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Synthesis and *in vitro* anti-proliferative evaluation of naphthalimide–chalcone/pyrazoline conjugates as potential SERMs with computational validation†

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A series of naphthalimide-chalcone/pyrazoline conjugates was prepared and evaluated for their anti-breast cancer potential against estrogen responsive, *i.e.* MCF-7 (ER+), and triple-negative, *i.e.* MDA-MB-231 (ER–), cell lines. The structure-activity-relationship (SAR) was deduced based on the influence of linker length, substituents on the phenyl ring and the generated functionalities, on anti-proliferative activities. Docking simulations further delineate the type of interactions of the designed molecules with the selected targets. This report discloses the scope of triazole tethered naphthalimide-chalcone/pyrazoline conjugates as anti breast cancer agents.

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Introduction

Cancer, an uncontrolled growth of cells, is the second leading cause of death worldwide. The most commonly diagnosed cancers are lung (12.6%), breast (11.9%), colorectal (9.8%), and prostate (7.7%).¹ Breast cancer is the most frequent cancer among women, affecting 2.1 million women each year. A World Health Organization (WHO) survey estimated that 627 000 women died from breast cancer worldwide in 2018.² Treatment for breast cancer may include surgery, radiation, chemotherapy, targeted therapy and hormone therapy. The proliferation of mammary cells is stimulated by the binding of hormone estrogen to estrogen receptors (ERs, existing in isomeric forms of ER α and ER β), the later has developed as target for breast cancer therapy.³ Consequently, selective inhibition of ER α has led to the emergence of drugs known as Selective Estrogen Receptor Modulators (SERMs) and is an important strategy in the treatment of breast cancer.⁴

Chalcones are a prolific class of flavonoids with significant occurrence in many edible fruits with medicinal properties.⁵ Chalcones, with an aromatic enone as a central core, showed anti-inflammatory,⁶ anti-histaminic,⁷ anti-oxidant,⁸ anti-protazoal,⁹ antibacterial,¹⁰ antimalarial,¹¹ antifungal,¹² anti-HIV¹³ activities. Moreover, chalcones have shown potent inhibition of

several anticancer targets, including thioredoxin reductase, and tubulin polymerization^{14,15} and by preferentially killing cancer cells over non-cancer cells.¹⁶ In this context, to improve the efficacy of chalcones while evading drug-resistance, molecular hybridization involving dual- or multi-target approach has been investigated. For instance, Lee *et al.*,¹⁷ synthesized quinoline-chalcones targeting MDR tumor and metastatic breast cancer cells, whereas Modzelewska *et al.*,¹⁸ reported a series of bis-chalcones with exceptional inhibitory potency in the growth of human breast and colon cancer cells.

Naphthalimides, owing to its flat aromatic structure and DNA-intercalating tendencies, have emerged as target motif in the area of cancer therapy.¹⁹ Some mono or bis-naphthalimide derivatives have displayed promising activities against tumor cells, among which, mitonafide, amonafide, elinafide and azonafide proceeded to clinical trials.^{20,21} However, poor therapeutic index, poor water solubility, and neurological toxicity impeded their further advancement in clinical phase.²² Thus, the ongoing efforts are geared towards modifying the molecular framework of naphthalimide based scaffolds to improve their therapeutic properties. A literary survey has disclosed better results of naphthalimide based scaffolds as anti tumor including anti-breast cancers.^{23–26}

1*H*-1,2,3-triazole core, on the other hand, presents desirable properties such as hydrogen bonding ability, resistance to oxidation, reduction, and enzymatic hydrolysis in physiological conditions. Besides, they have shown a broad spectrum of medicinal potential including anticancer.^{27–31} Therefore, on this backdrop and in continuation of our research venture,^{32–35} the present manuscript describes the hybridization of the chosen pharmacophores *i.e.* substituted chalcones/pyrazolines and 1,8-naphthalimides using triazole as a linker. Two human breast cancer cell lines *i.e.* MCF-7 (ER+) and MDA-MB-231 (ER–) were

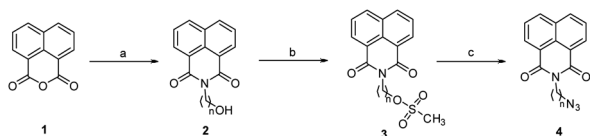
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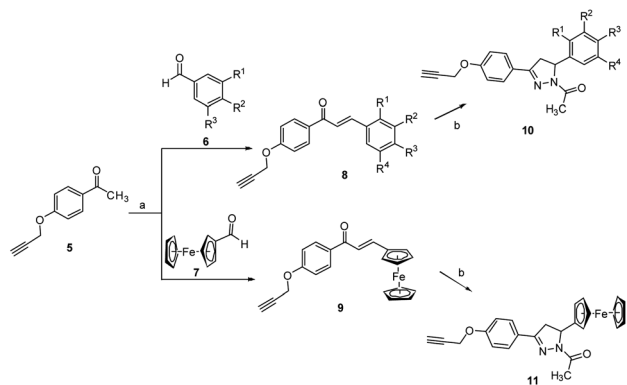
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† Electronic supplementary information (ESI) available: Scanned copies of ¹H and ¹³C spectra of few representative compounds *i.e.* 12b, 12g, 13b, 13d, 13g, 14b, 15b and Table S1. See DOI: 10.1039/d0ra01822h





Scheme 1 Reagents and conditions: (a) ethanol amine ($n = 2$)/propanol amine ($n = 3$), ethanol, reflux, 1 h (b) mesyl chloride, Et_3N , dry CHCl_3 , RT, 3 h (c) NaN_3 , DMF, 60°C , 2 h.



Scheme 2 Reagents and conditions: (a) NaOH (40% w/v), ethanol, RT, 1 h (b) hydrazine hydrate, acetic acid, 130°C , reflux, 2 h.

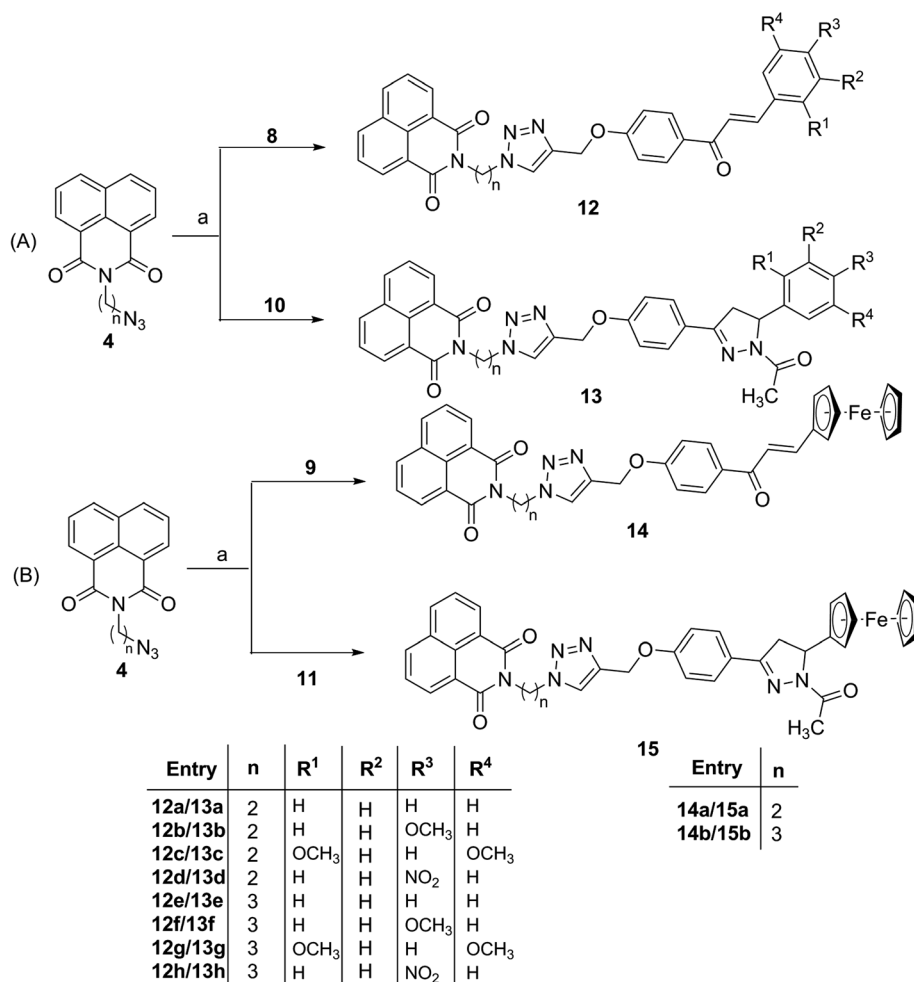
used to investigate the *in vitro* anti-proliferative potential of the synthesized compounds.

Results and discussion

Chemistry

The first set of precursors were obtained by treating 1,8-naphthalic anhydride **1** with ethanol/propanol amine under reflux to yield **2**. Further, compound **2** was treated with mesyl chloride and Et_3N in dry CHCl_3 to get the mesylated product **3**, followed by its treatment with NaN_3 in DMF to obtain the precursor **4**, (Scheme 1).

For second set of precursors, different aldehydes *viz.* **6** and **7** were treated with ketone **5**, formed by *O*-propargylation of 4-hydroxy acetophenone, under the basic condition to form the Claisen–Schmidt condensation products **8** and **9**. The introduction of pyrazoline rings is done by treating **8/9** with hydrazine hydrate in acetic acid at 130°C to afford **10** and **11** respectively, (Scheme 2). The rationale for introducing pyrazoline functionality in the designed conjugates is based on its favourable role in enhancing anti-proliferative potential.^{36,37}



Scheme 3 Reagents and condition: (a) CuSO_4 , sodium ascorbate, $\text{EtOH} : \text{H}_2\text{O}$ (90 : 10), RT, 10 h.



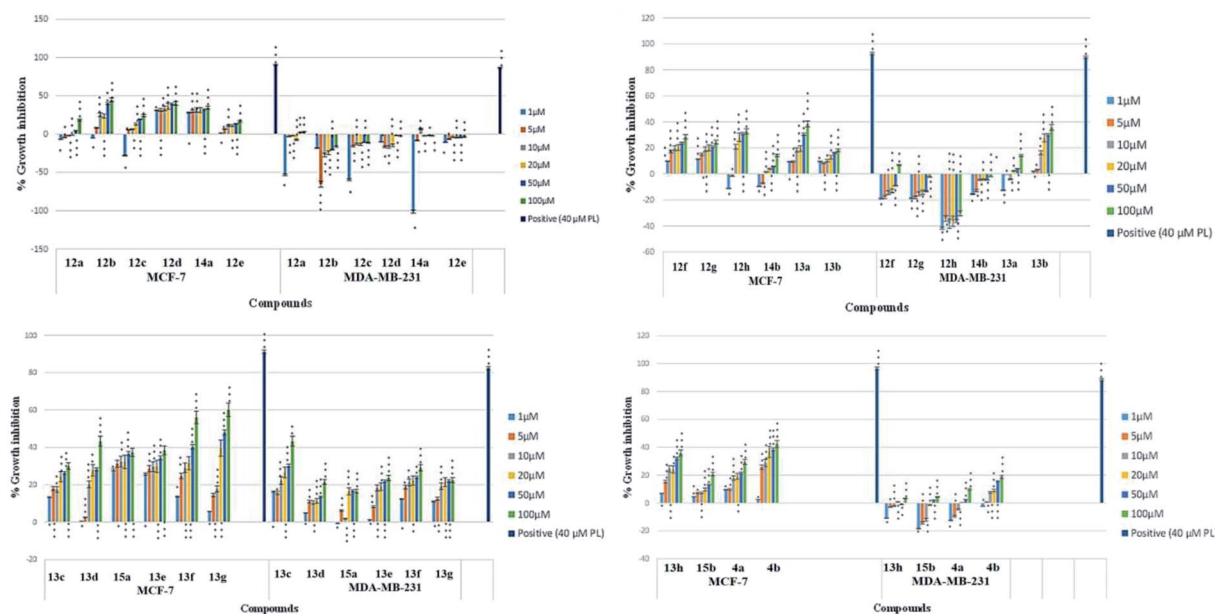


Fig. 1 Representative graphs comparing the percentage growth inhibition of MCF-7 and MDA-MB-231 cells at selected concentrations of test compounds. 40 μM Plumbagin was used as a positive control. Data are mean \pm standard deviation S.D ($n = 3$), where $*p < 0.05$, $**p < 0.01$ and $***p < 0.001$ significant difference to untreated control.

Subsequently, the target conjugates *viz.* **12a–h** and **13a–h** were synthesized by azide–alkyne cycloaddition reaction between **4** and **8/10** using CuSO_4 and sodium ascorbate in absolute ethanol, (Scheme 3A). The ferrocene based counterparts *viz.* **14a–b** and **15a–b** were synthesized analogously from the cycloaddition reaction between **4** and **9/11**, (Scheme 3B). The purification of the reaction mixture *via* column chromatography using ethyl acetate : hexane (10 : 90) mixture as eluent, afforded the desired conjugates in good to excellent yields. The structures of the synthesized molecules were assigned based on the spectral data and analytical evidences. For example, compound **12b**, showed molecular ion peak at m/z 559.1975 $[\text{M} + \text{H}]^+$ in its HRMS (ESI) spectrum. Its ^1H NMR spectrum exhibited characteristic singlets at δ 3.83 and 7.76 corresponding to methoxy and triazole ring proton respectively. The signals at δ 188.83, 161.91 and 161.63 corresponding to the carbonyl groups along with requisite signals in ^{13}C NMR spectrum further confirmed the assigned structure.

In vitro anti-proliferative evaluation and SAR

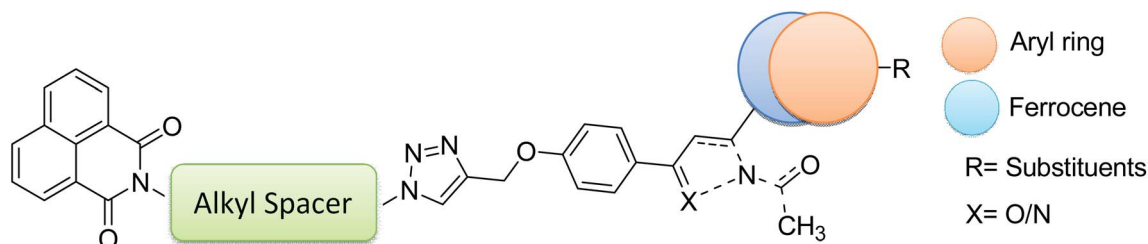
The synthesized series of triazole tagged naphthalimide-chalcone/pyrazoline conjugates was evaluated for their anti-proliferative activities against estrogen receptor positive and triple-negative human breast cancer cell lines, MCF-7 (ER⁺) and MDA-MB-231 (ER[−]) respectively, using MTT assay.³⁸ The percentage growth inhibition of MCF-7 and MDA-MB-231 cells were recorded at varying concentrations of the synthesized conjugates using Plumbagin as the positive control, and the results are depicted in Fig. 1. The IC_{50} values of the synthesized compounds, which is the concentration required to inhibit the growth of 50% of cells, are included in Table 1.

As evident from Table 1, naphthalimide–chalcone conjugates, **12a–h** failed to inhibit the growth of both the cell lines even at the highest tested concentration *i.e.* 100 μM . Replacement of chalcone core with pyrazoline ring did not result in the improvement of their anti-proliferative activities against MDA-MB-231 cells as evident from conjugates **13a–h**. Nevertheless,

Table 1 The IC_{50} values (in μM) of tested compounds against MCF-7 and MDA-MB-231 cells

Entries	MCF-7	MDA-MB-231
12a	>100	>100
12b	>100	>100
12c	>100	>100
12d	>100	>100
12e	>100	>100
12f	>100	>100
12g	>100	>100
12h	>100	>100
13a	>100	>100
13b	>100	>100
13c	>100	>100
13d	>100	>100
13e	>100	>100
13f	98.11	>100
13g	62.23	>100
13h	>100	>100
14a	>100	>100
14b	>100	>100
15a	>100	>100
15b	>100	>100
4a	>100	>100
4b	>100	>100
Tamoxifen	50	75
Plumbagin	3.5	4.4





Preference Order

- ❖ Pyrazoline ring over chalcone
- ❖ Propyl over ethyl as spacer
- ❖ Aryl ring over Ferrocene
- ❖ R= Electron donating over withdrawing

Fig. 2 Generalized SAR of the synthesized compounds.

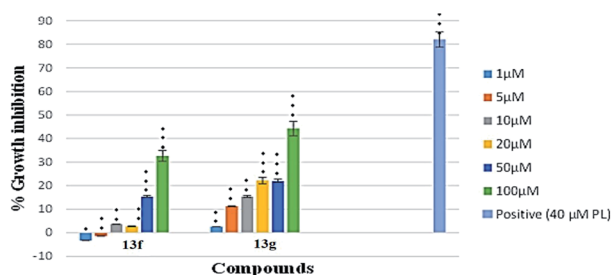


Fig. 3 Representative graph comparing the percentage growth inhibition of HEK-293 cells at selected concentrations of test compounds **13f** and **13g**. 40 μM Plumbagin was used as a positive control. Data are mean ± standard deviation S.D ($n = 3$), where * $p < 0.05$, ** $p < 0.01$ and *** $p < 0.001$ significant difference to untreated control.

Table 2 The IC_{50} values (in μM) of tested compounds against HEK-293 cells

Entries	HEK-293
13f	>100
13g	>100

conjugates **13f** and **13g** showed moderate cytotoxicity against MCF-7 cell lines. The conjugate **13f** bearing a spacer length of $n = 3$, pyrazoline ring and mono-methoxy substituted phenyl ring, showed IC_{50} value 98.11 μM whereas **13g** with $n = 3$ and dimethoxy substituted phenyl ring, exhibited an IC_{50} value 62.23 μM. Further, replacing the aryl core with ferrocene in naphthalimide-chalcones, **14a–b** as well as naphthalimide-pyrazolines, **15a–b** could not improve activity profiles. Thus, the SAR analysis demonstrated that propyl spacer, pyrazoline ring and electron donating substituents were requisite in order to impart activity in the designed scaffolds. In contrast, ethyl spacer, chalcone core and unsubstituted/nitro substituted/ferrocene tagged conjugates failed to elicit any activity. However, the observed anti-proliferative activities of the active compounds against MCF-7 cells is less as compared to the reference

cytotoxic agent, Plumbagin ($IC_{50} = 3.5$ μM against MCF-7 cells), but comparable to that of standard drug, Tamoxifen ($IC_{50} = 50$ μM against MCF-7 cells). The graphical representation of generalized SAR for the synthesized compounds is elucidated in Fig. 2.

The toxicity of moderately active compounds **13f** and **13g** was also tested against normal/non-tumor cell line *i.e.* Human Embryonic Kidney (HEK-293). The compounds were non-cytotoxic with $IC_{50} > 100$ μM (Fig. 3 and Table 2).

Molecular docking and ADMET properties prediction

In order to identify the type of interactions operating between the synthesized compounds (ligands) and the estrogen receptors

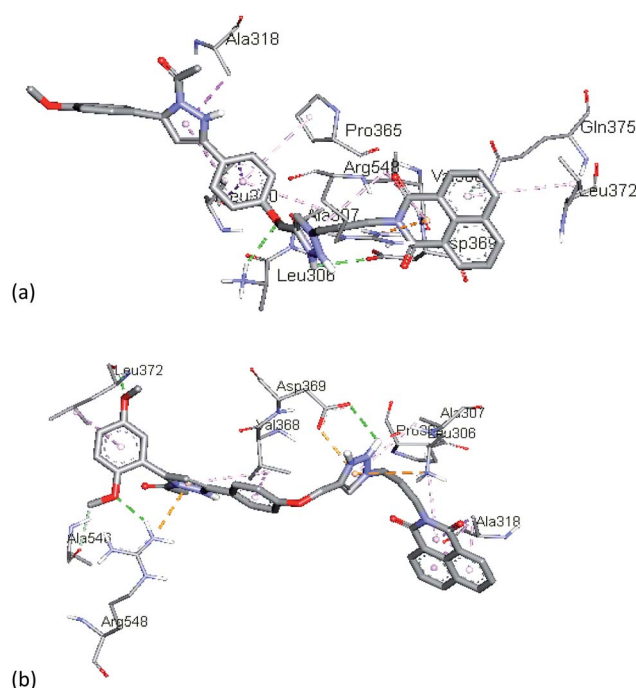


Fig. 4 The docked binding modes of (a) **13f** and (b) **13g** in the binding site of ER α (PDB:3ERT).



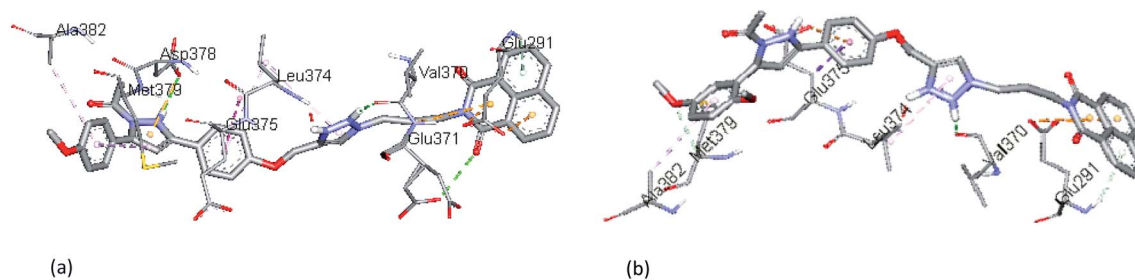


Fig. 5 The docked binding modes of (a) **13f** and (b) **13g** in the binding site of ER β (PDB:3LOS).

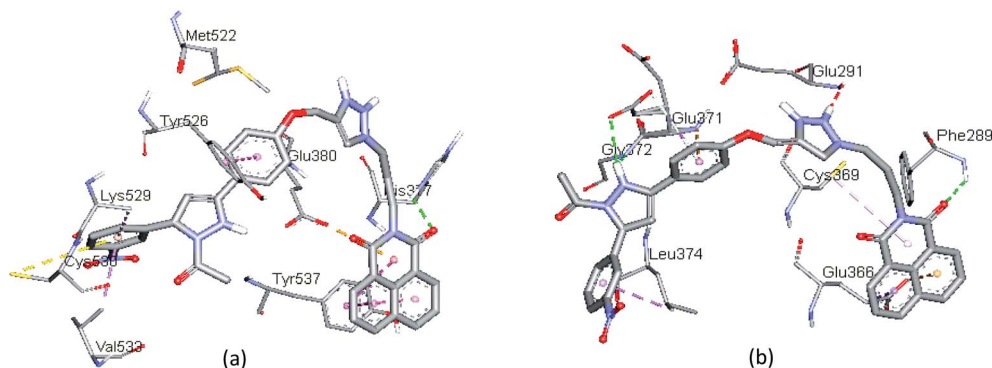


Fig. 6 The docked binding modes of **13h** in the binding site of (a) ER α and (b) ER β .

(protein), the active compounds *viz.* **13f** and **13g** were chosen for docking studies with ER α and ER β . Further an inactive compound **13h** was also included for comparison purpose.

The molecular docking analysis of compound **13f** (IC_{50} = 98.11 μ M; B.E. = -8.3 kcal mol $^{-1}$), presented hydrophobic interactions with Leu306, Ala307, Leu310, Ala318, Leu372, and Val368 of ER α receptor. The results also revealed that hydrophobic forces dominate the interactions between **13f** and the ER α receptor. Moreover, 1*H*-1,2,3-triazole ring forms π -alkyl interaction with Val368 while the protonated nitrogen in the triazole ring forms H-bonding interaction with Asp369. The naphthalene ring also forms hydrophobic interaction with residues, Val368 and Leu373 (Fig. 4a). The introduction of a 2,5-dimethoxy substituent in compound **13g** (B.E. = -8.4 kcal mol $^{-1}$) enhanced the anti-proliferative potency (IC_{50} = 62.23 μ M) compared to **13f** (IC_{50} = 98.11 μ M). Interestingly, this was reflected in the hydrogen bonding and hydrophobic interactions of compound **13g** with Leu372, Ala546, and Arg548 in the ligand-binding pocket of ER α . The *meta*-methoxy unit also featured a conventional H-bonding with the amino unit of Leu372 at a bond distance (b.d) of 2.14 Å and bond angle (b.a) of 138.73°. The *ortho*-methoxy unit on the other hand interacted with Arg548 and Ala546 *via* conventional and carbon-hydrogen bonding (b.d = 2.35 Å and 3.35 Å; b.a = 154.37°, respectively). The dimethoxy substituted phenyl ring showed π -alkyl interaction with Leu372. These binding modes helped to lock-in the ligand within the binding site of the ER α receptor for improved binding affinity. The H-bonding interaction of triazole linker with Asp369 is accompanied by π -cation and π -anion

interactions with Asp369 and Leu306 while the naphthalene ring is involved in π -alkyl and π -sigma hydrophobic interactions with Ala 318 and Pro365 (Fig. 4b). These results demonstrate that substituents on the phenyl ring can have drastic effects on the potency of these compounds as ER α antagonists.

The evaluation of potent compounds in the active region of ER β showed a binding mode for **13f** (B.E. = -7.8 kcal mol $^{-1}$) in which the triazole and pyrazoline NH units donate H-bond to the carbonyl oxygen atoms of Val370 and Asp378 respectively (b.d = 2.54 Å and 2.76 Å; b.a = 90.5° and 110° respectively). The naphthalene ring makes electrostatic contacts with Glu291 and H-bonding interaction with Glu372. Whereas the *para*-methoxy substituent forms hydrophobic interactions with Met379 and Ala382, respectively (Fig. 5a). Further, analysis of the docked complex of **13g** (B.E. = -8.3 kcal mol $^{-1}$) showed that the naphthalene unit interacts with Glu291 *via* electrostatic contact. The dimethoxy substituted phenyl ring formed π -alkyl interaction with Ala382 while the *meta*-methoxy unit forms a C-H bond with Met379. Strong conventional H-bonding was also observed between the NH group of triazole ring and Val370, (b.d = 2.11, b.a = 135°) (Fig. 5b).

The docked binding modes of **13h** in the binding site of ER α and ER β have been shown in Fig. 6. The compound featured three hydrogen bonds with Gly372, Glu371 and Phe289 (3.00 Å, 2.78 Å and 3.04 Å, respectively), and hydrophobic interactions with Leu374, Cys369, and Glu366 respectively, in the ligand-binding pocket of ER α . Compared to **13f** and **13g**, compound **13h** interacts with amino acid residues in a different active site of ER β ; it showed one hydrogen bond interaction with His377.



The binding profile is also accompanied by hydrophobic interactions with Tyr537, Tyr526, Val533, and Lys529, as well as electrostatic interactions with Glu380 and Cys530, respectively. The binding profile in the docked complex of compound **13h** with ER α and ER β partly justifies the low inhibitory activity observed (>100 μ M).

Furthermore, the predicted physicochemical properties and ADMET descriptors (Table S1, see ESI[†]) suggest good drug-like properties for compounds **13f** and **13g**. For instance, the compounds present good gastrointestinal absorption which correlates with the predicted water solubility. A closer look at the predicted metabolism interactions towards CYP (Cytochrome P450) isoforms revealed that the compounds are suitable inhibitors of cytochrome P450 and also CYP2D6/CYP3A4 substrate, although **13g** seems to be resistant to CYP1A2 inhibitor.

Conclusion

The current work disclosed the synthesis of naphthalimide-chalcone/pyrazoline conjugates tethered by triazole ring and their anti-proliferative effects against two breast cancer cell lines. The anti-proliferative evaluation identified two compounds, **13f** and **13g** to be explicitly targeting estrogen-responsive breast cancer cell line MCF-7 (ER⁺) while the whole series was inactive against estrogen non-responsive/triple-negative cell line MDA-MB-231 (ER⁻). The active compound displayed IC₅₀ value of 62.23 μ M which is comparable to the standard anti-breast cancer drug Tamoxifen. In order to confirm their safety profiles, the active compounds were evaluated against HEK-293 cell line and proved to be non-cytotoxic. The docking simulations as well as physicochemical properties were studied to get an insight into the type of interactions and drug likeness of the synthesized compounds. The only limitation is its moderate activity which needs further modification to get enhanced. Thus, the designed molecular framework can be set as therapeutic template to particularly target estrogen responsive breast cancer and thus can contribute in the venture of developing SERMs.

Experimental

General information

Stuart Digital Melting Point apparatus (SMP10) and an open capillary was used to determine melting points and are uncorrected. BRUKER AVANCE II (500 and 125 MHz) and JEOL (400 and 100 MHz) NMR spectrometers were used in order to record ¹H and ¹³C NMR spectra of the synthesized compounds dissolved in CDCl₃ (Sigma-Aldrich). Tetramethylsilane (TMS) was kept as reference and chemical shift values are expressed as parts per million (ppm) downfield from TMS. While the coupling constants represented by *J* values are expressed in hertz (Hz). Splitting patterns are indicated as s: singlet, d: doublet, t: triplet, m: multiplet, dd: double doublet, dt: doublet of a triplet and br: broad peak. Mass spectra were recorded on a Bruker micrOTOF-QII high resolution mass spectrometer.

General procedure for the synthesis of **4**

To a well stirred solution of 1,8-naphthalic anhydride, **1** (1 mmol) in absolute ethanol, ethanol/propanol amine (1.5 mmol) was added and the reaction mixture was refluxed for 1 h. On cooling, the reaction mixture was filtered to get solid **2**. The intermediate **2** (1 mmol) was treated with mesyl chloride (1.5 mmol) and Et₃N (2 mmol) in dry CHCl₃ for 3 h at room temperature. The resulting product **3** was further reacted with NaN₃ in dry DMF at 60 °C for 2 h to afford **4**.

General procedure for the synthesis of **8/9** and **10/11**

K₂CO₃ (2 mmol) was added to a stirred solution of 4-hydroxy acetophenone (1 mmol) in acetone. After the generation of anion, propargyl bromide (1.5 mmol) was added and the reaction mixture was allowed to stir for 5 h to yield **5**. The compound **5** (1 mmol) was then dissolved in alkaline ethanol and reacted with different aldehydes **6/7** (1.2 mmol) at room temperature for 1 h to obtain **8/9**. Further, hydrazine hydrate (5 mmol) and catalytic amount of acetic acid (0.5 mmol) was added to solution of **8/9** (1 mmol) in ethanol and the reaction mixture was refluxed for 1 h to afford **10/11**.

General procedure for the synthesis of **12/13** and **14/15**

To the solution of **4** (1 mmol) and **8/10** (1.2 mmol) in ethanol : water (90 : 10) catalytic amount of CuSO₄ and sodium ascorbate was added. The reaction was monitored on TLC and on its completion, the crude product was extracted using CHCl₃ : H₂O. The crude was then purified on column chromatography using ethylacetate : hexane (10 : 90) as eluent to afford final set of compounds **12/13**. The similar procedure was followed using **4** and **9/11** in order to produce the final hybrids **14/15** in good yields.

Physical and spectral data

2-(2-(4-((4-Cinnamoylphenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12a**)**. Yield-86%, bright yellow solid, MP = 148–150 °C, ¹H NMR (400 MHz, CDCl₃): δ 4.68–4.70 (m, 2H, CH₂), 4.78–4.80 (m, 2H, CH₂), 5.28 (s, 2H, OCH₂), 7.10 (d, *J* = 8.9 Hz, 2H, ArH), 7.30–7.34 (m, 3H, ArH), 7.39 (d, *J* = 15.5 Hz, 1H, olefinic H), 7.55 (d, *J* = 8.6 Hz, 2H, ArH), 7.65–7.70 (m, 3H, ArH), 7.75 (s, 1H, triazole-H), 7.95 (d, *J* = 8.8 Hz, 2H, ArH), 8.15 (dd, *J* = 0.9, 8.3 Hz, 2H, ArH), 8.40 (dd, *J* = 0.8, 7.2 Hz, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ 188.69, 163.94, 162.01, 144.02, 134.40, 131.63, 131.56, 131.34, 130.76, 130.33, 129.33, 128.91, 128.35, 128.21, 126.98, 123.40, 121.98, 121.93, 114.71, 114.60, 62.12, 48.02, 39.62. HRMS (ESI) calcd for C₃₂H₂₄N₄O₄ [M + H]⁺ 529.1876, found 529.1895.

(E)-2-(2-(4-((3-(4-Methoxyphenyl)acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12b**)**. Yield-85%, bright yellow solid, MP = 154–156 °C, ¹H NMR (400 MHz, CDCl₃): δ 3.83 (s, 3H, OCH₃), 4.64–4.67 (m, 2H, CH₂), 4.75–4.78 (m, 2H, CH₂), 5.24 (s, 2H, OCH₂), 6.91 (d, *J* = 8.8 Hz, 2H, ArH), 7.00 (d, *J* = 9.0 Hz, 2H, Ar H), 7.38 (d, *J* = 15.6 Hz, 1H, olefinic H), 7.57 (d, *J* = 8.6 Hz, 2H, ArH), 7.67–7.73 (m, 3H, ArH), 7.76 (s, 1H, triazole-H), 7.96 (d, *J* = 8.9 Hz, 2H,



ArH), 8.17 (dd, $J = 1.0, 8.4$ Hz, 2H, ArH), 8.46 (dd, $J = 1.1, 7.2$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 188.83, 164.04, 161.91, 161.63, 143.99, 143.79, 134.53, 131.76, 131.68, 131.67, 130.75, 130.21, 128.26, 127.84, 127.08, 123.54, 122.00, 119.57, 114.69, 114.48, 62.13, 55.49, 48.10, 39.70. HRMS (ESI) calcd for $\text{C}_{33}\text{H}_{26}\text{N}_4\text{O}_5$ $[\text{M} + \text{H}]^+$ 559.1981, found 559.1975.

(E)-2-(2-(4-((4-(3-(2,5-Dimethoxyphenyl)acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12c). Yield-85%, light brown solid, MP = 146–148 °C, ^1H NMR (400 MHz, CDCl_3): δ 3.80 (s, 3H, OCH_3), 3.86 (s, 3H, OCH_3), 4.60 (t, $J = 6.8$ Hz, 2H, CH_2), 4.71 (t, $J = 7.0$ Hz, 2H, CH_2), 5.25 (s, 2H, OCH_2), 6.85 (d, $J = 8.9$ Hz, 1H, ArH), 6.92 (dd, $J = 3.1, 9.1$ Hz, 1H, ArH), 7.08 (d, $J = 8.6$ Hz, 2H, ArH), 7.15 (d, $J = 3.0$ Hz, 1H, ArH), 7.58 (d, $J = 15.6$ Hz, 1H, olefinic H), 7.75–7.78 (m, 2H, ArH), 7.89 (s, 1H, triazole-H), 8.01 (d, $J = 8.6$ Hz, 2H, ArH), 8.05 (d, $J = 15.5$ Hz, 1H, olefinic H), 8.21 (d, $J = 8.1$ Hz, 2H, ArH), 8.58 (d, $J = 7.4$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 189.15, 164.40, 160.95, 153.45, 153.10, 143.65, 140.40, 135.21, 132.10, 131.95, 131.51, 130.90, 128.15, 127.45, 125.10, 123.40, 122.95, 122.35, 117.15, 114.51, 113.85, 112.50, 62.15, 56.20, 55.85, 48.21, 38.40. HRMS (ESI) calcd for $\text{C}_{34}\text{H}_{28}\text{N}_4\text{O}_6$ $[\text{M} + \text{H}]^+$ 589.2087, found 589.2072.

(E)-2-(2-(4-((4-(3-(4-Nitrophenyl)acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12d). Yield-85%, yellow solid, MP = 161–163 °C, ^1H NMR (400 MHz, CDCl_3): δ 4.65–4.68 (m, 2H, CH_2), 4.74–4.76 (m, 2H, CH_2), 5.25 (s, 2H, OCH_2), 6.95 (d, $J = 8.8$ Hz, 2H, ArH), 7.36 (d, $J = 8.7$ Hz, 2H, ArH), 7.40 (d, $J = 15.5$ Hz, 1H, olefinic H), 7.63 (d, $J = 8.8$ Hz, 2H, ArH), 7.70–7.74 (m, 3H, ArH), 7.76 (s, 1H, triazole-H), 8.10 (d, $J = 8.7$ Hz, 2H, ArH), 8.18 (dd, $J = 0.8, 8.1$ Hz, 2H, ArH), 8.50 (dd, $J = 0.7, 7.1$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 189.10, 164.10, 161.65, 143.92, 143.70, 141.30, 134.50, 131.91, 131.70, 131.60, 130.66, 130.25, 128.36, 127.95, 125.25, 123.85, 123.28, 122.15, 119.20, 114.40, 62.51, 48.15, 39.61. HRMS (ESI) calcd for $\text{C}_{32}\text{H}_{23}\text{N}_5\text{O}_6$ $[\text{M} + \text{H}]^+$ 574.1726, found 574.1737.

2-(3-(4-((4-(3-(4-Cinnamoylphenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12e). Yield-87%, off-white solid, MP = 135–137 °C, ^1H NMR (400 MHz, CDCl_3): δ 2.35–2.40 (m, 2H, CH_2), 4.60–4.64 (m, 2H, CH_2), 4.70–4.74 (m, 2H, CH_2), 5.24 (s, 2H, OCH_2), 7.12 (d, $J = 8.8$ Hz, 2H, ArH), 7.31–7.35 (m, 3H, ArH), 7.38 (d, $J = 15.6$ Hz, 1H, olefinic H), 7.56 (d, $J = 8.5$ Hz, 2H, ArH), 7.66–7.72 (m, 3H, ArH), 7.76 (s, 1H, triazole-H), 7.96 (d, $J = 8.7$ Hz, 2H, ArH), 8.16 (dd, $J = 1.0, 8.4$ Hz, 2H, ArH), 8.42 (dd, $J = 1.1, 7.3$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 188.65, 163.95, 162.10, 144.10, 134.42, 131.65, 131.57, 131.35, 130.72, 130.30, 129.35, 128.90, 128.40, 128.21, 126.95, 123.41, 121.95, 121.90, 114.71, 114.56, 62.20, 48.15, 38.90, 28.85. HRMS (ESI) calcd for $\text{C}_{33}\text{H}_{26}\text{N}_4\text{O}_4$ $[\text{M} + \text{H}]^+$ 543.2032, found 543.2045.

(E)-2-(3-(4-((4-(3-(4-Methoxyphenyl)acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12f). Yield-86%, off white solid, MP = 151–153 °C, ^1H NMR (400 MHz, CDCl_3): δ 2.37–2.43 (m, 2H, CH_2), 3.80 (s, 3H, OCH_3), 4.61–4.65 (m, 2H, CH_2), 4.72–4.75 (m, 2H, CH_2), 5.21 (s, 2H, OCH_2), 6.89 (d, $J = 8.7$ Hz, 2H, ArH), 6.95 (d, $J = 8.9$ Hz, 2H, ArH), 7.35 (d, $J = 15.5$ Hz, 1H, olefinic H), 7.51 (d, $J = 8.5$ Hz, 2H,

ArH), 7.65–7.72 (m, 3H, ArH), 7.75 (s, 1H, triazole-H), 7.95 (d, $J = 8.8$ Hz, 2H, ArH), 8.15 (dd, $J = 1.0, 8.5$ Hz, 2H, ArH), 8.42 (dd, $J = 1.1, 7.3$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 188.95, 164.10, 161.92, 161.61, 143.98, 143.75, 134.50, 131.76, 131.69, 131.60, 130.85, 130.40, 128.30, 127.91, 127.12, 123.51, 122.10, 119.51, 114.62, 114.45, 62.16, 56.01, 48.10, 39.45, 28.89. HRMS (ESI) calcd for $\text{C}_{34}\text{H}_{28}\text{N}_4\text{O}_5$ $[\text{M} + \text{H}]^+$ 573.2138, found 573.2152.

(E)-2-(3-(4-((4-(3-(2,5-Dimethoxyphenyl)acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12g). Yield-85%, bright yellow solid, MP = 142–144 °C, ^1H NMR (400 MHz, CDCl_3): δ 2.37–2.43 (m, 2H, CH_2), 3.79 (s, 3H, OCH_3), 3.84 (s, 3H, OCH_3), 4.25 (t, $J = 6.7$ Hz, 2H, CH_2), 4.48 (t, $J = 7.0$ Hz, 2H, CH_2), 5.24 (s, 2H, OCH_2), 6.84 (d, $J = 9.0$ Hz, 1H, ArH), 6.90 (dd, $J = 3.0, 9.0$ Hz, 1H, ArH), 7.05 (d, $J = 8.7$ Hz, 2H, ArH), 7.12 (d, $J = 2.9$ Hz, 1H, ArH), 7.56 (d, $J = 15.8$ Hz, 1H, olefinic H), 7.72–7.75 (m, 2H, ArH), 7.87 (s, 1H, triazole-H), 8.00 (d, $J = 8.7$ Hz, 2H, ArH), 8.02 (d, $J = 15.8$ Hz, 1H, olefinic H), 8.20 (d, $J = 8.2$ Hz, 2H, ArH), 8.57 (d, $J = 7.3$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 189.39, 164.43, 161.95, 153.52, 153.34, 143.44, 139.53, 134.43, 131.78, 131.67, 131.58, 130.93, 128.20, 127.13, 124.67, 123.33, 122.95, 122.35, 117.11, 114.67, 113.80, 112.49, 62.16, 56.19, 55.92, 48.49, 37.57, 28.99. HRMS (ESI) calcd for $\text{C}_{35}\text{H}_{30}\text{N}_4\text{O}_6$ $[\text{M} + \text{H}]^+$ 603.2243, found 603.2255.

(E)-2-(3-(4-((4-(3-(4-nitrophenyl)acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (12h). Yield-85%, bright yellow solid, MP = 140–142 °C, ^1H NMR (400 MHz, CDCl_3): δ 2.36–2.42 (m, 2H, CH_2), 4.26 (t, $J = 6.6$ Hz, 2H, CH_2), 4.45 (t, $J = 7.0$ Hz, 2H, CH_2), 5.25 (s, 2H, OCH_2), 6.94 (d, $J = 8.7$ Hz, 2H, ArH), 7.35 (d, $J = 8.8$ Hz, 2H, ArH), 7.41 (d, $J = 15.4$ Hz, 1H, olefinic H), 7.64 (d, $J = 8.7$ Hz, 2H, ArH), 7.71–7.75 (m, 3H, ArH), 7.75 (s, 1H, triazole-H), 8.12 (d, $J = 8.8$ Hz, 2H, ArH), 8.16 (dd, $J = 1.0, 8.0$ Hz, 2H, ArH), 8.45 (dd, $J = 1.1, 7.2$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 189.15, 164.12, 161.64, 143.90, 143.68, 141.28, 134.48, 131.92, 131.75, 131.61, 130.65, 130.21, 128.35, 127.90, 125.20, 123.81, 123.25, 122.10, 119.25, 114.41, 62.50, 48.20, 39.60, 28.90. HRMS (ESI) calcd for $\text{C}_{33}\text{H}_{25}\text{N}_5\text{O}_6$ $[\text{M} + \text{H}]^+$ 588.1883, found 588.1896.

2-(2-(4-((4-(1-Acetyl-5-phenyl-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13a). Yield-84%, light brown solid, MP = 82–84 °C, ^1H NMR (400 MHz, CDCl_3): δ 2.38 (s, 3H, CH_3), 2.94 (dd, $J = 4.5, 17.4$ Hz, 1H, CH_2), 3.62 (dd, $J = 10.9, 17.3$ Hz, 1H, CH_2), 4.65–4.68 (m, 2H, CH_2), 4.75–4.78 (m, 2H, CH_2), 5.21 (s, 2H, OCH_2), 5.56 (dd, $J = 4.5, 10.5$ Hz, 1H, CH), 7.12 (d, $J = 8.8$ Hz, 2H, ArH), 7.31–7.35 (m, 3H, ArH), 7.54 (d, $J = 8.5$ Hz, 2H, ArH), 7.65–7.69 (m, 2H, ArH), 7.75 (s, 1H, triazole-H), 7.94 (d, $J = 8.8$ Hz, 2H, ArH), 8.16 (dd, $J = 1.1, 8.4$ Hz, 2H, ArH), 8.39 (dd, $J = 1.0, 7.3$ Hz, 2H, ArH). ^{13}C NMR (101 MHz, CDCl_3) δ 168.75, 164.10, 159.95, 159.10, 153.65, 134.60, 134.25, 131.70, 131.65, 128.31, 128.20, 127.10, 126.95, 124.61, 123.45, 122.05, 115.10, 114.30, 62.10, 55.40, 48.10, 41.75, 39.60, 22.05. HRMS (ESI) calcd for $\text{C}_{34}\text{H}_{28}\text{N}_6\text{O}_4$ $[\text{M} + \text{H}]^+$ 585.2250, found 585.2272.

2-(2-(4-((4-(1-Acetyl-5-(4-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13b). Yield-85%, bright yellow solid, MP = 85–87 °C, ^1H NMR (400 MHz, CDCl_3): δ 2.37 (s, 3H, CH_3), 3.09 (dd, $J = 4.4, 17.6$ Hz, 1H, CH_2), 3.66 (dd, $J = 11.7,$



17.6 Hz, 1H, CH₂), 3.74 (s, 3H, OCH₃), 4.64–4.67 (m, 2H, CH₂), 4.74–4.77 (m, 2H, CH₂), 5.20 (s, 2H, OCH₂), 5.51 (dd, *J* = 4.4, 11.6 Hz, 1H, CH), 6.81 (d, *J* = 8.7 Hz, 2H, ArH), 6.97 (d, *J* = 8.9 Hz, 2H, ArH), 7.14 (d, *J* = 8.6 Hz, 2H, ArH), 7.64 (d, *J* = 8.9 Hz, 2H, ArH), 7.69–7.73 (m, 2H, ArH), 7.76 (s, 1H, triazole-H), 8.19 (dd, *J* = 1.0, 8.4 Hz, 2H, ArH), 8.50 (dd, *J* = 1.1, 7.3 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.74, 164.06, 159.99, 159.06, 153.68, 134.52, 134.27, 131.72, 131.68, 128.31, 128.25, 127.08, 126.99, 124.64, 123.45, 122.08, 115.13, 114.30, 62.13, 59.39, 55.35, 48.05, 42.44, 39.72, 22.05. HRMS (ESI) calcd for C₃₅H₃₀N₆O₅ [M + H]⁺ 615.2356, found 615.2368.

2-(2-(4-((4-(1-Acetyl-5-(2,5-dimethoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13c). Yield-84%, yellow solid, MP = 78–80 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.38 (s, 3H, CH₃), 2.94 (dd, *J* = 4.4, 17.5 Hz, 1H, CH₂), 3.60 (dd, *J* = 11.6, 17.5 Hz, 1H, CH₂), 3.69 (s, 3H, OCH₃), 3.80 (s, 3H, OCH₃), 4.25 (t, *J* = 6.7 Hz, 2H, CH₂), 4.47 (t, *J* = 7.0 Hz, 2H, CH₂), 5.20 (s, 2H, OCH₂), 5.76 (dd, *J* = 4.5, 11.5 Hz, 1H, CH), 6.54 (d, *J* = 3.0 Hz, 1H, ArH), 6.71 (dd, *J* = 2.9, 8.7 Hz, 1H, ArH), 6.76 (d, *J* = 8.7 Hz, 1H, ArH), 6.98 (d, *J* = 8.6 Hz, 2H, ArH), 7.64 (d, *J* = 8.7 Hz, 2H, ArH), 7.74–7.78 (m, 2H, ArH), 7.85 (s, 1H, triazole-H), 8.23 (d, *J* = 8.1 Hz, 2H, ArH), 8.60 (d, *J* = 7.3 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.76, 164.40, 159.85, 154.50, 153.71, 150.25, 143.60, 134.40, 131.70, 131.56, 130.61, 128.25, 128.20, 127.15, 124.75, 123.20, 122.35, 114.95, 112.45, 112.15, 111.85, 62.10, 56.12, 55.75, 55.40, 48.45, 41.65, 38.01, 22.05. HRMS (ESI) calcd for C₃₆H₃₂N₆O₆ [M + H]⁺ 645.2461, found 645.2475.

2-(2-(4-((4-(1-Acetyl-5-(4-nitrophenyl)-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13d). Yield-86%, light brown solid, MP = 94–96 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.40 (s, 3H, CH₃), 3.09 (dd, *J* = 4.9, 17.0 Hz, 1H, CH₂), 3.77 (dd, *J* = 12.0, 17.6 Hz, 1H, CH₂), 4.64–4.67 (m, 2H, CH₂), 4.74–4.77 (m, 2H, CH₂), 5.21 (s, 2H, OCH₂), 5.61 (dd, *J* = 4.8, 11.9 Hz, 1H, CH), 6.99 (d, *J* = 8.8 Hz, 2H, ArH), 7.38 (d, *J* = 8.7 Hz, 2H, ArH), 7.64 (d, *J* = 8.8 Hz, 2H, ArH), 7.71–7.75 (m, 2H, ArH), 7.77 (s, 1H, triazole-H), 8.16 (d, *J* = 8.7 Hz, 2H, ArH), 8.21 (dd, *J* = 0.7, 8.2 Hz, 2H, ArH), 8.51 (dd, *J* = 0.8, 7.2 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.99, 164.10, 160.26, 153.52, 149.03, 147.40, 143.81, 134.62, 131.74, 131.72, 128.37, 128.31, 127.12, 126.77, 124.40, 123.93, 123.51, 122.04, 115.22, 62.09, 59.40, 48.05, 42.21, 39.74, 21.96. HRMS (ESI) calcd for C₃₄H₂₇N₇O₆ [M + H]⁺ 630.2101, found 630.2115.

2-(3-(4-((4-(1-Acetyl-5-phenyl-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13e). Yield-87%, light brown solid, MP = 78–80 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.35–2.40 (m, 5H, CH₃ + CH₂), 2.95 (dd, *J* = 4.8, 17.9 Hz, 1H, CH₂), 3.61 (dd, *J* = 11.1, 17.8 Hz, 1H, CH₂), 4.65–4.68 (m, 2H, CH₂), 4.74–4.77 (m, 2H, CH₂), 5.25 (s, 2H, OCH₂), 5.56 (dd, *J* = 4.7, 11.5 Hz, 1H, CH), 7.12 (d, *J* = 8.7 Hz, 2H, ArH), 7.32–7.36 (m, 3H, ArH), 7.55 (d, *J* = 8.5 Hz, 2H, ArH), 7.65–7.68 (m, 2H, ArH), 7.76 (s, 1H, triazole-H), 7.95 (d, *J* = 8.7 Hz, 2H, ArH), 8.15 (dd, *J* = 0.9, 8.5 Hz, 2H, ArH), 8.40 (dd, *J* = 0.8, 7.5 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.72, 164.20, 159.90, 159.20, 153.60, 134.62, 134.26, 131.75, 131.66, 128.30, 128.21, 127.12, 126.91, 124.60, 123.41, 122.06, 115.12,

114.35, 62.12, 55.41, 48.15, 41.70, 39.42, 29.15, 22.09. HRMS (ESI) calcd for C₃₅H₃₀N₆O₄ [M + H]⁺ 599.2407, found 599.2419.

2-(3-(4-((4-(1-Acetyl-5-(4-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13f). Yield-85%, light brown solid, MP = 82–84 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.36–2.40 (m, 5H, CH₃ + CH₂), 3.10 (dd, *J* = 4.4, 17.5 Hz, 1H, CH₂), 3.65 (dd, *J* = 11.6, 17.5 Hz, 1H, CH₂), 3.75 (s, 3H, OCH₃), 4.63–4.66 (m, 2H, CH₂), 4.75–4.78 (m, 2H, CH₂), 5.21 (s, 2H, OCH₂), 5.50 (dd, *J* = 4.5, 11.3 Hz, 1H, CH), 6.82 (d, *J* = 8.6 Hz, 2H, ArH), 6.96 (d, *J* = 8.8 Hz, 2H, ArH), 7.15 (d, *J* = 8.5 Hz, 2H, ArH), 7.65 (d, *J* = 8.9 Hz, 2H, ArH), 7.70–7.75 (m, 2H, ArH), 7.77 (s, 1H, triazole-H), 8.20 (dd, *J* = 1.1, 8.5 Hz, 2H, ArH), 8.48 (dd, *J* = 1.2, 7.4 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.70, 164.10, 159.90, 159.10, 153.65, 134.53, 134.25, 131.75, 131.65, 128.35, 128.20, 127.10, 126.99, 124.65, 123.45, 122.10, 115.20, 114.30, 62.15, 59.40, 55.30, 48.12, 42.40, 39.70, 29.10, 22.04. HRMS (ESI) calcd for C₃₆H₃₂N₆O₅ [M + H]⁺ 629.2512, found 629.2525.

2-(3-(4-((4-(1-Acetyl-5-(2,5-dimethoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13g). Yield-88%, light brown solid, MP = 76–78 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.38–2.42 (m, 5H, CH₂ + CH₃), 2.95 (dd, *J* = 4.5, 17.6 Hz, 1H, CH₂), 3.61–3.66 (m, 1H, CH₂), 3.68 (s, 3H, OCH₃), 3.79 (s, 3H, OCH₃), 4.26 (t, *J* = 6.6 Hz, 2H, CH₂), 4.48 (t, *J* = 7.0 Hz, 2H, CH₂), 5.19 (s, 2H, OCH₂), 5.77 (dd, *J* = 4.4, 11.6 Hz, 1H, CH), 6.55 (d, *J* = 2.9 Hz, 1H, ArH), 6.70 (dd, *J* = 2.9, 8.8 Hz, 1H, ArH), 6.78 (d, *J* = 8.8 Hz, 1H, ArH), 6.97 (d, *J* = 8.7 Hz, 2H, ArH), 7.63 (d, *J* = 8.7 Hz, 2H, ArH), 7.73–7.77 (m, 2H, ArH), 7.84 (s, 1H, triazole-H), 8.22 (d, *J* = 8.1 Hz, 2H, ArH), 8.58 (d, *J* = 7.3 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.75, 164.44, 159.88, 154.55, 153.72, 150.23, 143.62, 134.45, 131.68, 131.59, 130.62, 128.27, 128.23, 127.14, 124.79, 123.21, 122.36, 114.98, 112.41, 112.14, 111.87, 62.09, 56.11, 55.71, 55.42, 48.48, 41.70, 37.58, 29.02, 22.04. HRMS (ESI) calcd for C₃₇H₃₄N₆O₆ [M + H]⁺ 659.2618, found 659.2627.

2-(3-(4-((4-(1-Acetyl-5-(4-nitrophenyl)-4,5-dihydro-1H-pyrazol-3-yl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)propyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (13h). Yield-85%, bright yellow solid, MP = 86–88 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.35–2.40 (m, 5H, CH₂ + CH₃), 3.10 (dd, *J* = 5.2, 16.9 Hz, 1H, CH₂), 3.76 (dd, *J* = 11.9, 16.8 Hz, 1H, CH₂), 4.63–4.66 (m, 2H, CH₂), 4.75–4.78 (m, 2H, CH₂), 5.23 (s, 2H, OCH₂), 5.60 (dd, *J* = 5.1, 11.8 Hz, 1H, CH), 6.95 (d, *J* = 8.7 Hz, 2H, ArH), 7.35 (d, *J* = 8.4 Hz, 2H, ArH), 7.64 (d, *J* = 8.8 Hz, 2H, ArH), 7.70–7.76 (m, 2H, ArH), 7.76 (s, 1H, triazole-H), 8.15 (d, *J* = 8.3 Hz, 2H, ArH), 8.20 (dd, *J* = 0.8, 8.1 Hz, 2H, ArH), 8.52 (dd, *J* = 0.9, 7.3 Hz, 2H, ArH). ¹³C NMR (101 MHz), δ 168.95, 164.20, 160.20, 153.50, 149.10, 147.38, 143.80, 134.62, 131.75, 131.70, 128.40, 128.30, 127.20, 126.71, 124.45, 123.90, 123.50, 122.10, 115.20, 62.10, 59.40, 48.10, 42.25, 38.01, 29.10, 21.95. HRMS (ESI) calcd for C₃₅H₂₉N₇O₆ [M + H]⁺ 644.2257, found 644.2265.

(E)-2-(2-(4-((4-(3-Ferrocenyl-acryloyl)phenoxy)methyl)-1H-1,2,3-triazol-1-yl)ethyl)-1H-benzo[de]isoquinoline-1,3(2H)-dione (14a). Yield-85%, red solid, MP = 220–222 °C, ¹H NMR (400 MHz, CDCl₃): δ 4.18 (s, 5H, FeH), 4.47 (s, 2H, FeH), 4.60 (s, 2H, FeH), 4.69 (t, *J* = 6.5 Hz, 2H, CH₂), 4.80 (t, *J* = 7.0 Hz, 2H, CH₂),



5.26 (s, 2H, OCH₂), 7.03 (d, *J* = 8.5 Hz, 2H, ArH), 7.13 (d, *J* = 15.1 Hz, 1H, olefinic H), 7.72–7.76 (m, 3H, ArH + olefinic H), 7.78 (s, 1H, triazole-H), 7.97 (d, *J* = 8.5 Hz, 2H, ArH), 8.22 (dd, *J* = 0.9, 8.1 Hz, 2H, ArH), 8.50 (dd, *J* = 0.8, 7.3 Hz, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ 188.20, 164.45, 161.72, 146.15, 143.41, 134.45, 131.89, 131.65, 131.50, 130.65, 128.20, 127.25, 123.30, 122.30, 118.80, 114.61, 79.54, 71.30, 69.92, 69.10, 62.31, 48.45, 39.12. HRMS (ESI) calcd for C₃₆H₂₈FeN₄O₄ [M + H]⁺ 637.1538, found 637.1549.

(*E*)-2-(2-(4-((4-(3-Ferrocenyl-acryloyl)phenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)propyl)-1*H*-benzo[de]isoquinoline-1,3(2*H*)-dione (**14b**). Yield-85%, brick red solid, MP = 210–212 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.37–2.44 (m, 2H, CH₂), 4.15 (s, 5H, FeH), 4.26 (t, *J* = 6.7 Hz, 2H, CH₂), 4.45 (t, *J* = 1.8 Hz, 2H, FeH), 4.49 (t, *J* = 7.1 Hz, 2H, CH₂), 4.56 (t, *J* = 1.8 Hz, 2H, FeH), 5.24 (s, 2H, OCH₂), 7.05 (d, *J* = 8.8 Hz, 2H, ArH), 7.11 (d, *J* = 15.3 Hz, 1H, olefinic H), 7.69–7.77 (m, 3H, ArH + olefinic H), 7.87 (s, 1H, triazole-H), 7.98 (d, *J* = 8.8 Hz, 2H, ArH), 8.21 (dd, *J* = 0.7, 8.2 Hz, 2H, ArH), 8.58 (dd, *J* = 0.8, 7.2 Hz, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ 188.18, 164.44, 161.78, 146.01, 143.46, 134.44, 131.87, 131.67, 131.59, 130.69, 128.21, 127.14, 123.29, 122.36, 118.85, 114.64, 79.43, 71.32, 69.85, 69.02, 62.16, 48.49, 37.57, 29.01. HRMS (ESI) calcd for C₃₇H₃₀FeN₄O₄ [M + H]⁺ 651.1694, found 651.1685.

2-(2-(4-((4-(1-Acetyl-5-ferrocenyl-4,5-dihydro-1*H*-pyrazol-3-yl)-phenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)ethyl)-1*H*-benzo[de]isoquinoline-1,3(2*H*)-dione (**15a**). Yield-85%, light brown solid, MP = 198–200 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.26 (s, 3H, CH₃), 3.45 (dd, *J* = 4.8, 17.3 Hz, 1H, CH₂), 3.56 (dd, *J* = 11.6, 17.6 Hz, 1H, CH₂), 3.94–3.96 (m, 2H, CH₂), 4.07–4.12 (m, 7H, FeH), 4.20–4.24 (m, 2H, CH₂), 4.47 (s, 2H, FeH), 5.25 (s, 2H, OCH₂), 5.45 (dd, *J* = 4.6, 11.5 Hz, 1H, CH), 7.05 (d, *J* = 8.1 Hz, 2H, ArH), 7.73–7.78 (m, 4H, ArH), 7.86 (s, 1H, triazole-H), 8.21 (d, *J* = 7.7 Hz, 2H, ArH), 8.56 (d, *J* = 6.8 Hz, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ 168.82, 164.42, 159.95, 153.81, 143.60, 134.40, 131.75, 131.60, 128.30, 128.20, 127.14, 124.70, 123.26, 122.35, 115.16, 87.42, 70.30, 68.69, 68.27, 62.10, 55.30, 48.45, 39.65, 37.56, 22.20. HRMS (ESI) calcd for C₃₈H₃₂FeN₄O₄ [M + H]⁺ 693.1912, found 693.1925.

2-(2-(4-((4-(1-Acetyl-5-ferrocenyl-4,5-dihydro-1*H*-pyrazol-3-yl)-phenoxy)methyl)-1*H*-1,2,3-triazol-1-yl)propyl)-1*H*-benzo[de]isoquinoline-1,3(2*H*)-dione (**15b**). Yield-85%, light brown solid, MP = 190–192 °C, ¹H NMR (400 MHz, CDCl₃): δ 2.27 (s, 3H, CH₃), 2.39–2.42 (m, 2H, CH₂), 3.47 (d, *J* = 15.5 Hz, 1H, CH₂), 3.59 (dd, *J* = 11.6, 16.3 Hz, 1H, CH₂), 3.97 (s, 1H, CH₂), 4.07–4.11 (m, 7H, FeH), 4.26 (s, 2H, CH₂), 4.46–4.49 (m, 3H, FeH + CH₂), 5.23 (s, 2H, OCH₂), 5.43 (d, *J* = 8.5 Hz, 1H, CH), 7.05 (d, *J* = 8.0 Hz, 2H, ArH), 7.72–7.77 (m, 4H, ArH), 7.87 (s, 1H, triazole-H), 8.22 (d, *J* = 7.8 Hz, 2H, ArH), 8.58 (d, *J* = 6.8 Hz, 2H, ArH). ¹³C NMR (101 MHz, CDCl₃) δ 168.81, 164.46, 159.99, 153.83, 143.61, 134.47, 131.69, 131.60, 128.25, 128.23, 127.16, 124.72, 123.28, 122.36, 115.17, 87.45, 70.35, 68.66, 68.27, 62.13, 55.29, 48.50, 39.68, 37.58, 29.03, 22.11. HRMS (ESI) calcd for C₃₉H₃₄FeN₄O₄ [M + H]⁺ 707.1991, found 707.1985.

Materials and methods

Cell culturing

MCF7 cells were cultured in DMEM media with 10% FBS, and 1% penicillin–streptomycin and MDA-MB-231 cells were

cultured in DMEM supplemented with 5% FBS and 1% penicillin–streptomycin, both incubated at 37 °C and 5% carbon dioxide.

MTT assay

Cells were seeded in 96 well plates at a density of 5000 cells per well in triplicate in media. After 24 hours, the test compounds diluted in complete Dulbecco's media Eagle's medium (DMEM) were added to each well. Cells were treated with a range of different concentrations of the compounds (1, 5, 10, 20, 50, 100 μM) for 24 hours at 37 °C and 5% carbon dioxide. Subsequently, sterile 5 μl of 5 mg mL⁻¹ MTT (Sigma-Aldrich) dissolved in PBS was added to each well and incubated with cells for 2 hours. Solubilization solution (10% SDS, 10 mM HCl) of equal volume to the wells was then added to each well, which was incubated with cells for 16 hours at 37 °C. The optical density of each well was read at 570 nm using a microtiter plate reader (Thermo Fisher Scientific Multiskan GO Microplate Reader, SkanIt™ software).

Statistical analysis

The statistical analysis was performed using Excel®, and IC₅₀ values were estimated using GraphpadPrism5 software (Hearne Scientific Software). The experiments were performed in duplicate, and the statistical significance was calculated using the student's *t*-test. A *p*-value of less than 0.05 was used to estimate the significance of the observations. A Z-factor was calculated for each 96-well plate and assays having Z-factor above >0.5 were included in the statistical analysis.³⁹

Molecular docking

Molecular docking was carried out using Auto dock Vina⁴⁰ and the crystal structures of human estrogen receptor alpha (ERα, PDB code 3ERT, 1.9 Å resolution) and estrogen receptor beta (ERβ, PDB code 3OLS, 2.2 Å resolution). The compatibility between the residue and the moderate active ligands was further checked using Protein–ligand Interaction Profiler software. SwissADME⁴¹ and pkCSM⁴² web-based applications were used to predict the ADMET properties of active compounds.

Abbreviations

ER	Estrogen receptor
SERMs	Selective estrogen receptor modulators
SAR	Structure activity relationship

Conflicts of interest

The authors declare no conflict of interest.

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Notes and references

- 1 Cancer Facts and Figures 2019, American Cancer Society, <https://www.cancer.org/research/cancer-facts-statistics/all-cancer-facts-figures/cancer-facts-figures-2019.html>.
- 2 <https://www.who.int/cancer/prevention/diagnosis-screening/breast-cancer/en/>.
- 3 W. F. Anderson, N. Chatterjee, W. B. Ershler and O. W. Brawley, *Breast Cancer Res. Treat.*, 2002, **76**, 27.
- 4 B. H. Mitlak and F. J. Cohen, *Drugs*, 1999, **57**, 653.
- 5 M. N. Gomes, E. N. Muratov, M. Pereira, J. C. Peixoto, L. P. Rosseto, P. V. L. Cravo, C. H. Andrade and B. J. Neves, *Molecules*, 2017, **22**, 1210.
- 6 D. Israf, T. Khaizurina, A. Syahida, N. Lajis and S. Khozirah, *Mol. Immunol.*, 2007, **44**, 673.
- 7 T. Yamamoto, M. Yoshimura, F. Yamaguchi, T. Kouchi, R. Tsuji, M. Saito, A. Obata and M. Kikuchi, *Biosci., Biotechnol., Biochem.*, 2004, **68**, 1706.
- 8 N. Aoki, M. Muko, E. Ohta and S. Ohta, *J. Nat. Prod.*, 2008, **71**, 1308.
- 9 M. Chen, S. B. Christensen, J. Blom, E. Lemmich, L. Nadelmann, K. Fich, T. G. Theander, A. Kharazmi and A. Licochalcone, *Antimicrob. Agents Chemother.*, 1993, **37**, 2550.
- 10 P. M. Sivakumar, S. Ganesan, P. Veluchamy and M. Doble, *Chem. Biol. Drug Des.*, 2010, **76**, 407.
- 11 H. Suwito, J. Mustofa, P. Pudjiastuti, M. Z. Fanani, Y. Kimata-Ariga, R. Katahira, T. Kawakami, T. Fujiwara, T. Hase, H. M. Sirat and N. N. T. Puspaningsih, *Molecules*, 2014, **19**, 21473.
- 12 P. Boeck, P. C. Leal, R. A. Yunes, V. Cechinel, S. Lopez, M. Sortino, A. Escalante, R. L. E. Furlan and S. Zacchino, *Arch. Pharm.*, 2005, **338**, 87.
- 13 S. Cheenpracha, C. Karalai, C. Ponglimanont, S. Subhadhirasakul and S. Tewtrakul, *Bioorg. Med. Chem.*, 2006, **14**, 1710.
- 14 S. Burmaoglu, O. Algul, D. Aktas, A. Gobek and G. Gulbol, *Bioorg. Med. Chem. Lett.*, 2016, **26**, 3172.
- 15 S. Ducki, *Adv. Anticancer Agents Med. Chem.*, 2009, **9**, 336.
- 16 V. R. Solomon and H. Lee, *Biomed. Pharmacother.*, 2012, **66**, 213.
- 17 I. K. Lindamulage, H.-Y. Vu, C. Karthikeyan, J. Knockleby, Y.-F. Lee, P. Trivedi and H. Lee, *Sci. Rep.*, 2017, **7**, 10298.
- 18 A. Modzelewska, C. Pettit, G. Achanta, N. E. Davidson, P. Huang and S. R. Khan, *Bioorg. Med. Chem.*, 2006, **14**, 3491.
- 19 H.-H. Gong, D. Addla, J.-S. Lv and C.-H. Zhou, *Curr. Top. Med. Chem.*, 2016, **16**, 3303.
- 20 T. Brider, B. Redko, F. Grynspan and G. Gellerman, *Tetrahedron Lett.*, 2014, **55**, 6675.
- 21 R. Seliga, M. Pilatova, M. Sarissky, V. Viglasky, M. Walko and J. Mojzis, *Mol. Biol. Rep.*, 2013, **40**, 4129.
- 22 E. Diaz-Rubio, M. Martin, J. M. López-Vega, A. Casado and A. Benavides, *Invest. New Drugs*, 1994, **12**, 277.
- 23 J. Noro, J. Maciel, D. Duarte, A. C. D. Olival, C. Baptista, A. C. D. Silva, M. J. Alves and P. Kong Thoo Lin, *Org. Chem.: Curr. Res.*, 2015, **4**, 144.
- 24 M. F. Braña, M. Cacho, M. A. Garcia, B. de Pascual-Teresa, A. Ramos, M. T. Dominguez, J. M. Pozuelo, C. Abradelo, M. F. Rey-Stolle, M. Yuste and M. Banez-Coronel, *J. Med. Chem.*, 2004, **47**, 1391.
- 25 X. Liang, K. Xu, Y. Xu, J. Liu and X. Qian, *Toxicol. Appl. Pharmacol.*, 2011, **256**, 52.
- 26 X. Liang, Y. Xu, K. Xu, J. Liu and X. Qian, *Mol. Cancer Res.*, 2010, **8**, 1619.
- 27 S. Nagarajan, P. Shanmugavelan, M. Sathishkumar, R. Selvi, A. Ponnuswamy, H. Harikrishnan and V. Shanmugaiah, *Chin. Chem. Lett.*, 2014, **25**, 419.
- 28 M. W. Pertino, C. Theoduloz, E. Butassi, S. Zacchino and G. Schmeda-Hirschmann, *Molecules*, 2015, **20**, 8666.
- 29 L. Zhao, L. Mao, G. Hong, X. Yang and T. Liu, *Bioorg. Med. Chem. Lett.*, 2015, **25**, 2540.
- 30 N. R. Penthala, L. Madhukuri, S. Thakkar, N. R. Madadi, G. Lamture, R. L. Eoff and P. A. Crooks, *MedChemComm*, 2015, **6**, 1535.
- 31 N. Pokhodylo, O. Shyyka and V. Matyichuk, *Sci. Pharm.*, 2013, **81**, 663.
- 32 S. Kumar, S. T. Saha, L. Gu, G. Palma, S. Perumal, A. S. Pillay, P. Singh, A. Anand, M. Kaur and V. Kumar, *ACS Omega*, 2018, **3**, 12106.
- 33 B. Sharma, A. Singh, L. Gu, S. T. Saha, A. S. Pillay, N. Cele, P. Singh, M. Kaur and V. Kumar, *RSC Adv.*, 2019, **9**, 9809.
- 34 S. Kumar, G. Palma, S. Perumal, M. Kaur, A. S. Pillay, R. Raj, P. Singh and V. Kumar, *RSC Adv.*, 2019, **9**, 42409.
- 35 A. Rani, G. I. Singh, R. Kaur, M. Kaur, G. Palma, S. Perumal, O. Ebenezer, P. Awolade, P. Singh and V. Kumar, *J. Organomet. Chem.*, 2020, **907**, 121072.
- 36 M. Karabacak, M. D. Altıntop, H. İ. Çiftçi, R. Koga, M. Otsuka, M. Fujita and A. Özdemir, *Molecules*, 2015, **20**, 19066.
- 37 H. Wang, J. Zheng, W. Xu, C. Chen, D. Wei, W. Ni and Y. Pan, *Molecules*, 2017, **22**, 1635.
- 38 S. Sagar, L. Esau, B. Moosa, N. M. Khashab, V. B. Bajic and M. Kaur, *Adv. Anticancer Agents Med. Chem.*, 2014, **14**, 170.
- 39 J. H. Zhang, T. D. Y. Chung and K. R. Oldenburg, *J. Biomol. Screen*, 1999, **4**, 67.
- 40 O. Trott and A. J. Olson, *J. Comput. Chem.*, 2009, **31**, 455.
- 41 <http://www.swissadme.ch/>.
- 42 <http://biosig.unimelb.edu.au/pkcsm/prediction>.

