


Cite this: *RSC Adv.*, 2020, 10, 11615

Microwave-assisted synthesis, biological evaluation and molecular docking studies of new coumarin-based 1,2,3-triazoles†

Ravinder Dharavath,^a Nalaparaju Nagaraju,^a M. Ram Reddy,^a D. Ashok,^{ID} ^{*a}
M. Sarasija,^b M. Vijjulatha,^c Vani T.,^c K. Jyothi^d and G. Prashanthi^d

Coumarin-based 1,4-disubstituted 1,2,3-triazole derivatives were synthesized using a highly efficient, eco-friendly protocol *via* a copper(I)-catalyzed click reaction between various substituted arylazides and terminal alkynes. The synthetic route was easy to access and gave excellent yields under microwave irradiation conditions compared to the conventional heating route. The structures of all the compounds were characterized by IR, ¹H NMR, ¹³C NMR spectroscopy and mass spectrometry. All the synthesized compounds were screened for their *in vitro* antimicrobial, antioxidant and anti-inflammatory activities; among all compounds, **8a**, **8j**, **8k** and **8l** exhibited better results with respect to standard drugs. Furthermore, molecular docking studies have been carried out with PDB IDs 2VCX (anti-inflammatory), 3VXI (antioxidant), 4GEE (antimicrobial) and 2XFH (antifungal) using the Glide module of the Schrödinger suite. The final compounds **8d**, **8e**, **8h**, and **8k** showed the highest hydrogen bond interactions with His-88 and Val-191 proteins and with water in all the proteins.

Received 4th February 2020
Accepted 24th February 2020

DOI: 10.1039/d0ra01052a

rsc.li/rsc-advances

1. Introduction

In spite of the extensive use of antibiotic and vaccination programs, infectious diseases continue to be a primary cause of morbidity and mortality worldwide. The continued emergence of antibiotic resistance is one of the utmost global health threats.¹ Multidrug resistant bacterial infections are increasing in frequency and require the use of more aggressive antibiotic therapies.² While the continued development of novel antibiotics is crucial, alternative strategies are also needed, such as the development of adjuvants that target bacterial pathways responsible for antibiotic tolerance or resistance.³ Inflammation is a general condition that occurs during infections in many number of diseases from hay fever, periodontitis, atherosclerosis, rheumatoid arthritis to cancer. The generally used non-steroidal anti-inflammatory drugs (NSAIDs) provide analgesic and antipyretic effects in addition to anti-inflammatory effects in higher doses. The therapeutics are prevailing, but they seriously increase vascular and

gastrointestinal risks.⁴ Although several drugs exist in the market, there is a serious need to required for the development of new antimicrobial drugs and NSAID (Nonsteroidal Anti-Inflammatory Drugs).

The chemistry of heterocyclic compounds has been an interesting field of study for a long period. Coumarin belongs to the class of flavonoids and are found in many natural as well as synthetic products. Coumarin, a well-known structural motif, shows many pharmacological activities such as anticancer,⁵ antioxidant,⁶ anti-inflammatory,⁷ antimicrobial,⁸ and anticoagulant⁹ effects; it is also proved to be a promising anti-acetylcholinesterase¹⁰ agent. Some of the marketed drugs like Warfarin,⁹ Phenprocoumon,¹⁰ Psoralen¹¹ and Angelicin¹² contain the coumarin skeleton in their structure and are found to exhibit various pharmacological activities (Fig. 1). Coumarins can be synthesized by using the Perkin,^{13,14} Pechmann^{15,16} and Knoevenagel¹⁷ reactions. Among the coumarin compounds, the 3-aryl substituted coumarins have gained attention due to their biological activities such as antimicrobial, antioxidant and anti-inflammatory activities. The 3-aryl coumarin derivatives can be synthesized by using reagents such as DCC, DDQ, NaOH, POCl₃ and Mukaiyama reagent (2-chloro-1-methylpyridiniumiodide).^{18,19} Many of these methods suffer limitations such as the formation of a complex mixture of products, usage of excess reagents and longer reaction times with poor yields. For this reason, a relatively more viable reagent such as 1,1-carbonyldiimidazole^{20,21} (CDI) is used for the synthesis of 3-aryl coumarin derivatives, which provides excellent yields by avoiding all the above-mentioned issues. On the other hand 1,2,3-triazole, an important heterocyclic motif, has gained much

^aGreen and Medicinal Chemistry Lab, Department of Chemistry, Osmania University, Hyderabad-500007, India. E-mail: ashokdou@gmail.com

^bDepartment of Chemistry, Satavahana University, Karimnagar-505001, India

^cMolecular Modelling and Medicinal Chemistry Group, Department of Chemistry, Osmania University, Hyderabad-500007, India

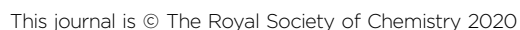
^dDepartment of Pharmaceutical Chemistry, St Mary's College of Pharmacy, Secunderabad-500025, India

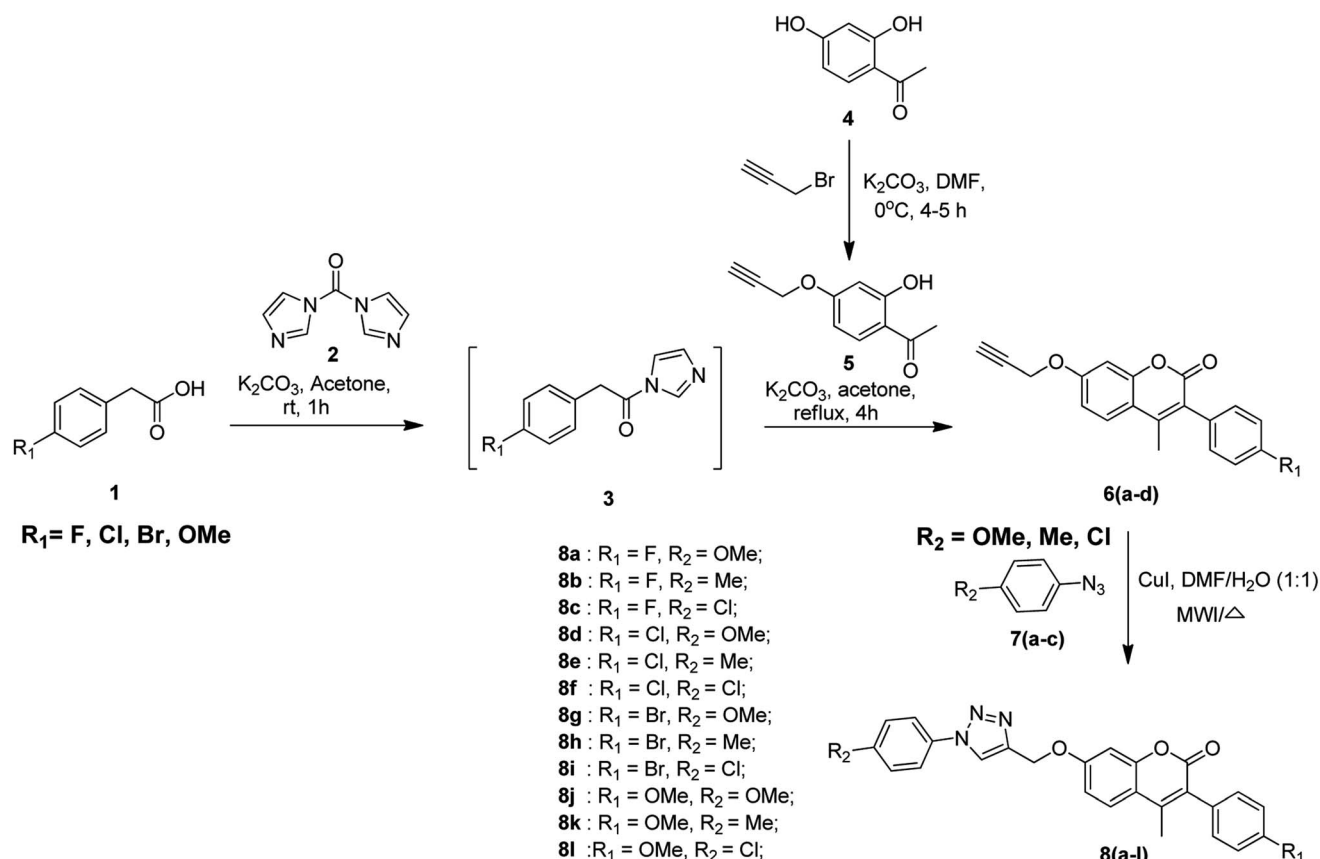
† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra01052a





Computational biology and bioinformatics play a major role in designing drug molecules and have the potential to speed up the drug discovery process. Molecular docking of the drug molecules with the receptor (target) gives significant information about drug-receptor interactions and is frequently used to find out the binding orientation of drug candidates to their protein targets in order to calculate the affinity and activity.³⁹ In view of the pharmaceutical activities of two or more pharmacophore moieties, *i.e.*, coumarin and 1,2,3-triazole, in order to find the combined effects of the newly synthesized scaffolds on biological potency, we herein report a protocol for the design and synthesis of some coumarin-based 1,2,3-triazole compounds.





Scheme 1 Synthetic route for coumarin-based 1,2,3-triazole compounds 8(a-l).

2. Results and discussion

2.1 Chemistry

The protocol for the synthesis of coumarin-based 1,2,3-triazoles is depicted in Scheme 1. The first step of the synthetic route involves the activation of substituted phenyl acetic acid **1** by using 1,1-carbonyldiimidazole **2** (CDI) using potassium carbonate as the base and acetone as the solvent at room temperature for 1 h to obtain the intermediate **3**. In the second step, the alkyne-substituted hydroxy acetophenone **5** is obtained by the selective propargylation of 2,4-dihydroxyacetophenone **4** using K_2CO_3 in DMF and it is further treated with the intermediate **3** under reflux conditions for 4 h using potassium carbonate and acetone to afford 3-aryl-substituted coumarin compounds **6(a-d)**. Finally, the alkyne coumarin intermediates **6(a-d)** undergo copper(i)-catalyzed Huisgen cycloaddition⁴¹ (click chemistry⁴²⁻⁴⁴) when treated with different aryl azides **7(a-c)** using copper iodide in the presence of DMF/ H_2O (1 : 1) to obtain coumarin-containing 1,2,3-triazole title compounds **8(a-l)**.

Initially, the activation of phenyl acetic acids was attempted using DABCO as the coupling agent by using a procedure reported in literature,⁴⁵ but it could not give promising results. Activation was then performed by treating an acid (1 eq.) with DBU (1 eq.) as a base using two different solvents, namely, dichloromethane and toluene by following a literature protocol.⁴⁶

Finally, the reaction conditions were successfully optimized with acid (1 eq.), CDI (1.2 eq.) using K_2CO_3 as the base and dry acetone as the solvent under inert atmospheric conditions.

Table 1 Comparison of the time and yields of the synthesized compounds **6(a-d)** and **8(a-l)** using conventional and microwave irradiation methods

Product	Conventional method		Microwave irradiation method	
	Time (h)	% yield	Time (min)	% yield
6a	5	75	5	86
6b	5.5	78	6	89
6c	4	79	7	90
6d	4.5	75	6	86
8a	5	72	7	85
8b	4.5	69	7	82
8c	5	75	6	84
8d	4.5	76	8	85
8e	5.5	75	7	85
8f	5	76	6	88
8g	5	70	8	82
8h	5.5	69	8	80
8i	4.5	72	7	85
8j	5	68	5.5	80
8k	6	72	6	85
8l	5.5	68	7	80



Table 2 Optimization of compound **8a** in various methods

Entry	Solvent	Conventional method	Microwave irradiation method
		(% yield)	(% yield)
1	THF	35	40
2	CH ₃ CN	20	27
3	1,4-Dioxane	17	24
4	DMF	58	62
5	DMF/H ₂ O (7 : 3)	60	65
6	DMF/H ₂ O (3 : 2)	68	75
7	DMF/H ₂ O (1 : 1)	72	85

The syntheses of alkyne-substituted 3-aryl coumarin intermediates **6(a-d)** and coumarin-based 1,2,3-triazole final compounds **8(a-l)** were carried out using both conventional and microwave (MW) irradiation methods. Among the two methods, the microwave irradiation method gave good yields (80–90%) when compared to the conventional method (68–79%). The reaction time and yields obtained for the intermediates **6(a-d)** and final compounds **8(a-l)** via both the methods are summarized in Table 1.

The optimized reaction conditions for the synthesis of the final compounds **8(a-l)** by the click reaction of different aryl azides **7(a-c)** with alkyne intermediates **6(a-d)** using copper(i) iodide as a catalyst under different solvent conditions in both conventional and MW irradiation methods are tabulated in Table 2. Among the reaction conditions performed, the best results were achieved when DMF/H₂O (1 : 1) was used as the solvent system in both the methods.

Table 3 Antioxidant activity of compounds **6(a-d)** and **8(a-l)**

Compound	Antioxidant activity	
	DPPH method IC ₅₀ (μM mL ⁻¹)	H ₂ O ₂ method IC ₅₀ (μM mL ⁻¹)
6a	2.24 ± 0.93	3.524 ± 0.47
6b	20.00 ± 0.34	23.57 ± 0.68
6c	293.18 ± 0.25	7.07 ± 0.02
6d	4.5 ± 0.47	1.35 ± 0.45
8a	3.5 ± 0.25	1.76 ± 0.06
8b	338.48 ± 0.47	124.48 ± 0.53
8c	42.50 ± 0.58	1.45 ± 0.45
8d	1.11 ± 0.43	71.21 ± 0.47
8e	343.33 ± 0.58	343.33 ± 0.57
8f	3.72 ± 0.69	1.72 ± 0.47
8g	18.57 ± 0.12	18.57 ± 0.69
8h	2.51 ± 0.36	42.51 ± 0.47
8i	4.8 ± 0.24	4.8 ± 0.56
8j	1.29 ± 0.35	1.269 ± 0.14
8k	0.061 ± 0.38	0.061 ± 0.41
8l	3.524 ± 0.65	3.06 ± 0.32
Ascorbic acid	1.46 ± 0.52	1.16 ± 0.89

2.2 Biological activity

All the synthesized compounds **6(a-d)** and **8(a-l)** were screened for antioxidant, anti-inflammatory and antimicrobial scavenging activities. Among all the screened compounds **8a**, **8j**, **8k** and **8l** showed better results compared with standard drugs.

2.3 SAR studies

2.3.1 Antioxidant activity. The *in vitro* antioxidant⁴⁷ activities of intermediates **6(a-d)** and final targets **8(a-l)** were determined by using two methods, namely, the DPPH⁴⁷ radical scavenging assay and H₂O₂⁴⁸ scavenging assay method³¹ with Ascorbic acid taken as the standard drug. All the synthesized compounds, *i.e.*, **6(a-d)** and **8(a-l)** were screened for *in vitro* scavenging activity. The IC₅₀ values ranged from 338.48 μM mL⁻¹ to −0.064 μM mL⁻¹ compared with that of standard Ascorbic acid (IC₅₀ value 1.46 μM mL⁻¹). Compounds **8k** (IC₅₀ value 0.06146 μM mL⁻¹), **8j** (IC₅₀ value 1.11 μM mL⁻¹) and **8d** (IC₅₀ value 1.29 μM mL⁻¹) showed excellent activity because the presence of methoxy and other electron releasing groups like methyl maintained the stability of the compounds and also contributed to the improved activity for compound **8k**. The presence of two methoxy groups in compound **8d** slightly decreased the activity. Further substitution of electron-withdrawing groups like chlorine and methoxy in compound **8l** decreased the activity. In addition, **8a** (3.50 μM mL⁻¹), **8l** (3.52 μM mL⁻¹) and **8f** (3.72 μM mL⁻¹) exhibited good activity than the standard drug in the DPPH method. In contrast, in the H₂O₂ method, compound **8k** (IC₅₀ value 0.06146 μM mL⁻¹) exhibited significant results and **8j** (IC₅₀ value 1.29 μM mL⁻¹), **8c** (IC₅₀ value 1.45 μM mL⁻¹), **8f** (IC₅₀ value 1.72 μM mL⁻¹) and **8a** (IC₅₀ value 1.76 μM mL⁻¹) revealed good activity than standard

Table 4 Anti-inflammatory activity of compounds **6(a-d)** and **8(a-l)**

Compound	Anti-inflammatory	
	Egg-albumin method IC ₅₀ (μM mL ⁻¹)	Heat-induced hemolytic method IC ₅₀ (μM mL ⁻¹)
6a	42.06 ± 0.57	12.06 ± 0.57
6b	22.91 ± 0.25	22.91 ± 0.25
6c	63.72 ± 0.32	63.72 ± 0.32
6d	16.50 ± 0.91	15.00 ± 0.91
8a	15.78 ± 0.52	17.78 ± 0.42
8b	59.809 ± 0.69	69.809 ± 0.69
8c	76.49 ± 0.14	66.49 ± 0.14
8d	238.13 ± 0.35	138.13 ± 0.35
8e	59.80 ± 0.63	69.80 ± 0.63
8f	31.21 ± 0.47	17.11 ± 0.47
8g	103.26 ± 0.69	13.16 ± 0.69
8h	53.77 ± 0.14	43.77 ± 0.14
8i	45.35 ± 0.87	15.35 ± 0.87
8j	18.90 ± 0.37	15.90 ± 0.37
8k	42.53 ± 0.41	60.67 ± 0.41
8l	28.90 ± 0.65	18.90 ± 0.65
Diclofenac	17.52 ± 0.98	17.52 ± 0.98



Table 5 Antibacterial activity of compounds **6(a–d)** and **8(a–l)** at different concentrations

Compound	Zone of inhibition (mm)							
	Gram positive bacteria				Gram negative bacteria			
	<i>Staphylococcus aureus</i>		<i>Bacillus subtilis</i>		<i>Escherichia coli</i>		<i>Klebsiella pneumonia</i>	
	10 $\mu\text{g mL}^{-1}$	20 $\mu\text{g mL}^{-1}$	10 $\mu\text{g mL}^{-1}$	20 $\mu\text{g mL}^{-1}$	10 $\mu\text{g mL}^{-1}$	20 $\mu\text{g mL}^{-1}$	10 $\mu\text{g mL}^{-1}$	20 $\mu\text{g mL}^{-1}$
6a	12	18	11	21	08	11	06	10
6b	09	19	12	19	10	12	04	11
6c	10	20	10	22	11	15	07	09
6d	12	17	09	18	09	12	12	14
8a	22	32	23	40	16	21	14	20
8b	15	24	16	32	13	16	08	14
8c	18	36	16	34	14	18	09	16
8d	23	32	22	42	18	23	13	20
8e	16	23	15	32	12	16	08	13
8f	18	25	19	38	15	19	11	17
8g	24	34	22	42	18	25	12	22
8h	15	24	16	28	14	17	06	13
8i	16	27	19	39	12	20	09	17
8j	26	34	27	39	21	29	14	22
8k	19	26	16	32	12	14	06	12
8l	19	27	18	37	14	25	08	15
Gatifloxacin	20	30	20	40	15	20	10	18

Ascorbic acid (IC_{50} value $1.16 \mu\text{M mL}^{-1}$). The antioxidant activity values are shown in Table 3.

2.3.2 Anti-inflammatory activity. The *in vitro* anti-inflammatory^{49,50} activities of all the synthesized compounds **6(a–d)** and **8(a–l)** were determined by using two methods, namely, the egg-albumin method and heat-induced hemolytic method by taking Diclofenac as the standard drug. In the egg-albumin method, **8a** (IC_{50} value $15.78 \mu\text{M mL}^{-1}$) and **6d** (IC_{50} value $16.50 \mu\text{M mL}^{-1}$) exhibited excellent activity than Diclofenac (IC_{50} value $17.52 \mu\text{M mL}^{-1}$). In the heat-induced hemolytic method, compounds **6a** (IC_{50} value $12.06 \mu\text{M mL}^{-1}$), **8g** (IC_{50} value $13.16 \mu\text{M mL}^{-1}$), **6d** (IC_{50} value $15.00 \mu\text{M mL}^{-1}$), **8i** (IC_{50} value $15.35 \mu\text{M mL}^{-1}$), **8j** (IC_{50} value $15.90 \mu\text{M mL}^{-1}$) and **8f** (IC_{50} value $17.11 \mu\text{M mL}^{-1}$) exhibited excellent activity; also, **8a** (IC_{50} value $17.78 \mu\text{M mL}^{-1}$) and **8l** (IC_{50} value $18.90 \mu\text{M mL}^{-1}$) demonstrated good activity compared to the standard Diclofenac drug (IC_{50} value $17.52 \mu\text{M mL}^{-1}$). The anti-inflammatory activity values are shown in Table 4.

2.3.3 Antibacterial activity. The newly synthesized compounds **6(a–d)** and **8(a–l)** were screened for their antibacterial activity against Gram-positive strains such as *Staphylococcus aureus* (MTCC 96) and *Bacillus subtilis* (MTCC 121) and Gram-negative strains such as *Escherichia coli* (MTCC 43) and *Klebsiella pneumonia* (MTCC 530) at various concentrations, i.e., $10 \mu\text{g mL}^{-1}$ and $20 \mu\text{g mL}^{-1}$ by using the agar disc diffusion method.⁵¹ The zone of inhibition was measured in mm and Gatifloxacin was used as the standard drug; the results are shown in Table 5. From the biological evaluation of the activity of the intermediates **6(a–d)** to the final compounds **8(a–l)**, activity further increased due to the presence of 1,2,3-triazole moieties. Among all the prepared compounds, **8a**, **8d**, **8g** and **8j** were highly potent due to the presence of the methoxy group in the triazole ring and also, compounds **8b**, **8c**, **8e**, **8f**, **8h**

and **8i** showed good activity against all bacterial strains due to the electron-withdrawing groups on the coumarin ring. Compound **8k** and compound **8l** both showed similar activity to that of the standard drug. The starting compounds **6(a–d)** did not show activity. The results also demonstrated that the activity of these compounds **6(a–d)** and **8(a–l)** was influenced by their structures. In conclusion, **8a** and **8j** showed very high potential antibacterial activity against tested organisms.

Table 6 Antifungal activities of the compounds **6(a–d)** and **8(a–l)** at a concentration $50 \mu\text{g mL}^{-1}$

Compound	Zone of inhibition (mm) at $50 \mu\text{g mL}^{-1}$ concentration		
	<i>Aspergillus niger</i>	<i>Aspergillus flavus</i>	<i>Fusarium oxysporum</i>
6a	07.2	06.4	07.3
6b	08.8	07.2	06.6
6c	07.2	08.6	09.2
6d	08.2	07.9	06.5
8a	18.8	17.6	18.6
8b	17.9	16.8	17.8
8c	18.2	16.3	18.9
8d	10.6	11.8	11.6
8e	13.6	14.2	14.8
8f	15.2	14.8	16.2
8g	10.3	12.5	12.6
8h	14.6	13.8	14.6
8i	15.9	14.5	15.8
8j	18.6	16.7	19.0
8k	16.5	15.8	16.2
8l	15.9	14.8	14.3
Clotrimazole	17.3	16.4	18.2



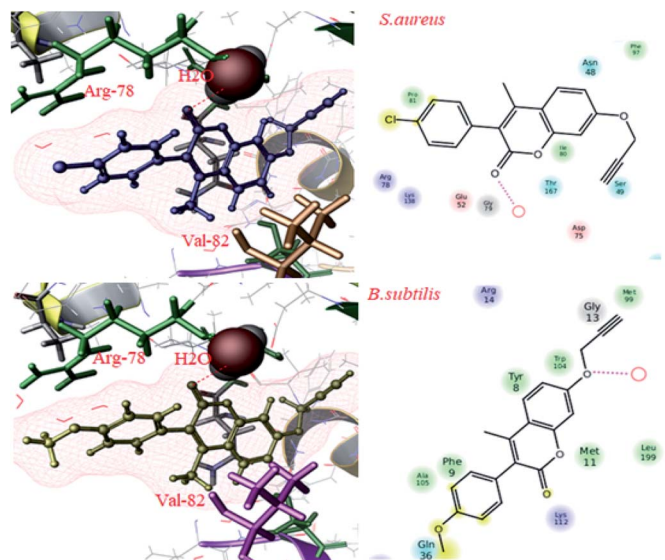


Fig. 3 Docking pose images of intermediate compounds **6b** and **6d** showing H-bond interaction with water in Gram positive bacteria by the protein 4GEE.

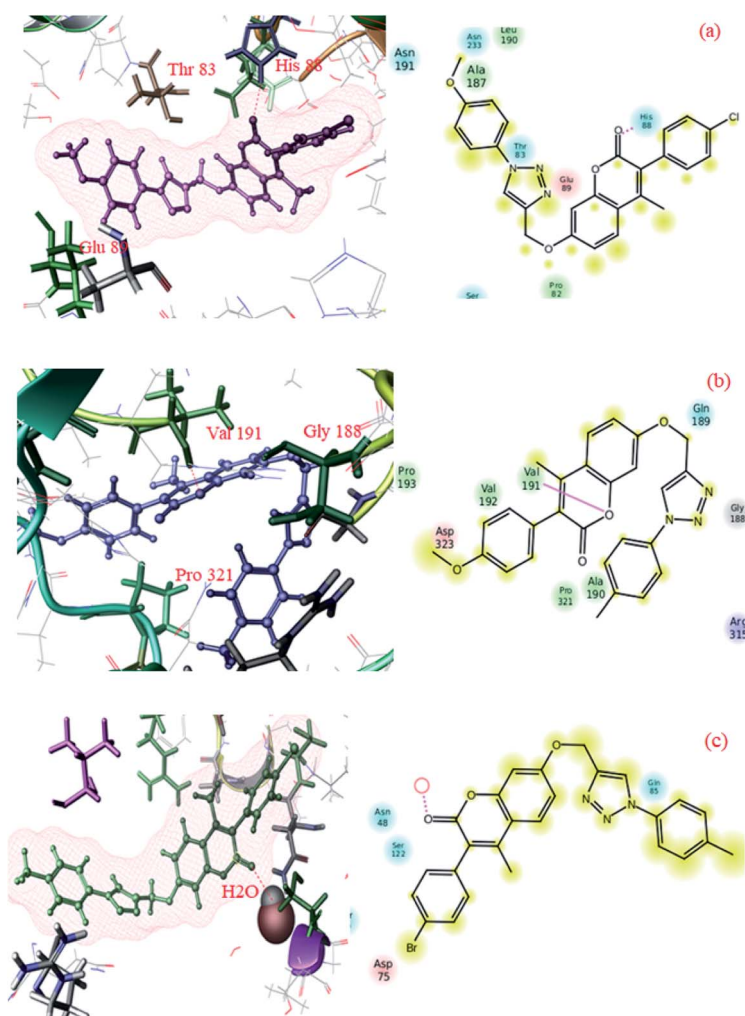


Fig. 4 Docking pose images of final compounds **8d**, **8h** and **8k** showing H-bond interaction with His-88 in 2VCX protein (a), Val-191 in 3VXI (b) and water in 2XFH and 4GEE proteins (c), respectively.



2.3.4 Antifungal activity. All the synthesized compounds **6(a–d)** and **8(a–l)** were screened for their *in vitro* antifungal activity against three fungal organisms, namely, *Aspergillus niger*, *Aspergillus flavus* and *Fusariumoxysporum* at a concentration of 50 $\mu\text{g mL}^{-1}$ by using the disc diffusion method⁵¹ and the results were compared with that of Clotrimazole, used as a standard drug. The study of antifungal activity is shown in Table 6; it was observed that among all the synthesized compounds, **8a**, **8b**, and **8c** showed better activity against the three pathogenic fungi due to the presence of fluorine group on coumarin. Compounds **8j**, **8k** and **8l** showed better activity due to the presence of the methoxy group in the triazole ring. The remaining compounds exhibited similar activity to that of the standard drug. Due to the methoxy group of the triazole compounds **8d** and **8g**, they showed poor activity. It was observed that the intermediate-to-product activity increased due to the presence of the 1,2,3-triazole moiety. Overall, the antifungal activities of the compounds were good against the tested fungal strains.

2.4 Molecular docking studies

To gain more insights into the interactions of coumarin-based 1,2,3-triazole derivatives **8(a–l)**, molecular docking studies were performed. Molecular docking was carried out using the Glide module of the Schrödinger suite using PDB IDs 2VCX (anti-inflammatory),⁵² 3VXI (antioxidant),⁵³ 4GEE (antimicrobial)⁵⁴ and 2XFH (antifungal)⁵⁵ to correlate the binding mode of the synthesized compounds with the proteins. The above-mentioned proteins were selected on the basis of the reference compound that existed as a co-crystal ligand on the target protein. In the current work, we have considered 2VCX for its

anti-inflammatory activity since it is a prostaglandin D2 synthase⁵⁶ protein involved in the cyclooxygenase pathway.⁵⁷ Similarly, 3VXI was used for its anti-oxidant activity due to its dye decolourising peroxidase (DyP) complex with Ascorbic acid as the crystal ligand.⁵⁸ The protein 4GEE was considered because of its DNA-gyrase B and topoisomerase IV-mediated broad spectrum activity.⁵⁹ For the identification of antifungal activity, we thereby considered 2XFH Cytochrome P450 EryK cocrystallized with the inhibitor Clotrimazole.⁶⁰ The potential binding free energies (ΔG) were evaluated and the rationality of the compounds was further evaluated through QikProp for their drug likeness.

In conclusion, comparative docking studies from all the proteins revealed that compound **8l** showed a high docking score of -7.792 and a binding energy of -117.95 in the protein 2VCX; also, their predicted activities were found to be slightly higher than that of the standard (provided in the ESI Table†). This clearly indicated that compound **8l** showed an explicit effect as a potential anti-inflammatory, antimicrobial and antioxidant molecule. The intermediate compounds **6b** and **6d** showed best H-bond interactions with water in all the proteins (2XFH, 4GEE and 3VXI) as the standard Ascorbic acid and Gatifloxacin predominantly (as illustrated in Fig. 1 and 3 of the ESI†), stating that these compounds could be further exploited to newer chemical agents possessing antioxidant and antimicrobial activities, as depicted in Fig. 4. The standard antifungal agent Clotrimazole shows hydrogen bond interactions with Arg-293 (depicted in Fig. 2 provided in the ESI†). The final compounds **8d**, **8e**, **8h** and **8k** showed hydrogen bond interactions with His-88 (Fig. 4a) and Val-191 (**4b**) amino acids and with water (**4c**) in all the proteins 2VCX, 3VXI, 4GEE and 2XFH.

Table 7 Quantified active site binding free energies along with the docking scores of the synthesized compounds and standard molecules

Compounds	Docking scores				Binding free energies			
	Anti-inflammatory activity (2VCX)	Anti-oxidant activity (3VXI)	Anti-microbial		Anti-inflammatory activity (2VCX)	Anti-oxidant activity (3VXI)	Anti-microbial	
			Anti-bacterial (4GEE)	Anti-fungal (2XFH)			Anti-bacterial (4GEE)	Anti-fungal (2XFH)
6a	−5.984	−4.882	−5.760	−6.657	−83.72	−54.41	−69.13	−77.31
6b	−6.315	−4.826	−6.748	−5.148	−86.94	−55.04	−70.79	−61.08
6c	−6.177	−4.559	−6.585	−4.857	−86.36	−55.68	−71.29	−61.61
6d	−6.232	−3.801	−6.580	−4.791	−83.57	−46.00	−78.56	−57.66
8a	−6.928	−3.355	−5.756	−3.180	−94.07	−52.26	−68.97	−52.0
8b	−6.587	−3.636	−4.994	−4.096	−88.37	−53.29	−57.01	−52.6
8c	−6.165	−2.949	−5.122	−5.676	−88.66	−59.42	−53.52	−69.37
8d	−6.962	−2.438	−5.970	−2.927	−92.74	−54.29	−71.94	−45.55
8e	−6.927	−3.740	−5.796	−2.368	−92.26	−60.44	−71.17	−38.23
8f	−6.051	−3.528	−4.818	−4.339	−92.16	−47.36	−62.39	−67.33
8g	−6.972	−3.720	−5.554	−2.792	−106.36	−57.00	−67.63	−44.93
8h	−6.725	−3.778	−5.654	−3.594	−92.263	−47.41	−72.09	−51.41
8i	−6.575	−2.823	−5.659	−4.112	−94.63	−53.35	−71.45	−68.46
8k	−6.587	−3.240	−5.006	−2.587	−94.638	−54.15	−62.72	−37.15
8l	−7.792	−2.868	−4.869	−3.984	−117.95	−50.68	−61.15	−47.79
Diclofenac	−5.553	—	—	—	−91.55	—	—	—
Ascorbic acid	—	−6.184	—	—	—	−85.23	—	—
Gatifloxacin	—	—	−5.553	—	—	—	−56.38	—
Clotrimazole	—	—	—	−6.288	—	—	—	−89.15



This signifies that the synthesized compounds possess anti-bacterial and antifungal activities in addition to antioxidant and anti-inflammatory activities. Docking scores along with the binding free energies for all the synthesized compounds have been quantified and tabulated in Table 7. The average predicted activities of different chemical methods have been calculated for the synthesized compounds (provided in the ESI from Tables 3d to 6d†). The pharmacokinetic properties (ADME) of the compounds exhibited excellent 100% human oral absorption and were found to be in the acceptable range, as shown in Table 2d (provided in the ESI†).

3. Conclusion

We herein, reported the new scaffolds **6(a–d)** and **8(a–l)** were synthesized by using a microwave irradiation method to obtained better yield as compared to conventional heating method. The use of a DMF : water (1 : 1) solvent system helped in increasing the product yield through traditional, conventional and microwave irradiation routes. However, we observed greater yields *via* the microwave irradiation route in lesser duration. Biological evaluations and molecular docking studies revealed an increase in the activity of the target compounds from their intermediates. Compounds **8d**, **8e**, **8h** and **8k** showed excellent hydrogen bonding interactions with His-88, Val-191 and water.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Author Ravinder Dharavath (File No. 09/132(0865)/2017-EMR-I) is grateful to CSIR, New Delhi, for financial assistance in the form of SRF. One of the authors D. Ashok is thankful to UGC-New Delhi, for the award of BSR Faculty Fellowship. Authors NN and MRR are thankful to UGC-New Delhi. The authors thankful to The Head, Department of Chemistry, Osmania University, Hyderabad, for providing laboratory facilities. We thank CFRD, Osmania University analytical team for providing spectral analysis facilities.

References

- 1 D. J. Payne, *Science*, 2008, **321**(5896), 1644–1645.
- 2 H. C. Neu, *Science*, 1992, **257**(5073), 1064–1073.
- 3 C. A. Arias and B. E. Murray, *N. Engl. J. Med.*, 2009, **360**(5), 439–443.
- 4 N. Bhala, J. Emberson, A. Merhi and S. Abramson, *Lancet*, 2013, **382**, 769–779.
- 5 G. Achar, C. R. Shahini, S. A. Patil, J. G. Małecki and S. Budagumpi, *New J. Chem.*, 2019, **43**(3), 1216–1229.
- 6 R. Nagamallu, B. Srinivasan, M. B. Ningappa and A. K. Kariyappa, *Bioorg. Med. Chem. Lett.*, 2016, **26**(2), 690–694.
- 7 P. Chandel, A. Kumar, N. Singla, A. Kumar, G. Singh and R. K. Gill, *MedChemComm*, 2019, **10**(3), 421–430.
- 8 K. Bhagat, J. Bhagat, M. K. Gupta, J. V. Singh, H. K. Gulati, A. Singh and H. Singh, *ACS Omega*, 2019, **4**(5), 8720–8730.
- 9 R. A. O'Reilly and P. M. Aggeler, *Circulation*, 1968, **38**(1), 169–177.
- 10 D. Viña, M. J. Matos, M. Yáñez, L. Santana and E. Uriarte, *MedChemComm*, 2012, **3**(2), 213–218.
- 11 M. L. Panno and F. Giordano, *World J. Clin. Oncol.*, 2014, **5**(3), 348.
- 12 Y. Wang, Y. Chen, X. Chen, Y. Liang, D. Yang, J. Dong and Z. Liang, *Exp. Ther. Med.*, 2019, **18**(5), 3365–3374.
- 13 M. Trković and Z. Ivezic, *J. Heterocycl. Chem.*, 2009, **37**, 137–141.
- 14 S. H. Mashraqui, D. Vashi and H. D. Mistry, *Synth. Commun.*, 2004, **34**(17), 3129–3134.
- 15 Y. Ming and D. W. Boykin, *Heterocycles*, 1987, **26**, 3229–3231.
- 16 R. S. Mali and S. G. Tilve, *Synth. Commun.*, 1990, **20**(12), 1781–1791.
- 17 N. Hans, M. Singhi, V. Sharma and S. K. Grover, *Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem.*, 1996, **35**, 1159–1162.
- 18 D. V. Kadnikov and R. C. Larock, *J. Organomet. Chem.*, 2003, **687**, 422–435.
- 19 D. Olmedo, R. Sancho, L. M. Bedoya, J. L. López-Pérez, E. D. Olmo, E. Muñoz, J. Alcamí, M. P. Gupta and A. S. Feliciano, *Molecules*, 2012, **17**(8), 9245–9257.
- 20 S. K. Verma, R. Ghorpade, A. Pratap and M. P. Kaushik, *Tetrahedron Lett.*, 2012, **53**(19), 2373–2376.
- 21 D. Ashok, E. V. L. Madhuri and M. Sarasija, *Asian J. Chem.*, 2020, **32**(1), 205–208.
- 22 N. Kuntala, J. R. Telu, V. Banothu, S. B. Nallapati, J. S. Anireddy and S. Pal, *MedChemComm*, 2015, **6**(9), 1612–1619.
- 23 P. Neeraja, S. Srinivas, K. Mukkanti, P. K. Dubey and S. Pal, *Bioorg. Med. Chem. Lett.*, 2016, **26**(21), 5212–5217.
- 24 N. Kuntala, J. R. Telu, V. Banothu, S. Balasubramanian, J. S. Anireddy and S. Pal, *Mini-Rev. Med. Chem.*, 2018, **18**(9), 803–809.
- 25 A. M. Macan, A. Harej, I. Cazin, M. Klobučar, V. Stepanić, K. Pavelić and S. Raić-Malić, *Eur. J. Med. Chem.*, 2019, **184**, 111739.
- 26 J. Mareddy, S. B. Nallapati, J. Anireddy, Y. P. Devi, L. N. Mangamoori, R. Kapavarapu and S. Pal, *Bioorg. Med. Chem. Lett.*, 2013, **23**(24), 6721–6727.
- 27 K. S. S. Praveena, S. Durgadas, N. S. Babu, S. Akkenapally, C. G. Kumar, G. S. Deora and S. Pal, *Bioorg. Chem.*, 2014, **53**, 8–14.
- 28 K. S. S. Praveena, E. V. V. S. Ramarao, N. Y. S. Murthy, S. Akkenapally, C. G. Kumar, R. Kapavarapu and S. Pal, *Bioorg. Med. Chem. Lett.*, 2015, **25**(5), 1057–1063.
- 29 J. Mareddy, N. Suresh, C. G. Kumar, R. Kapavarapu, A. Jayasree and S. Pal, *Bioorg. Med. Chem. Lett.*, 2017, **27**(3), 518–523.
- 30 R. Alvarez, S. Velazquez, A. San-Felix, S. Aquaro, E. D. Clercq, C. F. Perno and M. J. Camarasa, *J. Med. Chem.*, 1994, **37**(24), 4185–4194.



- 31 M. H. Shaikh, D. D. Subhedar, L. Nawale, D. Sarkar, F. A. K. Khan, J. N. Sangshetti and B. B. Shingate, *MedChemComm*, 2015, **6**(6), 1104–1116.
- 32 Z. Najafi, M. Mahdavi, M. Saeedi, E. Karimpour-Razkenari, N. Edraki, M. Sharifzadeh and T. Akbarzadeh, *Bioorg. Chem.*, 2019, **83**, 303–316.
- 33 M. Mohammadi-Khanaposhtani, K. Fahimi, E. Karimpour-Razkenari, M. Safavi, M. Mahdavi, M. Saeedi and T. Akbarzadeh, *Lett. Drug Des. Discovery*, 2019, **16**(7), 818–824.
- 34 A. V. Lipieva, D. O. Zakharov, L. G. Burova, T. S. Frolova, D. S. Baev, I. V. Shirokikh and E. E. Shults, *Molecules*, 2019, **24**(11), 2126.
- 35 F. C. Torres, G. A. Gonçalves, K. L. Vanzolini, A. A. Merlo, B. Gauer, M. Holzschuh and G. L. V. Poser, *J. Braz. Chem. Soc.*, 2016, **27**(9), 1541–1550.
- 36 K. Kushwaha, N. Kaushik and S. C. Jain, *Bioorg. Med. Chem. Lett.*, 2014, **24**, 1795–1801.
- 37 R. A. Kusanur and M. V. Kulkarni, *Indian J. Chem., Sect. B: Org. Chem. Incl. Med. Chem.*, 2005, **44**, 591–594.
- 38 M. H. Shaikh, D. D. Subhedar, F. A. K. Khan, J. N. Sangshetti and B. B. Shingate, *Chin. Chem. Lett.*, 2016, **27**(2), 295–301.
- 39 D. Ashok, K. Rangu, V. H. Rao, S. Gundu, B. Srilata and M. Vijjulatha, *Med. Chem. Res.*, 2016, **25**(3), 501–514.
- 40 D. Ashok, S. Gundu, V. K. Aamate, M. G. Devulapally, R. Bathini and V. Manga, *J. Mol. Struct.*, 2018, **1157**, 312–321.
- 41 V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2002, **41**(14), 2596–2599.
- 42 H. C. Kolb, M. G. Finn and K. B. Sharpless, *Angew. Chem., Int. Ed.*, 2001, **40**, 2004–2021.
- 43 Y. Hua and A. H. Flood, *Chem. Soc. Rev.*, 2010, **39**(4), 1262–1271.
- 44 P. Nouraie, S. Moradi Dehaghi and A. Foroumadi, *Synth. Commun.*, 2019, **49**(3), 386–394.
- 45 M. Roussaki, C. A. Kontogiorgis, D. Hadjipavlou-Litina, S. Hamilakis and A. Detsi, *Bioorg. Med. Chem. Lett.*, 2010, **20**(13), 3889–3892.
- 46 S. K. Verma, R. Ghorpade, A. Pratap and M. P. Kaushik, *Tetrahedron Lett.*, 2012, **53**(19), 2373–2376.
- 47 H. Barry, *Drugs*, 1991, **42**(2), 569–605.
- 48 V. Jacob and A. Micheal, *Nutrition*, 1999, **49**, 1–7.
- 49 S. Chandra, P. Chatterjee, P. Dey and S. Bhattacharya, *Asian Pac. J. Trop. Biomed.*, 2012, **2**(1), S178–S180.
- 50 S. C. Chippada and M. Vangalapati, *J. Chem., Biol. Phys. Sci.*, 2011, **1**(2), 260–269.
- 51 A. L. Barry, *Manual of clinical microbiology*, 1985, pp. 978–987.
- 52 M. Hohwy, L. Spadola, B. Lundquist, P. Hawtin, J. Dahmen, I. Groth-Clausen, E. Nilsson, S. Persdotter, K. Von Wachenfeldt, R. H. A. Folmer and K. Edman, *J. Med. Chem.*, 2008, **51**, 2178–2186.
- 53 Y. Sugano, T. Yoshida and H. Tsuge, *wwPDB X-ray Structure Validation Report*, 2018, pp. 1–12.
- 54 L. W. Tari, M. Trzoss, D. C. Bensen, X. Li, Z. Chen, T. Lam, J. Zhang, C. J. Creighton, M. L. Cunningham, B. Kwan, M. K. J. Shaw, F. C. Lightstone, S. E. Wong, T. B. Nguyen, J. Nix and J. Finn, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 1529–1536.
- 55 L. C. Montemiglio, S. Gianni, B. Vallone and C. Savino, *Biochemistry*, 2010, **49**, 9199.
- 56 R. G. Bellomo, S. M. Carmignano, T. Palermo, L. Cosenza, A. Saggini and R. Saggini, *Nonsteroidal Anti-Inflammatory Drugs*, 2017, vol. 67.
- 57 R. Emanuela and A. F. Garret, *Arterioscler., Thromb., Vasc. Biol.*, 2011, **31**(5), 986–1000.
- 58 J. Fabri, W. Dabelstein, A. Reglitzky, A. Schütze and K. Reders, *Ullmann's Encyclopedia of Industrial Chemistry*, 2000.
- 59 F. Collin, S. Karkare and A. Maxwell, *Appl. Microbiol. Biotechnol.*, 2011, **92**(3), 479–497.
- 60 American Society of Health-System Pharmacists Clotrimazole Monograph for Professionals, 8 February 2016.

