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One-pot oxime ligation from peptides bearing thiazolidine and aminooxyacetyl groups†

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One-pot oxime ligation under mild conditions using Pd(II) as a shared catalyst from an aldehyde precursor (Thz) and a protected aminooxyacetyl group (Proc-Aoa) is reported. Two complementary metal-free protocols using unmasked Aoa-peptide are also described. Acetoxime-peptide can proceed to the desired oxime through an additional transoximation step.

Oxime ligation is one of the hot topics in the field of bioconjugation of molecules due to its chemoselectivity and hydrolytic stability.^{1,2} It requires an electrophilic moiety such as a carbonyl group (aldehyde/ketone) and a nucleophilic amino-oxy partner (H₂N–O–R). Numerous methods have emerged over the past decade to insert these two components into peptide sequences.^{1,3} A critical step in oxime ligation is aldehyde/ketone formation. The functionalization strategies that have been developed using either chemical, enzymatic or bioengineering methods exhibit disparities in the selectivity and the facility of this electrophile formation.⁴

Thiazolidine (named Thz) was first used as an orthogonal cysteine protection strategy, especially in native chemical ligation (NCL) (Fig. 1a, right pathway, R = H).⁵ Such 4-carboxy thiazolidine can be introduced at the N-terminus of peptide by using commercially available protected thioproline (ThP) component. The cysteine residue can then be generated by treating the ThP-peptide with methoxyamine in buffer^{6,7} or by Pd(II) catalyst.⁸ Recent works used 2-carboxy thiazolidine to introduce a highly reactive α -oxo aldehyde for further bioconjugation.^{9,10} (Fig. 1a, left pathway, R' = H). This second thiazolidine can be appended into peptides with the precursor Boc-Thz-OH (Fig. 1b), then hydrolyzed with a metal-assisted ring opening using either silver or palladium catalysts.^{9,10} The scope of thiazolidine in bioconjugation was recently expanded with the use of thiazolidin-5-imine as a new linkage method in protein and live cell labelling.¹¹

In oxime conjugation, the other component is the amino-oxyacetyl peptide. Given the high nucleophilic reactivity of the amino-oxy moiety that traps laboratory ambient traces of

acetone, acetaldehyde or formaldehyde,^{12–14} Mezö *et al.* proposed to add 10-fold excess of free aminooxyacetic acid (Aoa-OH) as a carbonyl capture to overcome this undesired oxime bond.¹³ When this strategy is needed, it requires an additional purification step prior to oxime ligation to remove the excess of Aoa-OH. Thus, the use of an orthogonal protecting group for conventional Fmoc peptide chemistry can ease the purification and the storage of Aoa-peptides. Propargyloxycarbonyl (Proc) protecting group (Fig. 1b) was used to protect Aoa-OH in solid-phase peptide synthesis (SPPS) as it remains stable under harsh acidic condition like resin cleavage and can be selectively deprotected by several Pd(II) catalysts such as [Pd(allyl)Cl]₂ and Pd(dba)₂.^{8,15}

Several oxime conjugation methods were reported using Pd(II) or Ag(I) to uncage of 2-carboxy thiazolidine (Fig. 2a).^{9,10} These strategies have been used to label peptide or protein with biotin or fluorophore but they were not applied to peptide-peptide ligation. Proc group has been used to protect N-terminus cysteine for NCL purpose (Fig. 2b).⁸ All of these procedures required additional purification steps to isolate the intermediates from the high amount of catalysts. The need of

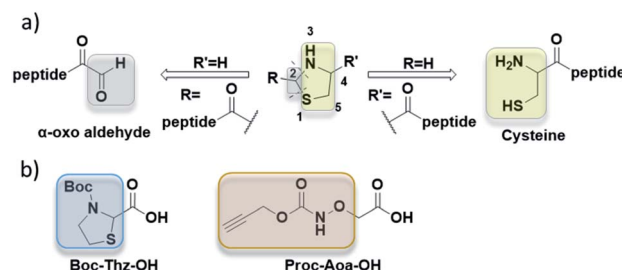


Fig. 1 (a) 2-Carboxy thiazolidine as a new highly reactive aldehyde precursor (Thz, left pathway). 4-Carboxy thiazolidine as N-term cysteine protection (thioproline analogue (ThP), right pathway). (b) Boc protected 1,3-thiazolidine-2-carboxylic acid (Boc-Thz-OH) and propargyloxycarbonyl (Proc) protected aminooxyacetic acid (Proc-Aoa-OH).

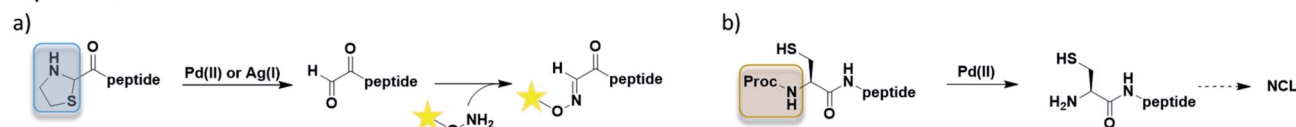
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Reported work



Present work

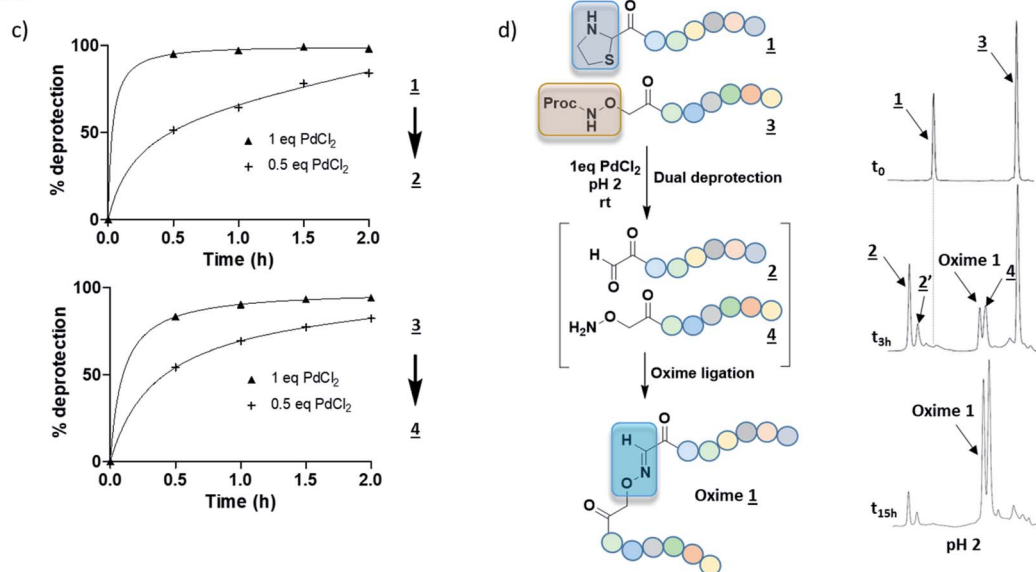


Fig. 2 (a) 2 steps synthesis with thiazolidine deprotection mediated by either [Pd] or [Ag] complex followed by a purification step and oxime coupling with nude aminoxy tag (biotine or dansyl).^{9,10} (b) Proc removal with [Pd] complex in NCL.⁸ (c) Kinetic studies of Pd-mediated deprotection of Thz or Proc groups in pH 2 solution. % deprotection was calculated from HPLC peak areas. (d) One-pot method starting from both protected partners to reach oxime ligation at pH 2 with HPLC monitoring. Peptides were numbered as in Table 1.

a precious care of the highly reactive functions α -oxo aldehyde and aminoxy groups before oxime conjugation led us to develop new methods.

In our project to deliver a cyclic 17-mer peptide into cells, the oxime bond was adopted to append a cell penetrating peptide (CPP).^{16,17} Boc-Thz-OH (Fig. 1b) was rapidly prepared according to the reported method⁹ and was introduced during SPPS at N-terminus of a peptide, with a flexible spacer Ahx (6-amino hexanoic acid) between Thz and the linear peptide of sequence CPSDSTRRKGGRCGRRL. The peptide was then cleaved, totally deprotected and finally cyclized between the side chains of Cys residues through an *o*-xylene moiety^{18,19} to give peptide 1 (formula shown in ESI[†]), the precursor of peptide 2 (Table 1). Among the available vectors, antennapedia (Antp) is well known for its fast delivery of cargo peptides into cells.^{20,21} Proc-Aoa-OH was prepared by protection of free Aoa-OH with propargyl chloroformate in basic condition (ESI[†]) then similarly introduced at N-terminus of an antennapedia derived 16-mer of sequence RQIKIWFQNRRMKWKK²² to give peptide 3. Peptide 4 was prepared by using commercial Boc-Aoa-OH (Table 1).

To better ease the conjugation between Thz-peptides and Aoa-peptides, we aimed to develop a convenient one-pot deprotection-conjugation procedure with both protected partners. Thiazolidine ring-opening can be mediated by Ag(I).¹⁰ Nonetheless, as Ag(I) cannot deprotect Proc group, we focused

our attention on Pd(II) as a shared catalyst for both Thz and Proc uncaging. Several Pd(II) complexes including [Pd(allyl)Cl]₂, Pd(OAc)₂ and PdCl₂ were first tested. In agreement with reported works,^{8,9} [Pd(allyl)Cl]₂ and PdCl₂ exhibited better results than Pd(OAc)₂ (data not shown). Proc group was easily removed by both PdCl₂ and [Pd(allyl)Cl]₂, whereas the latter was known for giving an unidentified by-product during Proc removal.⁸ Taken together, these preliminary results focused our method on PdCl₂ as a shared catalyst. Due to its limited aqueous solubility, a 1.25 mM PdCl₂ stock solution in HPLC mobile phase (aqueous 0.1% TFA, pH 2, named solution A) was prepared for deprotection experiments. Proc and Thz deprotection were first

Table 1 Peptide and oxime sequences with mass found in MALDI-TOFF experiments. Ahx: 6-amino hexanoic acid. *thioether *o*-xylene cyclization (formula shown in ESI)

Compound	Sequence	Mass found (Da)
Peptide 1	Thz-Ahx-C*PSDSTRRKGGRC*GRRL-NH ₂	2234
Peptide 2	CHO-(CO)-Ahx-C*PSDSTRRKGGRC*GRRL-NH ₂	2175
Peptide 3	Proc-Aoa-RQIKIWFQNRRMKWKK-NH ₂	2400
Peptide 4	Aoa-RQIKIWFQNRRMKWKK-NH ₂	2318
Oxime 1	Peptide 2-peptide 4 conjugate	4473
Oxime 2	Acetoxime of peptide 4	2358

investigated separately, using 0.5 and 1.0 equivalent of PdCl_2 in solution A at room temperature with HPLC monitoring during 0–2 h (Fig. 2c). Due to the release of β -mercaptoethylamine, Pd can be captured and not proceed in catalytic amounts. 0.5–1.0 eq. of PdCl_2 exhibited optimal Thz and Proc deprotection within few hours. Based on these results, a one-pot conjugation protocol was designed using 1.0 eq. PdCl_2 and a slight excess of Proc-Aoa-peptide 3 (about 1.5 eq.) in solution A. This convenient approach requiring less steps and a minimum amount of Pd would provide a powerful tool for peptide–peptide ligation. In agreement with the kinetic studies, after 3 h Thz-peptide 1 showed almost complete deprotection into 2 in comparison with Proc-peptide 3, which was partially deprotected (Fig. 2d). 2 was observed with its hydrated form 2' according to the reported work of Brabham *et al.* in Thz deprotection.⁹ We suppose through HSAB theory that the better Lewis base property of sulfur in Thz tends to faster its hydrolysis comparing to Proc group. A new peak corresponding to **Oxime 1** appeared. After an overnight stirring (15 h), a great conversion of the α -oxoaldehyde 2 into oxime was achieved and the excess of 3 was fully deprotected. PdCl_2 catalyst was then quenched by an aqueous solution of dithiothreitol (DTT) (50 eq.).²³ As our Aoa-peptide 4 and the resulting **Oxime 1** have similar retention times, the excess of free Aoa-peptide was easily converted into acetoxime derivative, named **Oxime 2** (Table 1) upon acetone addition. This retention time increase alleviated **Oxime 1** purification.

Although only 1.0 eq. of PdCl_2 is needed to remove both Thz and Proc, metal ions might be caged by Cys/Met/His rich proteins and may induce undesired biological effects.²⁴ Consequently, very stringent standards are applied in drug for limit content (EMA/CHMP/ICH/353369/2013). In our case, as a last

step prior to cell assays, metal-free conditions might be preferable. Recent strategies have emerged to promote oxime ligation using other electrophile precursors such as *N*-phenylglycine peptides.²⁵ Regarding the thiazolidine ring, metal-free ThP uncaging was described with a large excess of methoxyamine (>500 eq.) in 8–12 h under acidic conditions (Fig. 3a).^{6,7} The released formaldehyde was directly quenched into oxime by methoxyamine. Hence, further studies were carried out to investigate the propensity of our Aoa-peptide 3 to mimic this metal-free thiazolidine deprotection-quench. This first-in-class method would allow Aoa-peptide to deprotect and form the desired **Oxime 1** whilst releasing the β -mercaptoethylamine moiety. Two short model peptides, Thz-GYRMHK 5 and Aoa-GYRMHK 6, were thus prepared for this metal-free ligation. Among the screened conditions, solution A and standard ammonium acetate buffer at pH 4.5 with or without aniline catalyst²⁶ exhibited better results compared to conventional protein chemistry buffer PBS 1 \times (Table S1, ESI†). Additionally, a mild heat at 37 °C is strongly needed to complete the deprotection and thus the conjugation as recently described by Streefkerk *et al.*¹⁹ Numerous amine catalysts have been investigated to accelerate the reaction in oxime ligation.^{27,28} In our case, 10 mM aniline did not show catalyst property. More than 95% conversion was reached after 48 h treatment at 37 °C in solution A (Table S1, ESI†). Room temperature experiment gave 44% conversion in the same conditions (Table S1, ESI†). Taken together, these results exhibit the key roles played by both the low pH and the temperature in Thz uncaging and oxime formation without metal catalyst. We applied these conditions to our peptides of interest 1 and unprotected peptide 4 at 37 °C. After an overnight reaction, ~60% of peptide 1 were converted

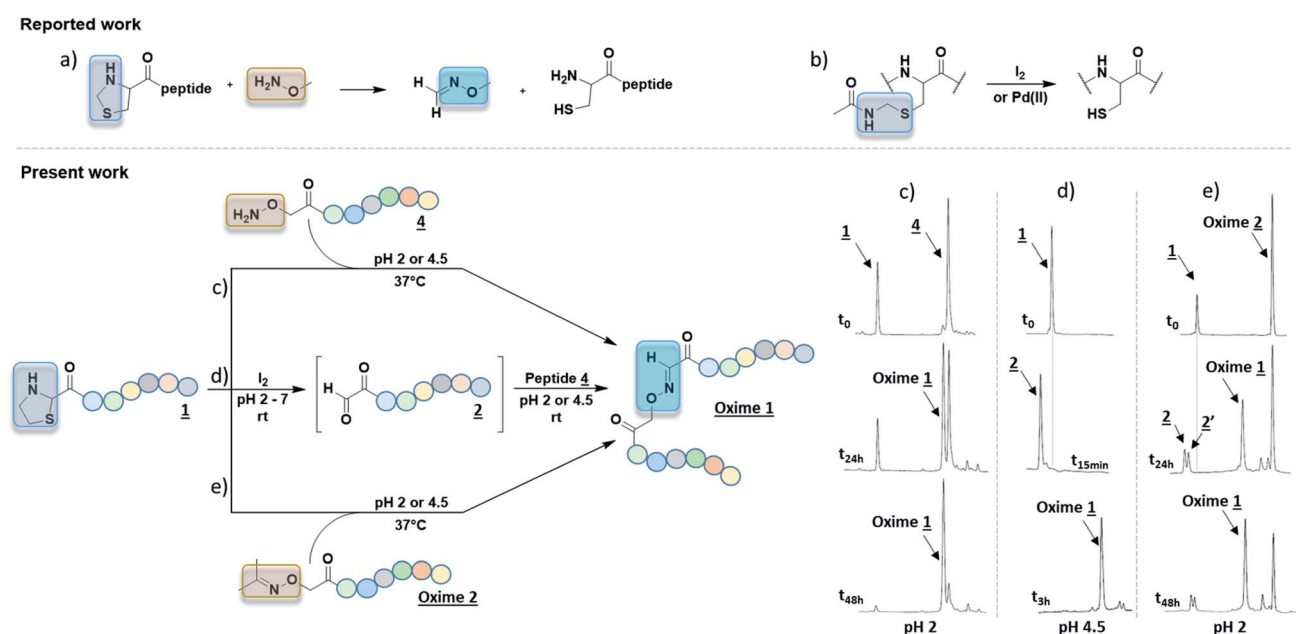


Fig. 3 (a) 4-Carboxy thiazolidine deprotected by methoxylamine for further NCL.^{6,7} (b) Cys(Acm) deprotection mediated by iodine or Pd(II) complex.³⁰ One-pot metal-free 2-carboxy thiazolidine deprotection using aminooxy assisted acidic hydrolysis at 37 °C (c); iodine at room temperature (d) or by transoximation (e) to reach peptide–peptide oxime bond. HPLC monitoring is depicted to illustrate these three different conditions.

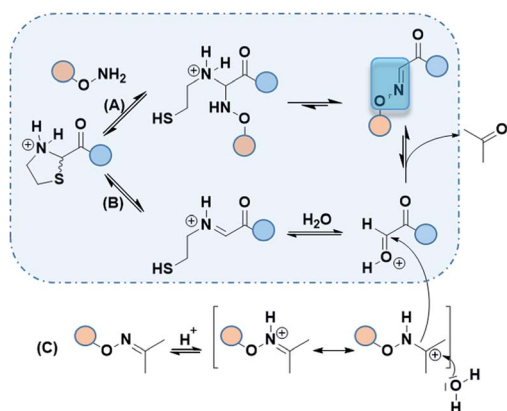
into oxime and one additional night resulted in >95% neat conversion at pH 2 (Fig. 3c). It has been reported in a mechanism study that 2-substituted thiazolidine proceeds to ring opening at 50–90 °C through an iminium intermediate.²⁹ Thz-peptide in solution A at 37 °C gave 45% and 65% of hydrolyzed α -oxo aldehyde after 24 h and 48 h respectively, meanwhile it remained stable at room temperature (data not shown). Combined, these results suggest that two pathways might be involved in this conjugation to reach >95% conversion. The slow acidic Thz hydrolysis releases the aldehyde that subsequently reacts with the aminooxy moiety to give the corresponding oxime (Scheme 1, pathway B). Interestingly, when comparing HPLC spectra in Fig. 2d and 3c, no aldehyde-peptide 2 was detected in the presence of free Aoa-peptide 4. Considering the slow rate of the Thz hydrolysis and the fact that aniline did not catalyze the process, we suppose that nucleophilic Aoa might assist the thiazolidine ring opening and directly proceed to oxime conjugation (Scheme 1, pathway A).

As Thz opening remains the slowest step, we focused our interest on promoting this deprotection. Recent work described the use of Pd(II) complex to deprotect Cys(Acm) compared to conventional iodine method (Fig. 3b).³⁰ Given the similarity between cysteine side chain protecting group acetamidomethyl (Acm) and thiazolidine function, we hypothesized that iodine could deprotect Thz as well as Acm. Thz-GYRMHK 5 was thus treated at room temperature with different amounts of iodine (0.2, 0.4, 0.6 and 1.0 eq.) at pH 2, 4.5 and 7 (Table S3, ESI†). 1.0 eq. iodine was found to achieve neat and rapid Thz deprotection within 15 min irrespective of the pH condition. In these conditions, peptide sequence was stable and Tyr was not iodinated. These conditions were used to afford **Oxime 1** from the peptides of interest 1 and 4. As in the case of model peptides, full Thz deprotection was achieved by 1.0 eq. of iodine in water or in pH 2, 4.5 and 7 buffers at room temperature. After iodine quench upon sodium thiosulfate addition (1.2 eq.), complete conversion into **Oxime 1** was obtained in 3 h at room temperature in pH 4.5 buffer (Fig. 3d). Interestingly, we could

reach >85% conversion in pH 7 buffer containing 100 mM aniline after an overnight stirring at 37 °C (Fig. S1, ESI†).

Although free aminooxy is frequently used in recent works, its high reactivity towards volatile carbonyl compounds makes purification and storage not easy. Based on the reported transoximation observed during tandem oxime ligation,^{12,14} we investigated the equilibrium between two oxime states to appreciate the role of acetoxime by-product in peptide ligation. Hyodo *et al.* reported the catalyst role of several Brønsted acids including HClO₄, HCO₂H, AcOH, H₂SO₄, and TFA in small organic molecules transoximation.³¹ In their work, TFA gave sufficient transoximation rate. Moreover, among the oxime starting material that were described for transoximation, acetoxime was pleasantly ranked among the most reactive ones. The authors described transoximation protocol using 5–30 mol% HClO₄ with 40 °C heat, conditions that are inappropriate for peptide chemistry. Yuen *et al.* described a trans-hydrazonation in small molecule for cellular aldehydes labelling³² with several amine-based catalyst to promote the reaction. Although ethyl acetimidate has been described as a slightly more reactive precursor in acid catalyzed transoximation,³¹ the formation of the starting oxime would not be as easy as in the case of acetoxime.³³ As no evidences were found in transoximation between two peptides and dual Thz deprotection–transoximation (Scheme 1, pathway B + C), Aoa-GYRMHK 6 was beforehand converted upon acetone addition into acetoxime. When mixed with Thz-GYRMHK 5 in solution A, acetoxime peptide gave 9% transoximation at room temperature contrary to 60% in sealed vial and 71% in open vial experiments at 37 °C (Table S3, ESI†). Standard oxime buffer gave similar results with aniline at pH 4.5 while higher amount of acetoxime did not increase transoximation rate (Table S3, ESI†). Using peptide 1 and **Oxime 2** (3.0 eq.) (Fig. 3e), 60% transoximation were obtained after an overnight reaction in solution A at 37 °C to give **Oxime 1**. As in the case of Thz deprotection mediated by Pd complex (Fig. 2d), the slow transoximation allows the formation of 2 and 2' by acidic hydrolysis. These results show that it is unnecessary to remove acetoxime by-product as it can also perform oxime conjugation in acidic conditions with moderate heating.

In summary, three complementary methods to perform one pot oxime ligation have been demonstrated. The first one consisting of a 1.0 eq. PdCl₂ catalyzed Thz and Proc removal and subsequent *in situ* oxime conjugation at room temperature in pH 2. The second with metal-free aminooxy-assisted Thz opening and oxime conjugation at 37 °C in pH 2 or pH 4.5. The third method using 1.0 eq. iodine allows the oxime ligation to be performed either at room temperature in pH 2–4.5 or at 37 °C in physiological pH with aniline catalyst. We also show that acetoxime-peptide can undergo a transoximation combined with Thz hydrolysis to reach the desired oxime at 37 °C in pH 2 or pH 4.5. The Pd(II) mediated conjugation using both stable protected partners can be run at room temperature while the metal-free ligation requires either a moderate heat (37 °C) or a treatment with iodine to enhance Thz opening. HPLC mobile phase can be used routinely as it remains efficient and convenient without any aromatic amine catalyst. These results were



Scheme 1 Suggested mechanism involved in Aoa-assisted Thz opening followed by oxime ligation (pathway A). Acetoxime peptide can also proceed to oxime coupling through an additional transoximation step (pathway B + C).



validated for small and medium-size peptides including a large array of amino acids. As far as we know, these methods have never been reported.

Conflicts of interest

There are no conflicts to declare.

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Notes and references

- 1 D. K. Kölmel and E. T. Kool, *Chem. Rev.*, 2017, **117**, 10358–10376.
- 2 S. Ulrich, D. Boturyn, A. Marra, O. Renaudet and P. Dumy, *Chem.–Eur. J.*, 2014, **20**, 34–41.
- 3 S. M. Agten, P. E. Dawson and T. M. Hackeng, *J. Pept. Sci.*, 2016, **22**, 271–279.
- 4 R. J. Spears and M. A. Fascione, *Org. Biomol. Chem.*, 2016, **14**, 7622–7638.
- 5 A. C. Conibear, E. E. Watson, R. J. Payne and C. F. W. Becker, *Chem. Soc. Rev.*, 2018, **47**, 9046–9068.
- 6 M. Villain, J. Vizzavona and K. Rose, *Chem. Biol.*, 2001, **8**, 673–679.
- 7 V. Y. Torbeev and S. B. H. Kent, *Angew. Chem., Int. Ed.*, 2007, **46**, 1667–1670.
- 8 M. Jbara, S. K. Maity, M. Seenaiha and A. Brik, *J. Am. Chem. Soc.*, 2016, **138**, 5069–5075.
- 9 R. L. Brabham, R. J. Spears, J. Walton, S. Tyagi, E. A. Lemke and M. A. Fascione, *Chem. Commun.*, 2018, **54**, 1501–1504.
- 10 X. Bi, K. K. Pasunooti, J. Lescar and C.-F. Liu, *Bioconjugate Chem.*, 2017, **28**, 325–329.
- 11 X. Bi, J. Yin, C. Rao, S. Balamkundu, B. Banerjee, D. Zhang, D. Zhang, P. C. Dedon and C.-F. Liu, *Org. Lett.*, 2018, **20**, 7790–7793.
- 12 C. Buré, D. Lelièvre and A. Delmas, *Rapid Commun. Mass Spectrom.*, 2000, **14**, 2158–2164.
- 13 G. Mezö, I. Szabó, I. Kertész, R. Hegedüs, E. Orbán, U. Leurs, S. Bösze, G. Halmos and M. Manea, *J. Pept. Sci.*, 2011, **17**, 39–46.
- 14 D. Lelièvre, C. Buré, F. Laot and A. Delmas, *Tetrahedron Lett.*, 2001, **42**, 235–238.
- 15 J. Li, J. Yu, J. Zhao, J. Wang, S. Zheng, S. Lin, L. Chen, M. Yang, S. Jia, X. Zhang and P. R. Chen, *Nat. Chem.*, 2014, **6**, 352–361.
- 16 G. Guidotti, L. Brambilla and D. Rossi, *Trends Pharmacol. Sci.*, 2017, **38**, 406–424.
- 17 A. Borrelli, A. Tornesello, M. Tornesello and F. Buonaguro, *Molecules*, 2018, **23**, 295.
- 18 T. Nishihara, H. Kitada, D. Fujiwara and I. Fujii, *Pept. Sci.*, 2016, **106**, 415–421.
- 19 D. E. Streefkerk, M. Schmidt, J. H. Ippel, T. M. Hackeng, T. Nuijens, P. Timmerman and J. H. van Maarseveen, *Org. Lett.*, 2019, **21**, 2095–2100.
- 20 D. Cussac, M. Vidal, C. Leprince, W. Liu, F. Cornille, G. Tiraboschi, B. P. Roques and C. Garbay, *FASEB J.*, 1999, **13**, 31–38.
- 21 J. Dourlat, W.-Q. Liu, F. Sancier, T. Edmonds, P. Pamonsinlapatham, F. Cruzalegui and C. Garbay, *Biochimie*, 2009, **91**, 996–1002.
- 22 D. Derossi, A. H. Joliot, G. Chassaing and A. Prochiantz, *J. Biol. Chem.*, 1994, **269**, 10444–10450.
- 23 A. Krężel, W. Leśniak, M. Jeżowska-Bojczuk, P. Młynarz, J. Brasuñ, H. Kozłowski and W. Bal, *J. Inorg. Biochem.*, 2001, **84**, 77–88.
- 24 J.-F. Gaucher, M. Reille-Seroussi, N. Gagey-Eilstein, S. Broussy, P. Coric, B. Seijo, M.-B. Lascombe, B. Gautier, W.-Q. Liu, F. Huguenot, N. Inguibert, S. Bouaziz, M. Vidal and I. Broutin, *PLoS One*, 2016, **11**, e0167755.
- 25 Q. A. E. Guthrie and C. Proulx, *Org. Lett.*, 2018, **20**, 2564–2567.
- 26 A. Dirksen and P. E. Dawson, *Bioconjugate Chem.*, 2008, **19**, 2543–2548.
- 27 M. Wendeler, L. Grinberg, X. Wang, P. E. Dawson and M. Baca, *Bioconjugate Chem.*, 2014, **25**, 93–101.
- 28 M. Rashidian, M. M. Mahmoodi, R. Shah, J. K. Dozier, C. R. Wagner and M. D. Distefano, *Bioconjugate Chem.*, 2013, **24**, 333–342.
- 29 T. H. Fife, R. Natarajan, C. C. Shen and R. Bembi, *J. Am. Chem. Soc.*, 1991, **113**, 3071–3079.
- 30 S. K. Maity, M. Jbara, S. Laps and A. Brik, *Angew. Chem., Int. Ed.*, 2016, **55**, 8108–8112.
- 31 K. Hyodo, K. Togashi, N. Oishi, G. Hasegawa and K. Uchida, *Green Chem.*, 2016, **18**, 5788–5793.
- 32 L. H. Yuen, N. S. Saxena, H. S. Park, K. Weinberg and E. T. Kool, *ACS Chem. Biol.*, 2016, **11**, 2312–2319.
- 33 V. Duléry, O. Renaudet and P. Dumy, *Tetrahedron*, 2007, **63**, 11952–11958.

