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A simple fluorescent probe for detection of Ag⁺ and Cd²⁺ and its Cd²⁺ complex for sequential recognition of S²⁻†

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In this study, we designed and synthesized a simple probe 2-(8-((8-methoxyquinolin-2-yl)methoxy) quinolin-2-yl)benzo[d]thiazole (DQT) for detection of Ag $^+$ and Cd $^{2+}$ in a CH $_3$ OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system. Its structure was characterized by NMR, ESI-HR-MS and DFT calculations, and its fluorescence performance was also investigated. Probe DQT showed fluorescence quenching in response to Ag $^+$ and Cd $^{2+}$ with low detection limits of 0.42 μ M and 0.26 μ M, respectively. Importantly, the complexation of the probe with Cd $^{2+}$ resulted in a red shift from blue to green, making it possible to detect Ag $^+$ and Cd $^{2+}$ by the naked eye under an ultraviolet lamp. The DQT-Cd $^{2+}$ complex could be used for sequential recognition of S $^{2-}$. The recovery response could be repeated 3 times by alternate addition of Cd $^{2+}$ and S $^{2-}$. A filter paper strip test further demonstrated the potential of probe DQT as a convenient and rapid assay.

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Introduction

Cd²⁺ and Ag⁺ ions are important transition metal ions involved in various biological and environmental processes.¹⁻⁴ However, excessive intake of Ag⁺ can cause serious harm to human health as it can inactivate thiolase and even have a carcinogenic effect.^{5,6} Cd²⁺ is one of the most serious air pollutants, and it is also highly toxic, causing serious health problems even at low concentrations.⁷⁻¹⁰ It is reported that Cd²⁺ ions are associated with the pathogenesis of many diseases including cancers.¹¹⁻¹⁴ Thus, it is necessary to detect Cd²⁺ and Ag⁺ in the environment. However, traditional methods such as capacity analysis,¹⁵ ICP-MS¹⁶ and atomic absorption method,¹⁷ may be greatly limited due to requirement of equipment, high cost, cumbersome operation and low accuracy.¹⁸ In comparison, the fluorescence detection method has advantages of high specific recognition ability, high sensitivity and short response time, making it

Two mechanisms namely PET and ICT are inclusively used as the basis for the design of fluorescent probes. ^{24–26} PET refers to the process of fluorescence quenching or enhancement caused by electron transfer between the excited electron donor or electron acceptor after photoexcitation. ²⁷ In addition, the recognition groups of ICT probes are usually a part of the pushpull electron system, and thus the combination of these groups with metal ions may affect the push-pull electron effect of the fluorophore and the intramolecular charge transfer, thus leading to changes in fluorescence spectrum. ²⁸

Many fluorescent probes have been developed based on the two mechanisms for the detection of a variety of ions, such as Cu^{2+}/Cd^{2+} , 29,30 Zn^{2+}/Cd^{2+} , $^{31-33}$ and $Cu^{2+}/Cd^{2+}/PPi$. 34 However, there are fewer probes for the detection of $Ag^+/Cd^{2+}/S^{2-}$. In this work, a simple fluorescent probe **DQT** was successfully synthesized based on quinolone³⁵⁻³⁸ and benzothiazole,³⁹⁻⁴¹ which showed high selectivity toward Ag^+ and Cd^{2+} in $CH_3OH/HEPES$ (9:1 v/v, pH = 7.30), and its Cd^{2+} complex could be

Scheme 1 Synthesis of probe DQT.

useful for the detection of many metal ions such as Hg^{2+} , Pb^{2+} , Cu^{2+} and Zn^{2+} . $^{19-23}$

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[†] Electronic supplementary information (ESI) available: Appendix A includes the Experimental section, ¹H NMR spectra, ¹³C NMR spectra, ESI-MS spectra and part of spectrums of probe **DQT**; Appendix B includes the detail crystal data, structure refinement, torsion angles and bond lengths and angles of probe **DQT**. CCDC 1975779. For ESI and crystallographic data in CIF or other electronic format see DOI: 10.1039/d0ra01768j

OH N S + O N Br K2CO3 N S DQT

used for sequential recognition of S^{2-} . The paper strip test was carried out to verify its practicability (Scheme 1).

Results and discussion

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X-ray structural analysis probe DQT

As shown in Fig. 1, the crystal structure without hydrogen atom of probe **DQT** was given. The cell parameters were a = 21.767(3) Å, $\alpha = 90$ deg; b = 4.8042(8) Å, $\beta = 103.866(7)$ deg; c = 42.042(7) Å, $\gamma = 90$ deg. The detail crystal data, structure refinement, torsion angles and bond lengths and angles for probe **DQT** were shown in Tables S1–S3 of Appendix B.† More information has been uploaded to the Cambridge Crystallographic Date Center (CCDC 1975779).

The fluorescence response of probe DQT towards Ag^+ and Cd^{2+}

To study the fluorescence selectivity of probe **DQT** toward metal ions, the fluorescence spectra of probe **DQT** upon the addition of 20 equiv. of K⁺, Na⁺, Zn²⁺, Cu²⁺, Hg²⁺, Pb²⁺, Ba²⁺, Cd²⁺, Cs⁺,

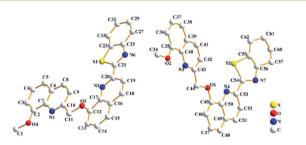


Fig. 1 The crystal structure without hydrogen atom of probe DQT.

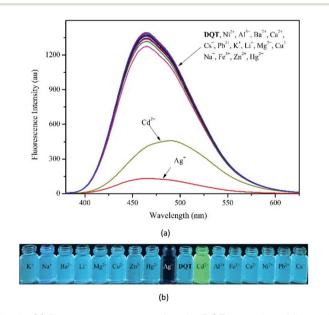


Fig. 2 (a) Fluorescence spectra of probe DQT upon the addition of various metal cations (2 \times 10 $^{-4}$ mol L $^{-1}$) and blank. (b) Color changes of probe DQT with various metal cations in CH $_3$ OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system under UV light.

Li⁺, Ag⁺, Mg²⁺, Ca²⁺, Al³⁺, Ni²⁺ and Fe³⁺ were recorded in CH₃OH/HEPES (9 : 1 v/v, pH = 7.30) (λ_{ex} = 344 nm). As shown in Fig. 2(a), no distinct changes were observed in fluorescence spectra of probe **DQT** ($\Phi = 0.308$) upon the containing of metal cations except that with the presence of Ag⁺ ($\Phi = 0.002$) and Cd^{2+} ($\Phi = 0.227$). Upon the addition of different metal ions, Ag^{+} caused a 88% fluorescence quenching and displayed an "onoff" behavior. With Cd2+, probe DQT showed a 33% fluorescence quenching with a spectral red-shift of 25 nm from 465 to 490 nm. This could be attributed to changes in the electronically conjugated structure of the molecule after the combination of the lone pair of electrons of N atoms with Cd2+. In addition, due to the special molecular configuration and electronic structure of the compound, complex DQT-Cd2+ was not interfered by other ions. Apparent color changes were observed, indicating that probe DQT could be used to detect Ag⁺ and Cd²⁺ by naked eye (Fig. 2(b) and (c)).

Competition experiments were performed to explore the effects of other metal ions on the recognition of Ag⁺ and Cd²⁺. It could be seen in Fig. 3(a) that there was significant change in fluorescence intensity after the addition of 20 equiv. of Ag⁺. Clearly, the recognition of Ag⁺ was affected by these metal ions. However, the recognition of Cd²⁺ was not affected especially Ag⁺ and the fluorescence color was still green under UV light, as

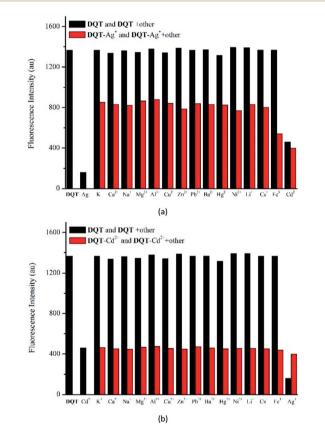


Fig. 3 Competitive selectivity of probe DQT (10 μ M) toward Ag⁺ (a) and Cd²⁺ (b) (20 equiv.) in the absence (black bars) or presence (red bars) of specified ions (20 equiv.).

shown in Fig. 3(b). Compared to Ag^+ , probe **DQT** had higher selectivity for Cd^{2+} .

To better understand the sensitivity of probe **DQT** toward Ag^+ and Cd^{2+} , the fluorescent titration of Ag^+ and Cd^{2+} was carried out in CH₃OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system at an excitation wavelength of 344 nm. As shown in Fig. 4(a), probe

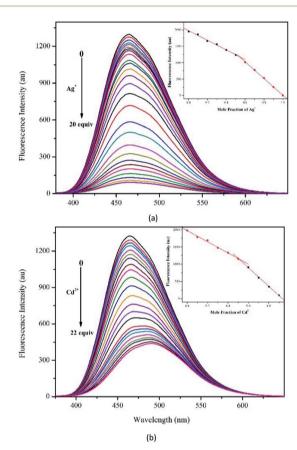


Fig. 4 The fluorescence intensity of probe DQT (10 μ M) in CH₃OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system upon incremental addition of Ag⁺ (a) and Cd²⁺ (b). Job's curves were shown at the upper right corner.

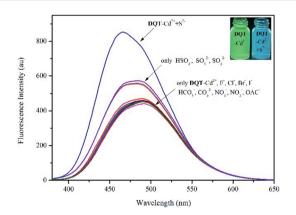


Fig. 5 Fluorescence spectra of DQT-Cd²⁺ (1×10^{-5} mol L⁻¹) with the addition of various anions (20 equiv.) in CH₃OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system.

DQT exhibited an emission at 465 nm with an enormous Stokes shift of 121 nm. Upon incremental addition of Ag+, the fluorescence emission gradually decreased until a plateau was reached at 20 equiv. of Ag⁺. The limit of detection (LOD) was determined to be 0.42 μ M based on LOD = $3\sigma/m$ (Fig. S6†). As shown in Fig. 4(b), the addition of Cd²⁺ resulted in a red shift of the fluorescence emission of probe DQT to 490 nm, and probe **DQT** showed fluorescence quenching with a LOD of 0.26 μM for Cd²⁺ (Fig. S7†). The stoichiometry determined by the Job's method was 1: 1 for both DQT-Ag⁺ and DQT-Cd²⁺ complexes, as shown at the upper right corner of Fig. 4(a) and (b). The complexation constant of probe **DQT** to Ag^+ ($K_a = 2.68 \times 10^3$ M^{-1}) (Fig. S4,† $Y = 1.75 \times 10^{-7} \times X - 4.69 \times 10^{-4}, R^2 = 0.9953$) obtained according to the Benesi-Hildebrand plots was lower than that to Cd^{2+} ($K_a = 2.23 \times 10^4 \text{ M}^{-1}$) (Fig. S6,† $Y = 3.91 \times 10^4 \text{ M}^{-1}$) $10^{-8} \times X + 8.72 \times 10^{-4}$, $R^2 = 0.9967$). The titration results were consistent with the interference results, indicating that probe **DQT** had a better sensitivity for Cd²⁺ than Ag⁺.

Fluorescence recognition of DQT-Cd²⁺ complex to S²⁻

In the interference test, the detection of Ag^+ was easily affected by other metal ions, and thus we focused on the response of Cd^{2+} and anions, including S^{2-} , HSO_3^- , SO_3^{2-} , SO_4^{2-} , Br^- , F^- , CO_3^{2-} , Cl^- , OAc^- , HCO_3^- , I^- , NO_2^- and NO_3^- in $CH_3OH/HEPES$ (9:1 v/v, pH = 7.30) buffer system ($\lambda_{ex}=344$ nm). In Fig. 5, only S^{2-} showed obvious fluorescence recovery with an enhancement efficiency of 64% and a blue shift from 490 to 465 nm. Competition experiments were performed to explore the effects of other anions on the recognition of S^{2-} . As exhibited in Fig. S8,† significant fluorescence recovery was observed upon the addition of 20 equiv. of S^{2-} . Obviously, these anions had no effect on the recognition of S^{2-} , especially sulfurcontaining anions. Thus, \mathbf{DQT} - \mathbf{Cd}^{2+} showed high specificity for S^{2-} .

The fluorescence titration with S^{2-} was recorded in $CH_3OH/HEPES$ (9 : 1 v/v, pH = 7.30) buffer system ($\lambda_{ex} = 344$ nm). In Fig. 6, the intensity of the strongest fluorescence emission peak gradually increased and blue-shifted as the S^{2-} concentration

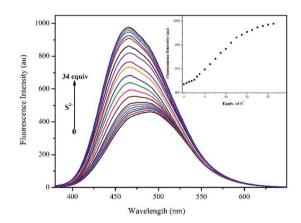


Fig. 6 Fluorescence spectra of DQT-Cd²⁺ (10 μ M) with S²⁻ (0–34 equiv.) in CH₃OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system (λ_{ex} = 344 nm). Inset: the plot of fluorescence intensity DQT-Cd²⁺ versus S²⁻.

was increased to 34 equiv., after which it remained relatively constant. The LOD was calculated to be 11.50 µM by eqn (1)

(Fig. S9,† $Y = 21.99 \times X + 420.56$, $R^2 = 0.9983$).

Reversibility of probe DQT for Cd2+ and S2-

The reversibility was determined by alternating addition of Cd^{2+} and S^{2-} . As illustrated in Fig. 7, the fluorescence intensity was quenched to 33% and the fluorescence colour changed from blue to green observed by naked eye with the addition of Cd^{2+} . However, the addition of S^{2-} resulted in the recovery of the fluorescence intensity to about 64% and fluorescence colour change to blue. As the number of cycles increased, the recognition performance decreased gradually but still remained high after 3 cycles.

Effect of pH

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As shown in Fig. 8, probe **DQT** exhibited excellent sequence selectivity and a red-shift at pH = 4–8 with the addition of Cd²⁺, and the fluorescence was replied with the addition of S²⁻. The selectivity of probe **DQT** was affected under highly acidic or alkaline conditions, probably due to changes in the structure of the probe and the loss of the coordination with ions.

DFT study

Gaussian 09 program was used for the density functional theory calculation at the level of B3LYP/6-31G (d, p) and B3LYP/LANL2DZ to gain a further insight into the photophysical properties and the nature of optical response of DQT, DQT-Ag⁺ and DQT-Cd²⁺. Fig. 9 showed the optimized structures and the electron dispersion of HOMO and LUMO. The molecular structure of probe DQT was planar, and most electrons in HOMO and LUMO were distributed on the 8-methoxyquinoline and quinolinylbenzothiazole groups, respectively. Its planarity was destroyed and the dihedral angles of quinolinobenzothiazole and 8-methoxyquinoline groups were 10° and 17° after coordinating with Ag⁺ and Cd²⁺, respectively. The electrons in HOMO were transferred to the binding site with Ag⁺, which

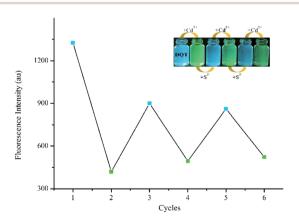


Fig. 7 Reversible changes in fluorescence intensity of probe DQT (10 μ M) in CH₃OH/HEPES (9 : 1 v/v, pH = 7.30) buffer system upon alternate addition of Cd²⁺ and S²⁻.

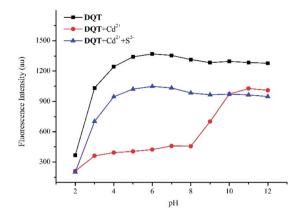


Fig. 8 Influences of the fluorescence intensity of probe DQT (10 μ M) toward Cd²⁺ (20 equiv.) and S²⁻ (20 equiv.) at pH = 2–12 in CH₃OH/HEPES (9 : 1 v/v) buffer system at room temperature.

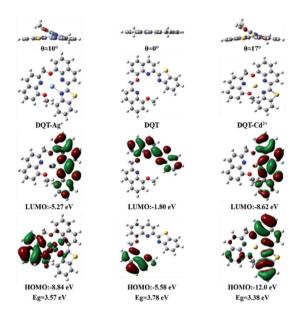


Fig. 9 Molecular orbital and electronic contributions of the relevant excitations.

promoted the PET process and quenched the fluorescence. Similarly, the electrons in HOMO and LUMO were mainly concentrated on the quinolinylbenzothiazole group after coordination with Cd²⁺, resulting in a red shift. The energy gaps of **DQT**, **DQT**-Ag⁺ and **DQT**-Cd²⁺ were 3.78 eV, 3.57 eV and 3.38 eV, respectively, indicating that the binding with Cd²⁺ was more stable.

The possible mechanism

The possible mechanism is proposed as follows. The stoichiometry of the $\mathbf{DQT}\text{-}\mathbf{Ag}^{^{+}}$ and $\mathbf{DQT}\text{-}\mathbf{Cd}^{2+}$ complex is determined to be 1:1. Given the fluorescence quenching and no red shift of emission wavelength, the binding mode of probe \mathbf{DQT} and $\mathbf{Ag}^{^{+}}$ conforms to the PET process. The coordination mechanism of probe \mathbf{DQT} and \mathbf{Cd}^{2+} can be attributed to ICT because the fluorescence emission wavelength is red-shifted after

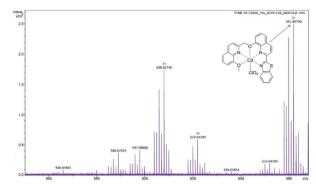


Fig. 10 ESI-MS spectra of DQT-Cd²⁺

coordination with Cd^{2^+} . 34,42 Moreover, the molecular ion peak (ESI-MS) at 661.96730 [DQT + Cd^{2^+} + ClO_4^- ; m/z calcd for 661.97167] (Fig. 10) indicates an 1 : 1 complexation between probe DQT and Cd^{2^+} , and thus it can be concluded that all nitrogen and oxygen atoms of the molecule participate in coordination. Due to the strong interaction between S^{2^-} and Cd^{2^+} , the overall fluorescence is recovered after the addition of S^{2^-} , suggesting that S^{2^-} can form CdS precipitate with Cd^{2^+} and remove Cd^{2^+} from DQT- Cd^{2^+} complex. In summary, the Scheme 2 manifested the possible coordination mode.

Practical application on test strips

Filter paper strip test was carried out to determine the practicability of probe **DQT**. Filter paper strips were immersed in $CH_3OH/HEPES$ (9:1 v/v, pH = 7.30) buffer system of **DQT** at a concentration of 1×10^{-3} M for 30 min and dried in air, and then smeared with Ag^+ and Cd^{2+} solution respectively. The fluorescence changes were observed under UV light. Fig. 11 showed that the fluorescence color of the test strip which treated with Cd^{2+} changed from blue (Fig. 11(b)) to green (Fig. 11(c)), while the fluorescence was quenched after treated



Scheme 2 The possible mechanism



Fig. 11 Color changes on test paper (a) DQT-Ag $^+$; (b) DQT; (c) DQT-Cd $^{2+}$ under UV light.

with Ag⁺ (Fig. 11(a)), suggesting that probe **DQT** could be used to distinguish for Ag⁺ and Cd²⁺ by naked eye under UV light.

Conclusions

Herein, probe **DQT** was successfully synthesized for recognition of multiple ions. It showed fluorescence quenching towards Ag^+ based on the PET mechanism, but the complexation constant was low and the recognition of Ag^+ could be interfered by other ions. Probe **DQT** could also be used to detect Cd^{2+} based on the ICT mechanism with a red shift of 25 nm of the fluorescence emission wavelength in the same $CH_3OH/HEPES$ (9 : 1 v/v, pH = 7.30) buffer system. The **DQT**- Cd^{2+} complex could be used for recognition of S^{2-} . Both Ag^+ and Cd^{2+} could form complexes with probe **DQT** with a coordination ratio of 1 : 1. Finally, filter paper strip test verified its practicability.

Conflicts of interest

There are no conflicts to declare.

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