


Cite this: *RSC Adv.*, 2020, 10, 5874

Synthesis of anti-allergic drugs

Shiyang Zhou^{ab} and Gangliang Huang^{ib}*^a

Histamine is formed by the decarboxylation of histidine catalyzed by enzymes. It is an endogenous biologically active substance involved in multiple complex physiological processes as an important chemical transmitter. Histamine receptors have four subtypes, H₁, H₂, H₃ and H₄, all of which are G protein coupling receptors (GPCRs) with different physiological functions. Histamine plays an important role in the pathophysiological mechanism of allergic diseases, and the antagonistic effect of histamine has become an important way to study anti-allergic drugs, wherein the anti-allergic drugs used in clinical practice are mainly H₁ receptor antagonists. Currently, there are many varieties of H₁ receptor antagonists in clinical applications, which can be divided into ethylenediamine antagonists, amino ether antagonists, propylamine antagonists, tricyclic antagonists, piperazine antagonists and piperidine antagonists depending on their chemical structures. This article mainly reviews the research progress of allergic reactions with histamine H₁ receptor antagonists and expounds the important aspects of the design and synthesis of various new compounds.

Received 18th December 2019
Accepted 20th January 2020

DOI: 10.1039/c9ra10659f

rsc.li/rsc-advances

1. Introduction

Histamine, chemically known as 4(5)-(2-aminoethyl)imidazole, is an endogenous biologically active substance involved in multiple complex physiological processes as an important chemical transmitter. Histamine is formed by the decarboxylation of histidine catalyzed by enzymes.^{1–6} It is widely found in a variety of plants, animals and microorganisms in nature.^{7–9} In animals, histamines are found in peripheral tissues and in peripheral and central nerves. Histamine is released under endogenous and exogenous stimulations and interacts with receptors. Histamine receptors have four subtypes, H₁, H₂, H₃ and H₄, all of which are G protein coupling receptors (GPCRs) with different physiological functions.^{10–13}

H₁ receptors are distributed in endothelial cells, smooth muscle, adrenal medulla, heart and central nervous system. Exogenous antigens can cause anaphylaxis, combine with antibody immunoglobulin on target cell mast cell nucleus and granulocyte, change the function of target cell membrane, release histamine and other allergic media, and make histamine distribute in H₁ receptor function of tissues and organs.¹⁴ After the activation of the H₁ receptor, phosphatidase C is activated through G proteins, which promotes the increase in Ca²⁺ concentration, thereby increasing the vasodilation and capillary permeability, leading to plasma exudation, local tissue redness and swelling, and allergic symptoms of bronchial and

gastrointestinal smooth muscle contraction.^{15–17} H₂ receptors are distributed in gastric wall cells, vascular smooth muscles, heart and central nervous system. H₂ receptor stimulation could promote gastric acid secretion, stimulate the heart and inhibit uterine contraction.¹⁸ Histamine, acetylcholine and gastrin receptors could be found on the surface of gastric mucosa cells. When the corresponding ligands interact with these receptors, they could activate the secretion of gastric acid. The H₃ receptor is located in the central and peripheral nerve endings of the presynaptic membrane, and it could function as their receptor, inhibiting histamine release and synthesis.^{19–23} H₃, as a different receptor, controls the release of other neurotransmitters. H₃ receptor has important physiological functions, such as cardiac function, gastric acid secretion, anaphylaxis, sleep wake cycle control, cognition and memory, convulsion, diarrhea or reducing gastric acid secretion.^{24–30} The H₄ receptor is only found in the small intestine, spleen, thyroid and immunocompetent cells. The H₄ receptor is related to the regulation of immune functions, such as allergic reactions, asthma, cancer and other immune diseases.^{31–35}

Anaphylaxis is an allergic disease that occurs when the body is exposed to specific allergens.^{36–39} It could be caused by a variety of exogenous substances like allergens that include heterologous serum (such as tetanus antitoxin), certain animal proteins (such as that of fish, shrimp, and crabs), bacteria, viruses, parasites, animal fur, plant pollen, dust mites in the air, and chemicals and drugs. Allergens could stimulate human B cells to produce immunoglobulin E, which combines with antibodies on human mast cells and sensitized cells, damages the cell membrane and leads to degranulation, and releases histamine, 5-hydroxytryptamine, leukotrienes, bradykinin and other active substances in the granules.^{40–46} The allergic reaction

^aChongqing Key Laboratory of Green Synthesis and Application, Active Carbohydrate Research Institute, Chongqing Normal University, Chongqing, 401331, China

^bKey Laboratory of Tropical Medicinal Resource Chemistry of Ministry of Education, College of Chemistry and Chemical Engineering, Hainan Normal University, Haikou, 571158, China. E-mail: huangdoctor226@163.com

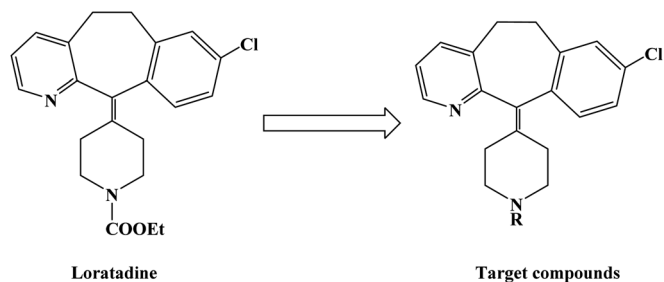



Fig. 1 The structural modification of loratadine analogues 4a–h.

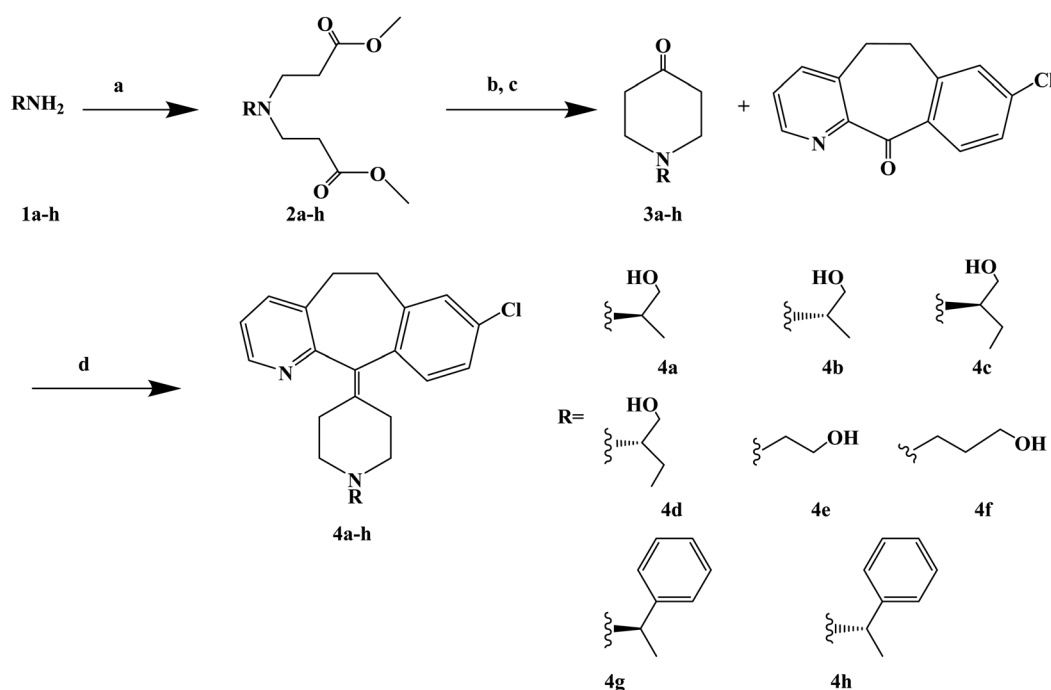
can manifest as skin redness and swelling, rash, itching, patches as well as allergic rhinitis, bronchial asthma, laryngeal edema and other symptoms. The clinical manifestations of anaphylaxis include decreased blood pressure, increased heart rate, pale skin, edema, and shock and even death in severe cases.^{47–51}

Histamine plays an important role in the pathophysiological mechanism of allergic diseases, and the antagonistic effect of histamine has become an important way to study anti-allergic drugs.^{52–55} Currently, the anti-allergic drugs used in clinical practice are mainly H₁ receptor antagonists.^{56–60} H₁ receptor antagonists were first reported in 1933 to relieve bronchospasm in animals induced by the inhalation of excessive histamine. Since the 1930s, scientists around the world have conducted a large number of structural modifications by using structure activity relationship (SAR) studies, and dozens of classic anti-allergy drugs have been listed for clinical use. In the history of drug development, they have been listed as the first generation

H₁ receptor antagonists. The second generation H₁ receptor antagonists were marketed after the 1980s. These compounds have characteristics of high selectivity towards the H₁ receptor, no sedation effect, separation of the antihistamine effect from the central nervous system and fewer side effects. They were known as non-sedation H₁ receptor antagonists.⁶¹ The third generation H₁ receptor antagonists are active optical isomers of the second generation H₁ receptor antagonists or their metabolites. This generation has greater safety and lower toxic and side effects; thus, their application scope has been expanding in recent years.^{62–65} Currently, there are many varieties of H₁ receptor antagonists in clinical application, which can be divided into ethylenediamine antagonists, amino ether antagonists, propylamine antagonists, tricyclic antagonists, piperazine antagonists and piperidine antagonists depending on their chemical structures.^{66–70} This article mainly reviews the research progress of allergic reactions with histamine H₁ receptor antagonists (2011–2017), including the research on the design and synthesis of various new compounds.

2. Synthesis

Loratadine is a second-generation antihistamine because it has fewer central nervous system and anticholinergic side effects. Loratadine has a strong therapeutic effect because of its semi-rigid conformation. Lewis *et al.* have reported a series of compounds based on loratadine that have both H₁ receptor antagonist activity and 5-lipoxygenase inhibitory activity. Liu *et al.* successfully synthesized a series of loratadine derivatives with C-alkyl butenamide, and found that several of them were able to inhibit histamine induction. Yue W. *et al.*⁷¹ further



Scheme 1 General synthesis of loratadine analogues 4a–h. Reagents and conditions: (a) methyl acrylate, MeOH, 40 °C, 12 h; (b) MeONa/MeOH 30% aq., toluene, 110 °C, 5 h; (c) HCl, 100 °C, 5 h; (d) TiCl₄/Zn, THF dry, 70 °C, 17 h.

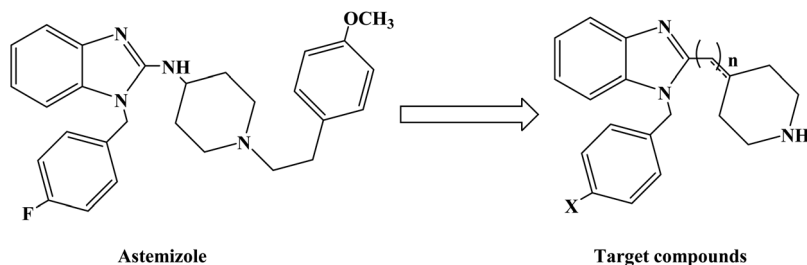


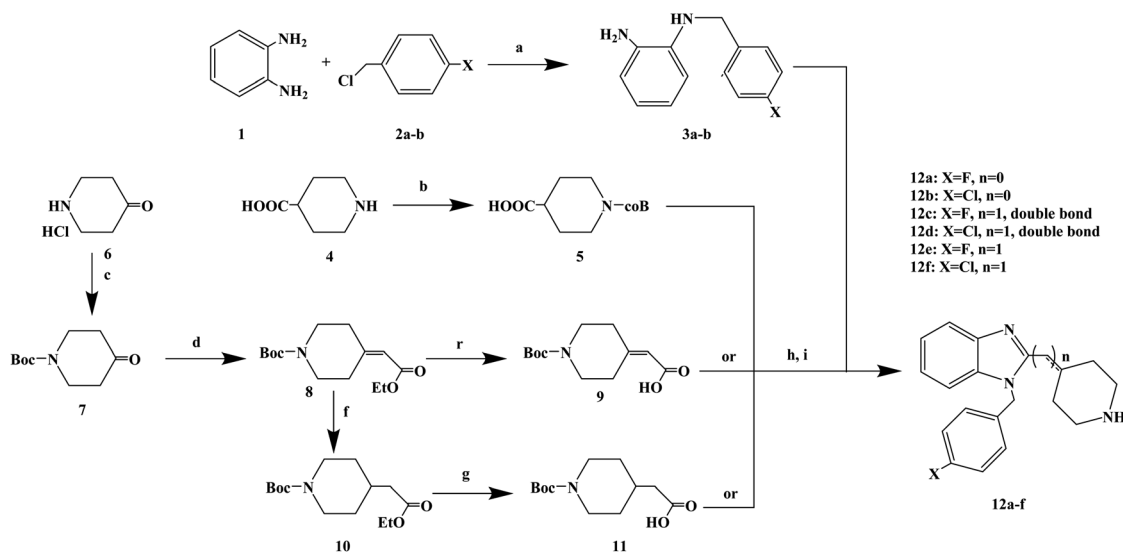
Fig. 2 The structural modification of benzimidazole derivatives 12a–f.

optimized the structure of loratadine based on previous studies, and designed and synthesized a series of loratadine derivatives (Fig. 1). In structural optimization, hydroxyl groups and chiral centers were introduced into the molecules to enhance their affinity for H_1 receptors. In the synthetic process (Scheme 1), a series of chiral hydroxyl groups containing *N*-substituted-4-piperidone were obtained by Michael addition. Subsequently, a Dieckmann cyclization and decarboxylation were performed with chiral or non-chiral aminoalcohols and methyl acrylate as raw materials, affording a yield of 35%. A variety of carbonyl groups were obtained by coupling piperidone with a tricyclic ketone using a low-priced titanium catalyst to obtain a series of target compounds at a yield of approximately 25%. The synthetic route is characterized by mild reaction conditions, good versatility and simple operation.

Astemizole (Fig. 2) is an effective histamine H_1 receptor antagonist ($K_i = 2 \text{ nmol L}^{-1}$). However, it could lead to arrhythmias with acquired long QT syndrome, which includes torsades DE pointes, and sudden death due to the blockage of rapidly activated delayed rectification K^+ current I_{Kr} and its underlying human ether-a-gogo related gene (hERG) K^+

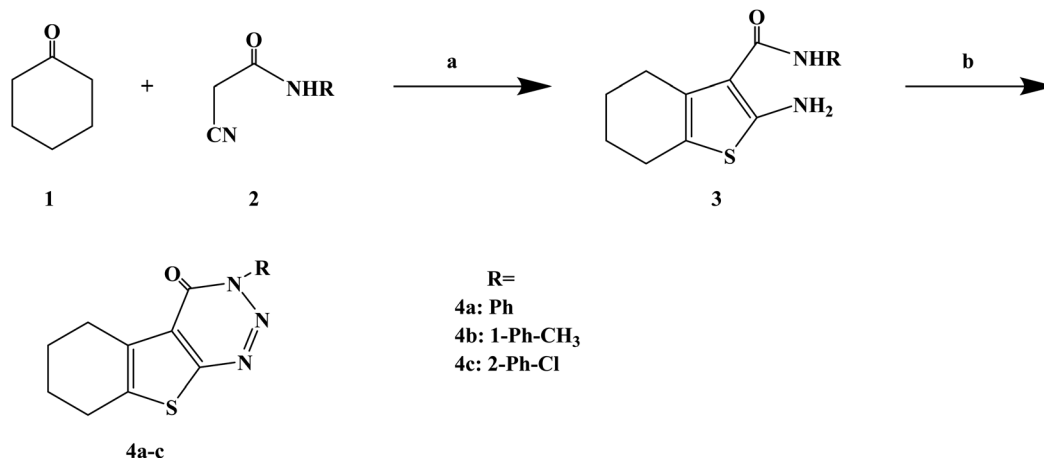
channel ($IC_{50} = 0.9 \text{ nmol L}^{-1}$). In order to find a new type of H_1 receptor antagonist with low hERG blocking activity, Xiao J. *et al.*⁷² carried out systematic structural modification based on astemizole (Fig. 2). The synthetic route of the benzimidazole derivative 12a–12f was summarized in Scheme 2. In addition, compounds 9 and 11 were synthesized from compound 6 according to previous literature. The compounds 12a–12f were produced by following acylation, cyclation, and deprotection procedures. The synthetic route was characterized by mild reaction conditions, high yield and good versatility.

Numerous uses of substituted triazines have been reported, such as anti-inflammatory, analgesic activity, purine confrontation, antitumoral and trypanocidal activity. Very little studies have been conducted on heterocyclic compounds such as 1, 2, 3-triazine and other triazines, this kind of compounds and similarity based on the structure of these compounds and other triazines like 1,2,4-triazine and 1,3,5-triazin. It was found that 1,2,3-triazine derivatives possess prostaglandin inhibition property, analgesic, anti-inflammatory and antihistaminic properties. Gollapalle L. *et al.*⁷³ further optimized 1,2,3-triazine



Scheme 2 General synthesis of benzimidazole derivatives 12a–f. Reagents and conditions: (a) K_2CO_3 , DMF, 50 °C, 6 h; (b) Boc_2O , 1 mol L^{-1} NaOH, THF, 25 °C, 2 h; (c) Boc_2O , 2 mol L^{-1} $NaHCO_3$, THF, 25 °C, 6 h; (d) ethyl diethoxyphosphoryl acetate, K_2CO_3 , DMF, 78 °C, 12 h; (e) 1 mol L^{-1} NaOH, CH_3OH , 25 °C, 30 min; (f) 10% Pd/C, H_2 , C_2H_5OH , 25 °C, 7 h; (g) 1 mol L^{-1} NaOH, CH_3OH , 25 °C, 30 min; (h) ethyl chloroformate, Et_3N , CH_2Cl_2 , 0 °C, 30 min; (i) AcOH, reflux, 2 h.





Scheme 3 General synthesis of 3H-benzo[4,5]thieno[2,3-d][1,2,3]triazin-4-ones **4a–c**. Reagents and conditions: (a) CH₃COOH/CH₃COONH₄, glacial acetic/cyclohexane, reflux 10 h; (b) NaNO₂/HCl, glacial acetic, 0–5 °C.

on the basis of previous studies, and designed and synthesized three new compounds with antihistamine effects. In the process of synthesis (Scheme 3), the starting compounds 2-amino-3-(*N*-substituted carboxamido)-4 and 5-tetramethylene thiophenes (compounds **3a–c**) were synthesized in three steps by adapting the Gewald reaction. Afterwards, compounds **3a–c** were diazotized to yield a series of 3-substituted amino-5,6-tetramethylene thieno[2,3-*d*][1,2,3]-triazin-4(3*H*)-ones (compounds **4a–c**). In this reaction, the starting compounds 2-amino-3-(*N*-substituted carboxamido)-4 and 5-tetramethylene thiophenes (compounds **3a–c**) react with NaNO₂ in the presence of HCl yielding the respective triazine-4-ones. The synthetic route was characterized by mild reaction conditions, simple operation and cheap and easy to obtain reagents.

1-(4-Fluorobenzyl)-2-(4-(4-methyl-1*H*-pyrazol-1-yl)piperidin-1-yl)-1*H*-benzo[*d*]imidazole (lead compound **1**, Fig. 3) is a high

affinity and selective H₁-antihistamine. The lead compound **1** exhibits limitations of poor metabolic stability and solubility. Further optimization of compound **1** led to the development of (*R*)-1-(4-fluorobenzyl)-2-(1-methylpiperidine-3-acyl)-1*H*-benzimidazole (lead compound **2**, Fig. 3) with high affinity for H₁-antihistamine. The lead compound **2** showed improved selectivity and stability for hERG channel when compared with lead compound **1**. Satheesh B. *et al.*⁷⁴ performed relevant structure activity relationship (SAR) studies, determining that a change in the direction of the basic center in lead compound **2** was a key step in establishing the required H₁ affinity and selectivity. Based on this discovery, Satheesh B. *et al.* designed and synthesized a series of H₁-antihistamines (Fig. 3) after structural optimization of the lead compounds **1** and **2**. In this process, different substituents R¹ were introduced into the imidazole ring. The polar group R² was added to the piperidine

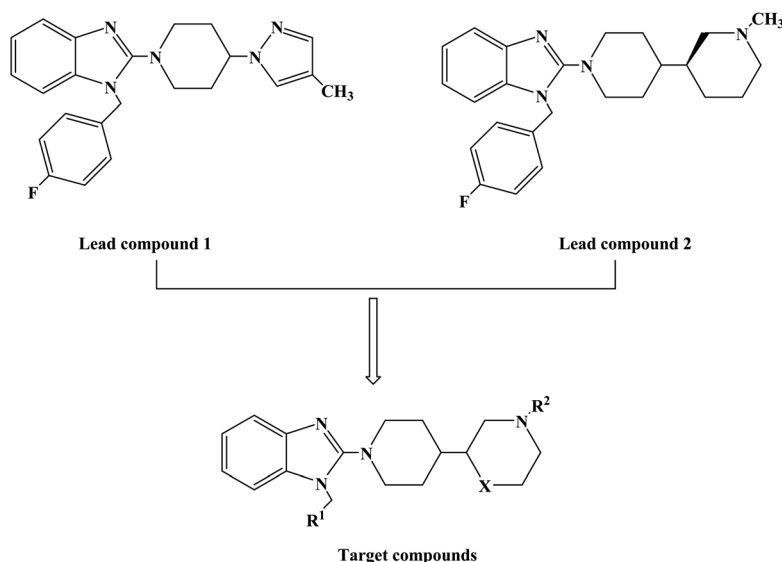
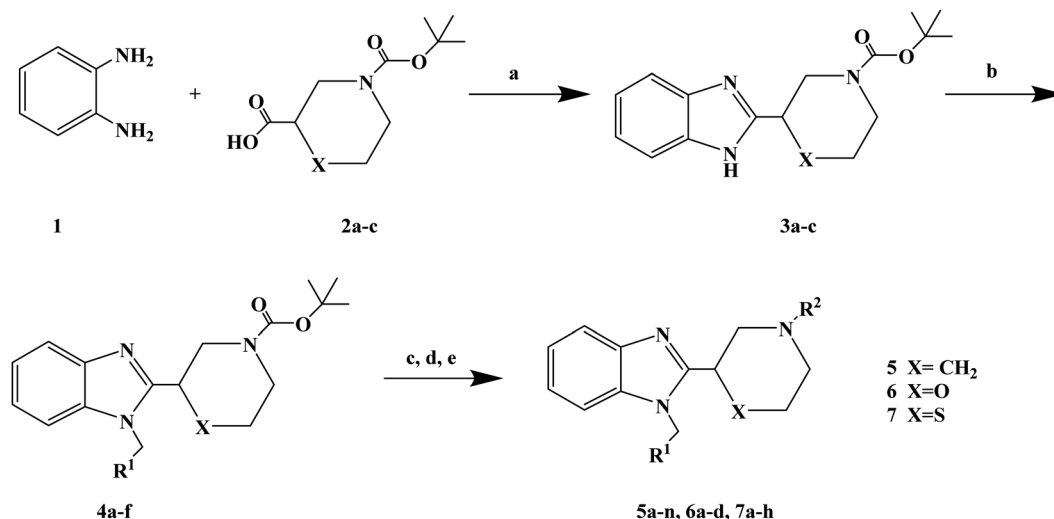


Fig. 3 The structural modification of 2-(piperidin-3-yl)-1*H*-benzimidazole analogues **5a–n**, **6a–d** and **7a–h**.

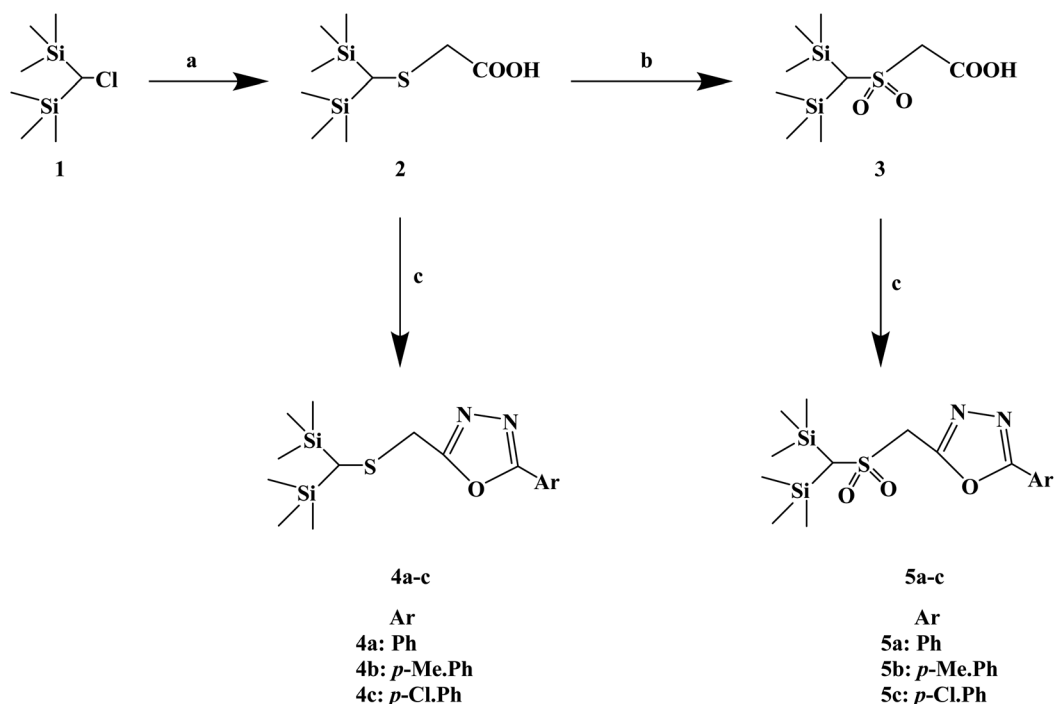


Scheme 4 General synthesis of 2-(piperidin-3-yl)-1H-benzimidazole analogues **5a–n**, **6a–d** and **7a–h**. Reagents and conditions: (a) EtOH, 120 °C; (b) R¹CH₂Br, K₂CO₃, DMF, 80 °C; (c) TFA, CH₂Cl₂; (d) R²–(=O)H, Na(OAc)₃BH or R₂X, Et₃N, THF; (e) separation of enantiomers by chiral SFC when chiral acids were not used.

ring nitrogen, hence, pK_a and/or log *P* were reduced by the electron attraction effect. The synthesis of analogues described in Scheme 4 relies on the formation of the benzimidazole core (compound **3**), after the coupling of a suitably protected amino acid (compound **1**) with *O*-phenylene diamine (compound **2**). Introduction of the appropriate arene-containing moiety R¹ was accomplished by alkylation of compound **3**. The final step included the alkylation or reductive amination and the removal

of Boc to obtain the final product (compounds **5a–n**, **6a–d** and **7a–h**). Further separation of enantiomers was achieved by chiral SFC when chiral acids were not used for the synthesis of benzimidazole. The synthetic route was characterized by mild reaction conditions, cheap and easy to obtain reagents, relatively high yield and good versatility.

Silicon substitution (C/Si exchange) has attracted extensive attention in drug-like scaffold materials due to its unique



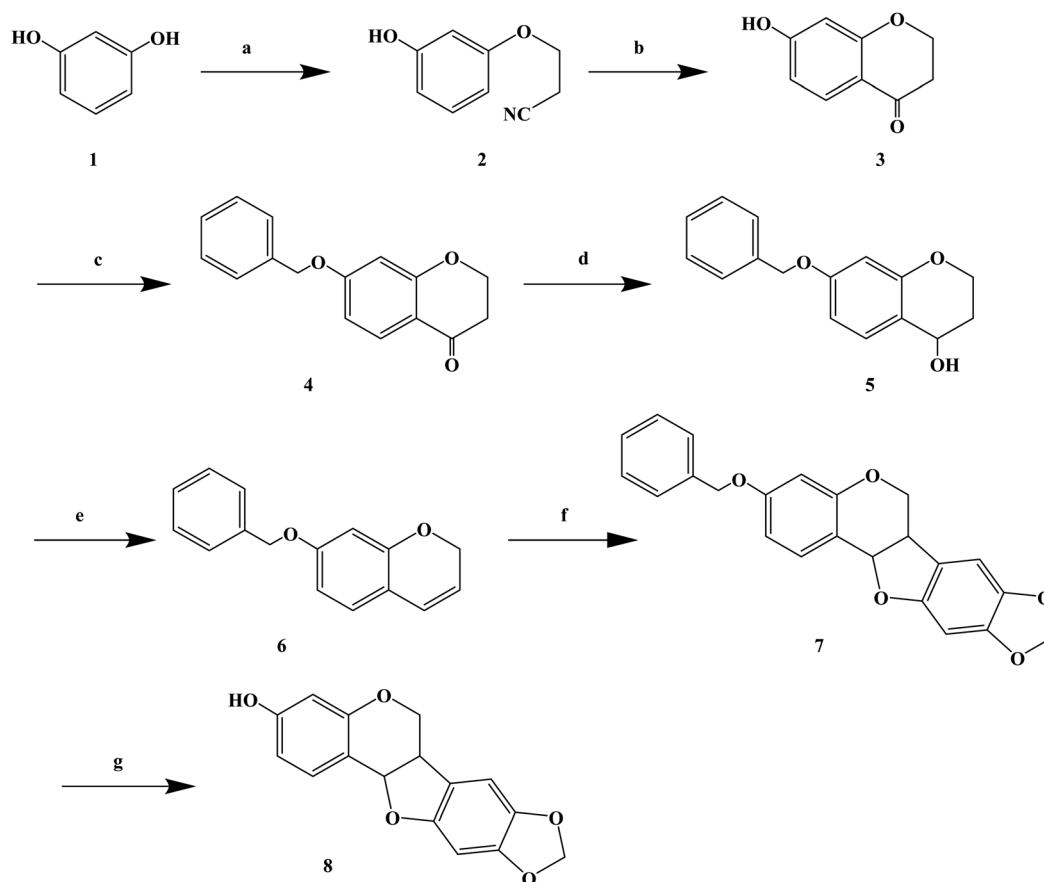
Scheme 5 General synthesis of 2-((bis(trimethylsilyl) methylthio/methylsulfonyl) methyl)-5-aryl-1,3,4-oxadiazoles. Reagents and conditions: (a) HSCH₂COOH, MeOH, NaOH, reflux 4 h; (b) H₂O₂, AcOH, 24 h; (c) ArCONHNH₂, POCl₃, reflux 6 h.



biological activity, chemical reactivity and physical and chemical properties compared with the parent carbon compounds. 1,3,4-Oxadiazole derivatives have been reported as promising antiallergic drugs. Dinneswara R. *et al.*⁷⁵ designed and synthesized a variety of substituted 1,3,4-oxadiazole compounds (Scheme 5) in a study related to silicon substitution drugs, and applied them to the study of antiallergic drugs. In the structural design process, -Ph, 4-Me-Ph and 4-Cl-Ph substituents were selected. In the process of synthesis, 2-(bis(trimethylsilyl)methylthio) acetic acid (compound 2) was synthesized by the reaction of bis(trimethylsilyl)chloromethane (compound 1) and mercaptoacetic acid in the presence of NaOH/MeOH. Compound 3 was subsequently oxidized in the presence of H₂O₂/CH₃COOH. The required heterocyclic compounds, 2-((bis(trimethylsilyl)methylthio)methyl)-5-aryl-1,3,4-oxadiazoles (compound 4) and 2-((bis(trimethylsilyl)methylsulfonyl)methyl)-5-aryl-1,3,4-oxadiazoles (compound 5) were synthesized by the reaction of compound 2/compound 3 with benzo-hydrazides in the presence of phosphorus oxychloride. The synthetic route has high reaction yields (79% to 88%), mild reaction conditions and conventional reagents.

Over the years, Kujin has been widely used to treat allergic diseases and many others. Phytochemical studies have shown that it contains quinolizidine, alkaloids, flavonoids, and

triterpenoids. Kujin and its active components have many biological properties, such as anti-inflammatory, antiasthmatic, antitumor and antibacterial. Hiroyuki M. *et al.*⁷⁶ found that the extract of Kujin has inhibited the up-regulation of H1R and IL-4 gene expression in rats sensitized by TDI, and confirmed (*R*)-maackiain as the main anti-allergy component of Kujin. (*R*)-Maackiain is found in low concentration in plants, which made it difficult to extract; so, it was obtained by chemical synthesis (Scheme 6). In the process of synthesis, maackiain bis(benzonitrile) palladium(II) dichloride [PdCl₂(PhCN)₂] was prepared as reported previously. The synthesis of (*R*)-3-benzylmaackiain was performed as follows. To a solution of 7-benzyloxy-2*H*-1-benzopyran and 2-bromo-4,5-methylene dioxypheyl in anhydrous DMF suspended in potassium acetate, PdCl₂(PhCN)₂ was added in one portion. The resulting suspension was stirred at 40 °C. After 24 h, the addition of PdCl₂(PhCN)₂ to the resulting reaction mixture was carried out. The latest procedure was repeated after 48 h and 72 h. Finally, the resulting mixture was stirred for additional 24 h. This synthetic route achieved the first chemical synthesis of (*R*)-maackiain, but it had disadvantages, such as long synthetic route, low yield, the use of precious metal Pd in the synthesis process, and long overall reaction time, which need to be further optimized.



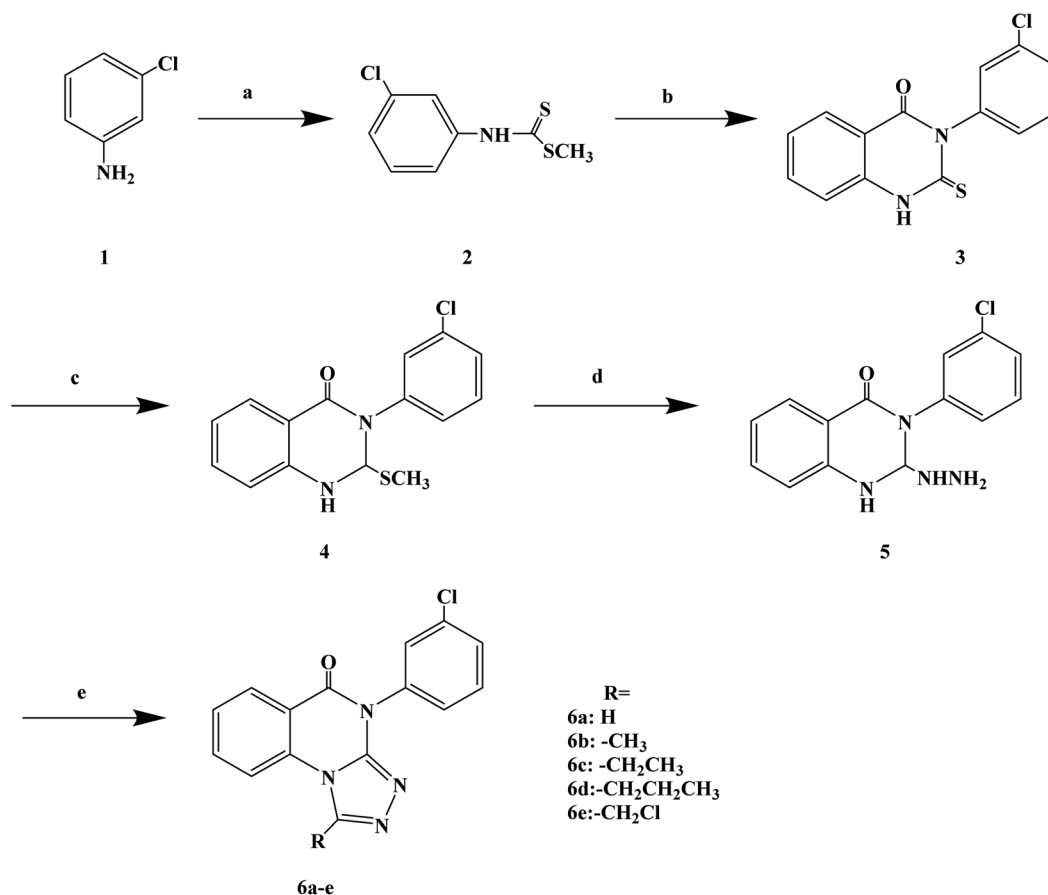
Scheme 6 General synthesis of (±)-maackiain. Reagents and conditions: (a) acrylonitrile, triton B, NaOH, 100 °C, 10 h; (b) H₂SO₄, CH₃COOH, H₂O, 100 °C, 8 h; (c) benzyl chloride, NaI, K₂CO₃, acetone, 60 °C, 18 h; (d) NaBH₄, ethanol, rt, 3 h; (e) *p*-toluenesulfonic acid, toluene, 110 °C, 2 h; (f) PdCl₂(PhCN)₂, KoAc, DMF, 40 °C, 4 days; (g) H₂-Pd/C, 3 h.



Quinazolinones have a wide range of biological and pharmacological effects, such as analgesic, anti-inflammatory, antifungal, antiviral and antihistamine activities. Veerachamy A. *et al.* proved that quinazolinone derivatives have strong antihistamine activity with low sedative effect. Manavalan G. *et al.*⁷⁷ designed and synthesized a series of 1-substituted-4-(3-chlorophenyl)-[1,2,4]triazolo[4,3-*a*]quinazolin-5(4*H*)-ones (Scheme 7) based on the structural modifications of quinazolinone. In that process, the triazolo group has been substituted with different groups (-H, -CH₃, -CH₂CH₃, -(CH₂)₂CH₃, -CH₂Cl) to obtain compounds with improved biological activity. In the process of synthesis, compound 3 (3-(3-chlorophenyl)-2-thioxo-quinazolin-4(3*H*)-one) was belonged to the key intermediate. Dithiocarbamic acid methyl ester was prepared with 3-chloro aniline, carbon disulfide and sodium hydroxide in dimethyl sulfoxide, then methylated with dimethyl sulfate, which was reacted to produce the required compound 4 in ethanol has a good yield (86%). Compound 3 was dissolved in a solution of 2% ethanol sodium hydroxide, stirred at room temperature, and methylated with dimethyl sulfate to give 3-(3-chlorophenyl)-2-(methylthio) quinazolin-4(3*H*)-one (compound 4). The nucleophilic displacement of the methylthio group of

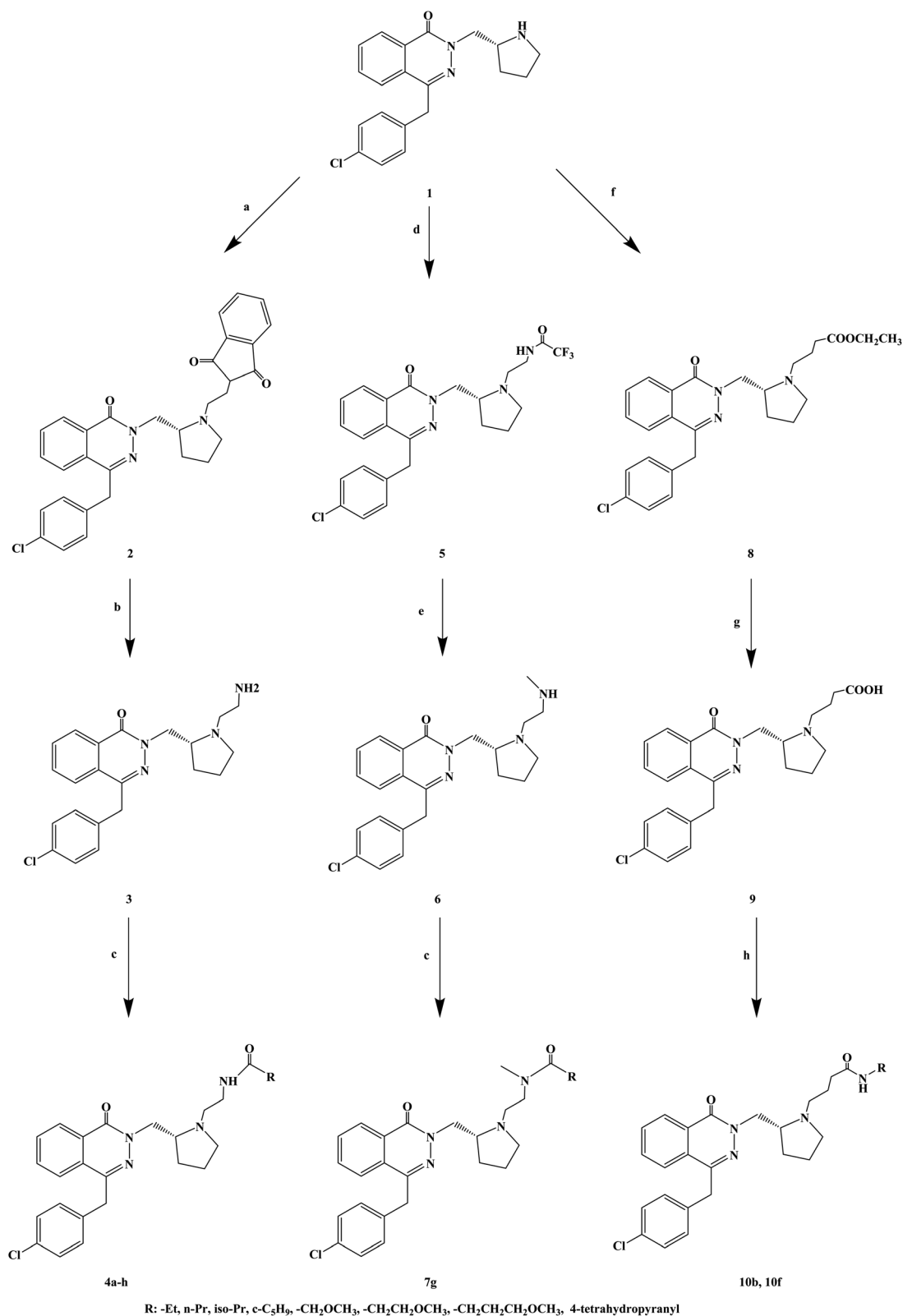
compound 4 with hydrazine hydrate was carried out using ethanol as the solvent, affording compound 5 (3-(3-chlorophenyl)-2-hydrazinyl-quinazolin-4(3*H*)-one). Finally, the compound 5 with a variety of one carbon donors such as acetic acid, propionic acid, formic acid, butyric acid and chloroacetyl chloride under refluxing provided 1-substituted-4-(3-chlorophenyl)-[1,2,4]triazolo[4,3-*a*]quinazolin-5(4*H*)-ones (compounds 6a-e). The synthetic route has high reaction yield (80% to 85%), mild reaction conditions and uses conventional reagents.

The compounds containing the phthalazinone scaffold were obtained through structural optimization and subsequently used as histamine H₁ receptor antagonists for the treatment of allergic reactions. These compounds bind to both H₁ and H₃ receptors. Based on the structure optimization of phthalazinone, Panayiotis A. *et al.*⁷⁸ introduced different substituents, and designed and synthesized a series of novel phthalazinone amide compounds (Scheme 8). In the molecular structure design, -Et, *n*-Pr, iso-Pr, *c*-C₅H₉, -CH₂OCH₃, -CH₂CH₂OCH₃, -CH₂CH₂CH₂OCH₃, 4-tetrahydropyranyl were selected as the substituent. During the synthetic procedure, compound 1 was alkylated with *N*-(2-bromoethyl) phthalimide to give compound 2 (yield 87%), and then, deprotected using



Scheme 7 General synthesis of 1-substituted-4-(3-chlorophenyl)-[1,2,4]triazolo[4,3-*a*]quinazolin-5(4*H*)-ones 6a-e. Reagents and conditions: (a) CS₂, NaOH, stirring 30 min, (CH₃)₂SO₄; (b) methyl anthranilate/EtOH, reflux 19 h; (c) NaOH/EtOH, string 2 h; (d) NH₂NH₂, H₂O/EtOH, reflux 24 h; (e) reflux 29 h.





Scheme 8 General synthesis of phthalazinone amide **3a–g**, **7a–g** and **10a–h**. Reagents and conditions: (a) *N*-(2-bromoethyl)phthalimide, K₂CO₃, 2-butanone, 80 °C, 18 h; (b) NH₂NH₂·H₂O, EtOH, 80 °C, 1.25 h; (c) RCO₂H, TBTU, Et₃N, DMF, 2 h, or RCOCl, Et₃N, DCM; (d) 2,2,2-trifluoro-*N*-(2-iodoethyl)acetamide, DIPEA, 2-butanone; (e) (i) MeI, K₂CO₃, DMF; (ii) K₂CO₃, H₂O, MeOH; (f) BrCH₂CH₂CH₂CO₂Et, K₂CO₃, DMF, 150 °C, microwave, 110 min; (g) NaOH, H₂O, MeOH, 20 °C, 1 h; (h) TBTU, Et₃N, DMF.

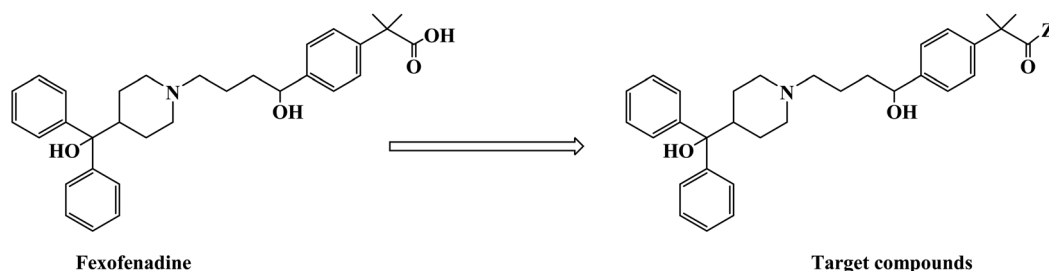
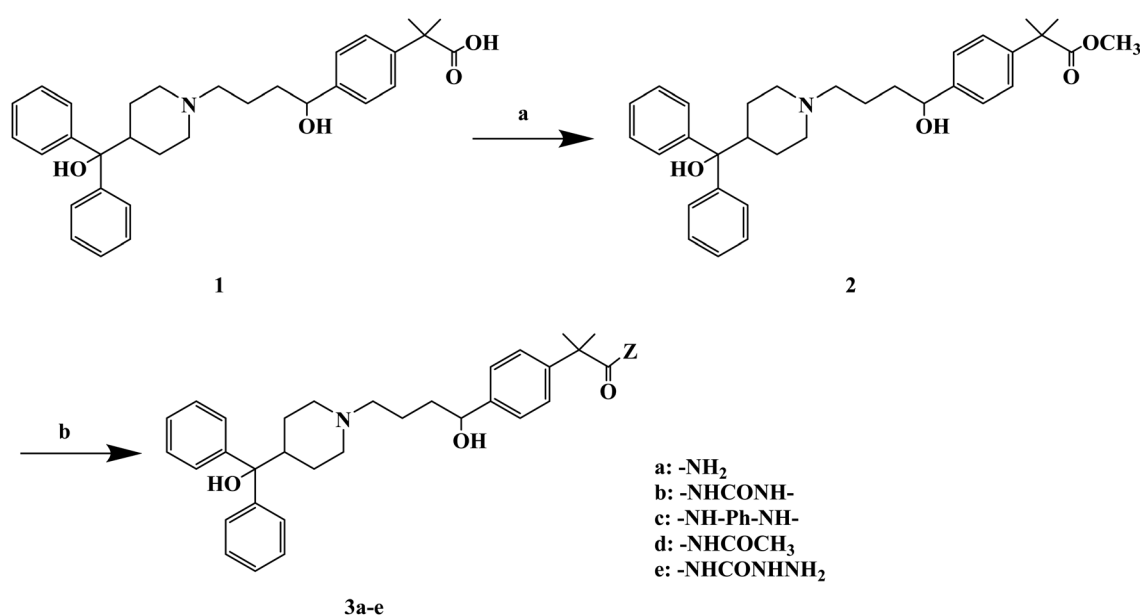


Fig. 4 The structural modification of fexofenadine derivatives **3a–e**.

hydrazine to afford compound **3** with 77% yield. Afterwards, compound **3** was acylated with a variety of carboxylic acids in the presence of TBTU to give compounds **4d**, **4f**, **4g** and **4h**, or with the corresponding acid chloride to give compounds **4a**, **4b**, **4c** and **4e**. Alkylation of compound **1** with 2,2,2-trifluoro-*N*-(2-iodoethyl)acetamide resulted in trifluoroacetamide (compound **5**), which was alkylated with iodomethane and deprotected, to obtain the secondary amine (compound **6**). The latter was acylated in the presence of 4-methoxybutanoic acid and TBTU to obtain tertiary amide (compound **7g**). Compound **1** reacted with ethyl 4-bromobutyrate to give compound **8**, which was converted to a carboxylic acid **9**, and finally, transformed into amides **10b** and **10f**. The synthetic route has good versatility and uses conventional reagents, but the partial yield was low (yield 29%), which should be further improved.

H₁ antagonists are competitive drugs that inhibit histamine action on tissues containing H₁. Fexofenadine (2-[4-[1-hydroxy-4-[4-(hydroxydiphenylmethyl)piperidino]butyl]phenyl]-2-methylpropanoic acid) (Fig. 4) is a selective and peripherally acting H₁ receptor antagonist and is a non-sedative active

metabolite of terfenadine. Fexofenadine was clinically used to treat seasonal allergic rhinitis, but there were associated adverse effects such as headache, throat irritation, viral infection, nausea, dysmenorrhea, drowsiness, dyspepsia and fatigue. Saeed A *et al.*⁷⁹ used fexofenadine as the lead compound (Fig. 4), and designed and synthesized fexofenadine derivatives with hydrazine hydrochloride, urea, *p*-phenylenediamine, acetamide and semicarbazide (Scheme 9). Herein, carboxylbenadine 2-methylpropionamide and benadrylamide derivatives were synthesized using carboxylbenadine as the nucleophilic substituent and tertiary C-19 was replaced by a benzene ring. The carboxyl group on the tertiary C-19 was esterified by a condensation reaction, and the nucleophilic substitution reaction on the acyl group formed a tetrahedral intermediate, which transformed back to the carbonyl group after the leaving group was eliminated. On the basis of the above reaction mechanism, various amines were reacted to produce amide compounds, and various reactions were identified. This synthetic route was characterized by a high yield, short reaction time and good versatility.



Scheme 9 General synthesis of fexofenadine derivatives **3a–e**. Reagents and conditions: (a) MeOH, H₂SO₄/H₂O, reflux 7–8 h; (b) amine, reflux 2–3 h.



3. Conclusions

Histamine plays an important role in the pathophysiological mechanism of allergic diseases, and the antagonistic effect of histamine has become an important way to study anti-allergic drugs. The anti-allergic drugs used in clinical practice were mainly H₁ receptor antagonists. This article mainly reviews the research progress of allergic reactions with histamine H₁ receptor antagonists and expounds the important aspects of design, synthesis and biological activity of various new compounds. These novel H₁ receptor antagonists, with regards to their chemical structure design, are mostly obtained by optimizing the structure of the reported compounds with antiallergic activity (including natural products, Schemes 6 and 9) or already sold drugs. These compounds have novel structures, and the design of H₁ receptor antagonists in the future will tend to optimize the structure of natural products as lead compounds. In the process of synthesis, most synthetic routes use conventional solvent reaction to synthesize target compounds. These synthetic routes were characterized by simple operation, mild reaction conditions, cheap reagents and a good overall yield, which will provide certain foundation for future industrialization. However, some synthetic routes use multiple catalysts or precious metals (such as Pd, Scheme 6), or the reaction time was too long (Scheme 7). In the selection of synthesis methods, researchers should focus on the modern synthesis methods, such as microwave synthesis, and the organic synthesis process will have more room for development. As can be seen from the reagent and reaction condition perspective, the choice direction in the future will be to use cheap and readily available raw materials, conventional solvents, no catalyst or conventional catalyst, and medium or low reaction temperature.

Conflicts of interest

The authors declare that they have no conflict of interest.

Acknowledgements

The project was sponsored by the Scientific Research Foundation for the Returned Overseas Chinese Scholars, State Education Ministry (No. 2015-1098). The work was also supported by Chongqing Key Research Project of Basic Science & Frontier Technology (No. cstc2017jcyjBX0012), Foundation Project of Chongqing Normal University (No. 14XY020), Chongqing General Research Program of Basic Research and Frontier Technology (No. cstc2015jcyjA10054), Chongqing Normal University Postgraduate's Research and Innovation Project (No. YKC17004), the National Natural Science Foundation (21662012, 41866005), Postgraduate Research and Innovation Project of Hainan Normal University (Hsyx2018-8), and Open Foundation Project of Key Laboratory of Tropical Medicinal Resource Chemistry of Ministry of Education (RDZH2019002), China.

References

- 1 R. Leurs, M. Church and M. Taglialatela, H₁-antihistamines: inverse agonism, anti-inflammatory actions and cardiac effects, *Clin. Exp. Allergy*, 2002, **32**(4), 489–498.
- 2 F. Simons, Advances in H₁-antihistamines, *N. Engl. J. Med.*, 2004, **351**(21), 2203–2217.
- 3 G. Khilnani and A. Khilnani, Inverse agonism and its therapeutic significance, *Indian J. Pharmacol.*, 2011, **43**(5), 492–501.
- 4 E. Nettis, M. Colanardi, L. Barra, *et al.*, Levocetirizine in the treatment of chronic idiopathic urticaria: a randomized, double-blind, placebo-controlled study, *Br. J. Dermatol.*, 2006, **154**(3), 533–558.
- 5 G. Howell, L. West, C. Jenkins, *et al.*, In vivo antimuscarinic actions of the third generation antihistaminergic agent, desloratadine, *BMC Pharmacol.*, 2005, **5**, 13.
- 6 G. Vena, N. Cassano, M. Filieri, *et al.*, Fexofenadine in chronic idiopathic urticaria: a clinical and immunohistochemical evaluation, *Int. J. Immunopathol. Pharmacol.*, 2002, **15**(3), 217–224.
- 7 M. Rosmilah, Characterization of major allergens of royal jelly *Apis mellifera*, *Trop. Biomed.*, 2008, **25**(3), 243–251.
- 8 M. Bublin, C. Radauer, I. Wilson, *et al.*, Cross-reactive N-glycans of Api g 5, a high molecular weight glycoprotein allergen from celery, are required for immunoglobulin E binding and activation of effector cells from allergic patients, *FASEB J.*, 2003, **17**(12), 1697–1699.
- 9 W. Horne, M. Lopez, J. Salvaggio, *et al.*, Basidiomycete allergy: identification and characterization of an important allergen from *Calvatia cyathiformis*, *Int. Arch. Allergy Appl. Immunol.*, 1991, **94**(1–4), 359–361.
- 10 E. Stephen, A. Raftery and P. Dowding, Forecasting spore concentrations: a time series approach, *Int. J. Biometeorol.*, 1990, **34**(2), 87–89.
- 11 M. Dhillon, Current status of mold immunotherapy, *Ann. Allergy*, 1991, **66**(5), 385–392.
- 12 S. Gupta, B. Pereira and A. Singh, Fomes pectinatis: an aeroallergen in India, *Asian Pac. J. Allergy Immunol.*, 1999, **17**(1), 1–7.
- 13 B. Fischer, N. Yawalkar, K. Brander, *et al.*, *Coprinus comatus* (shaggy cap) is a potential source of aeroallergen that may provoke atopic dermatitis, *J. Allergy Clin. Immunol.*, 1999, **104**(4 pt 1), 836–841.
- 14 K. Harley, J. Macher, M. Lipsett, *et al.*, Fungi and pollen exposure in the first months of life and risk of early childhood wheezing, *Thorax*, 2009, **64**(4), 353–358.
- 15 G. Canonica and M. Blaiss, Antihistaminic, anti-inflammatory, and antiallergic properties of the non-sedating second-generation antihistamine desloratadine: a review of the evidence, *World Allergy Organ. J.*, 2011, **4**(2), 47–53.
- 16 P. Panula, P. Chazot, M. Cowart, *et al.*, International Union of Basic and Clinical Pharmacology. XCVIII. Histamine Receptors, *Pharmacol. Rev.*, 2015, **67**(3), 601–655.

- 17 R. Leurs, M. Church and M. Taglialatela, H1-antihistamines: inverse agonism, anti-inflammatory actions and cardiac effects, *Clin. Exp. Allergy*, 2002, **32**(4), 489–498.
- 18 H. Lee, I. Chang, Y. Lee, *et al.*, Effect of antihistamine as an adjuvant treatment of isotretinoin in acne: a randomized, controlled comparative study, *J. Eur. Acad. Dermatol. Venereol.*, 2014, **28**(12), 1654–1660.
- 19 A. Layton, Top Ten List of Clinical Pearls in the Treatment of Acne Vulgaris, *Dermatol. Clin.*, 2016, **34**(2), 147–157.
- 20 H. Yoneyama, Efficient approaches to S-alkyl-N-alkylisothioureas: syntheses of histamine H3 antagonist clobenpropit and its analogues, *J. Org. Chem.*, 2008, **73**(6), 2096–2104.
- 21 T. Esbenshade, G. Fox, K. Krueger, *et al.*, Pharmacological and behavioral properties of A-349821, a selective and potent human histamine H3 receptor antagonist, *Biochem. Pharmacol.*, 2004, **68**(5), 933–945.
- 22 N. Ostrom, The history and progression of treatments for allergic rhinitis, *Allergy Asthma Proc.*, 2014, **35**(suppl. 3), S3–S10.
- 23 A. Jones, Perspectives in Drug Development and Clinical Pharmacology: The Discovery of Histamine H1 and H2 Antagonists, *Clin. Pharmacol. Drug Dev.*, 2016, **5**(1), 5–12.
- 24 S. Sicherer and H. Sampson, Food allergy: epidemiology, pathogenesis, diagnosis, and treatment, *J. Allergy Clin. Immunol.*, 2014, **133**(2), 291–307.
- 25 F. Simons, Anaphylaxis: recent advances in assessment and treatment, *J. Allergy Clin. Immunol.*, 2009, **124**(4), 625–636.
- 26 D. Finn and J. Walsh, Twenty-first century mast cell stabilizers, *Br. J. Pharmacol.*, 2013, **170**(1), 23–37.
- 27 J. May and W. Dolen, Management of Allergic Rhinitis: A Review for the Community Pharmacist, *Clin. Therapeut.*, 2017, **39**(12), 2410–2419.
- 28 L. Wheatley and A. Togias, Clinical practice. allergic rhinitis, *N. Engl. J. Med.*, 2015, **372**(5), 456–463.
- 29 S. Thomsen, Atopic dermatitis: natural history, diagnosis, and treatment, *ISRN Allergy*, 2014, **2014**, 1–7.
- 30 C. Anandan, U. Nurmatov, O. Schayck, *et al.*, Is the prevalence of asthma declining? Systematic review of epidemiological studies, *Allergy*, 2009, **65**(2), 152–167.
- 31 D. Golden, Insect sting anaphylaxis, *Immunol. Allergy Clin. North Am.*, 2007, **27**(2), 261–272.
- 32 J. Schafer, N. Mateo, G. Parlier, *et al.*, Penicillin allergy skin testing: what do we do now?, *Pharmacotherapy*, 2007, **27**(4), 542–545.
- 33 S. Ludman and R. Boyle, Stinging insect allergy: current perspectives on venom immunotherapy, *J. Asthma Allergy*, 2015, **8**, 75–86.
- 34 A. Grammatikos, The genetic and environmental basis of atopic diseases, *Ann. Med.*, 2008, **40**(7), 482–495.
- 35 R. Boyle, D. Ierodiakonou, T. Khan, *et al.*, Hydrolysed formula and risk of allergic or autoimmune disease: systematic review and meta-analysis, *BMJ*, 2015, **352**, i974.
- 36 K. Järvinen, K. Beyer, L. Vila, *et al.*, Specificity of IgE antibodies to sequential epitopes of hen's egg ovomucoid as a marker for persistence of egg allergy, *Allergy*, 2007, **62**(7), 758–765.
- 37 R. Panda, H. Ariyaratna, P. Amnuaycheewa, *et al.*, Challenges in testing genetically modified crops for potential increases in endogenous allergen expression for safety, *Allergy*, 2013, **68**(2), 142–151.
- 38 A. Gonzalez and C. Radojicic, Penicillin allergy: a practical guide for clinicians, *Cleve. Clin. J. Med.*, 2015, **82**(5), 295–300.
- 39 F. Marra, L. Lynd, M. Coombes, *et al.*, Does antibiotic exposure during infancy lead to development of asthma?: a systematic review and metaanalysis, *Chest*, 2006, **129**(3), 610–618.
- 40 S. Thavagnanam, J. Fleming, A. Bromley, *et al.*, A meta-analysis of the association between caesarean section and childhood asthma, *Clin. Exp. Allergy*, 2008, **38**(4), 629–633.
- 41 J. Zock, E. Plana, D. Jarvis, *et al.*, The use of household cleaning sprays and adult asthma: an international longitudinal study, *Am. J. Respir. Crit. Care Med.*, 2007, **176**(8), 735–741.
- 42 N. Dave, L. Xiang, K. Rehm, *et al.*, Stress and allergic diseases, *Immunol. Allergy Clin. North Am.*, 2011, **31**(1), 55–68.
- 43 C. Emanuelsson and M. Spangfort, Allergens as eukaryotic proteins lacking bacterial homologues, *Mol. Immunol.*, 2007, **44**(12), 3256–3260.
- 44 M. Grimaldeston, M. Metz, M. Yu, *et al.*, Effector and potential immunoregulatory roles of mast cells in IgE-associated acquired immune responses, *Curr. Opin. Immunol.*, 2006, **18**(6), 751–760.
- 45 J. Boyce, A. Assa'ad, A. Burks, *et al.*, Guidelines for the diagnosis and management of food allergy in the United States: report of the NIAID-sponsored expert panel, *J. Allergy Clin. Immunol.*, 2010, **126**(Suppl. 6), S1–S58.
- 46 F. Khan, A. Ueno, B. Serushago, *et al.*, Basophil activation test compared to skin prick test and fluorescence enzyme immunoassay for aeroallergen-specific Immunoglobulin-E, *Allergy, Asthma, Clin. Immunol.*, 2012, **8**(1), 1.
- 47 F. Greer, S. Sicherer and A. Burks, Effects of early nutritional interventions on the development of atopic disease in infants and children: the role of maternal dietary restriction, breastfeeding, timing of introduction of complementary foods, and hydrolyzed formulas, *Pediatrics*, 2008, **121**(1), 183–191.
- 48 D. Fleischer, J. Spergel, A. Assa'ad, *et al.*, Primary prevention of allergic disease through nutritional interventions, *J. Allergy Clin. Immunol.*, 2013, **1**(1), 29–36.
- 49 V. Garci, D. Ierodiakonou, K. Jarrold, *et al.*, Diet during pregnancy and infancy and risk of allergic or autoimmune disease: a systematic review and meta-analysis, *PLoS Med.*, 2018, **15**(2), e1002507.
- 50 C. Pelucchi, L. Chatenoud, F. Turati, *et al.*, Probiotics supplementation during pregnancy or infancy for the prevention of atopic dermatitis: a meta-analysis, *Epidemiology*, 2012, **23**(3), 402–414.
- 51 M. Netting, P. Middleton and M. Makrides, Does maternal diet during pregnancy and lactation affect outcomes in offspring? A systematic review of food-based approaches, *Nutrition*, 2014, **30**(11–12), 1225–1241.



- 52 G. Zhang, B. Liu, J. Li, *et al.*, Fish intake during pregnancy or infancy and allergic outcomes in children: a systematic review and meta-analysis, *Pediatr. Allergy Immunol.*, 2017, **28**(2), 152–161.
- 53 M. Frieri, Mast Cell Activation Syndrome, *Clin. Rev. Allergy Immunol.*, 2018, **54**(3), 353–365.
- 54 G. Canonica, J. Bousquet, T. Casale, *et al.*, Sub-lingual immunotherapy: World Allergy Organization Position Paper 2009, *Allergy*, 2009, **64**(suppl. 91), 1–59.
- 55 D. Bona, A. Plaia, M. Leto, *et al.*, Efficacy of Grass Pollen Allergen Sublingual Immunotherapy Tablets for Seasonal Allergic Rhinoconjunctivitis: A Systematic Review and Meta-analysis, *JAMA Intern. Med.*, 2015, **175**(8), 1301–1309.
- 56 C. Simpson, J. Newton, J. Hippisley, *et al.*, Incidence and prevalence of multiple allergic disorders recorded in a national primary care database, *J. R. Soc. Med.*, 2008, **101**(11), 558–563.
- 57 L. Cox, B. Williams, S. Sicherer, *et al.*, Pearls and pitfalls of allergy diagnostic testing: report from the American College of Allergy, Asthma and Immunology/American Academy of Allergy, Asthma and Immunology Specific IgE Test Task Force, *Ann. Allergy, Asthma, Immunol.*, 2008, **101**(6), 580–592.
- 58 S. Sicherer and D. Leung, Advances in allergic skin disease, anaphylaxis, and hypersensitivity reactions to foods, drugs, and insects, *J. Allergy Clin. Immunol.*, 2007, **119**(6), 1462–1469.
- 59 C. Galán, M. Smith, M. Thibaudon, *et al.*, Pollen monitoring: minimum requirements and reproducibility of analysis, *Aerobiologia*, 2014, **30**(4), 385–395.
- 60 J. Oteros, C. Galán, P. Alcázar, *et al.*, Quality control in bio-monitoring networks, Spanish Aerobiology Network, *Sci. Total Environ.*, 2013, **443**, 559–565.
- 61 G. Nieto, R. Márquez, A. García, *et al.*, The Histamine H3 Receptor: Structure, Pharmacology, and Function, *Mol. Pharmaceutics*, 2016, **90**(5), 649–673.
- 62 H. Andersen, J. Elberling and L. Arendt, Human surrogate models of histaminergic and non-histaminergic itch, *Acta Derm.-Venereol.*, 2015, **95**, 771–777.
- 63 P. Panula, P. Chazot, M. Cowart, *et al.*, International Union of Basic and Clinical Pharmacology. XCVIII. Histamine Receptors, *Pharmacol. Rev.*, 2015, **67**(3), 601–655.
- 64 M. Wouters, M. Vicario and J. Santos, The role of mast cells in functional GI disorders, *Gut*, 2015, **65**, 155–168.
- 65 P. Blandina, L. Munari, G. Provensi, *et al.*, Histamine neurons in the tuberomammillary nucleus: a whole center or distinct subpopulations, *Front. Syst. Neurosci.*, 2012, **6**, 1–6.
- 66 K. Yanai and M. Tashiro, The physiological and pathophysiological roles of neuronal histamine: an insight from human positron emission tomography studies, *Pharmacol. Therapeut.*, 2007, **113**(1), 1–15.
- 67 E. Alvarez, The role of histamine on cognition, *Behav. Brain Res.*, 2009, **199**(2), 183–189.
- 68 C. Ito, The role of the central histaminergic system on schizophrenia, *Drug News Perspect.*, 2004, **17**(6), 383–387.
- 69 F. Jadid and A. Mirshafiey, Histamine and histamine receptors in pathogenesis and treatment of multiple sclerosis, *Neuropharmacology*, 2010, **59**(3), 180–189.
- 70 P. Valent, H. Horny, L. Escribano, *et al.*, Diagnostic criteria and classification of mastocytosis: a consensus proposal, *Leuk. Res.*, 2001, **25**(7), 603–625.
- 71 W. Yue, W. Juan, L. Yan, *et al.*, Synthesis and antihistamine evaluations of novel loratadine analogues, *Bioorg. Med. Chem. Lett.*, 2011, **21**, 4454–4456.
- 72 J. Xiao, Y. Mei, F. Ji Hua, *et al.*, Synthesis, biological evaluation and SAR studies of benzimidazole derivatives as H1-antihistamine agents, *Chin. Chem. Lett.*, 2012, **23**, 707–710.
- 73 L. Gollapalle, P. Janaki, K. Nandakumar, *et al.*, Synthesis and antihistaminic activity of 3H-benzo [4,5] thieno [2,3-d] [1,2,3] triazin-4-ones, *Saudi Pharm. J.*, 2012, **20**, 45–52.
- 74 B. Satheesh, Y. Jinghua, A. Joe, *et al.*, Lead optimization of 2-(piperidin-3-yl)-1H-benzimidazoles: Identification of 2-morpholin- and 2-thiomorpholin-2-yl-1H-benzimidazoles as selective and CNS penetrating H1-antihistamines for insomnia, *Bioorg. Med. Chem. Lett.*, 2012, **22**, 421–426.
- 75 R. Dinneswara, J. Sae, W. Min, *et al.*, Syntheses and anti-allergic activity of 2-((bis(trimethylsilyl) methylthio/ methylsulfonyl) methyl)-5-aryl-1,3,4-oxadiazoles, *Eur. J. Med. Chem.*, 2013, **62**, 84–88.
- 76 M. Hiroyuki, N. Yuki, K. Shuhei, *et al.*, Maackiain is a novel antiallergic compound that suppresses transcriptional upregulation of the histamine H1 receptor and interleukin-4 genes, *Pharmacol. Res. Perspect.*, 2015, **3**(5), e00166.
- 77 M. Gobinath, S. Natesan, A. Veerachamy, *et al.*, Design, synthesis and H1-antihistaminic activity of novel 1-substituted-4-(3-chlorophenyl)-[1,2,4] triazolo [4,3-a] quinazolin-5 (4H)-ones, *J. Saudi Chem. Soc.*, 2015, **19**(3), 282–286.
- 78 A. Panayiotis, J. Alison, M. Paul, *et al.*, Design of phthalazinone amide histamine H1 receptor Antagonists for use in Rhinitis, *ACS Med. Chem. Lett.*, 2017, **8**(5), 577–581.
- 79 A. Saeed, S. Najma, S. Hina, *et al.*, Antihistaminic and other biological activities of 2-methylpropanamide and benzamide derivatives of carboxyterfenadine, *Arabian J. Chem.*, 2017, **10**(1), 114–120.

