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# A sensitive OFF–ON–OFF fluorescent probe for the cascade sensing of Al<sup>3+</sup> and F<sup>−</sup> ions in aqueous media and living cells†

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A simple Schiff-base ligand 2-hydroxy-1-naphthaldehyde semicarbazone (HNS) was synthesized and characterized. Based on the combined effect of inhibition of CH=N isomerization and chelation-enhanced fluorescence (CHEF), HNS functions as a fluorescence “turn on” sensor for Al<sup>3+</sup> in buffered aqueous media. Based on the strong affinity of Al<sup>3+</sup> to F<sup>−</sup> ions, the *in situ* generated Al<sup>3+</sup>–HNS complex can also be utilized as an effective chemosensor for F<sup>−</sup> sensing by metal displacement approach, ensuing quenching of fluorescence by the reversible return of HNS from Al<sup>3+</sup>–HNS complex. Thus a method using a single probe for the detection of both Al<sup>3+</sup> and F<sup>−</sup> ions is developed. The system exhibits high selectivity and sensitivity for Al<sup>3+</sup> and F<sup>−</sup> ions and the detection limits were found to be as low as 6.75 × 10<sup>−8</sup> M and 7.89 × 10<sup>−7</sup> M, respectively. Furthermore, the practical applicability of this probe has been examined in living cells.

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## 1. Introduction

Aluminum is the most abundant metal in the Earth's crust and is extensively used in every sphere of life from utensils to medicine. But aluminum in excessive amounts not only hampers plant growth but damages the human nervous system and has been reported to induce Alzheimer's disease, Parkinson's disease and amyotrophic lateral sclerosis, *etc.*<sup>1</sup> Therefore, detection of Al<sup>3+</sup> ions in natural environment and living organisms is of vital importance for human health. In this context, the development of fluorescence based probes attracted more attention because of the simplicity and high sensitivity of fluorescence assays. However, the lack of spectroscopic characteristics and poor coordination ability of Al<sup>3+</sup> (ref. 2) make it very challenging to develop a highly selective sensing system for Al<sup>3+</sup>.

Up to now, several fluorescent probes for Al<sup>3+</sup> have been designed by various workers.<sup>3–19</sup> And the structures of most

fluorescent probes for Al<sup>3+</sup> contain nitrogen–oxygen-rich coordination environments which provide a hard-base environment for the hard-acid Al<sup>3+</sup>. These probes, however, have some limitations such as poor water solubility and tedious synthetic procedure, which limited their practical applications. Thus, only few examples of Al<sup>3+</sup> probes can be used for cell imaging and biological samples.<sup>5,8,9,12–15</sup> Overall, it is still highly desirable to develop novel fluorescent probes for Al<sup>3+</sup> which satisfies several parameters *viz.* easy and economical synthetic procedure, moderate water solubility and being suitable for cell imaging studies.

Moderate fluoride for human has good performance in the treatment of osteoporosis and the prevention of dental caries. However, excess presence of fluoride for human body may lead to dental fluorosis, skeletal fluorosis, osteoporosis, and osteosclerosis.<sup>20</sup> Thus, the development of an efficient F<sup>−</sup> fluorescent sensor is quite important.<sup>21–23</sup> Interestingly, emissive probe + Al<sup>3+</sup> complexes can serve as anion sensors, especially to F<sup>−</sup>.<sup>24–29</sup> And these sensors that are capable of detecting Al<sup>3+</sup> and F<sup>−</sup> sequentially by fluorescence OFF–ON–OFF signaling response are highly effective when compared with the one-to-one probes.<sup>30–32</sup>

With these considerations in mind, we herein report an easily synthesizable Schiff base receptor 2-hydroxy-1-naphthaldehyde semicarbazone (HNS), which commendably supports the idea of the sequential sensing property for Al<sup>3+</sup> and F<sup>−</sup> in aqueous media and live cells. 2-Hydroxynaphthalene was selected as a signaling moiety for its low fluorescence quantum yield,<sup>33</sup> short fluorescence lifetime,<sup>34</sup> good binding sites and low cost. The binding of Al<sup>3+</sup> with the phenolic –OH group and Schiff base nitrogen atoms of HNS would inhibit the *cis–trans*

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 † Electronic supplementary information (ESI) available: <sup>1</sup>H NMR spectrum, <sup>13</sup>C NMR spectrum and HRMS spectrum of HNS. Job's plot for HNS and Al<sup>3+</sup> complexation in EtOH/H<sub>2</sub>O. Benesi–Hildebrand plot from fluorescence titration data of HNS with Al<sup>3+</sup>. FTIR spectra of HNS and Al<sup>3+</sup>–HNS complex. <sup>1</sup>H NMR spectrum, <sup>13</sup>C NMR spectrum, and HRMS spectrum of Al<sup>3+</sup>–HNS complex after adding F<sup>−</sup>. See DOI: 10.1039/d0ra02848g


isomerization around C=N and increase the rigidity of the molecular assembly to produce significant fluorescence enhancement, which shows a selective sensing property for Al<sup>3+</sup> in buffered aqueous media. Further, the formed Al<sup>3+</sup>-HNS complex can be utilized for the sensitive detecting of fluoride owing to the strong binding affinity of Al<sup>3+</sup> for F<sup>-</sup> ions.<sup>35</sup> Hence, the gradual addition of F<sup>-</sup> ions into a mixture of HNS and Al<sup>3+</sup> ions releases HNS, and shows fluorescence quenching. Thus, a single-probe sensory system was developed to detect both Al<sup>3+</sup> cation and F<sup>-</sup> anion with good detection reversibility, which would greatly decrease the cost for detection. What's more, detection of intracellular Al<sup>3+</sup> and F<sup>-</sup> in live SiHa cells by fluorescence imaging has been demonstrated in this study.

## 2. Experimental

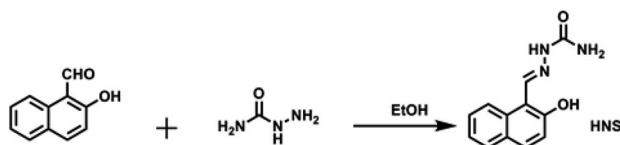
### 2.1 Chemicals and instrumentation

2-Hydroxy-1-naphthaldehyde (98%) was purchased from Energy Chemical (Shanghai, China). Aminourea hydrochloride was purchased from Aladdin Reagent Co. (Shanghai, China). All the other chemical reagents were of analytical grade and used as received without further purification. Stock solutions of metal ions were prepared by dissolving their nitrates in water and stock solutions of anions were prepared from the corresponding sodium salts. Hexamethylenetetramine-HCl buffer solution of pH 5.3 was prepared by mixing appropriate ratios of 0.01 mol L<sup>-1</sup> hexamethylenetetramine and 0.1 mol L<sup>-1</sup> HCl solutions. Double distilled water was used throughout. All the solutions have been stocked in the plastic centrifugal tubes.

The UV absorption spectra were recorded on a Puxi TU-1901 UV-vis absorption spectrophotometer (China). Fluorescence measurements were performed on a LS-55 spectrofluorimeter (PerkinElmer, USA). The samples were excited at 390 nm. The excitation and emission slits were set at 6 and 4 nm, respectively. IR spectra were taken as KBr pellets on a TENSOR II infrared spectrometer (Bruker, Germany). <sup>1</sup>H-NMR spectra were recorded on a DRX-300 spectrometer (Fällenden, Switzerland). The pH measurements were carried out on a pH-3C acidometer (Shanghai Precision & Scientific Instrument Co., Ltd, China). High resolution mass spectra (HRMS) were obtained on a Bruker micrOTOF-Q III mass spectrometer. The fluorescence intracellular images were obtained using an Olympus FV1000 confocal microscope (Tokyo, Japan) with a 40× objective (excited at 405 nm).

### 2.2 Synthesis of 2-hydroxy-1-naphthaldehyde semicarbazone (HNS)

Scheme 1 shows the facile one step synthesis of the ligand HNS. Aminourea hydrochloride (259 mg, 2.32 mmol) was dissolved in



Scheme 1 The synthesis of probe HNS.

8 mL of ethanol. The solution was neutralized to pH 7.0 with 1.0 M of NaOH and then was mixed with the ethanol solution of 2-hydroxy-1-naphthaldehyde (200 mg, 1.16 mmol). The mixture was heated under reflux for 4 h and then cooled to room temperature. Saturated sodium chloride solution was added dropwise and the precipitated solid was filtered off, washed with cold water and dried under vacuum and recrystallized from ethanol to give the product as a light-yellow solid (170 mg, 64% yield). IR  $\nu_{\max}$  (KBr): 3450, 3350, 1670, 1602, 1431, 1189 cm<sup>-1</sup>. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>)  $\delta$ : 6.37 (s, 2H, NH<sub>2</sub>), 7.20 (d, 1H, ArH), 7.34 (t, 1H, ArH), 7.52 (t, 1H, ArH), 7.85 (t, 2H, ArH), 8.38 (d, 1H, ArH), 8.86 (s, 1H, NH), 10.22 (s, 1H, CH=N), 11.24 (s, 1H, OH). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>): 161.51, 161.31, 145.24, 136.86, 136.74, 134.12, 133.38, 132.95, 128.72, 127.51, 123.87, 115.25. HRMS (ESI): calcd for C<sub>12</sub>H<sub>12</sub>N<sub>3</sub>O<sub>2</sub> (HNS + H) 230.0930, found 230.0927.

### 2.3 Cell imaging study

*In vitro* experiments were performed using SiHa cells. SiHa cells were cultured in DMEM medium, which was supplemented with 10% fetal bovine serum (FBS) in an atmosphere of 5% CO<sub>2</sub> at 37 °C. Immediately before the experiments, the cells were incubated with 50  $\mu$ L HNS (10<sup>-3</sup> M, ethanol solution) in 1 mL 0.1 M sterile PBS buffer for 30 min at room temperature. After incubation, the cells were washed with PBS buffer and incubated with Al<sup>3+</sup> (100  $\mu$ M) for additional 10 min at room temperature. Finally, the cells treated with HNS and Al<sup>3+</sup> were further incubated with NaF (500  $\mu$ M) for another 10 min. The fluorescence intracellular images were obtained using an Olympus FV1000 confocal microscope (Tokyo, Japan) with a 40× objective (excited at 405 nm).

## 3. Results and discussion

### 3.1 Spectral characteristics of HNS and HNS-Al<sup>3+</sup>

HNS could be dissolved in water when 10% (v/v) of ethanol was added. So, the spectroscopic properties of HNS were investigated in an ethanol-water (1 : 9, v/v) solution at pH 5.3. As shown in Fig. 1a, the absorption spectrum of HNS exhibits structured absorption spectrum corresponding to the  $\pi$ - $\pi^*$  transitions of naphthalene unit observed in naphthalene derivatives.<sup>36</sup> Upon addition of increasing amounts of Al<sup>3+</sup>, the longest-wavelength absorption band red shifted from 348 nm to 381 nm. The presence of two clear isosbestic points (330 and 363 nm) indicates the formation of a complex between HNS and Al<sup>3+</sup>.

Fig. 1b shows the changes in the emission intensity of HNS upon gradual increase of [Al<sup>3+</sup>]. HNS shows very weak broad emission in the range from 400 nm to 550 nm with a maximum at 449 nm on excitation at 390 nm. Upon addition of Al<sup>3+</sup>, HNS shows large fluorescence enhancement, accompanied with a blue-shift of emission maxima from 449 nm to 441 nm. The results suggest that Al<sup>3+</sup> most likely forms stable fluorescent complex with HNS. Schiff-bases usually have low emission due to the isomerization reaction around the imine linkage C=N.<sup>37</sup> This isomerization, however, is inhibited by the binding of Al<sup>3+</sup> ions leading to fluorescence switching of the receptor. In



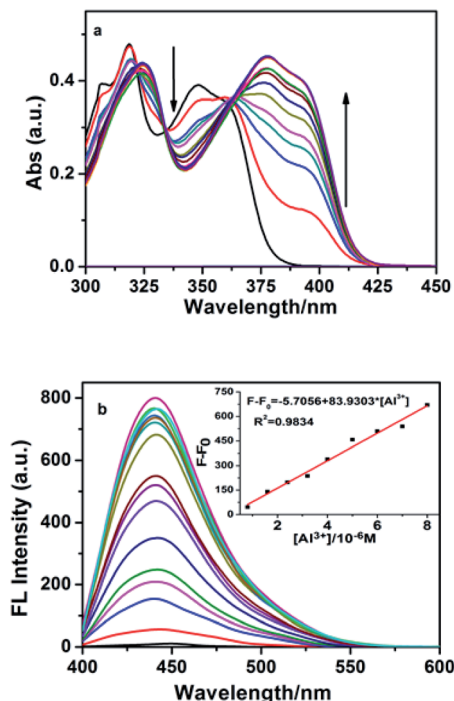


Fig. 1 (a) UV/vis spectra of HNS (40  $\mu\text{M}$ ) in the presence of different concentrations of  $\text{Al}^{3+}$  (0–10 equiv.) and (b) fluorescence titration spectra of HNS (4  $\mu\text{M}$ ) upon the addition of  $\text{Al}^{3+}$  ions (0–8 equiv.) in EtOH/ $\text{H}_2\text{O}$  (1 : 9, v/v, pH 5.3). The inset in (b) shows the fluorescence intensity of HNS as a function of  $\text{Al}^{3+}$  ion concentration) indicates the formation of a complex between HNS and  $\text{Al}^{3+}$ .

addition, the chelation of HNS with  $\text{Al}^{3+}$  increases the rigidity of the molecular assembly, resulting in enhancement of emission intensity ascribed as CHEF.

### 3.2 Fluorescence sensing of $\text{Al}^{3+}$

Optimization of pH on the efficiency of the sensor was essential. Fluorescence pH titrations were carried out for this purpose. Fig. 2 clearly demonstrates that at the pH range from 4.2 to 5.4, the HNS- $\text{Al}^{3+}$  system showed maximum emission intensity but decrease outside this range. Most plausibly, breakage of imine linkage of HNS at too low pH prevents coordination of  $\text{Al}^{3+}$  while at high pH values  $\text{OH}^-$  may succeed against HNS in binding to

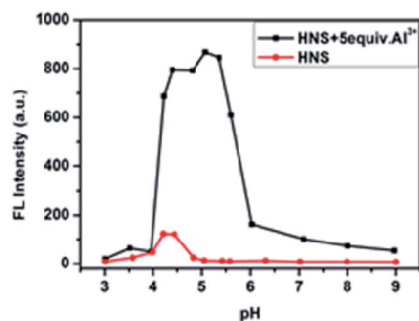


Fig. 2 Effect of pH on fluorescence intensity of the  $\text{Al}^{3+}$ -HNS complex in EtOH/ $\text{H}_2\text{O}$  (1 : 9, v/v),  $[\text{HNS}] = 4 \mu\text{M}$ ,  $[\text{Al}^{3+}] = 20 \mu\text{M}$ .

$\text{Al}^{3+}$ . In order to obtain a higher signal-to-noise, pH = 5.3 with hexamethylenetetramine-hydrochloric acid buffer solution was chosen throughout the experiment.

Selectivity of HNS as the fluorescence chemosensor for  $\text{Al}^{3+}$  was studied in the presence of various competing metal ions. Under the same conditions as used above for  $\text{Al}^{3+}$ , we tested the fluorescence responses of HNS toward other metal ions such as  $\text{Pb}^{2+}$ ,  $\text{Na}^+$ ,  $\text{Cd}^{2+}$ ,  $\text{K}^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Co}^{2+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Ni}^{2+}$ ,  $\text{Hg}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Fe}^{3+}$  and  $\text{In}^{3+}$ . As shown in Fig. 3a, significant spectral changes were observed only in the case of  $\text{Al}^{3+}$ . In contrast, treatment with other metal ions resulted in no significant changes, demonstrating that HNS is highly selective for  $\text{Al}^{3+}$  over competing metal ions. Furthermore, it was observed that the emission intensity of the  $\text{Al}^{3+}$ -HNS complex was not remarkably affected by most coexistent metal ions except  $\text{Cu}^{2+}$  and  $\text{Fe}^{3+}$  (Fig. 3b).  $\text{Cu}^{2+}$  showed complete quenching in fact, where luminescence was even less than that of HNS alone. A similar phenomenon was previously reported by Jang *et al.*<sup>38</sup> Thus HNS can be applied as an effective fluorescent sensor for  $\text{Al}^{3+}$  in the presence of most competing metal ions.

Based on the fluorometric spectra studies, a calibration curve was constructed by plotting the values of  $F - F_0$  against  $\text{Al}^{3+}$  concentration (inset of Fig. 1b). A good linear relationship between  $F - F_0$  and the concentration of  $\text{Al}^{3+}$  could be obtained in the range of 0.8–8  $\mu\text{M}$  ( $R^2 = 0.9834$ ). The limit of detection was measured to be  $6.75 \times 10^{-8} \text{ M}$  (signal to noise ratio of 3 : 1), which is satisfactory to the  $\text{Al}^{3+}$  detection in drinking water within World Health Organization (WHO) limit ( $200 \mu\text{g L}^{-1}$ ,  $7.41 \mu\text{M}$ ).<sup>39</sup> In terms of sensitivity, the detection limit of the proposed method is better than or comparable to those of

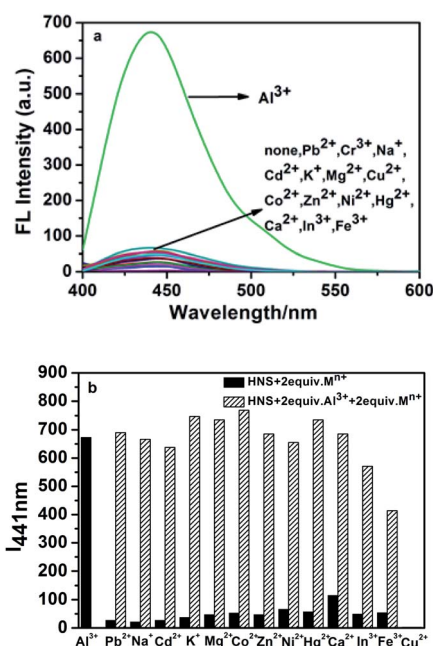


Fig. 3 (a) Fluorescence spectra changes of HNS (4  $\mu\text{M}$ ) in the presence of various metal ions (8  $\mu\text{M}$ ) in EtOH/ $\text{H}_2\text{O}$  (1 : 9, v/v, pH 5.3). (b) Fluorescence intensities of HNS (4  $\mu\text{M}$ ) at 441 nm ( $I_{441 \text{ nm}}$ ) upon addition of  $\text{Al}^{3+}$  in the presence of interfered metal ions.



previously reported Schiff base probes for  $\text{Al}^{3+}$  detection.<sup>8,9,15</sup> The relative standard deviation for five repeated measurements of  $8.0 \times 10^{-6}$  M of  $\text{Al}^{3+}$  was 1.08%, which illustrated that the response of **HNS** toward  $\text{Al}^{3+}$  was highly reproducible. It should be noted that this switch on sensing process could be easily observed by the naked eye. As shown in Fig. 7, **HNS** alone only exhibited very weak fluorescence, but gave a strong blue fluorescence in the presence of  $\text{Al}^{3+}$ .

### 3.3 Binding mode of **HNS** with $\text{Al}^{3+}$

A Job plot obtained from emission data showed a 2 : 1 stoichiometry of the complex formed between **HNS** and  $\text{Al}^{3+}$  (Fig. S4a†), which was corroborated by the mass spectra of  $\text{Al}^{3+}$ -**HNS** complex. The HRMS spectrum showed a peak at  $m/z$  483.1361 corresponding to  $2\text{HNS} + \text{Al}^{3+} - 2\text{H}^+$  [calcd,  $m/z$ : 483.1345] (Fig. 4), indicating that the binding of  $\text{Al}^{3+}$  to **HNS** induced deprotonation of the phenolic -OH group of **HNS**. Based on a 2 : 1 binding mode, the association constant ( $K$ ) was evaluated from the  $\text{Al}^{3+}$  ions titration curve (Fig. S4b†) using the modified Benesi-Hildebrand method<sup>19</sup> and was found to be  $1.14 \times 10^4 \text{ M}^{-1}$ .

Further evidence for the  $\text{Al}^{3+}$  coordination was obtained from the FT-IR and NMR spectral studies. In the IR spectrum of  $\text{Al}^{3+}$ -**HNS** complex shown in Fig. S5,† the sharp peaks at  $3450 \text{ cm}^{-1}$  and  $3350 \text{ cm}^{-1}$  may be attributed to N-H stretching of -CONH<sub>2</sub> group of **HNS**. On co-ordination with  $\text{Al}^{3+}$  ions, these peaks became a broad band, indicating that the primary amide participated in the binding with  $\text{Al}^{3+}$ . In addition, the stretching vibration band of C=N at  $1603 \text{ cm}^{-1}$  shifted to a lower frequency by  $31 \text{ cm}^{-1}$ , clearly indicating the coordination of imine nitrogen atom of **HNS** to aluminium ion.

<sup>1</sup>H NMR spectroscopy was used to ascertain the binding mode of **HNS** to  $\text{Al}^{3+}$ . <sup>1</sup>H NMR spectra of **HNS** were recorded in DMSO-*d*<sub>6</sub> with increasing concentrations of  $\text{Al}^{3+}$  (as its nitrate salt). Significant spectral changes were observed as shown in Fig. 5. After addition of  $\text{Al}^{3+}$ , the H<sub>a</sub> protons of the phenolic hydroxyl progressively disappeared, suggesting the occurrence

of deprotonation during the coordination to metal ions. The amino proton peak (H<sub>b</sub>) at around 6.37 ppm became broader, indicating the coordination of the amino nitrogen to  $\text{Al}^{3+}$ . Meanwhile, the imine proton peak (H<sub>c</sub>) at around 10.22 ppm became less intense and slightly downfield shifted by 0.02 ppm, supporting the coordination of the imine nitrogen to  $\text{Al}^{3+}$ . The overall changes in <sup>1</sup>H NMR spectra indicate the complexation between  $\text{Al}^{3+}$  and **HNS** through amino-N, azomethine-N and phenolic-OH. Based on the Job's plot, IR spectra, mass spectra and NMR titration, we proposed a rational coordinated mode that aluminium ion is hexa-coordinated with two tridentate ligand **HNS** shown in Fig. 5.

### 3.4 Fluorescence sensing of $\text{F}^-$

The sensor  $\text{Al}^{3+}$ -**HNS** for fluoride detection was prepared *in situ* by addition of 2.0 equiv. of  $\text{Al}^{3+}$  to C<sub>2</sub>H<sub>5</sub>OH/H<sub>2</sub>O (1 : 9, v/v) solution of **HNS**. The  $\text{F}^-$  sensing property of  $\text{Al}^{3+}$ -**HNS** was then evaluated by fluorescence titration experiments and the results were depicted in Fig. 6. As shown in Fig. 6a, the fluorescent intensity at 441 nm can be significantly quenched by  $\text{F}^-$  ions. The quenching effect of  $\text{F}^-$  on the fluorescence emission of the system is concentration dependent. There is a good linear relationship ( $R^2 = 0.9957$ ) between the fluorescence intensity and  $\text{F}^-$  concentration in the range of 0.8–20  $\mu\text{M}$ . Thus, probe **HNS** could not only act as a sensor for  $\text{Al}^{3+}$  but also for  $\text{F}^-$  in a successive manner.

Interferences from common anions have also been studied under the same conditions as used above for  $\text{F}^-$ . Out of the tested anions, significant quenching was observed only in case of fluoride (Fig. 6b). Furthermore, the other coexistent anions had a small and negligible effect on the emission intensity of the system, suggesting the high selectivity of the  $\text{Al}^{3+}$ -**HNS** ensemble for  $\text{F}^-$  over other anions. The detection limit of  $\text{Al}^{3+}$ -**HNS** complex as a fluorescent sensor for the analysis of  $\text{F}^-$  was obtained from a plot of fluorescence intensity as a function of

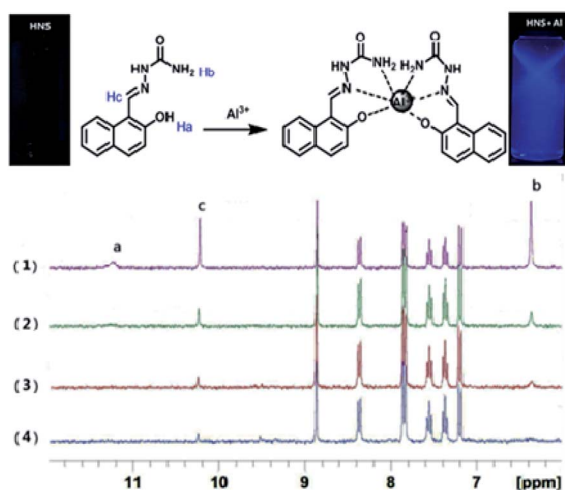


Fig. 4 The ESI-MS spectra of  $\text{Al}^{3+}$ -**HNS** complex.

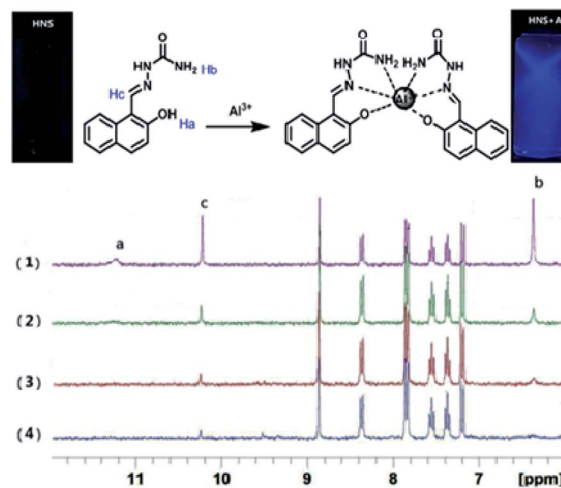


Fig. 5 Top: Binding mode of  $\text{Al}^{3+}$ -**HNS** system. Bottom: <sup>1</sup>H NMR spectra of **HNS** in the presence of different concentrations of  $\text{Al}^{3+}$  ( $\text{NO}_3$ )<sub>3</sub>·9H<sub>2</sub>O in *d*<sub>6</sub>-DMSO: (1) **HNS** only; (2) **HNS** +  $\text{Al}^{3+}$  (0.25 equiv.) and (3) **HNS** +  $\text{Al}^{3+}$  (0.5 equiv.) (4) **HNS** +  $\text{Al}^{3+}$  (1 equiv.).



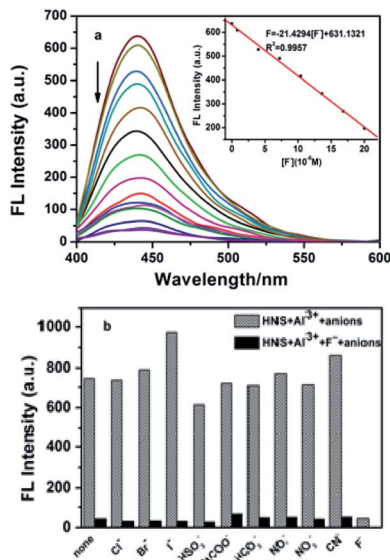
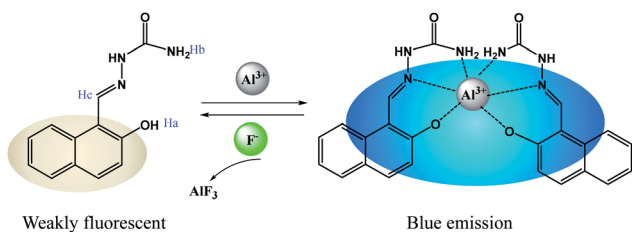


Fig. 6 (a) Fluorescent emission spectra of  $\text{Al}^{3+}$ -HNS in the presence of different concentrations of  $\text{F}^-$  in ethanol/water = 1 : 9 at pH 5.3.  $[\text{HNS}] = 4 \times 10^{-6}$  M,  $[\text{Al}^{3+}] = 8 \times 10^{-6}$  M, and (b) competitive signaling of fluoride ions by  $\text{Al}^{3+}$ -HNS in the absence (the grey) and presence (the black) of other anions as background in ethanol/water = 1 : 9 (pH = 5.3).  $[\text{HNS}] = 4 \times 10^{-6}$  M,  $[\text{anions}] = 8 \times 10^{-5}$  M.

the concentration of the different amounts of  $\text{F}^-$  added. It was found that  $\text{Al}^{3+}$ -HNS has a detection limit of  $7.89 \times 10^{-7}$  M for  $\text{F}^-$ , which is much lower than the maximum contaminant level defined by the U.S. Environmental Protection Agency ( $4.0 \text{ mg L}^{-1}$ ,  $211 \text{ } \mu\text{M}$ ).<sup>40</sup>

This phenomenon can be supposed that the  $\text{Al}^{3+}$  has been captured by  $\text{F}^-$  ions from the  $[\text{Al}^{3+}$ -HNS] complex to form a new more stable complex  $\text{AlF}_3$  and resulted in the release of free HNS (Scheme 2). To elucidate the nature of the strong binding of  $\text{F}^-$  to  $\text{Al}^{3+}$ , the UV-vis spectra,  $^1\text{H}$  NMR spectra and HRMS experiments have been conducted respectively. As shown in Fig. 7, toward adding  $\text{F}^-$ , the characteristic band at 381 nm of  $[\text{Al}^{3+}$ -HNS] complex is disappeared, while the absorption response is recovered to the original state of free HNS. The  $^1\text{H}$  NMR spectrum of  $[\text{Al}^{3+}$ -HNS] complex has been investigated in absence and in presence of  $\text{Al}^{3+}$  in  $\text{DMSO}-d_6$  (Fig. S6†). The broader amino proton peak ( $\text{H}_b$ ) at around 6.37 ppm became sharp and the imine proton peak ( $\text{H}_c$ ) at around 10.22 ppm became recovered, indicating the dissociation of the  $\text{Al}^{3+}$  to HNS after adding  $\text{F}^-$ . This was further confirmed by HRMS studies of



Scheme 2 Probable sensing mechanism for  $\text{Al}^{3+}$  and  $\text{F}^-$  by HNS.

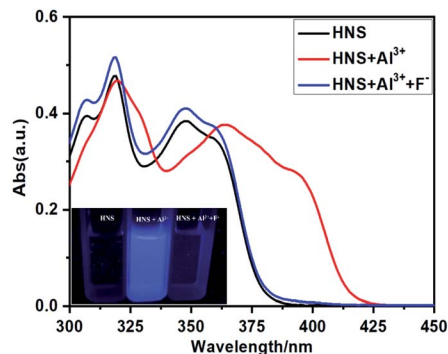


Fig. 7 UV-vis spectra of HNS ( $40 \text{ } \mu\text{M}$ ) in the presence of  $\text{Al}^{3+}$  (4 equiv.) and  $\text{Al}^{3+}$  (4 equiv.) +  $\text{F}^-$  (8 equiv.). Inset: the visual change exhibited for HNS in the presence of  $\text{Al}^{3+}$  and  $\text{F}^-$  under UV light.

HNS +  $\text{Al}(\text{NO}_3)_3$  (2.0 equiv.) + NaF (8.0 equiv.) (Fig. S7†). The peak of  $2\text{HNS} + \text{Al}^{3+} - 2\text{H}^+$  [calcd,  $m/z$ : 483.1345] at  $m/z = 483.1355$  became very weak and the peak at  $m/z = 230.0924$  corresponded to  $[\text{HNS} + \text{H}^+]$  (230.0930) displayed a prominent peak, which further proved that the  $[\text{Al}^{3+}$ -HNS] complex released the free HNS by addition of  $\text{F}^-$ . The overall changes in UV-vis spectra,  $^1\text{H}$  NMR spectra and HRMS analysis confirmed the dissociation between  $\text{Al}^{3+}$  and HNS, meanwhile, indirectly demonstrated the higher binding affinity of  $\text{Al}^{3+}$  toward  $\text{F}^-$  compared to that with HNS.

### 3.5 Cellular imaging by fluorescence microscopy

To demonstrate the potential of HNS to image  $\text{Al}^{3+}$  in living matrices, we carried out the experiments in SiHa cells. First, an MTT assay in SiHa cells has been performed to explore the cytotoxicity of HNS (Fig. S8†). The result confirmed that the HNS has no significant cytotoxicity on SiHa cells at a low concentration. As shown in Fig. 8, the cells showed no detectable fluorescence upon incubation only with HNS ( $50 \text{ } \mu\text{M}$ ) for 30 min. However, bright blue fluorescence was observed after the cells were incubated with receptor HNS ( $50 \text{ } \mu\text{M}$ ) and  $\text{Al}^{3+}$  ( $100 \text{ } \mu\text{M}$ ) simultaneously. Cells were subsequently incubated with a  $500 \text{ } \mu\text{M}$  solution of  $\text{F}^-$  for 10 min under the same conditions, whereupon a significant decrease in the fluorescence was observed. During the whole process, the cells remained in good cell morphological state, demonstrating the low cytotoxicity of the probe further. These results demonstrate that HNS has good cell-membrane permeability and can be used for sequential detection of  $\text{Al}^{3+}$  and  $\text{F}^-$  in an *in vitro* cellular system.

## 4. Conclusions

In conclusion, we have developed an “off-on” type chemosensor HNS for  $\text{Al}^{3+}$  in weak acid aqueous solution. The receptor showed a high sensitivity for  $\text{Al}^{3+}$  detection at concentrations ranging from  $0.8 \text{ } \mu\text{M}$  to  $8 \text{ } \mu\text{M}$  with a detection limit of  $67.5 \text{ nM}$ . Moreover, fluorescence of the  $\text{Al}^{3+}$ -HNS complex was drastically quenched by  $\text{F}^-$  via  $\text{Al}^{3+}$  displacement. Thus the  $\text{Al}^{3+}$ -HNS ensemble represents itself as an effective chemosensor for  $\text{F}^-$



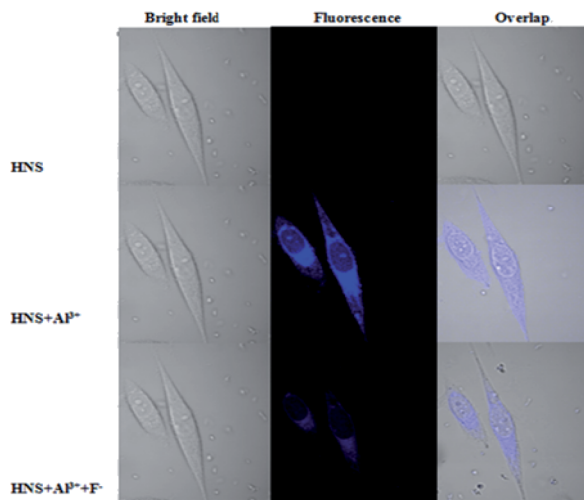


Fig. 8 Fluorescence images of SiHa cells treated with HNS,  $\text{Al}^{3+}$  and  $\text{F}^-$ . (Left) Bright field image; (middle) fluorescence image; (right) merged image.

too. The detection limit of  $\text{F}^-$  is quite low ( $0.789 \mu\text{M}$ ) using this ensemble. Furthermore, HNS has been utilized successfully for sequential detection of  $\text{Al}^{3+}$  and  $\text{F}^-$  ions in living cells. These merits of HNS, *i.e.*, requiring facile one-step synthesis, working in aqueous media, possessing low detection limits, displaying high selectivity, and being suitable for monitoring  $\text{Al}^{3+}$  in living cell, make HNS a promising candidate as a chemosensor for  $\text{Al}^{3+}$  and  $\text{F}^-$  ions.

## Conflicts of interest

There are no conflicts to declare.

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