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Development of TsDPEN based imine-containing ligands for the copper-catalysed asymmetric Kinugasa reaction†

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A novel class of chiral *N,N,N* imine-containing ligands derived from TsDPEN (*N*-(*p*-tosyl)-1,2-diphenylethylene-1,2-diamine) has been developed and applied to the copper-catalyzed asymmetric Kinugasa reaction. The copper(II) salt proved to be an efficient catalyst precursor, and it provides an efficient way to synthesize enantioenriched *cis*- β -lactam. The pathway is air-tolerant and easily manipulated, and the ligands are easy to synthesize. A working model is proposed in which the stereocontrolling step is the [2 + 2] cycloaddition between ketene and imine to explain the observed stereoselectivities.

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Introduction

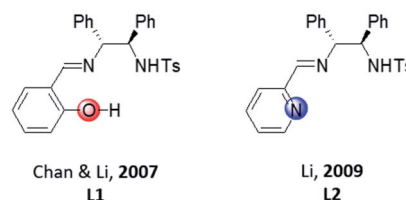
Since the discovery of penicillin in 1928, a number of β -lactam antibiotics have been developed.¹ In addition to antibacterial activity, β -lactam containing molecules also exhibit special biological activities,² such as Ezetimibe,^{2a} dual PPAR α/γ agonists,^{2b} a TRPV1 antagonist,^{2c} β -lactamic hFAAH inhibitors^{2d} and neuronal protective effect against brain ischemia.^{2e} At the same time, β -lactam compounds are also widely employed as useful intermediates in organic synthesis.³ Therefore, considerable efforts have been made in developing an efficient synthesis method for the preparation of chiral β -lactams.⁴

The Kinugasa reaction, developed in 1972,⁵ is a simple and direct reaction between alkynes and nitrones for the synthesis of β -lactams in the presence of copper and a base.⁶ The group of Miura reported the first catalytic asymmetric version by using CuI (10 mol%) and bis(oxazoline) (20 mol%) with 57% ee and 35% de in 1995.⁷ Fu *et al.* achieved the first diastereoselective and enantioselective catalytic Kinugasa reaction by employing 1–2.5 mol% CuCl and bis(azaferrocenes) (up to 93% ee and *cis/trans* up to 95/5) in 2002.⁸ Tang and co-workers demonstrated for the first time that a copper(II) catalyst could replace the previously performed air-sensitive copper(I) salts by using a C₃-symmetric tris(oxazoline) ligand in 2003.⁹ Since then, several ligands were developed for the catalytic asymmetric Kinugasa

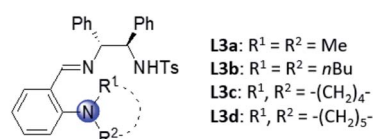
reaction, such as phosphaferrrocene-oxazoline,^{8b} HETPHOX,¹⁰ Inda-bis(oxazoline),¹¹ diamine,¹² *N*-PINAP,¹³ and Prolinol-Phosphine.¹⁴ Enders *et al.* developed a method to synthesis spirocyclic β -lactams through Kinugasa/Michael domino reactions by the utilizing of copper-BOX.¹⁵ In most cases the main products are *cis*- β -lactams, except for the use of Cu-diamine by Feng *et al.*¹² Although progress have been achieved, the scope of chiral ligands employed in the Kinugasa reaction is limited and the catalytic highly enantioselective Kinugasa reaction remains a challenge and less developed.

Chiral imino-containing ligands have emerged as a powerful tool in asymmetric catalysis.¹⁶ Despite the great success that has been achieved, there is still a need to develop more powerful novel chiral ligands with imino group for asymmetric catalysis.¹⁶ Chan & Li have designed two types of chiral tridentate *N*-

(a) Previously reported chiral imine-containing ligands derived from TsDPEN



(b) Novel *N,N,N* imine-containing ligands



Scheme 1 Chiral imine-containing ligands derived from TsDPEN.

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tosylated aminoimine ligands containing phenol and pyridine, respectively (Scheme 1a, **L1** and **L2**)¹⁷ for the addition of alkynes to imines. Recently, we have developed a new class of chiral polydentate *N,O* ligands derived from *N*-(*p*-tosyl)-1,2-diphenylethylene-1,2-diamine (TsDPEN).^{18,19} Herein, we successfully developed a novel class of chiral *N,N,N* imine-containing ligands derived from TsDPEN (Scheme 1b, **L3**) and applied them in Cu(II)-catalysed asymmetric Kinugasa reaction.

Results and discussion

At the outset, we investigated the Kinugasa reaction of alkyne **1a** and nitron **2a** in the presence of dicyclohexylamine as the base in CH₃CN at 0 °C, evaluating various copper catalysts prepared

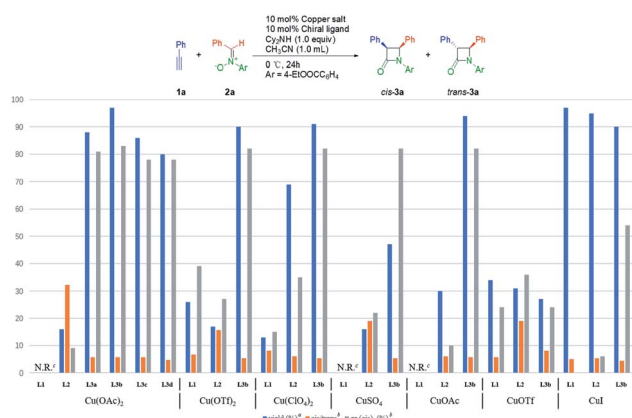


Fig. 1 Effect of copper salts and ligands on the asymmetric Kinugasa reaction. Reaction conditions: copper salt (10 mol%), ligand (10 mol%), **1a** (0.2 mmol), **2a** (0.22 mmol), dicyclohexylamine (1.0 equiv.), CH₃CN (1.0 mL), at 0 °C for 24 h. ^aTotal isolated yield of the *cis*- and *trans*-products. ^bThe ratios of *cis/trans* (x/1) and the ee of the *cis*-products were determined by HPLC on a chiral stationary phase. ^cN.R. = no reaction.

Table 1 Effect of the base on the asymmetric Kinugasa reaction^a

Entry	Base	Yield ^b (%)	<i>cis/trans</i> ^c	ee (<i>cis</i>) ^c (%)
1	—	Trace	—	—
2	Cy ₂ NH ^d	97	85 : 15	83
3	<i>i</i> Pr ₂ NEt	10	87 : 13	44
4	K ₂ CO ₃	9	61 : 39	71
5	<i>i</i> Pr ₂ NH	73	80 : 20	79
6	Cy ₂ NMe ^e	24	85 : 15	68
7	<i>n</i> Bu ₂ NH	90	59 : 41	80

^a Reaction conditions: Cu(OAc)₂ (10 mol%), **L3b** (10 mol%), **1a** (0.2 mmol), **2a** (0.22 mmol), base (1.0 equiv.), CH₃CN (1.0 mL), at 0 °C for 24 h. ^b Total isolated yield of the *cis*- and *trans*-products. ^c Determined by HPLC on a chiral stationary phase. ^d Dicyclohexylamine. ^e *N,N*-Dicyclohexylmethylethylamine.

Table 2 Effect of the solvent on the Kinugasa reaction^a

Entry	Solvent	Yield ^b (%)	<i>cis/trans</i> ^c	ee (<i>cis</i>) ^c (%)
1	CH ₃ CN	97	85 : 15	82
2	CH ₂ Cl ₂	69	84 : 16	81
3	THF	77	88 : 12	66
4	EtOAc	70	84 : 16	76
5	DMF	68	49 : 51	74
6	EtOH	60	87 : 13	74
7	Toluene	74	81 : 19	73

^a Reaction conditions: Cu(OAc)₂ (10 mol%), **L3b** (10 mol%), **1a** (0.2 mmol), **2a** (0.22 mmol), dicyclohexylamine (1.0 equiv.), solvent (1.0 mL), at 0 °C for 24 h. ^b Total isolated yield of the *cis*- and *trans*-products. ^c Determined by HPLC on a chiral stationary phase.

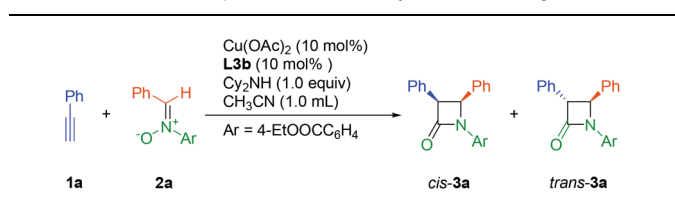
in situ from copper salts and chiral tridentate ligands **L1**–**L3** (Fig. 1).²⁰ Among them, Cu(OTf)₂/**L1** (26% yield, 87/13 *cis/trans*, 39% *cis* ee),²¹ CuOTf/**L2** (31% yield, 95/5 *cis/trans*, 36% *cis* ee),²² Cu(OAc)₂/**L3b** (97% yield, 85/15 *cis/trans*, 83% *cis* ee) turned out to be the efficient combination in terms of yield, dr and enantioselectivity. These results indicated that the dialkylamine substitution in aniline obviously affected the reactivity and the enantioselectivity. Then the substituent effect on the amino group of **L3** was examined and similar results were obtained (72–97% yield, ~85/15 *cis/trans*, 70–83% *cis* ee). Considering the reactivity and enantioselectivity, we selected **L3b** for further optimization.

Next, the influences of different bases were studied with the results shown in Table 1. Although the ligand contained a tertiary amine functional group, this transformation hardly proceeded without basic additives (Table 1, entry 1). Tertiary amines and inorganic bases gave poor yield (Table 1, entries 3, 4 and 6). Secondary amines afforded the desired products with better yield and enantioselectivity, and higher sterically hindered secondary amines achieved higher diastereoselectivity (Table 1, entries 2, 5 and 7). Dicyclohexylamine proved to be the best base.

Solvent effects were studied next, with the results summarized in Table 2. Only moderate enantioselectivity could be obtained in examined solvents other than CH₂Cl₂ and CH₃CN (Table 2, entries 1–7). No diastereoselectivity was observed in DMF (Table 2, entry 5). The best yield and enantioselectivity were obtained by CH₃CN (Table 2, entry 1).

Subsequently, the reaction temperature was examined. As shown in Table 3, both the diastereoselectivity and enantioselectivity were dependent on the reaction temperature. When the temperature was decreased from 0 to –30 °C, the diastereoselectivity and enantioselectivity increased (Table 3, entries 1–3). When the reaction temperature further decreased to –40 °C, the reaction almost stopped by about halfway through (Table 3, entry 4).

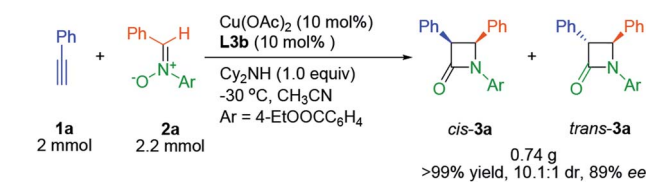


Table 3 Effect of temperature on the asymmetric Kinugasa reaction^a

Entry	T (°C)	Time (h)	Yield ^b (%)	cis/trans	ee (cis) ^c (%)
1	0	24	97	85 : 15 ^c	82
2	-15	48	>99	90 : 10 ^c	87
3	-30	48	>99	91 : 9 ^c (10.5 : 1) ^d	90
4	-40	48 or 72	46	92 : 8 ^c	90

^a Reaction conditions: Cu(OAc)₂ (10 mol%), **L3b** (10 mol%), **1a** (0.2 mmol), **2a** (0.22 mmol), dicyclohexylamine (1.0 equiv.), CH₃CN (1.0 mL). ^b Total isolated yield of the *cis*- and *trans*-products. ^c Determined by HPLC on a chiral stationary phase. ^d Determined by ¹H NMR.

Under the established optimal reaction conditions, the substrate scope of the cycloaddition reaction between various alkynes and nitrones was examined next (Table 4). First, different alkynes participated in the reaction (Table 4, entries 1–6, 12–15, and 21). Weakly electron donating methyl group did not pose a problem for the reaction, and the corresponding *cis*-β-lactams were isolated in good yields, diastereo- and



Scheme 2 mmol-scale reaction.

enantioselectivities (Table 4, entries 2, 3 and 12–15). The reaction conditions were also compatible with electron withdrawing group (Table 4, entries 4 and 21). Phenylacetylene with a strongly electron donating methoxy group had very low reactivity and only trace amount of corresponding product could be observed (Table 4, entry 5). The application of alkyl alkynes in the system is feasible, as exemplified by the formation of product **3e** with high diastereoselectivity (99 : 1), albeit with low enantioselectivity (12% ee). Next, effects of C-substituents of the nitron were investigated (Table 4, entries 7–15). Methyl group and fused aromatic substituents had no adverse effect on the reaction. While strong decrease in reactivity for electron withdrawing group and heterocyclic substituents was observed (Table 4, entries 10 and 11). The electronic character of the *N*-bound aromatic groups of nitrones was also investigated (Table 4, entries 16–20). Whether electron-deficient or electron-rich *N*-

Table 4 Substrate scope of the asymmetric Kinugasa reaction^a

Entry	R ¹ , R ² , R ³	T (°C)	Product	cis/trans ^b	Yield (cis) ^c (%)	ee (cis) ^d (%)
1	C ₆ H ₅ , C ₆ H ₅ , Ar	-30	3a	10.5 : 1	92	90
2	4-Me-C ₆ H ₄ , C ₆ H ₅ , Ar	-30	3b	13.1 : 1	93	88
3	3-Me-C ₆ H ₄ , C ₆ H ₅ , Ar	-30	3c	9.6 : 1	91	91
4	4-F-C ₆ H ₄ , C ₆ H ₅ , Ar	0	3d	6.2 : 1	74	88
5	4-MeOC ₆ H ₄ , C ₆ H ₅ , Ar	0	Trace			
6	Cyclohexyl, C ₆ H ₅ , Ar	0	3e	>99 : 1	68	12
7	C ₆ H ₅ , 3-Me-C ₆ H ₄ , Ar	-30	3f	14 : 1	80	90
8	C ₆ H ₅ , 4-Me-C ₆ H ₄ , Ar	0	3g	6.7 : 1	81	83
9	C ₆ H ₅ , 2-naphthyl, Ar	0	3h	5.3 : 1	70	77
10	C ₆ H ₅ , 4-Cl-C ₆ H ₄ , Ar	0	Trace			
11	C ₆ H ₅ , 2-furyl, Ar	0	Trace			
12	4-Me-C ₆ H ₄ , 3-Me-C ₆ H ₄ , Ar	-30	3i	17.5 : 1	91	89
13	3-Me-C ₆ H ₄ , 3-Me-C ₆ H ₄ , Ar	-30	3j	12.5 : 1	90	89
14	4-Me-C ₆ H ₄ , 4-Me-C ₆ H ₄ , Ar	0	3k	11 : 1	80	79
15	3-Me-C ₆ H ₄ , 4-Me-C ₆ H ₄ , Ar	0	3l	6.3 : 1	82	79
16	C ₆ H ₅ , C ₆ H ₅ , C ₆ H ₅	-30	3m	12.9 : 1	93	91
17	C ₆ H ₅ , C ₆ H ₅ , 4-Me-C ₆ H ₄	0	3n	14 : 1	71	85
18	C ₆ H ₅ , C ₆ H ₅ , 4-Cl-C ₆ H ₄	0	3o	12.5 : 1	80	87
19	C ₆ H ₅ , C ₆ H ₅ , 4-Br-C ₆ H ₄	0	3p	11.8 : 1	82	87
20	C ₆ H ₅ , C ₆ H ₅ , 4-F-C ₆ H ₄	0	3q	16.5 : 1	70	86
21	4-F-C ₆ H ₄ , C ₆ H ₅ , C ₆ H ₅	0	3r	16.3 : 1	75	84

^a Reaction conditions: Cu(OAc)₂ (10 mol%), **L3b** (10 mol%), **1** (0.2 mmol), **2** (0.22 mmol), dicyclohexylamine (1.0 equiv.), CH₃CN (1.0 mL), at -30 °C or 0 °C for 48 h. ^b Determined by ¹H NMR of the crude reaction mixture. ^c Isolated yield of *cis*-product by chromatography on silica gel.

^d Determined by HPLC on a chiral stationary phase.



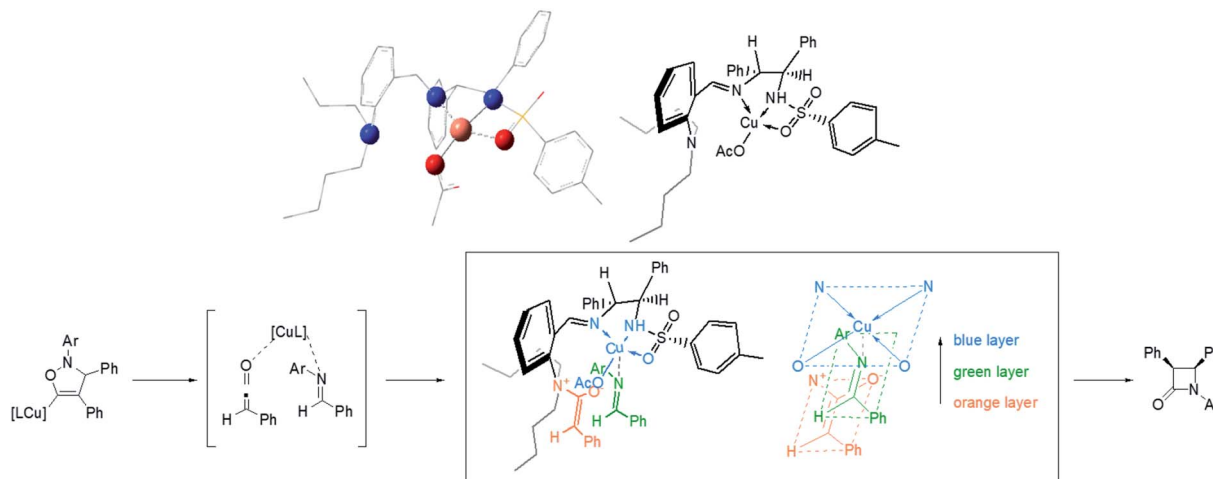


Fig. 2 Structure of the Cu complex optimized by DFT and proposed working model for stereoselectivity.

aryl nitrones were found to be highly suitable substrates for this asymmetric transformation.

To demonstrate the utility of this method, a mmol scale catalytic asymmetric Kinugasa reaction with 2.0 mmol **1a** and 2.2 mmol **2a** in the presence of 10 mol% chiral copper complex proceeded smoothly and **3a** was isolated with comparably high yield and stereoselectivity (Scheme 2).

The research of Kinugasa reaction mechanism has been an active research area.²³ Recently, Hein's group reported an improved mechanism based on the analysis of multiple byproducts and detailed kinetic analysis.^{23a} According to the modified mechanism, the stereocontrolling step is the [2 + 2] cycloaddition between ketene and imine, which are generated by reductive elimination and protonation from the initial [3 + 2] cycloaddition intermediate between nitron and σ -Cu acetylide. Based on this research and the geometry of Cu complex optimized by DFT,²⁴ a working model to explain the observed stereoselectivities is proposed in Fig. 2. The ketene combined with the *tert*-amine forming a zwitterionic enolate (phenyl away from N is favourable for the steric repulsion) and the imine coordinated to the Cu in a suitable position by avoiding the steric repulsion of *n*-butyl group on nitrogen.²⁵

Conclusions

In summary, we have successfully employed the novel chiral *N,N,N* imine-containing ligands derived from TsDPEN in the catalytic enantioselective Kinugasa reaction. It provides an efficient way to synthesize *cis*- β -lactam in good to excellent yields (up to 93%) and with good to excellent diastereo- and enantioselectivities (dr up to 17.5 : 1, ee up to 91%). This new protocol for the Kinugasa reaction is not effective for aliphatic alkynes and phenylacetylenes with a strongly electron donating group. A working model is proposed in which the stereocontrolling step is the [2 + 2] cycloaddition between ketene and imine to explain the observed stereoselectivities. The *N,N*-dialkylamine substitution is an unprecedented modification on the TsDPEN based imine containing ligand scaffold. The pathway is

air-tolerant and easily manipulated, and the ligands are easy to synthesize. Further investigations are underway in our laboratory including further geometry optimization of ligands, the detailed mechanism, and the extending of the substrate scope.

Experimental section

General

The solvents were dried by molecular sieve for more than 72 h before use. Other commercial reagents were used as purchased. NMR (Bruker 600 MHz) spectra were recorded in the deuterated solvents as stated, using residual non-deuterated solvent as internal standard. High resolution mass spectra were recorded with a Bruker Solaris XFT-ICR-MS system. The enantiomeric excess (ee) was determined by HPLC (Shimadzu LC-16) analysis using the corresponding commercial chiral column as stated in the experimental procedures at 23 °C with UV detector (254 nm). Optical rotations were measured on a commercial polarimeter (Rudolph Autopol I) and are reported as follows: $[\alpha]_D^{25}$ (c = g/100 mL, solvent). The diastereomeric ratios of the β -lactams were determined by integration of ¹H NMR spectra (the coupling constant for C3/C4 vicinal protons is 4.5–6 Hz for *cis* isomers and 2–2.5 Hz for *trans* isomers).^{8a} The absolute configuration of **3a**, **3e** and **3m** was determined by the comparison of the HPLC retention time and optical rotations with the literature data.^{8a,12} The absolute configuration of **3b–3d**, **3f–3l** and **3n–3r** was determined by analogy.

General preparation procedure of ligands

L1 and L2 are known compounds.¹⁷

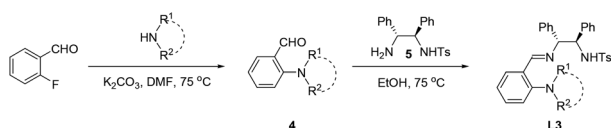
L1. Yellow solid, ¹H NMR (600 MHz, CDCl₃) δ 12.66 (s, 1H), 8.16 (s, 1H), 7.41 (d, J = 8.0 Hz, 2H), 7.34–7.31 (m, 1H), 7.19–7.07 (m, 9H), 7.00–6.95 (m, 3H), 6.90–6.83 (m, 3H), 5.32 (d, J = 7.4 Hz, 1H), 4.76 (t, J = 6.7 Hz, 1H), 4.56 (d, J = 5.9 Hz, 1H), 2.30 (s, 3H).

L2. Yellow solid, ¹H NMR (600 MHz, CDCl₃) δ 8.60 (d, J = 4.5 Hz, 1H), 8.00 (d, J = 7.7 Hz, 1H), 7.85–7.83 (m, 1H), 7.62 (d, J



= 8.2 Hz, 2H), 7.32–7.30 (m, 1H), 7.22–7.16 (m, 10H), 6.99 (d, J = 7.1 Hz, 1H), 5.94 (s, 1H), 4.64 (d, J = 6.6 Hz, 1H), 4.32 (d, J = 6.6 Hz, 1H), 2.43 (s, 3H).

General preparation procedure of L3. To a solution of *o*-fluorobenzaldehyde (10 mmol) and a secondary-amine (20 mmol) in 20 mL DMF was added K_2CO_3 (20 mmol), and the mixture was stirred under a N_2 atmosphere at 75 °C until the reaction proceed completely detected by TLC. The reaction mixture was filtered, the filtrate was diluted with CH_2Cl_2 (50 mL) and washed with saturated sodium bicarbonate solution (20 mL \times 3). The organic layers were dried (Na_2SO_4), and purified through column chromatography (petroleum ether/ether, 10 : 1) on silica gel to obtain a crude product. The crude product was dissolved in the mixture of petroleum ether/ether (v/v 10 : 1) and acidified to pH 2 with 1 M HCl. The resulting mixture was extracted twice with water, and the aqueous layer was basified to pH 9 with 1 M NaOH and extracted with CH_2Cl_2 . The dichloromethane layers were dried and the solvents were removed *in vacuo* to afford **4**. To a solution of **4** (1.2 mmol) in 30 mL ethanol was added **5** (1.2 mmol) and the mixture was stirred at 75 °C under a N_2 atmosphere until the reaction proceed completely detected by TLC. The EtOH was removed *in vacuo* to give **L3**.



L3a. White solid, (mixture of *E*- and *Z*-, E/Z = 5.6/1), 1H NMR (600 MHz, $CDCl_3$) δ 8.14 (*E*, s, 1H), 7.97 (*E*, d, J = 7.4 Hz, 1H), 7.91 (*Z*, d, J = 7.5 Hz, 0.18H), 7.56 (*Z*, d, J = 8.0 Hz, 0.37H), 7.51 (*Z*, d, J = 7.5 Hz, 0.38H), 7.32–7.29 (*E* + *Z*, m, 3.96H), 7.19–7.00 (*E* + *Z*, m, 11.37H), 6.91–6.89 (*E* + *Z*, m, 3.12H), 6.19 (*E*, d, J = 7.0 Hz, 1H) 4.84 (*E*, q, J = 3.3 Hz, 1H), 4.77 (*Z*, d, J = 5.2 Hz, 0.18H), 4.51 (*E*, d, J = 3.4 Hz, 1H), 4.26 (*Z*, d, J = 5.3 Hz, 0.18H), 2.35 (*E*, s, 6H), 2.32 (*Z*, s, 0.71H), 2.25 (*E*, s, 3H), 2.23 (*Z*, s, 0.20H). ^{13}C NMR (150 MHz, $CDCl_3$) δ *E*: 162.1, 154.1, 142.2, 140.9, 140.0, 137.2, 131.5, 129.0, 121.9, 117.8, 78.5, 63.6, 45.0, 21.3; *Z*: 162.1, 152.5, 143.4, 140.02, 139.4, 135.5, 133.4, 129.2, 124.0, 120.2, 75.2, 71.6, 68.5, 45.4, 21.4; *E* + *Z*: 128.27, 128.22, 127.98, 127.95, 127.84, 127.5, 127.4, 127.03, 127.02, 126.90, 126.98, 126.7, 126.5. ESI-HRMS calcd for $C_{30}H_{32}N_3O_2S^+$ [$M + H$] $^+$: 498.2210, found 498.2200.

L3b. White solid, (mixture of *E*- and *Z*-, E/Z = 9.1/1), 1H NMR (600 MHz, $CDCl_3$) δ 8.27 (*E*, s, 1H), 8.03 (*E*, d, J = 7.3 Hz, 1H), 7.65 (*Z*, d, J = 7.9 Hz, 0.23H), 7.42 (*Z*, d, J = 7.6 Hz, 0.25H), 7.34–7.33 (*E* + *Z*, m, 3H), 7.17–7.00 (*E* + *Z*, m, 12H), 6.93 (*E*, d, J = 7.9 Hz, 2H), 6.28, (*Z*, s, 0.11H), 6.20 (*E*, d, J = 6.5 Hz, 1H), 4.82 (*E*, q, J = 3.5 Hz, 1H), 4.64 (*Z*, d, J = 6.4 Hz, 0.11H), 4.51 (*E*, d, J = 3.7 Hz, 1H), 4.28 (*Z*, d, J = 6.4 Hz, 0.11H), 3.00–2.95 (*Z*, m, 0.49H), 2.73–2.64 (*E*, m, 4H), 2.28 (*E*, s, 3H), 2.27 (*Z*, s, 0.16H), 1.20–1.15 (*E*, m, 4H), 1.03–0.96 (*E*, m, 4H), 0.71 (*E*, t, J = 7.4 Hz, 6H). ^{13}C NMR (150 MHz, $CDCl_3$) δ *E*: 162.5, 152.5, 142.3, 141.1, 140.0, 137.4, 131.2, 78.0, 63.7, 54.2, 29.0, 21.4, 20.3, 13.9; *Z*: 162.5, 150.0, 143.5, 139.6, 139.2, 137.3, 130.7, 75.8, 71.9, 68.5, 54.6, 31.6, 29.4, 22.6, 21.5, 20.6, 20.3, 14.1, 14.0; *E* + *Z*: 129.3,

129.1, 128.3, 128.25, 128.20, 128.1, 128.0, 127.9, 127.7, 127.6, 127.3, 127.2, 127.08, 127.06, 126.9, 126.8, 124.2, 122.9, 122.8, 121.7. ESI-HRMS calcd for $C_{36}H_{44}N_3O_2S^+$ [$M + H$] $^+$: 582.3149, found 582.3140.

L3c. White solid, (mixture of *E*- and *Z*-, E/Z = 14.3/1), 1H NMR (600 MHz, $CDCl_3$) δ 8.10 (*E*, s, 1H), 7.88 (*E*, d, J = 7.7 Hz, 1H), 7.60 (*Z*, d, J = 7.8 Hz, 0.15H), 7.46 (*Z*, d, J = 7.5 Hz, 0.17H), 7.31–7.25 (*E* + *Z*, m, 3H), 7.23–7.09 (*E* + *Z*, m, 10H), 6.9–6.8 (*E* + *Z*, m, 3.1H), 6.73 (*E* + *Z*, d, J = 8.3 Hz, 1H), 6.19 (*E* + *Z*, s, 1.06H), 4.83 (*E*, q, J = 3.2 Hz, 1H), 4.69 (*Z*, d, J = 5.8 Hz, 0.07H), 4.47 (*E*, d, J = 3.2 Hz, 1H), 4.32 (*Z*, d, J = 5.8 Hz, 0.07H), 2.80–2.76 (*E* + *Z*, m, 4H), 2.37 (*Z*, s, 0.24H), 2.29 (*E*, s, 3H), 1.69–1.64 (*E* + *Z*, m, 4.19H). ^{13}C NMR (150 MHz, $CDCl_3$) δ *E*: 163.2, 150.4, 142.3, 141.0, 140.2, 137.3, 131.3, 129.2, 129.1, 128.3, 128.0, 127.06, 127.04, 126.98, 126.90, 126.7, 124.1, 118.9, 114.9, 78.7, 63.6, 52.6, 25.3, 21.4. ESI-HRMS calcd for $C_{32}H_{34}N_3O_2S^+$ [$M + H$] $^+$: 524.2366, found 524.2358.

L3d. White solid, (mixture of *E*- and *Z*-, E/Z = 3.7/1), 1H NMR (600 MHz, $CDCl_3$) δ 8.13 (*E*, s, 1H), 8.02 (*E*, d, J = 7.7 Hz, 1H), 7.66 (*Z*, d, J = 7.6 Hz, 0.57H), 7.58 (*Z*, d, J = 7.8 Hz, 0.58H), 7.42 (*E* + *Z*, t, J = 7.6 Hz, 0.61H), 7.32–7.31 (*E* + *Z*, m, 3.49H), 7.25–7.23 (*E* + *Z*, m, 1.87H), 7.18–7.05 (*E* + *Z*, m, 12H), 6.92–6.90 (*E* + *Z*, m, 3.14H), 6.20–6.19 (*E* + *Z*, m, 1.24H), 4.91 (*Z*, d, J = 3.4, 0.27H), 4.84–4.83 (*E*, m, 1H), 4.53 (*Z*, s, 0.27H), 4.48 (*E*, d, J = 3.5 Hz, 1H), 2.59–2.57 (*E* + *Z*, m, 2H), 2.49–2.46 (*E* + *Z*, m, 2H), 2.29 (*Z*, s, 0.9H), 2.26 (*E*, s, 3H), 1.37–1.30 (*E* + *Z*, m, 6H). ^{13}C NMR (150 MHz, $CDCl_3$) δ *E*: 162.0, 154.5, 142.2, 140.9, 140.0, 131.6, 78.5, 63.6, 54.5, 26.0, 23.9, 21.3; *Z*: 162.0, 152.9, 143.3, 141.1, 139.8, 137.7, 135.6, 134.8, 133.2, 75.0, 70.7, 68.4, 26.7, 26.1, 24.3, 21.4; *E* + *Z*: 129.1, 129.02, 128.98, 128.95, 128.5, 128.45, 128.43, 128.29, 128.26, 128.15, 128.07, 128.04, 127.95, 127.7, 127.5, 127.2, 127.1, 127.0, 127.95, 126.7, 126.6, 126.2, 125.2, 124.1, 122.3, 120.3, 118.6. ESI-HRMS calcd for $C_{33}H_{36}N_3O_2S^+$ [$M + H$] $^+$: 538.2523, found 538.2518.

General procedure for the asymmetric Kinugasa reaction

The mixture of $Cu(OAc)_2$ (0.02 mmol, 10 mol%) and **L3b** (0.02 mmol, 10 mol%) was stirring in CH_3CN (0.5 mL) under air atmosphere at 35 °C for 1 h. Then, dicyclohexylamine (0.2 mmol) and alkyne **1** (0.2 mmol) were sequentially added, and the test tube was rinsed with CH_3CN (0.2 mL), after stirring at room temperature for 10 minutes, the mixture was cooled to –30 °C or 0 °C. After stirring for 10 min at –30 °C or 0 °C, nitron **2** (0.22 mmol) and CH_3CN (0.3 mL) were added. The mixture was further stirred at –30 °C or 0 °C for the time indicated in Table 4. The resulting solution was purified by column chromatography (petroleum ether/AcOEt) on silica gel to afford the products.

(3*S*,4*S*)-1-(4-Carboethoxyphenyl)-3,4-diphenyl-2-azetidinone (3a). White solid, 68.3 mg, 92% yield (*cis*), 90% ee. $[\alpha]_D^{20}$ = –11.3 (*c* 0.80, $CHCl_3$). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min $^{-1}$, 254 nm, *cis*: t_r = 14.6 min (3*S*4*S*, major) and t_r = 15.9 min (3*R*4*R*, minor); *trans*: t_r = 17.5 min and t_r = 18.3 min (ref. 12: CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min $^{-1}$, *cis*: t_r = 17.5 min, t_r = 18.9 min; *trans*: t_r =



22.1 min (3S4R), t_r = 23.6 min (3R4S)). The ee was also determined by HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 95/5, flow rate = 1.0 mL min⁻¹, *cis*: t_r = 25.3 min (3S4S, major) and t_r = 35.8 min (3R4R, minor); *trans*: t_r = 12.8 min and t_r = 16.3 min (ref. 8a: Chiralcel OD, hexane/2-propanol 95/5, flow rate = 1.0 mL min⁻¹, *cis*: t_r = 19.8 min (3S4S), t_r = 38.6 min (3R4R)). ¹H NMR (600 MHz, CDCl₃) δ 7.97 (d, J = 8.7 Hz, 2H), 7.44 (d, J = 8.7 Hz, 2H), 7.11–7.03 (m, 10H), 5.50 (d, J = 6.2 Hz, 1H), 5.05 (d, J = 6.3 Hz, 1H), 4.35–4.31 (m, 2H), 1.35 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.0, 165.9, 141.2, 133.8, 131.7, 130.9, 128.8, 128.3, 128.2, 128.1, 127.3, 127.1, 125.9, 116.7, 60.9, 60.6, 60.5, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-3-(4-methylphenyl)-4-phenyl-2-azetidinone (3b). White solid, 71.7 mg, 93% yield (*cis*), 88% ee. $[\alpha]_D^{20}$ = -12.1 (c 0.72, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, t_r = 13.6 min (major) and t_r = 17.7 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.97 (d, J = 8.5 Hz, 2H), 7.43 (d, J = 8.5 Hz, 2H), 7.13–7.11 (m, 3H), 7.05–7.04 (m, 2H), 6.93–6.88 (m, 4H), 5.48 (d, J = 6.2 Hz, 1H), 5.02 (d, J = 6.2 Hz, 1H), 4.35–4.32 (m, 2H), 2.17 (s, 3H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.3, 166.0, 141.2, 137.0, 133.9, 130.9, 128.9, 128.7, 128.5, 128.4, 128.1, 127.1, 125.8, 116.7, 60.9, 60.6, 60.4, 21.1, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-3-(3-methylphenyl)-4-phenyl-2-azetidinone (3c). White solid, 70.2 mg, 91% yield (*cis*), 91% ee. $[\alpha]_D^{20}$ = -11.4 (c 0.69, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, t_r = 12.4 min (major) and t_r = 13.6 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.97 (d, J = 8.5 Hz, 2H), 7.44 (d, J = 8.5 Hz, 2H), 7.12–6.95 (m, 6H), 6.86–6.80 (m, 3H), 5.49 (d, J = 6.2 Hz, 1H), 5.02 (d, J = 6.2 Hz, 1H), 4.36–4.32 (m, 2H), 2.16 (s, 3H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.1, 166.0, 141.2, 137.7, 133.9, 131.5, 130.9, 129.5, 128.3, 128.1, 128.03, 128.00, 127.1, 125.9, 125.8, 116.7, 60.9, 60.6, 60.6, 21.2, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-3-(4-fluorophenyl)-4-phenyl-2-azetidinone (3d). White solid, 57.6 mg, 74% yield (*cis*), 88% ee. $[\alpha]_D^{20}$ = -5.6 (c 0.3, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 85/15, flow rate = 1.0 mL min⁻¹, t_r = 16.7 min (major) and t_r = 14.3 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.98 (d, J = 8.7 Hz, 2H), 7.43 (d, J = 8.7 Hz, 2H), 7.14–7.13 (m, 3H), 7.03–7.01 (m, 4H), 6.80–6.77 (m, 2H), 5.50 (d, J = 6.2 Hz, 2H), 5.03 (d, J = 6.2 Hz, 1H), 4.36–4.32 (m, 2H), 1.36 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.0, 165.7, 162.8, 161.1, 141.1, 133.7, 130.9, 130.5, 130.4, 128.51, 128.3, 127.0, 126.0, 116.8, 115.3, 115.1, 60.9, 60.5, 59.8, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-3-(cyclohexyl)-4-phenyl-2-azetidinone (3e). White solid, 51 mg, 68% yield (*cis*), 12% ee. $[\alpha]_D^{20}$ = -11.8 (c 0.57, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 95/5, flow rate = 1.0 mL min⁻¹, t_r = 17.8 min (3S4S, major) and t_r = 15.6 min (3R4R, minor) (ref. 12: CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *cis*: t_r = 14.4 min (3R4R), t_r = 16.6 min (3S4S); *trans*: t_r = 19.6 min, t_r = 25.9 min). ¹H NMR (600 MHz, CDCl₃) δ 7.91 (d, J = 8.1 Hz, 2H), 7.44–7.28 (m, 7H), 5.18 (d, J = 5.9 Hz, 1H), 4.31 (d, J = 7.0 Hz, 2H), 3.37–3.34 (m, 1H), 2.21 (s, 1H), 1.66 (s, 2H), 1.51 (s, 2H), 1.34 (t, J =

7.2 Hz, 3H), 1.18–1.10 (m, 4H), 0.76–0.66 (m, 2H). ¹³C NMR (150 MHz, CDCl₃) δ 167.8, 166.1, 141.1, 137.2, 134.6, 130.8, 129.0, 128.7, 128.6, 125.3, 124.9, 116.6, 115.9, 62.9, 60.8, 60.7, 60.7, 58.6, 34.9, 31.1, 30.7, 29.7, 29.7, 26.1, 25.5, 25.4, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-4-(3-methylphenyl)-3-phenyl-2-azetidinone (3f). White solid, 61.7 mg, 80% yield (*cis*), 90% ee. $[\alpha]_D^{20}$ = -11.5 (c 0.93, CHCl₃). The ee was determined by HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 90/10, flow rate = 0.8 mL min⁻¹, t_r = 15.4 min (major) and t_r = 20.0 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.97 (d, J = 8.5 Hz, 2H), 7.44 (d, J = 8.5 Hz, 2H), 7.10–7.04 (m, 5H), 6.98 (t, J = 7.8 Hz, 1H), 6.89 (d, J = 7.6 Hz, 1H), 6.83 (d, J = 6.3 Hz, 2H), 5.45 (d, J = 6.2 Hz, 1H), 5.03 (d, J = 6.2 Hz, 1H), 4.33 (q, J = 7.1 Hz, 2H), 3.14 (s, 3H), 1.35 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.04, 166.00, 141.3, 138.0, 134.0, 131.8, 130.8, 128.9, 128.8, 128.2, 128.1, 127.7, 125.8, 124.2, 116.7, 60.9, 60.6, 60.5, 21.2, 14.3. ESI-HRMS calcd for C₂₅H₂₄NO₃⁺ [M + H]⁺: 386.1751, found 386.1725.

(3S,4S)-1-(4-Carboethoxyphenyl)-4-(4-methylphenyl)-3-phenyl-2-azetidinone (3g). White solid, 62.4 mg, 81% yield (*cis*), 83% ee. $[\alpha]_D^{20}$ = -18.3 (c 0.9, CHCl₃). The ee was determined by HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 90/10, flow rate = 1 mL min⁻¹, t_r = 12.6 min (major) and t_r = 17.4 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.96 (d, J = 8.8 Hz, 2H), 7.43 (d, J = 8.6 Hz, 2H), 7.11–7.05 (m, 5H), 6.93–6.90 (m, 4H), 5.46 (d, J = 6.2 Hz, 1H), 5.03 (d, J = 6.2 Hz, 1H), 4.35–4.31 (m, 2H), 2.18 (s, 3H), 1.35 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.1, 166.0, 141.2, 137.9, 131.9, 130.8, 130.7, 129.1, 128.8, 128.2, 127.3, 127.0, 125.8, 116.8, 60.8, 60.5, 60.5, 21.1, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-4-(2-naphthyl)-3-phenyl-2-azetidinone (3h). Light yellow solid, 59 mg, 70% yield (*cis*), 77% ee. $[\alpha]_D^{20}$ = -25.5 (c 0.53, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, t_r = 18.11 min (major) and t_r = 21.51 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.95 (d, J = 8.8 Hz, 2H), 7.69–7.62 (m, 3H), 7.54 (d, J = 8.7 Hz, 1H), 7.47 (d, J = 8.8 Hz, 2H), 7.43–7.38 (m, 2H), 7.10–6.95 (m, 6H), 5.65 (d, J = 6.2 Hz, 1H), 5.13 (d, J = 6.2 Hz, 1H), 4.34–4.30 (m, 2H), 1.34 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.04, 165.96, 141.3, 132.9, 132.9, 131.6, 131.6, 130.9, 128.7, 128.2, 128.2, 127.8, 127.7, 127.4, 126.7, 126.3, 126.3, 125.9, 124.2, 116.7, 60.9, 60.8, 60.7, 14.3.

(3S,4S)-1-(4-Carboethoxyphenyl)-3-(4-methylphenyl)-4-(3-methylphenyl)-2-azetidinone (3i). White solid, 72.7 mg, 91% yield (*cis*), 89% ee. $[\alpha]_D^{20}$ = -16.0 (c 1.09, CHCl₃). The ee was determined by HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 90/10, flow rate = 1 mL min⁻¹, t_r = 11.86 min (major) and t_r = 14.50 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.96 (d, J = 8.7 Hz, 2H), 7.43 (d, J = 8.7 Hz, 2H), 6.99 (t, J = 7.9 Hz, 1H), 6.93–6.88 (m, 5H), 6.84 (d, J = 7.2 Hz, 2H), 5.42 (d, J = 6.2 Hz, 1H), 4.99 (d, J = 6.2 Hz, 1H), 4.35–4.31 (m, 2H), 2.17–2.15 (d, J = 9.0 Hz, 6H), 1.35 (t, J = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.4, 166.0, 141.3, 137.9, 136.9, 133.8, 130.8, 128.8, 128.6, 128.2, 127.8, 125.7, 124.2, 116.7, 60.8, 60.6, 60.4, 21.3, 21.0, 14.3. ESI-HRMS calcd for C₂₆H₂₆NO₃⁺ [M + H]⁺: 400.1907, found 400.1899.

(3S,4S)-1-(4-Carboethoxyphenyl)-3,4-(3-methylphenyl)-2-azetidinone (3j). Light yellow liquid, 71.9 mg, 90% yield (*cis*), 89% ee. $[\alpha]_D^{20}$ = -16.9 (c 1.32, CHCl₃). The ee was determined by



HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 90/10, flow rate = 1 mL min⁻¹, *t_r* = 12.24 min (major) and *t_r* = 15.45 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.97 (d, *J* = 8.8 Hz, 2H), 7.44 (d, *J* = 8.8 Hz, 2H), 7.00–6.95 (m, 2H), 6.90–6.80 (m, 6H), 5.43 (d, *J* = 6.2 Hz, 1H), 4.99 (d, *J* = 6.2 Hz, 1H), 4.35–4.31 (m, 2H), 2.15 (d, *J* = 5.0 Hz, 6H), 1.35 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.2, 166.0, 147.3, 137.9, 137.6, 133.8, 131.6, 130.8, 129.5, 128.8, 128.1, 128.0, 127.9, 127.7, 125.9, 125.7, 124.2, 116.7, 60.8, 60.6, 21.2, 21.2, 14.3. ESI-HRMS calcd for C₂₆H₂₆NO₃⁺ [*M* + *H*]⁺: 400.1907, found 400.1898.

(3*S*,4*S*)-1-(4-Carboethoxyphenyl)-3,4-(4-methylphenyl)-2-azetidinone (3k). Light yellow liquid, 63.9 mg, 80% yield (*cis*), 79% ee. [*α*]_D²⁰ = −27.2 (*c* 0.94, CHCl₃). The ee was determined by HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 90/10, flow rate = 1 mL min⁻¹, *t_r* = 8.84 min (major) and *t_r* = 11.77 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.95 (d, *J* = 8.8 Hz, 2H), 7.42 (d, *J* = 8.7 Hz, 2H), 6.94–6.89 (m, 8H), 5.44 (d, *J* = 6.2 Hz, 1H), 4.98 (d, *J* = 6.2 Hz, 1H), 4.35–4.31 (m, 2H), 2.18 (d, *J* = 7.2 Hz, 6H), 1.35 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.4, 166.0, 141.3, 137.8, 136.8, 130.8, 129.1, 128.9, 128.7, 128.7, 127.1, 125.7, 116.7, 60.8, 60.5, 60.3, 21.1, 21.1, 14.3. ESI-HRMS calcd for C₂₆H₂₆NO₃⁺ [*M* + *H*]⁺: 400.1907, found 400.1898.

(3*S*,4*S*)-1-(4-Carboethoxyphenyl)-3-(3-methylphenyl)-4-(4-methylphenyl)-2-azetidinone (3l). Light yellow liquid, 65.5 mg, 82% yield (*cis*), 79% ee. [*α*]_D²⁰ = −28.1 (*c* 0.89, CHCl₃). The ee was determined by HPLC analysis using a Chiralcel OD-H column, hexane/2-propanol 90/10, flow rate = 1 mL min⁻¹, *t_r* = 10.94 min (major) and *t_r* = 14.99 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.96 (d, *J* = 8.7 Hz, 2H), 7.43 (d, *J* = 8.7 Hz, 2H), 6.98–6.81 (m, 8H), 5.45 (d, *J* = 6.2 Hz, 1H), 4.98 (d, *J* = 6.2 Hz, 1H), 4.35–4.31 (m, 2H), 2.17 (d, *J* = 12.0 Hz, 6H), 1.35 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 166.3, 166.0, 141.3, 137.8, 137.7, 131.7, 130.8, 130.8, 129.5, 129.0, 127.99, 127.97, 127.0, 125.9, 125.7, 116.7, 60.8, 60.50, 60.47, 21.2, 21.1, 14.3. ESI-HRMS calcd for C₂₆H₂₆NO₃⁺ [*M* + *H*]⁺: 400.1907, found 400.1898.

(3*S*,4*S*)-1,3,4-Triphenyl-2-azetidinone (3m). White solid, 56.0 mg, 93% yield (*cis*), 91% ee. [*α*]_D²⁰ = −9.2 (*c* 0.54, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *t_r* = 10.35 min (3*S*4*S*, major) and *t_r* = 16.92 min (3*R*4*R*, minor). (ref. 12: CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *cis*: *t_r* = 11.4 min (3*S*4*S*), *t_r* = 19.2 min (3*R*4*R*); *trans*: *t_r* = 10.0 min, *t_r* = 13.2 min) ¹H NMR (600 MHz, CDCl₃) δ 7.42–7.41 (m, 2H), 7.30–7.25 (m, 2H), 7.11–7.04 (m, 11H), 5.47 (d, *J* = 6.2 Hz, 1H), 5.01 (d, *J* = 6.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 165.7, 137.7, 134.4, 132.1, 129.1, 128.9, 128.2, 128.1, 127.9, 127.2, 127.1, 124.1, 117.2, 60.3, 60.3.

(3*S*,4*S*)-1-(4-Methylphenyl)-3,4-diphenyl-2-azetidinone (3n). White solid, 44.5 mg, 71% yield (*cis*), 85% ee. [*α*]_D²⁰ = −12.19 (*c* 0.38, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *t_r* = 12.75 min (major) and *t_r* = 17.47 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.3 (d, *J* = 8.4 Hz, 2H), 7.10–7.03 (m, 12H), 5.43 (d, *J* = 6.1 Hz, 1H), 4.99 (d, *J* = 6.1 Hz,

1H), 2.29 (s, 3H). ¹³C NMR (150 MHz, CDCl₃) δ 165.4, 135.3, 134.5, 133.7, 132.2, 129.6, 128.9, 128.2, 128.1, 127.8, 127.13, 127.11, 117.2, 60.3, 60.2, 20.9.

(3*S*,4*S*)-1-(4-Chlorophenyl)-3,4-diphenyl-2-azetidinone (3o). White solid, 53.4 mg, 80% yield (*cis*), 87% ee. [*α*]_D²⁰ = −8.13 (*c* 0.57, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *t_r* = 10.66 min (major) and *t_r* = 14.34 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.35 (d, *J* = 8.8 Hz, 2H), 7.23 (d, *J* = 8.8 Hz, 2H), 7.11–7.03 (m, 10H), 5.44 (d, *J* = 6.1 Hz, 1H), 5.02 (d, *J* = 6.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 165.5, 136.2, 133.9, 131.8, 129.2, 129.1, 128.8, 128.3, 128.1, 128.0, 127.3, 127.1, 118.4, 60.6, 60.4.

(3*S*,4*S*)-1-(4-Bromophenyl)-3,4-diphenyl-2-azetidinone (3p). White solid, 62.0 mg, 82% yield (*cis*), 87% ee. [*α*]_D²⁰ = −10.23 (*c* 0.72, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *t_r* = 11.44 min (major) and *t_r* = 15.40 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.38 (d, *J* = 8.8 Hz, 2H), 7.29 (d, *J* = 8.8 Hz, 2H), 7.11–7.02 (m, 10H), 5.43 (d, *J* = 6.1 Hz, 1H), 5.02 (d, *J* = 6.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 165.6, 136.6, 133.9, 132.1, 131.8, 128.8, 128.3, 128.12, 128.06, 127.3, 127.1, 118.8, 116.7, 60.6, 60.4.

(3*S*,4*S*)-1-(4-Fluorophenyl)-3,4-diphenyl-2-azetidinone (3q). White solid, 44.4 mg, 70% yield (*cis*), 86% ee. [*α*]_D²⁰ = −7.0 (*c* 0.34, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 90/10, flow rate = 1.0 mL min⁻¹, *t_r* = 10.43 min (major) and *t_r* = 13.67 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.40–7.37 (m, 2H), 7.11–6.96 (m, 12H), 5.44 (d, *J* = 6.1 Hz, 1H), 5.02 (d, *J* = 6.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 165.3, 159.9, 158.3, 134.1, 133.9 (d, *J* = 10.7), 132.0, 128.9, 128.3, 128.1, 128.0, 127.2, 127.1, 118.66, 118.61, 116.0, 115.8, 60.53, 60.51. ESI-HRMS calcd for C₂₁H₁₇FNO⁺ [*M* + *H*]⁺: 318.1289, found 318.1284.

(3*S*,4*S*)-3-(4-Fluorophenyl)-1,4-diphenyl-2-azetidinone (3r). White solid, 47.6 mg, 75% yield (*cis*), 84% ee. [*α*]_D²⁰ = −5.4 (*c* 0.24, CHCl₃). The ee was determined by HPLC analysis using a CHIRALPAK IA column, hexane/2-propanol 85/15, flow rate = 1.0 mL min⁻¹, *t_r* = 8.42 min (major) and *t_r* = 15.66 min (minor). ¹H NMR (600 MHz, CDCl₃) δ 7.41–7.40 (m, 2H), 7.30–7.25 (m, 2H), 7.14–7.01 (m, 8H), 6.79–6.76 (m, 2H), 5.45 (d, *J* = 6.1 Hz, 1H), 4.97 (d, *J* = 6.1 Hz, 1H). ¹³C NMR (150 MHz, CDCl₃) δ 165.4, 162.7, 161.0, 137.6, 134.2, 130.5 (d, *J* = 32.5), 129.1, 128.4, 128.1, 127.0, 124.2, 117.2, 115.1, 115.0, 60.2, 59.5. ESI-HRMS calcd for C₂₁H₁₇FNO⁺ [*M* + *H*]⁺: 318.1289, found 318.1290.

Conflicts of interest

There are no conflicts to declare.

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