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# Development of a novel Maillard reaction-based time–temperature indicator for monitoring the fluorescent AGE content in reheated foods

Bei Hu,<sup>a</sup> Lin Li,<sup>ab</sup> Yi Hu,<sup>a</sup> Di Zhao,<sup>a</sup> Yuting Li,<sup>b</sup> Mingquan Yang,<sup>c</sup> Aijuan Jia,<sup>c</sup> Sui Chen,<sup>c</sup> Bing Li<sup>\*a</sup> and Xia Zhang <sup>\*a</sup>

Dietary advanced glycation end products (AGEs) are formed *via* the Maillard reaction in foods, especially in reheated foods, and can cause chronic diseases. In this study, a series of Maillard reaction-based time–temperature indicators (TTIs: TTI-1, TTI-2, and TTI-3) were developed with lysine and xylose for monitoring the dynamic formation of fluorescent AGEs in reheated foods. The discoloration kinetics of Maillard reaction-based TTIs and the dynamics of fluorescent AGE formation were explored. Formulas were derived to illustrate the relationship of the color change in the TTIs with time and temperature. The activation energies ( $E_a$  values) for generating the TTIs were 96.17, 87.98, and 83.55 kJ mol<sup>−1</sup>, respectively. TTI-1 was the optimal indicator for instant soy milk powder because it showed the lowest activation energy difference in this study. The results show that this series of Maillard reaction-based TTIs can be used to monitor the AGE contents in various reheated foods.

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## 1 Introduction

Advanced glycation end products (AGEs), known as glycotoxins, can direct processes that cause chronic diseases and related complications. The AGEs present in ingested food (dietary AGEs) contribute significantly to AGE levels in serum and organs after absorption in the small intestine.<sup>1</sup> Several animal and clinical studies have reported that absorbed dietary AGEs may be hazardous because of their ability to combine with their receptors (RAGEs) *in vivo*.<sup>2–4</sup> Dietary AGEs, which can accumulate *in vivo*, are associated with the generation and development of chronic diseases such as diabetes, cardiovascular disease and collagen disease.<sup>2,5–8</sup> It is more necessary for customers suffering diabetes and kidney dysfunction to reduce the dietary AGE intake.

The formation of AGEs in food is mainly attributed to the Maillard reaction; thus, factors such as temperature,<sup>9</sup> time,<sup>10</sup> ingredients,<sup>11,12</sup> pH,<sup>13,14</sup> and water activity,<sup>15</sup> which influence the Maillard reaction, can affect the formation of AGEs. Choosing the right kinds of food and changing processing conditions are effective methods to reduce the ingestion of dietary AGEs.<sup>16</sup>

At present, the most common methods for detecting AGEs in food products are gas or liquid chromatography and enzyme-linked immunosorbent assay (ELISA).<sup>17</sup> Some databases are available to help consumers select commercially available foods that have lower AGE contents.<sup>18,19</sup> However, these database cannot offer comprehensive information about AGEs in common foods, and the AGE levels in consumed foods are still unpredictable and indistinct. In particular, foods may be reprocessed by customers, such as by reheating or stewing, which would induce the formation of AGEs. Heat is a common means to reprocess purchased foods, such as soymilk and canned mixed congee. In this case, consumers (especially those with diabetes and kidney dysfunction) require a more convenient, visual and effective way to monitor AGEs in foods reheated at home.

Time–temperature indicators (TTIs) are simple, inexpensive devices that can be attached to a package surface for the real-time monitoring of the quality of foods or drugs.<sup>20,21</sup> TTIs show measurable and time–temperature dependent changes and can provide the temperature change history or indirectly indicate the actual quality of a product.<sup>21</sup> Generally, when visible response of a TTI (such as a color change or mechanical deformation) matches food quality parameters in the same temperature environment, the food quality can be predicted by the TTI changes.<sup>22</sup> TTIs greatly enhance consumer confidence in the quality of purchased foods due to their convenient, effective and observable characteristics.<sup>23</sup> Based on their different mechanisms, TTIs are generally categorized as diffusion-based,<sup>24</sup> chemical (polyreaction, photochromic reaction, redox

<sup>a</sup>School of Food Science and Engineering, Guangdong Province Key Laboratory for Green Processing of Natural Products and Product Safety, South China University of Technology, Guangzhou 510640, China. E-mail: bli@scut.edu.cn; cexzhang@scut.edu.cn; Tel: +86-20-8711-3252

<sup>b</sup>School of Chemical Engineering and Energy Technology, Dongguan University of Technology, College Road 1, Dongguan, 523808, China

<sup>c</sup>Guangdong MeiWeiXian Flavoring Foods Ltd. Co., Zhongshan, 528437, China



reaction, *etc.*),<sup>21</sup> enzymatic,<sup>25,26</sup> biological (microbial),<sup>27</sup> electronic<sup>28</sup> and so on.

Recently, Maillard reaction-based TTIs have been widely applied in monitoring food quality due to their good performance in warning of temperature increases in chilled foods and the growth of microbes in food and food materials.<sup>29–31</sup> In addition, Lee, Harada, Kawamura, & Koseki<sup>32</sup> used D-ribose (7.0 mol kg<sup>−1</sup>) and L-lysine (3.0 mol kg<sup>−1</sup>) to develop a Maillard reaction-based TTI to ensure microbiological safety during cooking processes. These studies have demonstrated the feasibility of the Maillard reaction in establishing TTIs. Bosch, Alegria, Farré, & Clemente<sup>33</sup> reported that fluorescent AGEs and color formation are markers of the advanced and final stages of the Maillard reaction. Both the brown compounds detected by the Maillard TTI and fluorescent AGEs are products of the Maillard reaction, suggesting the possibility of a Maillard reaction-based TTI for monitoring fluorescent AGEs in food. However, to the best of our knowledge, Maillard reaction-based TTIs of fluorescent AGEs in foods are currently scarce.

Therefore, in this study, a series of Maillard reaction-based TTIs (TTI-1, TTI-2, TTI-3) for monitoring AGEs were established using reducing sugars and amino acids. The kinetics of the TTI color change and fluorescent AGE formation in several kinds of food were explored. TTIs were investigated using instant soy milk as a model system to determine their practicality. This study may offer customers an easier option for determining the levels of AGEs in food during heat treatments (reheating, stewing, *etc.*). In addition, this work helps broaden the applicability of Maillard reaction-based TTIs and provides a theoretical basis for subsequent commercial applications.

## 2 Materials and methods

### 2.1 Materials

L-Lysine, L-arginine, glycine, L-alanine, D-(+)-glucose, D-fructose and D-(+)-galactose were purchased from Aladdin Bio-Chem Technology Co., Ltd. (Shanghai, China). Disodium phosphate dodecahydrate and sodium phosphate dibasic dihydrate were purchased from Sinopharm Chemical Reagent Co., Ltd. (Beijing, China). Instant soy milk powder was purchased from a supermarket in Canton (Guangzhou, China).

### 2.2 Establishment of the model Maillard reaction-based TTI

Four amino acids (glycine, alanine, serine, and lysine) and three common reducing sugars (glucose, xylose, and galactose) were selected to construct the model TTIs. Solutions of amino acid and reducing sugar (both at 0.25 M) were combined and then mixed in a water bath (HH-4, Changzhou Aohua Instrument Co., Ltd., China) at 80 °C for 60 min. The absorbance of the solution at a wavelength of 420 nm,<sup>34,35</sup> which reflects the major colored products of the Maillard reaction, was measured with a multimode reader (Infinite M200 Pro, Tecan, Switzerland) at 10 min intervals.

To determine the optimal concentration of materials for the model TTI, different concentrations of amino acids and reducing sugars were investigated. Then, quantitative amounts

of the amino acid and reducing sugar were dissolved in a series of 0.2 M PBS with pH values of 6.0, 6.5, 7.0, 7.5 and 8.0 (FiveEasy, Mettler Toledo, America) to determine the optimal pH of the reaction solution. All the mixed solutions were placed in a water bath at 80 °C for 30 min, and the UV absorbance was measured at 6 min intervals.

### 2.3 Determination of dynamic parameters of the model TTI

The model TTI was placed in a water bath at 70, 80, 90, 95 and 100 °C separately. The UV absorbance was monitored at a short intervals and recorded until the absorbance value went up to 1.2. Triplicate measurements were performed.

The relation between the response variable and apparent reaction rate constant (response value growth rate) of the model TTI can be expressed as follow:<sup>20</sup>

$$F(X) = kdT \quad (1)$$

where  $X$  is TTI response variable,  $F(X)$  is the function of TTI response variable,  $k$  is the Maillard reaction rate constant and  $t$  is the reaction time (heat-up period).

The color changes in Maillard reaction process can be characterized by UV absorbance. In this experiment, the TTI response variable  $X$  is the absorbance ( $A$ ) of the model TTI. With the linear regression of the  $A$  value to  $t$ , the constants,  $k$  values at different reaction temperatures can be calculated as the slopes of straight lines numerically.

The temperature dependence chemical reaction should be described by the Arrhenius formula, so does the Maillard reaction of TTI.<sup>36</sup> The activation energy of the model TTI can be gotten with the following formula:

$$k = k_0 \exp(-E_{a,TTI}/RT) \quad (2)$$

where  $k_0$  is the constant named pre-exponential factor,  $R$  is the molar gas constant (8.314 J (mol K)<sup>−1</sup>),  $T$  is the thermodynamic temperature (K), and  $E_{a,TTI}$  is the activation energy (kJ mol<sup>−1</sup>) of TTI.

The eqn (2) can be transformed into the following one:

$$\ln k = \ln k_0 - E_{a,TTI}/RT \quad (3)$$

With the linear regression analysis of the  $\ln k$  value to against  $1/T$ ,  $E_{a,TTI}$  and  $k_0$  can be calculated.  $E_{a,TTI}$  is equal to the slope of straight line numerically.

### 2.4 Case study: application of TTI in instant soy milk powder

Instant soy milk powder, which was identified as having a high AGEs content,<sup>37</sup> was selected as the model system for this work. The fluorescent AGEs in instant soy milk powder were detected according to a modified version of the method described by Ferrer *et al.*<sup>9</sup> Pronase (1 mL, 35 U mL<sup>−1</sup>) was added to 2 mL of instant soy milk powder solution (protein content: 0.02–0.05 g mL<sup>−1</sup>) and heated at 30 °C for 90 min to terminate enzymolysis. Then, the samples were maintained at 70, 80, 90, 95 and 100 °C in a water bath, respectively, for 15 min prior to analysis. All fluorometric experiments were conducted using a multimode



reader (Infinite M200 Pro, Tecan, Switzerland) at 3 min intervals. The excitation wavelength ( $\lambda_{\text{ex}}$ ) and the emission wavelength ( $\lambda_{\text{em}}$ ) of fluorescent AGEs were 370 nm and 440 nm, respectively.<sup>38,39</sup> The excitation and emission slit widths were set to 9.0 nm and 20.0 nm, respectively. The scan times were 25.

**2.4.1 Analysis of dynamic parameters and  $E_a$  of fluorescent AGEs.** The quality changes of food with time can be described by the following equation:<sup>40</sup>

$$Q(A) = k_{\text{food}} t \quad (4)$$

where  $Q(A)$  represents for flavor, color, nutrition, microorganisms or toxic component of food,  $k_{\text{food}}$  is the apparent reaction rate constant of food and related to temperature, and  $t$  is time.

In this study,  $Q(A)$  can be apparently characterized by the fluorescence intensity of AGEs. With the linear regression of scatter plot of fluorescence intensity of total fluorescence AGEs, the constant  $k_{\text{food}}$  at different reaction temperature can be calculated. Then, according to Arrhenius formula,  $E_a$  of fluorescent AGEs in instant soy milk powder can also be calculated.

**2.4.2 Prediction the content of fluorescent AGEs in instant soy milk powder by TTI.** The instant soy milk and suitable TTI were stored with the same temperature increasing from 70 °C to 100 °C in 15 min. The fluorescence intensity of samples and the color of TTI were monitored at 3 min intervals with a recorder. With the linear regression of the fluorescence intensity to the absorbance of TTI, the relationship between TTI color and AGEs level of sample could be demonstrated.

## 2.5 Statistical analysis

The averages, standard deviations, reaction constant  $k$  and  $E_a$  were calculated for each sample with Microsoft Excel. Regression analysis and curve fitting of scatter diagram were carried out with Origin V8.0. Data were analyzed using SPSS 22.0 software, one-way analyses of variance with Duncan's multiple range tests were performed to test if statistically significant differences existed for absorbance in reaction system of

different amino acids and reducing sugars release data at  $p < 0.05$ .

## 3 Results and discussion

### 3.1 Determination of the model TTI

The Maillard reaction is an intricate interaction between carbonyl compounds (mainly reducing sugars) and amino compounds (amino acids, peptides and proteins), ultimately resulting in melanoidins.<sup>41</sup> Melanoidins are a class of brown substances that can absorb 420 nm ultraviolet light.<sup>42,43</sup> The absorbances of melanoidins (at 420 nm) derived from amino acids (glycine, alanine, serine, and lysine) and reducing sugars (glucose, xylose, and galactose) are presented in Table 1. The UV absorbance of Groups 1, 2, 3, 4, 5 and 6 increased gradually with incubation time ( $p < 0.05$ ), and Group 1 (Xly + Lys) showed the strongest absorbance (Table 1). Kwak & Lim<sup>12</sup> also found that among the 60 Maillard reaction products from combinations of 12 amino acids and 5 reducing sugars, the xylose-glycated sample showed the highest UV absorbance. In the Maillard reaction, pentoses were more active than hexose and lysine, which also includes  $\epsilon$ -amidogen, resulting in more strongly colored products.<sup>44–46</sup> The UV absorptions of Groups 7, 8, 9, 10, 11 and 12 (without xylose or lysine) were close to zero. In addition, the color changes in Group 1 could be recognized by the naked eye. Therefore, lysine and xylose were selected as the materials for the model TTI in this study.

To determine the optimal concentrations of xylose and lysine for the model TTI, the effects of the xylose and lysine concentrations on the absorbance changes of the colored products (melanoidin) were assessed, and the results are shown in Fig. 1. As the concentrations of xylose and lysine were increased and the reaction time was prolonged, the overall absorbances rose (Fig. 1a and b). When the concentration of lysine was higher than 0.3 M, the absorbance changed substantially with time. However, the reaction rate plateaued after 24 min in a solution with 0.5 M lysine (Fig. 1a). The color change in the mixture should be easily recognized

**Table 1** Changes of absorbance (at 420 nm) in reaction systems with different amino acids and reducing sugars<sup>a</sup>

Group	Solution	0 min	10 min	20 min	30 min	40 min	50 min	60 min
1	Lys + Xyl	0.010 ± 0.001 <sup>ab</sup>	0.124 ± 0.014 <sup>a</sup>	0.394 ± 0.124 <sup>a</sup>	0.737 ± 0.016 <sup>a</sup>	1.314 ± 0.038 <sup>a</sup>	1.906 ± 0.085 <sup>a</sup>	2.257 ± 0.150 <sup>a</sup>
2	Gly + Xyl	0.000 ± 0.002 <sup>c</sup>	0.024 ± 0.009 <sup>c</sup>	0.082 ± 0.014 <sup>c</sup>	0.174 ± 0.002 <sup>c</sup>	0.418 ± 0.015 <sup>b</sup>	0.648 ± 0.004 <sup>b</sup>	1.121 ± 0.003 <sup>b</sup>
3	Lys + Gal	0.015 ± 0.001 <sup>a</sup>	0.056 ± 0.013 <sup>b</sup>	0.237 ± 0.094 <sup>b</sup>	0.358 ± 0.008 <sup>b</sup>	0.441 ± 0.193 <sup>b</sup>	0.563 ± 0.110 <sup>b</sup>	0.616 ± 0.024 <sup>c</sup>
4	Lys + Glu	0.001 ± 0.001 <sup>bc</sup>	0.019 ± 0.019 <sup>cd</sup>	0.026 ± 0.001 <sup>cd</sup>	0.099 ± 0.018 <sup>d</sup>	0.175 ± 0.016 <sup>c</sup>	0.219 ± 0.040 <sup>c</sup>	0.415 ± 0.058 <sup>cd</sup>
5	Ala + Xyl	0.000 ± 0.019 <sup>a</sup>	0.000 ± 0.004 <sup>e</sup>	0.019 ± 0.002 <sup>cd</sup>	0.048 ± 0.003 <sup>e</sup>	0.171 ± 0.003 <sup>b</sup>	0.207 ± 0.063 <sup>c</sup>	0.307 ± 0.008 <sup>de</sup>
6	Ser + Xyl	0.003 ± 0.002 <sup>bc</sup>	0.006 ± 0.005 <sup>de</sup>	0.011 ± 0.003 <sup>cd</sup>	0.042 ± 0.005 <sup>e</sup>	0.128 ± 0.018 <sup>c</sup>	0.162 ± 0.000 <sup>cd</sup>	0.230 ± 0.006 <sup>de</sup>
7	Gly + Gal	0.006 ± 0.001 <sup>ab</sup>	0.006 ± 0.001 <sup>cde</sup>	0.009 ± 0.002 <sup>cd</sup>	0.008 ± 0.004 <sup>f</sup>	0.022 ± 0.004 <sup>c</sup>	0.023 ± 0.001 <sup>de</sup>	0.030 ± 0.001 <sup>e</sup>
8	Ala + Glu	0.004 ± 0.000 <sup>bc</sup>	0.003 ± 0.000 <sup>e</sup>	0.003 ± 0.000 <sup>d</sup>	0.004 ± 0.000 <sup>f</sup>	0.009 ± 0.009 <sup>c</sup>	0.004 ± 0.001 <sup>e</sup>	0.030 ± 0.015 <sup>e</sup>
9	Ala + Gal	0.008 ± 0.002 <sup>ab</sup>	0.010 ± 0.002 <sup>cde</sup>	0.012 ± 0.002 <sup>cd</sup>	0.009 ± 0.000 <sup>f</sup>	0.013 ± 0.001 <sup>c</sup>	0.013 ± 0.000 <sup>de</sup>	0.021 ± 0.001 <sup>e</sup>
10	Ser + Gal	0.006 ± 0.000 <sup>ab</sup>	0.010 ± 0.001 <sup>cde</sup>	0.032 ± 0.003 <sup>cd</sup>	0.011 ± 0.003 <sup>f</sup>	0.010 ± 0.001 <sup>c</sup>	0.018 ± 0.004 <sup>de</sup>	0.018 ± 0.002 <sup>e</sup>
11	Ser + Glu	0.002 ± 0.000 <sup>bc</sup>	0.001 ± 0.001 <sup>e</sup>	0.002 ± 0.001 <sup>d</sup>	0.002 ± 0.001 <sup>f</sup>	0.061 ± 0.008 <sup>c</sup>	0.003 ± 0.001 <sup>e</sup>	0.005 ± 0.001 <sup>e</sup>
12	Gly + Glu	0.000 ± 0.007 <sup>abc</sup>	0.002 ± 0.007 <sup>e</sup>	0.005 ± 0.003 <sup>d</sup>	0.004 ± 0.001 <sup>f</sup>	0.001 ± 0.003 <sup>c</sup>	0.000 ± 0.000 <sup>e</sup>	0.002 ± 0.001 <sup>e</sup>

<sup>a</sup> Gly, Ser, Ala and Lys stand for glycine, serine, alanine and lysine separately; Xyl, Glu and Gal stand for xylose, glucose and galactose separately; the alphabet (a–f) expressed significant difference ( $p < 0.05$ ).



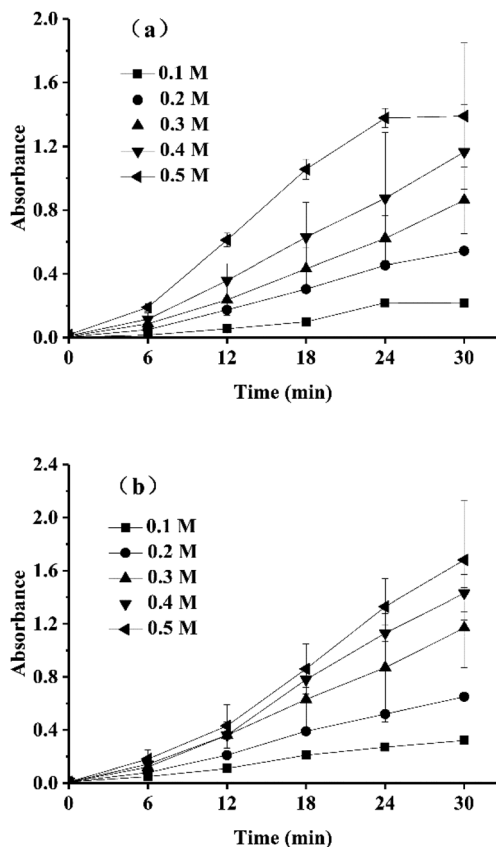


Fig. 1 Change in absorbance (at 420 nm) in reaction system with 0.3 M xylose and different concentrations of lysine (a); change in absorbance (at 420 nm) in reaction system with 0.4 M lysine and different concentrations of xylose (b).

by the naked eye, and this requires that the final absorbance values be greater than 1. Obviously, xylose concentrations of 0.3, 0.4 and 0.5 M were more suitable for establishing the TTI model (Fig. 1b). Thus, the optimal concentration of lysine was 0.4 M, and suitable concentrations of xylose were 0.3, 0.4 and 0.5 M.

Fig. 2 shows the changes in the formation of colored Maillard products at different pH values. There were no significant differences in the absorbances of the mixed solutions at different pH values (6–8). This is contrary to the findings that the nonenzymatic browning process of the Maillard reaction is facilitated by alkaline pH values<sup>47</sup> and inhibited by acidic conditions because the free amidogen of the amino acids would be blocked and the carbonyl-amine condensation products would be easily hydrolyzed.<sup>48–50</sup> This deviation might be due to the relatively low reaction temperature because higher temperatures afford Maillard reaction products with higher absorbances.<sup>47</sup> As shown in Fig. 2, the final absorbance values at all pH levels ranged from 0.9 to 1.4, and these changes were thus all visible to the naked eye. Considering that the model TTI would be used for food, a pH of 7.0 was selected for safety reasons. The terminal absorbance value of the reaction was 1.2.

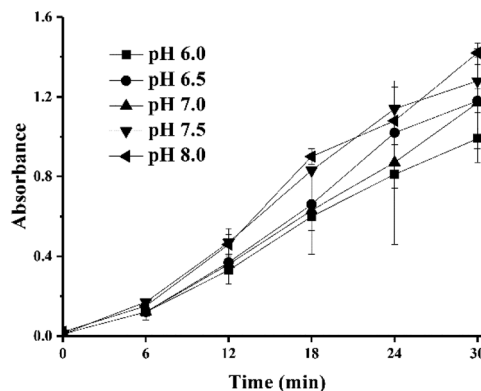


Fig. 2 Effect of pH on the progress of UV absorbance at 420 nm in sample with 0.4 M lysine and 0.4 M of xylose.

Table 2 TTI model with different concentration of xylose

TTI model	Lysine	Xylose	pH value of PBS
TTI-1	0.4 M	0.3 M	7.0
TTI-2	0.4 M	0.4 M	7.0
TTI-3	0.4 M	0.5 M	7.0

To summarize the above experiments, the materials and parameters of the model TTI were determined as follows: 0.4 M lysine and 0.3, 0.4 or 0.5 M xylose and PBS at pH 7.0. When the absorbance at a wavelength of 420 nm increased to 1.2, the reaction was complete. Table 2 shows a series of TTIs with different concentrations of lysine and xylose.

The change in color change in most TTIs can provide information on the cumulative time and temperature history,<sup>51</sup> and the color changes of the TTIs should be easily recognized by the naked eye.<sup>31</sup> Fig. 3 shows the changes in color of TTI-1 at 80 °C within 15 min. During the Maillard reaction, the color gradually changed from light yellow to dark brown, which could be easily recognized by the naked eye. In the Maillard reaction, higher molecular weight colored substances would be produced through the cross-linking of lower molecular weight chromophores and amidogens of amino acids or proteins.<sup>52</sup> Therefore, the colored Maillard products would accumulate, resulting in a darker color as the reaction proceeds. TTI-2 and TTI-3 also showed similar color changes; these results are not shown.

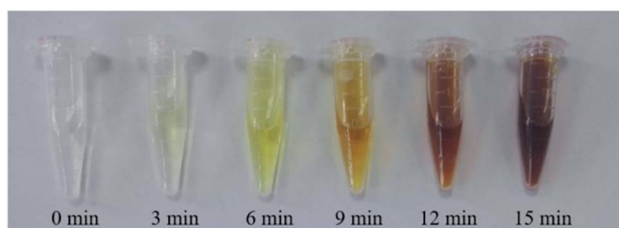


Fig. 3 Color changes of TTI-1 during 15 min of incubation at 80 °C.

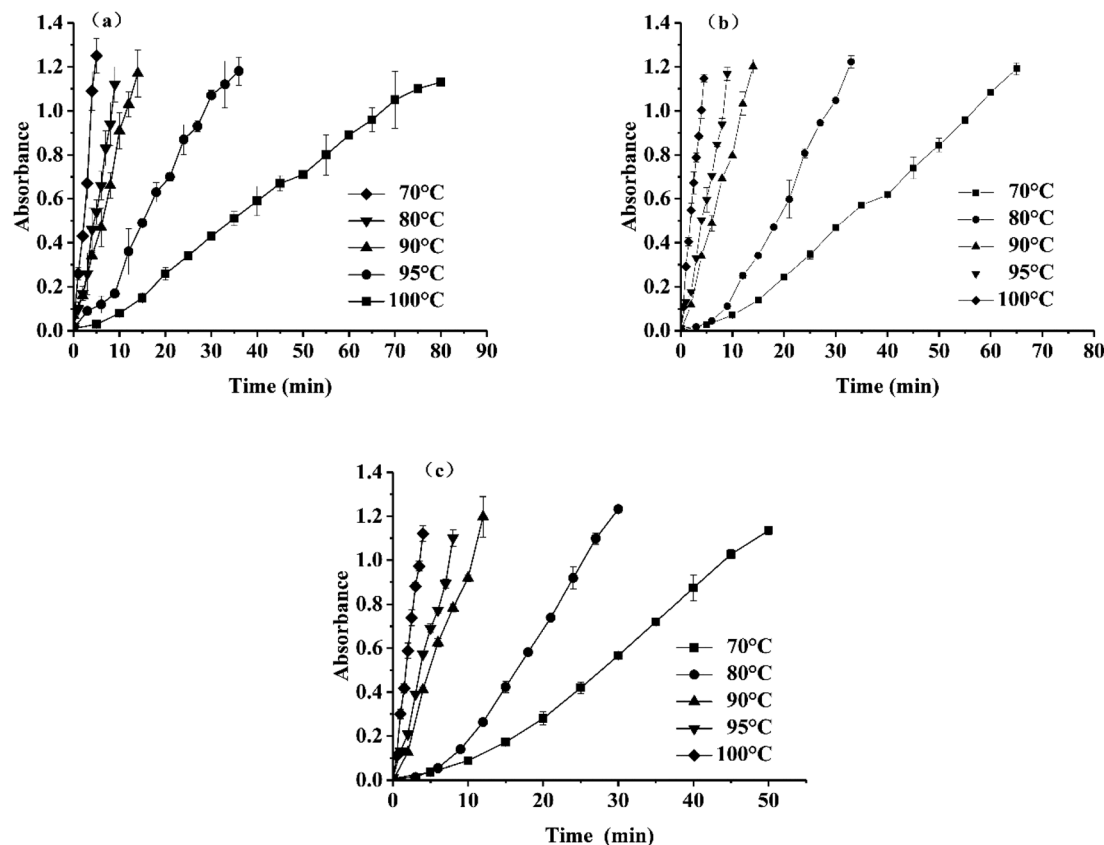


Fig. 4 Change of absorbance of TTIs at different temperature, (a) for TTI-1, (b) for TTI-2, (c) for TTI-3.

### 3.2 Kinetics analysis of the TTI

**3.2.1 The reaction rates at different temperatures.** The absorbances of the TTIs at different heating temperatures are shown in Fig. 4. The absorbances increased gradually with increasing temperature. Most of the data points were located on or close to the regression line, so linear regression equations between the absorbance of the TTIs and time at different

temperatures could be easily obtained and are shown in Table 3. The relation between absorbance ( $X$ ) and time ( $t$ ) can be expressed by eqn (1). The slopes of the fitting curves were the reaction rate constants,  $k$ . For TTI-1, TTI-2 and TTI-3, the reaction rate constants,  $k$  values, increased with increasing temperature. Comparing the reaction rate constants,  $k$  values, of TTI-1, TTI-2 and TTI-3 at the same temperature showed that

Table 3 Equations of linear regression and dynamic parameters for absorbance of TTIs at different temperature

TTIs	$T (^{\circ}\text{C})$	Equations	$k (\text{min}^{-1})$	$R^2$	$1/T \times 1000 (\text{K}^{-1})$	$\ln k$
TTI-1	70	$y = 0.0152x - 0.0370$	0.0152	0.9956	2.92	-4.19
	80	$y = 0.0356x - 0.4490$	0.0356	0.9858	2.83	-3.34
	90	$y = 0.0829x + 0.0203$	0.0829	0.9908	2.75	-2.49
	95	$y = 0.1235x + 0.0165$	0.1235	0.9873	2.72	-2.10
	100	$y = 0.2535x - 0.0134$	0.2535	0.9844	2.68	-1.37
TTI-2	70	$y = 0.0190x - 0.0960$	0.0190	0.9881	2.92	-3.96
	80	$y = 0.0392x - 0.1585$	0.0392	0.9658	2.83	-3.24
	90	$y = 0.0866x - 0.0233$	0.0866	0.9960	2.75	-2.45
	95	$y = 0.1250x - 0.0218$	0.1250	0.9908	2.72	-2.08
	100	$y = 0.2516x - 0.0198$	0.2516	0.9969	2.68	-1.38
TTI-3	70	$y = 0.0242x - 0.1221$	0.0242	0.9744	2.92	-3.72
	80	$y = 0.0442x - 0.1659$	0.0442	0.9645	2.83	-3.12
	90	$y = 0.0991x - 0.0148$	0.0991	0.9906	2.75	-2.31
	95	$y = 0.1346x - 0.0083$	0.1346	0.9918	2.72	-2.01
	100	$y = 0.2839x + 0.0031$	0.2839	0.9966	2.68	-1.26





$k_{\text{TTI-1}} < k_{\text{TTI-2}} < k_{\text{TTI-3}}$ , which was ascribed to the high concentration of xylose in the substrate.

For the various conditions, the fitting curves were approximately linear with correlation coefficients from 0.9844–0.9966, which indicated that the established linear regression equation was valid. Linear regression analysis confirmed a high linear relationship between the changes in the TTI absorbance and time. As derived from eqn (3), the dynamic parameters and the correlation coefficients under different temperatures are shown in Table 3.

**3.2.2 Activation energy ( $E_a$ ) of TTIs.** According to eqn (3), the regression line of  $\ln k$  versus  $1/T$  is shown in Fig. 5, and the activation energy ( $E_a$ ) and pre-exponential factor ( $k_0$ ) can be obtained from the slope and intercept of the linear equation.

Fig. 5 shows the Arrhenius plot of  $\ln k$  vs.  $1/T$ , and a high coefficient correlation for linearity ( $R^2 = 0.9610$ – $0.9826$ ) was obtained in each TTI. Based on the fitting equation in Fig. 5 and eqn (4), the  $E_a$  of TTI-1 is  $96.17 \text{ kJ mol}^{-1}$ , and the  $k_0$  value is  $23 \times 10^{12} \text{ min}^{-1}$ . Taoukis *et al.*<sup>20,53,54</sup> suggested that the activation energy difference between the food being monitored and the TTI should be less than  $25 \text{ kJ mol}^{-1}$  to ensure the relevance of the TTI response. Therefore, TTI-1 can be applied in food systems in which the activation energy of AGE formation ranges from  $71.17 \text{ kJ mol}^{-1}$  ( $96.17 - 25 \text{ kJ mol}^{-1}$ ) to  $121.17 \text{ kJ mol}^{-1}$  ( $96.17 + 25 \text{ kJ mol}^{-1}$ ). Similarly, the  $E_a$  of TTI-2 is  $87.98 \text{ kJ mol}^{-1}$  and  $k_0 = 4.36 \times 10^{11} \text{ min}^{-1}$ ; the  $E_a$  of TTI-3 was  $83.55 \text{ kJ mol}^{-1}$  and  $k_0 = 1.14 \times 10^{11} \text{ min}^{-1}$ . Thus, TTI-2 and TTI-3 are suitable

for food systems with activation energy ranges of  $62.98$ – $112.98 \text{ kJ mol}^{-1}$  and  $58.55$ – $108.55 \text{ kJ mol}^{-1}$ , respectively.

### 3.3 Fluorescent AGEs formation in instant soy milk and color change in the TTI

The color change in the TTI was compared with the formation of fluorescent AGEs in instant soy milk stored at different temperatures. The contents of fluorescent AGEs in instant soy milk at different temperatures are shown in Fig. 6. With increasing temperature, the fluorescence intensity increased

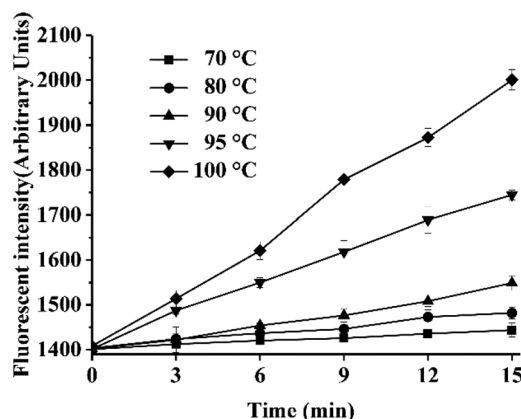


Fig. 6 Content of total fluorescent AGEs in instant soy milk when heated at different temperatures.

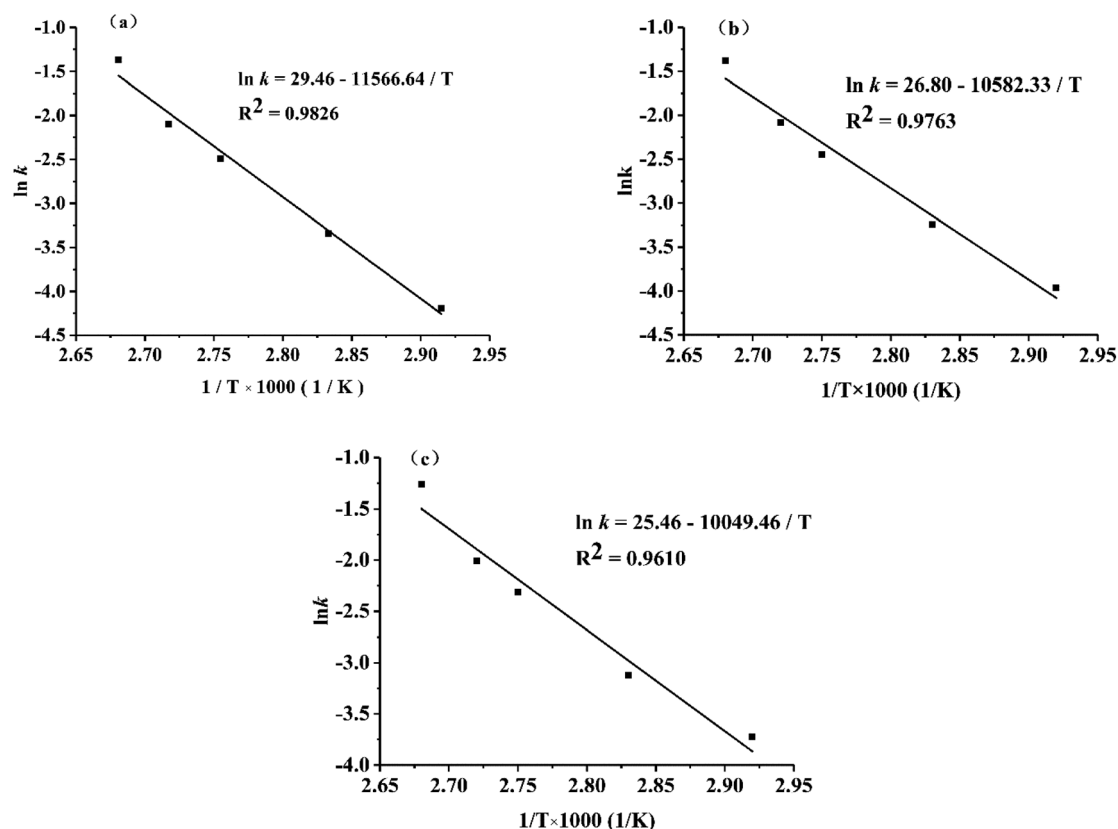


Fig. 5 Arrhenius plot of  $\ln k$  versus  $1/T$  for TTI-1 (a), TTI-2 (b) and TTI-3 (c).



gradually, and the fluorescence intensity changed faster at higher temperatures because the Maillard reaction is a typical endothermic reaction.<sup>41</sup> The changes in AGEs levels in food (Fig. 6) were in accordance with the increased UV absorbance of the TTIs (Fig. 4). Thus, the formation of AGEs could be stimulated by elevated temperatures and increases over time.

Subsequently, the activation energy of fluorescent AGEs formation in soy milk was calculated in the same way as the activation energy of the TTIs. The relationship between the fluorescence intensity of the soy milk powder and time at different temperatures is shown in Table 4. Similarly, as the rate of reaction increased with temperature, the rate constant,  $k$ , of the chemical reaction monotonically increased, and the chemical reaction kinetics of the formation of fluorescent AGEs and the colored Maillard reaction products were consistent. Under the various tested conditions, the fitting curves were approximately linear with very high correlation coefficients ( $R^2 > 0.98$ ), which indicated that the effect of the regression was remarkable and valid. On the basis of eqn (4) and the dynamic parameters ( $k$  values) shown in Table 4, the Arrhenius plot of the fluorescent AGEs in instant soy milk powder is shown in Fig. 7, and the activation energy ( $E_a$ ) of the fluorescent AGEs and the pre-exponential factor ( $k_0$ ) were calculated as  $93.78 \text{ kJ mol}^{-1}$  and  $4.32 \times 10^{14} \text{ min}^{-1}$ , respectively. The activation energy deviations between fluorescent AGE formation in heated soy milk and TTI-1, TTI-2 and TTI-3 are 2.39, 5.80 and  $10.23 \text{ kJ mol}^{-1}$ , respectively. These discrepancies are all less than  $25 \text{ kJ mol}^{-1}$ , indicating that all these TTIs could be used to predict the levels of fluorescent AGEs in instant soy milk during heating. TTI-1 was selected as the optimum system for indicating the levels of fluorescent AGEs in instant soy milk as it showed the smallest difference in activation energy.

The fitting curve of the fluorescence intensity of the AGEs and the UV absorbance of TTI-1 is shown in Fig. 8. The high correlation coefficient ( $R^2 = 0.9822$ ) demonstrates that TTI-1 should accurately indicate the level of fluorescent AGEs in instant soy milk during heating.

According to the color change dependence on time and temperature, a colorimetric card was developed based on the fitting equation to express the total fluorescent AGE level in instant soy milk powder. As shown in Fig. 9, customers can estimate the AGE level in soybean milk by using this colorimetric card.

In addition, a series of TTIs suitable for different kinds of foods have been developed, so this Maillard reaction-type TTI

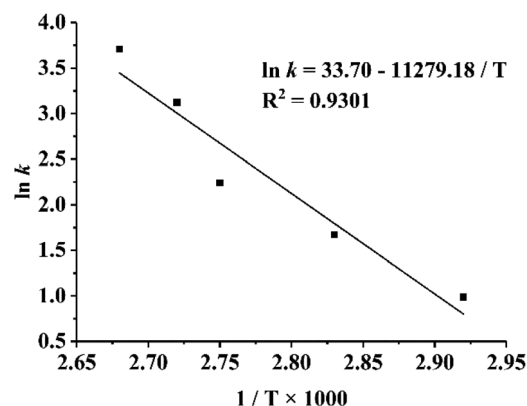


Fig. 7 Arrhenius plot of  $\ln k$  vs.  $1/T$  for fluorescent AGEs.

can be applied to various foods. Referring to the method of Ferrer, *et al.*<sup>9</sup> with some modification, the activation energy values of fluorescent AGE formation in soy sauce, milk powder, canned mixed congee and soybean paste were also determined, and they were  $75.55 \text{ kJ mol}^{-1}$ ,  $106.12 \text{ kJ mol}^{-1}$ ,  $62.95 \text{ kJ mol}^{-1}$  and  $81.04 \text{ kJ mol}^{-1}$ , respectively. According to the  $E_a$  differences between the TTIs and the fluorescent AGEs (which should be less than  $25 \text{ kJ mol}^{-1}$ ), TTI-3 could be used to indicate the levels of fluorescent AGEs in soy sauce, canned mixed congee and soybean paste, and TTI-1 could be used for milk powder.

### 3.4 The 3D packaging container model with TTI

The model TTI and the color changes in TTI-1 during 15 min of incubation at  $80^\circ\text{C}$  are shown in Fig. 10. The materials were sealed in a plastic bag. The selected plastic bag should be resistant to high temperatures ( $100^\circ\text{C}$ ), seal tightly and allow good heat transfer.

The instant soy milk powder was packaged in a cup (Fig. 11). The inside of the cup is a waterproof layer with good heat transfer, which can ensure that the TTI and the soy milk are not in direct contact with each other but are at the same temperature. The outside of the cup is a thermal insulating layer to prevent consumers from being burned. The TTI was embedded

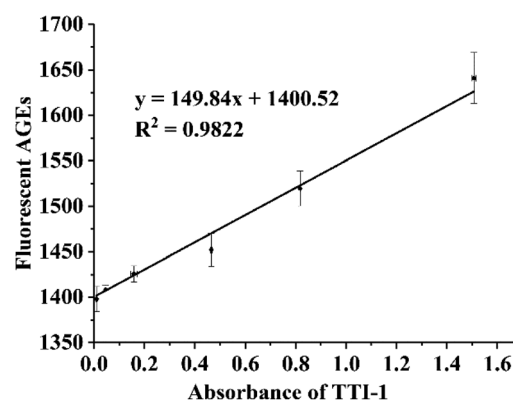


Fig. 8 Scatter plot and fitting curve of TTI-1 vs. total fluorescent AGEs in instant soy milk powder.

Table 4 Dynamic parameters for total fluorescent AGEs in instant soy milk powder at different temperatures

Equations	$T (^\circ\text{C})$	$k (\text{min}^{-1})$	$R^2$	$1/T \times 1000 (\text{K}^{-1})$	$\ln k$
$y = 2.68x + 1403.94$	70	2.68	0.9829	2.92	0.99
$y = 5.32x + 1403.94$	80	5.32	0.9900	2.83	1.67
$y = 9.39x + 1397.91$	90	9.39	0.9827	2.75	2.24
$y = 22.72x + 1410.06$	95	22.72	0.9946	2.72	3.12
$y = 40.64x + 1407.57$	100	40.64	0.9966	2.68	3.70



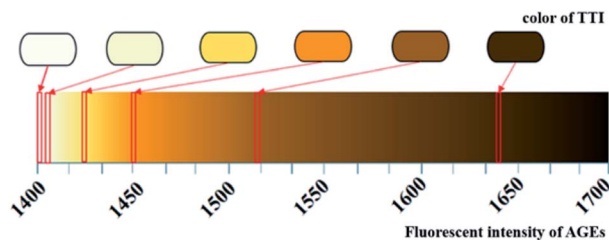


Fig. 9 Colorimetric card of TTI-1 in instant soy milk powder.

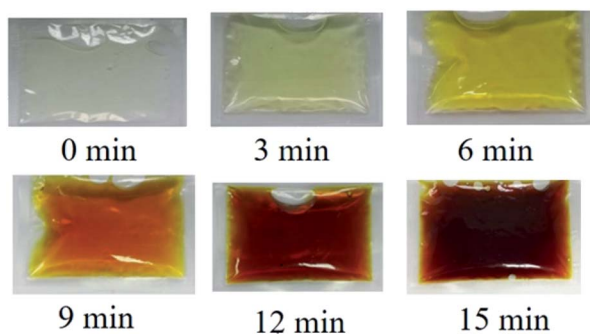


Fig. 10 The model of TTI and color changes of TTI-1 during 15 min of incubation at 80 °C.



Fig. 11 Package design of instant soy milk powder.

in hollowed-out part of the outer layer. When the soy milk powder was brewed with hot water, AGEs were continuously generated in the soy milk in the cup, and the embedded TTI changed color. The consumers observed the TTI color and then compared the final color to the colorimetric card to determine the content of AGEs in the soy milk.

Similarly, this system is compatible with other packaging shapes, such as cuboid and hemispherical packages, that would suit different foods that need to be reheated.

## 4 Conclusions

A prototype Maillard reaction-based TTI composed of xylose and lysine was developed in this study. The materials, response variables and Arrhenius behavior of the TTIs were evaluated. According to the  $E_a$  difference with various concentrations of lysine/xylose, a series of Maillard reaction-based TTIs were established for use with various foods, as the  $E_a$  difference between the TTI and the food should be less than 25 kJ mol<sup>-1</sup>. The results showed that the levels of AGEs in reheated foods could be characterized by the model TTI. Instant soy milk powder was used as a model system to demonstrate the practicality of using TTIs to indicate the level of fluorescent AGEs in foods, and this system provides a research basis for the practical application of the developed Maillard reaction-based TTIs for various reheated foods.

Further research is needed to determine suitable forms for the TTIs for different kinds of foods on the basis of the food processing and consumption methods. Increasing the indication accuracy and broadening the applied range would facilitate the commercialization of this Maillard reaction-based TTI.

## Author contributions

Data curation: Bei Hu, Yuting Li and Bing Li; formal analysis: Bei Hu, Lin Li, Mingquan Yang, Aijuan Jia, Sui Chen and Xia Zhang; methodology: Bei Hu, Yi Hu and Yuting Li; writing—original draft: Bei Hu; writing—review & editing: Lin Li, Di Zhao, Bing Li and Xia Zhang.

## Conflicts of interest

There are no conflicts to declare.

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## References

- 1 M. Li, M. Zeng, Z. He, Z. Zheng, F. Qin, G. Tao, S. Zhang and J. Chen, *J. Agric. Food Chem.*, 2015, **63**, 1658–1663.
- 2 M. W. Poulsen, R. V. Hedegaard, J. M. Andersen, C. B. De, S. Bügel, J. Nielsen, L. H. Skibsted and L. O. Dragsted, *Food Chem. Toxicol.*, 2013, **60**, 10–37.
- 3 N. Grossin, F. Auger, C. Niquet Leridon, N. Durieux, D. Montaigne, A. M. Schmidt, S. Susen, P. Jacolot, J. B. Beuscart, F. J. Tessier and E. Boulanger, *Mol. Nutr. Food Res.*, 2015, **59**, 927–938.





- 4 D. Zhao, L. Li, T. T. Le, L. B. Larsen, G. Su, Y. Liang and B. Li, *J. Agric. Food Chem.*, 2017, **65**, 5778–5788.
- 5 K. J. Bar, S. Franke, B. Wenda, S. Muller, R. Kientsch-Engel, G. Stein and H. Sauer, *Neurobiol. Aging*, 2003, **24**, 333–338.
- 6 A. Gautieri, F. S. Passini, U. Silván, M. Guizar-Sicairos, G. Carimati, P. Volpi, M. Moretti, A. Redaelli, M. Berli and J. G. Snedeker, *Matrix Biol.*, 2017, **59**, 95–108.
- 7 H. Vlassara and J. Uribarri, *Curr. Diabetes Rep.*, 2014, **14**, 453.
- 8 H. Vlassara, J. Uribarri, W. Cai, S. Goodman, R. Pyzik, J. Post, F. Grosjean, M. Woodward and G. E. Striker, *Clin. J. Am. Soc. Nephrol.*, 2012, **7**, 934–942.
- 9 E. Ferrer, A. Alegria, R. Farre, G. Clemente and C. Calvo, *J. Agric. Food Chem.*, 2005, **53**, 4911–4917.
- 10 T. Henle, *Amino Acids*, 2005, **29**, 313–322.
- 11 D. Laroque, C. Inisan, C. Berger, É. Voulard, L. Dufossé and F. Guérard, *Food Chem.*, 2008, **111**, 1032–1042.
- 12 E. J. Kwak and S. I. Lim, *Amino Acids*, 2004, **27**, 85–90.
- 13 S. I. F. S. Martins, W. M. F. Jongen and M. A. J. S. van Boekel, *Trends Food Sci. Technol.*, 2000, **11**, 364–373.
- 14 E. H. Ajandouz, V. Desseaux, S. Tazi and A. Puigserver, *Food Chem.*, 2008, **107**, 1244–1252.
- 15 L. Jiménez-Castaño, M. Villamiel, P. J. Martín-álvarez, A. Olano and R. López-Fandiño, *Food Hydrocolloids*, 2005, **19**, 831–837.
- 16 P. Chao, C. Hsu and M. Yin, *Food Chem.*, 2009, **113**, 262–266.
- 17 G. Chen and J. S. Smith, *Food Chem.*, 2015, **168**, 190–195.
- 18 J. L. J. M. Scheijen, E. Clevers, E. Lian, P. C. Dagnelie, F. Brouns, C. D. A. Stehouwer and C. G. Schalkwijk, *Food Chem.*, 2015, **190**, 1145–1150.
- 19 H. Glj, J. V. Woodside, J. M. Ames and G. J. Cuskelly, *Food Chem.*, 2012, **131**, 170–174.
- 20 P. S. Taoukis and T. P. Labuza, *J. Food Sci.*, 1989, **54**, 783–788.
- 21 S. Wang, X. Liu, M. Yang, Y. Zhang, K. Xiang and R. Tang, *Packag. Technol. Sci.*, 2015, **28**, 839–867.
- 22 M. J. Kim, H. W. Shin and S. J. Lee, *Food Control*, 2016, **70**, 167–173.
- 23 K. S. Pennanen, C. Focas, V. Kumpusalo Sanna, K. Keskitalo Vuokko, I. Matullat, M. Ellouze, S. Pentik Inen, M. Smolander, V. Korhonen and M. Ollila, *Packag. Technol. Sci.*, 2015, **28**, 303–323.
- 24 J. U. Kim, K. Ghafoor, J. Ahn, S. Shin, S. H. Lee, H. M. Shahbaz, H. H. Shin, S. Kim and J. Park, *LWT-Food Sci. Technol.*, 2016, **67**, 143–150.
- 25 A. P. Brizio and C. Prentice, *J. Food Sci.*, 2015, **80**, M1271–M1276.
- 26 D. Wu, S. Hou, J. Chen, Y. Sun, X. Ye, D. Liu, R. Meng and Y. Wang, *LWT-Food Sci. Technol.*, 2015, **60**, 1100–1104.
- 27 L. Young-Soon, S. Homma and K. Eun-Jung, *Food Sci. Technol. Res.*, 2013, **19**, 319–322.
- 28 A. W. Edwards, K. Blackler, R. D. Gill, E. V. D. Goot and J. Holm, *Elements*, 2015, **11**, 323–329.
- 29 T. Yamamoto and K. Isshiki, *Jpn. J. Food Chem. Saf.*, 2012, **19**, 84–87.
- 30 H. Fujikawa, T. Yoshikawa, S. Hasegawa, T. Hayashi, T. Aoyama, T. Ogata, H. Cho, T. Oshima, Y. Rino and S. Morita, *Ann. Surg. Oncol.*, 2014, **21**, 3002–3007.
- 31 H. Rokugawa and H. Fujikawa, *Food Control*, 2015, **57**, 355–361.
- 32 J. H. Lee, R. Harada, S. Kawamura and S. Koseki, *Food Bioprocess Technol.*, 2018, **11**, 185–193.
- 33 L. Bosch, A. Alegria, R. Farré and G. Clemente, *Food Chem.*, 2007, **105**, 1135–1143.
- 34 D. Zhao, L. Li, D. Xu, B. Sheng, D. Qin, J. Chen, B. Li and X. Zhang, *Food Hydrocolloids*, 2018, **80**, 122–129.
- 35 M. G. Huang, X. M. Zhang, K. Eric, S. Abbas, K. Hayat, P. Liu, S. Q. Xia and C. S. Jia, *J. Pept. Sci.*, 2012, **18**, 342–349.
- 36 B. Fu and T. P. Labuza, *Food Control*, 1993, **4**, 125–133.
- 37 H. W. Bian, Y. T. Li, B. Li, Q. Y. Fu and L. Li, *Mod. Food Sci. Technol.*, 2014, **30**, 223–228.
- 38 K. Yamaguchi, Y. Nomi, T. Homma, M. Kasai and Y. Otsuka, *Food Sci. Technol. Res.*, 2012, **18**, 67–76.
- 39 P. J. Beisswenger, S. Howell, T. Mackenzie, H. Corstjens, N. Muizzuddin and M. S. Matsui, *Diabetes Technol. Ther.*, 2012, **14**, 285–292.
- 40 B. Fu and T. P. Labuza, *J. Food Distrib. Res.*, 1992, **23**, 9–18.
- 41 L. C. Maillard, *C. R. Acad. Sci.*, 1912, **154**, 66–68.
- 42 J. A. Ruriánhenares and F. J. Morales, *J. Agric. Food Chem.*, 2008, **56**, 2357–2362.
- 43 F. J. Morales, V. Somoza and V. Fogliano, *Amino Acids*, 2012, **42**, 1097–1109.
- 44 A. N. Wijewickreme, D. D. Kitts and T. D. Durance, *J. Agric. Food Chem.*, 1998, **45**, 4577–4583.
- 45 M. J. Friedman, *Agric. Food Chem.*, 1996, **44**, 631–653.
- 46 M. Qian, W. Bai, W. Zhao, X. Liu and X. Zeng, *Food Sci.*, 2016, **37**, 31–35.
- 47 H. L. Wu, Z. Q. Wang, C. Han, Z. N. Peng and Y. Q. Chen, *Mod. Food Sci. Technol.*, 2010, **26**, 441–444.
- 48 E. H. Ajandouz, L. S. Tchiakpe, F. D. Ore, A. Benajiba and A. Puigserver, *J. Food Sci.*, 2001, **66**, 926–931.
- 49 E. H. Ajandouz and A. Puigserver, *J. Agric. Food Chem.*, 1999, **47**, 1786–1793.
- 50 H. E. Nursten, *The Maillard Reaction: Chemistry, Biochemistry and Implications*, 2005.
- 51 X. Wan and M. Knoll, *J. Food Eng.*, 2016, **168**, 79–83.
- 52 T. Hofmann, *J. Agric. Food Chem.*, 1998, **46**, 3891–3895.
- 53 P. S. Taoukis, in *Food Process Modelling*, 2001, pp. 402–431.
- 54 P. S. Taoukis and T. P. Labuza, *Novel Food Packaging Techniques*, 2003, pp. 103–126.

