to have neuroprotective and antioxidant activities.⁴⁸

25. Sargassum lacerifolium

Dimethylarsonioboside **51**, [deoxy(dimethylarsinoyl)ribosyl] mannitol **52**, and methyl 5-deoxy(dimethylarsinoyl) riboside **53** (Fig. 4) are arsenic-containing ribosides, isolated from the methanol extract of *S. lacerifolium* collected from Mullaloo Beach (Australia).⁴⁹

26. Sargassum latifolium

Polysaccharides (E1–E4) were detected in *S. latifolium* collected from in the Red Sea (Egypt). E1 and E4 were potent anti-initiators, where they lead not only to an inhibition in the carcinogen activator cytochrome P450 1A (IC₅₀ 2.54 and 10.30 μg mL⁻¹, respectively) but also to an induction in the carcinogen detoxification enzymes glutathione-*S*-transferases (144% and 225% of the control, respectively). E1 and E4 inhibited 59% and 63% of the induced-DNA damage. The anti-inflammatory activity of E1 and E4 enhanced the macrophage proliferation, inhibited the stimulated NO (30.7% and 59.3%), TNF- α (38.2% and 54.9), and COX-2 (20% and 18%), respectively. E3 showed also a selective cytotoxicity against lymphoblastic leukemia (1301 cells).⁵⁰

27. Sargassum linifolium

The carbohydrate moiety of sargassan involves a backbone chain of D-glucuronic acid and D-mannose residues, and side chains involving residues of D-galactose, D-xylose, and L-fucose with sulfate attached to some galactose and fucose residues were detected in *S. linifolium* collected from Alexandria (Egypt).⁵¹

28. Sargassum macrocarpum

Sargafuran 54 (Fig. 4) was detected in a methanol extract of S. macrocarpum, collected from north to south along the Japanese coastline, and showed activity against Propionibacterium acnes, with an MIC of 15 μg mL⁻¹.52 Sargachromenol 55, isolated from S. macrocarpum, had a marked nerve growth factor (NGF)dependent neurite outgrowth promoting activity to PC12D cells with a median effective dose (ED_{50}) of 9 μ M, against PC12D cells in the presence of 10 ng mL⁻¹ NGF. Sargachromenol 55 significantly promoted the survival of neuronal PC12D cells at 0-50 ng mL⁻¹ NGF in a serum-free medium.⁵³ Tuberatolide B **56** (Fig. 4) is a diastereomeric meroterpenoid detected in S. macrocarpum, and inhibits tumor growth in breast, lung, colon, prostate, and cervical cancer cells, and suppresses cancer progression by promoting the ROS-mediated inhibition of STAT3 signaling.54 The anti-inflammatory effects of an extract of S. macrocarpum (SME), collected from Jeju Island (Korea), in bone marrow-derived macrophages (BMDMs) and dendritic

cells (BMDCs) were detected. Primary BMDMs and BMDCs were used for cytokine production and western blot analysis. SME (0-50 μg mL⁻¹) pre-treatment led to a strong inhibitory effect against IL-12 p40, IL-6, and TNF-α, production in CpGstimulated BMDMs and BMDCs. SME pre-treatment caused a strong inhibitory effect against NF-κB activation.55

29. Sargassum mangarevense

S. mangarevense collected from Tahiti contained alginate (9.3% \pm 1.7% dw, M/G = 1.42 \pm 0.24), mannitol (12.2% \pm 2.1% dw), and phenolic contents (2.85% \pm 1.12% dw). The aqueous and ethanol extracts of S. mangarevense showed antimicrobial activity against S. aureus with MICs of 9.5 and 12.5 mm, respectively.56

Sargassum marginatum 30.

The growth inhibition of human pro-myelocytic leukemia (HL-60) cells by lipid extracts of S. marginatum collected from Goa (west coast of India) was detected with special reference to the fatty acid composition. PL exhibited cytotoxic activity at concentrations <20 μg mL⁻¹.⁵⁷

31. Sargassum micracanthum

Fucosterol 5, sargachromenol 55, and sargaquinoic acid 57 were detected in S. micracanthum collected on Jeju Island (Korea). Sargachromenol 55 and sargaquinoic acid 57 were found to scavenge DPPH (IC₅₀ = 49.3 and 100.2 μ M).⁵⁸ 2-[(2E,6E,10E)-15-Hydroxy-3,7,11,15-tetramethyl-14-oxohexadeca-2,6,10-trien-1-yl]-6-methylcy clohexa-2,5-diene-1,4-dione 58, (6E,10E,14E)-16-(2,5-dihy droxy-3-methylphenyl)-2-hydroxy-2,6,10,14-tetramethylhexadeca-6,10,14-trien-3-one **59**, 3-[(2E,6E,10E,14R)-14,15-dihydroxy-3,7,11,15tetramethylhexadeca-2,6,10-trien-1-yl]-4-hydroxy-5-meth ylphenyl octadecanoate 60, 3-[(2E,6E,10E,14R)-14,15-dihydroxy-3,7,11,15-tetramethylhexadeca-2,6,10-trien-1-yl]-4-hydroxy-5-methylphenyl(9Z)-octadec-9-enoate 61, and 2-geranylgeranyl-6methylhydroquinone 62 are plastoquinones detected in a methanol extract of S. micracanthum collected from the Toyama Bay coast (Japan). Compounds 59-61 have a reductive effect on DPPH (3.00%, 52.6%, and 32.3%, respectively), and cytotoxicity against colon 26-L5 $(IC_{50} 1.51, 17.5, and 1.69 \mu g mL^{-1}, respectively)$ at a dose of 100 mg mL⁻¹ of the sample.⁵⁹ 2-Geranylgeranyl-6-methylhydroquinone **62** and 2-geranylgeranyl-6-methylbenzoquinone 63 were detected also in a methanol extract of S. micracanthum collected from the Toyama Bay (Japan). Compounds 62 and 63 showed an inhibitory effect on lipid peroxidation with $IC_{50} = 0.11$ and 1.0 µg mL⁻¹, respectively. While their antiviral effects were detected against HSV-1, HSV-2, HCMV, mumps virus, measles virus, adeno virus, influenza virus, polio virus, and coxsackie virus, with IC₅₀ ranging from 7.7 to 35 μM.60 Sargassumol 64 was detected in a methanol extract of S. micracanthum collected from Wando County (Korea). Sargassumol 64 showed minimal scavenging activity against DPPH radical, but exhibited the potent ABTS radical-scavenging activity with an IC50 of 47 μM, comparable to that of trolox (IC₅₀ 45 μM).⁶¹ Dihydromonooxofranesylacetone and $\Delta 10$ -(11)-

Chemical structures of compounds 67-97.

dihydromonooxofranesylacetone 66 were detected in an acetone extract of S. micracanthum collected from the coast of Gosa (Japan) (Fig. 4).62 Dihydromonooxofarnesylacetone 67, (5Z,10E)-14-hydroxy-2,6,10-trimethylpentadeca-5,10-dien-4-one (5E)-6,10,14trimethylpentadec-5-ene-2,12-dione **69**, (5*E*,10*E*)-6,10,14-trimethyl pentadeca-5,10,13-triene-2,12-dione 70, (5E,9E)-6,10,14-trimethyl pentadeca-5,9,13-triene-2,12-dione 71, (10E)-14-hydroxy-2,6,10trimethylpentadec-10-en-4-one 72, and (6E,10E)-14-hydroxy-2,6,10trimethylpentadeca-6,10-dien-4-one 73 are farnesylacetone derivatives isolated from a methanol extract of Japanese S. micracanthum. 63 (2R,8'S)-7',8'-Dihydro-9'-oxo- δ -tocotrienol 74, and (2R)-9'-oxo- δ -tocotrienol 75 were detected in a methanol extract of S. micracanthum, collected from Toyama Bay (Japan) (Fig. 5).64

32. Sargassum muticum

1-Dodecene, 1-tetradecene, 1-pentadecene, n-pentadecene, 1hexadecene, n-heptadecane, 1-octadecene, (E)-3-eicosene, n-tricosane, n-tetracosane, n-pentacosane, n-hexacosane, and nheptacosane hydrocarbons, myristic acid, pentadecanoic acid, palmitic acid 7, stearic, palmitoleic, oleic, vaccenic acid, hexadecadienoic acid, linolenic, stearidonic, arachidonic, and eicosapentaenoic fatty acids were detected in GC-MS of a chloroform extract of S. muticum collected from Plouzané (France).65 Saringosterol 11 was detected in S. muticum, which is widely distributed in Korea, and exhibited an anti-obesity effect. The inhibitory effect of saringosterol 11 on adipogenesis was

detected via Oil Red O staining in 3T3-L1 preadipocytes in a dose-dependent manner, which inhibited adipocyte differentiation and expression of adipogenic marker genes.66

33. Sargassum myriocystum

Sulfated polysaccharides extracted from S. myriocystum (SMP) were investigated for gentamicin-induced nephrotoxicity in adult zebrafish. The SMP showed maximum carbohydrate, sulfate and L-fucose contents, which suppressed mRNA expression levels of KIM-1. NF-κB, TNF-α, and IL-6 in a dosedependent manner.67

34. Sargassum naozhouense

Glutamic acid (13.21 g/100 g protein), aspartic acid (8.39 g/100 g protein), alanine(5.27 g/100 g protein), glycine (4.38 g/100 g protein), tyrosine, threonine, phenylalanine, serine, histidine, lysine, proline, arginine, tryptophan, valine, methionine, cysteine, isoleucine, and leucine comprised the amino acid composition of cultivated S. naozhouense, collected from Techeng Island (China), while the main fatty acids in the cultivated S. naozhouense were myristic acid, palmitic acid, oleic acid, and arachidonic acid. Water-soluble polysaccharides were detected in an extract of S. naozhouense, which contained sulfate groups, and exhibited strong antiviral activity against HSV-1 strain, with an IC₅₀ of 8.92 µg mL⁻¹.68 Sargassumone 76, (2R,6S,8S,9S)-hexahydro-2,9-dihydroxy-4,4,8-trimethyl-6-acetyloxy-3(2H)-benzofuranone 77, (6S,8S,9R)-hexahydro-6,9dihydroxy-4,4,8-trimethyl-2(2H)-benzofuranone 78, (6S,8S,9R)hexahydro-6,9-dihydroxy-4,4,8-trimethyl-2(2H)-benzofuranone 79, loliolide 80, (+)-epiloliolide 81, and spheciospongones A 82 as norisoprenoid derivatives and a highly oxygenated cyclopentene were detected in S. naozhouense (Fig. 5).69

35. Sargassum natans

Sodium alginate was detected in S. natans collected from both Manzanilla and Mayaro Bays (Trinidad).70

36. Sargassum oligocystum

Fucosterol 5, cholesterol 6, 24-hydroperoxy-24-vinylcholesterol 24, a mixture of 24(S)-hydroxy-24-vinylcholesterol 38, and 24(R)-hydroxy-24-vinylcholesterol 39, 22-dehydrocholesterol 83, 29-hydroperoxystigmasta-5,24(28)-dien-3β-ol 84, and ostreasterol 85 were detected in S. oligocystum collected from the Persian Gulf (Fig. 5). A chloroform extract of S. oligocystum was moderately effective against A. salina nauplii (LC₅₀ = 159 μ g mL⁻¹).⁷¹ A water extract of S. oligocystum collected from the Persian Gulf was detected to have antitumor activity against K562 and Daudi cell lines at concentrations of 500 μg mL⁻¹, and 400 μg mL⁻¹, respectively.⁷²

Sargassum pallidum 37.

An ethanol crude extract of S. pallidum collected from Weihai (China) showed antioxidant activity against DPPH.73

Chemical structures of compounds 99–108.

38. Sargassum patens

The anti-inflammatory effect of an ethanol extract of *S. patens* (SPEE) collected from Busan (Korea) was detected. The production of NO was suppressed by SPEE to 28%, at 100 μg mL⁻¹, and the levels of IL-6, TNF- α , and IL-1 β were dose-dependently suppressed. *In vivo*, a croton oil-induced mouse ear edema was attenuated by SPEE and the infiltration of mast cells into the tissue was suppressed. ⁷⁴ A sulfated polysaccharide was detected in *S. patens*, and was reported to inhibit the replication of herpes simplex virus type 2 (HSV-2) in a dose-dependent manner by 38.5–96.1%, of the control level, after incubation with 0.78–12.5 μg mL⁻¹ of the polysaccharide. The sulfated polysaccharide exhibited extracellular virucidal activity only in high concentrations (\geq 12.5 μg mL⁻¹), but suppressed the virus attachment to its host cells by 45.1%, at a concentration < 1 μg mL⁻¹.⁷⁵

39. Sargassum paradoxum

Fallahydroquinone **30**, fallaquinone **31**, sargahydroquinoic acid **86**, chabrolohydroxybenzaquinone A **87**, chabrolohydroxybenzaquinone C **88**, paradoxhydroquinone **89**, 2-[11-(hydroxymethyl)-3,7,15-trimethyl-2,6,10,14-hexadecatetraen-1-yl]-6-methyl-1,4-benzenediol **90**, sargaquinal **91**, sargahydroquinal **92**, paradoxquinol **93**, sargahydroquinoic acid **94**, 2-[11-(hydroxymethyl)-3,7,15-trimethyl-2,6,10,14-hexadecatetraen-1-yl]-6-methyl-1,4-benzoquinone, **95**, sargaquinone **96**, paradoxquinone **97**, and (3'E,7'Z)-9-(6-hydroxy-2,8-dimethyl-2*H*-1-benzopyran-2-yl)-6-methyl-2-(4-methyl-3penten-1-yl)-2,6-nonadi enal **98** were detected in a crude extract of *S. paradoxum* collected from Port Phillip (Australia) (Fig. 5).⁷⁶

40. Sargassum parvivesiculosum

1,3-Di-O-[2',2'-di-(p-phenylene) isopropylidene]glycerol **99**, (2S)-1-O-heptatriacontanoyl glycerol **100**, and (2S)-1,2-di-O-palmitoyl-3-O-(6-sulfo- α -D-quinovopyranosyl)glycerol **101** are glycerol derivatives isolated from an ethanol extract of S. parvivesiculosum collected from Sanya (China) (Fig. 6).

41. Sargassum polycystum

The total yield of fucoidan isolated from *S. polycystum* was 4.51 \pm 0.24%, and contained 46.8% of L-fucose and 22.35% \pm 0.23% of sulfate respectively. The cytotoxicity effect of fucoidan, showed a higher percentage (90.4% \pm 0.25%) of inhibition against the MCF-7 cell line, at 150 μg mL⁻¹, with an IC₅₀ of 50 μg mL⁻¹.78

42. Sargassum ringgoldianum

An extract of S. ringgoldianum collected from Jeju Island (Korea), showed the inhibition of α -glucosidase and α -amylase, and alleviated postprandial hyperglycemia in streptozotocininduced diabetic mice. The IC $_{50}$ values of S. ringgoldianum extract against α -glucosidase and α -amylase were 0.12, and 0.18 mg mL $^{-1}$, respectively. The blood glucose levels of the S.

ringgoldianum extract in the administered group were significantly lower in the streptozotocin-induced diabetic mice compared to in the control group.⁷⁹

43. Sargassum sagamianum

An ethanol extract of S. sagamianum collected from Yeonhwari (Korea) was detected to include an anti-inflammatory agent, whereby the rate of formation of edema in a mouse ear model was reduced by 46%, using a dose of 250 mg kg⁻¹. The ethanol extract showed potent inhibitory effects on the LPS-induced expression of inflammatory mediators, such as NO, iNOS, COX-2, and cytokines, in macrophages through suppression of the NF-κB p65 pathway.80 An ethanol extract of S. sagamianum collected from Jeju Island (Korea) was investigated on INS-1 pancreatic β cells against high glucose-induced oxidative stress and apoptosis at high concentrations (30 mM) causing β cell apoptosis, whereby treatment with SSE protected the β cells from high glucose-induced damage by recovering the cell viability. Treatment with the ethanol extract at concentrations of 10-100 µg mL⁻¹ decreased lipid peroxidation, intracellular ROS, and nitric oxide levels, as well as increased cell viability and insulin secretion in high glucose pre-treated INS-1 cells, in a dose-dependent manner.81 1-Octadecatetraenoyl glycerol 102 is a polyunsaturated fatty acid-derived monoglyceride detected in S. sagamianum collected from Jeju Island (Korea). A series of monoglycerides were synthesized using glycerol and various fatty acids. Several synthesized compounds showed a moderate to significant inhibition of phospholipase A2 and cyclooxygenase-2.82 Sargachromenol 55, sargaquinoic acid 57, dihydromonofarnesylacetone 65, and monooxofarnesylacetone 103 were isolated from a methanol extract of S. sagamianum. These compounds showed moderate acetylcholinesterase (AChE) inhibitory activity in a micromole range.83 In vitro, treatment with sargachromenol 55 and sargaquinoic acid 57 promoted cell death and the activation of caspase-3, caspase-8, caspase-9, and poly (ADP-ribose) polymerase (PARP) in a concentrationdependent manner. Sargaquinoic acid- or sargachromenolinduced apoptosis was enhanced by co-treatment with UVB irradiation. The topical application of sargaquinoic acid (1 mg mL⁻¹) before UVB irradiation (2.5 kJ m⁻²) to hairless mice also enhanced apoptosis, including the activation of caspase-3.84 15'-Hydroxysargaquinolide 104, 11'-hydroxysargachromelide 105, 15'-methylenesargaquinolide 106, chromequinolide 107, and (2'E,5'E)-2-methyl-6-(7'-oxo-3'-methylocta-2',5'-dienyl)-1,4benzoquinone 108 were detected in S. sagamianum collected from Manazuru in the Kanagawa prefecture. Compounds 104, 105, and 108, had antibacterial activity against S. aureus, (MIC 16, 128, and 2 μg mL⁻¹, respectively) (Fig. 6).85

44. Sargassum serratifolium

An ethanol extract of *S. serratifolium* collected from Busan (Korea) ameliorated paw swelling, reduced the arthritis score, decreased the secretion of pro-inflammatory cytokines in the serum and joint tissue, and suppressed the collagen-induced rheumatoid arthritis of mice. The ethanol extract showed a downregulated

This article is licensed under a Creative Commons Attribution 3.0 Unported Licence.

Open Access Article. Published on 01 July 2020. Downloaded on 12/10/2025 12:33:11 AM.

RSC Advances Review

NF-kB signaling pathway by suppressing the phosphorylation of protein kinase B, c-Jun N-terminal kinase, and p38 mitogenactivated protein kinases.86 An ethanol extract of S. serratifolium exhibited potential antimicrobial activity against pathogenic commensal bacteria related to acne vulgaris (P. acnes, S. epidermidis, S. aureus, and P. aeruginosa), and C. albicans, which causes cutaneous candidiasis. Among the solvent-soluble fractions from the ethanol extract, the *n*-hexane fraction showed the strongest antimicrobial activity against all the tested human skin pathogens (MIC from 32 to 512 μg mL⁻¹).87 An ethanol extract of S. serratifolium, collected from Tongyoung (Korea), which mainly contained sargachromenol 55 and sargaquinoic acid 57, efficiently suppressed adipocyte differentiation and lipid accumulation in 3T3-L1 cells by downregulating the proteins involved in cell cycle progression and adipogenesis, and also downregulated the key transcription factors, such as C/EBPβ, C/EBPα, PPARγ, RXRα, SREBP1c, and STAT3, which were responsible for the observed suppression of lipid accumulation upon treatment with an ethanol extract of S. serratifolium.88

45. Sargassum siliquastrum

Sargachromanol A–P **109–124** are meroterpenoids detected in *S. siliquastrum*. These compounds exhibited significant antioxidant activity in the DPPH assay. Compounds **115** and **123** also

showed inhibitory activity toward butylcholine esterase.89 Sargachromanol O 125 and sargachromanol R 126 were detected in S. siliquastrum collected from Cheju Island (Korea). Sargachromanol R 126 had potent cytotoxic activity against AGS, HT-29, and HT-1080 cell lines, with IC50 values of 6.5, 3.4, and 13.9 μg mL⁻¹, respectively, compared with paclitaxel and doxorubicin.90 Mojabanchromanol 127 was detected in an ethanol extract of S. siliquastrum collected from the seashore of Pusan (Fig. 7).91 Isonahocol E3 128, detected in S. siliquastrum, has functional antagonistic activities against ET-1 induced inflammatory and pro-angiogenic effects, through inhibition of ET-1induced cell proliferation, as well as inflammatory mediators, like IL-6, IL-8, and TNF-α, and pro-angiogenic factors, including metalloproteinases in immortalized human keratinocytes. Isonahocol E3 128 also reduced the expression level of endothelin ETA receptor and endothelin ETB receptor, as well as suppressed ET-1-induced extracellular signal-regulated kinase (ERK) phosphorylation.92 Sargachromanol D 112, sargachromanol E 113, sargachromanol K 119, sargachromanol P 124, (9S,10S)-13-(3,4-dihydro-6-hydroxy-2,8-dimethy-2H-1-benzopyran-2-yl)-2,6,10-trimethyl-trideca-(2E,6E)-diene-4,5,10-triol (9S,10R)-13-(3,4-dihydro-6-hydroxy-2,8-dimethy-2H-1benzo-pyran-2-yl)-2,6,10-trimethyl-trideca-(2E,6E)-diene-4,5,10triol 130, and 9-(3,4-dihydro-6-hydroxy-2,8-dimethy-2H-1-benzopyran-2-yl)-2,6-dimethyl-(6E)-nonenoic acid 131 were detected

Fig. 7 Chemical structures of compounds 109-127.

in S. siliquastrum collected from JeJu Island (Korea). Sargachromanols A 109, B 110, and K 119, had strong cytotoxicity against AGS, HT-29, HT-1080, and MCF-7 cell lines, with IC50 values of 0.7, 6.1, 0.7, and 28.1 μg mL; 0.5, 1.0, 3.3, and 23.6 μg mL^{-1} and 5.7, 0.8, 1.8, and 10.3 $\mu g mL^{-1}$, respectively, comparable with paclitaxel and doxorubicin.93 (6E,10E)-16-(2,5-Dihydroxy-3-methylphenyl)-4,14-dihydroxy-2,6,10,14tetramethylhexadeca-2,6,10-trien-5-one 132, methyl(5-hydroxy-2-{[(6E,10E,13S)-13-hydroxy-3,7,11,15-tetramethyl-12oxohexadeca-1,6,10,14-tetraen-3-yl]oxy}phenyl)acetate 133, methyl(5-hydroxy-2-{[(6E,10E,12S)-12-hydroxy-3,7,11,15tetramethyl-13-oxohexadeca-1,6,10,14-tetraen-3-yl]oxy}phenyl) acetate 134, methyl(5-hydroxy-2-{[(6E,10Z,12R)-12-hydroxy-3,7,11,15-tetramethyl-13-oxohexadeca-1,6,10,14-tetraen-3-yl] oxy\phenyl)acetate 135, methyl(5-hydroxy-2-{[(6E,13E)-12hydroxy-3,7,11,15-tetramethylhexadeca-1,6,13,15-tetraen-3-yl] oxyphenyl)acetate 136, methyl $[2-({(6E)-3,7-dimethyl-8-[(1R,5R)$ 3-methyl-5-(2-methylprop-1-en-1-yl)-4-oxocyclopent-2-en-1-yl] octa-1,6-dien-3-yl}oxy)-5-hydroxyphenyl]acetate 137, methyl{2,5dihydroxy-3-[(2E,6E,10E,13S)-13-hydroxy-3,7,11,15-tetramethyl-12-oxohexadeca-2,6,10,14-tetraen-1-yl]phenyl}acetate methyl{2,5-dihydroxy-3-[(2E,6E,13S)-13-hydroxy-3,7,11,15-

tetramethyl-12-oxohexadeca-2,6,14-trien-1-yl]phenyl}acetate

139, methyl{2,5-dihydroxy-3-[(2Z,10E,13S)-13-hydroxy-3,7,11,15-

tetramethyl-6-oxohexadeca-2,10,14-trien-1-yl]phenyl}acetate **140** and methyl{1-[(2*E*,6*E*,10*E*,12*R*,13*S*)-12,13-dihydroxy-3,7,11,15-tetramethylhexadeca-2,6,10,14-tetraen-1-yl]-2,5-dioxocyclohex-3-en-1-yl}acetate **141** were detected in *S. siliquastrum*. Compounds **132–141** were revealed to have a common tetraprenyl hydroquinone structure, which belonged to the nahocol, isonahocol, and sargahydroquinoic acid classes. The dihydroquinone moiety of **141** was unique and unprecedented in a brown alga. These compounds exhibited moderate to significant radical-scavenging activity as well as weak inhibitory activities against sortase A, and isocitrate lyase (Fig. 8).⁹⁴

46. Sargassum siliquosum

Methanol extracts of *S. siliquosum* (ME) and a fucoxanthin-rich fraction (FRF) showed antioxidant activity, with IC₅₀ values of 0.2, and 0.04 mg mL⁻¹, respectively. Also, the IC₅₀ values for the angiotensin-converting enzyme (ACE) inhibitory activity of ME (IC₅₀ = 0.03–0.42 mg mL⁻¹) was lower than that of FRF (0.94–1.53 mg mL⁻¹). CE and FRF also showed α-glucosidase inhibitory activity, with IC₅₀ values of 0.57, and 0.50 mg mL⁻¹, respectively.⁹⁵

Fig. 8 Chemical structures of compounds 128-141.

47. Sargassum spinuligerum

Deshydroxytetrafuhalol-A, tetrafuhalol-A, pentafuhalol-A, hexafuhalol-A, heptafuhalol-A, octafuhalol-A, nonafuhalol-A and undecafuhalol-A, the acetylated derivatives of deshydro xytetrafuhalol-C, deshydroxyhexafuhalol-A, deshydroxyhexafuhaloland deshydroxyhexafuhalol-D, deshydroxyoctafuhalol-C, octafuhalol-B, decafuhalol-A, dodecafuhalol-A, tetradecafuhalol-A, hexadecafuhalol-A, octadecafuhalol-A and eicosafuhalol-A were the largest phlorotannins found in an ethanol extract of S. spinuligerum, collected from Whangaparoa Island, (Auckland).96 Phlorotannins, fucophlorethol-B octa-acetate, fucodiphlorethol-B, fucodiphlorethol-D and fucodiphlorethol-F deca-acetate, hydroxyfucodiphlorethol-A undeca-acetate, bisfucotriphlorethol-A pentadeca-acetate, hydroxybisfucophlorethol-A hexadeca-acetate, bisfucotetra phlorethol-A heptadeca-acetate, dihydroxyfucotriphlorethol-A and dihydroxyfucotriphlorethol-B tetradeca-acetate, bisfucopenta phlorethol-B nonadeca-acetate, chlorobisfucopentaphlorethol-A nonadeca-acetate, difucodiphlorethol-A trideca-acetate, fucodi fucotetraphlorethol-A icosa-acetate, bisfucotriphlorethol-A penta deca-acetate, chlorobisfucopentaphlorethol-A nonadeca-acetate , and fucodifucotetraphlorethol-A icosa-acetate were detected in S. spinuligerum.97 Pseudotrifuhalol-A, pseudotetrafuhalol-A, pseudopentafuhalols-A-D, pseudohexafuhols-A-C, pseudoheptafuhalols-A-D and psuedooctafuhalols-B-D were detected in an ethanolic extract of S. spinuligerum.98

48. Sargassum swartzii

Sulfated polysaccharide extracted (SPE) from *S. swartzii* was collected from a coastal region of Mandapam (India), with a molecular weight of 50 kDa, and with a high percentage of carbohydrate (7.40% \pm 0.63%), followed by sulfate (5.3% \pm 1.54%). The antioxidant activity of SPE detected for ABTS, $\rm H_2O_2$ and DPPH, was 55% \pm 3.61%, 47.23% \pm 2.81%, and 25.33% \pm 2.52%, respectively. Bioactive fucoidan fractions (CFF, FF1, and FF2) were detected in *S. swartzii* collected from the Gulf of Mannar (India), which contained mainly sugars, sulfate, and uronic acid. Fraction FF2 was found to exhibit significant anti-HIV-1 activity at concentrations of 1.56 μg mL $^{-1}$ as observed by a >50% reduction in HIV-1 p24 antigen levels and reverse transcriptase activity. 100

49. Sargassum tenerrimum

Sulfated polysaccharides, detected in *S. tenerrimum* collected from Mandapam (India), showed a higher percentage of carbohydrate (8.20% \pm 1.23%) followed by sulfate (6.6% \pm 1.42%) and protein (0.86% \pm 0.42%). The free radical scavenging potential was found to be higher in ABTS (70.33% \pm 2.33%) followed by DPPH (64.66% \pm 2.08%) and H₂O₂ (61.56% \pm 2.05%). The total antioxidant capacity (TAC) was found to be 62.55% \pm 1.40%. ¹⁰¹

50. Sargassum thunbergii

3-{[5-Deoxy-5-(trimethylarsonio) pentofuranosyl]oxy}-2-hydroxy propyl sulfate **142**, arseno-sugar was detected in *S. thunbergii* collected from Nakaminato (Japan). Sargachrominol 55, sargaquinoic acid 57, sargahydroquinoic acid 94, and sargathunbergol A **143** were detected in *S. thunbergii* collected from Youngdo Island (Korea), which exhibited antioxidant activities (EC₅₀ values of 32, 27,20, and 38 μ g mL⁻¹ respectively), compared to BHT (EC₅₀ = 42 μ g mL⁻¹) and α -tocopherol (EC₅₀ 23 μ g mL⁻¹). Sargahydroquinoic acid **95**, sargaquinoic acid **2**, and sargachromenol 55, thunbergol A **144**, and thunbergol B **145**, were detected in *S. thunbergii* collected from Busan (Korea). The evaluating capacity of compounds **144**, and **145** to scavenge DPPH radical, showed they exhibited EC₅₀ values of 30 and 31 μ g mL⁻¹, respectively.

(2S)-1-O-(5Z,8Z,11Z,14Z,17Z-Eicosapentaenoyl)-2-O-(9Z,12Z,15Z -octadecatrienoyl)-3-O-β-D-galactopyranosyl-sn-glycerol 146 and (2S)-1-O-(9Z,12Z,15Z-octadecatrienoyl)-2-O-(6Z,9Z,12Z,15Z-octade catetraenoyl)-3-O-β-D-galactopyranosyl-sn-glycerol 147 were glycolipids detected in the methanol extract of S. thunbergii, collected from the West Sea in Korea.¹⁰⁵ Thunberol 148, along with 24ethylcholesta-4,24(28)-dien-3-one 149, stigmasta-5,28-dien-3β-ol 150, cholesta-5,23-dien-3β,25-diol 151, and cholesta-5,14-dien-3β-ol 152, were detected in S. thunbergii, collected from Nanji Island (China). Thunberol 148 exhibited significant inhibitory activity against protein tyrosine phosphatase 1B, a potential compound target for the treatment of Type-II diabetes and obesity, with an IC50 value of 2.24 µg ml⁻¹.106 (STCs)—indole-2-carboxaldehyde (STC-1) 153, indole-3-carboxaldehyde (STC-2) 154, indole-4-carboxaldehyde (STC-3) 155, indole-5-carboxaldehyde (STC-4) 156, indole-6carboxaldehyde (STC-5) 157, and indole-7-carboxaldehyde (STC-6) 158 were detected in S. thunbergii and their inhibitory effects were evaluated on adipocyte differentiation in 3T3-L1 cells. STC-1 and STC-5 showed non-toxic inhibition of the differentiation of 3T3-L1 adipocytes. STC-1 and STC-5 significantly inhibited lipid accumulation and downregulated the expression of peroxisome proliferatoractivated receptor-γ (PPARγ), CCAAT/enhancer-binding protein α (C/EBPα), and sterol regulatory element-binding protein 1c (SREBP-1c) in a dose-dependent manner. The specific mechanism mediating the effects of STC-1 and STC-5 was shown to be AMPactivated protein kinase (AMPK) activation. The inhibitory effect of STC-1, and STC-5 on adipogenesis was through activation of the AMPK signal pathway. STC-1 and STC-5 may be effective candidates for the prevention of obesity or obesity-related diseases. 107 (+)-Epiloliolide 81, (-)-loliolide 80, and apo-9'-fucoxanthinone 159 norisoprenoids were detected in S. thunbergii, collected from Korea. 108 1-(5-Acetyl-2,4-dihydroxyphenyl)-3-methylbutan-1-one 160 and 1-(5acetyl-2-hydroxy-4-methoxyphenyl)-3-methylbutan-1-one 161 were detected in a methanol extract of S. thunbergii, collected from Weihai City (China).109 Saringosterol 11, chlorophyll a 162, and isofucosterol 163, were also isolated from the chloroform fraction of S. thunbergii extract (Fig. 9).110

51. Sargassum trichophyllum

Laminaran, alginate, and fucoidan were detected in *S. tricho*phyllum. Laminaran was found to be a β -glucan consisting of terminal- (14.5 mol%), 1,3- (69.4%), 1,6- (33.4%), and 1,3,6linked (12.8%) residues. Alginate was found to be a mannuronic acid-rich alginate (M/G = 1.88). Fucoidan consisted of L-fucose (79.1 mol%) and p-galactose (19.9 mol%), and its sulfate content was estimated to be 25.5%, with antiviral activity against the herpes simplex virus type 2.111

52. Sargassum turbinarioides

Alginate was detected in S. turbinarioides collected from Nosy Bey (Madagascar), with a molecular weight of $5.528 \times 10^5 \mathrm{g}$ mol^{-1} .112

Sargassum vachellianum 53.

Fucoidan-rich polysaccharide extract (SPS) and polyphenol-rich extract (SPP) from S. vachellianum, collected from Nan'ao Island (China), showed a protective effect on the skin from UV damage. SPP showed good free radical scavenging ability, antimicrobial activity against E. coli and S. aureus, and effectively absorbed UVB and UVA rays. Whereas SPS hardly absorbs UVA and UVB rays and showed weak free radical scavenging ability and no antimicrobial activity. SPS showed considerable inhibition of tyrosinase (51.21%) and had better moisture absorption (52.1%) and retention (63.24%) abilities than SPP. 113 (4Z,9Z)-4-Methyl-

Chemical structures of compounds 142-162.

1,2,6,8-tetraazacycloundeca-4,9-diene-3,7,11-trione **164** is an eleven-membered macrocyclic hydrazide detected in *S. vachellianum* collected in the South China Sea (Fig. 10).¹¹⁴

54. Sargassum wightii

1-0 Palmitoyl-3-0(6'-sulfo-alpha-quinovopyranosyl)-glycerol **165**, sulfoglycerolipid, was detected in a methanol extract of *S. wightii*, and showed antibacterial activity against *Xanthomonas oryzae* pv. *oryzae*, which causes bacterial blight of rice. ¹¹⁵ 4-(8-Ethyl-tetrahydro-7-oxo-2*H*-pyran-5-yl)-propyl-4'-methylbenzoate **166** and methyl-2-(12-oxo-7-phenyl-8-vinyl-1-oxa-4,9-cyclododecadien-3-yl)-acetate **167**, detected in the methanol extract of *S. wightii*, showed antioxidant activities against DPPH radical scavenging, with (IC so values of 0.24–0.32 mg mL $^{-1}$), compared to α -tocopherol (IC 50 0.63 mg mL $^{-1}$). Also, **166** and **167** showed 5-lipoxygenase

inhibitory activity with IC₅₀ values of 0.56 and 0.29 mg mL⁻¹, respectively. 116 2\alpha-Hydroxy-(28,29)-frido-olean-12 13, 21(22)-dien-20-propyl-21-hex-40(E)-enoate 168, and 2α-hydroxy-(28,29)-fridoolean-12 13, 21(22)-dien-20-prop-2-(E)-en-21-butanoate 169, fridooleanene triterpenoids derivatives, 2α-hydroxy-8(17),12E,14-labdatriene 170, and 3β , 6β , 13α -trihydroxy-8(17), 12E, 14-labdatriene 171, labdane diterpenoids, were detected in an extract of S. wightii, were shown to be potential anti-hyperglycaemic pharmacophore leads to reduce the risk of elevated postprandial glucose levels. The oxygenated labdane diterpenoids displayed significantly lesser antioxidant and tyrosine phosphatase-1B inhibitory properties than those exhibited by the frido-oleanenes. 117 3"-Isopropyl-3c-{3b-[(2-oxo-3,4-dihydro-2*H*-chromen-3-yl)methyl]butyl}-2"-butenyl-3'hydroxy-2'-(2'b-methoxy-2'-oxoethyl)-3',4'-dihydro-2H-pyran-4'carboxylate 172, 2c-methylbutyl-6-[6c-(benzoyloxy)propyl]-6-methyltetrahydro-2H-pyran-2-carboxylate 173, 6-{6b-[3'-(5'a-methyl

Fig. 10 Chemical structures of compounds 163-179.

Review

Oxygenated cyclopentene ■ Farnesvlacetone derivatives ■ Glycerol derivatives Skin protection ■ Plastoquinones ■ Anti-diabetic ■ Hvdrocarbons Anti-inflammatory Fatty acids Antibacterial Amino acids Meroditerpenoids Antioxidant ■ Meroterpenoids Antiproliferative Sterols Other ■ Polyphenolic Other В

Fig. 11 Secondary metabolites (A) and their reported bioactivity (B) produced by Sargassum species

propyl)-3',4'-dihydro-2*H*-pyran-6'-yl]ethyl}-tetrahydro-2*H*-pyran-2one 174 and 7-(7c-methylpentyl)-3,4,6,7,8,8a-hexahydro-2Hchromen-2-one 175 are hitherto undescribed O-heterocyclic derivatives with use natural antioxidant and antihypertensive functional food supplements and are also utilized as therapeutic leads in antihypertensive management as angiotensin-converting enzyme inhibitors (Fig. 10).118

55. Sargassum yezoense

Meroterphenol A 176, meroterphenol B 177, meroterphenol C 178, and meroterphenol D 179, plastoquinones, were detected in S. yezoense, with the structures of these compounds characterized by a 6-methyl-1,4-benzohydroquinone moiety with an oxygenated diterpenoic acid chain. Compounds 176-179 showed potent activation effects on the peroxisome proliferatoractivated receptor gamma (PPARy) (Fig. 10).119

56. Conclusion

Sargassum is an important seaweed that is widely and excessively distributed in tropical and subtropical regions and has been reported to contain 537 species, with 358 of them accepted taxonomically. Sargassum species are used in many folk applications in human nutrition and are considered a rich source of vitamins, carotenoids, proteins, and minerals. Significant progress has been detected in the publication rates of studies mentioning the genus Sargassum over the past five years (Fig. 11), with many bioactive compounds reported with diverse chemical structures, from oxygenated cyclopentene, farnesylacetone, glycerol derivatives, plastoquinones, hydrocarbons, fatty acids, amino acids, to meroditerpenoids, meroterpenoids, sterols, polyphenolic, and sulfated polysaccharides (Fig. 2-10). According to the reported data concerning the phytochemical metabolites identified in the different Sargassum spp., the sulfated polysaccharides and polyphenolic compounds represent 27% and 20% of the major isolates, followed by meroterpenoids, sterols 9%, meroditerpenoids 8%, and amino acids 6%, reflecting their history as a nutrient in folk use (Fig. 11). These isolated compounds and/or extracts exhibit

diverse biological activities; such as skin protection for the polysaccharide rich extract of S. vachellianum, and S. fusiforme, whereas the polysaccharide hardly absorbs UVA, UVB rays, and has shown considerable inhibition on tyrosinase (51.21%), as well as better moisture absorption (52.1%) and retention (63.24%) abilities. Also, Sargassum spp. contains mainly polysaccharides, and polyphenolic compounds sulfated (Fig. 11), reflecting major reports of their analgesic, antiinflammatory 15%, antioxidant 18%, neuroprotective, antimicrobial 15%, anti-tumor 23% activities. The polysaccharide extracted from S. asperifolium, S. hemiphyllum, and S. latifolium, with S. fulvellum, S. macrocarpum, S. horneri, S. patens, S. sagamianum, and S. serratifolium possess anti-inflammatory activity, through a significant inhibition of nitric oxide generation, and LPS-induced TNF-α, IL-12 p40, IL-6, causing a strong inhibitory effect against NF-kB activation. While isonahocol E3 128, detected in S. siliquastrum, has functional antagonistic activities against ET-1 induced inflammation. Moreover, S. angustifolium, S. boveanum, and fucoidan S. polycystum extracts have been reported to have a cytotoxic effect against HeLa (cervical cancer) and MCF-7 (breast cancer) cells, where the cell survival is inversely proportional to the increase in the concentration of the extracts. Polysaccharides extracted from S. latifolium showed a selective cytotoxicity against lymphoblastic leukemia. 28S-Epoxy-24-ethylcholesterol 26 showed cytotoxic activity against MCF-7, HCT-8, 1A9, HOS, and PC-3 with IC50 values of 4.0, 8.8, 10.0, 10.0, and 7.2 μg mL⁻¹, respectively. Sargachromanols A 109, B 110, and K 119 had strong cytotoxicity against AGS, HT-29, HT-1080, and MCF-7 cell lines, with IC50 values of 0.7, 6.1, 0.7, and 28.1 μ g mL; 0.5, 1.0, 3.3, and 23.6 μ g mL⁻¹, and 5.7, 0.8, 1.8, and 10.3 $\mu g \text{ mL}^{-1}$, respectively. 28 ξ -Dihydroxy-24ethylcholesta-5,23*Z*-dien 21, and 24ξ-hydroperoxy-24vinylcholesterol 24 were reported to have cytotoxic activities against HL-60, with IC₅₀ values of 7.8 and 8.5 μg mL⁻¹, respectively. While, fucosterol 5 and 24-ethylcholesta-4,24(28)dien-3,6-dione 23 showed potent cytotoxic activities against P-388, with IC_{50} values of 0.7 and 0.8 μg mL⁻¹, respectively. Sargachromanol R 126 had potent cytotoxic activity against AGS, HT-29, and HT-1080 cell lines, with IC50 values of 6.5, 3.4, and 13.9 μg mL⁻¹, respectively, compared with paclitaxel and

RSC Advances Review

doxorubicin. Also, fucoidan extracted from S. glaucescens, S. siliquosum, S. swartzii, S. tenerrimum, and S. pallidum exhibited antioxidant activities in a dose-dependent manner against DPPH. A hot-water extract of S. hemiphyllum collected from the coast of Penghu County (Taiwan) had antioxidant activity and an immune-stimulating one. The antioxidant activity was showed to be increased related to a concentration <3.5 mg mL⁻¹. Heteropolysaccharides extracted from S. integerrimum were shown to have neuroprotective and antioxidant activities. Sargachromanol A-P 109-124, isolated from S. siliquastrum, exhibited significant antioxidant activity in the DPPH assay. Sargachrominol 55, sargaquinoic acid 57, sargahydroquinoic acid 94, and sargathunbergol A 143 were detected in S. thunbergii, and showed antioxidant activities (EC₅₀ values of 32, 27, 20, and 38 $\mu g \text{ mL}^{-1}$ respectively), compared to BHT (EC₅₀ = 42 $\mu g \text{ mL}^{-1}$) and α -tocopherol (EC₅₀ = 23 $\mu g \text{ mL}^{-1}$). Also, thunbergol A 144, and thunbergol B 145, isolated from S. thunbergii, were showed to scavenge DPPH radicals, with EC50 values of 30 and 31 µg mL⁻¹, respectively. Additionally, an extract of S. fulvellum (SFEE) and grasshopper ketone 32 was shown to have an inhibitory effect on atopic dermatitis (AD)-like skin lesions in BALB/c mice by regulating immune mediators and cells and may be a potentially effective alternative therapy for AD. The sulfated polysaccharides from a hot-water crude extract of S. kjellmanianum showed an immune-regulatory effect, through the enhanced immune function of immunocytes and increased the proliferation rate of spleen lymphocytes and peritoneal macrophages up to 53.56% and 51.41%. Sargassum spp., isolated compounds and/or extracts, also exhibited anti-diabetic, fibrinolytic, anti-coagulant, hepatoprotective, and anti-viral activities. Despite the wide diversity in the reported biological activates for Sargassum spp., only 54/537 species, representing about 10.05% from the species accepted taxonomically, were detected as having chemically and pharmacologically relevant activities. Consequently, further studies on the remaining species, their constituents, and biological activities are needed to exploit their maximum therapeutic potential in the field of medicinal and pharmaceutical sciences for novel and fruitful applications.

In 2007, the genus Sargassum extracts were the focus of a patent as a cosmetic product (KR1020090064079A) for their activities in lowering photo-induced cytotoxicity, whitening effect, inhibiting MMP-1 biosynthesis in human fibroblasts, and promoting the synthesis of biosynthesis type 1 procollagen, thereby improving skin elasticity and fine wrinkles, and effectively inhibiting the photo-aging phenomenon promoted by ultraviolet rays, $etc.^{120}$ Specially Sargassum horneri (KR1020120065631A), which has a deep relation to the photoaging inhibitory effect on skin cells using the chromenes compound derived from it, increased the expression of collagen synthesis markers, such as procollagen and type I collagen in UV-A treated fibroblasts, and also enhanced elastin by modulating elastase activity, and thus could be used as a material and cosmetic for improving skin wrinkles and preventing wrinkles by UV.121 Also, Sargassum natans was part of a patent for removing gold ions from aqueous solution or suspension using a biomass-derived from it, and this process can be utilized to remove gold from industrial or natural waters.¹²²

Conflicts of interest

The authors declare there were no conflicts of interest.

References

- 1 L. Liu, M. Heinrich, S. Myers and S. A. Dworjanyn, Towards a better understanding of medicinal uses of the brown seaweed *Sargassum* in traditional Chinese medicine: a phytochemical and pharmacological review, *J. Ethnopharmacol.*, 2012, 142, 591–619, https://www.sciencedirect.com/science/article/pii/S0378874112003686.
- 2 L. Mattio and C. E. Payri, 190 Years of *Sargassum* Taxonomy, Facing the Advent of DNA Phylogenies, *Bot. Rev.*, 2011, 77, 31–70, https://link.springer.com/article/10.1007/s12229-010-9060-x.
- 3 S. R. Yende, U. N. Harle and B. B. Chaugule, Therapeutic potential and health benefits of *Sargassum* species, *Pharmacogn. Rev.*, 2014, **8**, 1, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3931196/.
- 4 G. Vaseghi, M. Sharifi, N. Dana, A. Ghasemi and A. Yegdaneh, Cytotoxicity of *Sargassum angustifolium* Partitions against Breast and Cervical Cancer Cell Lines, *Adv. Biomed. Res.*, 2018, 7, 43, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5887695/.
- 5 N. J. Borazjani, M. Tabarsa, S. You and M. Rezaei, Purification, molecular properties, structural characterization, and immunomodulatory activities of water soluble polysaccharides from *Sargassum angustifolium*, *Int. J. Biol. Macromol.*, 2018, **109**, 793–802, https://www.sciencedirect.com/science/article/pii/S0141813017313624.
- 6 S. B. Nasab, A. Homaei and L. Karami, Kinetic of α-amylase inhibition by *Gracilaria corticata* and *Sargassum angustifolium* extracts and zinc oxide nanoparticles, *Biocatal. Agric. Biotechnol.*, 2020, 23, 101478, https://www.sciencedirect.com/science/article/pii/S1878818119318109.
- 7 S. S. Moni, M. F. Alam, H. A. Makeen, H. A. Alhazmi, M. Sultan, R. Siddiqui, A. Jabeen, S. Sanobar, M. S. Alam, Z. U. Rehman and M. E. Elmobark, Solvent extraction, spectral analysis and antibacterial activity of the bioactive crystals of *Sargassum aquifolium* (Turner) C. Agardh from Red Sea, *Nat. Prod. Res.*, 2019, 1–5, https://www.tandfonline.com/doi/abs/10.1080/14786419.2019.1645659.
- 8 M. I. Bilan, N. E. Ustyuzhanina, A. S. Shashkov, T. T. T. Thanh, M. L. Bui, T. T. Van Tran and A. I. Usov, Sulfated polysaccharides of the Vietnamese brown alga *Sargassum aquifolium* (Fucales, Sargassaceae), *Carbohydr. Res.*, 2017, 449, 23–31, https://www.sciencedirect.com/science/article/abs/pii/S0008621517303555.

Review

9 M. Camero, M. Marinaro, A. Lovero, G. Elia, M. Losurdo, C. Buonavoglia and M. Tempesta, Polysaccharide extracts of the brown alga *Sargassum asperifolium* possess in vitro cancer chemopreventive properties, *Nat. Prod. Res.*, 2014, 28, 2304–2311, https://www.tandfonline.com/doi/abs/ 10.1080/14786419.2014.918120.

- 10 A. A. Matloub and N. E. Awad, Phycochemistry of some *Sargassum* spp. and their cytotoxic and antimicrobial activities, *Egypt. Pharm. J.*, 2012, **11**, 99–108, http://epj.eg.net/article.asp?issn=1687-4315;year=2012;volume=11; issue=2;spage=99;epage=108;aulast=Matloub.
- 11 S. E. N. Ayyad, S. Z. Sowellim, M. S. El-Hosini and A. Abo-Atia, The structural determination of a new steroidal metabolite from the brown alga *Sargassum asperifolium*, *Z. Naturforsch.*, 2003, 58, 333–336, https://www.degruyter.com/view/journals/znc/58/5-6/article-p333.xml.
- 12 N. Tsuchiya, A. Sato, H. Haruyama, T. Watanabe and Y. Iijima, Nahocols and isonahocols, endothelin antagonists from the brown alga, *Sargassum autumnale*, *Phytochemistry*, 1998, 48, 1003–1011, https:// www.sciencedirect.com/science/article/abs/pii/ S0031942297004767.
- 13 D. Noviendri, I. Jaswir, H. M. Salleh, M. Taher, K. Miyashita and N. Ramli, Fucoxanthin extraction and fatty acid analysis of *Sargassum binderi* and *S. duplicatum*, *J. Med. Plants Res.*, 2011, 5, 2405–2412, https://www.researchgate.net/profile/Dedi_Noviendri/publication/287564183_Fucoxanthin_extraction_and_fatty_acid_a nalysis_of_Sargassum_binderi_and_S_duplicatum/links/587c379308ae4445c0643765/Fucoxanthin-extraction-and-fatty-acid-analysis-of-Sargassum-binderi-and-S-duplicatum.pdf.
- 14 V. Akbari, S. Zafari and A. Yegdaneh, Anti-tuberculosis and cytotoxic evaluation of the seaweed *Sargassum boveanum*, *Res. Pharm. Sci.*, 2018, **13**, 30–37, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5772079/.
- 15 H. F. Tang, Y. H. Yi, X. S. Yao, Q. Z. Xu, S. Y. Zhang and H. W. Lin, Bioactive steroids from the brown alga *Sargassum carpophyllum*, *J. Asian Nat. Prod. Res.*, 2002, 4, 95–101, https://www.tandfonline.com/doi/abs/10.1080/10286020290027362.
- 16 H. F. Tang, Y. H. Yi, X. S. Yao, Q. Xu, S. Zhang and H. Lin, A novel steroid for *Sargassum carpophyllum*, *Zhongguo Haiyang Yaowu*, 2003, **22**, 28–30, http://en.cnki.com.cn/Article_en/CJFDTotal-HYYW200302007.htm.
- 17 S. N. Somasundaram, S. Shanmugam, B. Subramanian and R. Jaganathan, Cytotoxic effect of fucoidan extracted from Sargassum cinereum on colon cancer cell line HCT-15, Int. J. Biol. Macromol., 2016, 91, 1215–1223, https:// www.sciencedirect.com/science/article/pii/ S0141813016306432.
- 18 S. S. Narayani, S. Saravanan, J. Ravindran, M. S. Ramasamy and J. Chitra, In vitro anticancer activity of fucoidan extracted from *Sargassum cinereum* against Caco-2 cells, *Int. J. Biol. Macromol.*, 2019, 138, 618–628, https://

- www.sciencedirect.com/science/article/pii/S0141813019321683.
- 19 F. N. Baleta, J. M. Bolaños, O. C. Ruma, A. N. Baleta and J. D. Cairel, Phytochemicals screening and antimicrobial properties of *Sargassum oligocystum* and *Sargassum crassifolium* extracts, *J. Med. Plant*, 2017, 5, 382–387, https://pdfs.semanticscholar.org/b4b6/ 8e0ba1c29a53b0a47fcc05210d12932feee1.pdf.
- 20 S. Sugiono, M. Masruri, T. Estiasih and S. B. Widjarnako, Structural and Rheological Characteristics of Alginate from *Sargassum cristaefolium* Extracted by Twin Screw Extruder, *J. Aquat. Food Prod. Technol.*, 2019, 28, 944–959, https://www.tandfonline.com/doi/abs/10.1080/ 10498850.2019.1665603.
- 21 A. M. Gamal-Eldeen, M. A. Abo-Zeid and E. F. Ahmed, Antigenotoxic effect of the *Sargassum dentifolium* extracts: prevention of chromosomal aberrations, micronuclei, and DNA fragmentation, *Exp. Toxicol. Pathol.*, 2013, 65, 27–34, https://www.sciencedirect.com/science/article/abs/pii/S0940299311000807.
- 22 F. F. Madkour, W. F. Khalil and A. A. Dessouki, Protective effect of ethanol extract of *Sargassum dentifolium* (Phaeophyceae) in carbon tetrachloride-induced hepatitis in rats, *Int. J. Pharm. Pharm. Sci.*, 2012, 4, 637–641, https://www.researchgate.net/profile/Waleed_Khalil/publication/2 57138802_Protective_effect_of_ethanol_extract_of_Sargas sum_Dentifolium_Phaeophyceae_in_carbon_tetrachloride-induced_hepatitis_in_rats/links/ 0deec5247322b65f0a000000/Protective-effect-of-ethanol-extract-of-Sargassum-Dentifolium-Phaeophyceae-in-carbon-tetrachloride-induced-hepatitis-in-rats.pdf.
- 23 P. Reddy and S. Urban, Meroditerpenoids from the southern Australian marine brown alga *Sargassum fallax*, *Phytochem*, 2009, **70**, 250–255, https://www.sciencedirect.com/science/article/pii/S0031942208005979.
- 24 L. Silva Costa, C. B. Silva Telles, R. Medeiros Oliveira, L. T. Duarte Barreto Nobre, N. Dantas-Santos, R. Barros Gomes Camara, M. Santana Santos Pereira Costa, J. Almeida-Lima, R. F. Melo-Silveira, I. R. Lopes Albuquerque and E. L. Leite, Heterofucan from Sargassum filipendula Induces Apoptosis in HeLa Cells, Mar. Drugs, 2011, 9, 603–614, https://www.mdpi.com/1660-3397/9/4/ 603.
- 25 J. Chale-Dzul, R. P. C. de Vaca, C. Quintal-Novelo, L. Olivera-Castillo and R. Moo-Puc, Hepatoprotective effect of a fucoidan extract from *Sargassum fluitans* Borgesen against CCl₄-induced toxicity in rats, *Int. J. Biol. Macromol.*, 2020, 145, 500–509, https://www.sciencedirect.com/science/article/pii/S0141813019338309.
- 26 B. K. Kang, M. J. Kim and D. H. Ahn, In vivo and in vitro inhibitory activity of an ethanolic extract of *Sargassum* fulvellum and its component grasshopper ketone on atopic dermatitis, *Int. Immunopharmacol.*, 2016, 40, 176– 183, https://www.sciencedirect.com/science/article/pii/ S1567576916302855.

RSC Advances Review

- 27 J. Y. Kang, M. N. A. Khan, N. H. Park, J. Y. Cho, M. C. Lee, H. Fujii and Y. K. Hong, Antipyretic, analgesic, and antiinflammatory activities of the seaweed Sargassum fulvellum and Sargassum thunbergii in mice, J. Ethnopharmacol., 2008, 116, 187-190, https:// www.sciencedirect.com/science/article/abs/pii/ S0378874107005612.
- 28 A. Ina, K. I. Hayashi, H. Nozaki and Y. Kamei, Pheophytin a, a low molecular weight compound found in the marine Sargassum fulvellum, promotes differentiation of PC12 cells, Int. J. Dev. Neurosci., 2007, 25, 63-68, https://www.sciencedirect.com/science/article/ abs/pii/S0736574806004151.
- 29 W. Wu, K. Hasumi, H. Peng, X. Hu, X. Wang and B. Bao, Fibrinolytic Compounds Isolated from a Brown Alga, Sargassum fulvellum, Mar. Drugs, 2009, 7, 85-94, https:// www.mdpi.com/1660-3397/7/2/85.
- 30 T. Kusumi, M. Ishitsuka, T. Iwashita, H. Naoki, T. Konno and H. Kakisawa, A novel type of glycerides bearing a methacrylic acid moiety from the brown alga, Sargassum fulvellum, Chem. Lett., 1981, 1393-1396, www.journal.csj.jp/doi/abs/10.1246/cl.1981.1393.
- 31 R. B. Jia, Z. R. Li, J. Wu, Z. R. Ou, Q. Zhu, B. Sun, L. Lin and M. Zhao, Physicochemical properties of polysaccharide from Sargassum fusiforme hypoglycemic and hypolipidemic activities in type 2 diabetic rats, Int. J. Biol. Macromol., 2019, DOI: 10.1016/ j.ijbiomac.2019.12.243, https://www.sciencedirect.com/ science/article/pii/S0141813019399982.
- 32 Y. Ye, D. Ji, L. You, L. Zhou, Z. Zhao and C. Brennan, Structural properties and protective effect of Sargassum fusiforme polysaccharides against ultraviolet B radiation in hairless Kun Ming mice, J. Funct. Foods, 2018, 43, 8-16, https://www.sciencedirect.com/science/article/abs/pii/ S175646461830032X
- 33 N. Payghami, S. Jamili, A. Rustaiyan, S. Saeidnia, M. Nikan and A. R. Gohari, Alpha-amylase inhibitory activity and sterol composition of the marine algae, Sargassum glaucescens, Pharmacogn. Res., 2014, 7, 314-321, https:// www.ncbi.nlm.nih.gov/pmc/articles/PMC4660509/.
- 34 C. Y. Huang, S. J. Wu, W. N. Yang, A. W. Kuan and C. Y. Chen, Antioxidant activities of crude extracts of fucoidan extracted from Sargassum glaucescens by a compressional-puffing-hydrothermal extraction process, Food Chem., 2016, 197, 1121-1129, https:// www.sciencedirect.com/science/article/pii/ S0308814615302454.
- 35 P. A. Hwang, C. H. Wu, S. Y. Gau, S. Y. Chien and D. F. Hwang, Antioxidant and immune-stimulating activities of hot-water extract from seaweed Sargassum hemiphyllum, J. Mar. Sci. Technol., 2010, 18, 41-46, https:// jmst.ntou.edu.tw/marine/18-1/41-46.pdf.
- 36 P. A. Hwang, Y. L. Hung, Y. K. Tsai, S. Y. Chien and Z. L. Kong, The brown seaweed Sargassum hemiphyllum exhibits α-amylase and α-glucosidase inhibitory activity and enhances insulin release in vitro, Cytotechnology,

- 653-660, https://link.springer.com/article/ 2015. 67, 10.1007/s10616-014-9745-9.
- 37 P. A. Hwang, S. Y. Chien, Y. L. Chan, M. K. Lu, C. H. Wu, Z. L. Kong and C. J. Wu, Inhibition of lipopolysaccharide (LPS)-induced inflammatory responses by Sargassum hemiphyllum sulfated polysaccharide extract in RAW 264.7 macrophage cells, J. Agric. Food Chem., 2011, 59, 2062-2068, https://pubs.acs.org/doi/abs/10.1021/jf1043647.
- 38 M. Wei, S. Li, J. Chen and Y. Lin, Chemical constituents of the brown alga Sargassum henslowianum collected from the South China Sea, Chem. Nat. Compd., 2012, 48, 677-678, https://link.springer.com/article/10.1007/s10600-012-0346-
- 39 H. D. Cuong, T. T. T. Thuy, T. T. Huong, B. M. Ly and T. T. Van, Structure and hypolipidaemic activity of fucoidan extracted from brown seaweed Sargassum henslowianum, Nat. Prod. Res., 2015, 29, 411-415, https:// www.tandfonline.com/doi/abs/10.1080/ 14786419.2014.948436.
- 40 M. T. Ale, H. Maruyama, H. Tamauchi, J. D. Mikkelsen and A. S. Meyer, Fucose-Containing Sulfated Polysaccharides from Brown Seaweeds Inhibit Proliferation of Melanoma Cells and Induce Apoptosis by Activation of Caspase-3 in Mar. *Drugs*, 2011, 9, 2605–2621, https:// www.mdpi.com/1660-3397/9/12/2605/htm.
- 41 M. E. Kim, Y. C. Jung, I. Jung, H. W. Lee, H. Y. Youn and J. S. Lee, Anti-Inflammatory Effects of Ethanolic Extract from Sargassum horneri (Turner) C. Agardh on Lipopolysaccharide-Stimulated Macrophage Activation via NF-κB Pathway Regulation, Immunol. Invest., 2015, 44, https://www.tandfonline.com/doi/abs/10.3109/ 137-146,08820139.2014.942459.
- 42 T. U. Jayawardena, H. S. Kim, K. A. Sanjeewa, S. Y. Kim, J. R. Rho, Y. Jee, G. Ahn and Y. J. Jeon, Sargassum horneri and 6-hydroxy-4,4,7a-trimethyl-5,6,7,7atetrahydrobenzofuran-2(4H)-one (HTT); LPS-induced inflammation attenuation via suppressing NF-κB, MAPK and oxidative stress through Nrf2/HO-1 pathways in RAW 264.7 macrophages, Algal Res., 2019, 40, 101513, https:// www.sciencedirect.com/science/article/pii/ S2211926419300177.
- 43 M. Nakayama, Y. Fukuoka, H. Nozaki, A. Matsuo and S. Hayashi, Structure of (+)-kjellmanianone, a highly oxygenated cyclopentenone from the marine alga, Sargassum kjellmanianum, Chem. Lett., 1980, 9, 1243-1246, https://www.journal.csj.jp/doi/abs/10.1246/cl.1980.1243.
- 44 H. Nozaki, Y. Fukuoka, A. Matsuo, O. Soga and M. Nakayama, Structure of sargassumlactam, a new β , γ unsaturated- γ -lactam, from the marine alga *Sargassum* kjellmanianum, Chem. Lett., 1980, 9, 1453-1454, https:// www.journal.csj.jp/doi/abs/10.1246/cl.1980.1453.
- 45 I. Yamamoto, M. Takahashi, T. Suzuki, H. Seino and H. Mori, Antitumor effect of seaweeds. IV. Enhancement of antitumor activity by sulfation of a crude fucoidan fraction from Sargassum kjellmanianum, Ipn. J. Exp. Med., 1984, 54, 143-151, https://europepmc.org/article/med/ 6513098.

Review

46 L. I. L. Ma Ww and G. Zhou, *In vitro* immunoregulatory and antitumor activity of sulfated polysaccharides from *Sargassum kjellmanianum*, *Food Sci.*, 2013, 7, 270–274, http://en.cnki.com.cn/Article_en/CJFDTotal-SPKX201307058.htm.

- 47 S. R. Yende, U. N. Harle, S. K. Arora and V. B. Pande, Phytochemical screening and anticonvulsant activity of *Sargassum ilicifolium* (brown algae) in mice, *J. Phytopharm.*, 2018, 7, 25–28, http://www.phytopharmajournal.com/Vol7 Issue1 06.pdf.
- 48 W. Jin, W. Zhang, J. Wang, J. Yao, E. Xie, D. Liu, D. Duan and Q. Zhang, A study of neuroprotective and antioxidant activities of heteropolysaccharides from six *Sargassum* species, *Int. J. Biol. Macromol.*, 2014, 67, 336–342, https://www.sciencedirect.com/science/article/pii/S0141813014001846.
- 49 K. A. Francesconi, J. S. Edmonds, R. V. Stick, B. W. Skelton and A. H. White, Arsenic-containing ribosides from the brown alga *Sargassum lacerifolium*: X-ray molecular structure of 2-amino-3-[5'-deoxy-5'-(dimethylarsinoyl) ribosyloxy]propane-1-sulphonic acid, *J. Chem. Soc., Perkin Trans.* 1, 1991, 2707–2716, https://pubs.rsc.org/en/content/articlehtml/1991/p1/p19910002707.
- 50 A. M. Gamal-Eldeen, E. F. Ahmed and M. A. Abo-Zeid, *In vitro* cancer chemopreventive properties of polysaccharide extract from the brown alga, *Sargassum latifolium*, *Food Chem. Toxicol.*, 2009, 47, 1378–1384, https://www.sciencedirect.com/science/article/pii/S0278691509001215.
- 51 A. F. Abdel-Fattah, M. M. D. Hussein and H. M. Salem, Some structural features of sargassan, a sulphated heteropolysaccharide from *Sargassum linifolium*, *Carbohydr. Res.*, 1974, 33, 19–24, https://www.sciencedirect.com/science/article/abs/pii/S0008621500829365.
- 52 Y. Kamei, M. Sueyoshi, K. I. Hayashi, R. Terada and H. Nozaki, The novel anti-propionibacterium acnes compound, sargafuran, found in the marine brown alga *Sargassum macrocarpum*, *J. Antibiot.*, 2009, **62**, 259–263, https://www.nature.com/articles/ja200925.
- 53 C. K. Tsang, A. Ina, T. Goto and Y. Kamei, Sargachromenol, a novel nerve growth factor-potentiating substance isolated from *Sargassum macrocarpum*, promotes neurite outgrowth and survival via distinct signaling pathways in PC12D cells, *Neuroscience*, 2005, 132, 633–643, https://www.sciencedirect.com/science/article/abs/pii/S0306452205001284.
- 54 Y. K. Choi, J. Kim, K. M. Lee, Y. J. Choi, B. R. Ye, M. S. Kim, S. G. Ko, S. H. Lee, D. H. Kang and S. J. Heo, Tuberatolide B suppresses cancer progression by promoting ROS-mediated inhibition of STAT3 signaling, *Mar. Drugs*, 2017, 15, 55, https://www.mdpi.com/1660-3397/15/3/55.
- 55 Z. Manzoor, V. B. Mathema, D. Chae, E. S. Yoo, H. K. Kang, J. W. Hyun, N. H. Lee, M. H. Ko and Y. S. Koh, Extracts of the seaweed *Sargassum macrocarpum* inhibit the CpG-induced inflammatory response by attenuating the NF-κB pathway, *Food Sci. Biotechnol.*, 2014, 23, 293–297, https://

- idp.springer.com/authorize/casa?redirect_uri=https://link.springer.com/article/10.1007/s10068-014-0041-4and casa_token=KPIG5pTg4T4AAAAA:lY2pSCZFNPucfDFsjzvEjJi2k_WfOAxWRBFIeFLpGK7mXL2BGkoRNMXtAw6ps-HctBwg0M79c-mlo1cw.
- 56 M. Zubia, C. Payri and E. Deslandes, Alginate, mannitol, phenolic compounds and biological activities of two range-extending brown algae, *Sargassum mangarevense* and *Turbinaria ornata* (Phaeophyta: Fucales), from Tahiti (French Polynesia), *J. Appl. Phycol.*, 2008, 20, 1033–1043, https://link.springer.com/article/10.1007/s10811-007-9303-3.
- 57 N. Bhaskar, Growth inhibition of human pro-myelocytic leukemia (HL-60) cells by lipid extracts of marine alga *Sargassum marginatum* (Fucales, Phaeophyta) harvested off Goa (west coast of India) with special reference to fatty acid composition, *Indian J. Mar. Sci.*, 2004, 33, 355–360, http://nopr.niscair.res.in/handle/123456789/1690.
- 58 Y. M. Ham, K. N. Kim, W. J. Lee, N. H. Lee and C. G. Hyun, Chemical constituents from *Sargassum micracanthum* and antioxidant activity, *Int. J. Pharmacol.*, 2010, **6**, 147–151, https://scialert.net/fulltextmobile/?doi=ijp.2010.147.151.
- 59 J. Mori, M. Iwashima, H. Wakasugi, H. Saito, T. Matsunaga, M. Ogasawara, S. Takahashi, H. Suzuki and T. Hayashi, New plastoquinones isolated from the brown alga, *Sargassum micracanthum*, *Chem. Pharm. Bull.*, 2005, 53, 1159–1163, https://www.jstage.jst.go.jp/article/cpb/53/9/53_9_1159/ _article/-char/ja/.
- 60 M. Iwashima, J. Mori, X. Ting, T. Matsunaga, K. Hayashi, D. Shinoda, H. Saito, U. Sankawa and T. Hayashi, Antioxidant and Antiviral Activities of Plastoquinones from the Brown Alga Sargassum micracanthum, and a New Chromene Derivative Converted from the Plastoquinones, Biol. Pharm. Bull., 2005, 28, 374–377, https://www.jstage.jst.go.jp/article/bpb/28/2/28_2_374/_article/char/ja/.
- 61 C. Kim, I. K. Lee, G. Y. Cho, K. H. Oh, Y. W. Lim and B. S. Yun, Sargassumol, a novel antioxidant from the brown alga *Sargassum micracanthum*, *J. Antibiot.*, 2012, 65, 87–89, https://www.nature.com/articles/ja2011107/.
- 62 Y. Shizuri, S. Matsukawa, M. Ojika and K. Yamada, Two new farnesylacetone derivatives from the brown alga *Sargassum micracanthum*, *Phytochem*, 1982, 21, 1808–1809, https://www.sciencedirect.com/science/article/abs/pii/S0031942282850747.
- 63 T. Kusumi, M. Ishitsuka, Y. Nomura, T. Konno and H. Kakisawa, New farnesylacetone derivatives from *Sargassum micracanthum*, *Chem. Lett.*, 1979, **8**, 1181–1184, https://www.journal.csj.jp/doi/pdf/10.1246/cl.1979.1181.
- 64 M. M. Iwashima, N. Tako, T. Hayakawa, T. Matsunaga, J. Mori and H. Saito, New chromane derivatives isolated from the brown alga, *Sargassum micracanthum*, *Chem. Pharm. Bull.*, 2008, **56**, 124–128, https://www.jstage.jst.go.jp/article/cpb/56/1/56_1_124/_article/char/ja/.
- 65 E. Plouguerné, E. Ioannou, P. Georgantea, C. Vagias, V. Roussis, C. Hellio, E. Kraffe and V. Stiger-Pouvreau,

Anti-microfouling activity of lipidic metabolites from the invasive brown alga *Sargassum muticum* (Yendo) Fensholt, *Mar. Biotechnol.*, 2010, 12, 52–61, https://link.springer.com/article/10.1007/s10126-009-9199-9.

RSC Advances

- 66 J. A. Lee, Y. R. Cho, S. S. Hong and E. K. Ahn, Anti-obesity activity of saringosterol isolated from *Sargassum muticum* (Yendo) Fensholt extract in 3T3-L1 cells, *Phytother. Res.*, 2017, 31, 1694–1701, https://onlinelibrary.wiley.com/doi/abs/10.1002/ptr.5892.
- 67 D. V. Devi and P. Viswanathan, Sulphated polysaccharide from *Sargassum myriocystum* confers protection against gentamicin-induced nephrotoxicity in adult zebrafish, *Environ. Toxicol. Pharmacol.*, 2019, 72, 103269, https://www.sciencedirect.com/science/article/pii/S1382668919301437.
- 68 Y. Peng, E. Xie, K. Zheng, M. Fredimoses, X. Yang, X. Zhou, Y. Wang, B. Yang, X. Lin, J. Liu and Y. Liu, Nutritional and Chemical Composition and Antiviral Activity of Cultivated Seaweed *Sargassum naozhouense* Tseng et Lu, *Mar. Drugs*, 2013, 11, 20–32, https://www.mdpi.com/1660-3397/11/1/20.
- 69 Y. Peng, R. M. Huang, X. P. Lin and Y. H. Liu, Norisoprenoids from the brown alga *Sargassum naozhouense* Tseng et Lu, *Molecules*, 2018, 23, 348, https://www.mdpi.com/1420-3049/23/2/348.
- 70 A. Mohammed, R. Bissoon, E. Bajnath, K. Mohammed, T. Lee, M. Bissram, N. John, N. K. Jalsa, K. Y. Lee and K. Ward, Multistage extraction and purification of waste *Sargassum natans* to produce sodium alginate: an optimization approach, *Carbohydr. Polym.*, 2018, 198, 109–118, https://www.sciencedirect.com/science/article/pii/S0144861718307215.
- 71 P. Permeh, S. Saeidnia, A. Mashinchian-Moradi and A. R. Gohari, Sterols from *Sargassum oligocystum*, a brown algae from the Persian Gulf, and their bioactivity, *Nat. Prod. Res.*, 2012, **26**, 774–777, https://www.tandfonline.com/doi/abs/10.1080/14786419.2010.548812.
- 72 K. Zandi, S. Ahmadzadeh, S. Tajbakhsh, Z. Rastian, F. Yousefi, F. Farshadpour and K. Sartavi, Anticancer activity of *Sargassum oligocystum* water extract against human cancer cell lines, *Eur. Rev. Med. Pharmacol. Sci.*, 2010, 14, 669–673, http://eprints.bpums.ac.ir/6196/.
- 73 H. Ye, C. Zhou, Y. Sun, X. Zhang, J. Liu, Q. Hu and X. Zeng, Antioxidant activities in vitro of ethanol extract from brown seaweed Sargassum pallidum, Eur. Food Res. Technol., 2009, 230, 101, https://link.springer.com/article/10.1007/s00217-009-1147-4.
- 74 M. J. Kim, M. J. Kim, K. B. W. R. Kim, S. H. Park, H. D. Choi, S. Y. Park, J. H. Kim, M. R. Jang, M. H. Im and D. H. Ahn, Anti-Inflammatory effect of *Sargassum patens* C. Agardh ethanol extract in LPS-induced RAW264. 7 cells and mouse ear edema, *Microbiol. Biotechnol. Lett.*, 2017, 45, 110–117, http://www.koreascience.or.kr/article/JAKO201720136710783.page.
- 75 W. Zhu, L. C. M. Chiu, V. E. C. Ooi, P. K. S. Chan and P. O. Ang Jr, Antiviral property and mode of action of a sulphated polysaccharide from *Sargassum patens* against

- herpes simplex virus type 2, *Int. J. Antimicrob. Agents*, 2004, 24, 279–283, https://www.sciencedirect.com/science/article/abs/pii/S0924857904001657.
- 76 R. Brkljača and S. Urban, Chemical Profiling (HPLC-NMR andamp; HPLC-MS), Isolation, and Identification of Bioactive Meroditerpenoids from the Southern Australian Marine Brown Alga Sargassum paradoxum, Mar. Drugs, 2015, 13, 102–127, https://www.mdpi.com/1660-3397/13/1/102/htm.
- 77 S. H. Qi, S. Zhang, J. S. Huang, Z. H. Xiao, J. Wu and L. J. Long, Glycerol derivatives and sterols from *Sargassum parvivesiculosum*, *Chem. Pharm. Bull.*, 2004, **52**, 986–988, https://www.jstage.jst.go.jp/article/cpb/52/8/52_8_986/article/-char/ja/.
- 78 S. Palanisamy, M. Vinosha, T. Marudhupandi, P. Rajasekar and N. M. Prabhu, Isolation of fucoidan from *Sargassum polycystum* brown algae: structural characterization, *in vitro* antioxidant and anticancer activity, *Int. J. Biol. Macromol.*, 2017, 102, 405–412, https://www.sciencedirect.com/science/article/pii/S0141813017300958.
- 79 C. W. Lee and J. S. Han, Hypoglycemic effect of Sargassum ringgoldianum extract in STZ-induced diabetic mice, Prev. Nutr. Food Sci., 2012, 17, 8, https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3866760/.
- 80 M. J. Kim, D. H. Jeong and D. H. Ahn, Anti-inflammatory activity of ethanolic extract of *Sargassum sagamianum* in RAW 264.7 cells, *Food Sci. Biotechnol.*, 2013, 22, 1113–1120, https://idp.springer.com/authorize/casa? redirect_uri=https://link.springer.com/article/10.1007/s10 068-013-0191-9andcasa_token=JhHbNWjs8FAAAAA A:uNeDYQd0IB4YDgdR7PThZ_yhI0KWOaGb62w6SGWC-4K7STlbgY0DTzgK_n3JULFyDGKR8zIawcJXVZHEmA.
- 81 J. S. Lee and H. A. Lee, *Sargassum sagamianum* extract protects INS-1 pancreatic β cells against high glucose-induced apoptosis, *Cytotechnology*, 2019, 71, 389–399, https://link.springer.com/article/10.1007/s10616-019-00295-5.
- 82 H. W. Chang, K. H. Jang, D. Lee, H. R. Kang, T. Y. Kim, B. H. Lee, B. W. Choi, S. Kim and J. Shin, Monoglycerides from the brown alga *Sargassum sagamianum*: isolation, synthesis, and biological activity, *Bioorg. Med. Chem. Lett.*, 2008, 18, 3589–3592, https://www.sciencedirect.com/science/article/pii/S0960894X08004915.
- 83 B. W. Choi, G. Ryu, S. H. Park, E. S. Kim, J. Shin, S. S. Roh, H. C. Shin and B. H. Lee, Anticholinesterase activity of plastoquinones from *Sargassum sagamianum*: lead compounds for Alzheimer's disease therapy, *Phytother. Res.*, 2007, **21**, 423–426, https://onlinelibrary.wiley.com/doi/abs/10.1002/ptr.2090.
- 84 S. Hur, H. Lee, Y. Kim, B. H. Lee, J. Shin and T. Y. Kim, Sargaquinoic acid and sargachromenol, extracts of Sargassum sagamianum, induce apoptosis in HaCaT cells and mice skin: Its potentiation of UVB-induced apoptosis, Eur. J. Pharmacol., 2008, 582, 1–11, https:// www.sciencedirect.com/science/article/abs/pii/ S0014299907013283.

Review

85 S. Horie, S. Tsutsumi, Y. Takada and J. Kimura, Antibacterial Quinone Metabolites from the Brown Alga, *Sargassum sagamianum*, *Bull. Chem. Soc. Jpn.*, 2008, **81**, 1125–1130, https://www.journal.csj.jp/doi/abs/10.1246/bcsj.81.1125.

- 86 E. J. Joung, M. Kwon, W. G. Gwon, L. Cao, S. G. Lee, T. Utsuki, N. Wakamatsu, J. I. Kim and H. R. Kim, Meroterpenoid-rich fraction of the ethanol extract of Sargassum serratifolium suppresses collagen-induced rheumatoid arthritis in DBA/1J mice via inhibition of nuclear factor κB activation, Mol. Nutr. Food Res., 2020, 64, 1900373, https://onlinelibrary.wiley.com/doi/full/ 10.1002/mnfr.201900373.
- 87 Y. H. Kim, J. H. Kim, D. H. Kim, S. H. Kim, H. R. Kim and Y. M. Kim, Synergistic antimicrobial effect of *Sargassum serratifolium* (C. Agardh) C. Agardh extract against human skin pathogens, *Korean J. Food Sci. Technol.*, 2016, 48, 241–246, http://www.koreascience.or.kr/article/JAKO201620341074982.page.
- 88 M. Kwon, S. J. Lim, B. Lee, T. Shin and H. R. Kim, Ethanolic extract of *Sargassum serratifolium* inhibits adipogenesis in 3T3-L1 preadipocytes by cell cycle arrest, *J. Appl. Phycol.*, 2018, **30**, 559–568, https://link.springer.com/article/10.1007/s10811-017-1215-2.
- 89 K. H. Jang, B. H. Lee, B. W. Choi, H. S. Lee and J. Shin, Chromenes from the brown alga *Sargassum siliquastrum*, *J. Nat. Prod.*, 2005, **68**, 716–723, https://pubs.acs.org/doi/abs/10.1021/np058003i.
- 90 J. I. Lee, B. J. Park, H. Kim and Y. Seo, Isolation of Two New Meroterpenoids from *Sargassum siliquastrum*, *Bull. Korean Chem. Soc.*, 2014, 35, 2867–2869, https://pdfs.semanticscholar.org/b2d2/e68f33fe57d1e70beba69e2022661c4f54ca.pdf.
- 91 S. H. Cho, J. Y. Cho, S. E. Kang, Y. K. Hong and D. H. Ahn, Antioxidant activity of mojabanchromanol, a novel chromene, isolated from brown alga *Sargassum siliquastrum*, *J. Environ. Biol.*, 2008, **29**, 479–484, http://www.jeb.co.in/journal_issues/200807_jul08_spl/paper_12.pdf.
- 92 S. K. Sah, B. H. Kim, G. T. Park, S. Kim, K. H. Jang, J. E. Jeon, J. Shin and T. Y. Kim, Novel isonahocol E3 exhibits anti-inflammatory and anti-angiogenic effects in endothelin-1-stimulated human keratinocytes, *Eur. J. Pharmacol.*, 2013, 720, 205–211, https://www.sciencedirect.com/science/article/abs/pii/S0014299913007838.
- 93 J. I. Lee, M. K. Kwak, H. Y. Park and Y. Seo, Cytotoxicity of meroterpenoids from *Sargassum siliquastrum* against human cancer cells, *Nat. Prod. Commun.*, 2013, 8, 1934578X1300800403, https://journals.sagepub.com/doi/ abs/10.1177/1934578X1300800403.
- 94 M. Jung, K. H. Jang, B. Kim, B. H. Lee, B. W. Choi, K. B. Oh and J. Shin, Meroditerpenoids from the brown alga *Sargassum siliquastrum*, *J. Nat. Prod.*, 2008, **71**, 1714–1719, https://pubs.acs.org/doi/abs/10.1021/np800321y.
- 95 H. Nagappan, P. P. Pee, S. H. Y. Kee, J. T. Ow, S. W. Yan, L. Y. Chew and K. W. Kong, Malaysian brown seaweeds Sargassum siliquosum and Sargassum polycystum: Low

- density lipoprotein (LDL) oxidation, angiotensin converting enzyme (ACE), α-amylase, and α-glucosidase inhibition activities, *Food Res. Int.*, 2017, **99**, 950–958, https://www.sciencedirect.com/science/article/abs/pii/S0963996917300339.
- 96 K. W. Glombitza and M. Keusgen, Fuhalols and deshydroxyfuhalols from the brown alga *Sargassum spinuligerum*, *Phytochem*, 1995, **38**, 987–995, https://www.sciencedirect.com/science/article/abs/pii/003194229400735C.
- 97 K. W. Glombitza, M. Keusgen and S. Hauperich, Fucophlorethols from the brown algae *Sargassum spinuligerum* and *Cystophora torulosa*, *Phytochem*, 1997, **46**, 1417–1422, https://www.sciencedirect.com/science/article/abs/pii/S0031942297004998.
- 98 M. Keusgen and K. W. Glombitza, Pseudofuhalols from the brown alga *Sargassum spinuligerum*, *Phytochem*, 1997, **46**, 1403–1415, https://www.sciencedirect.com/science/article/abs/pii/S0031942297005104.
- 99 P. Vijayabaskar, N. Vaseela and G. Thirumaran, Potential antibacterial and antioxidant properties of a sulfated polysaccharide from the brown marine algae *Sargassum swartzii*, *Chin. J. Nat. Med.*, 2012, **10**, 421–428, https://www.sciencedirect.com/science/article/pii/S187553641260082X.
- 100 S. Dinesh, T. Menon, L. E. Hanna, V. Suresh, M. Sathuvan and M. Manikannan, In vitro anti-HIV-1 activity of fucoidan from *Sargassum swartzii*, *Int. J. Biol. Macromol.*, 2016, 82, 83–88, https://www.sciencedirect.com/science/ article/pii/S0141813015300039.
- 101 P. Vijayabaskar and N. Vaseela, In vitro antioxidant properties of sulfated polysaccharide from brown marine algae *Sargassum tenerrimum*, *Asian Pac. J. Trop. Dis.*, 2012,
 2, S890–S896, https://www.sciencedirect.com/science/article/pii/S2222180812602874.
- 102 Y. Shibata and M. Morita, A novel, trimethylated arsenosugar isolated from the brown alga *Sargassum thunbergii*, *Agric. Biol. Chem.*, 1988, 52, 1087–1089, https:// www.tandfonline.com/doi/abs/10.1080/ 00021369,1988,10868781.
- 103 Y. W. Seo, K. E. Park and T. J. Nam, Isolation of a new chromene from the brown alga *Sargassum thunbergii*, *Bull. Korean Chem. Soc.*, 2007, **28**, 1831–1833, http://www.koreascience.or.kr/article/ArticleFullRecord.jsp?cn=JCGMCS 2007 v28n10 1831.
- 104 Y. Seo, K. E. Park, Y. A. Kim, H. J. Lee, J. S. Yoo, J. W. Ahn and B. J. Lee, Isolation of tetraprenyltoluquinols from the brownalga *Sargassum thunbergii*, *Chem. Pharm. Bull.*, 2006, 54, 1730–1733, https://www.jstage.jst.go.jp/article/cpb/54/12/54_12_1730/_article/-char/ja/.
- 105 Y. H. Kim, E. H. Kim, C. Lee, M. H. Kim and J. R. Rho, Two new monogalactosyl diacylglycerols from brown alga *Sargassum thunbergii*, *Lipids*, 2007, 42, 395–399, https:// aocs.onlinelibrary.wiley.com/doi/abs/10.1007/s11745-007-3035-7.
- 106 W. F. He, L. G. Yao, H. L. Liu and Y. W. Guo, Thunberol, a new sterol from the Chinese brown alga *Sargassum*

RSC Advances Review

- thunbergii, J. Asian Nat. Prod. Res., 2014, 16, 685-689, https://www.tandfonline.com/doi/abs/10.1080/ 10286020.2014.924511.
- 107 M. C. Kang, Y. Ding, E. A. Kim, Y. K. Choi, T. De Araujo, S. J. Heo and S. H. Lee, Indole derivatives isolated from brown alga Sargassum thunbergii inhibit adipogenesis through AMPK activation in 3T3-L1 preadipocytes, Mar. Drugs, 2017, 15, 119, https://www.mdpi.com/1660-3397/15/ 4/119.
- 108 K. E. Park, Y. A. Kim, H. A. Jung, H. J. Lee, J. W. Ahn, B. J. Lee and Y. W. Seo, Three norisoprenoids from the brown alga Sargassum thunbergii, J. Korean Chem. Soc., 2004, 48, 394–398, http://www.koreascience.or.kr/article/ ArticleFullRecord.jsp?cn=JCGMDC_2004_v48n4_394.
- 109 Y. P. Cai, C. B. Xie, B. C. Wang, P. L. Li and B. F. Li, Two new resorcinols from Sargassum thunbergii, J. Asian Nat. Prod. Res., 2010, 12, 1001–1004, https://www.tandfonline.com/ doi/abs/10.1080/10286020.2010.522179.
- 110 K. B. W. R. Kim, M. J. Kim and D. H. Ahn, Lipase inhibitory activity of chlorophyll a, isofucosterol and saringosterol isolated from chloroform fraction of Sargassum thunbergii, Prod. Res., 2014, 28, 1310–1312, www.tandfonline.com/doi/abs/10.1080/ 14786419.2014.900769.
- 111 J. B. Lee, A. Takeshita, K. Hayashi and T. Hayashi, Structures and antiviral activities of polysaccharides from Sargassum trichophyllum, Carbohydr. Polym., 2011, 86, 995-999. https://www.sciencedirect.com/science/article/ abs/pii/S0144861711004395.
- 112 T. A. Fenoradosoa, G. Ali, C. Delattre, C. Laroche, E. Petit, A. Wadouachi and P. Michaud, Extraction and characterization of an alginate from the brown seaweed Sargassum turbinarioides Grunow, J. Appl. Phycol., 2010, 131–137, https://link.springer.com/article/10.1007/ s10811-009-9432-v.
- 113 V. Jesumani, H. Du, P. Pei, M. Aslam and N. Huang, Comparative study on skin protection activity of polyphenol-rich extract and polysaccharide-rich extract from Sargassum vachellianum, PLoS One, 2020, 15, 1-17,

- https://journals.plos.org/plosone/article?id=10.1371/ journal.pone.0227308.
- 114 S. H. Xu, Y. Z. Cen, Y. L. Li and S. Y. Xu, A novel elevenmembered heterocyclic compound from algae Sargassum vachellianum CA131:283698, Chin. Chem. Lett., 1999, 10, 401-402.
- 115 K. Arunkumar, N. Selvapalam and R. Rengasamy, The antibacterial compound sulphoglycerolipid 1-0 palmitoyl-3-0(6'-sulpho-alpha-quinovopyranosyl)-glycerol from Sargassum wightii Greville (Phaeophyceae), Bot. Mar., 2005, **48**, 441–445, https://www.degruyter.com/view/j/ bot.2005.48.issue-5/bot.2005.058/bot.2005.058.xml.
- Maneesh and K. Chakraborty, Unprecedented antioxidative and anti-inflammatory aryl polyketides from the brown seaweed Sargassum wightii, Food Res. Int., 2017, https://www.sciencedirect.com/science/ 100, 640-649, article/abs/pii/S096399691730323X.
- 117 A. Maneesh and K. Chakraborty, Previously undescribed frido oleanenes and oxygenated labdanes from the brown seaweed Sargassum wightii and their protein tyrosine phosphatase-1B inhibitory activity, Phytochem, 2017, 144, 19-32, https://www.sciencedirect.com/science/article/abs/ pii/S0031942217302868.
- 118 A. Maneesh and K. Chakraborty, Previously undescribed antioxidative O-heterocyclic angiotensin enzyme inhibitors from the intertidal seaweed Sargassum wightii as potential antihypertensives, Food Res. Int., 2018, 474-486, https://www.sciencedirect.com/science/ article/abs/pii/S0963996918305738.
- 119 M. C. Kim, H. C. Kwon, S. N. Kim, H. S. Kim and B. H. Um, Plastoquinones from Sargassum yezoense; Chemical Structures and Effects on the Activation of Peroxisome Proliferator-Activated Receptor Gamma, Chem. Pharm. Bull., 2011, 59, 834-838, https://www.jstage.jst.go.jp/ article/cpb/59/7/59 7 834/ article/-char/ja/.
- 120 https://patents.google.com/patent/KR20110006444A/en.
- 121 https://patents.google.com/patent/KR20130142424A/en.
- 122 https://patents.justia.com/patent/4769223#history.