


Cite this: *RSC Adv.*, 2020, 10, 30953

Received 8th April 2020
Accepted 11th August 2020

DOI: 10.1039/d0ra03160g

rsc.li/rsc-advances

Organic–inorganic nanocrystal reductase to promote green asymmetric synthesis†

Kotchakorn T. Sriwong, Afifa Ayu Koesoema and Tomoko Matsuda*

An acetophenone reductase from *Geotrichum candidum* (GcAPRD) was immobilized by the organic–inorganic nanocrystal method. The GcAPRD nanocrystal presented improved stability and recyclability compared with those of the free GcAPRD. Moreover, the GcAPRD nanocrystal reduced broad kinds of ketones with excellent enantioselectivities to produce beneficial chiral alcohols such as (S)-1-(3',4'-dichlorophenyl)ethanol with >99% yield and >99% ee. The robust and versatile properties of the GcAPRD nanocrystal demonstrated an approach to promote green asymmetric synthesis and sustainable chemistry.

1. Introduction

Biocatalysts have presented superior properties for green and sustainable synthesis with their high selectivity, turnover rate, and efficiency.^{1,2} For example, an alcohol dehydrogenase (ADH) from *Rhodococcus ruber* DSM 44541 expressed in *Arxula adeninivorans* is able to enantioselectively reduce *p*-chloroacetophenone with a remarkable turnover rate (K_{cat}) of 432 s^{−1}.^{3,4} Thus, the biocatalysts have played an essential role in an extensive area of industries such as fine and bulk chemical, pharmaceutical, food and beverage, cosmetic, textile, and pulp and paper.⁵ Meanwhile, the needs for chiral compounds have risen continuously, especially in the pharmaceutical fields, due to the different biological activities and toxicities that may be exhibited by each enantiomer of the chiral compounds.^{6,7}

Asymmetric reduction of ketones by chemical and biological catalysts has been considered as a promising method to produce chiral alcohols.^{8–10} Enzymes catalyzing asymmetric reduction reactions have been widely studied due to their high specificity property, and the well-known enzyme used for this purpose is ADH.¹⁰ An acetophenone reductase from *Geotrichum candidum* NBRC 4597 (GcAPRD) is a novel ADH that reduces ketones to their corresponding (S)-alcohols.¹¹ The GcAPRD has versatile and robust properties such as excellent enantioselectivity, broad substrate specificity, and non-aqueous solvents tolerance that make it potential for industrial applications.^{12–18} However, the free form of GcAPRD has limitations to recycle and operate in a flow process.

Enzyme immobilization is a method that improves stability and recyclability of the enzyme allowing the enzyme to operate

under wide conditions.² In most cases, the enzyme is confined with the immobilization supports that limits their mobility.¹⁹ Various techniques have been addressed to immobilize the enzyme, including the organic–inorganic nanocrystal method, first reported by Ge *et al.* in 2012.^{20–22} This method has been proven to be effective in immobilizing various biomolecules such as proteins, peptides, and antibodies.^{21,22} Moreover, limitations faced by other immobilization methods, such as insufficient binding of protein on support and severe loss of activities, have been overcome by the organic–inorganic nanocrystal method.^{19,23} Thereby, several studies have applied this method to immobilize broader kinds of biomolecules.²¹ Enzymes have been one of the most widespread biomolecules immobilized by this method for examples α -lactalbumin,²² laccase,²² carbonic anhydrase,²² lipases,^{22,24–26} α -amylases,²⁷ papain,^{28,29} and α -acetolactate decarboxylases.³⁰ However, only a few studies have focused on the immobilization of alcohol dehydrogenase (ADH) by this method,³¹ particularly for asymmetric synthesis,³² and there is no example yet of versatile ADH nanocrystals with broad substrate specificities.

Thus, this research aimed to immobilize the GcAPRD by using the organic–inorganic nanocrystal method. The preparation conditions of GcAPRD nanocrystal were optimized, and the properties of the GcAPRD nanocrystal were characterized. The GcAPRD nanocrystal showed improved stability and recyclability while retaining excellent enantioselectivity. To the best of our knowledge, this is the first study to report an ADH nanocrystal with broad substrate specificity.

2. Materials and methods

2.1 Reagents and materials

GcAPRD wild type was prepared according to the previous procedure.¹² Chemicals for cell cultivation, enzyme purification, and preparation of the GcAPRD nanocrystal were supplied from Nacalai Tesque (Japan), except for 4-(2-hydroxyethyl)-1-

Department of Life Science and Technology, School of Life Science and Technology, Tokyo Institute of Technology, 4259 Nagatsuta-cho, Midori-ku, Yokohama, 226-8501, Japan. E-mail: tmatsuda@bio.titech.ac.jp; Fax: +81-45-924-5757; Tel: +81-45-924-5757

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra03160g



piperazine ethanesulfonic acid (HEPES) and dithiothreitol (DTT), which were purchased from Sigma Aldrich (USA) and Wako (Japan), respectively. Protein concentration measurement reagent was supplied from Bio-Rad (USA). Commercial grade solvents, ketones, and alcohol standards (**1b–3b**) were purchased from Nacalai Tesque (Japan). Previously prepared alcohol (**4b–7b**) and chiral ester (**2b–4b** and **7b**) standards were used in this study.^{33–35}

2.2 Apparatus

Ultraviolet-visible spectroscopic analysis was conducted in a UV-1900-UV-Visible spectrophotometer from Shimadzu (Japan). Chiral gas chromatography (GC) was performed on a GC-14B equipped with flame ionization detector and a CP-Chirasil-Dex-CB column (Varian 0.32 mm × 0.25 μm × 50 m) using He carrier gas (5 mL min^{−1}, head pressure: 274 kPa, injector: 180 °C, detector: 180 °C) (Shimadzu, Japan). The morphology and energy dispersive X-ray spectroscopy (EDX) analysis were studied by Bench-top Scanning Electron Microscope proX supplied by Phenom-World (Netherlands). Thermo Gravimetric Analysis (TGA) was performed on TG-DTA, DTG-60 (Shimadzu, Japan). The optical rotation value was measured by JASCO P-2200 Polarimeter with a 10 cm path-length cell. The ¹H-NMR analysis was performed on a Bruker Biospin AVANCE III 400 spectrometer at 400 MHz in CDCl₃.

2.3 GcAPRD nanocrystal preparation

The phosphate-buffered saline (PBS) containing GcAPRD was mixed with a metal ion by pipetting for 5 times gently, incubated at 4 °C for 24 h, and centrifuged at 4 °C, 12 000 rpm for 5 min. The detailed experimental conditions are shown in the ESI (Table S1†). The supernatant was removed to measure the residual protein amount by Bradford method,³⁶ and to calculate the immobilization yield (1). The precipitant was washed by distilled water and resuspended in PBS (GcAPRD nanocrystal).

Immobilization yield (%)

$$= \left(\frac{[\text{Protein}]_i - [\text{Protein}]_R}{[\text{Protein}]_i} \right) \times 100 \quad (1)$$

[Protein]_i = initial protein concentration (mg mL^{−1}), [Protein]_R = concentration of residual protein in the supernatant after the GcAPRD nanocrystal formation and centrifugation (mg mL^{−1}).

2.4 Relative reaction yield and activity measurement of enzyme

The relative reaction yield measurement to investigate the optimum immobilization conditions was conducted by mixing acetophenone **1a** (5.0 mM), 2-propanol (15% v/v), free GcAPRD (8.30–16.50 μg of protein per mL) or GcAPRD nanocrystal (8.30–33.0 μg of protein per mL), NAD⁺ (0.20 mM), and HEPES–NaOH buffer (0.10 M, pH 7.2) up to 3 mL. For the investigation of the optimum immobilization conditions, the reaction was performed at 37 °C, 200 rpm for 10 min. The product formed (**1b**) was extracted with diethyl ether and analyzed by GC, using 3-methyl-1-butanol as an internal standard.

Activity measurement to investigate the temperature and pH profile, and the stability of GcAPRD nanocrystal were assayed by using UV-spectrophotometer. All assays were performed at 37 °C in 1.0 mL HEPES–NaOH buffer (0.10 M, pH 7.2), consisting of **1a** (5.0 mM) except for the pH profile experiment (5.6 mM), NADH (0.28 mM), and GcAPRD nanocrystal (0.80–4.00 μg of protein per mL). Initial velocity was determined by detecting the decrease of NADH at 340 nm for 132 s.

The recyclability was assessed by batch process. The reaction mixture consisted of **1a** (3.0 mM), 2-propanol (15% v/v), GcAPRD nanocrystal (630 μg of protein), NAD⁺ (0.20 mM), and HEPES–NaOH buffer (0.10 M, pH 7.2) up to 1.5 mL. The reaction in each cycle was conducted at 30 °C, 200 rpm for up to 180 min. Afterward, the reaction mixture was centrifuged at 4 °C, 16 000 rpm for 5 min. The supernatant was removed, extracted with diethyl ether, and analyzed by GC. The precipitant was washed by the reaction mixture with the absence of NAD⁺, and proceeded the next reaction cycle.

2.5 Micromole scale asymmetric reduction of ketones by GcAPRD nanocrystal

The experiment was conducted based on the previously reported procedure.¹³ Micromole scale reductions were performed in a 3.0 mL reaction mixture, which consists of **1a–7a** (5.0 mM), 2-propanol (15% v/v), NAD⁺ (5.0 mM), HEPES–NaOH buffer (0.10 M, pH 7.2), and GcAPRD nanocrystal with the amount needed to achieve 1.0 μmol min^{−1} mL^{−1} reduction of each substrate to its corresponding alcohol for **1a–6a**, and 350 μg of protein/mL for **7a**. The mixture was incubated at 30 °C, 200 rpm for 3 h. A portion of the mixture was extracted with diethyl ether to determine the reaction yield by GC analysis. The enantioselectivity excess (ee) values and the absolute configurations of **1b**, **5b**, and **6b** were determined by comparing their GC retention times with the corresponding authentic samples prepared before.³⁵ Meanwhile, for **2b–4b** and **7b**, the reaction mixtures were extracted by dichloromethane, dried over MgSO₄, and used for propionylation by propionyl chloride reaction. ee values and absolute configurations were determined by comparing the GC retention times of propionates of **2b–4b** and **7b** with the corresponding authentic samples prepared before.^{33,34} The chiral GC retention time of ketone, alcohol, and ester standards are given in Table S2.†

2.6 Preparative scale asymmetric reduction of a ketone by GcAPRD nanocrystal

The reaction was performed in a 100 mL scale consisting of **6a** (197 mg, 10.0 mM), 2-propanol (3.0% v/v), NAD⁺ (1.4 mM), HEPES–NaOH buffer (0.10 M, pH 7.2), and GcAPRD nanocrystal (2 mg of protein). The mixture was incubated at 30 °C, 130 rpm for 18 h, and extracted by diethyl ether for 3 times, dried over MgSO₄, filtered, and evaporated. The corresponding alcohol was purified by silica gel column chromatography (hexane : ethyl acetate, 3 : 1) to afford (*S*)-**6b** (165 mg, 0.87 mmol, 84%), and characterized by ¹H-NMR analysis. The ¹H-NMR spectra of alcohol (*S*)-**6b** was in agreement with that reported in the literature.³⁷ ee was determined by chiral GC analysis, and



the absolute configuration was determined by comparing the optical rotation value of product with the literature.³⁸ $[\alpha]_{\text{D}}^{20} = -38.19$ ($c = 1.04$, CHCl_3 , $ee = >99\%$); lit³⁸ (*S*)-**6b**: $[\alpha]_{\text{D}}^{20} = -33.70$ ($c = 1.44$, CHCl_3 , $ee = 91\%$). ¹H-NMR (400 MHz, CDCl_3 , 25 °C, TMS): $\delta = 7.48$ (d, $J = 2.0$ Hz, 1H, CH), 7.41 (d, $J = 8.3$ Hz, 1H, CH), 7.20 (dd, $J = 8.2$ Hz, 2.0 Hz, 1H, CH), 4.84–4.90 (m, 1H, CH), 1.82–1.83 (m, 1H, OH), 1.48 (d, $J = 6.48$ Hz, 3H, CH_3).

2.7 GcAPRD nanocrystal characterization by scanning electron microscopy (SEM) and EDX analysis, and TGA

The prepared GcAPRD nanocrystal was washed by distilled water several times to remove the residual phosphate and metal ion prior to drying at room temperature. The dried GcAPRD nanocrystal was used for SEM and EDX analysis, and TGA.

3. Results and discussions

3.1 Optimization of GcAPRD nanocrystal synthesis

Several parameters for the GcAPRD nanocrystal preparation, such as kinds of metal ions, metal concentration, PBS concentration, PBS pH, and protein concentration, were varied. The immobilization yields and relative reaction yields are shown in Fig. 1.

Different amino acid residues in each biomolecule present different affinities with various metal ions, and the appropriate metal ion concentration is necessary for the nucleation.²¹ In this research, 6 metal ions were used to prepare the GcAPRD nanocrystal (Fig. 1a). It was found that the immobilization yield was high when using Co^{2+} or Ni^{2+} as a metal ion. Nonetheless, only the GcAPRD nanocrystal synthesized by Co^{2+} retained sufficient activity. It was assumed that the His-tag located in the C-terminus of the GcAPRD acted as a binder with Co^{2+} or Ni^{2+} ion, which was comparable to the discovery of López-Gallego and Yate's.³¹ They reported that the presence of His-tagged protein is mandatory as the driving force for the crystal formation. The His-tag binds Co^{2+} ions *via* the coordination bond and forms a nucleation site, which initiates the crystal formation.³¹ Although Ni^{2+} could form the crystal with His-tagged protein, high concentration of nickel could cause the enzyme deactivation (Fig. S1†). To investigate the importance of Co^{2+} and His-tagged GcAPRD bonding, the formed GcAPRD nanocrystal was suspended in 500 mM imidazole in 5 mM PBS to break the Co^{2+} and His-tag bond, thus eluting the protein. After the elution, the GcAPRD nanocrystal was apparently detached, suggested that Co^{2+} and His-tag bonding played an important role in the GcAPRD nanocrystal formation (Fig. S2†). Additionally, the metal concentration was optimized to 10 mM (Fig. 1b), comparable with Kim *et al.*'s study (10 mM of cobalt for the formation of BSA-inorganic hybrid nanoflower).³⁹

PBS concentration is another important parameter for the nanocrystal formation.²⁸ Thereby, we investigated the effect of PBS concentration on the GcAPRD nanocrystal formation (Fig. 1c). The prepared GcAPRD nanocrystal performed the highest relative reaction yield at 3.75 mM PBS. However, the optimum PBS concentration is different depending on the biomolecule and other synthesis parameters.^{40,41} Synthesis pH

is an essential parameter influencing the nanocrystal formation.^{29,41,42} Thus, this study investigated the effect of PBS pH on the GcAPRD nanocrystal formation (Fig. 1d). The immobilization yield of GcAPRD was low in acidic conditions (pH 5.5), and higher in higher pH (pH 7.4, 9.2, and 11.0). It was proposed that in acidic conditions, the biomolecule is highly protonated, causing strong repulsions with the positive charges of the transition metal (*i.e.*, Cu^{2+} and Co^{2+}).⁴³ Joshua *et al.*, reported that protonating the imidazole nitrogen atom of the histidine residue (pKa 6.0) by lowering the pH to 6.0 can disrupt the coordination bond between histidine and Co^{2+} .⁴⁴ Altinkaynak *et al.* also reported an easier synthesis of lactoperoxidase hybrid nanoflower at pH 9.0 and 10.0.¹⁹ The GcAPRD nanocrystal showed the best relative reaction yield in pH 9.2, and pH 11.0.

The presence of biomolecule is mandatory for the hybrid nanocrystal formation.^{21,22} In this study, the protein concentration in the GcAPRD nanocrystal formation was varied to 0.02, 0.10, 0.05, 1.00, and 2.00 mg mL^{-1} (Fig. 1e). The experiments demonstrated that at the protein concentration of 0.05 mg mL^{-1} and below, the enzyme was entirely immobilized (immobilization yield >99%). On the other hand, the protein concentration of more than 1.00 mg mL^{-1} resulted in a decrease in immobilization yield, but protein immobilized per metal amount was high. Nevertheless, the relative reaction yield result showed that the reaction proceeded better when the synthesized protein concentrations were more than 0.50 mg mL^{-1} , and the greatest relative reaction yield was found on the 2.00 mg mL^{-1} of protein. Ge *et al.* reported that different concentration of protein has a different nanocrystal formation mechanism. The lower concentration of protein causes fewer nucleation sites, leading to a larger nanocrystal with a more complex structure.²² At last, it was confirmed that the active GcAPRD was mandatory to the GcAPRD nanocrystal activity (ESI, section 4†). To understand this phenomenon clearly, SEM analysis on GcAPRD nanocrystal synthesized with various protein concentrations were performed as described in section 3.2.

3.2 Morphology analysis by SEM

SEM analysis on GcAPRD nanocrystal synthesized with various protein concentrations was performed to understand its effect in the immobilization yield and relative reaction yield. As shown in Fig. 2, it was found that without enzyme, the cobalt-phosphate crystal was formed (Fig. 2a and b). In the presence of 0.02 mg mL^{-1} (Fig. 2c and d) and 0.10 mg mL^{-1} (Fig. 2e and f) of protein, the surfaces of the crystals were similar to the control, assuming that the small amount of protein was buried underneath the cobalt-phosphate crystals. Therefore, the mass transfer between the hindered enzyme and the substrate was limited, and low relative reaction yields were observed for the nanocrystals synthesized with 0.02 and 0.10 mg mL^{-1} of protein. The surfaces of GcAPRD nanocrystals synthesized with 0.50 mg mL^{-1} (Fig. 2g–h) and 2.00 mg mL^{-1} (Fig. 2i–j) protein were different from the control and displaying a parallel hexahedron structure assembled to a flower-like structure mixed with sponge-like structure. The proposed formation mechanism of the GcAPRD nanocrystal synthesized with 0.50 and



2.00 mg mL⁻¹ of protein is presented in Fig. 3. There may be 3 steps, comprised of nucleation, aggregation, and anisotropic growth. A similar structure was reported previously by Wang *et al.* in the formation of nanocrystals with the presence of 2.00 mg mL⁻¹ protein.^{27,31} The high relative reaction yield

suggested that the GcAPRD was gathered on the surface of the nanostructure, and the enzymes were not buried. In addition, the GcAPRD nanocrystal was characterized by EDX analysis and TGA to examine the chemical composition and structure of the GcAPRD nanocrystal as in Fig. S3–S5 (ESI,† section 5).

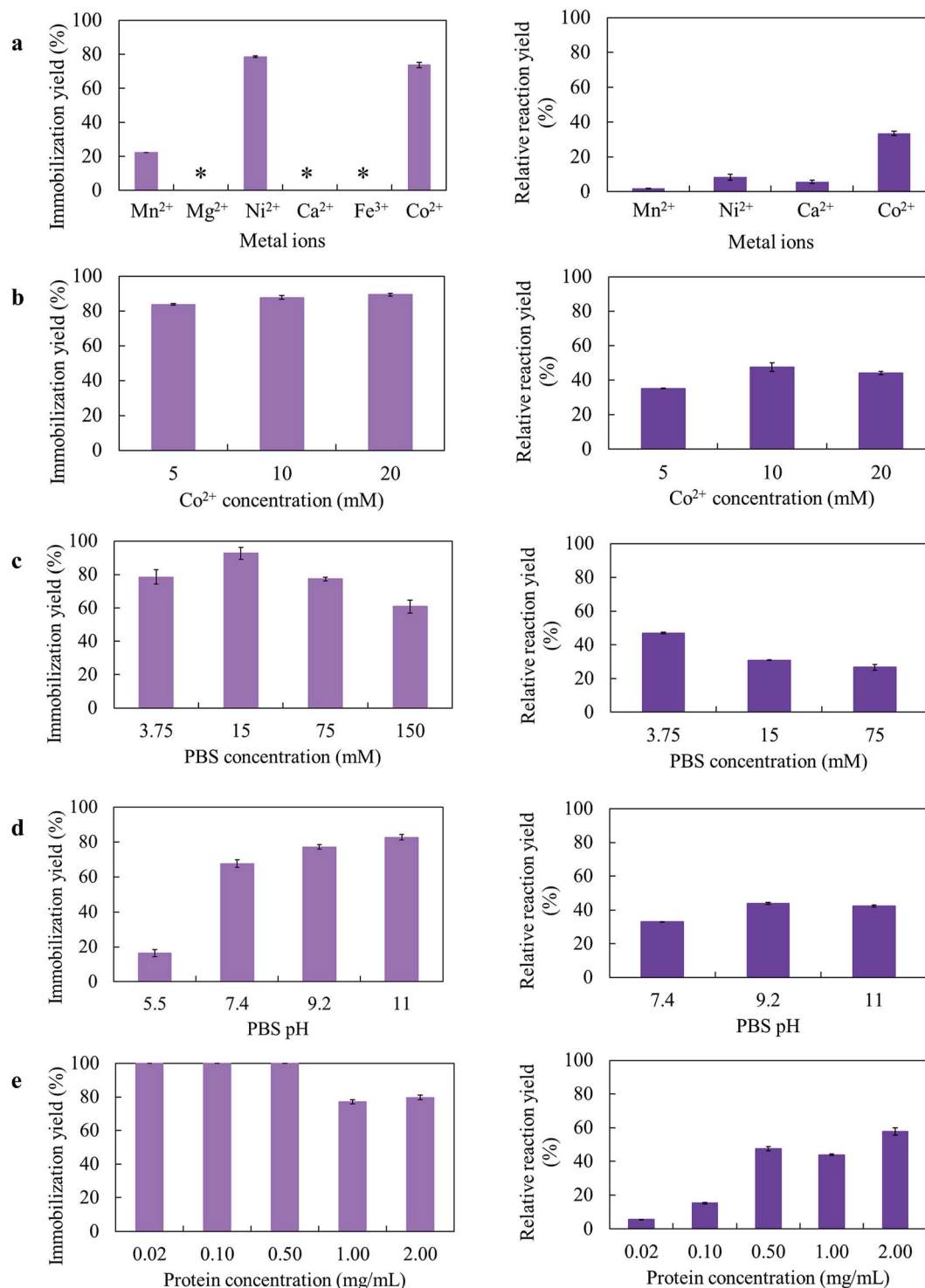


Fig. 1 Optimization of GcAPRD nanocrystal preparation (a) metal ions, (b) metal ion (Co²⁺) concentration, (c) PBS concentration, (d) PBS pH, and (e) protein concentration. Left and right column depict the immobilization yields and relative reaction yields, respectively. The reaction yield of free GcAPRD was set to be 100%. *Very small precipitation was found. The preparation and reaction conditions are described in the section 2.4.



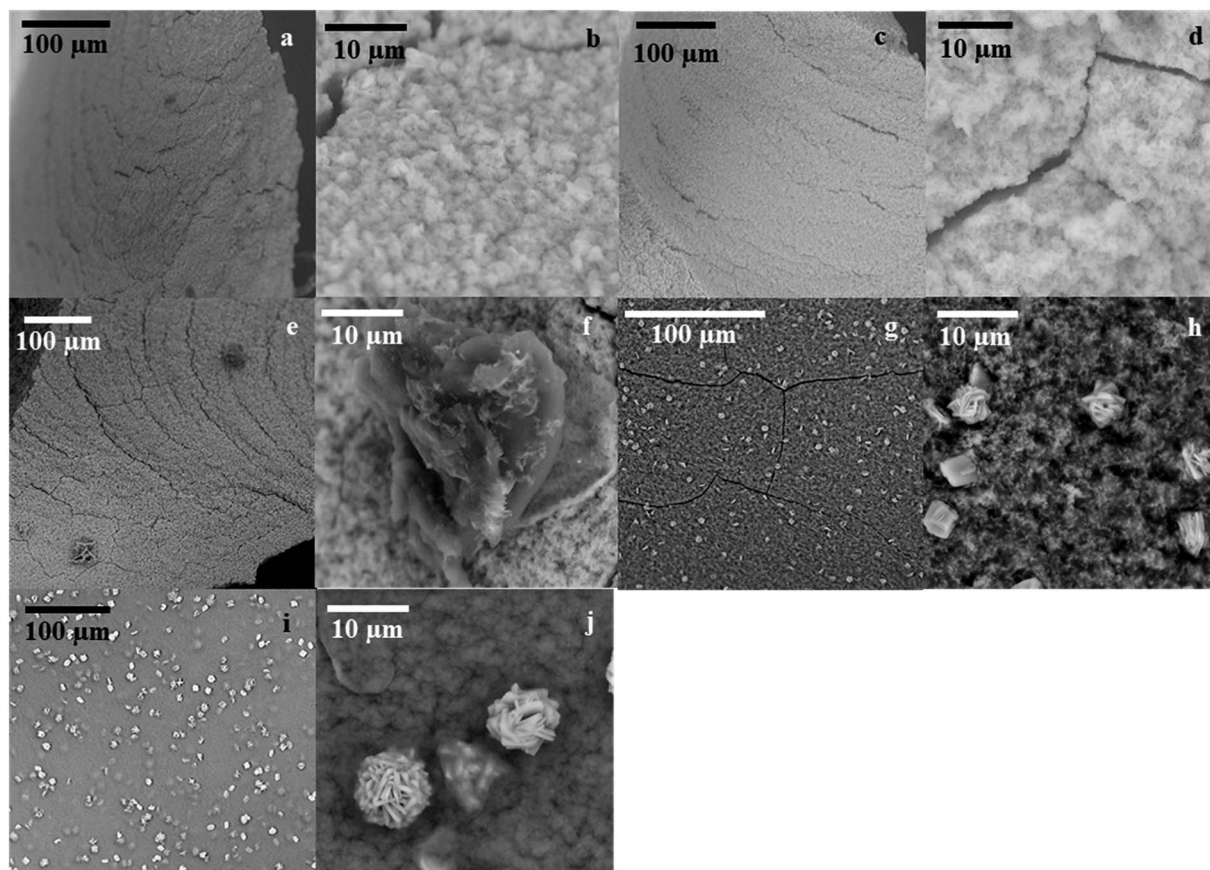


Fig. 2 SEM images of GcAPRD nanocrystal formed under various protein concentrations, (a and b) control (no enzyme), (c and d) 0.02 mg mL^{-1} , (e and f) 0.10 mg mL^{-1} , (g and h) 0.50 mg mL^{-1} , and (i and j) 2.00 mg mL^{-1} .

3.3 Temperature and pH profile, and stability of GcAPRD nanocrystal

Immobilization has been reported to improve the temperature and pH profile, as well as the stability of the enzyme.^{2,31,40,41} The temperature profile of GcAPRD nanocrystal and free GcAPRD was examined (Fig. 4a). Both free GcAPRD and GcAPRD nanocrystal demonstrated poor activity at low temperatures (20–30 °C). The activities of both enzymes were increased in higher temperatures. The free GcAPRD performed the highest relative

activity at 50 °C (170%), and the activity gradually decreased at the temperature above 60 °C. To our delight, the GcAPRD nanocrystal presented the improved temperature profile as the highest relative activity was observed at 60 °C (189%). At the same time, the effect of pH on the GcAPRD nanocrystal activity was investigated, and it showed less sensitivity than the free GcAPRD in acidic conditions (Fig. 4b). The relative activity of the free GcAPRD dramatically decreased from 99% to 18% when the pH was lowered from 5.0 to 4.5, while the GcAPRD nanocrystal's

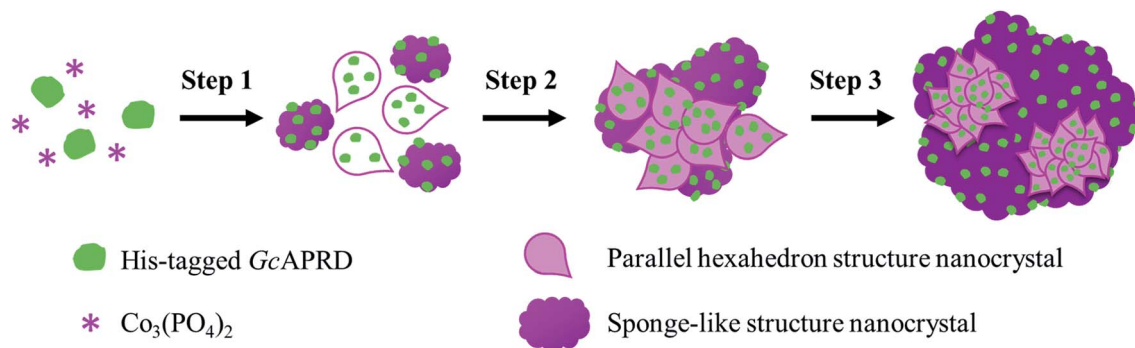


Fig. 3 The proposed formation mechanism of GcAPRD nanocrystal, comprised of 3 steps: (1) nucleation, (2) aggregation, and (3) anisotropic growth.



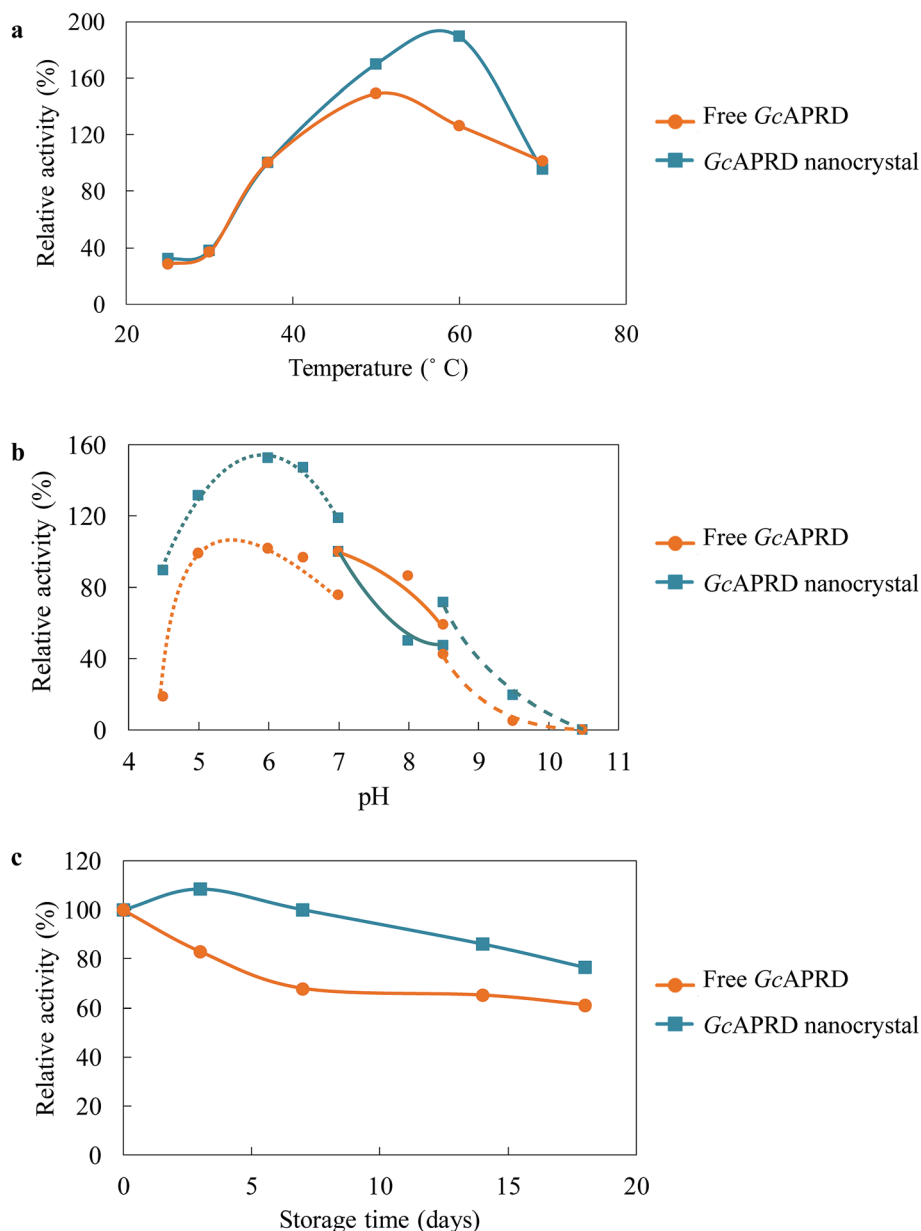


Fig. 4 Temperature and pH profile, and stability of free GcAPRD (orange-circle line) and GcAPRD nanocrystal (blue-square line). (a) temperature profile, (b) pH profile, and (c) storage stability. Enzyme activities of free GcAPRD and GcAPRD nanocrystal at 37 °C for (a), HEPES–NaOH buffer (pH 7.0) for (b), and day 0 for (c) were set to be 100%. For the pH profile, the enzyme activity was measured in MES–NaOH buffer (0.10 M) (dotted line), HEPES–NaOH buffer (0.10 M) (line), and glycine–NaOH buffer (0.10 M) (broken line). The activity assay is described in the section 2.4.

relative activity moderately decreased from 132% to 89%. The storage stability of the enzymes at 4 °C was investigated. As shown in Fig. 4c, after 7 days, the residual activity of the free GcAPRD was 68%, while the GcAPRD nanocrystal still retained 100% activity. Also, after 18 days, the free GcAPRD's residual activity was 61%, while the GcAPRD nanocrystal had 76% of residual activity.

3.4 Recyclability of GcAPRD nanocrystal

We investigated the recyclability of the GcAPRD nanocrystal by performing the reduction of **1a** in batch process. The GcAPRD

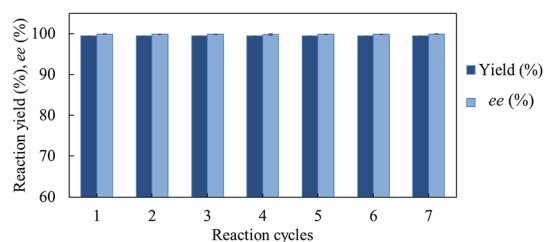


Fig. 5 Recyclability of GcAPRD nanocrystal. The reaction condition is described in the section 2.4.



nanocrystal was able to be recycled with noticeable high reduction yield (>99%) and excellent enantioselectivity (>99%) up to 7 reaction cycles (Fig. 5). This result proved the superior recyclable properties of the *GcAPRD* nanocrystal, which is a promising approach to further applications.

3.5 Substrate specificity of *GcAPRD* nanocrystal

Excellent enantioselectivity and broad substrate specificity are one of the most crucial aspects of the enzyme utilization for industrial processes.¹⁰ The *GcAPRD* nanocrystal was tested to reduce broad kinds of ketones (Table 1). **1a**, as a model substrate for the *GcAPRD* reduction, was reduced by *GcAPRD* nanocrystal with >99% yield and >99% ee (*S*). The *GcAPRD* nanocrystal also reduced challenging aliphatic ketones **2a–4a** with high yields and enantioselectivities (up to >99% ee (*S*)). Moreover, the *GcAPRD* nanocrystal was able to enantioselectively reduce “difficult to resolve” ketone **3a** with only one

carbon difference between side chains adjacent to the carbonyl carbon. The enantioselectivities reported for the free enzyme were retained perfectly.^{9,12,13}

The *GcAPRD* nanocrystal produced beneficial chiral alcohols with excellent enantioselectivities (ee up to >99% (*S*)) such as (*S*)-**5b** (intermediate of EGFR and HER2 kinase inhibitors, a potential cancer treatment agent),⁴⁵ (*S*)-**6b** (intermediate of sertraline, an obsessive-compulsive disorder medication),⁴⁶ and (*S*)-**7b** (intermediate of aminotetralin, which can be used to stimulate dopamine, serotonin, and melatonin receptors).^{47,48} We were also able to isolate (*S*)-**6b** with 84% isolated yield, in a preparative scale reaction catalyzed by the *GcAPRD* nanocrystal. To the best of our knowledge, this is the first study to report an ADH nanocrystal with broad substrate specificity.

4. Conclusions

We successfully immobilized a novel ADH, *GcAPRD*, by the organic–inorganic nanocrystal method. The *GcAPRD* nanocrystal presented improved temperature and pH profile, and stability compared with those of the free *GcAPRD*. The *GcAPRD* nanocrystal overcame the recyclability limitation of the free *GcAPRD*. It could be recycled up to 7 cycles with remarkably high yield and excellent enantioselectivity. The *GcAPRD* nanocrystal reduced broad kinds of ketones to produce valuable chiral alcohols. The robust and versatile properties of the *GcAPRD* nanocrystal made it a promising and sustainable catalyst for industrial applications. Future studies will investigate the reaction of *GcAPRD* nanocrystal in non-aqueous solvent and perform cascade reactions to allow the production of broader kinds of beneficial compounds.

Conflicts of interest

There are no conflicts to declare.


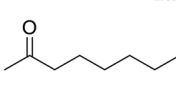
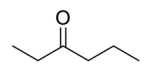
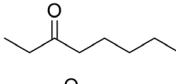
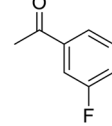
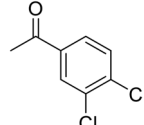
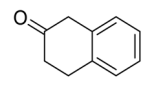
Acknowledgements

The authors are grateful to the Open Research Facilities for Life Science and Technology, School of Life Science and Technology, Tokyo Institute of Technology, for the support of SEM and EDX analysis, and Open Facility Center, Materials Analysis Division, Tokyo Institute of Technology, for the support of TGA. This work was partially supported by the Japan Society for the Promotion of Science, Japan (grant number JP16K05864 and JP19K05560) to Tomoko Matsuda.

Notes and references

- 1 K. Nakamura, R. Yamanaka, T. Matsuda and T. Harada, *Tetrahedron: Asymmetry*, 2003, **14**, 2659–2681.
- 2 A. Madhavan, R. Sindhu, P. Binod, R. K. Sukumaran and A. Pandey, *Bioresour. Technol.*, 2017, **245**, 1304–1313.
- 3 W. Stampfer, B. Kosjek, K. Faber and W. Kroutil, *J. Org. Chem.*, 2003, **68**, 402–406.
- 4 M. Giersberg, A. Degelmann, R. Bode, M. Piontek and G. Kunze, *J. Ind. Microbiol. Biotechnol.*, 2012, **39**, 1385–1396.

Table 1 Reduction of ketones by *GcAPRD* nanocrystal^a

Substrate	Yield (%)	ee (%)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	>99	>99 (<i>S</i>)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	>99	>99 (<i>S</i>)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	74	99 (<i>S</i>)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	77	>99 (<i>S</i>)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	>99	>99 (<i>S</i>)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	>99	>99 (<i>S</i>)
 R_1 has lower Cahn-Ingold-Prelog priority than R_2	91	96 (<i>S</i>)

^a The reaction conditions are described in the section 2.5.



- 5 J.-M. Choi, S.-S. Han and H.-S. Kim, *Biotechnol. Adv.*, 2015, **33**, 1443–1454.
- 6 R. N. Patel, *Bioorg. Med. Chem.*, 2018, **26**, 1252–1274.
- 7 D. Ballesteros-vivas, G. Alvarez-rivera, M. Bueno and A. Cifuentes, *TrAC, Trends Anal. Chem.*, 2020, **123**, 115761.
- 8 N. Itoh, *Appl. Microbiol. Biotechnol.*, 2014, **98**, 3889–3904.
- 9 A. A. Koesoema, D. M. Standley, T. Senda and T. Matsuda, *Appl. Microbiol. Biotechnol.*, 2020, **104**, 2897–2909.
- 10 H. Gröger, W. Hummel and R. Metzner, in *Comprehensive Chirality*, 2012.
- 11 T. Matsuda, S. Kanamaru, Y. Nakata, R. Kanamori and T. Fukae, *Appl. Microbiol. Biotechnol.*, 2009, **86**, 625–631.
- 12 T. Yamamoto, Y. Nakata, C. Cao, Y. Sugiyama, Y. Asanuma, S. Kanamaru and T. Matsuda, *Appl. Microbiol. Biotechnol.*, 2013, **97**, 10413–10421.
- 13 A. A. Koesoema, Y. Sugiyama, Z. Xu, D. M. Standley, M. Senda, T. Senda and T. Matsuda, *Appl. Microbiol. Biotechnol.*, 2019, **103**, 9543–9553.
- 14 K. Nakamura, Y. Inoue, T. Matsuda and I. Misawa, *J. Chem. Soc., Perkin Trans. 1*, 1999, 2397–2402.
- 15 T. Matsuda, T. Harada and K. Nakamura, *Chem. Commun.*, 2000, 1367–1368.
- 16 T. Matsuda, K. Watanabe, T. Kamitanaka, T. Harada and K. Nakamura, *Chem. Commun.*, 2003, **3**, 1198–1199.
- 17 T. Matsuda, R. Marukado, M. Mukouyama, T. Harada and K. Nakamura, *Tetrahedron: Asymmetry*, 2008, **19**, 2272–2275.
- 18 T. Matsuda, Y. Yamagishi, S. Koguchi, N. Iwai and T. Kitazume, *Tetrahedron Lett.*, 2006, **47**, 4619–4622.
- 19 C. Altinkaynak, S. Tavlasioglu, N. Özdemir and I. Ocsoy, *Enzyme Microb. Technol.*, 2016, **93–94**, 105–112.
- 20 N. R. Mohamad, N. H. C. Marzuki, N. A. Buang, F. Huyop and R. A. Wahab, *Biotechnol. Biotechnol. Equip.*, 2015, **29**, 205–220.
- 21 J. Cui and S. Jia, *Coord. Chem. Rev.*, 2017, **352**, 249–263.
- 22 J. Ge, J. Lei and R. N. Zare, *Nat. Nanotechnol.*, 2012, **7**, 428–432.
- 23 C. Altinkaynak, S. Tavlasioglu, N. Özdemir and I. Ocsoy, *Enzyme Microb. Technol.*, 2016, **93–94**, 105–112.
- 24 S. Escobar, S. Velasco-Lozano, C. H. Lu, Y. F. Lin, M. Mesa, C. Bernal and F. López-Gallego, *J. Mater. Chem. B*, 2017, **5**, 4478–4486.
- 25 Y. Zhang, W. Sun, N. M. Elfeky, Y. Wang, D. Zhao, H. Zhou, J. Wang and Y. Bao, *Enzyme Microb. Technol.*, 2020, **132**, 109408.
- 26 S. S. Nadar and V. K. Rathod, *Enzyme Microb. Technol.*, 2018, **108**, 11–20.
- 27 L. B. Wang, Y. C. Wang, R. He, A. Zhuang, X. Wang, J. Zeng and J. G. Hou, *J. Am. Chem. Soc.*, 2013, **135**, 1272–1275.
- 28 J. Yu, X. Chen, M. Jiang, A. Wang, L. Yang, X. Pei, P. Zhang and S. G. Wu, *RSC Adv.*, 2018, **8**, 2357–2364.
- 29 Y. Li, X. Fei, L. Liang, J. Tian, L. Xu, X. Wang and Y. Wang, *J. Mol. Catal. B: Enzym.*, 2016, **133**, 92–97.
- 30 F. Zhao, Q. Wang, J. Dong, M. Xian, J. Yu, H. Yin, Z. Chang, X. Mu, T. Hou and J. Wang, *Process Biochem.*, 2017, **57**, 87–94.
- 31 F. López-Gallego and L. Yate, *Chem. Commun.*, 2015, **51**, 8753–8756.
- 32 X. Chen, L. Xu, A. Wang, H. Li, C. Wang, X. Pei, P. Zhang and S. G. Wu, *J. Chem. Technol. Biotechnol.*, 2019, **94**, 236–243.
- 33 A. A. Koesoema, Y. Sugiyama, Z. Xu, D. M. Standley, M. Senda, T. Senda and T. Matsuda, *Appl. Microbiol. Biotechnol.*, 2019, **103**, 9543–9553.
- 34 A. A. Koesoema, D. M. Standley, T. Kotchakorn, M. Tamura and T. Matsuda, *Tetrahedron Lett.*, 2020, **61**, 151682.
- 35 A. A. Koesoema, D. M. Standley, S. Ohshima, M. Tamura and T. Matsuda, *Tetrahedron Lett.*, 2020, 151820.
- 36 M. M. Bradford, *Anal. Biochem.*, 1976, **72**, 248–254.
- 37 R. Wang, J. Qi, Y. Yue, Z. Lian, H. Xiao, S. Zhuo and L. Xing, *Tetrahedron*, 2019, **75**, 130463.
- 38 Y. Li, S. Yu, X. Wu, J. Xiao, W. Shen, Z. Dong and J. Gao, *J. Am. Chem. Soc.*, 2014, **136**, 4031–4039.
- 39 K. H. Kim, J. M. Jeong, S. J. Lee, B. G. Choi and K. G. Lee, *J. Colloid Interface Sci.*, 2016, **484**, 44–50.
- 40 S. S. Nadar, S. D. Gawas and V. K. Rathod, *Int. J. Biol. Macromol.*, 2016, **92**, 660–669.
- 41 B. Somturk, M. Hancer, I. Ocsoy and N. Özdemir, *Dalton Trans.*, 2015, **44**, 13845–13852.
- 42 B. Somturk, I. Yilmaz, C. Altinkaynak, A. Karatepe, N. Özdemir and I. Ocsoy, *Enzyme Microb. Technol.*, 2016, **86**, 134–142.
- 43 C. Altinkaynak, I. Yilmaz, Z. Koksall, H. Özdemir, I. Ocsoy and N. Özdemir, *Int. J. Biol. Macromol.*, 2016, **84**, 402–409.
- 44 J. A. Bornhorst and J. J. Falke, *Methods Enzymol.*, 2000, **326**, 245–254.
- 45 H. Mastalerz, M. Chang, P. Chen, P. Dextraze, B. E. Fink, A. Gavai, B. Goyal, W. C. Han, W. Johnson, D. Langley, F. Y. Lee, P. Marathe, A. Mathur, S. Oppenheimer, E. Ruediger, J. Tarrant, J. S. Tokarski, G. D. Vite, D. M. Vyas, H. Wong, T. W. Wong, H. Zhang and G. Zhang, *Bioorg. Med. Chem. Lett.*, 2007, **17**, 2036–2042.
- 46 C. Barbieri, E. Caruso, P. D'Arrigo, G. Pedrocchi Fantoni and S. Servi, *Tetrahedron: Asymmetry*, 1999, **10**, 3931–3937.
- 47 P. Bonomi, P. Cairoli, D. Ubiali, C. F. Morelli, M. Filice, I. Nieto, M. Pregnotato, P. Manitto, M. Terreni and G. Speranza, *Tetrahedron: Asymmetry*, 2009, **20**, 467–472.
- 48 F. Orsini, G. Sello, E. Travaini and P. Di Gennaro, *Tetrahedron: Asymmetry*, 2002, **13**, 253–259.

