


 Cite this: *RSC Adv.*, 2020, 10, 14531

Effects of temperature and polyols on the ciprofloxacin hydrochloride-mediated micellization of sodium dodecyl sulfate†

 Shamim Mahbub,^{ab} Sayma Akter,^a Luthfunnessa,^a Parul Akter,^a Md. Anamul Hoque,^b Malik Abdul Rub,^{cd} Dileep Kumar,^{ef} Yousef G. Alghamdi,^c Abdullah M. Asiri^{cd} and Hurija Džudžević-Čančar^g

Herein, a conductivity method was engaged to explore the effects of a fluoroquinolone drug, namely ciprofloxacin hydrochloride (CFH)/CFH + polyols (organic compounds with multiple hydroxyl groups (glucose and fructose)), on the aggregation phenomenon of sodium dodecyl sulfate (SDS) at different temperatures (298.15–318.15 K) while maintaining a gap of 5 K. In this study, the critical micelle concentration (cmc) of the SDS/SDS + CFH mixture in water and polyols media was determined from plots of the specific conductivity *versus* the concentration of SDS to gain knowledge of the effects of CFH/CFH + polyols on the micelle formation behavior of SDS. The cmc value of the surfactant decreases in the presence of CFH in an aqueous medium; thus, CFH favors the micellization of SDS. The cmc values of SDS and the SDS + CFH mixture were enhanced in polyols media. The cmc values of SDS/SDS + CFH show a U-shaped behavior with temperature. The counterion dissociation (α) of the pure surfactant is higher in the presence of the drug and is further enhanced through an increase in the CFH concentration in water/polyols media. Different thermodynamic parameters, such as the Gibbs free energy of micellization (ΔG_m°), standard enthalpy (ΔH_m°), entropy (ΔS_m°), different transfer energies and enthalpy–entropy compensation parameters of micellization were determined and illustrated in detail to compare these parameters between the pure SDS and SDS + CFH mixture in polyols media. The negative values of ΔG_m° for the SDS/SDS + CFH mixture in all cases indicate spontaneous micelle formation. The ΔH_m° and ΔS_m° values indicate the presence of both hydrophobic and electrostatic interactions amongst the studied components.

 Received 8th January 2020
 Accepted 17th March 2020

DOI: 10.1039/d0ra00213e

rsc.li/rsc-advances

1. Introduction

A surfactant with a structure that contains both hydrophilic and hydrophobic moieties in the same molecule possesses unique solubility properties, *i.e.*, solubility in both polar and nonpolar media. In an aqueous medium, a surfactant can form

aggregates at a certain concentration due to a delicate balance of hydrophilic interactions and hydrophobic interactions.¹ This surfactant aggregate is known as a micelle, and the concentration at which a surfactant micelle formed is known as cmc.^{2–4} This micellar aggregation of an amphiphilic substance can augment the solubility of weakly soluble organic compounds by altering their microenvironment properties *e.g.* polarity, surface tension, viscosity *etc.*⁵ The solubility enhancement property of surfactant micelles is the key factor in their wide use in industrial, commercial and technological applications.^{6–11} Surfactants are extensively found in products (medicines, food, cosmetics, paints *etc.*) are used in daily life. Surfactants are widely used in the textile and pharmaceutical industries as solubility enhancers, diluents, emulsifying agents, and stabilizing agents.^{12–16} In drug formulation and development as well as drug delivery, the study of the molecular interactions between the surfactant and desired drug is interesting and important because the efficiency of drug release and drug delivery depend on the molecular interactions of the surfactants and drug. The release of sparingly water-soluble drugs is enhanced in the presence of surfactant. In the presence of

^aDepartment of Chemistry & Physics, Gono Bishwabidyalay, Savar, Dhaka-1344, Bangladesh

^bDepartment of Chemistry, Jahangirnagar University, Savar, Dhaka-1342, Bangladesh
^cChemistry Department, Faculty of Science, King Abdulaziz University, Jeddah-21589, Saudi Arabia

^dCenter of Excellence for Advanced Materials Research, King Abdulaziz University, Jeddah-21589, Saudi Arabia

^eDivision of Computational Physics, Institute for Computational Science, Ton Duc Thang University, Ho Chi Minh City, Vietnam. E-mail: dileepkumar@tdtu.edu.vn; Tel: +84943720085

^fFaculty of Applied Sciences, Ton Duc Thang University, Ho Chi Minh City, Vietnam

^gDepartment of Natural Science in Pharmacy, Faculty of Pharmacy, University of Sarajevo, Zmaja od Bosne 8, 71 000 Sarajevo, Bosnia and Herzegovina

† Electronic supplementary information (ESI) available. See DOI: 10.1039/d0ra00213e



a lower concentration of surfactant, penetration of water molecules into the drug mass is facilitated due to the lower surface tension and thus wettability of drug increases. At higher concentrations (above cmc), the dissolution of the drug is higher due to the solubilization phenomenon of the surfactant micelles. This solubilization phenomenon enhances the rate of entry of drug molecules into the bloodstream; however, excess use of surfactant lowers the drug absorption level by lowering the chemical potential of the drug. This occurs when the amount of surfactant exceeds the required amount to solubilize the drug,¹⁵ *i.e.*, it exceeds the cmc under the corresponding conditions. Thus, it is important to accurately determine the cmc of a surfactant in the presence of a drug in different media for drug formulation. Again, the structures of surfactants are comparable with the biological membrane; therefore, they can be considered as model biomembranes, and the interactions of drugs with surfactants in the presence of different additives provide important information about the behavior of drugs with biological membranes.^{17,18} In different biomedical applications, fluorescent organic nanoparticles (FON) are significantly employed and have advantages over conventional fluorescent inorganic nanoparticles.^{19–23} However, a drawback of FON is their hydrophobic properties and insolubility in water.^{19,20} Thus, the synthesis of FON with amphiphilic properties as well as increased hydrophilicity is desirable in the field of bioengineering. In the synthesis of amphiphilic FON, surfactants with a combination of aggregation-induced emission components play a significant role.^{19,20}

SDS is an anionic surfactant; it is the most common surfactant used in detergents and is highly effective to remove oily stains. It is used as a food additive and is generally recognized as a safe ingredient. SDS is potentially effective to hinder and avert infections caused by numerous viruses, such as HIV, herpes simplex and the Semliki Forest virus.^{24,25} It is also utilized as a cell lysis agent to extract DNA/RNA and to denature proteins. Aqueous solutions of SDS with Triton X-100 and sodium dodecylbenzene sulfonate are popular for suspending nanotubes, such as carbon nanotubes.²⁶ The currently employed drug CFH is a fluoroquinolone antibiotic drug that is utilized to treat different bacterial diseases, such as skin, respiratory/sinus, bone and joint and urinary tract infections. It can also be exploited for the treatment of gonorrhoea and to treat persons affected by anthrax or plague. However, CFH tends to reduce blood sugar levels. Diabetic patients are at very high risk for numerous bacterial infections, *e.g.* skin infections, respiratory infections, and urinary tract infections.^{27,28} Skin infection, as well as delay of wound healing, are very common in diabetic patients;^{29,30} interestingly, CFH is used for the treatment of these infections.³¹ Thus, the interaction of CFH with a model surfactant such as SDS (or other pharmaceutical ingredients) in the presence of different sugars such as glucose and fructose, which are also known as polyols, is important. The transportation of hydrophobic drugs in the human body is accomplished by the incorporation of surfactant micelles. Again, the micelle formation of a surfactant is a function of the additive concentration. The uptake of CFH by a patient can alter the cmc of the used surfactant; thus, the drug transportation properties

can be altered. Thus, the amount of surfactant that needs to be used in a drug formulation can be understood from the cmc values obtained at varying conditions.

Rub *et al.*³² investigated the behavior of SDS with the amphiphilic drug promazine hydrochloride in the presence of electrolyte and urea; they reported that the cmc was reduced in electrolyte medium and enhanced in urea medium. Our group also investigated the interactions between SDS and CFH in H₂O/electrolyte solutions at different temperatures and observed favorable micellization in the electrolyte medium.³³ The behavior of tetradecyltrimethylammonium bromide (TTAB) and an antibiotic drug, levofloxacin hemihydrate (LFH), in the presence of monohydroxy/polyhydroxy organic compounds was also investigated by our group.³⁴ Although a large number of investigations about the interactions of different ionic surfactants with various drugs have been reported,^{32–35} studies of the interactions of SDS with CFH in the presence of polyols are rare. Accordingly, we planned here to investigate the association behavior of SDS in media containing CFH/CFH + polyols. In the current study, different physico-chemical parameters, such as cmc, α , standard free energy change (ΔG_m°), enthalpy change (ΔH_m°), entropy change (ΔS_m°), and intrinsic enthalpy gain ($\Delta H_m^{*,*}$), have been assessed for SDS aggregation in the presence of CFH/CFH + polyols mixtures to understand the effects of CFH and polyols as well as to elucidate the modes of interaction between the employed components at different temperatures. Here, we apply the surfactant as a drug carrier in glucose or fructose medium as a model drug delivery system. Glucose and fructose are also found in the human body; therefore, their presence may affect the micellization tendencies of surfactant and surfactant–drug mixtures because surfactants are usually used as drug carriers.

2. Experimental

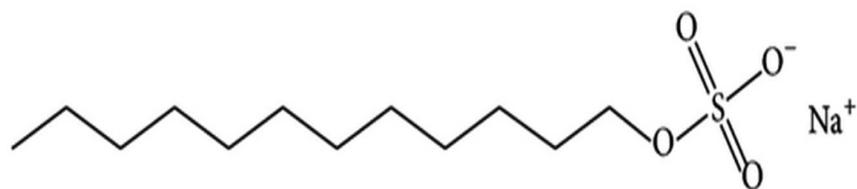
2.1 Materials

In this study, we utilized analytical grade chemicals without any purification. SDS (CAS number: 151-21-3) with a mass fraction purity of 0.98 was collected from Scharlau Chemie S. A. (E.U.). CFH (CAS number: 86483-48-9) was collected from Gonoshasthaya Pharmaceuticals Ltd., Bangladesh. Glucose (CAS number: 50-99-7) and fructose (CAS number: 57-48-7) were collected from Merck, Germany. The mass fraction purities of CFH, glucose, and fructose were 0.98, 0.99 and 0.98, respectively. All employed system solutions were prepared using double-distilled deionized water with a specific conductivity of less than 2 $\mu\text{S cm}^{-1}$ over the temperature range from 298.15 to 318.15 K.

2.2 Conductivity method

Conductivity measurements were performed to elucidate the interactions between SDS (shown in Scheme 1) and CFH (shown in Scheme 2) in the presence/absence of polyols according to a process mentioned in the literature.^{36–40} The specific conductivities of the SDS and SDS + CFH mixed systems with different concentrations of CFH in H₂O and polyols (glucose/fructose)





Scheme 1 Molecular structure of SDS.

media were recorded with a conductivity meter (digital HI 2315, Hanna, Germany) with a dip cell; the cell constant provided by the company was 1.0 cm^{-1} . The precision of the employed conductivity meter was about $\pm 0.5\%$. The temperature of the system was sustained at the desired value with an RM6 Lauda H_2O thermostatted bath with a precision of $\pm 0.2 \text{ K}$. For calibration of the conductivity meter, 0.1 N KCl solution was utilized prior to each experiment. 50 mL of solvent ($\text{H}_2\text{O}/10 \text{ mmol kg}^{-1}$ glucose/ 10 mmol kg^{-1} fructose) in the absence/presence of CFH was placed in a large test tube, and the specific conductivity of the corresponding system was measured. Later, a fixed concentration of surfactant solution prepared in the corresponding solvent was progressively added to the solvent. After appropriate mixing and sufficient time to attain temperature equilibration, the conductivity of the ensuing solution was noted. After that, the observed specific conductivity (κ) reading *versus* [SDS] was plotted by Origin 17 software, and the cmc of that system was estimated from the sharp contravention point.

3. Results and discussion

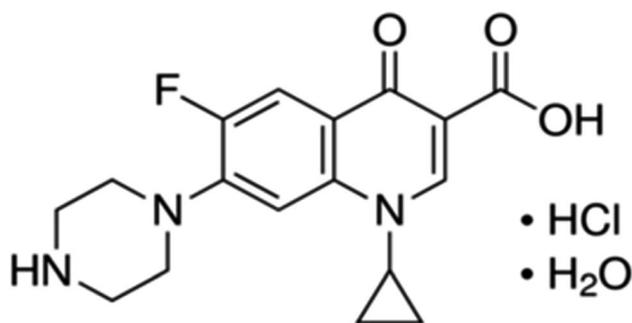
3.1 Critical micelle concentration (cmc) and extent of counterion dissociation (α)

Self-assembly is a general route for the formation of nanoparticles and surfactants that are used in smart materials. Thus, the effects of the concentration of the surfactant, such as sodium dodecyl sulfate (SDS), on the formation of self-assemblies should be evaluated, *i.e.* it is necessary to evaluate the critical concentration of self-assembly (cmc) of SDS in different conditions. For this purpose, in the current study, the cmc values of pure surfactant/surfactant + CFH mixed systems in the absence/presence of polyols (glucose or fructose) at

different temperatures were evaluated. A simple and reliable conductometric technique was employed to determine the cmc of SDS in different additive media. The specific conductivity of the solution of the ionic surfactant shows a linear relationship with the concentration of the surfactant. The conductivity of SDS solution increases with increasing concentration of SDS because the concentrations of Na^+ and DS^- (dodecyl sulfate ion) originate from the increasing disintegration of SDS. However, disruption of the linear relationship between the conductivity and the concentration of SDS was observed at a certain concentration due to micelle formation of SDS because SDS micelles has lower mobility than monomeric SDS molecules. Thus, the contravention point in the specific conductivity (κ) *vs.* [SDS] is considered to be the cmc of the corresponding surfactant system.^{34,41,42} Fig. 1 shows the plots of κ *vs.* surfactant concentration.

The observed cmc of pure SDS at 298.15 K by conductivity measurements was $8.44 \text{ mmol kg}^{-1}$, which is comparable with the value estimated by NMR spectroscopy ($8.22 \text{ mmol kg}^{-1}$).⁴³ The observed cmc of pure SDS at 313.15 K by conductivity measurements was $7.08 \text{ mmol kg}^{-1}$, whereas in the literature, the cmc was observed to be 8.0 mmol kg^{-1} by dynamic light scattering.⁴⁴ Obtained cmc values of pure SDS in the range of 7.75 to 8.25 mM were reported using different techniques by Baloch *et al.*⁴⁵ The cmc of pure SDS reported by Kumar *et al.* was 8.1 mmol kg^{-1} .⁴⁶ All the literature values support the experimental values obtained herein.

From Fig. 1, it can be observed that the slope in the post-micellar region is lower than that in the pre-micellar region; this occurs due to reduced counterion binding in the stern layer after micellization. Thus, these two slopes can be exploited for the calculation of α by the equation $\alpha = S_2/S_1$, where S_1 and S_2 are the slopes in the pre- and post-micellar regions, correspondingly.^{47,48} Buckingham *et al.*⁴⁹ established the α value valuation *via* the conductivity method, which was confirmed by another research group (Kale *et al.*)⁵⁰ as well as by Bandhopadhyay and Moulik⁵¹ through an ion-selective electrode method. The measurement of the value of α is significant to distinguish the micellar behavior of surfactants. The stability along with the shape changeover of micelles from spherical to rod-like structures is responsible for the viscoelastic behavior of the surfactant and depends on the α value.⁵²⁻⁵⁴ Again, in real applications, where the charge of the micelle surface plays a vital role, *e.g.* DNA transportation, estimation of the α value is important.⁵⁵ The rate of reaction of an organic molecule with a hydrophilic ion while maintaining the binding capability to



Scheme 2 Molecular structure of CFH.



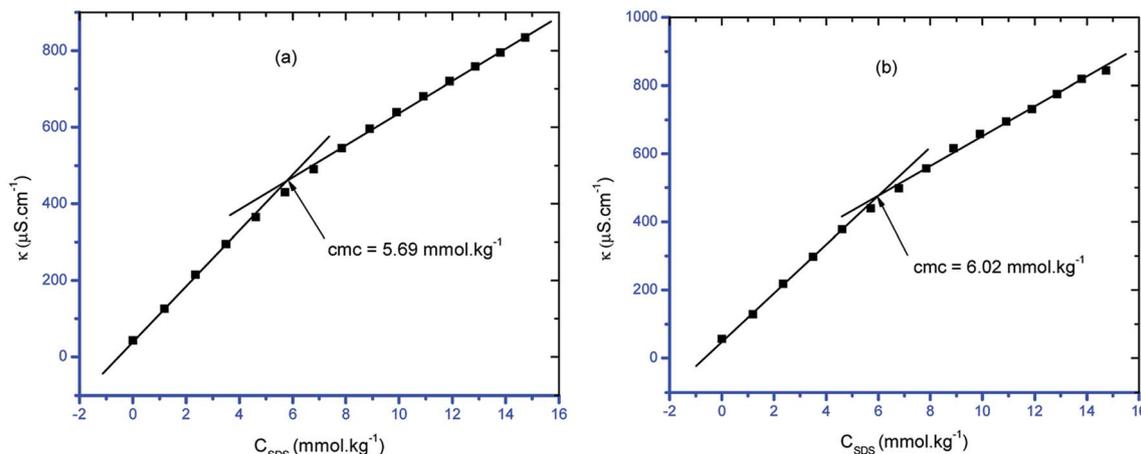


Fig. 1 Representative graphs of κ vs. [SDS] containing 0.5 mmol kg^{-1} CFH in (a) glucose (10 mmol kg^{-1}) and (b) fructose (10 mmol kg^{-1}) at temperature = 303.15 K .

the micelle is critically dependent on the α value.^{52,53} Again, the thermodynamic parameters of the micellization phenomenon are critically dependent on the α value.⁵⁶ The fraction of counterion binding (β) is estimated from the relation $\beta = (1 - \alpha)$.^{57,58}

The cmc and α values observed in our experiments are outlined in Tables 1 and 2. The observed cmc values of SDS were found to dwindle in the presence of CFH; this reduction proceeds *via* enhancement of the CFH concentration in the

Table 1 The various physico-chemical parameters of the studied systems in water and in the presence of polyols^a

C_{CFH} (mmol kg^{-1})	Water			10 mmol kg^{-1} glucose			10 mmol kg^{-1} fructose		
	cmc, mmol kg^{-1}	α	β	cmc, mmol kg^{-1}	α	β	cmc, mmol kg^{-1}	α	β
$T = 298.15 \text{ K}$									
0.0	8.44	0.59	0.41	4.99	0.54	0.46	5.67	0.57	0.43
0.5	8.02	0.60	0.40	5.92	0.58	0.42	6.34	0.61	0.39
1.0	7.67	0.66	0.34	6.33	0.59	0.41	6.89	0.63	0.37
2.0	6.99	0.75	0.25	6.95	0.61	0.39	7.41	0.64	0.36
$T = 303.15 \text{ K}$									
0.0	7.97	0.58	0.42	4.48	0.53	0.47	4.98	0.55	0.45
0.5	7.62	0.61	0.39	5.69	0.57	0.43	6.02	0.60	0.40
1.0	7.40	0.65	0.35	6.01	0.58	0.42	6.43	0.62	0.38
2.0	5.92	0.74	0.26	6.72	0.60	0.40	6.98	0.63	0.37
$T = 308.15 \text{ K}$									
0.0	7.33	0.57	0.43	4.04	0.52	0.48	4.62	0.54	0.46
0.5	7.02	0.60	0.40	5.13	0.56	0.44	5.26	0.59	0.41
1.0	6.81	0.62	0.38	5.66	0.57	0.43	5.92	0.60	0.40
2.0	6.21	0.71	0.29	6.15	0.58	0.42	6.47	0.61	0.39
$T = 313.15 \text{ K}$									
0.0	7.08	0.57	0.43	4.55	0.54	0.46	4.99	0.55	0.45
0.5	6.63	0.59	0.41	5.49	0.57	0.43	5.97	0.60	0.40
1.0	6.42	0.64	0.36	6.04	0.59	0.41	6.51	0.61	0.39
2.0	5.94	0.69	0.31	6.61	0.60	0.40	6.98	0.63	0.37
$T = 318.15 \text{ K}$									
0.0	7.47	0.56	0.44	4.97	0.55	0.45	5.51	0.56	0.44
0.5	6.91	0.62	0.38	5.89	0.59	0.41	6.39	0.61	0.39
1.0	6.56	0.66	0.34	6.46	0.60	0.40	6.82	0.62	0.38
2.0	6.32	0.75	0.25	6.93	0.62	0.38	7.41	0.64	0.36

^a Relative standard uncertainty (u_r) limits are $u_r(\text{cmc}) = \pm 3\%$, $u_r(\alpha) = \pm 4\%$ and $u_r(\beta) = \pm 4\%$.



Table 2 Variations of cmc (mmol kg^{-1}) and degrees of dissociation of SDS + CFH systems with the concentration of polyols^a

C_{Polyols} , mmol kg^{-1}	C_{CFH} , mmol kg^{-1}	Glucose			Fructose		
		cmc/ mmol kg^{-1}	α	β	cmc/ mmol kg^{-1}	α	β
1.00	0.00	5.16	0.57	0.43	5.57	0.58	0.42
5.00	0.00	4.79	0.56	0.44	5.31	0.57	0.43
10.00	0.00	4.48	0.53	0.47	4.98	0.55	0.45
15.00	0.00	4.23	0.52	0.48	4.87	0.54	0.46
20.00	0.00	4.06	0.51	0.49	4.75	0.52	0.48
1.00	0.50	6.83	0.63	0.37	6.97	0.59	0.41
5.00	0.50	6.33	0.61	0.39	6.48	0.59	0.41
10.00	0.50	5.69	0.57	0.43	6.02	0.60	0.40
15.00	0.50	5.35	0.56	0.44	6.91	0.61	0.39
20.00	0.50	5.07	0.55	0.45	7.49	0.62	0.38

^a Relative standard uncertainty (u_r) limits are $u_r(\text{cmc}) = \pm 3\%$, $u_r(\alpha) = \pm 4\%$ and $u_r(\beta) = \pm 4\%$.

water system in the entire studied temperature range. However, the cmc values were found to be enhanced in the presence of CFH in the cases of polyols (glucose or fructose) medium; this continued with further increase of [CFH]. There is an opportunity to obtain a positively charged N atom (N^+ ion) in the structure of CFH by rearrangement, which neutralizes the micellar surface charge and favors micellization. On the other hand, the oxygen of the quinolone group of CFH repels the SO_4^{2-} group of the surfactant; thus, the presence of CFH disfavors micellization. In water, the first factor predominates over the second; thus, the cmc decreases in the presence of CFH and increases in the presence of polyols. For ionic surfactants, the cmc decreases as the temperature augments until it reaches a minimum; subsequently, it increases with further increment of the temperature.⁵⁹

The variation of cmc of SDS/SDS + CFH as a function of temperature was also observed to be U-shaped, *e.g.* cmc

decreases as the temperature increases, reaches a minimum, and then increases with the successive upsurge of temperature (Fig. 2). In aqueous medium, the minimum was observed at 313.15 K, whereas in polyols medium, it was observed at 308.15 K (Table 1). The obtained alteration of cmc of amphiphiles *via* temperature can be explained in the following two ways: (a) the enhanced dehydration of the hydrophilic heads at elevated temperature favors micellization; (b) the enhanced solubility of the surfactant at elevated temperature opposes micellization. These two opposite effects determine whether the cmc of the ionic surfactant will increase or decline at a certain temperature. The obtained U-shaped change of cmc with changing temperature reveals that the first factor is predominant at lower temperature, and reduction of cmc is observed. At elevated temperature, the second factor is predominant over the first one; thus, the cmc is enhanced.

The observed cmc values of SDS and SDS + CFH dwindled in the presence of polyols (glucose/fructose). The hydroxyl groups of the polyols strongly attract water molecules; thus, the solubility of the surfactants is effectively reduced, which increases the hydrophobic interaction amongst the surfactant monomers. Thus, micellization starts to occur at lower concentration, *i.e.* cmc is lower in polyols solution. The attained cmc values follow the order $\text{cmc}_{\text{water}} > \text{cmc}_{\text{fructose}} > \text{cmc}_{\text{glucose}}$ (Table 1 and Fig. 2). The decrease of cmc of the amphiphile in the presence of polyols was also reported by other researchers.³⁴ Again, the observed cmc values of SDS/SDS + CFH decrease as the glucose concentration increases (Fig. 3), whereas the cmc of pure SDS decreases with the enhancement of fructose concentration; however, for SDS + CFH, the cmc value shows U-shaped behavior with changing fructose concentration (Fig. 3). Because the cmc of the SDS/SDS + CFH systems decreases with increasing glucose concentration and dissolution of the drug decreases above cmc, a smaller amount of SDS should be used to formulate the drug for diabetic patients to obtain better drug activity. The attained outcomes indicate that micellization is favored in the presence of CFH in water but disfavored in polyols media. Polyols that are water-soluble can be employed as

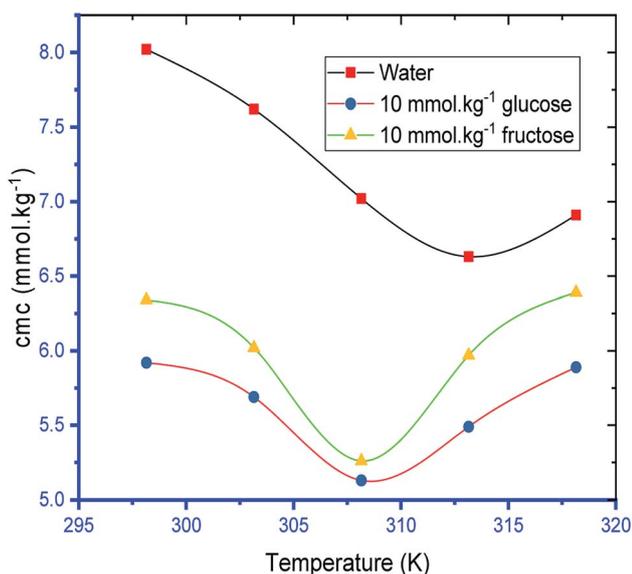


Fig. 2 cmc vs. T for SDS + 0.5 mmol kg^{-1} CFH in different media.



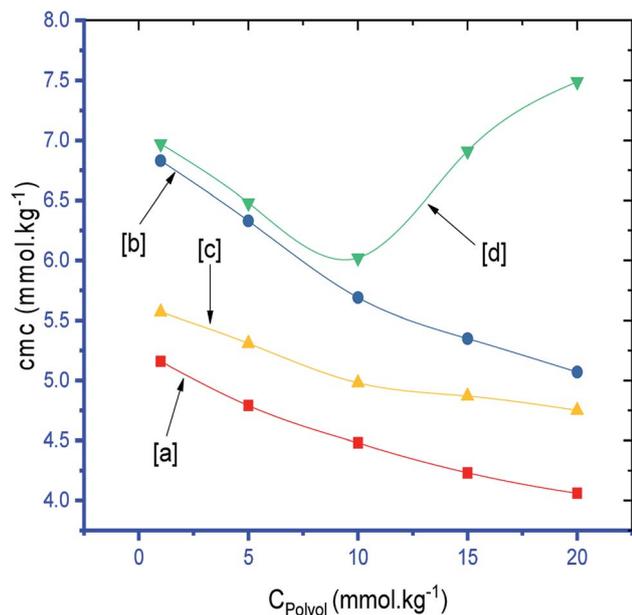


Fig. 3 Effects of polyols concentration on the cmc of the studied systems: (a) pure SDS in glucose, (b) SDS + 0.5 mmol kg⁻¹ CFH in glucose, (c) pure SDS in fructose, (d) SDS + 0.5 mmol kg⁻¹ CFH in fructose.

cosolvents, and these polyols are not incorporated in the micelles. Thus, the addition of polyols to water is the cause of modification of the aqueous phase, which alters the micellar properties of the surfactants. The higher density of polyols compared to that of water is the cause of the increment of the volume fraction of surfactant in polyols solvent, which rises with increasing concentration of polyols. The interlayer spacing of the surfactant is reduced with augmentation of the volume fraction of the surfactant, which reduces the value of cmc in the presence of polyols (Table 1).

Table 1 shows that the values of α of the SDS and SDS + CFH mixed systems were reduced in the presence of polyols in almost all cases. The value of α of the surfactant was perceived to be greater in CFH solution, and the value increased further with increasing [CFH] in water/polyols solvents (Table 1). For pure SDS, the values of α were found to decrease monotonically in aqueous medium, whereas a U-shaped trend was observed in polyols medium with temperature variation. For the SDS + CFH mixture in the aqueous system, the values of α did not show any trend with temperature; however, their values in the presence of polyols decreased initially with temperature, after which the values increased as the temperature increased further (Table 1). The α value decreased with increasing glucose content for both SDS and the SDS + CFH mixture. However, in the presence of fructose, the α value decreased with increasing fructose content in pure SDS solution but was enhanced for the SDS + CFH mixture (Table 2 and Fig. 4).

Because there are no reported values about the behavior of CFH with surfactant in the presence of polyols, especially glucose, the formulation of drugs for diabetic patients is difficult. Again, excess use of surfactants in drug formulations can

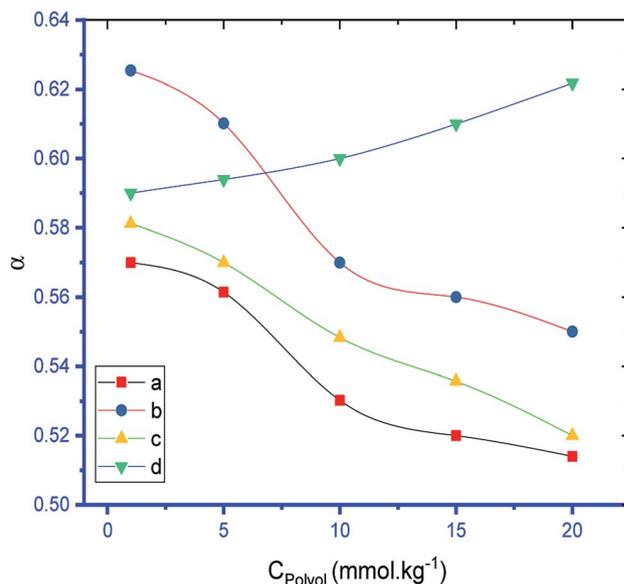


Fig. 4 Variation of the degree of dissociation with the variation of concentration of polyols for (a) SDS in glucose, (b) SDS + 0.5 mmol kg⁻¹ CFH in glucose, (c) pure SDS in fructose, (d) SDS + 0.5 mmol kg⁻¹ CFH in fructose.

cause problems; thus, more study is required to obtain equal activity using less surfactant. The cmc values of the currently employed surfactant along with its mixture with CFH were reduced in polyols solutions; this helps to achieve better drug delivery of a hydrophobic drug by incorporating it in micelles at a lower surfactant concentration and minimizing the use of surfactants. Thus, the findings of our current study provide ideas for drug formulation, especially for diabetic patients, which is very rare in the existing literature in this field.

3.2 Thermodynamic properties of SDS and the SDS + CFH mixture

Thermodynamic parameters are important tools to understand the micellization phenomenon, the interactions between the drug and surfactant and the influence of different additives. Also, the drug delivery and release rate are functions of the molecular interaction of a drug with surfactants; thus, these molecular interactions can be explained in terms of thermodynamic parameters, and the values of different thermodynamic parameters can be utilized in drug formulation to achieve better drug delivery and drug release rates. The spontaneity or non-spontaneity of micellization can be measured from the ΔG_m° values of micellization, which can be executed based on the pseudo-phase partition model⁶⁰⁻⁶³ through the following relation:

$$\Delta G_m^\circ = (1 + \beta)RT \ln X_{\text{cmc}} \quad (1)$$

In eqn (1), X_{cmc} represents the mole fraction of cmc regarding the employed surfactant, while R and T elicit their usual meanings. The enthalpy of micellization (ΔH_m°) for pure SDS along with CFH-mediated micellization of SDS were estimated utilizing the subsequent equation:



$$\Delta H_m^\circ = -(1 + \beta)RT^2(\partial \ln X_{\text{cmc}})/\partial T \quad (2)$$

The alteration of X_{cmc} , which is dependent on temperature, is demonstrated to be a parabolic arc through relation (3):

$$\ln X_{\text{cmc}} = A + BT + CT^2 \quad (3)$$

where the constants A , B and C are obtained from the regression assessment of least squares. Fig. 5 shows the plot of the polynomial fitting arc of $\ln X_{\text{cmc}}$ vs. T , which was subsequently exploited to measure ΔH_m° of the currently studied system. The estimated constant values attained from eqn (3) are summarized in Table S1 (ESI)[†] and were exploited accordingly to measure the values of ΔH_m° through the following relation:^{64–66}

$$\Delta H_m^\circ = -(1 + \beta)RT^2[B + 2CT] \quad (4)$$

The estimated ΔG_m° and ΔH_m° values were subsequently used for the measurement of the entropy (ΔS_m°) under analogous conditions utilizing the following equation:

$$\Delta S_m^\circ = (\Delta H_m^\circ - \Delta G_m^\circ)/T \quad (5)$$

All the thermodynamic parameters evaluated in the current study are summarized in Table 3. The ΔG_m° values for all systems (SDS/SDS + CFH in aqueous as well as in polyols (glucose or fructose) media) were negative, which shows that the micellization phenomena are thermodynamically spontaneous.^{2,11,67} The observed ΔG_m° value of SDS alone was found to be higher than those of the SDS + CFH mixed system both in aqueous and polyols (glucose or fructose) media, which signifies that pure SDS undergoes micellization more spontaneously than the SDS + CFH mixture. The negative ΔG_m° for the micellization of individual SDS in aqueous medium is

enhanced as the temperature elevates, indicating that the association phenomena are additionally spontaneous at the higher studied temperatures; therefore, cmc is lower at higher temperature (Table 1). However, for the SDS + CFH mixture in H₂O, the negative values of ΔG_m° increase initially with temperature, reach a maximum, and then dwindle with the successive upsurge of the temperature. In polyols media, the negativity of ΔG_m° in the cases of the surfactant and the surfactant and CFH mixture increase initially with increasing temperature; after a certain temperature, their values start to decrease with the subsequent increase in temperature, with few exceptions (Table 3). In the aqueous system, the estimated value of ΔH_m° of SDS alone was found to be positive at subordinate temperature; however, on elevating the temperature, the value became negative, which signifies that micellization of SDS in aqueous medium is endothermic and exothermic at lower and higher temperature, respectively. In the case of SDS alone, this type of variation of ΔH_m° can also be found in the literature.³⁹ The ΔH_m° value in the case of CFH-mediated micellization of SDS in H₂O was positive at both lower and higher CFH concentrations, whereas at the intermediate employed CFH concentration, a negative value was obtained; this signifies that the micellization phenomenon is endothermic at lesser and greater concentrations and exothermic at the intermediate concentration (Table 3). The ΔH_m° values for the SDS and SDS + CFH mixed systems in the presence of polyols were found to be positive and negative at lower and elevated temperature, respectively, in almost all cases (Table 3); this implies that micellization of SDS/SDS + CFH is endothermic and exothermic at lower and elevated temperature, respectively. The ΔH_m° value is the outcome of different types of interactions, e.g. hydrophobic as well as hydrophilic interactions, counterion binding and hydration of the polar head groups of the surfactants. Negative values of ΔH_m° arise when hydration of the hydrophilic portion (head groups) of the surfactant dominates the disruption of the H₂O structure around the hydrophobic chains of the monomeric surfactant and *vice versa*. The attained ΔS_m° value for SDS alone was positive in H₂O and declined as the temperature increased; this implies that SDS molecules are arranged in a more orderly fashion at higher temperatures, and therefore micellization is favored and cmc is lowered (Table 1). The ΔS_m° value for SDS + CFH was positive at the lower and higher employed CFH concentrations and negative at the intermediate concentration. Again, in glucose solution, at a lower selected temperature, the ΔS_m° value for the surfactant alone was found to be positive; meanwhile, the value was negative at a higher temperature and positive in all cases for SDS in fructose solution. The positive ΔS_m° values in the presence of polyols decreased with elevation of the temperature. The ΔS_m° values for the SDS + CFH mixed system in the presence of polyols (glucose or fructose) were found to be positive in almost all cases. The attained ΔS_m° values for the SDS + CFH mixture decreased as the temperature was elevated, signifying more ordered SDS + CFH systems at elevated temperature. Positive values of ΔS_m° arise when the hydrophobic portion of the drug transfers from the aqueous vicinity to the micelle interior.⁶⁸ It is well known that the H-bonding of water molecules in the immediate vicinity of

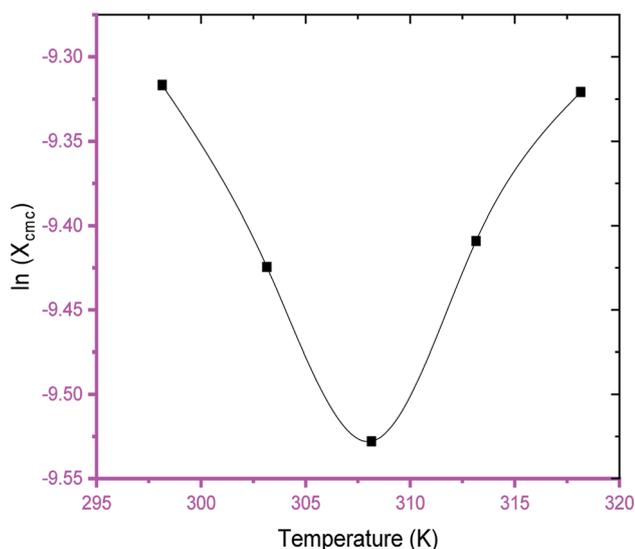


Fig. 5 Representative plot of $\ln X_{\text{cmc}}$ vs. T for pure SDS in 10 mmol kg^{-1} glucose.



Table 3 Thermodynamic parameters (ΔG_m° (kJ mol⁻¹), ΔH_m° (kJ mol⁻¹) and ΔS_m° (J K⁻¹ mol⁻¹) of all studied systems^a

	$\Delta G_m^\circ/\Delta H_m^\circ/\Delta S_m^\circ$				
C_{CFH} mmol kg ⁻¹	298.15 K	303.15 K	308.15 K	313.15 K	318.15 K
Water					
0.0	-30.73/7.49/128.19	-31.67/0.21/105.15	-32.72/-7.69/81.25	-33.38/-16.10/55.19	-33.95/-25.22/27.45
0.5	-7.18/36.58/146.70	-7.41/32.23/130.77	-7.85/28.02/116.39	-8.22/23.39/100.95	-8.04/17.8/81.29
1.0	-7.01/-14.18/-24.06	-7.30/-18.93/-38.37	-7.80/-24.24/-53.32	-8.05/-29.27/-67.76	-7.99/-34.27/-82.62
2.0	-6.79/38.27/151.15	-7.41/31.12/127.09	-7.58/23.84/101.96	-7.95/15.38/74.50	-7.54/5.71/41.65
10 mmol kg⁻¹ glucose					
0.0	-33.72/9.13/143.71	-34.92/-10.71/79.84	-36.13/-32.18/12.82	-35.77/-54.21/-58.89	-35.75/-77.53/-131.34
0.5	-32.19/35.65/227.55	-33.11/25.10/191.99	-34.27/13.61/155.37	-34.33/1.13/113.24	-34.13/-11.90/69.87
1.0	-31.52/-7.97/77.69	-32.68/34.94/223.04	-33.67/25.06/190.60	-33.50/14.02/151.77	-33.55/2.59/113.59
2.0	-30.96/32.70/213.51	-31.82/24.42/185.54	-33.13/15.50/157.84	-32.93/5.51/122.78	-32.81/-4.84/87.91
10 mmol kg⁻¹ fructose					
0.0	-32.57/64.67/326.15	-34.06/48.96/273.84	-35.14/31.34/215.73	-35.17/-12.05/150.79	-35.11/-8.25/84.42
0.5	-31.28/48.67/268.15	-32.21/35.71/224.04	-33.47/21.57/178.61	-33.31/6.14/125.97	-33.35/-10.08/73.12
1.0	-31.73/43.98/253.93	-32.68/34.94/223.04	-32.80/-21.62/36.30	-32.75/-35.77/-9.62	-32.87/-50.59/55.69
2.0	-30.08/32.44/209.66	-31.01/22.26/175.74	-32.25/11.27/141.24	-32.03/-0.82/99.69	-32.09/-13.42/58.68

^a Relative standard uncertainty (u_r) limits are $u_r(\Delta G_m^\circ) = \pm 3\%$, $u_r(\Delta H_m^\circ) = \pm 4\%$ and $u_r(\Delta S_m^\circ) = \pm 5\%$.

a hydrophobic portion is stronger than that of normal water, *i.e.* the H₂O molecules in the immediate vicinity of a hydrophobic moiety attract each other more strongly than normal H₂O molecules; due to tightening of the H₂O structure,⁶⁹ the internal torsional vibration of the hydrophobic chain is reduced. This highly ordered H₂O structure along with the reduced internal torsional vibration leads to the reduction of entropy. The removal of a non-polar moiety (hydrophobic chain) from the aqueous vicinity is entropically favorable, which disrupts the highly ordered H₂O structure.⁶⁸

Taken together, the magnitudes of ΔH_m° and ΔS_m° indicate that micellization of pure SDS is entropically controlled at lower temperature and both entropically and enthalpically controlled at greater temperature in H₂O/fructose solution. In glucose medium, the micellization is governed by both entropy and enthalpy at lower temperature, whereas it is entirely enthalpically controlled at elevated temperature. In the aqueous system, the magnitudes of ΔH_m° and ΔS_m° for the SDS + CFH mixed system indicate that micellization is entropically governed at both lower and higher drug concentrations, but enthalpically governed at the intermediate concentration of the drug. The ΔH_m° and ΔS_m° values for the SDS + CFH mixed system in the presence of glucose/fructose elicits that micellization is governed by entropy at lower temperature but becomes governed by both enthalpy and entropy at elevated temperature. Negative ΔH_m° and positive ΔS_m° values were also observed for SDS in hexanediol + water medium in a microcalorimetric study.⁷⁰ It is reported that positive enthalpy and entropy values of a system reveal the presence of hydrophobic bonding, while negative enthalpy and entropy values are indicative of both hydrogen bonding and electrostatic interactions.^{71,72} Other researchers have reported the presence of hydrophobic interactions between the surfactant and solutes based on negative

enthalpies and positive entropies.⁷³ Thus, the binding forces between SDS and CFH involve hydrophobic interactions as well as electrostatic interactions such as hydrogen bonding and ion-dipole interactions.

3.3 Thermodynamic transfer properties of the SDS/SDS + CFH mixed systems

Diverse thermodynamic transfer properties, *e.g.* free energy of transfer ($\Delta G_{m,tr}^\circ$), enthalpy of transfer ($\Delta H_{m,tr}^\circ$) and entropy of transfer ($\Delta S_{m,tr}^\circ$), during the micellization of SDS/SDS + CFH mixtures in the different employed solvents can be measured from the following equations:^{74,75}

$$\Delta G_{m,tr}^\circ = \Delta G_m^\circ(\text{aq. additive}) - \Delta G_m^\circ(\text{aq.}) \quad (6)$$

$$\Delta H_{m,tr}^\circ = \Delta H_m^\circ(\text{aq. additive}) - \Delta H_m^\circ(\text{aq.}) \quad (7)$$

$$\Delta S_{m,tr}^\circ = \Delta S_m^\circ(\text{aq. additive}) - \Delta S_m^\circ(\text{aq.}) \quad (8)$$

All measured $\Delta G_{m,tr}^\circ$, $\Delta H_{m,tr}^\circ$ and $\Delta S_{m,tr}^\circ$ values in all exploited solvents are provided in Table 4. In the aqueous medium, in the presence of CFH, the magnitude of $\Delta G_{m,tr}^\circ$ was found to be positive; this illustrates the lower spontaneity of micelle formation in the presence of CFH. The $\Delta H_{m,tr}^\circ$ and $\Delta S_{m,tr}^\circ$ values in H₂O were found to be positive at lower and greater concentrations of the drug and negative at an intermediate concentration of the drug. In the presence of polyols (glucose or fructose), the $\Delta G_{m,tr}^\circ$ values obtained were negative for SDS alone at all investigated temperatures, implying the greater spontaneity of micelle formation in the presence of polyols (glucose or fructose). The $\Delta H_{m,tr}^\circ$ and $\Delta S_{m,tr}^\circ$ values for SDS alone in glucose medium were negative at almost all temperatures;



Table 4 Evaluated thermodynamic transfer parameters [$\Delta G_{m,tr}^\circ$ (kJ mol⁻¹), $\Delta H_{m,tr}^\circ$ (kJ mol⁻¹) and $\Delta S_{m,tr}^\circ$ (J K⁻¹ mol⁻¹)] of the currently studied systems^a

C _{CFH} (mmol kg ⁻¹)	T (K)	$\Delta G_{m,tr}^\circ/\Delta H_{m,tr}^\circ/\Delta S_{m,tr}^\circ$	$\Delta G_{m,tr}^\circ/\Delta H_{m,tr}^\circ/\Delta S_{m,tr}^\circ$	$\Delta G_{m,tr}^\circ/\Delta H_{m,tr}^\circ/\Delta S_{m,tr}^\circ$
		Water	10 mmol kg ⁻¹ glucose	10 mmol kg ⁻¹ fructose
0.0	298.15		-2.99/1.64/15.52	-1.85/57.18/197.96
0.0	303.15		-3.25/-10.92/-25.31	-2.39/48.75/168.69
0.0	308.15		-3.40/-24.49/-68.43	-2.41/39.03/134.48
0.0	313.15		-2.38/-38.11/-114.09	-1.79/28.15/95.59
0.0	318.15		-1.80/-52.32/-158.78	-1.16/16.97/56.97
0.5	298.15	23.54/29.08/18.59	-1.47/28.16/99.36	-0.55/41.18/139.96
0.5	303.15	24.26/32.03/25.63	-1.44/24.89/86.85	-0.54/35.50/118.90
0.5	308.15	24.88/35.71/35.14	-1.55/21.30/74.12	-0.74/29.26/97.36
0.5	313.15	25.17/39.49/45.75	-0.95/17.23/58.05	0.08/22.24/70.77
0.5	318.15	25.91/43.04/53.84	-0.18/13.32/42.42	0.60/15.13/45.67
1.0	298.15	23.72/-21.67/-152.25	-1.01/36.4/125.74	0.18/-3.00/-10.66
1.0	303.15	24.37/-19.14/-143.52	-1.01/34.73/117.89	0.15/-8.18/-27.45
1.0	308.15	24.92/-16.55/-134.57	-0.95/32.75/109.35	-0.08/-13.93/-44.95
1.0	313.15	25.33/-13.17/-122.96	-0.12/30.13/96.57	0.63/-19.67/-64.81
1.0	318.15	25.96/-9.06/-110.07	0.41/27.81/86.14	1.08/-25.37/-83.14
2.0	298.15	23.94/30.78/22.96	-0.23/25.21/85.33	0.65/24.94/81.48
2.0	303.15	24.26/30.91/21.94	-0.16/24.21/80.39	0.66/22.06/70.60
2.0	308.15	25.14/31.52/20.71	-0.41/23.19/76.59	0.47/18.96/59.99
2.0	313.15	25.43/31.48/19.31	0.45/21.61/67.58	1.35/15.29/44.50
2.0	318.15	26.41/30.93/14.21	1.14/20.38/60.46	1.86/11.79/31.23

^a Relative standard uncertainty (u_r) limits are $u_r(\Delta G_{m,tr}^\circ) = \pm 3\%$, $u_r(\Delta H_{m,tr}^\circ) = \pm 4\%$ and $u_r(\Delta S_{m,tr}^\circ) = \pm 5\%$.

however, these values were positive in fructose solution. The $\Delta H_{m,tr}^\circ$ and $\Delta S_{m,tr}^\circ$ values for SDS + CFH were positive in glucose medium at all temperatures and CFH concentrations employed. Again, the $\Delta H_{m,tr}^\circ$ and $\Delta S_{m,tr}^\circ$ values for the SDS + CFH mixtures were positive at lower and greater concentrations of CFH and negative at intermediate concentration (Table 4).

3.4 Enthalpy–entropy compensation

The linear relationship between ΔH_m° and ΔS_m° is called enthalpy–entropy compensation and can be assessed by exploiting the following relation:^{76,77}

$$\Delta H_m^\circ = \Delta H_m^{\circ,*} + T_C \Delta S_m^\circ \quad (9)$$

In eqn (9), the intercept $\Delta H_m^{\circ,*}$ and the slope T_C indicate the intrinsic enthalpy and compensation temperature, respectively. A representative graph of ΔH_m° vs. ΔS_m° is presented in Fig. 6. The solute–solute and solute–solvent interactions can be explained by the assessed magnitudes of $\Delta H_m^{\circ,*}$ and T_C , respectively. Enthalpy–entropy compensation was observed both for the surfactant alone and for its mixture with CFH in water/polyols media, and the attained values of the compensation parameters are summarized in Table S2 (ESI).† The estimated $\Delta H_m^{\circ,*}$ value was found to be negative in all systems (SDS/SDS + CFH) in the absence/presence of polyols (Table S2 (ESI)†). The obtained negative $\Delta H_m^{\circ,*}$ value indicates that micellization is privileged even at $\Delta S_m^\circ = 0$. The augmentation of the negative

$\Delta H_m^{\circ,*}$ value signifies enhancement of the micelle stability.^{78,79} The estimated values of T_C were in the range of 288 K to 345 K. The obtained values of T_C in the range of 270 K to 330 K can be exploited to study the effects of H₂O in the protein solution.⁸⁰ Thus, the estimated values are comparable with biological fluids in almost all cases.

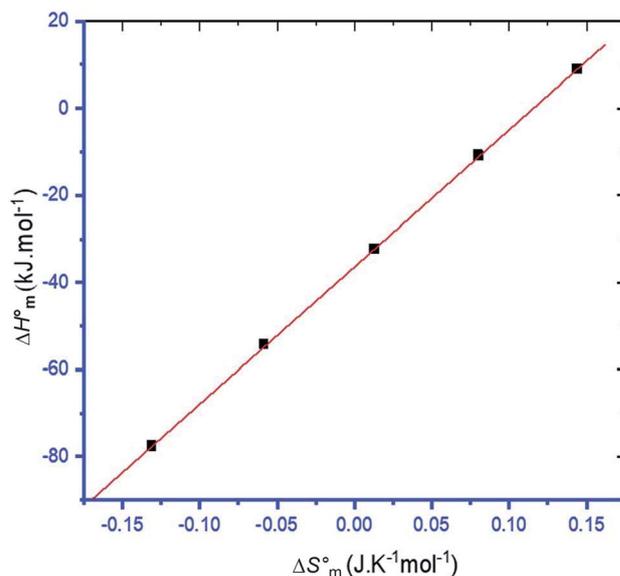


Fig. 6 Enthalpy–entropy compensation plot for pure SDS in 10 mmol kg⁻¹ glucose solution.



4. Conclusions

A conductometric study has been performed herein to study the self-aggregation properties of SDS and a mixture of SDS + CFH in the absence/presence of polyols (glucose or fructose) at different temperatures. The decrease in cmc in the presence of CFH is due to the establishment of additional hydrophobic interactions between the hydrophobic moieties of SDS and CFH. The addition of glucose/fructose enhances the self-aggregation of SDS + CFH. Micellar parameters such as cmc and α of the SDS + CFH mixture were observed to be dependent on the additive (polyols) and temperature variation. The ΔC_m^o , ΔH_m^o and ΔS_m^o values for SDS and the SDS + CFH mixture reveal that the micellization process is thermodynamically spontaneous and that the interaction forces between SDS and CFH are hydrophobic, hydrogen bonding and ion-dipole type. The greater negative values of ΔH_m^{o*} disclose the formation of more stable micelles in glucose/fructose compared to in aqueous medium. This study will aid the formulation of the drug CFH using the most commonly utilized surfactant, SDS, while considering different target patients, including diabetic patients, to achieve maximum activity of the drug; however, more study is still required to consider all the ingredients in body fluid during drug formulation. Because glucose reduces the cmc of SDS and the SDS + CFH mixture, the use of SDS can be reduced for a particular set of formulations. Also, the SDS + CFH mixture can be further studied in additives media which are present in body fluid using SEM/TEM for morphological studies and MD simulations to observe the interaction sites.

Conflicts of interest

There are no conflicts of interest to declare.

Acknowledgements

This project was funded by the Deanship of Scientific Research (DSR), King Abdulaziz University, Jeddah, Saudi Arabia under grant no. (KEP-51-130-38). The authors, therefore, acknowledge with thanks DSR technical and financial support.

References

- D. Kumar and M. A. Rub, *RSC Adv.*, 2019, **9**, 22129.
- D. Kumar and M. A. Rub, *J. Phys. Org. Chem.*, 2016, **29**, 394.
- D. Kumar and M. A. Rub, *J. Surfactants Deterg.*, 2019, **22**, 1299.
- U. Ashraf, O. A. Chat, M. Maswal, S. Jabeen and A. A. Dar, *RSC Adv.*, 2015, **5**, 83608.
- M. Akram, S. Anwar, F. Ansari, I. A. Bhat and Kabir-ud-Din, *RSC Adv.*, 2016, **6**, 21697.
- M. Akram, I. A. Bhat and Kabir-ud-Din, *RSC Adv.*, 2015, **5**, 102780.
- D. Kumar, S. Hidayathulla and M. A. Rub, *J. Mol. Liq.*, 2018, **271**, 254.
- S. Das, S. Mondal and S. Ghosh, *RSC Adv.*, 2016, **6**, 30795.
- P. Shi, H. Zhang, L. Lin, C. Song, Q. Chen and Z. Li, *RSC Adv.*, 2019, **9**, 3224.
- D. Kumar and M. A. Rub, *J. Mol. Liq.*, 2019, **274**, 639.
- M. A. Rub, N. Azum, F. Khan and A. M. Asiri, *J. Chem. Thermodyn.*, 2018, **121**, 199.
- S. He, X. Liu, P. Yan, A. Wang, J. Su and X. Su, *RSC Adv.*, 2019, **9**, 4908.
- D. Kumar and M. A. Rub, *J. Mol. Liq.*, 2018, **250**, 329.
- D. Kumar and M. A. Rub, *J. Phys. Org. Chem.*, 2019, **32**, e3918.
- V. Bhardwaj, T. Bhardwaj, K. Sharma, A. Gupta, S. Chauhan, S. S. Cameotra, S. Sharma, R. Guptad and P. Sharma, *RSC Adv.*, 2014, **4**, 24935.
- M. F. Ahmed, M. R. Molla, M. Saha, I. Shahriar, M. S. Rahman, M. A. Halim, M. A. Rub, M. A. Hoque and A. M. Asiri, *RSC Adv.*, 2019, **9**, 6556.
- M. Fresta, S. Guccione, A. R. Beccari, P. M. Furneri and G. Puglisi, *Bioorg. Med. Chem.*, 2002, **10**, 3871.
- J. N. Israelachvili, *Intermolecular and Surface Forces*, Academic Press, New York, 1995.
- Z. Huang, X. Zhang, X. Zhang, C. Fu, K. Wang, J. Yuan, L. Tao and Y. Wei, *Polym. Chem.*, 2015, **5**, 607.
- H. Huang, M. Liu, R. Jiang, J. Chen, L. Mao, Y. Wen, J. Tian, N. Zhou, X. Zhang and Y. Wei, *J. Colloid Interface Sci.*, 2018, **513**, 198.
- J. Chen, M. Liu, Q. Huang, L. Huang, H. Huang, F. Deng, Y. Wen, J. Tian, X. Zhang and Y. Wei, *Chem. Eng. J.*, 2018, **337**, 82.
- R. Jiang, M. Liu, T. Chen, H. Huang, Q. Huang, J. Tian, Y. Wen, Q. Cao, X. Zhang and Y. Wei, *Dyes Pigm.*, 2017, **148**, 52.
- X. Zhang, X. Zhang, B. Yang, M. Liu, W. Liu, Y. Chen and Y. Wei, *Polym. Chem.*, 2014, **5**, 356.
- J. Piret, A. Désormeaux and M. G. Bergeron, *Curr. Drug Targets*, 2002, **3**, 17.
- J. Piret, J. Lamontagne, J. Bestman-Smith, S. Roy, P. Gourde, A. Désormeaux, R. F. Omar, J. Juhász and M. G. Bergeron, *J. Clin. Microbiol.*, 2000, **38**, 110.
- M. F. Islam, E. Rojas, D. M. Bergey, A. T. Johnson and A. G. Yodh, *Nano Lett.*, 2003, **3**, 269.
- S. Knapp, *Gerontology*, 2013, **59**, 99.
- G. C. Koh, S. J. Peacock, T. van der Poll and W. J. Wiersinga, *Eur. J. Clin. Microbiol. Infect. Dis.*, 2012, **31**, 379.
- H. Liao, J. Zakhaleva and W. Chen, *Biomaterials*, 2009, **30**, 1689.
- S. Mori, H. K. Takahashi, K. Liu, H. Wake, J. Zhang, R. Liu, T. Yoshino and M. Nishibori, *Br. J. Pharmacol.*, 2010, **161**, 229.
- L. Boyanova and I. Mitov, *Expert Rev. Anti-Infect. Ther.*, 2013, **11**, 411.
- M. A. Rub, N. Azum, S. B. Khan, H. M. Marwani and A. M. Asiri, *J. Mol. Liq.*, 2015, **212**, 532.
- M. A. Ahsan, M. R. Amin, S. Mahbub, M. R. Molla, S. Aktar, M. N. Arshad, M. A. Hoque, M. A. Rub and M. A. Khan, *Chin. J. Chem. Eng.*, 2020, **28**, 216.
- M. R. Amin, S. Mahbub, S. Hidayathulla, M. M. Alam, M. A. Hoque and M. A. Rub, *J. Mol. Liq.*, 2018, **269**, 417.



- 35 M. A. Hoque, M. M. Alam, M. R. Molla, S. Rana, M. A. Rub, M. A. Halim, M. A. Khan and A. Ahmed, *J. Mol. Liq.*, 2017, **244**, 512.
- 36 S. Mahbub, M. A. Rub, M. A. Hoque and M. A. Khan, *J. Phys. Org. Chem.*, 2018, **31**, e3872.
- 37 S. Mahbub, M. Rahman, S. Rana, M. A. Rub, M. A. Hoque, M. A. Khan and A. M. Asiri, *J. Surfactants Deterg.*, 2018, **22**, 137.
- 38 S. Mahbub, M. A. Rub, M. A. Hoque and M. A. Khan, *J. Phys. Org. Chem.*, 2019, **32**, e3917.
- 39 S. Mahbub, M. R. Molla, M. Saha, I. Shahriar, M. A. Hoque, M. A. Halim, M. A. Rub and M. A. Khan, *J. Mol. Liq.*, 2019, **283**, 263.
- 40 S. Mahbub, M. A. Rub, M. A. Hoque, M. A. Khan and D. Kumar, *J. Phys. Org. Chem.*, 2019, **32**, e3967.
- 41 M. A. Hoque, M.-O.-F. Patoary, M. M. Rashid, M. R. Molla and M. A. Rub, *J. Solution Chem.*, 2017, **46**, 682.
- 42 F. Akhtar, M. A. Hoque and M. A. Khan, *J. Chem. Thermodyn.*, 2008, **40**, 1082.
- 43 G. Manzo, M. Carboni, A. C. Rinaldi, M. Casu and M. A. Scorciapino, *Magn. Reson. Chem.*, 2013, **51**, 176.
- 44 K. Ogino, T. Kubota, H. Uchiyama and M. Abe, *J. Jpn. Oil Chem. Soc.*, 1988, **37**, 588.
- 45 M. K. Baloch, G. Hameed and A. Bano, *J. Chem. Soc. Pak.*, 2002, **24**, 77.
- 46 S. Kumar, Z. A. Khan, N. Parveen and Kabir-ud-Din, *Colloids Surf., A*, 2005, **268**, 45.
- 47 D. Kumar and M. A. Rub, *J. Mol. Liq.*, 2017, **238**, 389.
- 48 S. M. I. H. Sristy, S. Mahbub, M. M. Alam, M. A. Rub and M. A. Hoque, *J. Mol. Liq.*, 2019, **284**, 12.
- 49 S. A. Buckingham, C. J. Garve and G. G. Warr, *J. Phys. Chem.*, 1993, **97**, 10236.
- 50 K. M. Kale, E. L. Cussler and D. F. Evans, *J. Phys. Chem.*, 1980, **84**, 593.
- 51 A. Bandhopadhyay and S. P. Moulik, *Colloid Polym. Sci.*, 1988, **266**, 455.
- 52 L. S. Romsted, *Rate Enhancements in Micellar Systems*, PhD thesis, Indiana University, Bloomington, IN, 1975.
- 53 V. Soldi, J. Keiper, L. S. Romsted, I. M. Cuccovia and H. Chaimovich, *Langmuir*, 2000, **16**, 59.
- 54 R. Oda, J. Narayanan, P. A. Hassan, C. Manohar, R. A. Salkar, F. Kern and S. J. Candau, *Langmuir*, 1998, **14**, 4364.
- 55 Y. Wang, P. L. Dubin and H. Zhang, *Langmuir*, 2001, **17**, 1670.
- 56 D. G. Hall, *J. Chem. Soc., Faraday Trans. 1*, 1981, **77**, 1121.
- 57 J. M. Kuiper, R. T. Buwalda, R. Hulstand and J. B. F. N. Engberts, *Langmuir*, 2001, **17**, 5216.
- 58 L. Wang, Y. Zhang, L. Ding, J. Liu, B. Zhao, Q. Deng and T. Yan, *RSC Adv.*, 2015, **5**, 74764.
- 59 G. C. Kresheck, *Water: A Comprehensive Treatise*, ed. F. Franks, Plenum, New York, vol. 4, 1975.
- 60 M. Rahman, M. A. Khan, M. A. Rub and M. A. Hoque, *J. Mol. Liq.*, 2016, **223**, 716.
- 61 Z. Medoš and M. B. Rogač, *J. Chem. Thermodyn.*, 2015, **83**, 117.
- 62 F. Khan, M. A. Rub, N. Azum, D. Kumar and A. M. Asiri, *J. Solution Chem.*, 2015, **44**, 1937.
- 63 M. R. Molla, S. Rana, M. A. Rub, A. Ahmed and M. A. Hoque, *J. Surfactants Deterg.*, 2018, **21**, 231.
- 64 H.-U. Kim and K.-H. Lim, *Colloids Surf., A*, 2004, **235**, 121.
- 65 A. Beesley, D. F. Evans and R. G. Laughlin, *J. Phys. Chem.*, 1988, **92**, 791.
- 66 B. Bergenstaahl and P. Stenius, *J. Phys. Chem.*, 1987, **91**, 5944.
- 67 L. Qin and X.-H. Wang, *RSC Adv.*, 2017, **7**, 51426.
- 68 D. Attwood and A. T. Florence, *Surfactant Systems*, Chapman and Hall London, New York, 1985.
- 69 P. Taboada, P. M. Landeira, J. M. Ruso, M. Garcia and V. Mosquera, *Colloids Surf., A*, 2002, **197**, 95.
- 70 J. W. Comeau, A. A. McLachlan and D. G. Marangoni, *J. Dispersion Sci. Technol.*, 2009, **30**, 1288.
- 71 E. Pramauro and E. Pelizzetti, *Surfactants in Analytical Chemistry: Applications of Organized Media*, in *Comprehensive Analytical Chemistry*, ed. S. G. Weber, Elsevier, Amsterdam, 1996.
- 72 A. Beesley, D. F. Evans and R. G. Laughlin, *J. Phys. Chem.*, 1988, **92**, 791.
- 73 V. Bhardwaja, P. Sharma, M. S. Chauhan and S. Chauhan, *J. Saudi Chem. Soc.*, 2016, **20**, S109.
- 74 S. K. Shivaji and A. K. Rakshit, *J. Surfactants Deterg.*, 2004, **7**, 305.
- 75 M. R. Amin, S. Mahbub, M. R. Molla, M. M. Alam, M. F. Hossain, S. Rana, M. A. Rub, M. A. Hoque and D. Kumar, *J. Chem. Eng. Data*, 2019, **64**, 2750.
- 76 L. J. Chen, S. Y. Lin and C. C. Huang, *J. Phys. Chem.*, 1998, **102**, 4350.
- 77 S. Mahbub, M. A. Rub and M. A. Hoque, *J. Chem. Eng. Data*, 2019, **64**, 4181.
- 78 G. Sugihara and M. Hisatomi, *J. Colloid Interface Sci.*, 1999, **219**, 31.
- 79 M. A. Hoque, M. O. F. Patoary, M. R. Molla, M. A. Halim, M. A. Khan and M. A. Rub, *J. Dispersion Sci. Technol.*, 2017, **38**, 1578.
- 80 R. Lumry and S. Rajender, *Biopolymers*, 1970, **9**, 1125.

